Copper(II) gluconate (a non-toxic food supplement/dietary aid) as a precursor catalyst for effective photo-induced living radical polymerisation of acrylates

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Figure S1: Typical set up for photo-induced polymerisation.



Figure S2: Molecular weight distribution of poly(methyl acrylate), M_n = 5500g/mol; D = 2.30; 96% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% v/v.



Figure S3: Molecular weight distribution of poly(methyl acrylate), M_n = 5400g/mol; D = 1.80; 98% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (pure)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% v/v.



Figure S4: Molecular weight distribution of poly(methyl acrylate), M_n = 5000g/mol; D = 1.50; 98% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h.



Figure S5a: Molecular weight distribution of poly(methyl acrylate), M_n = 5500g/mol; D = 1.19; 96% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 weeks.



Figure S5b: MALDI-ToF-MS reflectron mode spectrum of poly(methyl acrylate),obtained from photo-mediated polymerisation: [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 weeks.



Figure S5c: Monitoring effect of UV irradiation on Cu^(II) gluconate/Me₆-Tren in DMSO complex as a function of time by UV-vis spectroscopy.



Figure S6: Molecular weight distribution of poly(methyl acrylate), M_n = 5400g/mol; D = 1.38; 97% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% v/v, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 1 week.



Figure S7a: Molecular weight distribution of poly(methyl acrylate), M_n = 4900g/mol; D = 1.15; 97% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (pure)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h under UV irradiation.





Figure S7b: MALDI-ToF-MS reflectron mode spectrum of poly(methyl acrylate), obtained from the photo-mediated polymerisation: [MA]:[EBiB]:[Cu^(II) gluconate (pure)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h under UV irradiation.



Figure S8a: Molecular weight distribution of poly(methyl acrylate), M_n = 5600g/mol; D = 1.16; 98% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h under UV irradiation.



Figure S8b: ¹H NMR (400MHz, CDCl₃) of poly(methyl acrylate) obtained from UV experiment: [MA]:[EBiB]:[Cu^(II) gluconate]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% v/v.



Figure S9: Molecular weight distribution of poly(methyl acrylate), M_n = 3900g/mol; D = 1.33; 70% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (pure)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h under UV irradiation at 15 °C.



Figure S10: Molecular weight distribution of poly(methyl acrylate), M_n = 4200g/mol; D = 1.40; 75% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h under UV irradiation at 15 °C.



Figure S11a: Molecular weight distribution of poly(methyl acrylate), M_n = 4300g/mol; D = 1.18; 90% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (pure)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h at 60 °C.



Figure S11b: MALDI-ToF-MS reflectron mode spectrum, of poly(methyl acrylate), obtained for the photo-mediated polymerisation: [MA]:[EBiB]:[Cu^(II) gluconate (pure)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h at 60 °C.



Figure S12a: Molecular weight distribution of poly(methyl acrylate), M_n = 5200g/mol; D = 1.19; 95% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h at 60 °C.





Figure S12b: MALDI-ToF-MS reflectron mode spectrum, of poly(methyl acrylate), obtained for the photo-mediated polymerisation: [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren] = [50]:[1]:[0.02]:[0.12] in DMSO 50% *v/v*, pre-mixing of the Cu^(II) gluconate/Me₆-Tren complex for 2 h at 60 °C.



Figure S13a: Molecular weight distribution of poly(methyl acrylate), M_n = 5400g/mol; D = 1.15; 99% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren]:[NaBr] = [50]:[1]:[0.02]:[0.12];[0.04] in DMSO 50% v/v.



Figure S13b: MALDI-ToF-MS reflectron mode spectrum, of poly(methyl acrylate), obtained for the photo-mediated polymerisation: $[MA]:[EBiB]:[Cu^{(II)} \text{ gluconate (tablet)}]:[Me_6-Tren]:[NaBr] = [50]:[1]:[0.02]:[0.12]:[0.04] in DMSO 50% v/v.$



Figure S14a: Molecular weight distribution of poly(methyl acrylate), M_n = 5100g/mol; D = 1.12; 98% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (pure)]:[Me₆-Tren]:[NaBr] = [50]:[1]:[0.02]:[0.12]:[0.04] in DMSO 50% v/v.





Figure S14b: MALDI-ToF-MS reflectron mode spectrum, obtained from the photo-mediated polymerisation: $[MA]:[EBiB]:[Cu^{(II)} gluconate (pure)]:[Me_6-Tren]:[NaBr] = [50]:[1]:[0.02]:[0.12]:[0.04] in DMSO 50% v/v.$



Figure S15: ¹**H NMR** poly(methyl acrylate), 95% conversion. [MA]:[EBiB]:[Cu^(II) gluconate (tablet)]:[Me₆-Tren]:[NaBr] = [**200**]:[1]:[0.02]:[0.12]:[0.04] in DMSO 50% v/v.



