# Polyglycolic Acid from the Direct Polymerization of Renewable C1 Feedstocks

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# **Electronic Supplementary Information (ESI)**

**Supplementary Information Available:** Synthetic details and complete polymer characterization data.

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#### **General Considerations and Instrumentation**

Unless otherwise noted, all solvents were purified by stirring over calcium hydride for 24 hours and then vacuum transfer into an oven dried Straus flask. Xylenes were purchased from Sigma Aldrich and stored over molecular sieves. Solvents were purchased from Sigma Aldrich and utilized after further purification. Paraformaldehyde was purchased from Sigma Aldrich. Trioxane was purchased from Acros Organics and used after recrystallization in CHCl<sub>3</sub>. The initiators *para*-toluenesulfonic acid (*p*-TSA) and BF<sub>3</sub>•OEt<sub>2</sub> were purchased from Sigma Aldrich and used as received. Triflic acid (TfOH) was purchased from Oakwood Chemical and used as received. A Parr 4768Q 600 mL high pressure autoclave was used for the experiments with a magnetic stir bar.

Nuclear magnetic resonance (<sup>1</sup>H NMR and <sup>13</sup>C NMR) spectra were recorded using an Inova 500 MHz spectrometer. NMR sample preparation: 0.5 g of sample was dissolved in 0.4 g of HFIP, and 1.0 g of CDCl<sub>3</sub> (or C<sub>6</sub>D<sub>6</sub>) was added to the mixture.; HFIP- $d_2$  was also used for some NMR spectra. For <sup>1</sup>H NMR, number of scans = 32 with a 5 s relaxation delay. For <sup>13</sup>C NMR (125 MHz), number of scans = 10,000 with a 3 s relaxation delay. Chemical shifts are reported in parts per million (ppm) downfield relative to tetramethylsilane (TMS, 0.0 ppm) or residual protons in the specified solvent.

Differential scanning calorimetry thermograms were obtained with a DSC Q1000 from TA instruments. About 1.5-3 mg of each sample was weighed and sealed in a pan. Thermal history was established by a heat/cool/heat cycle at 10 °C/min, and the data were obtained for the second heating ramp.

Thermogravimetric analyses were performed under nitrogen with a TGA Q5000 from TA Instruments. About 5-10 mg of each sample was heated at 10 °C/min from RT to 500 °C.

Gel permeation chromatography (GPC) was performed at 40 °C using an Agilent Technologies 1260 Infinity Series liquid chromatography system with an internal differential refractive index detector, and two Waters Styragel HR-5E columns (7.8 mm i.d., 300 mm length, guard column 7.8 mm i.d., 25 mm length) using a solution of 0.1% potassium triflate (K(OTf)) in HPLC grade hexafluoroisopropanol (HFIP) as the mobile phase at a flow rate of 0.5 mL/min. Calibration was performed with narrow polydispersity polymethyl methacrylate (PMMA) standards.

**S**2

#### **Representative Polymerization Procedure for Table S1**

Under a nitrogen atmosphere, a mixture of trioxane (9.03 g, 0.1 mol), and an acid initiator (1 mmol) in 100 mL of a solvent was placed in a 600 mL high pressure reactor, charged with the desired pressure of CO, and heated to the desired reaction temperature. The reaction was stirred under CO pressure for the desired reaction time at the same temperature. After cooling to room temperature, the pressure was released and the mixture was poured into cold basic methanol. The product was isolated by filtration, washed with methanol and dichloromethane (DCM), and dried under vacuum.

PGA formation (Table S1 entry 17)= <sup>1</sup>H NMR (500 MHz, HFIP-CDCl<sub>3</sub>)  $\delta$  ppm 4.84 (s, 2H). <sup>13</sup>C NMR (125 MHz, HFIP-CDCl<sub>3</sub>)  $\delta$  ppm 168.4, 61.4.

