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Multifaceted Glycodendrimers With Programmable Bioactivity Through Convergent, Divergent, and Accelerated Approaches Using Polyfunctional Cyclotriphosphazenes

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Contents

1.	Materials and Methods	S2
2.	Synthetic protocols, Characterization, NMR, IR spectra, Mass, GPC, DLS traces	S4
3.	Diffusion NMR experiments	S182
4.	Competitive Surface Plasmon Resonance Studies and Sensorgrams	S185
5.	X-Ray crystallographic analysis	S195

1. Materials and methods

All reactions in organic medium were performed in standard oven dried glassware under an inert atmosphere of nitrogen using freshly distilled solvents. CH₂Cl₂ was distilled from CaH₂ and DMF from ninhydrin, and kept over molecular sieves. Solvents and reagents were deoxygenated when necessary by purging with nitrogen. Water used for lyophilization of final dendrimers was nanopure grade, purified through Barnstead NANOPure II Filter with Barnstead MegOhm-CM Sybron meter. All reagents were used as supplied without prior purification unless otherwise stated, and obtained from Sigma-Aldrich Chemical Co. Ltd.

LecA (Pseudomonas aeruginosa lectin-I) was purchased from Sigma-Aldrich (L9895-1MG, Lot 051M4011V).

Reactions were monitored by analytical thin-layer chromatography using silica gel 60 F254 precoated plates (E. Merck) and compounds were visualized by 254 nm light, a mixture of lodine/silica gel and/or mixture of Ceric Ammonium Molybdate solution (100 ml H_2SO_4 , 900 ml H_2O , 25g (NH₄)₆Mo₇O₂₄H₂O, 10g Ce(SO₄)₂) and subsequent development by gentle warming with a heat-gun. Purifications were performed by flash column chromatography using silica gel from Silicycle (60 Å, 40-63 µm) with the indicated eluent.

¹H NMR and ¹³C NMR spectra were recorded at 300 or 600 MHz and 75 or 150 MHz, respectively, on a Bruker spectrometer (300 MHz) and Varian spectrometer (600 MHz). All NMR spectra were measured at 25℃ in indicated deuterated solvents. Proton and carbon chemical shifts (δ) are reported in ppm and coupling constants (J) are reported in Hertz (Hz). The resonance multiplicities in the ¹H NMR spectra are described as "s" (singlet), "d" (doublet), "t" (triplet), "quint" (quintuplet) and "m" (multiplet) and broad resonances are indicated by "br". Residual protic solvent of CDCl₃ (¹H, δ 7.27 ppm; ¹³C, δ 77.0 ppm (central resonance of the triplet)), D₂O (¹H, δ4.79 ppm and 30.9 ppm for CH₃ of Acetone for ¹³C spectra of de-O-acetylated compounds), MeOD (¹H, δ 3.31 ppm and ¹³C, δ 49.0 ppm. 2D Homonuclear correlation ¹H-¹H COSY and Heteronuclear correlation ¹H-¹³C HETCOR experiments were used to confirm NMR peak assignments. Characteristic signals of protected and deprotected peripheric lactosides at the glycodendrimers' periphery were assigned in comparison with corresponding monovalent reference previously described.¹ 2D Gel Permeation Chromatography (GPC) was performed using THF or CHCl₃/Et₃N (1%) as the eluent, at 40°C with a 1 mL/min flow rate on a Viscotek VE 2001 GPCmax (SEC System) with Wyatt DSP/Dawn EOS and refractive index RI/LS system as detectors. 2 PLGel mixed B LS (10 µm, 300×7.5 mm) and LS-MALLS detection with performances verified with polystyrene 100 kDa and 2000 kDa were used to determine the number-average molecular weight (M_n) and polydispersity index (M_W/M_n). Calculations were performed with Zimm Plot (model). Fourier transform infrared (FTIR) spectra were obtained with Thermo-scientific, Nicolet model 6700 equipped with ATR. The absorptions are given in wavenumbers (cm⁻¹). The intensity of the bands is described as s (strong), m (medium) or w (weak). Melting points were measured on a Electrothermal MEL-TEMP apparatus and are uncorrected.

Accurate mass measurements (HRMS) were performed on a LC-MSD-TOF instrument from Agilent Technologies in positive electrospray mode. Low-resolution mass spectra were performed on the same apparatus or on a LCQ

¹ V. Percec et al., J. Am. Chem. Soc., **2013**, 135, 9055-9077.

Advantage ion trap instrument from Thermo Fisher Scientific in positive electrospray mode (Mass Spectrometry Laboratory (Université de Montréal), or Plateforme analytique pour molécules organiques (Université du Québec à Montréal), Québec, Canada). Either protonated molecular ions [*M*+nH]ⁿ⁺ or adducts [*M*+nX]ⁿ⁺ (X = Na, K, NH₄) were used for empirical formula confirmation. MALDI-TOF analyses were performed in either reflectron or linear mode on an Ultraflextreme TOF/TOF instrument from Bruker Daltonics. Mass spectra were acquired over the appropriate mass range for every particular sample. Each mass spectrum represents the sum of minimum 1000 laser shots. Samples were solubilized in either dichloromethane or water to an approximate final concentration of 6 mg/mL. Dithranol and DHB at 10 mg/mL in methanol containing 0.1 % TFA were used as matrices while NaTFA at 2 mg/mL in methanol was used as ionizing agent. A mixture of 20 uL matrix, 20 uL sample and 10 uL ionizing agent was prepared in a 600 uL Eppendorf tube. Aliquots of 1 uL of the above solution were applied on the MALDI plate and allowed to dry. All solvents for Mass Analyses (Water, Dichlorometane, Methanol and Acetonitrile) were HPLC grade, from de J.T. Baker (Phillipsburg, NJ). Sodium trifluoroacetate (NaTFA), dithranol andtrifluoroacetic (TFA) acid were from Sigma (St Louis, MO) while DHB was from Bruker Daltonics (Billerica, MA).

2. Synthetic protocols, Characterization, NMR, IR spectra, Mass, GPC, DLS traces

Hexapropargylated cyclotriphosphazene 1



Hexachlorophosphazene $N_3P_3Cl_6$ (freshly recrystallized from Hexanes, 29.3 mg, 84.4 µmol, 1.0 eq.) and 4propargyloxyphenol 4^2 (150.0 mg, 1.010 mmol, 12.0 eq.) were dissolved in 4 mL of anhydrous THF. Under nitrogen atmosphere, Cs_2CO_3 (620.0 mg, 1.900 mmol, 22.5 eq.) was added and the mixture was stirred at reflux temperature (66°C) for 18 hours. The solution was filtered and washed with DCM. The filtrate was concentrated under reduced pressure. Column chromatography on silica (EtOAc/Hexanes 10:90 to 70:30) afforded the desired compound **1** (71.1 mg, 69.9 µmol, **83%**) as a white solid.

Crystallization conditions: 130 mg of amorphous **1** (white solid) are dissolved in a $Et_2O/EtOH_{abs.}$ mixture (5 mL/1.5 mL) at room temperature and 6 mL of Hexanes are added. The mixture containing the resulting white precipitate was refluxing at 75°C for 30 minutes (total dissolution was observed at 65°C). After gentle cooling, colorless plates were obtained at room temperature.

 $\mathbf{R}_{f} = 0.23$, EtOAc/Hexanes 35:65.

m.p. = 64° C (recryst. from Et₂O/EtOH_{abs}/Hexanes (5:1.5:6)).

¹**H NMR** (300 MHz, CDCl₃, δ ppm): 6.81 (dd, 24H, *J* = 9.0 Hz, C*H*_b, C*H*_c), 4.65 (d, 12H, *J* = 2.4 Hz; OC*H*₂C≡CH), 2.54 (t, 6H, *J* = 2.4 Hz; OCH₂C≡C*H*).

¹³C NMR (75 MHz, CDCl₃, δ ppm): 154.5 (C_a), 144.8 (C_d), 121.8 (C_c), 115.5 (C_b), 78.5 (C≡CH), 75.7 (C≡CH), 56.1 (OCH₂).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.9 (s, 3P).

m/z (MALDI-MS) for C₅₄H₄₂N₃O₁₂P₃ = 1017.9; found 1017.2 (ESI⁺-HRMS) 1018.20541 [*M*+H]⁺; found 1018.20476; 1040.18735 [*M*+Na]⁺; found 1040.18735.

² M. Srinivasan, S. Sankararaman, H. Hopf, I. Dix and P.G. Jones, J. Org. Chem., 2001, 66, 4299–4303.



Figure S1. ¹H NMR spectrum of compound 1 (CDCl₃, 300MHz)



Figure S2. gCOSY spectrum of compound 1



Figure S3. $^{\rm 13}\text{C}$ NMR spectrum of compound 1 (CDCl_3, 75MHz)



Figure S4. ³¹P NMR spectrum of compound 1 (CDCl₃, 122MHz)



Figure S5. Mass spectrometry of compound 1 (up: MALDI-TOF MS and bottom: ESI⁺-HRMS)



Hexachlorophosphazene (freshly recrystallized from Hexanes, 1.200 g, 3.450 mmol, 2.0 eq.) and *N*Boc-protected *p*-aminophenol derivative 2^3 (361.0 mg, 1.725 mmol, 1.0 eq.) were dissolved in 125 mL of anhydrous THF. Under nitrogen atmosphere, Cs₂CO₃ (2.810 g, 8.630 mmol, 5.0 eq.) was added and the mixture was stirred at reflux temperature (66°C) for 24 hours. The solution was filtered and washed with DCM. The filtrate was concentrated under reduced pressure. Column chromatography on silica (DCM/Hexanes 10:90 to 70:30) afforded the desired compound **3** (538.0 mg, 1.034 mmol, **60%**) as a colorless oil.

 $\mathbf{R}_{f} = 0.61$, EtOAc/Hexanes 20:80.

¹**H NMR** (300 MHz, CDCl₃, δ ppm): 7.40 (d, 2H, *J* = 9.0 Hz, C*H*_b), 7.18 (d, 2H, *J* = 9.0 Hz, C*H*_c), 6.62 (br s, 1H, N*H*), 1.52 (s, 9H, C*H*₃).

¹³**C NMR** (75 MHz, CDCl₃, δ ppm): 152.5 (*C*=O), 144.4 (*C*_d, d, *J*_{P-C} = 10.4 Hz), 137.0 (*C*_a, d, *J*_{P-C} = 2.7 Hz), 121.8 (*C*_b, d, *J*_{P-C} = 5.0 Hz), 119.5 (*C*_c), 80.9 (*C*_q), 28.3 (*C*H₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 22.4 (d, 2P, ²*J*(P,P) = 59.3 Hz, *P*Cl₂), 12.8 (t, 1P, ²*J*(P,P) = 59.3 Hz, Cl-*P*-O). *m/z* (ESI⁺-HRMS) for C₁₁H₁₄Cl₅N₄O₃P₃ = 540.8614 [*M*+Na]⁺; found 540.8628.



³ N. Jain, Y. Arntz, V. Goldschmidt, G. Duportail, Y. Mely and A. S. Klymchenko, *Bioconjugate Chem.*, 2010, **21**, 2110–2118.





– S9 –



Figure S9. ESI⁺-HRMS spectrum of compound 3



Monofunctionalized derivative **3** (50.0 mg, 0.0961 mmol, 1.0 eq.) and 4-propargyloxyphenol **4** (113.9 mg, 0.7690 mmol, 8.0 eq.) were dissolved in 8 mL of anhydrous THF. Under nitrogen atmosphere, Cs_2CO_3 (1.50 g, 4.81 mmol, 48.0 eq.) was added and the mixture was stirred at reflux temperature (66°C) for 6 hours. The solution was filtered and washed with DCM. The filtrate was concentrated under reduced pressure. Column chromatography on silica (EtOAc/Hexanes 10:90 to 35:65) afforded the desired compound **5** (88.0 mg, 0.0816 mmol, **87%**) as a colorless oil.

 $\mathbf{R}_{f} = 0.36$, EtOAc/Hexanes 35:65.

¹**H NMR** (300 MHz, CDCl₃, δ ppm): 7.18 (d, 2H, J = 9.0 Hz, CH_b), 6.87-6.76 (m, 22H, CH_b, CH_c, CH_c), 6.54 (br s, 1H, NH), 4.65 (t_{app}, 10H, OCH₂C≡CH), 2.55 (t_{app}, 5H, C≡CH), 1.52 (s, 9H, CH₃).

¹³**C NMR** (75 MHz, CDCl₃, δ ppm): 154.4 (C_a), 152.6 (C=O), 145.8 (C_d), 144.7 (C_d), 135.1 (C_a), 121.8 (C_c), 121.3 (C_b), 119.5 (C_c), 115.4 (C_b), 80.5 (C_q), 78.5 (C=CH), 78.4 (C=CH), 75.7 (C=CH), 56.1 (OCH₂), 28.3 (CH₃).

 $^{31}\textbf{P}$ NMR (122 MHz, CDCl₃, δ ppm): 9.81 (s_{app}, 3P).

m/z (ESI⁺-HRMS) for C₅₆H₄₉N₄O₁₃P₃ = 1079.2582 [*M*+H]⁺; found 1079.2556.



Figure S12. ³¹P NMR spectrum of compound 5 (CDCl₃, 122MHz)



Figure S13. ESI⁺-HRMS spectrum of compound 5



Dissymetric hexasubstitued protected phosphazene derivative **5** (158.0 mg, 0.1460 mmol, 1.0 eq.) was dissolved in 3.5 mL of anhydrous DCM under nitrogen atmosphere and at 0°C was added dropwise 550 μ L of trifluoroacetic acid (TFA) over a 30 minutes period. After stirring 3 hours at rt, the solvent was removed under reduced pressure and coevaporated with toluene. The residue was dissolved in 3 mL of anhydrous DCM and, under nitrogen atmosphere, was added 100 μ L of DIPEA (74.7 mg, 0.578 mmol, 4.0 eq.). At 0°C was added dropwise chloroacetylchoride^{*} (34.5 μ L, 49.0 mg, 0.433 mmol, 3.0 eq.) in 1 mL of anhydrous DCM over a 15 minutes period. After stiring at rt for 18 hours, the solvent was removed under reduced pressure and the residue was dissolved in 25 mL of EtOAc and washed with HCl 1M (2×20 mL), NH₄Cl (20 mL), then water (10 mL). The organic phase was dried over Na₂SO₄, and concentrated under reduced pressure. Column chromatography on silica (EtOAc/Hexanes 20:80 to 45:55) afforded the desired compound **6** (112.0 mg, 0.1061 mmol, **72%**) as a yellowish oil.

 $\mathbf{R}_{\mathbf{f}} = 0.39$, EtOAc/Hexanes 50:50.

¹**H NMR** (300 MHz, CDCl₃, δ ppm): 8.24 (br s, 1H, N*H*), 7.33 (d, 2H, J = 9.0 Hz, CH_b), 6.88-6.76 (m, 22H, CH_b , CH_c , CH_c), 4.65 (t_{app} , 10H, OC H_2 C=CH), 4.19 (s, 2H, CH_2 Cl), 2.55 (t_{app} , 5H, C=CH).

¹³C NMR (75 MHz, CDCl₃, δ ppm): 163.7 (*C*=O), 154.4 (*C*_a), 147.3 (*C*_d), 144.7 (*C*_d), 133.4 (*C*_a), 121.7 (*C*_c), 121.5 (*C*_b), 121.2 (*C*_c), 115.4 (*C*_b), 78.5 (*C*≡CH), 78.7 (*C*≡CH), 75.7 (C≡CH), 56.1 (OCH₂), 42.8 (CH₂Cl).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.69 (s_{app}, 3P).

m/z (ESI⁺-HRMS) for C₅₃H₄₂CIN₄O₁₂P₃ = 1055.1773 [*M*+H]⁺; found 1055.1758.

^{*} The use of bromoacetylchloride at this step provides a mixture of halogenated (Cl/Br) derivatives after the same steps of purification.











Figure S17. ³¹P NMR spectrum of compound 6 (CDCl₃, 122MHz)



Figure S18. ESI⁺-HRMS spectrum of compound 6

Mixture Br/Cl (ratio of ~ 20:80) with the use of bromoacetylchloride:



Figure S19. ¹H NMR spectrum of the Cl/Br mixture of compound 6 (CDCl₃, 300MHz)



Les calculs sont effectués sur les adduits de sodium, [M+Na]⁺, des 2 composés, soit à m/z 1077 pour le produit chloré et à m/z 1121 pour le produit bromé.



En utilisant la somme des intensités pour les 5 premiers pics de chaque patron isotopique, nous obtenons un ratio de 78.6% de produit chloré pour 21.4% de produit bromé.

Figure S21. Calculation of CI/Br ratio from ESI⁺-HRMS of compound 6 when using bromoacetyl chloride as reactant



Figure S22. ESI⁺-HRMS spectrum of compound 6 when using bromoacetyl chloride as reactant



Hexachlorophosphazene (freshly recrystallized from Hexanes, 518.6 mg, 1.490 mmol, 2.0 eq.) and 4propargyloxyphenol **4** (110.5 mg, 0.745 mmol, 1.0 eq.) were dissolved in 50 mL of anhydrous THF. Under nitrogen atmosphere, Cs_2CO_3 (2.42 g, 7.44 mmol, 10.0 eq.) was added and the mixture was stirred at reflux temperature (66°C) for 22 hours. The solution was filtered and washed with DCM. The filtrate was concentrated under reduced pressure. Column chromatography on silica (DCM/Hexanes 20:80 to 75:25) afforded the desired compound **7** (243.6 mg, 0.530 mmol, **71%**) as a colorless oil.

 $\mathbf{R}_{f} = 0.66$, EtOAc/Hexanes 20:80.

¹**H NMR** (300 MHz, CDCl_3 , δ ppm): 7.15-7.11 (m, 2H, CH_b), 6.93-6.89 (m, 2H, CH_c), 4.61 (d, J = 2.4 Hz, 2H, $\text{OCH}_2\text{C}\text{=C}$), 2.46 (t, 1H, J = 2.4 Hz, C=CH).

¹³**C** NMR (75 MHz, CDCl₃, δ ppm): 155.8 (C_a , d, $J_{P-C} = 2.6$ Hz), 143.3 (C_d , d, $J_{P-C} = 10.5$ Hz), 122.4 ($C_{c,}$ d, $J_{P-C} = 5.0$ Hz), 116.0 (C_b , d, $J_{P-C} = 2.2$ Hz), 78.1 ($C \equiv CH$), 76.0 ($C \equiv CH$), 56.3 ($OCH_2C \equiv C$).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 22.4 (d, 2P, ²*J*(P,P) = 59.2 Hz, *P*Cl₂), 12.9 (t, 1P, ²*J*(P,P) = 59.3 Hz, Cl-*P*-O).

m/z (ESI⁺-HRMS) for C₉H₇Cl₅N₃O₂P₃ = 457.8267 [*M*+H]⁺; found 457.8280, 479.8086 [*M*+Na]⁺, found 479.8093.







Figure S25. ³¹P NMR spectrum of compound 7 (CDCl₃, 122MHz)



Figure S26. ESI⁺-HRMS spectrum of compound 7



The monofunctionalized derivative **7** (74.7 mg, 0.163 mmol, 1.0 eq.) and protected *p*-aminophenol derivative **2** (278.0 mg, 1.329 mmol, 8.1 eq.) were dissolved in 10 mL of anhydrous THF. Under nitrogen atmosphere, Cs_2CO_3 (2.13 g, 6.53 mmol, 40.0 eq.) was added and the mixture was stirred at reflux temperature (66°C) for 18 hours. The solution was filtered and washed with DCM. The filtrate was concentrated under reduced pressure. Column chromatography on silica (EtOAc/Hexanes 5:95 to 35:65) afforded the desired compound **8** (180.0 mg, 0.136 mmol, **83%**) as an off-white foam.

 $\mathbf{R}_{\mathbf{f}} = 0.42$, EtOAc/Hexanes 4:6.

¹**H NMR** (300 MHz, CDCl₃, δ ppm): 7.07-7.00 (m, 10H, C*H*_b), 6.82-6.64 (m, 19H, C*H*_b, C*H*_c, C*H*_c', N*H*), 4.58 (d, J = 2.4 Hz, 2H, OC*H*₂C≡CH), 2.46 (t, 1H, J = 2.4 Hz, C≡C*H*), 1.46 (s, 27H, C*H*₃), 1.45 (s, 18H, C*H*₃).

¹³C NMR (75 MHz, CDCl₃, δ ppm): 154.6 (*C*_a), 153.3 (*C*=O), 153.1 (*C*=O), 146.0 (*C*_d), 144.8 (*C*_d), 135.1 (*C*_a), 135.0 (*C*_a), 122.0 (*C*_c), 121.3 (*C*_c), 121.2 (*C*_c), 120.7 (*C*_b), 120.3 (*C*_b), 115.5 (*C*_b), 80.5 (*C*_q), 78.7 (*C*=CH), 75.7 (C=CH), 56.2 (OCH₂), 28.5 (CH₃), 28.5 (CH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.9 (t, ${}^{2}J$ = 9.3 Hz, 3P).

m/z (ESI⁺-HRMS) for C₆₄H₇₇N₈O₁₇P₃ = 1323.4692 [M+H]⁺; found 1323.4664, 1345.4512 [M+Na]⁺, found 1345.4484.







Figure S29. ³¹P NMR spectrum of compound 8 (CDCl₃, 122MHz)







Figure S31. ESI⁺-HRMS spectrum of compound 8



Dissymetric hexasubstitued protected phosphazene derivative **8** (100.5 mg, 0.076 mmol, 1.0 eq.) was dissolved in 5 mL of anhydrous DCM under nitrogen atmosphere and at 0°C. Trifluoroacetic acid (TFA, 3 mL) was added dropwise over a 35 minutes period. After stirring 5 hours at rt, the solvent was removed under reduced pressure and coevaporated with toluene. The residue was dissolved in 2 mL of anhydrous DCM and, under nitrogen atmosphere was added 159 μ L of DIPEA (117.8 mg, 0.911 mmol, 12.0 eq.). At 0°C was added dropwise chloroacetylchoride (60.4 μ L, 85.8 mg, 0.760 mmol, 10.0 eq.) in 1.5 mL of anhydrous DCM over a 2 hours period. After stiring at rt for 13 hours, the solvent was removed under reduced pressure and the residue was dissolved in 25 mL of EtOAc and washed with HCl 1M (2x20 mL), NH₄Cl (20 mL), then water (10 mL). The organic phase was dried over Na₂SO₄, and concentrated under reduced pressure. Column chromatography on silica (EtOAc/Hexanes 40:60 to 100:0) afforded the desired compound **9** (44.0 mg, 0.0370 mmol, **48%**).

R_f = 0.56, DCM/MeOH 10:90.

¹**H NMR** (300 MHz, MeOD, δ ppm): 7.46-7.37 (dd, 10H, J = 11.2 Hz, J = 9.0 Hz, CH_b), 6.90-6.78 (m, 14H, CH_b , CH_c , CH_c), 4.69 (d, 2H, J = 2.4 Hz, OCH_2C =CH), 4.25 (2×s, 10H, CH_2C I), 2.95 (t, J = 2.4 Hz, 1H, C=CH).

¹³C NMR (75 MHz, MeOD, δ ppm): 167.4 (*C*=O), 156.4 (*C*_a), 148.1 (*C*_{d'}), 148.0 (*C*_d), 136.5 (*C*_{a'}), 122.8 (*C*_c), 122.9 (*C*_{b'}), 122.7 (*C*_{b'}), 122.4 (*C*_{c'}), 116.8 (*C*_b), 79.9 (*C*=CH), 77.1 (C=CH), 57.2 (OCH₂), 44.3 (CH₂CI), 44.2 (CH₂CI). ³¹P NMR (122 MHz, MeOD, δ ppm): 9.6 (t, ²*J* = 17.9 Hz, 3P).

m/z (ESI⁺-HRMS) for C₄₉H₄₂Cl₅N₈O₁₂P₃ = 624.0181 [*M*+2Na]²⁺, found 624.0190, 1225.0470 [*M*+Na]⁺; found 1225.0463.



