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SUPPLEMENTARY INFORMATION

Incorporation of alkyl-halide initiator moiety into the frustule of *T. weissflogii*. Synchronized *T. weissflogii* cultures were inoculated in artificial seawater media enriched with F/2 Guillards media at either 1 x 10⁴ cells/mL or 1 x 10⁵ cells/mL in a final volume of 200 ml. Cultures received TEOS and CTEOS at a ratio of 3 TEOS:1 CTEOS such that the silicic acid concentration in culture was 200 μ M at each addition. Cultures received TEOS/CTEOS precursors at time of inoculation and at 48 hour intervals until time of collection at 192 hours post inoculation. Control cultures received Na₂SiO₃ at a final concentration of 200 μ M at each dosing time-point. Cell density was determined using a haemocytometer. Six flasks were inoculated for each treatment. Cultures were harvested by centrifugation 192 hours post inoculation. The organic casing of the diatom was removed by successive washes with de-ionized water and methanol. Diatoms were washed three times in de-ionized water, followed by three washes in de-ionized water heated to 60^oC. The final cleaning step involved a minimum of three washes in methanol until the pellet appeared white in color.

Characterization of TEOS/CTEOS *T. weissflogii* by energy dispersive X-ray spectroscopy coupled to scanning electron microscopy (EDX-SEM) and transmission electron microscopy (TEM). EDX-SEM analysis was performed using Hitachi S-4700 SEM with INCA® software. Cleaned diatoms suspended in methanol were allowed to air dry on a carbon stub and were subsequently gold coated. Diatoms were analyzed if the valve view was clearly visible. TEM images of frustules were collected using Hitachi H-7500 TEM with AMT image capture software. Cleaned diatoms suspended in methanol were allowed to air dry on a copper grid. Pore parameters (perimeter, width, length and area) were quantified using ImageJ software. The resolution of images analyzed was 0.5 nm/pixel. The mean pore parameters were calculated as follows; five sections per diatom were analyzed to calculate the mean pore parameters per diatom, three separate diatoms were analyzed to calculate the mean pore parameters for both *T. weissflogii* and TEOS/CTEOS *T. weissflogii*.

Statistical analysis. GraphPad Prism® software was used to perform statistical analysis. Details of statistical analysis performed are detailed in sections above. A two-way ANOVA followed by Bonferroni *post-hoc* analysis

was performed to determine statistical differences between treatment groups at p<0.05. One-way ANOVA was performed to investigate determine statistical differences between treatment groups at p<0.05.

Investigating the effect of ethanol on the growth profile of T. weissflogii

Methods: Synchronised *T. weissflogii* cultures were inoculated at 1 x 10^5 cells/mL in a final volume of 200 ml. Cultures received either Na₂SiO₃ or Na₂SiO₃ and ethanol at time of inoculation and at 48 hour intervals until time of collection. Na₂SiO₃ was added at a final concentration of 200 µM. Ethanol was added at a final concentration of 750 µM. Six flasks were inoculated for each treatment. A two-way ANOVA followed by Bonferroni *post-hoc* analysis was performed to determine statistical differences between treatment groups at p<0.05.

Results: As the hydrolysis of alkoxysilanes and organoalkoxysilanes releases alcohol, the potential toxicity of ethanol to *T. weissflogii* growth was investigated. Addition of TEOS/CTEOS mix to the culture media at a final silicic acid concentration of 200 μ M has the potential to generate a concentration of 750 μ M ethanol. Thus, the effect of addition of ethanol at a concentration of 750 μ M at 48 hour intervals on the growth profile of *T. weissflogii* was investigated. The presence of 750 μ M ethanol in the culture media did not adversely affect the growth profile of *T. weissflogii* (Supplementary Figure 1).



Fig. S1 - The growth profile of *T. weissflogii* grown in the presence of Na₂SiO₃ or Na₂SiO₃ + ethanol added at 48 hour intervals. Two-way ANOVA followed by Bonferroni *post-hoc* analysis revealed statistical difference. Data are shown as mean \pm sem (n = 6) ***p<0.001.