POM formation (Table S1 entry 2)= <sup>1</sup>H NMR (500 MHz, HFIP-CDCl<sub>3</sub>) δ ppm 5.18 (s, 2H), 4.98 (s, 2H), 4.94 (s, 2H). <sup>13</sup>C NMR (125 MHz, HFIP-CDCl<sub>3</sub>) δ ppm 89.9.

### **General Polymerization Procedure for Table S2**

Under a nitrogen atmosphere, a mixture of trioxane (9.03 g, 0.1 mol), glycerol (0.073 mL, 1 mmol) and triflic acid (0.088 mL, 1 mmol) in 100 mL of DCM was placed in a 600 mL high pressure reactor, charged with 800 psi CO, and heated to the desired reaction temperature. The reaction was stirred under CO pressure for one day at the same temperature. After cooling to room temperature, the pressure was released and the mixture was poured into cold basic methanol. The product was isolated by filtration, washed with methanol and DCM, and dried under vacuum. <sup>1</sup>H NMR (500 MHz, HFIP-CDCl<sub>3</sub>) δ ppm 4.86 (s, 2H). <sup>13</sup>C NMR (125 MHz, HFIP-CDCl<sub>3</sub>) δ ppm 168.2, 60.9.

### **General Polymerization Procedure for Table S3**

Under a nitrogen atmosphere, a mixture of the desired amount of paraformaldehyde and triflic acid (1 mol%) in 100 mL of DCM was placed in a 600 mL high pressure reactor, charged with 800 psi CO, and heated to the desired reaction temperature. The reaction was stirred under CO pressure for three days at the same temperature. After cooling to room temperature, the pressure was released and the mixture was poured into cold basic methanol. The product was isolated by filtration, washed with methanol and DCM, and dried under vacuum.

### The Production of PGA from POM (entry 27)

Under a nitrogen atmosphere, a mixture of trioxane (9.03 g, 0.1 mol), and triflic acid (0.088 mL, 1 mmol) in 100 mL of DCM was placed in a 600 mL high pressure reactor. The reaction was stirred at room temperature for 12 hours. Then, without terminating the polymerization, the vessel was charged with 800 psi CO, and heated to 170 °C. The reaction

was stirred under CO pressure for three days. After cooling to room temperature, the pressure was released and the mixture was poured into cold basic methanol. The product was isolated by filtration, washed with methanol and DCM, and dried under vacuum. <sup>1</sup>H NMR (500 MHz, HFIP-CDCl<sub>3</sub>)  $\delta$  ppm 4.39 (s, 2H). <sup>13</sup>C NMR (125 MHz, HFIP-CDCl<sub>3</sub>)  $\delta$  ppm 168.2, 60.9.

## Polycondensation of Oligomeric Glycolic Acid (OGA) (entry 28)

A 2.0 g amount of OGA (from Table S1 entry 17) was weighed into a 50 mL reaction flask equipped with a magnetic stirrer. Then, 0.5 weight % of  $Zn(OAc)_2 \cdot 2H_2O$  relative to OGA was added. The mixture was heated at 200 °C with stirring under nitrogen for 2h, then heating continued under reduced pressure for 12 h. After cooling to room temperature, the mixture was poured into cold basic methanol. The product was isolated by filtration, washed with methanol and DCM to remove oligomeric residue, and dried under vacuum. Yield: 1.55 g brown solid. <sup>1</sup>H NMR (500 MHz, HFIP-CDCl<sub>3</sub>)  $\delta$  ppm 4.89 (s, 2H). <sup>13</sup>C NMR (125 MHz, HFIP-CDCl<sub>3</sub>)  $\delta$  ppm 168.3, 60.9.