Figure S32. ¹H NMR spectrum of compound 9 (CD₃OD, 300MHz)



Figure S33. gCOSY spectrum of compound 9







									<u> </u>		
1			1		1				1	1	1
650	700	750	800	850	900	950	1000	1050	1100	1150	1200
			Co	ounts vs	. Mass-t	o-Charg	je (m/z)				

MS Spectrum Peak List

Ion	Formula	Abund	Observed m/z	Calc m/z	Diff(ppm)
(M+2Na)+2	C49H42Cl5N8Na2O12P3	1074.44	624.01899	624.0181	0.89
(M+Na)+	C49H42Cl5N8NaO12P3	9275.75	1225.04633	1225.04699	-0.66

Figure S36. ESI⁺-HRMS spectrum of compound 9

Trivalent dendron 12



To a stirring solution of 10^1 (570 mg, 680 mmol, 3.45 eq.) and tripropargylated synthon 11^4 (70.0 mg, 197 mmol, 1.00 eq.) in dry THF (6 mL) were added 6 mL of H₂O and a mixture of CuSO₄·5H₂O (44.7 mg, 179 mmol, 0.90 eq.) and Sodium ascorbate (35.5 mg, 179 mmol, 0.90 eq.). After stirring for 3 hours at 50°C in a 20 mL vial, the reaction was left stirring overnight at room temperature. EtOAc (50 mL) was added and the solution was washed successively with a saturated aqueous solution of NH₄Cl (3×25 mL), water (2×20 mL) and brine (1×10 mL). The organic phase was then dried over MgSO₄ and concentrated under reduced pressure. Column chromatography on silica (DCM/MeOH 98:2 to 94:6) afforded the desired compound **12** (300 mg, 108 mmol, **55%**) as a colorless oil.

R_f = 0.17, DCM/MeOH 95:5

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 7.71 (s, 3H, $H_{triazole}$), 7.05 (br s, 1H, N*H*COCH₂Br), 6.80 (br s, 1H, N*H*COCH₂Cl), 5.33 (d_{app}, 3H, H_{4gal}), 5.17 (dd, ³*J*_{4,3} = 9.4 Hz, ³*J*_{3,2} = 9.1 Hz, 3H, H_{3glc}), 5.10 (dd, ³*J*_{2,1} = 10.5 Hz, ³*J*_{3,2} = 8.0 Hz, 3H, H_{2gal}), 4.97 (dd, ³*J*_{2,3} = 7.0 Hz, ³*J*_{3,4} = 3.4 Hz, 3H, H_{3gal}), 4.88 (dd, ³*J*_{2,1} = 9.4 Hz, ³*J*_{3,2} = 8.0 Hz, 3H, H_{2glc}), 4.57-4.47 (m, 21H, C_qC*H*₂O, OCH₂C*H*₂N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.12-4.08 (m, 9H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.89-3.57 (m, 59H, C*H*₂Br/Cl, OC*H*₂C*H*₂N, OC*H*₂, H_{4glc} , H_{5glc}), 2.18-1.97 (m, 63H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.3, 170.1, 170.0, 169.7, 169.6, 169.0 (7×s, COCH₃), 166.7 (CO), 144.4 ($C_{triazole}$ =CH), 123.7 (C= $C_{triazole}$ H), 101.0 (C_{1gal}), 100.5 (C_{1glc}), 76.2 (C_{4glc}), 72.7 (C_{3glc}), 72.5 (C_{5glc}), 71.5 (C_{2glc}), 70.9 (C_{3gal}), 70.6 (C_{5gal}), 70.5, 70.4, 70.2, 69.4 (OCH₂), 69.0 (C_{2gal}), 69.0 (OCH₂), 68.6 ($C_{q}CH_{2}O$), 66.5 (C_{4gal}), 64.6 (OCH₂C=C), 61.9 (C_{6glc}), 60.7 (C_{6gal}), 60.2 (C_{q}), 50.2 ($N_{triazole}CH_{2}$), 42.3 (CH₂Cl), 29.8 (CH₂Br), 20.8, 20.8, 20.7, 20.6, 20.6, 20.5 (7×s, COCH₃).

m/z (ESI⁺ HRMS) for C₁₁₇H₁₇₁BrN₁₀O₆₇ = 1434.4805 [*M*+2H]²⁺, found 1434.4843, 1456.4624 [*M*+2Na]²⁺, found 1456.4624.

⁴ Y. M. Chabre, C. Contino-Pépin, V. Placide, T. C. Shiao and R. Roy, J. Org. Chem., 2008, **73**, 5602–5605.



Figure S37. ¹H NMR spectrum of compound 12 (CDCl₃, 600MHz)



Figure S38. ¹³C NMR spectrum of compound 12 (CDCl₃, 150MHz)



Figure S39. ESI⁺-HRMS spectrum of compound 12

Pentavalent dendron 13



To a solution of pentapropargylated cyclotriphosphazene derivative **6** (55.8 mg, 0.053 mmol, 1.0 eq.) in a 1:1 mixture of H_2O/THF_{anh} (8 mL), were added azido derivative **10** (456.7 mg, 0.545 mmol, 10.0 eq.), CuSO₄·5H₂O (102.8 mg, 0.412 mmol, 7.8 eq.) and sodium ascorbate (83.5 mg, 0.421 mmol, 8.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room tem perature for additional 18 hours. Ethyl acetate (30 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (20 mL), washed with saturated aqueous NH₄Cl (2×20 mL), water (20 mL) and brine (10 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 90:10) afforded desired multivalent compound **13** (230.5 mg, 0.044 mmol, **83%**).

R_f = 0.59, DCM/MeOH 90:10

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 9.43 (s, 1H, N*H*), 7.88 (s, 2H, *H*_{triazole}), 7.82 (2×s, 3H, *H*_{triazole}), 7.34 (d, ³*J* = 8.6 Hz, 2H, *CH*_{*b*}), 6.91-6.75 (m, 20H, *CH*_{*b*}, 6.57 (d, ³*J* = 8.6 Hz, 2H, *CH*_{*c*}), 5.31 (d_{app}, 5H, *H*_{4gal}), 5.16-5.05 (m, 20H, *H*_{3glc}, C_qC*H*₂O, *H*_{2gal}), 4.92 (dd, ³*J*_{2,3} = 2.3 Hz, ³*J*_{3,4} = 3.5 Hz, 5H, *H*_{3gal}), 4.84 (t_{app}, 6H, *H*_{2glc}), 4.53-4.45 (m, 25H, *CH*₂N, *H*_{1glc}, *H*_{6aglc}, *H*_{1gal}), 4.22 (s, 2H C*H*₂Cl), 4.11-4.03 (m, 15H, *H*_{6bglc}, *H*_{6agal}, *H*_{6bgal}), 3.87-3.54 (m, 80H, *H*_{5gal}, *H*_{5glc}, *H*_{4glc}, OC*H*₂), 2.14-1.93 (m, 105H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.2, 170.2, 170.0, 169.9, 169.6, 169.5, 169.0 (7×s, COCH₃), 164.6 (*C*=O), 155.2 + 155.2 (*C*_a), 146.6 (*C*_d), 144.4 (*C*_{triazole}=CH), 144.1 (*C*_d), 143.5 (*C*_{triazole}=CH), 143.3 (*C*_{triazole}=CH), 134.5 (*C*_a),

124.4 (C_{triazole}=CH), 124.0 (C_{triazole}=CH), 121.8 (C_c), 121.8 (C_c), 121.7(C_b), 121.0 (C_c), 120.8 (C_c), 115.2 (C_b), 115.0 (C_b), 100.9 (C_{1gal}), 100.4 (C_{1glc}), 76.1 (C_{4glc}), 72.7 (C_{3glc}), 72.5 (C_{5glc}), 72.5 (C_{2glc}), 71.5 (C_{3gal}), 70.8 (C_{5gal}), 70.5, 70.4, 70.3, 70.3, 70.1, 70.1, 69.2, 69.2 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 66.5 (C_{4gal}), 62.2 (OCH₂C=C), 61.9 (C_{6glc}), 61.7 (C_{6gal}), 60.6, 50.2 (NCH₂), 50.1 (NCH₂), 43.4 (CH₂Cl), 20.7, 20.7, 20.6, 20.5, 20.5, 20.5, 20.4 (7×s, COCH₃). ³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.2-9.4 (m, 3P).

IR (neat, v cm⁻¹) 2878 (m), 1743 (s), 1501 (m), 1367 (m), 1216 (s), 1042 (m), 833 (w).

m/z (ESI⁺-HRMS) for C₂₂₃H₂₉₇CIN₁₉O₁₁₇P₃= 1747.8998 [*M*+3H]³⁺; found 1747.8958, 1769.8818 [*M*+3Na]³⁺, found 1769.8798.



Figure S40. ¹H NMR spectrum of compound 13 (CDCl₃, 600MHz)





Figure S43. ¹³C NMR spectrum of compound 13 (CDCl₃, 150MHz)



Figure S44. FT-IR spectrum of compound 13


Figure S45. ESI⁺-HRMS spectrum of compound 13

Trivalent dendron 14



To a stirring solution of **12** (443.9 mg, 0.155 mmol, 1.0 eq) in dry DMF (6 mL) under a nitrogen atmosphere were added sodium azide (25.1 mg, 0.386 mmol, 2.5 eq.) and sodium iodide (2.3 mg, 0.015 mmol, 0.1 eq.). After stirring overnight at 70°C, the solvent was removed and EtOAC was added, then the solution was washed successively with water (4x50 mL) and brine (3x25 mL). The organic phase was then dried over Na_2SO_4 and concentrated under reduced pressure. Column chromatography on silica (EtOAc/Acetone 90:10 to 30:70) afforded the desired compound **27** (339.1 mg, 0.120 mmol, **77%**) as a colorless oil.

R_f = 0.30, DCM/MeOH 94:6

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 7.70 (s, 3H, H_{triazole}), 6.80 (br s, 1H, N*H*), 5.33 (d_{app}, 3H, $H_{4\text{gal}}$), 5.17 (dd, ³ $J_{4,3}$ = 9.4 Hz, ³ $J_{3,2}$ = 9.1 Hz, 3H, $H_{3\text{glc}}$), 5.10 (dd, ³ $J_{2,1}$ = 10.5 Hz, ³ $J_{3,2}$ = 8.0 Hz, 3H, $H_{2\text{gal}}$), 4.97 (dd, ³ $J_{2,3}$ = 7.0 Hz, ³ $J_{3,4}$ = 3.4 Hz, 3H, $H_{3\text{gal}}$), 4.88 (dd, ³ $J_{2,1}$ = 9.4 Hz, ³ $J_{3,2}$ = 8.0 Hz, 3H, $H_{2\text{glc}}$), 4.57-4.47 (m, 21H, C_qCH₂O, OCH₂CH₂N, $H_{1\text{glc}}$, $H_{6\text{aglc}}$, $H_{1\text{gal}}$), 4.12-4.08 (m, 9H, $H_{6\text{bglc}}$, $H_{6\text{bgal}}$), 3.89-3.57 (m, 59H, CH₂N₃, OCH₂CH₂N, OCH₂, $H_{4\text{glc}}$, $H_{5\text{gal}}$, $H_{5\text{glc}}$), 2.18-1.97 (m, 63H, COCH₃).

¹³**C NMR** (150 MHz, CDCl₃, δ ppm): 170.3, 170.3, 170.1, 170.0, 169.7, 169.6, 169.0 (7×s, COCH₃), 166.7 (CO), 144.4 ($C_{triazole}$ =CH), 123.7 (C= $C_{triazole}$ H), 101.0 (C_{1gal}), 100.5 (C_{1glc}), 76.2 (C_{4glc}), 72.7 (C_{3glc}), 72.5 (C_{5glc}), 71.5 (C_{2glc}), 70.9 (C_{3gal}), 70.6 (C_{5gal}), 70.5, 70.4, 70.2, 69.4 (OCH₂), 69.0 (C_{2gal}), 69.0 (OCH₂), 68.6 (C_{q} CH₂O), 66.5 (C_{4gal}), 64.6 (OCH₂C=C), 61.9 (C_{6glc}), 60.7 (C_{6gal}), 60.0 (C_{q}), 52.6 (CH₂N₃), 50.2 (N_{triazole}CH₂), 20.8, 20.8, 20.7, 20.6, 20.6, 20.6, 20.5 (7×s, COCH₃).

m/z (ESI⁺ HRMS) for C₁₁₇H₁₇₁N₁₃O₆₇ = 1416.0259 [M+2H]²⁺, found 1416.0296, 1438.0079 [M+2Na]²⁺, found 1438.0117.

GPC measurements (THF): $M_w = 1776$; $M_n = 1671$, PDI (M_w/M_n) = 1.063



Figure S46. ¹H NMR spectrum of compound 14 (CDCl₃, 600MHz)



Figure S47. gCOSY spectrum of compound 14





Figure S49. ESI⁺-HRMS spectrum of compound 14



Figure S50. GPC trace (in THF) of compound 14

Pentavalent dendron 15



To a stirring solution of **13** (224.7 mg, 0.043 mmol, 1.0 eq) in dry DMF (3 mL) under a nitrogen atmosphere were added sodium azide (8.4 mg, 0.129 mmol, 3.0 eq.) and sodium iodide (1.3 mg, 0.009 mmol, 0.2 eq.). After stirring overnight (15 h) at 70°C, the solvent was removed and EtOAc was added, then the solution was washed successively with water (4x50 mL) and brine (3x25 mL). The organic phase was then dried over Na₂SO₄ and concentrated under reduced pressure. Column chromatography on silica (DCM/MeOH 99:1 to 90:10) afforded the desired compound **15** (209.8 mg, 0.040 mmol, **93%**) as a colorless oil.

R_f = 0.59, DCM/MeOH 90:10

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 9.43 (s, 1H, N*H*), 7.88 (s, 2H, *H*_{triazole}), 7.82 (2×s, 3H, *H*_{triazole}), 7.32 (d, ³*J* = 8.6 Hz, 2H, *CH*_b), 6.91-6.75 (m, 20H, *CH*_b, *CH*_c), 6.53 (d, ³*J* = 8.6 Hz, 2H, *CH*_c), 5.30 (d_{app}, 5H, *H*_{4gal}), 5.16-5.04 (m, 20H, *H*_{3glc}, C_qC*H*₂O, *H*_{2gal}), 4.91 (dd, ³*J*_{2,3} = 2.3 Hz, ³*J*_{3,4} = 3.5 Hz, 5H, *H*_{3gal}), 4.84 (t_{app}, 5H, *H*_{2glc}), 4.53-4.45 (m, 25H, *CH*₂N, *H*_{1glc}, *H*_{6aglc}, *H*_{1gal}), 4.10-4.02 (m, 17H, *CH*₂N₃, *H*_{6bglc}, *H*_{6bgal}), 3.86-3.54 (m, 85H, *H*_{5gal}, *H*_{5glc}, *H*_{4glc}, OC*H*₂), 2.17-1.92 (m, 105H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.2, 170.2, 170.0, 169.9, 169.6, 169.5, 169.0 (7×s, COCH₃), 165.8 (C=O), 155.2 + 155.2 (C_a), 146.6 (C_d), 144.5 (C_d), 144.4 (C_d), 143.5 ($C_{triazole}$), 143.3 ($C_{triazole}$), 134.5 (C_a), 124.4 ($CH_{triazole}$), 124.0 ($CH_{triazole}$), 121.9 (C_c), 121.8 (C_c), 121.7 (C_b), 121.6 (C_c), 120.9 (C_c), 120.9 (C_c), 120.7 (C_c), 115.2 (C_b), 115.0 (C_b), 100.9 (C_{1gal}), 100.4 (C_{1glc}), 76.1 (C_{4glc}), 72.6 (C_{3glc}), 72.5 (C_{5glc}), 72.4 (C_{2glc}), 71.5 (C_{3gal}), 70.8 (C_{5gal}), 70.5, 70.4,

– S42 –

70.4, 70.3, 70.3, 70.3, 70.1, 70.1, 69.2, 69.2 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 66.5 (C_{4gal}), 62.1 (OCH₂C=C), 61.8 (C_{6glc}), 61.6 (C_{6gal}), 60.6, 52.3 (CH₂N₃), 50.2 (NCH₂), 50.1 (NCH₂), 20.7, 20.7, 20.6, 20.5, 20.5, 20.4 (COCH₃). ³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.3-9.4 (m, 3P).

IR (neat, *v* cm⁻¹) 2882 (m), 2113 (w, N₃), 1740 (s), 1502 (m), 1366 (m), 1216 (s), 1041 (m), 833 (w)

m/z (ESI⁺-HRMS) for C₂₂₃H₂₉₇N₂₂O₁₁₇P₃= 1750.2466 [*M*+3H]³⁺; found 1750.2448, 1772.2285 [*M*+3Na]³⁺, found 1772.2226.

NMR diffusion studies (CDCl₃): $D = 3.10 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 2.6 \text{ nm}$

GPC measurements (THF): M_w = 5462 ; M_n = 5367 , PDI (M_w/M_n) = 1.018 (CHCl₃/Et₃N (1%)): M_w = 4784 ; M_n = 4609 , PDI (M_w/M_n) = 1.038



Figure S51. ¹H NMR spectrum of compound 15 (CDCl₃, 600MHz)





Figure S54. ³¹P NMR spectrum of compound 15 (CDCI₃, 122MHz)



Figure S55. FT-IR spectrum of compound 15



Figure S56. ESI⁺-HRMS spectrum of compound 15



Figure S57. GPC trace (in THF) of compound 15



Figure S58. GPC trace (in CHCl₃/Et₃N (1%)) of compound 15

Protected hexavalent glycocluster 16



To a solution of propargylated derivative **1** (13.5 mg, 0.013 mmol, 1.0 eq.) in a 1:1 mixture of H₂O/THF (5 mL), were added azido derivative **10** (95.6 mg, 0.114 mmol, 8.8 eq.), CuSO₄·5H₂O (26.9 mg, 0.101 mmol, 7.8 eq.) and sodium ascorbate (22.1 mg, 0.111 mmol, 8.6 eq.). While stirring, the mixture was first heated at 50°C for 3 h ours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 99:1 to 96:4) afforded desired multivalent compound **16** (67 mg, 0.011 mmol, **84%**).

R_f = 0.42, DCM/MeOH 95:5.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 7.82 (s, 6H, $H_{triazole}$), 6.79 (d, ³*J* = 8.9 Hz, 12H, C*H*_b), 6.75 (d, ³*J* = 8.9 Hz, 12H, C*H*_c), 5.30 (d_{app}, 6H, H_{4gal}), 5.14 (dd, ³*J*_{4,3} = 9.4 Hz, ³*J*_{3,2} = 9.1 Hz, 3H, H_{3glc}), 5.09-5.04 (m, 18H, C_qC*H*₂O, *H*_{2gal}), 4.91 (dd, ³*J*_{2,3} = 7.1 Hz, ³*J*_{3,4} = 3.3 Hz, 6H, H_{3gal}), 4.84 (t_{app}, 6H, H_{2glc}), 4.53-4.43 (m, 30H, C*H*₂N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.08-4.03 (m, 18H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.86-3.83 (m, 24H, OC*H*₂CH₂N, LacOC*H*₂), 3.75 (t_{app}, 6H, H_{4glc}), 3.60-3.53 (m, 72H, OC*H*₂, H_{5gal} , H_{5gal} , 2.11-1.92 (7×s, 126H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.5, 170.5, 170.3, 170.2, 170.0, 169.8, 169.3 (7s, COCH₃), 155.5 (C_a), 144.7 (C_d), 143.7 ($C_{triazole}$ =CH), 124.4 ($C_{triazole}$ =CH), 122.0 (C_c), 115.5 (C_b), 101.2 (C_{1gal}), 100.8 (C_{1glc}), 76.5 (C_{4glc}), 73.0 (C_{3glc}), 72.8 (C_{5glc}), 71.8 (C_{2glc}), 71.2 (C_{3gal}), 70.8 (C_{5gal}), 70.8, 70.7, 70.6, 70.4, 69.6 (OCH₂), 69.3 (C_{2gal}), 69.2 (OCH₂),

– S48 –

66.8 (C_{4gal}), 62.4 (OCH₂C=C), 62.2 (C_{6glc}), 61.0 (C_{6gal}), 50.4 (NCH₂), 21.1, 21.0, 20.9, 20.8, 20.8, 20.8, 20.7 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.7 (s, 3P).

m/z (ESI⁺-HRMS) for C₂₅₈H₃₄₈N₂₁O₁₃₈P₃ = 1511.5099 [*M*+4H]⁴⁺; found 1511.5123, 1533.2410 [*M*+4Na]⁴⁺, found 1533.2391.

NMR diffusion studies (CDCl₃): $D = 2.76 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 2.9 \text{ nm}$.

GPC measurements (THF): $M_{\rm w} = 6882$; $M_{\rm n} = 6764$, PDI ($M_{\rm w}/M_{\rm n}$) = 1.017.



Figure S59. ¹H NMR spectrum of compound 16 (CDCl₃, 600MHz)





Figure S62. ³¹P NMR spectrum of compound 16 (CDCl₃, 122MHz)



Figure S63. ESI⁺-HRMS spectrum of compound 16



Figure S64. GPC trace (in THF) of compound 16

Protected octadecavalent dendrimer 17



To a solution of hexapropargylated cyclotriphosphazene derivative **1** (6.91 mg, 6.79 µmol, 1.0 eq.) in a 1:1 mixture of H_2O/THF_{anh} (5 mL), were added azido derivative **14** (150 mg, 53.0 µmol, 7.8 eq.), CuSO₄·5H₂O (13.2 mg, 53.0 µmol, 7.8 eq.) and sodium ascorbate (10.4 mg, 53.0 µmol, 7.8 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (30 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (20 mL), washed with saturated aqueous NH₄Cl (2×20 mL), water (20 mL) and brine (10 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 90:10) afforded desired multivalent compound **17** (79.0 mg, 4.39 µmol, **65%**).

R_f = 0.28, DCM/MeOH 92:8

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 7.89 (s, 6H, $H_{int-triazole}$), 7.68 (s, 18H, $H_{ext-triazole}$), 6.88-6.69 (m, 24H, CH_b ; CH_b , CH_c , $CH_{c'} + NH$ not visible), 5.30 (d_{app}, 18H, H_{4gal}), 5.14 (t_{app}, 18H, H_{3glc}) 5.07-5.02 (m, 30H, C_qCH_2O , H_{2gal}), 4.91 (dd, ³ $J_{2,3}$ = 3.4 Hz, ³ $J_{3,4}$ = 7.0 Hz, 18H, H_{3gal}), 4.83 (t_{app}, 18H, H_{2glc}), 4.54-4.43 (m, 126H, C_qCH_2O , OCH_2CH_2N , H_{1glc} , H_{6aglc} , H_{1gal}),

4.10-4.02 (m, 54H, *H*_{6bglc}, *H*_{6agal}, *H*_{6bgal}), 3.86-3.53 (m, 354H, NHCOC*H*₂N_{triazole}, OC*H*₂CH₂N, *H*_{5gal}, *H*_{5glc}, *H*_{4glc}, OC*H*₂, HNC_qC*H*₂O), 2.11-1.92 (m, 378H, COC*H*₃).

¹³**C** NMR (150 MHz, CDCl₃, δ ppm): 170.2, 170.2, 170.0, 169.9, 169.6, 169.5, 168.9 (7×s, COCH₃), 165.1 (*C*O), 155.2 (*C*_a), 144.2 (*C*_d), 144.2 (*C*_{triazole}=CH), 143.3 (*C*_{triazole}=CH), 125.2 (*C*_{triazole}=CH), 123.7 (*C*_{triazole}=CH), 121.6 (*C*_c), 115.1 (*C*_b), 100.9 (*C*_{1gal}), 100.4 (*C*_{1glc}), 76.1 (*C*_{4glc}), 72.6 (*C*_{3glc}), 72.5 (*C*_{5glc}), 71.4 (*C*_{2glc}), 70.8 (*C*_{3gal}), 70.5 (*C*_{5gal}), 70.4, 70.3, 70.2, 70.2, 70.0, 69.2 (OCH₂), 68.9 (*C*_{2gal}), 68.9 (OCH₂), 68.4 (*C*_qCH₂O), 66.5 (*C*_{4gal}), 64.4 (OCH₂C=C), 62.0 (*C*_{6glc}), 61.8 (*C*_q), 60.6 (*C*_{6gal}), 60.3 (NCH₂), 52.5 (HNCOCH₂N_{triazole}), 49.9 (OCH₂), 20.7, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.0-9.6 (m, 3P).

m/z (ESI⁺-MS) for C₇₅₆H₁₀₆₈N₈₁O₄₁₄P₃= 18008.6 [*M*+H]⁺; found 18008.5 (after deconvolution).

NMR diffusion studies (CDCl₃): $D = 1.48 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 5.5 \text{ nm}$.

GPC measurements (THF): $M_w = 23790$; $M_n = 23540$, PDI (M_w/M_n) = 1.011 (CHCl₃/Et₃N (1%)): $M_w = 21740$; $M_n = 21350$, PDI (M_w/M_n) = 1.018.