Figure S16: ¹**H NMR for the block copolymerization** from a PMA macroinitiator. Initial conditions: $[MA]:[EBiB]:[Cu^{(II)} gluconate (tablet)]:[Me_6-Tren]:[NaBr] = [50]:[1]:[0.02]:[0.12]:[0.04], DMSO (50%,$ *v/v*). Chain extension achieved upon addition of an aliquot of PEGA (15 equiv.) in DMSO (33%,*v/v*).

Experimental

Materials

All materials were purchased from Sigma Aldrich or Fisher Scientific unless otherwise stated. The dietary supplement (purchased on the internet from "BioCare" with a stated 1.1 mg Cu per poll (110% RDA)), the analytical pure copper^(II) gluconate and ethyl 2-bromoisobutyrate (EBiB) were used as received. Methyl acrylate was passed through a basic Al₂O₃ chromatographic column prior to use. Tris-(2-(dimethylamino)ethyl)amine (Me₆-Tren) was synthesised according to previously reported literature.¹

Apparatus

¹H NMR spectra were recorded on Bruker DPX-300 or DPX-400 spectrometers in CDCl₃ unless otherwise stated. Chemical shifts are given in ppm downfield from the internal

standard tetramethylsilane. Size exclusion chromatography (SEC) measurements were conducted using an Agilent 1260 SEC-MDS fitted with differential refractive index (DRI), light scattering (LS) and viscometry (VS) detectors equipped with $2 \times PLgel 5 \text{ mm mixed-D}$ columns (300 \times 7.5 mm), 1 \times PLgel 5 mm guard column (50 \times 7.5 mm) and autosampler. Narrow linear poly(methyl methacrylate) standards in the range of 200 to 1.0×10^6 g·mol⁻¹ were used to calibrate the system. All samples were passed through 0.45 µm PTFE filter before analysis. The mobile phase was chloroform with 2% triethylamine eluent at a flow rate of 1.0 mL/min. SEC data was analysed using Cirrus v3.3 software with calibration curves produced using Varian Polymer laboratories Easi-Vials linear poly(methyl methacrylate) standards (200-4.7×10⁵ g/mol). MALDI-ToF mass spectrometry was conducted using a Bruker Daltonics Ultraflex II MALDI-ToF mass spectrometer, equipped with a nitrogen laser delivering 2 ns laser pulses at 337 nm with positive ion ToF detection performed using an accelerating voltage of 25 kV. Solutions in tetrahydrofuran (50 µL) of trans-2-[3-(4-tertbutylphenyl)-2-methyl-2-propylidene] malonitrile (DCTB) as a matrix (saturated solution), sodium iodide as cationisation agent (1.0 mg/mL) and sample (1.0 mg/mL) were mixed, and 0.7 µL of the mixture was applied to the target plate. Spectra were recorded in reflector mode calibrating PEG-Me 1100 kDa. UV/Vis spectra were recorded on Agilent Technologies Cary 60 UV-Vis spectrophotometer in the range of 200-1100 nm using a cuvette with 10 mm path length. A nail lamp was purchased online (λ ~365 nm) and used as the main UV source.

General procedure for the homopolymerisation of MA

Appropriate amounts of EBiB (1 eq.), MA (DP_n eq), Cu^(II) gluconate (0.02 eq.), Me₆-Tren (0.12eq.) and DMSO (50% ν/ν) were placed in a polymerisation flask, which was equipped with a magnetic stir bar and fitted with a rubber septum. The reaction mixture was degassed *via* bubbling with nitrogen for 20 min. The polymerization was allowed to proceed for 2 h under irradiation at λ ~365 nm. Samples were taken periodically for conversion and molecular weight analyses. The polymerisation mixture was initially dissolved in THF and then passed through a small basic Al₂O₃ chromatographic column to remove the copper salts. The resulting solution was precipitated in methanol.

In situ block copolymerisation

Filtered MA (1 mL, 11.1 mmol, 50 eq), EBiB (32 μ L, 0.22 mmol, 1 eq), Cu^(II) gluconate (tablet) (1.0 mg, 4.4 μ mol, 0.02 eq), Me₆-Tren (7 μ L, 22.0 μ mol, 0.12 eq) and DMSO (1 mL)

were added to a septum sealed vial and degassed by purging with nitrogen for 15 mins. Polymerisation commenced upon addition of the degassed reaction mixture to the UV lamp. After 90 min a 1: 0.5 mixture of degassed PEGA (15 eq) used for block copolymerization and DMSO was added to the reaction mixture *via* degassed syringe. Samples were taken periodically and conversions were measured using ¹H NMR and SEC analysis.

References

1. M. Ciampolini and N. Nardi, Inorg. Chem., 1966, 5, 41-44.