## **Summary of Polymerization Data**

Entry <sup>a</sup>	CO (psi)	Initiator <sup>b</sup>	Solvent	τ <sub>ρ</sub> (°C)	Time	Yield (g)	Yield (%)	<i>M</i> <sub>w</sub> (g/mol)	M <sub>n</sub> (g/mol)	PDI	τ <sub>g</sub> (°C)	Т <sub>т</sub> (°С)
1	250	BF <sub>3</sub> •OEt <sub>2</sub>	DCM	RT	1 day	4.76	28	13,000	9,200	1.42	С	С
2	480	BF <sub>3</sub> •OEt <sub>2</sub>	DCM	RT	1 day	7.03	41	15,000	8,700	1.76	С	С
3	800	BF <sub>3</sub> •OEt <sub>2</sub>	DCM	100	1 day	NR	0	-	-	-	-	-
4	800	<i>p</i> -TSA	DCM	120	2 days	5.45	31	1,100	720	1.48	-22	133
5	800	TfOH	DCM	50	1 day	4.09	24	34,000	13,400	2.54	С	С
6	800	TfOH	DCM	60	1 day	4.26	25	38,000	12,400	3.05	С	С
7	800	TfOH	DCM	80	1 day	2.63	15	11,000	4,000	2.64	С	С
8	800	TfOH	DCM	100	1 day	2.51	14	1,000	740	1.43	С	115
9	800	TfOH	DCE	100	1 day	2.00	11	1,100	900	1.23	С	141
10	800	TfOH	Heptane	100	1 day	3.50	20	1,100	830	1.32	-18	119
11	800	TfOH	DCM	105	1 day	2.58	15	1,100	550	2.04	17	С
12	800	TfOH	DCM	130	1 day	3.53	20	1,600	1,100	1.50	3	С
13	800	TfOH	DCM	120	2 days	6.70	39	1,500	1,000	1.48	-39	143
14	800	TfOH	DCM	110	3 days	8.25	47	2,300	1,400	1.69	37	161
15	800	TfOH	DCM	130	3 days	11.00	63	1,500	1,000	1.48	С	164
16	800	TfOH	DCM	150	3 days	14.40	83	2,400	1,600	1.52	С	181
17	800	TfOH	DCM	170	3 days	16.00	92	3,200	1,800	1.73	13	192
18	800	TfOH	DCM	180	3 days	9.80	56	1,500	1,000	1.43	15	С
19 <sup>d</sup>	800	TfOH	HFIP	170	3 days	2.44	56	2,200	1,400	1.59	38	С
20	800	TfOH	Xylenes	130	3 days	NR	0	_	_	-	_	_

Table S1. Polymerization results: trioxane and carbon monoxide.

<sup>a</sup>Reactions conducted with 0.100 mol (9.03 g) of trioxane. <sup>b</sup>Initiator loading was 1 mol % of trioxane. <sup>c</sup>Missing thermal response ( $T_g$ ) or low decomposition temperatures ( $T_m$ ) prevented conclusive thermal analysis by DSC. <sup>d</sup>0.020 mol (1.8 g) of trioxane employed.

**Table S2.** Polymerization results: trioxane, carbon monoxide, and glycerol.



Entry <sup>a</sup>	<i>Τ</i> <sub>p</sub> ( <sup>o</sup> C)	Time	Yield (g)	Yield (%)	<i>M</i> <sub>n</sub> (g/mol)	PDI	<i>T</i> <sub>g</sub> (°C)	<i>T</i> <sub>m</sub> (°C)
21	100	1 day	7.12	40	1,700	1.56	45	172
22	150	1 day	14.2	81	3,100	1.83	50	206
23	150	2 days	14.0	80	3,700	2.46	47	205
24	170	1 day	5.02	28	1,600	1.37	51	b

<sup>a</sup>Reactions conducted with 0.100 mol (9.03 g) of trioxane. Initiator (TfOH) loading was 1.0 mol % of trioxane. <sup>b</sup>Low decomposition temperature prevented conclusive  $T_m$  analysis by DSC.