Figure S68. ³¹P NMR spectrum of compound 17 (CDCl₃, 122MHz)





Figure S69. ESI⁺-MS (deconvolution) spectrum of compound 17



Figure S70. GPC trace of compound 17



Figure S71. GPC trace (in $CHCI_3/Et_3N$ (1%)) of compound 17

Hydroxylated hexavalent glycocluster 18



Acetylated compound **16** (43.0 mg, 7.11 µmol) was dissolved in a dry mixture of MeOH/DCM (5 mL, 4:1) and a solution of sodium methoxide (1M in MeOH, 5 µL every 20 minutes until precipitation) was added. An additional 100 µL was then injected and the heterogeneous reaction mixture was stirred at room temperature for 24 h... The solvent was then removed with a Pasteur pipette and another dry mixture of MeOH/DCM (5 mL, 4:1) is added to the residual oil. A vigorous agitation is maintained for an additional 15 min. period. After removal of the solvents with a Pasteur pipette, 2 mL of dry MeOH were added to the viscous residue under agitation. After 15 min., the MeOH was removed and the residue was dissolved in 3 mL of H₂O, and the pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected hexamer **18** as a white solid (25.0 mg, 5.84 µmol) in an 82% yield.

¹H NMR (600 MHz, D₂O, δ ppm): 7.99 (s, 6H, $H_{triazole}$), 6.73 (br s, 24H, C H_{ar}), 5.03 (br s, 12H, C_qC H_2 O), 4.51 (br s, 12H, C H_2 N), 4.46-4.43 (m, 12H, H_{1glc} , H_{1gal}), 3.98-3.84 (m, 18H, OC H_2 CH₂N, LacOCHHCH₂), 3.82-3.47 (m, 132H, H_{3glc} , H_{2gal} , H_{3gal} , H_{6bglc} , H_{4gal} , H_{6agal} , H_{6agal} , H_{6agal} , H_{5gal} , H_{5glc} , H_{4glc} , LacOCHHCH₂, OCH₂), 3.33 (t_{app}, 6H, H_{2glc}). ¹³C NMR (150 MHz, CDCl₃, δ ppm): 155.6 (C_a), 144.6 (C_d), 143.7 ($C_{triazole}$ =CH), 125.9 ($C_{triazole}$ =CH), 122.4 (C_c), 116.3 (C_b), 103.7 (C_{1gal}), 102.8 (C_{1glc}), 79.1 (C_{4glc}), 76.0 (C_{3glc}), 75.4 (C_{5glc}), 75.0 (C_{5gal}), 73.5 (C_{2glc}), 73.2 (C_{3gal}), 71.6 (C_{2gal}), 70.3, 70.2, 70.2, 69.4, (OCH₂), 69.3 (C_{4gal}), 69.2 (OCH₂), 62.0 (OCH₂C=C), 61.7 (C_{6glc}), 60.8 (C_{6gal}), 50.7 (NCH₂). ³¹P NMR (122 MHz, D₂O, δ ppm): 10.2 (s, 3P). m/z (ESI⁺-HRMS) for C₁₇₄H₂₆₄N₂₁O₉₆P₃ = 1070.1481 [*M*+4H]⁴⁺, found 1070.1473; 2139.2890 [*M*+2H]²⁺, found 2139.2886.

NMR diffusion studies (D₂O): $D = 1.10 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 3.6 \text{ nm}$.





Figure S73. gCOSY spectrum of compound 18





Figure S75. ³¹P NMR spectrum of compound 18 (D₂O, 122MHz)







MS Spectrum Peak List

Ion	Formula	Abund	Observed m/z	Calc m/z	Diff (ppm)
(M+5Na)+5	C174H264N21Na5O96P3	963.12	878.30523	878.30191	3.78
(M+4H)+4	C174H268N21O96P3	2430.45	1070.14729	1070.14814	-0.79
(M+4Na)+4	C174H264N21Na4O96P3	22308.58	1092.12983	1092.13008	-0.23
(M+3H)+3	C174H267N21O96P3	21826.74	1426.5281	1426.52843	-0.23
(M+3Na)+3	C174H264N21Na3O96P3	26206.53	1448.50953	1448.51037	-0.58
(M+2H)+2	C174H266N21O96P3	1185.92	2139.28859	2139.289	-0.19
(M+2Na)+2	C174H264N21Na2O96P3	859.67	2161.26644	2161.27095	-2.08

Figure S76. ESI⁺-HRMS spectrum of compound 18

Hydroxylated octadecavalent glycodendrimer 19



Acetylated compound **17** (60.0 mg, 3.33 µmol) was dissolved in a dry mixture of MeOH/DCM (5 mL, 4:1) and a solution of sodium methoxide (1M in MeOH, 5 µL every 20 minutes until precipitation) was added. An additional 100 µL was then injected and the heterogeneous reaction mixture was stirred at room temperature for 18 hours. The solvent was then removed with a Pasteur pipette and another dry mixture of MeOH/DCM (5 mL, 4:1) is added to the residual oil. A vigorous agitation is maintained for an additional 15 min. period. After removal of the solvents with a Pasteur pipette, 2 mL of dry MeOH were added to the viscous residue under agitation. After 15 min., the MeOH was removed and the residue was dissolved in 3 mL of H_2O , and the pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected derivative **19** as a white solid (39.0 mg, 3.07 µmol, **90%**).

¹**H** NMR (600 MHz, D₂O, δ ppm): 8.06-7.96 (m, 24H, $H_{triazole}$), 6.84-6.77 (m, 24H, CH_{ar}), 5.20-5.14 (m, 24H, NHCOC H_2 N, C_qC H_2 O), 4.57-4.50 (m, 72H, C H_2 N, C_{q-triazole}C H_2 O), 4.49-4.44 (m, 36H, H_{1glc} , H_{1gal}), 4.02-3.55 (m, 486,

 OCH_2CH_2N , H_{3glc} , H_{2gal} , H_{3gal} , H_{6bglc} , H_{6agal} , H_{6aglc} , H_{5gal} , H_{5glc} , H_{4glc} , H_{4gal} , $LacOCH_2CH_2$, OCH_2 , NHC_qCH_2O), 3.22 (t_{app}, 18H, H_{2glc}).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 167.1 (CONH), 156.6 (C_a), 144.6 (C_d), 143.7 ($C_{triazole}$ =CH), 127.0 ($C_{triazole}$ =CH), 125.9 ($C_{triazole}$ =CH), 122.4 (C_c), 116.3 (C_b), 103.6 (C_{1gal}), 102.8 (C_{1glc}), 79.1 (C_{4glc}), 76.0 (C_{3glc}), 75.4 (C_{5glc}), 75.0 (C_{5gal}), 73.5 (C_{2glc}), 73.2 (C_{3gal}), 71.6 (C_{2gal}), 70.3, 70.2, 70.2, 70.2, 70.1 (OCH₂), 69.3 (C_{4gal}), 69.2 (OCH₂), 68.1 ($C_{q}CH_{2}O$), 64.2 (OCH₂C=C), 62.0 (C_{q}), 61.7 (C_{6glc}), 60.9 (C_{6gal}), 60.8 (OCH₂), 52.9 (N_{triazole}CH₂CO), 50.6 (CH₂N). ³¹P NMR (122 MHz, D₂O, δ ppm): 10.2 (s, 3P).

m/z (ESI⁺-HRMS) for C₅₀₄H₈₁₆N₈₁O₂₈₈P₃ = 12712.2, found 12711.7 ([*M*+H]⁺, after deconvolution). NMR diffusion studies (D₂O): $D = 0.67 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 5.9 \text{ nm}$.



Figure S77. ¹H NMR spectrum of compound **19** (D₂O, 600MHz)





Figure S80. ³¹P NMR spectrum of compound 19 (D₂O, 122MHz)





Protected dumbbell-shape decavalent glycocluster 21



To a solution of known dipropargylated tretra(ethylene)glycol 20^5 (1.52 mg, 5.63 µmol, 1.0 eq.) in a 1:1 mixture of H₂O/THF_{anh} (4 mL), were added azido dendron derivative **15** (65.0 mg, 12.4 µmol, 2.2 eq.), CuSO₄·5H₂O (8.4 mg, 33.0 µmol, 6.0 eq.) and sodium ascorbate (6.7 mg, 33.0 µmol, 6.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 85:15) afforded desired multivalent compound **21** (44.0 mg, 4.08 µmol, **73%**) as a yellowish foam.

R_f = 0.50, DCM/MeOH 90:10.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 10.12 (s, 2H, N*H*), 7.89 (s, 3H, *H*_{triazole}), 7.85-7.80 (2×s, 9H, *H*_{triazole}), 7.33 (d, ³*J* = 8.6 Hz, 4H, C*H*_b), 6.96-6.69 (m, 40H, *CH*_b, *CH*_c), 6.53 (d, ³*J* = 8.6 Hz, 4H, *CH*_c), 5.40 (br s, 4H, NC*H*₂CO), 5.30 (d_{app}, 10H, *H*_{4gal}), 5.18-5.07 (m, 40H, *H*_{3glc}, C_qC*H*₂O, *H*_{2gal}), 4.94 (dd, ³*J*_{2,3} = 2.3 Hz, ³*J*_{3,4} = 3.5 Hz, 10H, *H*_{3gal}), 4.84 (t_{app}, 10H, *H*_{2glc}), 4.64 (br s, 4H, C_{q-triazole}C*H*₂O), 4.55-4.46 (m, 50H, C*H*₂N, *H*_{1glc}, *H*_{6aglc}, *H*_{1gal}), 4.13-4.05 (m, 30H, *H*_{6bglc}, *H*_{6agal}, *H*_{6bgal}), 3.86-3.54 (m, 186H, *H*_{5gla}, *H*_{5glc}, *H*_{4glc}, OC*H*₂), 2.17-1.92 (7×s, 210H, COC*H*₃).

¹³**C NMR** (75 MHz, CDCl₃, δ ppm): 170.2, 170.2, 170.0, 169.9, 169.6, 169.5, 169.0 (7×s, COCH₃), 164.0 (*C*=O), 155.3 + 155.2 (*C*_a), 146.6 (*C*_d), 144.5 (*C*_d), 144.4 (*C*_d), 143.8 (*C*_{triazole}=CH), 143.5 (*C*_{triazole}=CH), 143.3 (*C*_{triazole}=CH), 134.5 (*C*_a), 124.7 (*C*H_{triazole}), 124.0 (*C*H_{triazole}), 121.9 (*C*_c), 121.8 (*C*_c), 121.7 (*C*_b), 121.6 (*C*_c), 120.9 (*C*_c), 120.7 (*C*_c), 115.3 (*C*_b), 115.1 (*C*_b), 100.9 (*C*_{1gal}), 100.4 (*C*_{1glc}), 76.2 (*C*_{4glc}), 72.7 (*C*_{3glc}), 72.6 (*C*_{5glc}), 72.5 (*C*_{2glc}), 71.5 (*C*_{3gal}), 70.8 (*C*_{5gal}), 70.6, 70.5, 70.4, 70.4, 70.3, 70.3, 70.3, 70.2, 70.1, 70.1, 69.4, 69.2, 69.2 (OCH₂),

⁵ X. Sheng, T. C. Mauldin and M. R. Kessler, J. Polym. Sci.: Part A: Polym. Chem., 2010, 48, 4093–4102.

68.9 (C_{2gal}), 68.9 (OCH₂), 66.5 (C_{4gal}), 64.3 ($C_{q-triazole}CH_2OCH_2$), 62.1 (OCH₂C=C), 61.8 (C_{6glc}), 61.6 (C_{6gal}), 60.6, 52.4 (NHCOCH₂N_{triazole}), 50.2 (NCH₂), 50.1 (NCH₂), 20.8, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃). ³¹P NMR (122 MHz, CDCl₃, δ ppm): 10.8-9.4 (m, 6P).

m/z (ESI⁺-MS) for C₄₆₀H₆₁₆N₄₄O₂₃₉P₆= 10772.8 [*M*+H]⁺; found 10772.5 (After deconvolution).

NMR diffusion studies (CDCl₃): $D = 2.47 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 3.3 \text{ nm}$.

GPC measurements (THF): $M_w = 14710$; $M_n = 13860$, PDI (M_w/M_n) = 1.061 (CHCl₃/Et₃N (1%)): $M_w = 10410$; $M_n = 10180$, PDI (M_w/M_n) = 1.023







Figure S83. gCOSY spectrum of compound 21





Figure S86. ESI⁺-MS spectrum (deconvolution) of compound 21



Figure S87. GPC trace (in THF) for compound 21



Figure S88. GPC trace (in $CHCI_3/Et_3N$ (1%)) for compound 21

Hydroxylated dumbbell-shape decavalent glycocluster 22



Acetylated compound **21** (28.0 mg, 2.60 µmol) was dissolved in a dry mixture of MeOH/DCM (5 mL, 4:1) and a solution of sodium methoxide (1M in MeOH, 5 µL every 20 minutes until precipitation) was added. An additional 100 µL was then injected and the heterogeneous reaction mixture was stirred at room temperature for 18 hours. The solvent was then removed with a Pasteur pipette and another dry mixture of MeOH/DCM (5 mL, 4:1) is added to the residual oil. A vigorous agitation is maintained for an additional 15 min. period. After removal of the solvents with a Pasteur pipette, 2 mL of dry MeOH were added to the viscous residue under agitation. After 15 min., the MeOH was removed and the residue was dissolved in 3 mL of H_2O , and the pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected derivative **22** as an off-white solid (18.5 mg, 2.55 µmol, **98%**).

¹**H NMR** (600 MHz, D₂O, δ ppm): 7.99 (s, 12H, $H_{triazole}$), 7.28 (br s, 4H, $CH_{b'}$), 6.75-6.50 (m, 44H, CH_{ar}), 5.29 (br s, 4H, NC H_2 CO), 4.96 (br s, 20H, C_qC H_2 O), 4.53-4.39 (m, 44H, N_{triazole}C H_2 CH₂, C_{q-triazole}C H_2 O, H_{1glc} , H_{1gal}), 3.90 (m, 30H, OC H_2 CH₂N, LacOCHHCH₂), 3.82-3.47 (m, 236H, H_{3glc} , H_{2gal} , H_{3gal} , H_{6bglc} , H_{4gal} , H_{6agal} , H_{6bgal} , H_{6aglc} , H_{5gal} , H_{5glc} , H_{4glc} , LacOCHHCH₂, OC H_2), 3.33 (t_{app}, 10H, H_{2glc}).

¹³C NMR (75 MHz, D₂O, δ ppm): 165.3 (C=O), 156.3 (C_a), 144.8 (C_{d'}), 144.5 (C_d), 143.7 (C_{triazole}=CH), 143.7 (C_{triazole}=CH), 125.9 (C_{triazole}=CH), 122.4 (C_{triazole}=CH), 121.8 (C_c + C_{b'} + C_{c'}), 116.2 (C_b), 115.1 (C_b), 103.7 (C_{triazole}], 102.8 (C_{1glc}), 79.1 (C_{4glc}), 76.0 (C_{3glc}), 75.4 (C_{5glc}), 75.0 (C_{5gal}), 73.5 (C_{2glc}), 73.2 (C_{3gal}), 71.6 (C_{2gal}), 70.3, 70.2, 70.0, 69.4, (OCH₂), 69.3 (C_{4gal}), 69.2 (OCH₂), 63.8 (C_{q-triazole}CH₂OCH₂), 62.0 (OCH₂C=C), 61.7 (C_{6glc}), 60.7 (C_{6gal}), 53.2 (N_{triazole}CH₂CO), 50.7 (NCH₂).

³¹**P NMR** (122 MHz, D₂O, δ ppm): 10.2 (s, 6P).

m/z (ESI⁺-MS) for C₃₂₀H₄₇₆N₄₄O₁₆₉P₃ = 7830.3 [*M*+H]⁺, found 7829.9 (After deconvolution).

NMR diffusion studies (D₂O): $D = 0.75 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 5.3 \text{ nm}$.

– S72 –


Figure S89. ¹H NMR spectrum of compound 22 (D₂O, 600MHz)



Figure S90. gCOSY spectrum of compound 22







Figure S93. ESI⁺-MS spectrum (deconvolution) of compound 22





To a solution of propargylated derivative **9** (11.4 mg, 9.46 μ mol, 1.1 eq.) in a 1:1 mixture of H₂O/THF_{anh} (4 mL), were added azido dendron **15** (45.0 mg, 8.57 μ mol, 1.0 eq.), CuSO₄·5H₂O (23.0 mg, 92.0 μ mol, 11.1 eq.) and sodium ascorbate (11.9 mg, 92.0 μ mol, 11.1 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 85:15) afforded desired multivalent compound **23** (40 mg, 6.2 μ mol, **72%**) as an off-white foam

R_f = 0.29, DCM/MeOH 94:6.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 10.24 (s, 1H, N*H*_{core}), 8.96 (s, 2H, N*H*_{ext}), 8.60 (s, 2H, N*H*_{ext}), 8.59 (s, 1H, N*H*_{ext}), 8.05 (s, 1H, *H*_{triazole int}), 7.90 (s; 2H, *H*_{triazole ext}), 7.85 (s, 1H, *H*_{triazole ext}), 7.84 (s, 2H, *H*_{triazole ext}), 7.32 (d, 2H, *CH*_b, ³*J* = 8.6 Hz), 7.23 (m, 10H, *CH*_b^{-,}), 6.98 (d, 2H, *CH*_c⁻), 6.91-6.44 (m, 34H, *CH*_b, *CH*_c, *CH*_c^{-,}), 5.46 (s, 2H, N_{triazole}*CH*₂CONH), 5.33 (br s, 5H, *H*_{4gal}), 5.18-5.08 (m, 22H, *H*_{3glc}, *C*_{q triazole}*CH*₂O, *H*_{2gal}), 4.94 (d_{app}, 5H, *H*_{3gal}), 4.86 (m, 5H, *H*_{2glc}), 4.55-4.46 (m, 25H, *CH*₂N, *H*_{1glc}, *H*_{6aglc}, *H*_{1gal}), 4.22 (s, 6H; *CH*₂Cl), 4.18 (s, 4H; *CH*₂Cl), 4.11-4.06 (m, 15H, *H*_{6bglc}, *H*_{6agal}, *H*_{6bgal}), 3.87-3.57 (m, 85H, *H*_{5gla}, *H*_{5glc}, *H*_{4glc}, OC*H*₂), 2.13-1.95 (m, 105H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.2, 170.0, 169.9, 169.6, 169.5, 169.0 (7×s, COCH₃), 164.6 (CONH), 164.5 (CONH), 163.8 (C=O), 155.3 ($C_{a^{"}}$), 155.2 + 155.2 (C_{a}), 147.0 ($C_{d^{"}}$), 146.8 ($C_{d^{'}}$), 144.5 (C_{d}), 144.5 ($C_{d^{"}}$), 144.2 (C_{d}), 143.8 ($C_{triazole}$ =CH), 143.7 ($C_{triazole}$ =CH), 143.2 ($C_{triazole}$ =CH), 134.6 ($C_{a^{"}}$), 134.5 ($C_{a^{'}}$), 124.7 ($C_{triazole}$ int=CH), 124.0 ($C_{triazole}$ =CH), 122.0 ($C_{b^{"}}$), 121.9 ($C_{b^{"}}$), 121.8 (C_{c}), 121.7 ($C_{b^{'}}$), 121.6 ($C_{c^{'}}$), 121.2

 (C_{c}) , 121.1 (C_{c}) , 120.9 (C_{c}) , 120.4 (C_{c}) , 120.3 (C_{c}) , 115.3 (C_{b}) , 115.2 (C_{b}) , 115.0 (C_{b}) , 100.9 (C_{1gal}) , 100.4 (C_{1glc}) , 76.1 (C_{4glc}) , 72.6 (C_{3glc}) , 72.5 (C_{5glc}) , 72.4 (C_{2glc}) , 71.5 (C_{3gal}) , 70.8 (C_{5gal}) , 70.5, 70.4, 70.4, 70.3, 70.3, 70.3, 70.1, 70.1, 69.2, 69.2 (OCH_2) , 68.9 (C_{2gal}) , 68.9 (OCH_2) , 66.5 (C_{4gal}) , 62.1 $(OCH_2C=C)$, 61.8 (C_{6glc}) , 61.6 (C_{6gal}) , 60.6 (OCH_2) , 52.9 $(N_{triazole}CH_2CONH)$, 50.3 (NCH_2) , 50.2 (NCH_2) , 43.2 (CH_2CI) , 43.1 (CH_2CI) , 20.7, 20.7, 20.6, 20.5, 20.5, 20.4 $(7 \times s, COCH_3)$.

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.9-8.5 (m, 6P, *P*-O).

m/z (ESI⁺-HRMS) for C₂₇₂H₃₃₉Cl₅N₃₀O₁₂₉P₆= 1636.6848 [*M*+4Na]⁴⁺; found 1636.6855, 2151.9342 [*M*+3Na]³⁺, found 2151.9363.

GPC measurements (THF): $M_w = 7981$; $M_n = 7889$, PDI (M_w/M_n) = 1.012.











Figure S98. ESI⁺-HRMS spectrum of compound 23



Figure S99. GPC trace (in THF) for compound 23

Protected pentadecavalent glycodendrimer 25



To a solution of known tripropargylated core 24^6 (1.27 mg, 3.91 µmol, 1.0 eq.) in a 1:1 mixture of H₂O/THF_{anh.} (4 mL), were added azido dendron derivative **15** (80 mg, 0.015 mmol, 3.9 eq.), CuSO₄·5H₂O (7.8 mg, 0.031 mmol, 8.0 eq.) and sodium ascorbate (6.2 mg, 0.031 mmol, 8.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness under vacuum with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 85:15) afforded desired multivalent compound **25** (54 mg, 3.4 µmol, **86%**) as a white foam.

⁶ Y. M. Chabre, D. Giguère, B. Blanchard, J. Rodrigue, S. Rocheleau, M. Neault, S. Rauthu, A. Papadopoulos, A. A. Arnold, A. Imberty and R. Roy, *Chem. Eur. J.*, 2011, **17**, 6545–6562.

R_f = 0.38, DCM/MeOH 93:7.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 10.1 (br s, 3H, N*H*), 8.49 (br s, 3H, N*H*), 7.98 (s, 3H, C*H*_{core}), 7.91-7.83 (m, 18H, *H*_{triazole}), 7.36-7.30 (m, 6H, C*H*_b), 6.97-6.48 (m, 66H, C*H*_b, C*H*_c, C*H*_c), 5.40 (br s, 6H, N_{triazole}C*H*₂CONH), 5.32 (d_{app}, 15H, *H*_{4gal}), 5.20-5.04 (m, 60H, *H*_{3glc}, C_qC*H*₂O, *H*_{2gal}), 4.93 (dd, ³*J*_{2,3} = 3.3 Hz, ³*J*_{3,4} = 7.1 Hz, 15H, *H*_{3gal}), 4.87-4.82 (m, 15H, *H*_{2glc}), 4.63 (br s, 6H, CONHC*H*₂C_q), 4.54-4.42 (m, 75H, C*H*₂N, *H*_{1glc}, *H*_{6aglc}, *H*_{1gal}), 4.12-4.04 (m, 45H, *H*_{6bglc}, *H*_{6agal}, *H*_{6bgal}), 3.86-3.54 (m, 255H, *H*_{5gla}, *H*_{5glc}, *H*_{4glc}, OC*H*₂), 2.11-1.92 (m, 315H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.4, 170.3, 170.2, 170.1, 169.9, 169.7, 169.5 (7×s, COCH₃), 165.0 (NHC=O_{core}), 163.9 (NHC=O), 155.3 (C_a), 155.3 (C_a), 146.5 (C_d), 144.4 (C_d), 144.1 (C_d), 143.5 ($C_{triazole}$ =CH), 143.3 ($C_{triazole}$ =CH), 133.8 (C_{q-core}), 129.1 (CH_{core}), 124.7 ($C_{triazole}$ =CH), 124.5 (C_a), 124.1 ($C_{triazole}$ =CH), 121.8 (C_c), 120.9 (C_c), 120.7 (C_b), 115.2 (C_b), 115.1 (C_b), 101.0 (C_{1gal}), 100.5 (C_{1glc}), 76.2 (C_{4glc}), 72.7 (C_{3glc}), 72.5 (C_{5glc}), 71.5 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.4, 70.4, 70.3, 70.2, 70.1, 70.1, 69.3, 69.1, 69.0 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 66.5 (C_{4gal}), 62.1 (OCH₂C=C), 61.9 (C_{6glc}), 61.6 (OCH₂), 60.7 (C_{6gal}), 52.7 ($N_{triazole}$ CH₂CONH), 50.2 (NCH₂), 50.1 (NCH₂), 35.8 (C_{core} CONHCH₂C_{triazole}), 29.6, 20.8, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.9-9.5 (m, 9P).

m/z (ESI⁺-MS) for C₆₈₇H₉₀₆N₆₉O₃₅₄P₉= 16074.6 [*M*+H]⁺; found 16074.1 (After deconvolution).

NMR diffusion studies (CDCl₃): $D = 1.65 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 5.1 \text{ nm}$.

GPC measurements (THF): $M_w = 18970$; $M_n = 18820$, PDI (M_w/M_n) = 1.008.