**Table S3.** Polymerization results: paraformaldehyde and carbon monoxide.

$$(CH_2O)_n$$
  $\xrightarrow{1 \text{ mol }\% \text{ TfOH}}$   $\xrightarrow{O}$ 

Entry	Paraformaldehyde (g)	<i>Τ</i> <sub>p</sub> ( <sup>°</sup> C)	Time	Yield (g)	<i>M</i> <sub>n</sub> (g/mol)	PDI	<i>T</i> <sub>g</sub> (°C)	<i>T</i> <sub>m</sub> (°C)	
25	3.0	100	1 day	4.4	1,000	1.30	-19	162	
26	9.0	170	3 days	6.2	1,500	1.49	51	а	
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<sup>a</sup>Low decomposition temperature prevented conclusive *T*<sub>m</sub> analysis by DSC







Scheme S2. High molecular weight PGA (entry 28) production from polycondensation of oligomeric glyocolic acid (OGA).

## **FTIR Spectra**







Figure S2. FTIR spectrum of polymer 1 (Table S1 entry 1).



Figure S3. FTIR spectrum of polymer 2 (Table S1 entry 2).



Figure S4. FTIR spectrum of polymer 4 (Table S1 entry 4).



Figure S5. FTIR spectrum of polymer 5 (Table S1 entry 5).



Figure S6. FTIR spectrum of polymer 6 (Table S1 entry 6).







Figure S8. FTIR spectrum of polymer 8 (Table S1 entry 8).



Figure S9. FTIR spectrum of polymer 9 (Table S1 entry 9).







Figure S11. FTIR spectrum of polymer 11 (Table S1 entry 11).



Figure S12. FTIR spectrum of polymer 12 (Table S1 entry 12).







Figure S14. FTIR spectrum of polymer 14 (Table S1 entry 14).



Figure S15. FTIR spectrum of polymer 15 (Table S1 entry 15).







Figure S17. FTIR spectrum of polymer 17 (Table S1 entry 17).



Figure S18. FTIR spectrum of polymer 18 (Table S1 entry 18).



Figure S19. FTIR spectrum of polymer 19 (Table S1 entry 19).



Figure S20. FTIR spectrum of polymer 21 (Table S2 entry 21).



Figure S21. FTIR spectrum of polymer 22 (Table S2 entry 22).







Figure S23. FTIR spectrum of polymer 24 (Table S2 entry 24).



Figure S24. FTIR spectrum of polymer 25 (Table S3 entry 25).







Figure S26. FTIR spectrum of polymer 27 (Scheme S1 entry 27).



Figure S27. FTIR spectrum of polymer 28 (Scheme S2 entry 28).

# **Thermogravimetric Analyses**



Figure S29. TGA Thermogram of polymer 1 (Table S1 entry 1).



Figure S30. TGA Thermogram of polymer 2 (Table S1 entry 2).



















Figure S38. TGA Thermogram of polymer 11 (Table S1 entry 11).





Figure S40. TGA Thermogram of polymer 13 (Table S1 entry 13).



Figure S41. TGA Thermogram of polymer 14 (Table S1 entry 14).











Figure S45. TGA Thermogram of polymer 18 (Table S1 entry 18).





Figure S47. TGA Thermogram of polymer 21 (Table S2 entry 21).











Figure S52. TGA Thermogram of polymer 26 (Table S3 entry 26).



Figure S53. TGA Thermogram of polymer 27 (Scheme S1 entry 27).



Figure S54. TGA Thermogram of polymer 28 (Scheme S2 entry 28).

# **Differential Scanning Calorimetry (DSC) Thermograms**



Figure S55. DSC Thermogram of commercial polyglycolic acid.



Figure S56. DSC Thermogram of polymer 1 (Table S1 entry 1).



Figure S57. DSC Thermogram of polymer 2 (Table S1 entry 2).



Figure S58. DSC Thermogram of polymer 4 (Table S1 entry 4).



Figure S59. DSC Thermogram of polymer 5 (Table S1 entry 5).





Figure S61. DSC Thermogram of polymer 7 (Table S1 entry 7).



Figure S62. DSC Thermogram of polymer 8 (Table S1 entry 8).



Figure S63. DSC Thermogram of polymer 9 (Table S1 entry 9).



Figure S64. DSC Thermogram of polymer 10 (Table S1 entry 10).



Figure S65. DSC Thermogram of polymer 11 (Table S1 entry 11).





Figure S67. DSC Thermogram of polymer 13 (Table S1 entry 13).



Figure S68. DSC Thermogram of polymer 14 (Table S1 entry 14).


Figure S69. DSC Thermogram of polymer 15 (Table S1 entry 15).



Figure S70. DSC Thermogram of polymer 16 (Table S1 entry 16).



Figure S71. DSC Thermogram of polymer 17 (Table S1 entry 17).





Figure S73. DSC Thermogram of polymer 19 (Table S1 entry 19).





Figure S75. DSC Thermogram of polymer 22 (Table S2 entry 22).





Figure S77. DSC Thermogram of polymer 24 (Table S2 entry 24).



Figure S78. DSC Thermogram of polymer 25 (Table S3 entry 25).



Figure S79. DSC Thermogram of polymer 26 (Table S3 entry 26).





Figure S81. DSC Thermogram of polymer 28 (Scheme S2 entry 28).

## <sup>1</sup>H NMR Spectra



Figure S82. <sup>1</sup>H NMR spectrum of HFIP solvent in CDCl<sub>3</sub>.



Figure S83. <sup>1</sup>H NMR spectrum of commercial polyglycolic acid in HFIP solvent and CDCl<sub>3</sub>.



**Figure S84.** <sup>1</sup>H NMR spectrum of polymer **1** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 1).



Figure S85. <sup>1</sup>H NMR spectrum of polymer 2 in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 2).



Figure S86. <sup>1</sup>H NMR spectrum of polymer **4** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 4).







Figure S88. <sup>1</sup>H NMR spectrum of polymer 6 in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 6).



Figure S89. <sup>1</sup>H NMR spectrum of polymer **7** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 7).



Figure S90. <sup>1</sup>H NMR spectrum of polymer 8 in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 8).



**Figure S91.** <sup>1</sup>H NMR spectrum of polymer **9** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 9).



Figure S92. <sup>1</sup>H NMR spectrum of polymer **10** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 10).



Figure S93. <sup>1</sup>H NMR spectrum of polymer **11** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 11).



Figure S94. <sup>1</sup>H NMR spectrum of polymer **12** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 12).



Figure S95. <sup>1</sup>H NMR spectrum of polymer **13** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 13).



Figure S96. <sup>1</sup>H NMR spectrum of polymer **14** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 14).



Figure S97. <sup>1</sup>H NMR spectrum of polymer **15** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 15).



Figure S98. <sup>1</sup>H NMR spectrum of polymer **16** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 16).