– S83 –



Figure S103. ³¹P NMR spectrum of compound 25 (CDCl₃, 122MHz)





Figure S104. ESI⁺-MS spectra (deconvolution) of compound 25



Figure S105. GPC trace (in THF) for compound 25

Protected tricontavalent glycodendrimer 26



To a solution of hexapropargylated cyclotriphosphazene derivative **1** (2.40 mg, 2.36 μ mol, 1.0 eq.) in a 1:1 mixture of H₂O/THF_{anh} (5 mL), were added azido derivative **15** (100 mg, 19.0 μ mol, 8.1 eq.), CuSO₄·5H₂O (4.71 mg, 18.8 μ mol, 8.0 eq.) and sodium ascorbate (3.74 mg, 18.8 μ mol, 8.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 85:15) afforded desired multivalent compound **26** (59 mg, 1.81 μ mol, **77%**) as a white foam.

R_f = 0.42, DCM/MeOH 92:8.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 10.1 (br s, 6H, N*H*), 7.98 (s, 6H, $H_{triazole}$), 7.86-7.80 (m, 30H, $H_{triazole}$), 7.36-7.30 (m, 12H, C*H*_b), 6.97-6.54 (m, 156H, C*H*_b, C*H*_c, C*H*_c), 5.40 (br s, 12H, N_{triazole}C*H*₂CONH), 5.32 (d_{app}, 30H, *H*_{4gal}), 5.17-5.04 (m, 132H, H_{3glc} , C_qC*H*₂O, H_{2gal}), 4.93 (dd, ³*J*_{2,3} = 3.3 Hz, ³*J*_{3,4} = 7.1 Hz, 30H, H_{3gal}), 4.87-4.82 (m, 30H, H_{2glc}), 4.54-4.42 (m, 150H, C*H*₂N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.12-4.04 (m, 90H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.90-3.45 (510H, H_{5gal} , H_{5glc} , H_{4glc} , OC*H*₂), 2.11-1.92 (m, 630H, COC*H*₃).

¹³**C** NMR (75 MHz, CDCl₃, δ ppm): 170.2, 170.2, 170.0, 169.9, 169.9, 169.7, 169.5 (7×s, COCH₃), 163.9 (*C*=O), 155.2 (*C*_a), 146.5 (*C*_d'), 144.4 (*C*_d), 144.1 (*C*_d), 143.4 (*C*_{triazole}=CH), 143.2 (*C*_{triazole}=CH), 134.8 (*C*_a'), 125.2 (*C*_{triazole}=CH), 124.6 (*C*_{triazole}=CH), 124.1 (*C*_{triazole}=CH), 121.9 (*C*_c'), 121.7 (*C*_b'), 120.9 (*C*_c), 120.7 (*C*_c), 115.9 (*C*_b), 115.2 (*C*_b), 115.1 (*C*_b), 101.0 (*C*_{1gal}), 100.5 (*C*_{1glc}), 76.2 (*C*_{4glc}), 72.7 (*C*_{3glc}), 72.5 (*C*_{5glc}), 71.5 (*C*_{2glc}), 70.9 (*C*_{3gal}), 70.5 (*C*_{5gal}), 70.5, 70.4, 70.4, 70.3, 70.2, 70.1, 70.1, 69.3, 69.1, 69.0 (OCH₂), 68.9 (*C*_{2gal}), 68.9 (OCH₂), 66.5 (*C*_{4gal}), 62.1 (OCH₂C=C), 61.9 (*C*_{6glc}), 61.6 (OCH₂), 60.7 (*C*_{6gal}), 52.7 (N_{triazole}CH₂CONH), 50.2 (NCH₂), 50.1 (NCH₂), 29.6, 20.8, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.2-9.7 (m, 21P).

m/z (ESI⁺-MS) for C₁₃₉₂H₁₈₂₄N₁₃₅O₇₁₄P₂₁= 32523.3 [M+H]⁺; found 32523.7 (After deconvolution) with signals corresponding to losses of monomers (~876) and dendron(s) (~5290)).

MALDI-TOF (DHB matrix) : 32550.1, with signals corresponding to losses of monomers (~876) and dendron(s) (~5290)).

NMR diffusion studies (CDCl₃): $D = 1.08 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 7.5 \text{ nm}$.

GPC measurements (THF): M_w = 33560 ; M_n = 33090 , PDI (M_w/M_n) = 1.014 (CHCl₃/Et₃N (1%)): M_w = 35320 ; M_n = 34480 , PDI (M_w/M_n) = 1.025.



Figure S106. ¹H NMR spectrum of compound 26 (CDCI₃, 600MHz)



– S88 –



User Spectra



Figure S111. ESI⁺-MS spectrum (deconvolution) of compound 26



Figure S112. MALDI-TOF (DHB matrix) of compound 26



Figure S113. GPC trace (in THF) for compound 26



Figure S114. GPC trace (in CHCl₃/Et₃N (1%)) for compound 26

Hydroxylated pentadecavalent glycodendrimer 27



Acetylated compound **25** (50.0 mg, 3.11 µmol) was dissolved in a dry mixture of MeOH/DCM (5 mL, 4:1) and a solution of sodium methoxide (1M in MeOH, 5 µL every 20 minutes until precipitation) was added. An additional 100 µL was then injected and the heterogeneous reaction mixture was stirred at room temperature for 18 hours. The solvent was then removed with a Pasteur pipette and another dry mixture of MeOH/DCM (5 mL, 4:1) is added to the residual oil. A vigorous agitation is maintained for an additional 15 min. period. After removal of the solvents with a Pasteur pipette, 2 mL of dry MeOH were added to the viscous residue under agitation. After 15 min., the MeOH was removed and the residue was dissolved in 3 mL of H₂O, and the pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected derivative **27** as an off-white solid (29.0 mg, 2.49 µmol, **80%**).

¹**H NMR** (600 MHz, D₂O, δ ppm): 8.27 (s, 3H, CH_{core}), 7.86-7.75 (m, 18H, $H_{triazole}$), 7.10 (br s, 6H, CH_b), 6.57-6.47 (m, 66H, CH_b, CH_c, CH_c), 5.17 (br s, 6H, N_{triazole}CH₂CO), 4.96-4.82 (br s, 36H, C_{q-triazole}CH₂O + CONHCH₂C_q), 4.40-4.30 (m, 60H, N_{triazole}CH₂CH₂, H_{1glc} , H_{1gal}), 3.82 (m, 45H, OCH₂CH₂N, LacOCHHCH₂), 3.75-3.27 (m, 330H, H_{3glc} , H_{2gal} , H_{3gal} , H_{6bglc} , H_{4gal} , H_{6agal} , H_{5gal} , H_{5glc} , H_{4glc} , LacOCHHCH₂, OCH₂), 3.33 (t_{app}, 15H, H_{2glc}).

¹³C NMR (150 MHz, D₂O, δ ppm): 167.1 (C=O), 165.2 (C=O), 156.6 (C_a), 146.8 (C_{d'}), 145.5 (C_d), 144.5 (C_d), 143.6 (C_{triazole}=CH), 143.7 (C_{triazole}=CH), 133.8 (C_{q-core}), 129.9 (CH_{core}), 125.9 (C_{triazole}=CH), 125.4 (C_{a'}), 122.4 (C_{triazole}=CH), 121.7 (C_c + C_{b'} + C_{c'}), 116.2 (C_b), 103.7 (C_{1gal}), 102.8 (C_{1glc}), 79.1 (C_{4glc}), 76.0 (C_{3glc}), 75.4 (C_{5glc}), 75.0 (C_{5gal}), 73.5 (C_{2glc}), 73.2 (C_{3gal}), 71.6 (C_{2gal}), 70.3, 70.2, 70.0, 69.4, (OCH₂), 69.4 (C_{4gal}), 69.2 (OCH₂), 62.0 (OCH₂C=C), 61.7 (C_{6glc}), 60.7 (C_{6gal}), 53.2 (N_{triazole}CH₂CO), 50.7 (NCH₂), 36.8 (C_{core}CONHCH₂C_q).

³¹**P NMR** (122 MHz, D₂O, δ ppm): 10.2 (s, 9P).

m/z (ESI⁺-HRMS) for C₄₇₇H₆₉₆N₆₉O₂₄₉P₉ = 11660.7 [*M*+H]⁺, found 11660.6 (After deconvolution). NMR diffusion studies (D₂O): $D = 0.70 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 5.7 \text{ nm}$.





Figure S117. ¹³C NMR spectrum of compound **27** (D_2O , 150MHz, acetone as reference) (*insert*: Zoom of the aromatic section)







Figure S119. ESI⁺-MS spectrum of compound 27

Hydroxylated tricontavalent glycodendrimer 28



Acetylated compound **26** (50.0 mg, 1.54 μ mol) was dissolved in a dry mixture of MeOH/DCM (5 mL, 4:1) and a solution of sodium methoxide (1M in MeOH, 5 μ L every 20 minutes until precipitation) was added. An additional 100 μ L was then injected and the heterogeneous reaction mixture was stirred at room temperature for 18 hours. The solvent was then removed with a Pasteur pipette and another dry mixture of MeOH/DCM (5 mL, 4:1) is added to the residual oil. A vigorous agitation is maintained for an additional 15 min. period. After removal of the solvents with a Pasteur pipette, 2 mL of dry MeOH were added to the viscous residue under agitation. After 15 min., the MeOH was removed and the residue was dissolved in 3 mL of H₂O, and the pH was adjusted to 7 with addition of ion-exchange

resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected derivative **28** as an off-white solid (29.0 mg, 1.22 μ mol, **93%**).

¹**H NMR** (600 MHz, D₂O, δ ppm): 7.99 (m, 36H, $H_{triazole}$), 7.28 (br s, 12H, C H_b), 6.99 (m, 156H, C H_b , C H_c , C H_c , C H_{arcore}), 5.10-4.90 (br s, 72H, C_{q-triazole}C H_2 O), 4.56-4.41 (m, 132H, N_{triazole}C H_2 CH₂, N_{triazole}C H_2 CO, H_{1glc} , H_{1gal}), 3.93 (m, 90H, OC H_2 CH₂N, LacOCHHCH₂), 3.81-3.34 (m, 690H, H_{3glc} , H_{2gal} , H_{3gal} , H_{6bglc} , H_{4gal} , H_{6agal} , H_{6agal} , H_{6aglc} , H_{5gal} , H_{5gal} , H_{5gal} , H_{5gal} , H_{5gal} , H_{5gal} , H_{2gal} , H_{2gal} , H_{2gal} , H_{2gal} , H_{4gal} , H_{6agal} , H_{6bgal} , H_{6agal} , H_{6ag

¹³C NMR (150 MHz, D₂O, δ ppm): 167.1 (*C*=O) not visible, 165.2 (*C*=O) not visible, 156.6 (*C*_a), 154.3 (*C*_{a-core}), 146.8 (*C*_{d'}), 145.5 (*C*_d), 144.6 (*C*_d), 143.7 (*C*_{triazole}=CH), 143.7 (*C*_{triazole}=CH), 136.5 (*C*_{a'}), 125.9 (*C*_{triazole}=CH), 122.4 (*C*_{triazole}=CH), 121.0, 120.7 (*C*_c + *C*_{b'} + *C*_{c'}), 116.2 (*C*_b, *C*_{b-core}), 103.7 (*C*_{1gal}), 102.8 (*C*_{1glc}), 79.1 (*C*_{4glc}), 76.0 (*C*_{3glc}), 75.4 (*C*_{5glc}), 75.0 (*C*_{5gal}), 73.5 (*C*_{2glc}), 73.2 (*C*_{3gal}), 71.6 (*C*_{2gal}), 70.3, 70.2, (OCH₂), 69.4 (*C*_{4gal}), 69.2 (OCH₂), 62.1 (OCH₂C=C), 61.7 (*C*_{6glc}), 60.8 (*C*_{6gal}), 53.2 (N_{triazole}CH₂CO), 50.7 (NCH₂).

³¹**P NMR** (122 MHz, D₂O, δ ppm): 10.2 (s, 21P).

m/z (MALDI-TOF) for C₉₇₂H₁₄₀₄N₁₃₅O₅₀₄P₂₁ = 23694.6, found 23755.9 (*with signals corresponding to losses of dendron(s)*).

NMR diffusion studies (D₂O): $D = 0.56 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 7.2 \text{ nm}$.



Figure S120. ¹H NMR spectrum of compound 28 (D₂O, 600MHz)



Figure S122. ¹³C NMR spectrum of compound **28** (D₂O, 150MHz, acetone as reference)



Figure S123. 31 P NMR spectrum of compound 28 (D₂O, 122MHz)



Figure S124. MALDI-TOF (DHB matrix) spectrum for compound 26 (D₂O, 600MHz)

AB₁₅ glycodendron with a focal CI 29



To a stirring solution of **6** (9.8 mg, 9.3 µmol, 1.0 eq.) and dendron **14** (174.2 mg, 60.7 µmol, 6.5 eq.) in dry THF (2.5 mL) were added 2.5 mL of H₂O and a mixture of CuSO₄·5H₂O (13.2 mg, 52.8 µmol, 5.7 eq.) and sodium ascorbate (10.5 mg, 52.8 µmol, 5.7 eq.). After stirring for 3 hours at 50°C in a 20 mL vial, the reaction was left stirring at room temperature for 18 hours (additional CuSO₄·5H₂O (7.0 mg) and sodium ascorbate (4.0 mg) were incorporated in the mixture after 5 hours of reaction). EtOAc (50 mL) was added and the solution was washed successively with a saturated aqueous solution of NH₄Cl (3×25 mL), water (2×20 mL) and brine (10 mL). The organic phase was then dried over MgSO₄ and concentrated under reduced pressure. Column chromatography on silica (DCM/MeOH 100:0 to 93:7) afforded the desired compound **29** (103.0 mg, 6.77 µmol, **73%**) as a colorless oil.

R_f = 0.18, DCM/MeOH 94:6

¹**H NMR** (600 MHz, CDCI₃, δ ppm): 10.1 (5H, N*H*) *not visible*, 9.48 (br s, 1H, N*H*), 7.93-7.90 (m, 5H, *H*_{triazole int}), 7.72-7.68 (s, 15H, *H*_{triazole ext}), 7.35 (m, 2H, C*H*_b), 6.97-6.57 (m, 22H, C*H*_b, C*H*_c, C*H*_c), 5.30 (d_{app}, 15H, *H*_{4gal}), 5.15 (t_{app}, 15H, *H*_{3glc}), 5.08-5.05 (m, 25H, C_qC*H*₂O, *H*_{2gal}), 4.93 (dd, ³*J*_{2,3} = 3.4 Hz, ³*J*_{3,4} = 7.0 Hz, 15H, *H*_{3gal}), 4.84 (t_{app}, 15H, *H*_{2glc}),

– S100 –

4.53-4.44 (m, 105H, C_qCH_2O , OCH_2CH_2N , H_{1glc} , H_{6aglc} , H_{1gal}), 4.17 (s, 2H, CH_2CI), 4.11-4.03 (m, 45H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.88-3.55 (m, 295H, NHCOC $H_2N_{triazole}$, OCH_2CH_2N , H_{5gal} , H_{5glc} , H_{4glc} , OCH_2 , HNC_qCH_2O), 2.11-1.92 (m, 315H, COC H_3).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.2, 170.0, 169.9, 169.7, 169.5, 169.0 (7×s, COCH₃), 167.1 (CONH), 165.1 (CONH), 155.3 (C_a), 147.3 (C_d), 144.6 (C_d), 144.3 (C_d), 144.3, 144.2 ($C_{triazole}$ =CH), 143.5 ($C_{triazole}$ =CH), 134.6 ($C_{a'}$), 125.4 ($C_{triazole}$ =CH), 125.1 ($C_{triazole}$ =CH), 123.7 ($C_{triazole}$ =CH), 121.8 (C_c), 120.9 (C_b), 120.8 (C_c), 115.9, 115.1 (C_b), 100.9 (C_{1gal}), 100.5 (C_{1glc}), 76.1 (C_{4glc}), 72.7 (C_{3glc}), 72.5 (C_{5glc}), 71.5 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.5, 70.4, 70.3, 70.1, 69.2 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 68.5 (C_qCH_2O), 66.5 (C_{4gal}), 64.5 (OCH₂C=C), 62.0 (C_q), 61.9 (C_{6glc} + C_qCH_2O), 60.7 (C_{6gal}), 60.3 (OCH₂), 52.5 (N_{triazole}CH₂CONH), 49.9 (NCH₂), 43.5 (COCH₂Cl), 20.7, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.8 (m, 3P).

m/z (ESI⁺ MS) for C₆₃₈H₈₉₇CIN₆₉O₃₄₇P₃ = 15214.6 [*M*+H]⁺, found 15214.3 (After deconvolution). NMR diffusion studies (CDCI₃): $D = 1.57 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 5.1 \text{ nm}$.



Figure S125. ¹H NMR spectrum of compound 29 (CDCl₃, 600MHz)







Figure S129. ESI⁺-MS spectrum (deconvolution) of compound 29

AB₁₅ glycodendron with a focal N₃ 30



To a stirring solution of **29** (129 mg, 8.48 μ mol, 1.0 eq) in dry DMF (2 mL) under a nitrogen atmosphere were added sodium azide (5.0 mg, 76 μ mol, 9.0 eq.) and sodium iodide (1.0 mg, 1.7 μ mol, 0.2 eq.). After stirring at 70°C for 5 hours under a nitrogen atmosphere, the mixture was stirred at room temperature for additional 15 hours. The solvent was removed under reduced pressure and EtOAc was added. Then the organic was washed successively with water (4×20 mL) and brine (3×10 mL). The organic phase was then dried over Na₂SO₄ and concentrated under reduced pressure. Column chromatography on silica (DCM/MeOH 100:0 to 90:10) afforded the desired compound **30** (98.0 mg, 6.44 μ mol, **76%**) as a colorless oil.

R_f = 0.18, DCM/MeOH 94:6

¹**H NMR** (600 MHz, CDCI₃, δ ppm): 10.1 (5H, N*H*) *not visible*, 9.48 (br s, 1H, N*H*), 7.93-7.90 (m, 5H, *H*_{triazole int}), 7.73-7.67 (s, 15H, *H*_{triazole ext}), 7.33 (m, 2H, C*H*_b'), 6.99-6.57 (m, 22H, C*H*_b, C*H*_c, C*H*_c'), 5.30 (d_{app}, 15H, *H*_{4gal}), 5.16 (t_{app}, 15H, *H*_{3glc}), 5.09-5.04 (m, 25H, C_qC*H*₂O, *H*_{2gal}), 4.92 (dd, ³*J*_{2,3} = 3.4 Hz, ³*J*_{3,4} = 7.0 Hz, 15H, *H*_{3gal}), 4.84 (t_{app}, 15H, *H*_{2glc}),

4.53-4.44 (m, 105H, C_qC*H*₂O, OCH₂C*H*₂N, *H*_{1glc}, *H*_{6aglc}, *H*_{1gal}), 4.11-4.03 (m, 47H, *H*_{6bglc}, *H*_{6agal}, *H*_{6bgal}, C*H*₂N₃), 3.84-3.52 (m, 295H, NHCOC*H*₂N_{triazole}, OC*H*₂CH₂N, *H*_{5qal}, *H*_{5qglc}, *H*_{4qlc}, OC*H*₂, HNC_qC*H*₂O), 2.11-1.92 (m, 315H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.2, 170.2, 170.0, 169.9, 169.6, 169.5, 168.9 (7×s, COCH₃), 166.0 (CONH), 165.0 (CONH), 155.2 (C_a), 147.3 (C_d), 144.6 (C_d), 144.5 (C_d), 144.3, 144.2 ($C_{triazole}$ =CH), 143.4 ($C_{triazole}$ =CH), 134.6 ($C_{a'}$), 125.3 ($C_{triazole}$ =CH), 125.1 ($C_{triazole}$ =CH), 123.7 ($C_{triazole}$ =CH), 121.7 (C_c), 120.8 (C_b), 120.8 (C_c), 115.8, 115.1 (C_b), 100.9 (C_{1gal}), 100.4 (C_{1glc}), 76.1 (C_{4glc}), 72.6 (C_{3glc}), 72.4 (C_{5glc}), 71.4 (C_{2glc}), 70.8 (C_{3gal}), 70.4 (C_{5gal}), 70.4, 70.3, 70.2, 69.2 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 68.4 (C_qCH_2O), 66.4 (C_{4gal}), 64.4 (OCH₂C=C), 62.0 (C_q), 61.9 ($C_{6glc} + C_qCH_2O$), 60.7 (C_{6gal}), 60.3 (OCH₂), 52.5 (N_{triazole}CH₂CONH), 52.1 (COCH₂N₃), 49.9 (NCH₂), 20.7, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.8 (m, 3P).

m/z (ESI⁺ MS) for C₆₃₈H₈₉₇CIN₆₉O₃₄₇P₃ = 15221.1 [*M*+H]⁺, found 15220.6 (After deconvolution). GPC measurements (THF): $M_w = 16580$; $M_n = 15400$, PDI (M_w/M_n) = 1.077.









Figure S133. ³¹P NMR spectrum of compound **30** (CDCl₃, 122MHz)



Figure S134. ESI+-MS spectrum (deconvolution) of compound 30



Figure S135. GPC trace (in THF) for compound 30
AB₂₅ glycodendron with a focal N₃ 31



To a solution of cyclotriphosphazene core **6** (1.86 mg, 1.76 µmol, 1.0 eq.) in a 1:1 mixture of H_2O/THF_{anh} (5 mL), were added azido derivative **15** (60 mg, 11.4 µmol, 6.5 eq.), $CuSO_4 \cdot 5H_2O$ (2.86 mg, 11.4 µmol, 6.5 eq.) and sodium ascorbate (2.23 mg, 11.4 µmol, 6.5 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 85:15) afforded desired multivalent compound **31** (40.0 mg, 1.46 µmol, **83%**) as a yellowish foam.

R_f = 0.32, DCM/MeOH 92:8.

¹**H NMR** (600 MHz, $CDCI_3$, δ ppm): 10.15 (br s, 5H, N*H*), 9.45 (br s, 1H, N*H*COCH₂Cl), 8.03 + 7.98 (2×s, 5H, *H*_{int.triazole}), 7.89-7.83 (m, 25H, *H*_{ext.triazole}), 7.35-7.30 (m, 12H, *CH*_b), 6.97-6.54 (m, 132H, *CH*_c), *CH*_c, *CH*_c), 5.40 (br s, 1.4) (br s, 1

– S109 –

10H, N_{triazole}CH₂CONH), 5.32 (d_{app}, 25H, H_{4gal}), 5.17-5.04 (m, 110H, H_{3glc} , C_qCH₂O, H_{2gal}), 4.93 (dd, ${}^{3}J_{2,3}$ = 3.3 Hz, ${}^{3}J_{3,4}$ = 7.1 Hz, 25H, H_{3gal}), 4.87-4.82 (m, 25H, H_{2glc}), 4.54-4.42 (m, 125H, CH₂N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.22 (br s, 2H, NHCOCH₂Cl), 4.11-4.02 (m, 75H, H_{6aglc} , H_{6agal} , H_{6bgal}), 3.92-3.44 (m, 425H, H_{5gal} , H_{5glc} , H_{4glc} , OCH₂), 2.11-1.92 (m, 525H, COCH₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.2, 170.0, 169.9, 169.7, 169.5, 169.0 (7×s, COCH₃), 164.4 (*C*=O), 163.9 (*C*=O), 155.2 (*C*_a), 146.6 (*C*_{d'}), 144.5 (*C*_d), 144.2 (*C*_d), 143.5 (*C*_{triazole}=CH), 143.3 (*C*_{triazole}=CH), 134.8 (*C*_{a'}), 125.3 (*C*_{triazole}=CH), 124.6 (*C*_{triazole}=CH), 124.1 (*C*_{triazole}=CH), 121.9 (*C*_c), 121.8 (*C*_{b'}), 120.9 (*C*_c), 120.8 (*C*_c), 116.0 (*C*_b), 115.3 (*C*_b), 115.1 (*C*_b), 101.0 (*C*_{1gal}), 100.5 (*C*_{1glc}), 76.2 (*C*_{4glc}), 72.7 (*C*_{3glc}), 72.5 (*C*_{5glc}), 71.5 (*C*_{2glc}), 70.9 (*C*_{3gal}), 70.5 (*C*_{5gal}), 70.5, 70.4, 70.4, 70.3, 70.2, 70.1, 70.1, 69.3, 69.1, 69.0 (OCH₂), 68.9 (*C*_{2gal}), 68.9 (OCH₂), 66.5 (*C*_{4gal}), 62.1 (OCH₂C=C), 61.9 (*C*_{6glc}), 61.6 (OCH₂), 60.7 (*C*_{6gal}), 52.7 (N_{triazole}CH₂CONH), 50.3 (NCH₂), 50.1 (NCH₂), 43.5 (NHCOCH₂Cl), 20.8, 20.7, 20.6, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.2-9.7 (m, 18P).

m/z MALDI-TOF (DHB matrix) for C₁₁₆₈H₁₅₂₇CIN₁₁₄O₅₉₇P₁₈= 27309.0 ; found 27241.4.

NMR diffusion studies (CDCl₃): $D = 1.19 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 6.8 \text{ nm}$.

GPC measurements (CHCl₃/Et₃N (1%)): $M_w = 26040$; $M_n = 25230$, PDI (M_w/M_n) = 1.032.



Figure S136. ¹H NMR spectrum of compound **31** (CDCI₃, 600MHz)



Figure S138. ¹³C NMR spectrum of compound 31 (CDCl₃, 150MHz)



Figure S140. ³¹P NMR spectrum of compound **31** (CDCl₃, 122MHz)



Figure S141. MALDI-TOF spectrum (DHB matrix) of compound 31



Figure S142. GPC trace (CHCl₃/Et₃N (1%)) for compound 31

Hypercore with 20 peripheral propargylic functions 34



To a solution of tetrathioacetylated pentaerythritol core **32** (3.42 mg, 9.28 µmol, 1.0 eq.) and chloroacetamide dendron **6** (51.0 mg, 48.3 µmol, 5.2 eq.) in dry EtOH (1.5 mL) were added at room temperature finely ground NaOH (2.97 mg, 74.3 µmol, 8.0 eq.) and NaBH₄ (3.00 mg, 74.4 µmol, 8.0 eq.) under a nitrogen atmosphere. The white solution was warmed up to 35°C for 3 h.. Insoluble brown oil quickly formed corresponding to the desired compound. The solvent was then removed *via* syringe and the residual oil was rinsed once with cold EtOH while stirring (2×2.5 mL). Finally, the oil was solubilized in CH₂Cl₂ and purified by column chromatography (SiO₂, CH₂Cl₂/MeOH 100:0 to 99:1) to afford desired icosapropargylated core **34** (31.0 mg, 7.71 µmol, **83%**) as an off-white foam.

$R_{f} = 0.29, CH_{2}CI_{2}/MeOH 98:2$

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 8.53 (br s, 4H, N*H*), 7.39 (d, 8H, J = 9.0 Hz, CH_b), 6.88-6.76 (m, 88H, CH_b , CH_c , CH_c), 4.63 (m, 40H, $OCH_2C\equiv CH$), 3.39 (br s, 8H, SCH_2CONH), 2.84 (br s, 8H, C_qCH_2S), 2.54 (m, 20H, $C\equiv CH$).

¹³**C NMR** (150 MHz, CDCl₃, δ ppm): 166.4 (*C*=O), 154.5 (*C*_a), 147.1 (*C*_d'), 144.7 (*C*_d (×2)), 134.3 (*C*_a'), 121.8 (*C*_c (×2)), 121.4 (*C*_b'), 121.0 (*C*_c'), 115.4 (*C*_b (×2)), 78.6 (*C*=CH), 78.4 (*C*=CH), 76.0 (C=CH), 75.8 (C=CH (×2)), 56.2 (OCH₂ (×3)), 44.0 (*C*_q), 38.5 (SCH₂CONH), 37.4 (*C*_qCH₂S).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.71 (t_{app}, 12P).

m/z (ESI⁺-MS) for C₂₁₇H₁₇₆N₁₆O₄₈P₁₂S₄ = 2137.3851 [*M*+2H]²⁺; found 2137.3857; 1425.2592 [*M*+3H]³⁺; found 1425.2595.

GPC measurements (CHCl₃): $M_w = 4625$; $M_n = 4540$, PDI (M_w/M_n) = 1.019.



















Abund (% first)

100 247.58

332.95

318.32

240.47

151.78

82.84

17.38

6.88

2.5

0.84

0.27

0.08

40

Figure S148. ESI⁺-HRMS spectrum of compound 34





Hypercore with 30 peripheral propargylic functions 35

To a solution of hexakis(thioacetyl)benzene core 33^4 (3.67 mg, 6.02 µmol, 1.0 eq.) and chloroacetamide dendron **6** (50.0 mg, 47.4 µmol, 7.9 eq.) in dry EtOH (2.0 mL) were added at room temperature finely ground NaOH (3.50 mg, 87.5 µmol, 14.5 eq.) and NaBH₄ (3.50 mg, 92.5 µmol, 15.3 eq.) under a nitrogen atmosphere. The white solution was warmed up to 35°C for 3 h. and an insoluble oil quickly formed. The solvent was removed *via* syringe and the residual oil was rinced with EtOAc while stirring (3×10 mL). Finally, the oil was solubilized in CH₂Cl₂ and purified by column chromatography (SiO₂, CH₂Cl₂/MeOH 100:0 to 99.5:0.5) to afford desired tricontapropargylated core **35** (29 mg, 4.48 µmol, **74%**) as an off-white foam. *Degradation occurs after 3 days at -20°C*.

$R_{f} = 0.11, CH_{2}CI_{2}/MeOH 98:2$

¹**H NMR** (300 MHz, CDCl₃, δ ppm): 8.58 (br s, 6H, N*H*), 7.42 (d, 12H, J = 9.0 Hz, C*H*_b), 6.90-6.73 (m, 132H, C*H*_b, C*H*_c, C*H*_c), 4.60 (m, 60H, OC*H*₂C≡CH), 4.12 (br s, 12H, C_{core}C*H*₂S), 3.28 (br s, 12H, SC*H*₂CONH), 2.53 (m, 30H, C≡C*H*).

¹³C NMR (75 MHz, CDCl₃, δ ppm): 167.7 (C=O), 154.6 (C_a), 147.2 (C_d), 144.7 (C_d), 136.2 (C_{q core}), 134.6 (C_a), 121.7 (C_c), 121.5 (C_b), 121.2 (C_c), 115.4 (C_b), 78.7 (C=CH), 78.6 (C=CH), 75.9 (C=CH), 75.9 (C=CH), 56.3 (OCH₂), 56.2 (OCH₂), 37.8 (SCH₂CONH), 31.2 (C_{core}CH₂S).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.69 (s_{app}, 18P).

m/z (MALDI-TOF/DHB matrix) for C₃₃₀H₂₆₄N₂₄O₇₂P₁₈S₆ = 6467.6; found 6468.9.

GPC measurements (THF): $M_w = 7152$; $M_n = 6399$, PDI (M_w/M_n) = 1.12.







Figure S154. MALDI-TOF (DHB matrix) spectrum of compound 35 (insert: MALDI-TOF of 35, 3 days after synthesis)



Figure S155. GPC trace (in THF) for compound 35

Icosavalent glycodendrimer 36



To a solution of icosapropargylated core **34** (11.0 mg, 2.58 µmol, 1.0 eq.) in a 1:1 mixture of H₂O/THF (4.0 mL) were added azido derivative **10** (72.0 mg, 0.086 mmol, 33.3 eq.), CuSO₄·5H₂O (17.4 mg, 0.069 mmol, 27.0 eq.) and sodium ascorbate (13.7 mg, 0.069 mmol, 27.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Noteworthy is the fact that 5 mg of CuSO₄·5H₂O and 4 mg of sodium ascorbate were added to the solution after the 3 hours of heating. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 85:15) afforded desired multivalent compound **36** (43.0 mg, 2.00 µmol, **74%**) as a yellowish foam.

R_f = 0.40, DCM/MeOH 92:8

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 9.70 (br s, 4H, N*H*), 7.85-7.84 (2×s, 20H, $H_{triazole}$), 7.44-7.40 (m, 8H, C H_b), 6.90-6.56 (m, 88H, C H_b , C H_c , C H_c), 5.32 (d_{app}, 20H, H_{4gal}), 5.18-5.06 (m, 80H, H_{3glc} , C_qC H_2 O, H_{2gal}), 4.95 (dd, ³ $J_{2,3}$ = 3.3 Hz, ³ $J_{3,4}$ = 7.1 Hz, 20H, H_{3gal}), 4.88-4.83 (m, 20H, H_{2glc}), 4.54-4.42 (m, 100H, C H_2 N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.13-4.05 (m, 60H,

– S123 –

*H*_{6bglc}, *H*_{6agal}, *H*_{6bgal}), 3.88-3.45 (m, 348H, *H*_{5gal}, *H*_{5glc}, *H*_{4glc}, OC*H*₂, SC*H*₂CONH), 2.81 (br s, 8H, C_qC*H*₂S), 2.11-1.92 (m, 420H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.2, 170.1, 170.0, 169.7, 169.5, 169.0 (7×s, COCH₃), 167.7 (NHC=O), 155.2 (C_a), 146.5 (C_d), 144.4 (C_d), 144.3 (C_d), 143.4 ($C_{triazole}$ =CH), 143.3 ($C_{triazole}$ =CH), 135.4 (C_a), 124.5 ($C_{triazole}$ =CH), 124.1 ($C_{triazole}$ =CH), 121.8 (C_c), 121.7 (C_c), 121.0 (C_c), 120.9 (C_b), 115.2 (C_b), 101.0 (C_{1gal}), 100.5 (C_{1glc}), 76.2 (C_{4glc}), 72.7 (C_{3glc}), 72.5 (C_{5glc}), 71.5 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.4, 70.4, 70.3, 70.2, 70.1, 70.1, 69.3, 69.1, 69.0 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 66.5 (C_{4gal}), 62.2 (OCH₂C=C), 61.9 (C_{6glc}), 61.8 (OCH₂), 60.8 (C_{6gal}), 50.2 (NCH₂), 50.1 (NCH₂), 44.0 (C_q , not *visible*), 38.4 (SCH₂CONH), 37.6 (C_qCH_2S), 29.6, 20.8, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.8 (t_{app}, 12P).

m/z (MALDI-TOF-MS/DHB matrix) for C₈₉₇H₁₁₉₆N₇₆O₄₆₈P₁₂S₄= 21031.3; found 21098.1 (*with successive losses of monomers and dendrons*).

NMR diffusion studies (CDCl₃): $D = 1.17 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 7.0 \text{ nm}$.

GPC measurements (CHCl₃/Et₃N (1%)): $M_w = 25870$; $M_n = 24610$, PDI (M_w/M_n) = 1.051.



Figure S156. ¹H NMR spectrum of compound 36 (CDCl₃, 600MHz)





Figure S159. ³¹P NMR spectrum of compound 35 (CDCI₃, 122MHz)



Figure S160. MALDI-TOF spectrum of compound 36 with successive fragmentations.



Figure S161. GPC trace (CHCl₃/Et₃N (1%)) for compound 36



Defectuous glycodendrimer 36a (location of propargylic functions is arbitrary)

To a solution of icosapropargylated core **34** (10.2 mg, 2.39 µmol, 1.0 eq.) in a 1:1 mixture of H₂O/THF_{anh} (4.0 mL) were added azido derivative **10** (32.0 mg, 0.038 mmol, 16.0 eq.), CuSO₄·5H₂O (12.0 mg, 0.048 mmol, 20.0 eq.) and sodium ascorbate (9.5 mg, 0.048 mmol, 20.0 eq.). While stirring, the mixture was first heated at 50°C for 3 h ours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 85:15) afforded desired mixture of defectuous multivalent compounds containing 4-5 propargylic functions on average **36a** (36.0 mg, 2.04 µmol, **85%**) as an off-white foam.

$R_{f} = 0.40$, DCM/MeOH 92:8.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 9.70 (br s, 4H, N*H*), 7.85-7.84 (2×s, 16H, $H_{triazole}$), 7.44-7.40 (m, 8H, C H_b), 6.90-6.56 (m, 88H, C H_b , C H_c , C H_c), 5.32 (d_{app}, 16H, H_{4gal}), 5.18-5.06 (m, 64H, H_{3glc} , C_qC H_2 O, H_{2gal}), 4.95 (dd, ³ $J_{2,3}$ = 3.3 Hz, ³ $J_{3,4}$ = 7.1 Hz, 16H, H_{3gal}), 4.88-4.83 (m, 16H, H_{2glc}), **4.65 (m, ~8H, CH₂OC=CH)**, 4.54-4.42 (m, 80H, C H_2 N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.13-4.05 (m, 48H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.88-3.45 (m, 280H, H_{5gal} , H_{5glc} , H_{4glc} , OC H_2 , SC H_2 CONH), 2.81 (br s, 8H, C_qC H_2 S), **2.53 (br s, ~4H, CH₂OC=CH)**, 2.11-1.92 (m, 336H, COC H_3).

– S128 –

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.2, 170.1, 170.0, 169.7, 169.5, 169.0 (7×s, COCH₃), 167.7 (NHC=O), 155.3 (C_a), 154.4 (C_a), 146.5 (C_d), 144.5 (C_d), 144.3 (C_d), 143.4 ($C_{triazole}$ =CH), 143.3 ($C_{triazole}$ =CH), 135.4 (C_a), 124.5 ($C_{triazole}$ =CH), 124.5 ($C_{triazole}$ =CH), 124.1 ($C_{triazole}$ =CH), 121.8 (C_c), 121.7 (C_c), 121.0 (C_c), 120.9 (C_b), 115.5 (C_b), 115.2 (C_b), 101.0 (C_{1gal}), 100.5 (C_{1glc}), **78.7 (C=CH)**, **78.5 (C=CH)**, 76.2 (C_{4glc}), **76.0 (C=CH)**, **75.8 (C=CH)**, 72.8 (C_{3glc}), 72.5 (C_{5glc}), 71.6 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.5, 70.4, 70.4, 70.3, 70.2, 70.1, 70.1, 69.3, 69.1, 69.0 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 66.5 (C_{4gal}), 62.2 (OCH₂C=C), 61.9 (C_{6glc}), 61.8 (OCH₂), 60.7 (C_{6gal}), **56.1 (OCH₂C=CH)**, 50.1 (NCH₂), 50.1 (NCH₂), 44.2 (C_q), 38.4 (SCH₂CONH), 37.4 (C_qCH_2S), 29.6, 20.8, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.8 (t_{app} , 12P).

m/z (MALDI-TOF-MS/DHB matrix) : From 21005.2 (click completion) to 7574 with signals corresponding to successive losses of monomers (~876 MW).



Figure S162. ¹H NMR spectrum of compound 36a (CDCl₃, 600MHz)



– S130 –



Figure S166. ³¹P NMR spectrum of compound 36a (CDCI₃, 122MHz)



Figure S167. MALDI-TOF (DHB matrix) spectrum of compound 36a with successive losses of monomers.

Protected tricontavalent glycodendrimer 37



To a solution of tricontapropargylated core **35** (11.0 mg, 1.70 µmol, 1.0 eq.) in a 1:1 mixture of H₂O/THF_{anh} (4.0 mL) were added azido derivative **10** (71.2 mg, 0.085 mmol, 50.0 eq.), CuSO₄·5H₂O (17.0 mg, 0.068 mmol, 40.0 eq.) and sodium ascorbate (13.5 mg, 0.068 mmol, 40.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Noteworthy is the fact that 6mg of CuSO₄·5H₂O and 4mg of sodium ascorbate were added to the solution after the 3 hours of heating. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 85:15) afforded desired multivalent compound **37** (40.0 mg, 1.27 µmol, **74%**) as a yellowish foam.

R_f = 0.27, DCM/MeOH 94:6

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 9.82 (br s, 6H, N*H*), 7.85-7.84 (br s, 30H, *H*_{triazole}), 7.46-7.40 (m, 12H, C*H*_b), 6.95-6.50 (m, 132H, C*H*_b, C*H*_c, C*H*_c), 5.33 (d_{app}, 30H, *H*_{4gal}), 5.19-5.07 (m, 120H, *H*_{3glc}, C_qC*H*₂O, *H*_{2gal}), 4.94 (dd, ³*J*_{2,3} = 3.3

Hz, ${}^{3}J_{3,4} = 7.1$ Hz, 30H, H_{3gal}), 4.88-4.83 (m, 30H, H_{2glc}), 4.55-4.46 (m, 150H, $CH_{2}N$, H_{1glc} , H_{6aglc} , H_{1gal}), 4.20 (br s, 12H, $C_{core}CH_{2}S$), 4.13-4.05 (m, 90H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.89-3.40 (m, 522H, H_{5gal} , H_{5glc} , H_{4glc} , OCH₂, SCH₂CONH), 2.11-1.92 (m, 630H, COCH₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.2, 170.1, 170.0, 169.7, 169.5, 169.0 (7×s, COCH₃), 169.3 (NHC=O), 155.2 (C_a), 154.5 (C_a), 146.5 (C_d) (not visible), 144.4 (C_d), 144.3 (C_d), 143.4 ($C_{triazole}$ =CH), 143.3 ($C_{triazole}$ =CH), 136.2 (C_q core), 135.4 (C_a), 124.5 ($C_{triazole}$ =CH), 124.1 ($C_{triazole}$ =CH), 121.8 (C_c), 121.7 (C_c), 121.0 (C_c), 120.9 (C_b), 115.2 (C_b), 101.0 (C_{1gal}), 100.5 (C_{1glc}), 76.2 (C_{4glc}), 72.7 (C_{3glc}), 72.5 (C_{5glc}), 71.5 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.5, 70.4, 70.4, 70.3, 70.2, 70.1, 70.1, 69.3, 69.1, 69.0 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 66.5 (C_{4gal}), 62.1 (OCH₂C=C), 61.9 (C_{6glc}), 61.8 (OCH₂), 60.7 (C_{6gal}), 50.2 (NCH₂), 50.1 (NCH₂), 37.8 (SCH₂CONH), 31.2 (C_{core} CH₂S) (not visible), 29.6, 20.8, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 9.8 (m, 18P).

m/z (MALDI-TOF-MS/DHB matrix) for C₁₃₅₀H₁₇₉₄N₁₁₄O₇₀₂P₁₈S₆ = 31600; found 31478 (*with signals corresponding to successive losses of monomer*(s).

NMR diffusion studies (CDCl₃): $D = 1.34 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 6.2 \text{ nm}$.

GPC measurements (CHCl₃/Et₃N (1%)): M_w = 42710 ; M_n = 39580 , PDI (M_w/M_n) = 1.079.



Figure S168. ¹H NMR spectrum of compound 37 (CDCl₃, 600MHz)





-50 f1 (ppm) 140 120 100 80 60 20 -230 40 0 -10 -30 -70 -90 -110 -140 -170 -200 Figure S171. ³¹P NMR spectrum of compound 37 (CDCl₃, 122MHz)



Figure S172. GPC trace (CHCl₃/Et₃N (1%)) for compound 37



Figure S173. MALDI-TOF (DHB matrix) spectrum of compound 37 with successive losses of monomers

Hydroxylated tricontavalent glycodendrimer 38



Acetylated compound **37** (34.0 mg, 1.08 µmol) was dissolved in a dry mixture of MeOH/DCM (5 mL, 4:1) and a solution of sodium methoxide (1M in MeOH, 5 µL every 20 minutes until precipitation) was added. An additional 100 µL was then injected and the heterogeneous reaction mixture was stirred at room temperature for 18 hours. The solvent was then removed with a Pasteur pipette and another dry mixture of MeOH/DCM (4 mL, 3:1) is added to the residual oil. A vigorous agitation is maintained for an additional 15 min. period. After removal of the solvents with a Pasteur pipette, 2 mL of dry MeOH were added to the viscous residue under agitation. After 15 min., the MeOH was removed and the residue was dissolved in 3 mL of H_2O , and the pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected tricontavalent derivative **38** as an off-white solid (20.0 mg, 0.88 µmol, **83%**).

¹**H NMR** (600 MHz, D₂O, δ ppm): 7.99 (m, 30H, $H_{triazole}$), 7.28 (br s, 12H, CH_b), 6.99 (br s, 132H, CH_b, CH_c, CH_c), 5.10-4.90 (br s, 60H, C_{q-triazole}CH₂O), 4.58-4.32 (m, 132H, N_{triazole}CH₂CH₂, C_{core}CH₂S, H_{1glc}, H_{1gal}), 3.97-3.25 (m, 792H,

 $OCH_2CH_2N, LacOCHHCH_2, H_{3glc}, H_{2gal}, H_{3gal}, H_{6bglc}, H_{4gal}, H_{6agal}, H_{6agal}, H_{6agal}, H_{5gal}, H_{5glc}, H_{4glc}, LacOCHHCH_2, OCH_2, H_{2glc}, SCH_2CONH).$

¹³**C NMR** (150 MHz, D₂O, δ ppm): ~165.0 (C=O) (not visible), 155.6 (C_a), 145.0 (C_d) (not visible), 144.6 (C_d), 143.7 (C_{triazole}=CH), 136.2 (C_{q core}) (not visible), 130.7 (C_a) (not visible), 126.0 (C_{triazole}=CH), 122.4 (C_c + C_b + C_c), 116.2 (C_b), 103.7 (C_{1gal}), 102.8 (C_{1glc}), 79.1 (C_{4glc}), 76.0 (C_{3glc}), 75.4 (C_{5glc}), 75.0 (C_{5gal}), 73.5 (C_{2glc}), 73.2 (C_{3gal}), 71.6 (C_{2gal}), 70.2, (OCH₂), 69.4 (C_{4gal}), 69.2 (OCH₂), 62.1 (OCH₂C=C), 61.7 (C_{6glc}), 60.8 (C_{6gal}), 50.7 (NCH₂), 37.8 (SCH₂CONH) (not visible), 31.2 (C_{core}CH₂S) (not visible).

³¹**P NMR** (122 MHz, D₂O, δ ppm): 10.1 (br s, 18P).

m/z (MALDI-TOF/ DHB matrix) for $C_{930}H_{1374}N_{114}O_{492}P_{18}S_6 = 22773.2$, found 22862.0.

NMR diffusion studies (D₂O): $D = 0.65 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 6.2 \text{ nm}$.



Figure S174. ¹H NMR spectrum of compound 38 (D₂O, 600MHz)



Figure S176. ¹³C NMR spectrum of compound 38 (D_2O , 150MHz, acetone as reference)



Figure S177. 31 P NMR spectrum of compound 38 (D₂O, 122MHz)



Figure S178. MALDI-TOF (DHB matrix) spectrum of compound 38 with successive losses of monomers

Hexacontavalent glycodendrimer 39



To a solution of icosapropargylated cyclotriphosphazene derivative **34** (2.35 mg, 0.55 μ mol, 1.0 eq.) in a 1:1 mixture of H₂O/THF_{anh} (3 mL), were added azido derivative **14** (51.8 mg, 18.3 μ mol, 33.3 eq.), CuSO₄·5H₂O (4.7 mg, 18.6 μ mol, 34.0 eq.) and sodium ascorbate (3.7 mg, 18.6 μ mol, 34.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Noteworthy is the fact that 2.5 mg of CuSO₄·5H₂O and 2mg of sodium ascorbate were added to the solution after the 3 hours of heating. Ethyl acetate (30 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (20 mL), washed with saturated aqueous NH₄Cl (2×20 mL), water (20 mL) and brine (10 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 90:10) afforded desired multivalent compound **39** (26.0 mg, 0.43 μ mol, **74%**) as a yellowish oil.

R_f = 0.15, DCM/MeOH 94:6.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 7.93 (s, 20H, $H_{int-triazole}$), 7.73 (s, 60H, $H_{ext-triazole}$), 7.28 (br s, 12H, $CH_{b'}$ + NH_{int}), 6.99-6.60 (br s, 108H, CH_b , CH_c , CH_c' + NH_{ext}), 5.33 (br s, 60H, H_{4gal}), 5.19 (t_{app}, 60H, H_{3glc}), 5.10-5.02 (m, 100H, CqtriazoleCH₂O, H_{2gal}), 4.95 (dd, ³J_{2,3} = 3.4 Hz, ³J_{3,4} = 7.0 Hz, 60H, H_{3gal}), 4.83 (t_{app}, 60H, H_{2glc}), 4.54-4.47 (m, 420H, CqtriazoleCH₂OCH₂, OCH₂CH₂N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.10-4.02 (m, 180H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.86-3.53 (m, 1188H, CqtriazoleCH₂OCH₂Ctriazole, NHCOCH₂N_{triazole}, OCH₂CH₂N, H_{5gal} , H_{5glc} , H_{4glc} , OCH₂, HNCOCH₂S), 2.82 (br s, 8H, CqCH₂S), 2.11-1.92 (m, 1260H, COCH₃). ¹³C NMR (150 MHz, CDCl₃, δ ppm (<u>145000 scans</u>)): 170.2, 170.2, 170.0, 169.9, 169.6, 169.5, 168.9 (7×s, COCH₃), 166.3 (CONH_{int}), 165.2 (CONH_{ext}), 155.3 (C_a), 154.0 (C_a), 149.9 (C_d), 144.3 (C_d), 144.2 (C_{triazole}=CH), 143.3 (C_{triazole}=CH), 135.2 (C_a), 125.4 (C_{triazole}=CH), 123.9 (C_{triazole}=CH), 121.8 (C_c), 121.4 (C_b), 116.1 (C_b), 115.2 (C_b), 101.0 (C_{1gal}), 100.4 (C_{1glc}), 76.2 (C_{4glc}), 72.8 (C_{3glc}), 72.6 (C_{5glc}), 71.6 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.4, 70.3, 70.2, 70.2, 70.0, 69.3 (OCH₂), 69.0 (C_{2gal}), 68.9 (OCH₂), 68.6 (C_qCH₂O), 66.6 (C_{4gal}), 64.5 (OCH₂C=C), 62.0 (C_{6glc}), 61.9 (C_q), 60.6 (C_{6gal}), 60.3 (NCH₂), 52.5 (HNCOCH₂N_{triazole}), 49.9 (OCH₂), 45.9 (C_q), 37.4 (SCH₂CONH), 37.0 (C_qCH₂S), 20.9, 20.8, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.0-9.6 (m, 12P).

m/z (MALDI-TOF/DHB matrix) for C₂₅₅₇H₃₅₉₆N₂₇₆O₁₃₈₈P₁₂S₄= 60908.9; found: 59682-centered Gaussian.