**Figure S99.** <sup>1</sup>H NMR spectrum of polymer **17** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 17).



**Figure S100.** <sup>1</sup>H NMR spectrum of polymer **18** in HFIP solvent and  $CDCI_3$  (Table S1 entry 18).



**Figure S101.** <sup>1</sup>H NMR spectrum of polymer **19** in HFIP solvent and  $CDCI_3$  (Table S1 entry 19).



**Figure S102.** <sup>1</sup>H NMR spectrum of polymer **21** in HFIP solvent and CDCl<sub>3</sub> (Table S2 entry 21).



**Figure S103.** <sup>1</sup>H NMR spectrum of polymer **22** in HFIP solvent and CDCl<sub>3</sub> (Table S2 entry 22).



**Figure S104.** <sup>1</sup>H NMR spectrum of polymer **23** in HFIP solvent and CDCl<sub>3</sub> (Table S2 entry 23).



**Figure S105.** <sup>1</sup>H NMR spectrum of polymer **24** in HFIP solvent and CDCl<sub>3</sub> (Table S2 entry 24).



**Figure S106.** <sup>1</sup>H NMR spectrum of polymer **25** in HFIP solvent and CDCl<sub>3</sub> (Table S3 entry 25).



**Figure S107.** <sup>1</sup>H NMR spectrum of polymer **26** in HFIP solvent and CDCl<sub>3</sub> (Table S3 entry 26).



**Figure S108.** <sup>1</sup>H NMR spectrum of polymer **27** in HFIP solvent and CDCl<sub>3</sub> (Scheme S1 entry 27).



**Figure S109.** <sup>1</sup>H NMR spectrum of polymer **28** in HFIP solvent and CDCl<sub>3</sub> (Scheme S2 entry 28).

## <sup>13</sup>C NMR Spectra





Figure S111. <sup>13</sup>C NMR spectrum of commercial polyglycolic acid in HFIP solvent and CDCl<sub>3</sub>.





Figure S113. <sup>13</sup>C NMR spectrum of polymer **2** in HFIP solvent and  $C_6D_6$  (Table S1 entry 2).



Figure S114. <sup>13</sup>C NMR spectrum of polymer 4 in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 4).



**Figure S115.** <sup>13</sup>C NMR spectrum of polymer **5** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 5).



Figure S116. <sup>13</sup>C NMR spectrum of polymer 6 in HFIP solvent and  $CDCl_3$  (Table S1 entry 6).



Figure S117. <sup>13</sup>C NMR spectrum of polymer 7 in HFIP solvent and  $CDCl_3$  (Table S1 entry 7).



Figure S118. <sup>13</sup>C NMR spectrum of polymer 8 in HFIP solvent and  $CDCl_3$  (Table S1 entry 8).



Figure S119. <sup>13</sup>C NMR spectrum of polymer 9 in HFIP solvent and  $CDCl_3$  (Table S1 entry 9).



Figure S120. <sup>13</sup>C NMR spectrum of polymer **10** in HFIP solvent and C<sub>6</sub>D<sub>6</sub> (Table S1 entry 10).



**Figure S121.** <sup>13</sup>C NMR spectrum of polymer **11** in HFIP solvent and  $CDCI_3$  (Table S1 entry 11).



**Figure S122.** <sup>13</sup>C NMR spectrum of polymer **12** in HFIP solvent and  $CDCI_3$  (Table S1 entry 12).



**Figure S123.** <sup>13</sup>C NMR spectrum of polymer **13** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 13).



**Figure S124.** <sup>13</sup>C NMR spectrum of polymer **14** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 14).



**Figure S125.** <sup>13</sup>C NMR spectrum of polymer **15** in HFIP solvent and  $CDCl_3$  (Table S1 entry 15).



**Figure S126.** <sup>13</sup>C NMR spectrum of polymer **16** in HFIP solvent and  $CDCI_3$  (Table S1 entry 16).



**Figure S127.** <sup>13</sup>C NMR spectrum of polymer **17** in HFIP solvent and CDCl<sub>3</sub> (Table S1 entry 17).



**Figure S128.** <sup>13</sup>C NMR spectrum of polymer **18** in HFIP solvent and  $C_6D_6$  (Table S1 entry 18).



**Figure S129.** <sup>13</sup>C NMR spectrum of polymer **19** in HFIP solvent and  $CDCI_3$  (Table S1 entry 19).



**Figure S130.** <sup>13</sup>C NMR spectrum of polymer **21** in HFIP solvent and CDCl<sub>3</sub> (Table S2 entry 21).



**Figure S131.** <sup>13</sup>C NMR spectrum of polymer **22** in HFIP solvent and CDCl<sub>3</sub> (Table S2 entry 22).



**Figure S132.** <sup>13</sup>C NMR spectrum of polymer **23** in HFIP solvent and CDCl<sub>3</sub> (Table S2 entry 23).



**Figure S133.** <sup>13</sup>C NMR spectrum of polymer **24** in HFIP solvent and  $CDCI_3$  (Table S2 entry 24).



Figure S134. <sup>13</sup>C NMR spectrum of polymer 25 in HFIP- $d_2$  (Table S3 entry 25).



**Figure S135.** <sup>13</sup>C NMR spectrum of polymer **26** in HFIP solvent and CDCl<sub>3</sub> (Table S3 entry 26).



**Figure S136.** <sup>13</sup>C NMR spectrum of polymer **27** in HFIP solvent and CDCl<sub>3</sub> (Scheme S1 entry 27).



Figure S137. <sup>13</sup>C NMR spectrum of polymer **28** in HFIP solvent and CDCl<sub>3</sub> (Scheme S2 entry 28).









Figure S140. GPC Chromatogram of polymer 2 in HFIP solvent (Table S1 entry 2).



Figure S141. GPC Chromatogram of polymer 4 in HFIP solvent (Table S1 entry 4).



Figure S142. GPC Chromatogram of polymer 5 in HFIP solvent (Table S1 entry 5).



Figure S143. GPC Chromatogram of polymer 6 in HFIP solvent (Table S1 entry 6).



Figure S144. GPC Chromatogram of polymer 7 in HFIP solvent (Table S1 entry 7).



Figure S145. GPC Chromatogram of polymer 8 in HFIP solvent (Table S1 entry 8).



Figure S146. GPC Chromatogram of polymer 9 in HFIP solvent (Table S1 entry 9).



Figure S147. GPC Chromatogram of polymer 10 in HFIP solvent (Table S1 entry 10).



Figure S148. GPC Chromatogram of polymer 11 in HFIP solvent (Table S1 entry 11).



Figure S149. GPC Chromatogram of polymer 12 in HFIP solvent (Table S1 entry 12).



Figure S150. GPC Chromatogram of polymer 13 in HFIP solvent (Table S1 entry 13).



Figure S151. GPC Chromatogram of polymer 14 in HFIP solvent (Table S1 entry 14).



Figure S152. GPC Chromatogram of polymer 15 in HFIP solvent (Table S1 entry 15).







Figure S154. GPC Chromatogram of polymer 17 in HFIP solvent (Table S1 entry 17).



Figure S155. GPC Chromatogram of polymer 18 in HFIP solvent (Table S1 entry 18).



Figure S156. GPC Chromatogram of polymer 19 in HFIP solvent (Table S1 entry 19).



Figure S157. GPC Chromatogram of polymer 21 in HFIP solvent (Table S2 entry 21).



Figure S158. GPC Chromatogram of polymer 22 in HFIP solvent (Table S2 entry 22).



Figure S159. GPC Chromatogram of polymer 23 in HFIP solvent (Table S2 entry 23).



Figure S160. GPC Chromatogram of polymer 24 in HFIP solvent (Table S2 entry 24).



Figure S161. GPC Chromatogram of polymer 25 in HFIP solvent (Table S3 entry 25).



Figure S162. GPC Chromatogram of polymer 26 in HFIP solvent (Table S3 entry 26).







Figure S164. GPC Chromatogram of polymer 28 in HFIP solvent (Scheme S2 entry 28).

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## **Computational Study for Copolymerization Thermodynamics**

Computational studies were performed with Spartan `10 for Macintosh, version 1.1.0.  $\Delta H$  values for the reactions were determined from heat of formations (HOF), determined by the **G3** (MP2) method. Entropy (*S*) and  $\Delta S$  values were determined by DFT B3LYP 6-311++G\*\*. The units for HOF,  $\Delta H$ , and  $\Delta G$  are kcal/mol and the units for *S* and  $\Delta S$  are cal/molK.





## Polymer Photographs



Figure S165. Photograph of commercial PGA (left) and entry 17 from Table S1 (right).



**Figure S166.** Photograph of mostly waxy, amorphous, oligomeric material (Table S1 entry 8) obtained according to the trioxane/CO copolymerization method described by G. Cevidalli, M. Ragazzini, and M. Modena, *U.S. Patent* 3,673,156, **1972**.