NMR diffusion studies (CDCl₃): $D = 1.07 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 7.6 \text{ nm}$.

GPC measurements (CHCl₃/Et₃N (1%)): M_w = 76570 ; M_n = 75530 , PDI (M_w/M_n) = 1.014.







Figure S181. ¹³C NMR spectrum of compound **39** (CDCI₃, 150MHz, 145000 scans)


Figure S182. ³¹P NMR spectrum of compound **39** (CDCl₃, 122MHz)



Figure S183. MALDI-TOF (DHB matrix) of compound 39



Figure S184. GPC trace (CHCl₃/Et₃N (1%)) for compound 39

Defectuous glycodendrimer 39a (location of propargylic functions is arbitrary)



To a solution of icosapropargylated cyclotriphosphazene derivative **34** (2.66 mg, 0.62 µmol, 1.0 eq.) in a 1:1 mixture of H_2O/THF_{anh} (3 mL), were added azido derivative **14** (30.0 mg, 10.6 µmol, 17.0 eq.), CuSO₄·5H₂O (5.3 mg, 21.1 µmol, 34.0 eq.) and sodium ascorbate (4.2 mg, 21.1 µmol, 34.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (30 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (20 mL), washed with saturated aqueous NH₄Cl (2×20 mL), water (20 mL) and brine (10 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 90:10) afforded desired mixture of compounds presenting an average of 3 remaining propargylic functionalities **39a** (24.0 mg, 0.47 µmol, **75%**) (*based on a MW* = *52414 for 17 grafted dendrons*) as a colorless oil.

R_f = 0.15, DCM/MeOH 94:6.

¹H NMR (600 MHz, CDCl₃, δ ppm): 7.93 (s, 17H, $H_{int-triazole}$), 7.73 (s, 48H, $H_{ext-triazole}$), 7.28 (br s, 12H, $CH_{b'}$ + NH_{int}), 6.99-6.60 (br s, 105H, CH_{b} , CH_{c} , $CH_{c'}$ + NH_{ext}), 5.33 (br s, 51H, H_{4gal}), 5.19 (t_{app} , 51H, H_{3glc}), 5.10-5.02 (m, 85H, $C_{q-triazole}CH_2O$, H_{2gal}), 4.95 (dd, ${}^{3}J_{2,3}$ = 3.4 Hz, ${}^{3}J_{3,4}$ = 7.0 Hz, 51H, H_{3gal}), 4.83 (t_{app} , 51H, H_{2glc}), **4.70-4.60 (br s, 6H, OCH₂C=CH res.)**, 4.54-4.47 (m, 357H, $C_{q-triazole}CH_2OCH_2$, OCH₂CH₂N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.10-4.02 (m, 153H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.86-3.53 (m, 1011H, $C_{q-triazole}CH_2OCH_2C_{triazole}$, NHCOCH₂N_{triazole}, OCH₂CH₂N, H_{5gal} , H_{5glc} , H_{4glc} , OCH₂, HNCOCH₂S), 2.82 (br s, 8H, C_qCH_2 S), **2.65 (2×br s, 3H, OCH₂C=CH res.)**, 2.11-1.92 (m, 1071H, COCH₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.2, 170.2, 170.0, 169.9, 169.6, 169.5, 168.9 (7×s, COCH₃), 166.3 (CONH_{int}), 165.2 (CONH_{ext}), 155.3 (C_a), 154.0 (C_a), 149.9 (C_d), 144.3 (C_d), 144.2 (C_{triazole}=CH), 143.3 (C_{triazole}=CH), 135.2 (C_a),

125.4 ($C_{triazole}=CH$), 123.9 ($C_{triazole}=CH$), 121.8 (C_c), 121.4 (C_b), 116.1 (C_b), 115.2 (C_b), 101.0 (C_{1gal}), 100.4 (C_{1glc}), 76.2 (C_{4glc}), 72.8 (C_{3glc}), 72.6 (C_{5glc}), 71.6 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.4, 70.3, 70.2, 70.2, 70.0, 69.3 (OCH₂), 69.0 (C_{2gal}), 68.9 (OCH₂), 68.6 (C_qCH_2O), 66.6 (C_{4gal}), 64.5 (OCH₂C=C), 62.0 (C_{6glc}), 61.9 (C_q), 60.6 (C_{6gal}), 60.3 (NCH₂), **56.1 (OCH₂C=CH res.)**, 52.5 (HNCOCH₂N_{triazole}), 49.9 (OCH₂), 45.9 (C_q), 37.4 (SCH₂CONH), 37.0 (C_qCH_2S), 20.9, 20.8, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

m/z (MALDI-TOF/DHB matrix), found: 50696-centered Gaussian.

GPC measurements (CHCl₃/Et₃N (1%)): $M_w = 73780$; $M_n = 72270$, PDI (M_w/M_n) = 1.021



Figure S185. ¹H NMR spectrum of compound **39a** (CDCl₃, 600MHz)



Figure S187. ¹³C NMR spectrum of compound **39a** (CDCl₃, 150MHz, 58000 scans)



Figure S188. MALDI-TOF (DHB matrix) spectrum of compound 39a



Protected nonacontavalent glycodendrimer 40



To a solution of tricontapropargylated cyclotriphosphazene derivative **35** (3.80 mg, 0.59 µmol, 1.0 eq.) in a 1:1 mixture of H_2O/THF_{anh} (4 mL), were added azido dendron **14** (83.2 mg, 29.4 µmol, 50.0 eq.), CuSO₄·5H₂O (6.10 mg, 23.5 µmol, 40.0 eq.) and sodium ascorbate (4.80 mg, 23.5 µmol, 40.0 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temper ature for additional 18 hours. Noteworthy is the fact that 3.0 mg of CuSO₄·5H₂O and 2.0 mg of sodium ascorbate were added to the solution after the 3 hours of heating. Ethyl acetate (5 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (20 mL), washed with saturated aqueous NH₄Cl (2×20 mL), water (20 mL) and brine (10 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 90:10) afforded desired multivalent compound **40** (38.0 mg, 0.42 µmol, **72%**) as a pale oil.

R_f = 0.13, DCM/MeOH 94:6.

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 7.93 (s, 30H, $H_{int-triazole}$), 7.73 (s, 90H, $H_{ext-triazole}$), 7.28 (br s, 42H, $CH_{b'}$ + NH_{ext}), 6.99-6.60 (br s, 138H, CH_b , CH_c , $CH_{c'}$ + NH_{int}), 5.32 (br s, 90H, H_{4gal}), 5.16 (t_{app} , 90H, H_{3glc}), 5.10-5.02 (m, 150H, $C_{q-triazole}CH_2O$, H_{2gal}), 4.95 (dd, ${}^{3}J_{2,3}$ = 3.4 Hz, ${}^{3}J_{3,4}$ = 7.0 Hz, 90H, H_{3gal}), 4.86 (t_{app} , 90H, H_{2glc}), 4.54-4.47 (m, 630H, $C_{q-triazole}CH_2OCH_2$, OCH₂CH₂N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.14-4.02 (m, 282H, $C_{core}CH_2S$, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.95-3.25 (m, 1782H, $C_{q-triazole}CH_2OCH_2Ct_{riazole}$, NHCOCH₂N_{triazole}, OCH₂CH₂N, H_{5gal} , H_{5glc} , H_{4glc} , OCH₂, HNCOCH₂S), 2.11-1.92 (m, 1890H, COCH₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.2, 170.2, 170.0, 169.9, 169.6, 169.5, 168.9 (7×s, COCH₃), 165.9 (CONH_{int}), 165.6 (CONH_{ext}), 155.3 (C_a), 149.9 (C_d) (not visible), 144.3 (C_d), 144.2 (C_{triazole}=CH), 143.3 (C_{triazole}=CH), 136.2 (C_{q core})

– S150 –

(not visible), 135.2 ($C_{a'}$) (not visible), 125.5 ($C_{triazole}=CH$), 123.9 ($C_{triazole}=CH$), 121.8 (C_c), 121.4 (C_b), 116.1 (C_b), 115.1 (C_b), 101.0 (C_{1gal}), 100.5 (C_{1glc}), 76.2 (C_{4glc}), 72.8 (C_{3glc}), 72.6 (C_{5glc}), 71.6 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.4, 70.3, 70.2, 70.2, 70.0, 69.3 (OCH₂), 69.0 (C_{2gal}), 68.9 (OCH₂), 68.6 (C_qCH_2O), 66.6 (C_{4gal}), 64.5 (OCH₂C=C), 62.0 (C_{6glc}), 61.9 (C_q), 60.6 (C_{6gal}), 60.3 (NCH₂), 52.5 (HNCOCH₂N_{triazole}), 49.9 (OCH₂), 37.3 (SCH₂CONH), 31.2 ($C_{core}CH_2S$), 20.9, 20.8, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.0-9.2 (m, 18P).

m/z (MALDI-TOF/DHB matrix) for $C_{3840}H_{5394}N_{414}O_{2082}P_{18}S_6$ = 91417.4; found: 79824-centered Gaussian. NMR diffusion studies (CDCl₃): *D* = 0.81×10⁻¹⁰ m²/s; *d*_s = 10.1 nm.



Figure S190. ¹H NMR spectrum of compound 40 (CDCl₃, 600MHz)



Figure S192. ¹³C NMR spectrum of compound 40 (CDCl₃, 150MHz, 60000 scans)



Figure S193. ³¹P NMR spectrum of compound 40 (CDCl₃, 122MHz)



Figure S194. MALDI-TOF (DHB matrix) spectrum of compound 40

Hydroxylated nonacontavalent glycodendrimer 41



Acetylated compound **40** (22.0 mg, 0.24 µmol) was dissolved in dry MeOH/DCM (3+0.5 mL) and a solution of sodium methoxide (1M in MeOH, 5 µL every 20 minutes until precipitation) was added. An additional 100 µL was then injected and the heterogeneous reaction mixture was stirred at room temperature for 18 hours. The solvent was then removed with a Pasteur pipette and another dry mixture of MeOH/DCM (5 mL, 4:1) is added to the residual oil. A vigorous agitation is maintained for an additional 15 min. period. After removal of the solvents with a Pasteur pipette, 2 mL of dry MeOH were added to the viscous residue under agitation. After 15 min., the MeOH was removed and the residue was dissolved in 3 mL of H₂O, and the pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected compound **41** as a white solid (14.0 mg, 0.22 µmol) in an 85% yield.

¹**H NMR** (600 MHz, D₂O, δ ppm): 8.15-7.89 (m, 120H, $H_{triazole}$), 7.55-7.28 (br s, 12H, C H_{b}), 6.84-6.77 (m, 132H, C H_{ar}), 5.20-5.14 (m, 120H, NHCOC H_2 N, C_qC H_2 O), 4.62-4.44 (m, 540H, C H_2 N, C_{q-triazole}C H_2 O, 36H, H_{1glc} , H_{1gal}), 4.02-3.55 (m, 2430, OC H_2 CH₂N, H_{3glc} , H_{2gal} , H_{3gal} , H_{6bglc} , H_{6agal} , H_{6aglc} , H_{5gal} , H_{5glc} , H_{4glc} , H_{4gal} , LacOC H_2 CH₂, OC H_2 , NHC_qC H_2 O), 3.22 (t_{app}, 90H, H_{2glc}).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 167.1 (C_{ext} ONH), 165.0 (C_{int} ONH) (*not visible*), 155.8-155.7 (2× C_a), 144.6 (C_d), 143.7 ($C_{triazole}$ =CH), 136.2 ($C_{ar-core}$) (*not visible*), 127.0 ($C_{triazole}$ =CH), 125.9 ($C_{triazole}$ =CH), 122.4 (C_c), 116.3 (C_b), 103.6 (C_{1gal}), 102.8 (C_{1glc}), 79.1 (C_{4glc}), 76.0 (C_{3glc}), 75.4 (C_{5glc}), 75.0 (C_{5gal}), 73.5 (C_{2glc}), 73.2 (C_{3gal}), 71.6 (C_{2gal}), 70.3, 70.2, 70.2, 70.1 (OCH₂), 69.3 (C_{4gal}), 69.2 (OCH₂), 68.1 (C_q CH₂O), 64.2 (OCH₂C=C), 62.0 (C_q) (*not visible*), 61.7

 (C_{6glc}) , 60.9 (C_{6gal}) , 60.8 (OCH_2) , 53.0 $(N_{triazole}CH_2CO)$, 50.6 (CH_2N) , 37.8 (SCH_2CONH) (not visible), 31.2 $(C_{ar}CH_2S)$ (not visible).

 $^{31}\textbf{P}$ NMR (122 MHz, D2O, δ ppm): 10.7-9.5 (m, 18P).

 $\textit{m/z} \ (\text{MALDI-TOF/DHB matrix}) \ \text{for} \ C_{2580}H_{4134}N_{414}O_{1452}P_{18}S_6 = 64934.3, \ \text{found} \ 56530 \text{-centered} \ Gaussian.$

NMR diffusion studies (D₂O): $D = 0.60 \times 10^{-10} \text{ m}^2/\text{s}$; $d_s = 6.6 \text{ nm}$



Figure S195. ¹H NMR spectrum of compound 41 (D₂O, 600MHz)



Figure S197. ¹³C NMR spectrum of compound **41** (D₂O, 150MHz, acetone as reference, 81000 scans)



-50 f1 (ppm) . 140 120 100 . 80 40 0 -10 -30 60 20 -70 -90 -110 -140 -170 -200 -230

Figure S198. ³¹P NMR spectrum of compound 41 (D₂O, 122MHz)



Figure S199. MALDI-TOF (DHB matrix) spectrum of compound 41

Synthesis of monomeric reference 43



To a solution of 2-(2-{2-[2-(2-azido-ethoxy)-ethoxy]-ethoxy}-ethyl) 2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-acetyl- β -D-glucopyranoside **10**¹ (100.0 mg, 119.3 µmol, 1.0 eq.) in a 1:1 mixture of H₂O/THF_{anh} (5 mL), were added propargyl alcohol (28.1 mg, 29.1 µL, 501.3 µmol, 4.2 eq.), CuSO₄·5H₂O (14.9 mg, 59.7 µmol, 0.5 eq.) and sodium ascorbate (11.8 mg, 59.7 µmol, 0.5 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 92:8) afforded desired compound **S43** (86.0 mg, 96.6 µmol, **81%**) as a white foam.

R_f = 0.30, DCM/MeOH 94:6

¹**H NMR** (600 MHz, CDCl₃, δ ppm): 7.75 (s, 1H, $H_{triazole}$), 5.29 (d_{app}, 1H, H_{4gal}), 5.14 (dd, ${}^{3}J_{4,3} = 9.4$ Hz, ${}^{3}J_{3,2} = 9.1$ Hz, 1H, H_{3glc}), 5.05 (dd, ${}^{3}J_{2,1} = 10.5$ Hz, ${}^{3}J_{3,2} = 8.0$ Hz, 1H, H_{2gal}), 4.93 (dd, ${}^{3}J_{2,3} = 10.5$ Hz, ${}^{3}J_{3,4} = 3.4$ Hz, 1H, H_{3gal}), 4.83 (dd, ${}^{3}J_{2,1} = 9.4$ Hz, ${}^{3}J_{3,2} = 8.0$ Hz, 1H, H_{2glc}), 4.73 (br s, 2H, C H_{2} OH), 4.53 (d, ${}^{3}J_{1,2} = 9.4$ Hz, 1H, H_{1glc}), 4.50 (t_{app}, 2H, C H_{2} N), 4.48 (dd, ${}^{2}J_{6a.6b} = 12.0$ Hz, ${}^{3}J_{5.6a} = 2.1$ Hz, 1H, H_{6aglc}), 4.47 (d, ${}^{3}J_{1,2} = 7.9$ Hz, 1H, H_{1gal}), 4.12-4.00 (m, 3H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.90-3.52 (m, 17H, OC H_{2} , H_{4glc} , H_{5gal} , H_{5glc} ,), 3.30 (br s, 1H, OH), 2.15 (s, 3H, COC H_{3}), 2.12 (s, 3H, COC H_{3}), 2.06 (s, 3H, COC H_{3}), 2.04 (3s, 9H, 3×COC H_{3}), 1.96 (s, 3H, COC H_{3}).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.2, 170.0, 169.9, 169.7, 169.6, 169.0 (7×s, COCH₃), 147.7 ($C_{q \text{ triazole}}$), 122.9 ($CH_{triazole}$), 100.9 (C_{1gal}), 100.4 (C_{1glc}), 76.6 (C_{4glc}), 72.6 (C_{3glc}), 72.5 (C_{5glc}), 71.5 (C_{2glc}), 70.8 (C_{3gal}), 70.5 (C_{5gal}), 70.3, 70.3, 70.2, 70.2 (OCH₂), 69.3 (C_{2gal}), 69.0, 68.9 (OCH₂), 66.5 (C_{4gal}), 61.8 (C_{6glc}), 60.7 (C_{6gal}), 56.3 ($CH_{2}OH$), 50.0 ($CH_{2}N$), 20.8, 20.8, 20.7, 20.6, 20.6, 20.6, 20.5 (7×s, COCH₃).

m/z (ESI⁺-HRMS) for C₃₇H₅₅N₃O₂₂ = 894.3350 [*M*+H]⁺; found 894.3361 ; 916.3169 [*M*+Na]⁺; found 916.3181.



Figure S200. ¹H NMR spectrum of compound S43 (CDCI₃, 600MHz)



Figure S201. gCOSY spectrum of compound S43





Figure S203. ESI⁺-HRMS spectrum of compound S43



Acetylated compound **S43** (86.0 mg, 96.2 μ mol) was dissolved in dry MeOH (4 mL) and a solution of sodium methoxide (1M in MeOH, 150 μ L) was added. The reaction mixture was stirred at room temperature for 24 hours. The pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected reference **43** as a white solid (52.5 mg, 87.6 μ mol, **91%**).

¹**H NMR** (300 MHz, D₂O, δ ppm): 8.03 (s, 1H, $H_{triazole}$), 4.73 (s, 2H, OC H_2 C_{triazole}), 4.64 (t, J = 5.0 Hz, 2H, C H_2 N), 4.52 (d, J = 7.9 Hz, 1H, H_{1glc}), 4.46 (d, J = 7.7 Hz, 1H, H_{1gal}), 4.08-3.53 (m, 25H, OC H_2 CH₂N, OCH₂, H_{3glc} , H_{2gal} , H_{3gal} , H_{6bglc} , H_{4gal} , H_{6agal} , H_{6agal} , H_{6agal} , H_{6aglc} , H_{5glc} , H_{5glc} , H_{4glc}), 3.36 (m, 6H, H_{2glc}).

¹³C NMR (75 MHz, D₂O, δ ppm): 147.5 ($C_{triazole}$ =CH), 125.1 ($C_{triazole}$ =CH), 103.6 (C_{1gal}), 102.7 (C_{1glc}), 79.0 (C_{4glc}), 76.0 (C_{3glc}), 75.4 (C_{5glc}), 75.0 (C_{5gal}), 73.5 (C_{2glc}), 73.2 (C_{3gal}), 71.6 (C_{2gal}), 70.3, 70.2, 70.1, 70.1, 69.4, (OCH₂), 69.2 (C_{4gal}), 61.7 (C_{6glc}), 60.7 (C_{6gal}), 55.3 (OCH₂C_{triazole}), 50.7 (NCH₂).

m/z (ESI⁺-HRMS) for C₂₃H₄₁N₃O₁₅ = 600.2610 [*M*+H]⁺; found 600.2618; 622.2430 [*M*+Na]⁺; found 622.2438.







•				-
Figure S20	7. ESI ⁺ -HRMS :	spectrum of comp	ound 43	

Synthesis of monomeric reference lacking PEG chain 44



Acetylated compound **S44**⁷ (100.0 mg, 139.0 μ mol) was dissolved in dry MeOH (4 mL) and a solution of sodium methoxide (1M in MeOH, 150 μ L) was added. The reaction mixture was stirred at room temperature for 24 hours. The pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator, lyophilized to yield the fully deprotected reference **44** as a white solid (45.6 mg, 108.0 μ mol, **78%**).

¹**H NMR** (300 MHz, D₂O, δ ppm): 8.19 (s, 1H, *H*_{triazole}), 5.76 (d, *J* = 9.2 Hz, 1H, *H*_{1glc}), 4.72 (s, 2H, C*H*₂OH), 4.48 (d, *J* = 7.8 Hz, 1H, *H*_{1gal}), 4.04 (t_{app}, 1H, *H*_{2glc}), 3.95–3.64 (m,10H, *H*_{3glc}, *H*_{3gal}, *H*_{4glc}, *H*_{4gal}, *H*_{5glc}, *H*_{5gal}, *H*_{6glc}, *H*_{6gal}), 3.54 (dd, *J* = 7.7 Hz, *J* = 7.8 Hz, 2H, *H*_{2gal}).

¹³**C** NMR (75 MHz, D₂O, δ ppm): 148.2 ($C_{triazole}$), 120.2 ($CH_{triazole}$), 101.1 (C_{1gal}), 85.5 (C_{1glc}), 75.9 (C_{4glc}), 75.5 (C_{3glc}), 72.5 (C_{5glc}), 70.9 (C_{2glc}), 70.8 (C_{3gal}), 70.5 (C_{5gal}), 69.0 (C_{2gal}), 66.5 (C_{4gal}), 61.7 (C_{6gal}), 60.8 (C_{6glc}), 56.3 (CH_2OH). *m/z* (ESI⁺ HRMS) for C₁₅H₂₅N₃O₁₁ = 424.1562 [*M*+H]⁺, found: 424.1556; 446.1381 [*M*+Na]⁺, found: 446.1372.



Figure S208. ¹H NMR spectrum of compound 44 (D₂O, 600MHz)

⁷ R. Kumar, P. R. Maulik and A. K. Misra, *Glycoconj. J.* 2008, **25**, 595–602.



Figure S210. ¹³C NMR spectrum of compound 44 (D₂O, 150MHz)



Figure S211. ESI⁺-HRMS spectrum of compound 44

Negative control 45



To a solution of propargylated derivative **1** (30.0 mg, 29.4 µmol, 1.0 eq.) in a 1:1 mixture of H₂O/THF_{anh} (5 mL), were added acetylated tetra(ethylene)glycol azide **S45a** (92.4 mg, 354 µmol, 12.0 eq., *previously obtained from acetylation of hydroxylated derivative in classical conditions (Ac₂O/Pyridine/DMAP_{cat.} at 25^oC, o.n., R_{=}0.6 (3%MeOH in DCM))), CuSO₄·5H₂O (80.0 mg, 320 µmol, 10.8 eq.) and sodium ascorbate (70.0 mg, 320 µmol, 10.8 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2x10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness <i>in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 99:1 to 96:4) afforded desired acetylated multivalent compound **S45b** which was directly dissolved in dry MeOH (4 mL). A solution of sodium methoxide (1M in MeOH, 150 µL)) was added. The reaction mixture was stirred at room temperature for 18 hours. The pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator and purification by flash chromatography (SiO₂, DCM/MeOH 85:15) to afford pure reference **S45** as viscous oil (48.8 mg, 20.9 µmol, **71%** (2 steps)).

R_f = 0.12, DCM/MeOH 85:15.

¹H NMR (300 MHz, CD₃OD, δ ppm): 8.1 (s, 6H, $H_{triazole}$), 6.87-6.77 (m, 24H, C H_{ar}), 5.14 (br s, 12H, OC $H_2C_{triazole}$), 4.55 (t, J = 5.0 Hz, 12H, C $H_2N_{triazole}$), 3.85 (t, J = 5.0 Hz, 12H, C $H_2CH_2N_{triazole}$), 3.63-3.48 (m, 72H, OC H_2). ¹³C NMR (75 MHz, CD₃OD, δ ppm): 156.9 (C_a), 145.8 (C_{triazole}=CH), 144.7 (C_d), 126.3 (C_{triazole}=CH), 123.0 (C_c), 116.8

 (C_b) , 73.7 $(C_{triazole}CH_2O)$, 71.5, 71.5, 71.4, 70.3, 62.9, 62.2 (OCH_2) , 51.4 (NCH_2) . ³¹**P NMR** (122 MHz, CD₃OD, δ ppm): 10.1 (m, 3P). m/z (ESI⁺-HRMS) for C₁₀₂H₁₄₄N₂₁₀O₃₆P₃= 1166.9721 [M+2H]⁺; found 1166.9663; 1188.9540 [M+2Na]²⁺; found 1188.9547.



Figure S213. gCOSY spectrum of compound 45













Ion	Formula	Abund	Observed m/z	Calc m/z	Diff (ppm)
(M+2H)+2	C114H156N21O42P3	23800.17	1292.99974	1293.00376	3.11
(M+2Na)+2	C114H156N21O42P3	44595.1	1314.98672	1314.9857	-0.77





Figure S217. ESI⁺-HRMS spectrum of compound 45



Synthesis of hexavalent analog of 1 lacking PEG chains

To a solution of propargylated derivative **1** (13.5 mg, 13.1 µmol, 1.0 eq.) dissolved in a vial in a 1:1 mixture of H_2O/THF_{anh} (5 mL), were added lactosyl azide⁸ (72.6 mg, 110.0 µmol, 8.4 eq.), CuSO₄·5H₂O (23.5 mg, 94.1 µmol, 7.2 eq.) and sodium ascorbate (18.6 mg, 94.1 µmol, 7.2 eq.). While stirring, the mixture was first heated at 50°C for 3 hours in a 20 mL vial and at room temperature for additional 18 hours. Ethyl acetate (15 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (2×10 mL), water (10 mL) and brine (5 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 99:1 to 96:4) afforded desired acetylated multivalent compound **1a** (57.2 mg, 9.99 µmol, 76%) which was directly dissolved in dry MeOH (4 mL). A solution of sodium methoxide (1M in MeOH, 150 µL)) was added. The reaction mixture was stirred at room temperature for 18 hours. The solvent was then removed with a Pasteur pipette and dry MeOH (3 mL) is added to the residual solid. A vigorous agitation is maintained for an additional 15 min. period. After removal of the solvent with a Pasteur pipette, 3 mL of H₂O were added to the solid residue. The pH was adjusted to 7 with addition of ion-exchange resin (Amberlite IR 120 H⁺). After filtration, the solvent was removed under *vacuum* with rotary evaporator to afford pure reference **1b** as viscous colorless oil (30.5 mg, 9.30 µmol, **75%** (2 steps)).

¹**H NMR** (300 MHz, D₂O, δ ppm): 7.94 (s, 6H, *H*_{Triazole}), 6.49 (br s, 24H, *CH*_{arom}), 5.49 (d, *J* = 9.2 Hz, 6H, *H*_{1glc}), 4.48 (br s, 6H, *H*_{1gal}), 3.85-3.43 (m, 90H, *H*_{2glc}, *H*_{3glc}, *H*_{3gal}, *H*_{4glc}, *H*_{4gal}, *H*_{5glc}, *H*_{5gal}, *H*_{6glc}, *H*_{6gal}, OC*H*₂C_{q-triazole}, *H*_{2gal}). ¹³**C NMR** (75 MHz, D₂O, δ ppm): 154.5 (*C*_a), 143.5 (*C*_{Triazole}), 142.8 (*C*_d), 123.9 (*C*H_{Triazole}), 121.3 (*C*_c), 116.5 (*C*_b), 102.6 (*C*_{1gal}), 86.9 (*C*_{1glc}), 77.2 (*C*_{4glc}), 74.9 (*C*_{5glc}), 74.2 (*C*_{2glc}), 72.1 (*C*_{3gal}), 71.6 (*C*_{5gal}), 70.5 (*C*_{2gal}), 68.1 (*C*_{4gal}),

60.7 (C_{6gal}), 60.6 (OCH₂), 59.3 (C_{6glc}).

⁸ F. D. Tropper, F. O. Andersson, S. Braun and R. Roy, Synthesis 1992, 618–620.

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.3 (s, 3P).

m/z (ESI⁺-HRMS) for $C_{126}H_{168}N_{21}O_{72}P_3 = 1632.9564 [M+2Na]^{2+}$; found 1632.9574.



Figure S218. ¹H NMR spectrum of compound 1b (D₂O, 300MHz)



Figure S219. gCOSY spectrum of compound 1b



Figure S222. ESI⁺-HRMS spectrum of compound 1b

Synthetic sequence for the construction of "onion peel" glycodendrimer with 45 peripheral protected lactosides



A highly accelerated convergent approach based on the three-fold coupling of AB₁₅ hypermonomer around an extended core was investigated (Scheme 10). To this end, three tetra(ethylene)glycol chains containing one terminal propargylic function (S2)⁹ were introduced onto commercial phloroglucinol S1. NMR spectra of S3 clearly indicated desired O-alkylations with characteristic signal of CH_{ar} in ¹H NMR at δ 6.03 ppm integrating for three protons, together with two distinctive signals in ¹³C NMR at δ 160.6 and 94.4 ppm for C_{ar} O and CH_{ar} , respectively. Relative integrations corresponding to terminal propargylic signals are in addition in full agreement with those from the aromatic section, thus confirming the three-fold substitution. Application of Cu-catalyzed click reaction in the presence of dendron **30** and core S3 furnished the desired "onion peel" glycodendrimer S4 containing 45 peripheral lactoside units, in a satisfactory yield of 53%. Once again, NMR analyses clearly indicated the absence of characteristic signals from precursor **30**, notably the triplet at δ 2.43 ppm in ¹H NMR for $-CH_2C\equiv CH$ and the distinctive signal at δ 58.4 ppm in ¹³C NMR for $-CH_2C\equiv CH$. On the other hand, expected central aromatic's signals mentioned earlier clearly pointed out, confirming the integrity of the macromolecule S4 (*See SI*). Unfortunately, in deep contrast with other structures presented herein (*See the following section*), analytical efforts towards the complete characterization of compound S4 were unsuccessful, especially for MALDI-TOF MS and GPC techniques. Protocoles' optimization concerning Mass Spectrometry analyses (matrix) for S4 is currently under investigation.

⁹ X.-L. Sun, C. L. Stabler, C. S. Cazalis and E. L. Chaikof, *Bioconjugate Chem.*, 2006, **17**, 52–57.

PEGylated core S3



To a solution of phloroglucinol **S1** (12.4 mg, 98.3 µmol, 1.0 eq.) in DMF_{anh.} (2 mL), were added K₂CO₃ (dried at 200°C under vacuum) (81.5 mg, 590 µmol, 6.0 eq.) and NaI (1.47 mg, 9.83 µmol, 0.1 eq.) under a N₂ atmosphere. A solution of di-functional PEG **S2**⁹ in 0.5 mL of DMF_{anh.} was then slowly added and the resulting mixture was heated at 70°C for 22 hours. Ethyl acetate (25 mL) was added and the solution was poured into a separatory funnel containing ethyl acetate (10 mL), washed with saturated aqueous NH₄Cl (3×30 mL), water (20 mL) and brine (15 mL). Organics were collected, dried over Na₂SO₄ and concentrated to dryness *in vacuo* with rotary evaporator. Purification by flash chromatography (SiO₂, DCM/MeOH 100:0 to 99:1) afforded desired core **S3** (20.4 mg, 26.5 µmol, **27%**) as a yellowish foam.

R_f = 0.20, DCM/MeOH 97:3

¹**H NMR** (300 MHz, CDCl₃, δ ppm): 6.11 (s, 3H, C H_{ar}), 4.20 (d, J = 1.9 Hz, 6H, C H_2 C=CH), 4.07 (t, J = 4.9 Hz, 6H, C $_{ar}$ OC H_2 CH $_2$), 3.83 (t, J = 4.9 Hz, 6H, C $_{ar}$ OCH $_2$ CH $_2$), 3.70-3.67 (m, 36H, OC H_2 CH $_2$ O), 2.44 (d, J = 1.9 Hz, 3H, CH $_2$ C=CH).

¹³**C NMR** (75 MHz, CDCl₃, δ ppm): 160.5 (C_{ar} O), 94.4 (CH_{ar}), 79.7 (C=CH), 74.5 (C=CH), 70.8, 70.6, 70.4, 69.6, 69.1, 67.4 (OCH₂CH₂O), 58.4 (CH₂C=CH).

m/z (ESI⁺-HRMS) for C₃₉H₆₀O₁₅ = 769.4005 [M+H]⁺; found 769.3993 ; 791.3824 [M+Na]⁺; found 791.3813.



Figure S223. ¹H NMR spectrum of compound S3 (CDCI₃, 300MHz)



Figure S224. gCOSY spectrum of compound S3







Figure S226. ESI+HRMS spectrum of compound S3



Protected glycodendrimer with 45 peripheral lactosides S4

To a stirring solution of core **S3** (0.96 mg, 1.25 μ mol, 1.0 eq.) and hypermonomer **30** (80.2 mg, 5.27 μ mol, 4.2 eq.) in dry THF (2.5 mL) were added 2.5 mL of H₂O and a mixture of CuSO₄·5H₂O (2.84 mg, 11.3 μ mol, 9.0 eq.) and sodium ascorbate (2.23 mg, 1.13 μ mol, 9.0 eq.). After stirring for 3 hours at 50°C in a 20 mL vial, the reaction was left stirring at room temperature for 18 hours (additional CuSO₄·5H₂O (1.00 mg) and sodium ascorbate (0.80 mg) were incorporated in the mixture after 5 hours of reaction). EtOAc (25 mL) was added and the solution was washed successively with a saturated aqueous solution of NH₄Cl (3×15 mL), water (2×10 mL) and brine (10 mL). The organic phase was then dried over MgSO₄ and concentrated under reduced pressure. Column chromatography on silica (DCM/MeOH 100:0 to 93:7) afforded the desired compound **S4** (30.7 mg, 0.66 μ mol, **53%**) as a colorless oil.

R_f = 0.15, DCM/MeOH 94:6

¹**H NMR** (600 MHz, CDCl₃, δ ppm): ~8.00 (3H, N*H*) (*not visible*), 7.92-7.90 (m, 18H, $H_{\text{triazole int}}$), 7.74-7.69 (s, 45H, $H_{\text{triazole ext}}$), 7.40-7.28 (br s, 21H, N*H* + C*H*_b), 7.03-6.50 (m, 66H, C*H*_b, C*H*_c, C*H*_c), 6.07 (s, 3H, C*H*_{ar}), 5.34 (d_{app}, 45H, H_{4gal} + br s (6H, N_{triazole}C*H*₂CONH)), 5.17 (t_{app}, 45H, H_{3glc}), 5.10-5.07 (m, 75H, C_qC*H*₂O, H_{2gal}), 4.93 (dd, ³*J*_{2,3} = 3.4 Hz, ³*J*_{3,4} = 7.0 Hz, 45H, H_{3gal}), 4.84 (t_{app}, 45H, H_{2glc}), 4.55-4.46 (m, 315H, C_qC*H*₂O, OCH₂C*H*₂N, H_{1glc} , H_{6aglc} , H_{1gal}), 4.13-4.05 (m, 135H, H_{6bglc} , H_{6agal} , H_{6bgal}), 3.89-3.55 (m, 933H, NHCOC*H*₂N_{triazole}, OC*H*₂CH₂N, H_{5gal} , H_{5glc} , H_{4glc} , OC*H*₂, HNC_qC*H*₂O), 2.11-1.92 (m, 945H, COC*H*₃).

¹³C NMR (150 MHz, CDCl₃, δ ppm): 170.3, 170.3, 170.1, 170.0, 169.7, 169.6, 169.0 (7×s, COCH₃), 166.1 (CONH), 165.1 (CONH), 160.4 ($C_{ar}O$), 155.4 (C_{a}), 147.3 (C_{d}), 144.3 (C_{d}), 144.3, 144.2 ($C_{triazole}$ =CH), 143.5 ($C_{triazole}$ =CH), 134.7 ($C_{a'}$), 125.2 ($C_{triazole}$ =CH), 125.2 ($C_{triazole}$ =CH), 123.8 ($C_{triazole}$ =CH), 121.8 (C_{c}), 121.1 ($C_{b'}$), 120.9 ($C_{c'}$), 115.9, 115.3 (C_{b}), 100.9 (C_{1gal}), 100.5 (C_{1glc}), 94.2 (CH_{ar}), 76.2 (C_{4glc}), 72.6 (C_{3glc}), 72.5 (C_{5glc}), 71.5 (C_{2glc}), 70.9 (C_{3gal}), 70.5 (C_{5gal}), 70.5, 70.4, 70.3, 70.1, 69.2 (OCH₂), 68.9 (C_{2gal}), 68.9 (OCH₂), 68.5 (C_{q} CH₂O), 67.3 (CH₂O), 66.5 (C_{4gal}), 64.5 (OCH₂C=C), 62.0 (C_{q}), 61.9 ($C_{6glc} + C_{q}$ CH₂O), 60.7 (C_{6gal}), 60.3 (OCH₂), 52.5 (N_{triazole}CH₂CONH), 50.1 (NCH₂), 20.7, 20.7, 20.6, 20.5, 20.5, 20.4 (7×s, COCH₃).

³¹**P NMR** (122 MHz, CDCl₃, δ ppm): 10.2-9.8 (m, 9P).



Figure S227. ¹H NMR spectrum of compound S4 (CDCl₃, 600MHz)



Figure S229. ¹³C NMR spectrum of compound S4 (CDCl₃, 150MHz). Arrows indicated central C_{ar} O and C_{ar} H.


Figure S230. ^{31}P NMR spectrum of compound S4 (CDCl_3, 122MHz)

3. Diffusion NMR experiments

NMR diffusion experiments: NMR diffusion measurements were performed at 25°C on a Varian Inova Unity 600 spectrometer (Agilent, Santa Clara, CA, USA) operating at a frequency of 599.95 MHz for ¹H using a 5mm broadband z-gradient temperature-regulated probe. The temperature was calibrated with 1,2-ethanediol according to a standard procedure.¹⁰ The diffusion experiment employed a bipolar pulse-field gradient stimulated echo sequence as proposed by Wu et al.¹¹ Gradient pulse durations δ were set between 3 and 5 ms while diffusion times (Δ) were 30 to 150 ms to ensure that the echo intensities were attenuated by at least 80%. A complete attenuation curve was obtained by measuring 30 gradient strengths, which were linearly incremented between 1.8 and 54.2 G/cm. Hard 90° 1H pulses of 15µs were used and 36 k data points were recorded with 16 scans acquired for each gradient strength. A recycle delay of 3.0 s was used. The gradient strength was calibrated by back calculation of the coil constant from diffusion experiments on a 20% H₂O/80% D₂O standard ($D = 1.97 \times 10^{-9} \text{ m}^2 \text{ s}^{-1}$).¹²

Diffusion rates were extracted from the slope of the straight lines obtained by plotting ln(*I*) against the gradientpulse power squared according to the following equation: $\ln(I) = -D\gamma^2 G^2 \delta^2 (\Delta - \delta' 3 - \tau/2) + \ln(I_0)$ where *I* is the relative intensity of a chosen resonance ($I = I_0 \exp - [D\gamma^2 G^2 \delta^2 (\Delta - \delta' 3 - \tau/2)]$), *G*=gradient strength, γ =proton gyromagnetic ratio, *D*=diffusion rate, δ =gradient duration, Δ =diffusion delay, and τ =pulse length for bipolar pulses. All diffusion spectra were processed in MatNMR.¹³

The diffusion rates (*D*) were calculated from the decay of the signal intensity with increasing field gradient strength of the H_{4gal} proton (δ = 5.30 ppm) for the samples in CDCl₃ (acetylated conjugates) or the $H_{triazole}$ proton (δ =7.90 ppm) for the samples in D₂O (hydroxylated derivatives). In all cases, mono-exponential behavior was observed, resulting in a linear decay of the logarithm of the signal intensity as a function of the gradient strength squared. The linearity of the data was consistent with a monomolecular behavior in each case, confirming the absence of aggregation phenomena in aqueous solution.

The measurement of the diffusion rate (D) allows calculating the solvodynamic diameter of a molecule.¹⁴ The dendrimers are considered as spherical molecular objects, and characterized by an apparent diffusion coefficient D. The application of the Stokes-Einstein equation gives an estimate of the diameter of the molecule.

Stokes-Einstein equation:

$D = K_B T / 6\pi \eta r_s$

D: Diffusion rate (m².s⁻¹); K_B: Boltzmann's constant (K_B = $1.38 \times 10^{-23} \text{ m}^2 \text{ kg.s}^{-2} \text{ K}^{-1}$); T: Temperature (K) (T = 298.15 K); η : solvent viscosity en Pa s; r_s : Solvodynamic radius of the species.

¹⁰ S. Berger and S. Braun, 200 and More NMR Experiments-A Practical Course; Wiley-VCH: Weinheim, 2004; pp 145–148.

¹¹ D. H. Wu, A. D. Chen, C. S. and Johnson Jr., *J. Magn. Reson., Ser. A* 1995, **115**, 260–264.

¹² M. Holz and H. Weingärtner, *J. Magn. Reson.* 1991, **92**, 115–125.

¹³ J. D. van Beek, *J. Magn. Reson.* 2007, **187**, 19–26.

¹⁴ M. D. Diaz and Berger, S. Carbohydr. Res. 2000, **329**, 1–5.



Figure S231. a) Decay of normalized ¹H signal for the pentadecavalent dendron **31** in CDCl₃ during the PFGSTE experiment. The gradient strength is increased linearly between 1.8 and 54.2 G.cm⁻¹; b) Characteristic echo decays of the H_{4gal} resonances ($\delta = 5.30$ ppm) as a function of squared gradient strength located in dendrons **15**, **29** and **31**. Such linear behavior was also obtained for the decay of the signal intensities of other protons located either in internal regions of the conjugates, including the dendritic core and connecting branches, or in the peripheral saccharidic belt (results not shown). Linear fits, which are plotted as solid lines and used to calculate self-diffusion coefficients, are characteristic of a single molecule and therefore rule out aggregation.



Figure S232. a) Decay of normalized ¹H signal for the tricontavalent glycodendrimer **26** in CDCl₃ during the PFGSTE experiment. The gradient strength is increased linearly between 1.8 and 54.2 G.cm⁻¹; b) Characteristic echo decays of the H_{4gal} resonances ($\delta = 5.30$ ppm) as a function of squared gradient strength located in lactosylated derivatives **26**, **16**, **17**, **21**, **25**, **37**, **40**, **39**, and **36**. Such linear behavior was also obtained for the decay of the signal intensities of other protons located either in internal regions of the conjugates, including the dendritic core and connecting branches, or in the peripheral saccharidic belt (results not shown). Linear fits, which are plotted as solid lines and used to calculate self-diffusion coefficients, are characteristic of a single molecule and therefore rule out aggregation.



Figure S233. a) Decay of normalized ¹H signal for the octadecavalent hydroxylated glycodendrimer **19** in D₂O during the PFGSTE experiment. The gradient strength is increased linearly between 1.8 and 54.2 G.cm⁻¹; b) Characteristic echo decays of the $H_{triazole}$ resonances (δ = 7.90 ppm) as a function of squared gradient strength located in lactosylated derivatives **19**, **28**, **18**, **27**, **22**, **38**, and **41**. Such linear behavior was also obtained for the decay of the signal intensities of other protons located either in internal regions of the conjugates, including the dendritic core and connecting branches, or in the peripheral saccharidic belt (results not shown). Linear fits, which are plotted as solid lines and used to calculate self-diffusion coefficients, are characteristic of a single molecule and therefore rule out aggregation.

4. Competitive Surface Plasmon Resonance Studies and Sensorgrams

For LecA: The studies were conducted using a Biacore T200 SPR instrument with a CM5 sensor chip. A continuous flow of HEPES buffer (10 mm HEPES and 150 mm NaCl, 2 mM CaCl₂, pH 7.4) was maintained over the sensor surface at a flow rate of 10 µl/min. The CM5 sensor chip was activated with an injection of a solution containing Nethyl-N'-(3-diethylaminopropyl) carbodiimide (EDC) (0.2 M) and N-hydroxysuccinimide (NHS) (0.05 M) for 7 minutes. Lactoside 42 (200 µg/mL) and Et₃N (1 mM) in NaOAc buffer (pH 4.5) was injected over the activated flow cell at flow rate of 10 µl/min for 2 minute to achieve a ~230 RU immobilization. The immobilization procedure was completed by an injection of ethanolamine hydrochloride (1 M) (70 µL), followed by a flow of the buffer (100 µL/min.), in order to eliminate physically adsorbed compounds. Ethanol amine alone was used in one of the flow-cell as a reference. The solutions of pre incubated (1 h) mixtures of glycodendrimers or monomers (with the various concentrations) and a LecA lectin (1.5 µM) in running HEPES buffer are passed over flow cells of the lactoside and ethanol amine (Association: 3 min and dissociation: 3 min). The sensor chip was regenerated with the serial injections of D-lactose (0.25 M, 3 min), buffer (3 min), D-lactose (0.25 M, 3 min) and buffer (3 min). For each inhibition assay, LecA lectin (1.5 µM) without inhibitor was injected to observe the full adhesion of the lectin onto the sugar-coated surface (0% inhibition). Response units from the surface of lactoside were subtracted from the surface of ethanol amine to eliminate non-specific interactions, as well as, bulk change in RU due to variation in refractive index of the medium. The primary subtracted sensorgrams were analyzed by 1:1 Langmuir model fitting, using the BIAevaluation software. For IC₅₀ evaluation, the response units at the equilibrium was considered as the amount of lectin bound to the sugar surface in the presence of a defined concentration of inhibitor. Inhibition curves were obtained by plotting the percentage of inhibition against the inhibitor concentration (on a logarithmic scale) by using Origin 7.0 software (OriginLab Corp.) and IC_{50} values were extracted from a sigmoidal fit of the inhibition curve. The error values are obtained from the fitting of exponential curve.

For truncated hGal-3: The studies were conducted using a Biacore T200 SPR instrument with a CM5 sensor chip. A continuous flow of standard PBS buffer (HyClone®, Phosphate Buffered Saline (10X) 0.067M (PO₄), pH 7.4) was maintained over the sensor surface at a flow rate of 10 µl/min. The CM5 sensor chip was activated with an injection of a solution containing *N*-ethyl-*N'*-(3-diethylaminopropyl) carbodiimide (EDC) (0.2 M) and *N*-hydroxysuccinimide (NHS) (0.05 M) for 7 minutes. Lactoside **42** (200 µg/mL) and Et₃N (1 mM) in NaOAc buffer (pH 4.5) was injected over the activated flow cell at flow rate of 10 µl/min for 2 minute to achieve a ~230 RU immobilization. The immobilization procedure was completed by an injection of ethanolamine hydrochloride (1 M) (70 µL), followed by a flow of the buffer (100 µL/min.), in order to eliminate physically adsorbed compounds. Ethanol amine alone was used in one of the flow-cell as a reference. The solutions of pre incubated (15 min) mixtures of glycodendrimer or monomers (with the various concentrations) and a truncated galectin-3 (7.5 µM) in running PBS buffer are passed over flow cells of the lactoside and ethanol amine (Association: 3 min), buffer (3 min), D-lactose (0.25 M, 3 min) and buffer (3 min). For each inhibition assay, truncated hGal-3 (7.5 µM) without inhibitor was injected to observe the full adhesion of the lectin onto the sugar-coated surface (0% inhibition). Response units from the surface of lactoside were subtracted from the surface of

ethanol amine to eliminate non-specific interactions, as well as, bulk change in RU due to variation in refractive index of the medium. The primary subtracted sensorgrams were analyzed by 1:1 Langmuir model fitting, using the BIAevaluation software. For IC₅₀ evaluation, the response units at the equilibrium was considered as the amount of lectin bound to the sugar surface in the presence of a defined concentration of inhibitor. Inhibition curves were obtained by plotting the percentage of inhibition against the inhibitor concentration (on a logarithmic scale) by using Origin 7.0 software (OriginLab Corp.) and IC₅₀ values were extracted from a sigmoidal fit of the inhibition curve. The error values are obtained from the fitting of exponential curve.

Sensorgrams and inhibitory curves with LecA



Figure S234. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of monomeric **43** varying from 18 μ M (top curve) to 4.60 mM (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **43**.



Figure S235. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of hexavalent **18** varying from 0.306 μ M (top curve) to 40 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **18**.

– S186 –



Figure S236. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of decavalent **22** varying from 0.075 μ M (top curve) to 10 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **22**.



Figure S237. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of pentadecavalent **27** varying from 0.153 μ M (top curve) to 20 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **27**.



Figure S238. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of octadecavalent **19** varying from 0.153 μ M (top curve) to 20 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **19**.



Figure S239. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of tricontavalent **28** varying from 0.5 μ M (top curve) to 16 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **28**.



Figure S240. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of tricontavalent **38** varying from 0.306 μ M (top curve) to 20 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **38**.



Figure S241. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of nonacontavalent **41** varying from 0.125 μ M (top curve) to 8 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **41**.



Figure S242. (*left*) Sensorgrams obtained by injection of LecA (1.5 μ M) lectin incubated with different concentrations of "short" hexavalent reference (containing 6 Lactoside but without PEG chains) varying from 3.75 μ M (top curve) to 240 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for this compound.

Sensorgrams and inhibitory curves with truncated hGal-3



Figure S243. (*left*) Sensorgrams obtained by injection of truncated hGal-3 (7.5 μ M) lectin incubated with different concentrations of monomeric **43** varying from 37.5 μ M (top curve) to 1.2 mM (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **43**.



Figure S244. (*left*) Sensorgrams obtained by injection of truncated hGal-3 (7.5 μ M) lectin incubated with different concentrations of hexavalent **18** varying from 0.25 μ M (top curve) to 8 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **18**.



Figure S245. (*left*) Sensorgrams obtained by injection of truncated hGal-3 (7.5 μ M) lectin incubated with different concentrations of decavalent **22** varying from 0.062 μ M (top curve) to 2 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **22**.



Figure S246. (*left*) Sensorgrams obtained by injection of truncated hGal-3 (7.5 μ M) lectin incubated with different concentrations of pentadecavalent **27** varying from 0.1 μ M (top curve) to 3.2 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **27**.



Figure S247. (*left*) Sensorgrams obtained by injection of truncated hGal-3 (7.5 μ M) lectin incubated with different concentrations of octadecavalent **19** varying from 0.1 μ M (top curve) to 3.2 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **19**.



Figure S248. (*left*) Sensorgrams obtained by injection of truncated hGal-3 (7.5 μ M) lectin incubated with different concentrations of tricontavalent **28** varying from 0.1 μ M (top curve) to 3.2 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **28**.



Figure S249. (*left*) Sensorgrams obtained by injection of truncated hGal-3 (7.5 μ M) lectin incubated with different concentrations of tricontavalent **38** varying from 0.1 μ M (top curve) to 1.6 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **38**.



Figure S250. (*left*) Sensorgrams obtained by injection of truncated hGal-3 (7.5 μ M) lectin incubated with different concentrations of nonacontavalent **41** varying from 0.05 μ M (top curve) to 1.6 μ M (bottom curve) on the surface of immobilized lactoside **42**. (*right*) The inhibitory curve for the compound **41**.

5. X-Ray crystallographic analysis



Figure S251. ORTEP at 50% thermal ellipsoid probability of the elementary unit cell of **1**. Hydrogen atoms are omitted for clarity.

Crystal Structure Report for 1

A colorless plate-like specimen of $C_{27}H_{21}N_{1.50}O_6P_{1.50}$, approximate dimensions 0.120 mm x 0.418 mm x 0.583 mm, was used for X-ray crystallographic analysis on a Bruker APEX DUOusing Molybdenum radiation (0.71073Å wavelength) at 150K. A total of 1464 frames were collected. The total exposure time was 4.07 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 14096 reflections to a maximum θ angle of 27.67° (0.77 Å resolution), of which 5478 were independent (average redundancy 2.573, completeness = 99.3%, R_{int} = 2.64%) and 5004 (91.35%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 18.901(2) Å, <u>b</u> = 7.5595(9) Å, <u>c</u> = 17.833(2) Å, β = 111.235(2)°, volume = 2375.0(5) Å³, are based upon the refinement of the XYZ-centroids of 5832 reflections above 20 $\sigma(I)$ with 4.624° < 2 θ < 55.03°. Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.948. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.7066 and 0.7456. The structure was solved and refined using the Bruker

SHELXTL Software Package, using the space group C 1 2 1 (*C*2, #5), with Z = 4 for the formula unit, $C_{27}H_{21}N_{1.50}O_6P_{1.50}$. The final anisotropic full-matrix least-squares refinement on F² with 326 variables converged at R1 = 3.10%, for the observed data and wR2 = 6.93% for all data. The goodness-of-fit was 1.022. The largest peak in the final difference electron density synthesis was 0.213 e⁻/Å³ and the largest hole was -0.296 e⁻/Å³ with an RMS deviation of 0.041 e⁻/Å³. On the basis of the final model, the calculated density was 1.423 g/cm³ and F(000), 1056 e⁻.



 Table S1.Information on sample, data collection and structure refinement for 1.

Chemical formula	$C_{27}H_{21}N_{1.50}O_6P_{1.50}$	
Formula weight	508.91 g/mol	
Temperature	150(2) K	
Wavelength	0.71073 Å	
Crystal size	0.120 x 0.418 x 0.583 mm	
Crystal habit	colorless plate	
Crystal system	monoclinic	
Space group	C 1 2 1	
Unit cell dimensions	a = 18.901(2) Å	$\alpha = 90^{\circ}$
	b = 7.5595(9) Å	β = 111.235(2)°
	c = 17.833(2) Å	γ = 90°
Volume	2375.0(5) Å ³	
Z	4	
Density (calculated)	1.423 g/cm ³	
Absorption coefficient	0.196 mm ⁻¹	
F(000)	1056	
Theta range for data collection	1.23 to 27.67°	
Index ranges	-24<=h<=24, -9<=k<=9, -23<=l<=23	
Reflections collected	14096	
Independent reflections	5478 [R(int) = 0.0264]	
Coverage of independent reflections	99.3%	
Absorption correction	multi-scan	
Max. and min. transmission	0.7456 and 0.7066	
Structure solution technique	direct methods	
Structure solution program	SHELXS-97 (Sheldrick, 2008)	
Refinement method	Full-matrix least-squares on F ²	
Refinement program	SHELXL-2013 (Sheldrick, 2013)	
Function minimized	$\Sigma w(F_o^2 - F_c^2)^2$	
Data / restraints / parameters	5478 / 1 / 326	
Goodness-of-fit on F ²	1.022	
$\Delta \sigma_{max}$	0.001	
Final R indices	5004 data; I>2σ(I)	R1 = 0.0310, wR2 = 0.0667
	all data	R1 = 0.0366, wR2 = 0.0693
Weighting scheme	$w=1/[\sigma^{2}(F_{o})+(0.0329P)^{2}+0.6412P]$	
Absolute structure parameter	0.1(0)	
Largest diff. peak and hole	0.213 and -0.296 eA ⁻³	
R.M.S. deviation from mean	0.041 eA ~	

	x/a	y/b	z/c	U(eq)
P1	0.5	0.44796(10)	0.5	0.01529(18)
P2	0.56920(3)	0.12904(8)	0.56291(3)	0.01511(13)
O1	0.43407(11)	0.5366(3)	0.84159(11)	0.0392(5)
O2	0.47457(9)	0.5908(2)	0.55014(9)	0.0208(4)
O3	0.57376(9)	0.0657(2)	0.64912(9)	0.0189(4)
O4	0.73128(10)	0.3698(3)	0.94575(10)	0.0307(4)
O5	0.64702(9)	0.0459(2)	0.56383(9)	0.0185(3)
O6	0.73530(9)	0.0805(2)	0.30021(10)	0.0250(4)
N1	0.57058(11)	0.3380(2)	0.55677(12)	0.0173(4)
N2	0.5	0.0275(4)	0.5	0.0176(6)
C1	0.4482(2)	0.0901(7)	0.8859(2)	0.0698(13)
C2	0.41918(16)	0.2305(5)	0.87093(18)	0.0413(7)
C3	0.38492(17)	0.4062(4)	0.85576(17)	0.0353(7)
C4	0.44249(14)	0.5345(4)	0.76777(15)	0.0241(5)
C5	0.49528(13)	0.6538(3)	0.75968(14)	0.0230(5)
C6	0.50670(13)	0.6666(3)	0.68761(14)	0.0196(5)
C7	0.46537(13)	0.5594(3)	0.62387(13)	0.0173(5)
C8	0.61731(12)	0.1472(3)	0.72243(13)	0.0185(5)
C9	0.58092(13)	0.1761(3)	0.77627(14)	0.0232(5)
C10	0.62149(14)	0.2494(4)	0.85069(15)	0.0275(6)
C11	0.69723(15)	0.2937(3)	0.87079(14)	0.0228(5)
C12	0.80545(16)	0.4430(4)	0.96475(16)	0.0323(6)
C13	0.86563(16)	0.3102(4)	0.99559(15)	0.0304(6)
C14	0.91374(17)	0.2044(5)	0.02319(16)	0.0406(7)
C15	0.66949(13)	0.0541(3)	0.49653(14)	0.0173(5)
C16	0.62500(13)	0.9803(3)	0.42358(14)	0.0195(5)
C17	0.65023(13)	0.9886(3)	0.35964(15)	0.0209(5)
C18	0.71914(13)	0.0683(3)	0.36929(14)	0.0199(5)
C19	0.80514(14)	0.1635(4)	0.30645(15)	0.0263(6)
C20	0.80208(14)	0.2071(3)	0.22548(15)	0.0258(6)
C21	0.80047(16)	0.2484(4)	0.16158(17)	0.0362(7)
C22	0.76514(12)	0.1327(4)	0.44404(13)	0.0218(5)
C23	0.73919(12)	0.1275(4)	0.50782(14)	0.0204(5)
C24	0.73338(14)	0.2623(3)	0.81697(14)	0.0241(5)
C25	0.69281(14)	0.1885(3)	0.74190(14)	0.0231(5)
C26	0.40214(14)	0.4266(3)	0.70432(14)	0.0237(5)
C27	0.41313(14)	0.4403(3)	0.63109(14)	0.0225(5)

Table S2. Atomic coordinates and equivalent isotropic atomic displacement parameters ($Å^2$) for **1**. U(eq) is defined as one third of the trace of the orthogonalized U_{ii} tensor.

Table S3. Bond lengths (Å) for 1.				
P1-O2	1.5839(16)	P1-O2	1.5839(16)	
P1-N1	1.587(2)	P1-N1	1.587(2)	
P2-N2	1.5799(14)	P2-O3	1.5825(16)	
P2-N1	1.584(2)	P2-O5	1.5943(16)	
O1-C4	1.382(3)	O1-C3	1.438(3)	
O2-C7	1.407(3)	O3-C8	1.410(3)	
O4-C11	1.382(3)	O4-C12	1.429(3)	
O5-C15	1.412(3)	O6-C18	1.375(3)	
O6-C19	1.429(3)	N2-P2	1.5798(14)	
C1-C2	1.180(5)	C1-H1	0.95	
C2-C3	1.459(5)	C3-H19	0.99	
C3-H18	0.99	C4-C26	1.379(3)	
C4-C5	1.391(3)	C5-C6	1.382(3)	
C5-H3	0.95	C6-C7	1.384(3)	
C6-H4	0.95	C7-C27	1.376(3)	
C8-C25	1.377(3)	C8-C9	1.386(3)	
C9-C10	1.385(3)	C9-H12	0.95	
C10-C11	1.385(4)	C10-H17	0.95	
C11-C24	1.385(3)	C12-C13	1.466(4)	
C12-H15	0.99	C12-H16	0.99	
C13-C14	1.177(4)	C14-H2	0.95	
C15-C23	1.375(3)	C15-C16	1.385(3)	
C16-C17	1.387(3)	C16-H5	0.95	
C17-C18	1.388(3)	C17-H6	0.95	
C18-C22	1.389(3)	C19-C20	1.462(3)	
C19-H9	0.99	C19-H8	0.99	
C20-C21	1.171(4)	C21-H7	0.95	
C22-C23	1.393(3)	C22-H11	0.95	
C23-H10	0.95	C24-C25	1.396(3)	
C24-H13	0.95	C25-H14	0.95	
C26-C27	1.398(3)	C26-H20	0.95	
C27-H21	0.95			

Table S4. Bond angles (°) for 1.

O2-P1-O2	94.00(12)	O2-P1-N1	111.14(9)
O2-P1-N1	110.74(9)	O2-P1-N1	110.74(9)
O2-P1-N1	111.14(9)	N1-P1-N1	116.79(15)
N2-P2-O3	106.41(8)	N2-P2-N1	117.75(12)
O3-P2-N1	111.87(10)	N2-P2-O5	109.82(9)
O3-P2-O5	98.35(9)	N1-P2-O5	110.84(10)
C4-O1-C3	117.7(2)	C7-O2-P1	125.28(14)
C8-O3-P2	125.21(14)	C11-O4-C12	117.6(2)
C15-O5-P2	122.29(14)	C18-O6-C19	117.27(18)
P2-N1-P1	122.38(13)	P2-N2-P2	121.84(18)
C2-C1-H1	180.0	C1-C2-C3	177.7(4)
O1-C3-C2	112.6(2)	O1-C3-H19	109.1
C2-C3-H19	109.1	O1-C3-H18	109.1
C2-C3-H18	109.1	H19-C3-H18	107.8
C26-C4-O1	124.6(2)	C26-C4-C5	120.2(2)
O1-C4-C5	115.2(2)	C6-C5-C4	120.1(2)
C6-C5-H3	120.0	C4-C5-H3	120.0
C5-C6-C7	119.4(2)	C5-C6-H4	120.3
C7-C6-H4	120.3	C27-C7-C6	121.1(2)
C27-C7-O2	122.2(2)	C6-C7-O2	116.4(2)
C25-C8-C9	121.2(2)	C25-C8-O3	122.5(2)
C9-C8-O3	116.2(2)	C10-C9-C8	119.0(2)
C10-C9-H12	120.5	C8-C9-H12	120.5
C11-C10-C9	120.4(2)	C11-C10-H17	119.8
C9-C10-H17	119.8	O4-C11-C10	115.3(2)
O4-C11-C24	124.6(2)	C10-C11-C24	120.1(2)
O4-C12-C13	112.7(2)	O4-C12-H15	109.1
C13-C12-H15	109.1	O4-C12-H16	109.1
C13-C12-H16	109.1	H15-C12-H16	107.8
C14-C13-C12	177.4(3)	C13-C14-H2	180.0
C23-C15-C16	121.6(2)	C23-C15-O5	117.4(2)
C16-C15-O5	120.9(2)	C15-C16-C17	118.8(2)
C15-C16-H5	120.6	C17-C16-H5	120.6
C16-C17-C18	120.3(2)	C16-C17-H6	119.9
C18-C17-H6	119.9	O6-C18-C17	114.9(2)
O6-C18-C22	124.8(2)	C17-C18-C22	120.3(2)
O6-C19-C20	108.5(2)	O6-C19-H9	110.0
C20-C19-H9	110.0	O6-C19-H8	110.0
C20-C19-H8	110.0	H9-C19-H8	108.4
C21-C20-C19	177.5(3)	C20-C21-H7	180.0
C18-C22-C23	119.4(2)	C18-C22-H11	120.3
C23-C22-H11	120.3	C15-C23-C22	119.6(2)
C15-C23-H10	120.2	C22-C23-H10	120.2
C11-C24-C25	119.7(2)	C11-C24-H13	120.1
C25-C24-H13	120.1	C8-C25-C24	119.5(2)
C8-C25-H14	120.3	C24-C25-H14	120.3
C4-C26-C27	119.8(2)	C4-C26-H20	120.1
C27-C26-H20	120.1	C7-C27-C26	119.3(2)
			/-/

C7-C27-H21

120.3

Table S5.Torsion angles (9 for 1.

02-P1-02-C7	-160.5(2)	N1-P1-O2-C7	-46.1(2)
N1-P1-O2-C7	85.43(19)	N2-P2-O3-C8	-166.90(17)
N1-P2-O3-C8	-37.0(2)	O5-P2-O3-C8	79.50(18)
N2-P2-O5-C15	64.96(18)	O3-P2-O5-C15	175.84(17)
N1-P2-O5-C15	-66.86(19)	N2-P2-N1-P1	9.92(18)
O3-P2-N1-P1	-113.81(14)	O5-P2-N1-P1	137.52(13)
O2-P1-N1-P2	-133.46(13)	O2-P1-N1-P2	123.45(14)
N1-P1-N1-P2	-5.10(9)	O3-P2-N2-P2	121.66(7)
N1-P2-N2-P2	-4.76(9)	O5-P2-N2-P2	-132.85(7)
C4-O1-C3-C2	75.2(3)	C3-O1-C4-C26	5.9(4)
C3-O1-C4-C5	-175.0(2)	C26-C4-C5-C6	1.0(4)
O1-C4-C5-C6	-178.2(2)	C4-C5-C6-C7	-0.2(4)
C5-C6-C7-C27	0.0(3)	C5-C6-C7-O2	174.2(2)
P1-O2-C7-C27	-61.8(3)	P1-O2-C7-C6	124.1(2)
P2-O3-C8-C25	-49.7(3)	P2-O3-C8-C9	133.50(19)
C25-C8-C9-C10	0.6(4)	O3-C8-C9-C10	177.5(2)
C8-C9-C10-C11	0.2(4)	C12-O4-C11-C10	-170.6(2)
C12-O4-C11-C24	8.9(4)	C9-C10-C11-O4	178.5(2)
C9-C10-C11-C24	-1.1(4)	C11-O4-C12-C13	-84.8(3)
P2-O5-C15-C23	124.8(2)	P2-O5-C15-C16	-59.4(3)
C23-C15-C16-C17	-3.1(3)	O5-C15-C16-C17	-178.7(2)
C15-C16-C17-C18	0.5(3)	C19-O6-C18-C17	179.2(2)
C19-O6-C18-C22	0.9(3)	C16-C17-C18-O6	-175.2(2)
C16-C17-C18-C22	3.2(4)	C18-O6-C19-C20	-164.5(2)
O6-C18-C22-C23	173.9(2)	C17-C18-C22-C23	-4.3(4)
C16-C15-C23-C22	1.9(4)	O5-C15-C23-C22	177.6(2)
C18-C22-C23-C15	1.9(4)	O4-C11-C24-C25	-178.4(2)
C10-C11-C24-C25	1.1(4)	C9-C8-C25-C24	-0.6(4)
O3-C8-C25-C24	-177.3(2)	C11-C24-C25-C8	-0.3(4)
O1-C4-C26-C27	177.5(2)	C5-C4-C26-C27	-1.5(4)
C6-C7-C27-C26	-0.5(4)	O2-C7-C27-C26	-174.4(2)
C4-C26-C27-C7	1.2(4)		

The anisotropic atomic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U_{11} + + 2 h k a^* b^* U_{12}]$						
	U 11	U ₂₂	U ₃₃	U ₂₃	U ₁₃	U ₁₂
P1	0.0195(4)	0.0119(4)	0.0153(4)	0	0.0074(3)	0
P2	0.0151(3)	0.0138(3)	0.0161(3)	0.0011(2)	0.0052(2)	0.0002(2)
O1	0.0473(12)	0.0541(14)	0.0242(10)	-0.0145(9)	0.0224(9)	-0.0265(11)
O2	0.0321(9)	0.0151(9)	0.0181(8)	0.0014(6)	0.0125(7)	0.0047(7)
O3	0.0202(8)	0.0186(8)	0.0176(8)	0.0012(7)	0.0064(7)	-0.0032(7)
O4	0.0303(10)	0.0401(11)	0.0205(9)	-0.0051(9)	0.0076(8)	-0.0025(9)
O5	0.0157(8)	0.0205(9)	0.0195(8)	0.0015(7)	0.0065(6)	0.0026(7)
O6	0.0216(8)	0.0328(11)	0.0236(9)	-0.0052(7)	0.0118(7)	-0.0064(7)
N1	0.0175(10)	0.0168(10)	0.0173(10)	0.0003(8)	0.0059(8)	-0.0019(8)
N2	0.0163(14)	0.0144(14)	0.0203(15)	0	0.0043(11)	0
C1	0.056(2)	0.090(3)	0.077(3)	0.046(3)	0.040(2)	0.025(2)
C2	0.0310(15)	0.063(2)	0.0332(15)	0.0135(17)	0.0159(12)	-0.0039(17)
C3	0.0364(15)	0.0485(19)	0.0286(15)	-0.0052(13)	0.0210(13)	-0.0161(14)
C4	0.0236(13)	0.0301(14)	0.0202(12)	-0.0033(11)	0.0098(10)	-0.0053(11)
C5	0.0219(12)	0.0262(14)	0.0191(11)	-0.0057(10)	0.0053(9)	-0.0062(10)
C6	0.0176(11)	0.0174(12)	0.0232(12)	0.0000(9)	0.0066(9)	-0.0010(9)
C7	0.0225(12)	0.0157(11)	0.0141(11)	0.0019(9)	0.0072(9)	0.0059(10)
C8	0.0210(11)	0.0160(12)	0.0163(11)	0.0031(9)	0.0041(9)	0.0010(10)
C9	0.0176(11)	0.0300(14)	0.0219(12)	0.0042(10)	0.0071(9)	0.0040(10)
C10	0.0264(13)	0.0362(15)	0.0220(12)	0.0007(12)	0.0112(10)	0.0064(12)
C11	0.0276(13)	0.0225(13)	0.0161(12)	0.0016(9)	0.0052(10)	0.0015(10)
C12	0.0362(15)	0.0326(15)	0.0221(13)	-0.0056(12)	0.0033(11)	-0.0064(13)
C13	0.0323(15)	0.0400(16)	0.0168(13)	-0.0047(12)	0.0063(12)	-0.0070(13)
C14	0.0386(16)	0.054(2)	0.0251(14)	-0.0030(14)	0.0068(12)	0.0074(16)
C15	0.0175(11)	0.0155(11)	0.0204(12)	0.0026(9)	0.0086(9)	0.0050(9)
C16	0.0182(12)	0.0161(12)	0.0245(13)	-0.0015(10)	0.0081(10)	-0.0006(9)
C17	0.0187(12)	0.0200(12)	0.0221(12)	-0.0039(10)	0.0052(10)	-0.0004(9)
C18	0.0212(12)	0.0174(12)	0.0230(12)	-0.0006(9)	0.0104(10)	0.0031(9)
C19	0.0218(12)	0.0298(15)	0.0277(13)	0.0001(11)	0.0096(10)	-0.0054(10)
C20	0.0236(13)	0.0239(14)	0.0323(14)	-0.0018(11)	0.0131(11)	-0.0016(11)
C21	0.0400(16)	0.0404(17)	0.0323(15)	0.0003(14)	0.0180(13)	-0.0071(14)
C22	0.0160(11)	0.0218(12)	0.0276(12)	-0.0013(11)	0.0080(9)	-0.0015(11)
C23	0.0175(11)	0.0210(12)	0.0197(11)	-0.0017(11)	0.0033(9)	-0.0001(11)
C24	0.0207(12)	0.0295(14)	0.0219(12)	0.0010(11)	0.0075(10)	-0.0059(11)
C25	0.0242(12)	0.0295(14)	0.0179(11)	0.0004(10)	0.0105(10)	-0.0016(10)
C26	0.0244(12)	0.0242(13)	0.0244(13)	-0.0050(10)	0.0111(10)	-0.0092(11)
C27	0.0261(13)	0.0197(12)	0.0200(12)	-0.0045(10)	0.0063(10)	-0.0011(11)

Table S6. Anisotropic atomic displacement parameters (Å $^2)$ for 1.

	, , , , , , , , , , , , , , , , , , , ,				
	x/a	y/b	z/c	U(eq)	
H1	0.4715	-0.0230	0.8979	0.084	
H19	0.3371	0.4007	0.8083	0.042	
H18	0.3721	0.4430	0.9026	0.042	
H3	0.5235	0.7266	0.8038	0.028	
H4	0.5426	0.7482	0.6819	0.024	
H12	0.5289	0.1460	0.7624	0.028	
H17	0.5972	0.2694	0.8882	0.033	
H15	0.8134	0.5372	1.0055	0.039	
H16	0.8090	0.4978	0.9158	0.039	
H2	0.9526	0.1190	1.0455	0.049	
H5	0.5781	-0.0750	0.4174	0.023	
H6	0.6202	-0.0605	0.3091	0.025	
H9	0.8482	0.0825	0.3329	0.032	
H8	0.8127	0.2724	0.3392	0.032	
H7	0.7992	0.2820	0.1098	0.043	
H11	0.8138	0.1799	0.4516	0.026	
H10	0.7695	0.1743	0.5588	0.024	
H13	0.7856	0.2908	0.8311	0.029	
H14	0.7171	0.1669	0.7045	0.028	
H20	0.3670	0.3433	0.7103	0.028	
H21	0.3848	0.3681	0.5868	0.027	

Table S7. Hydrogen atomic coordinates and isotropic atomic displacement parameters (Å²) for 1.