

Supporting Information

Synthesis and biological application of BKT-140 peptide modified polymer micelles for treating tumor metastasis with an enhanced cell internalization efficiency

*Xuan Liu^a, Bolin Cheng^a, Tingting Meng^a, Jian You^a, Yun Zhu^a, Binbin Lu^a, Hong Yuan^a, Xuan Huang^b, Fuqiang Hu^{a,*1}*

^aCollege of Pharmaceutical Science, Zhejiang University, 866 Yuhangtang Road, Hangzhou 310058, People's Republic of China

^bDepartment of Pharmacy, School of Medicine Science, Jiaxing University, Zhejiang 314001, People's Republic of China.

¹ *Corresponding author: Fuqiang Hu, PhD, professor

Address: College of Pharmaceutical Science, Zhejiang University, 866 Yuhangtang Road, Hangzhou 310058, P.R.China

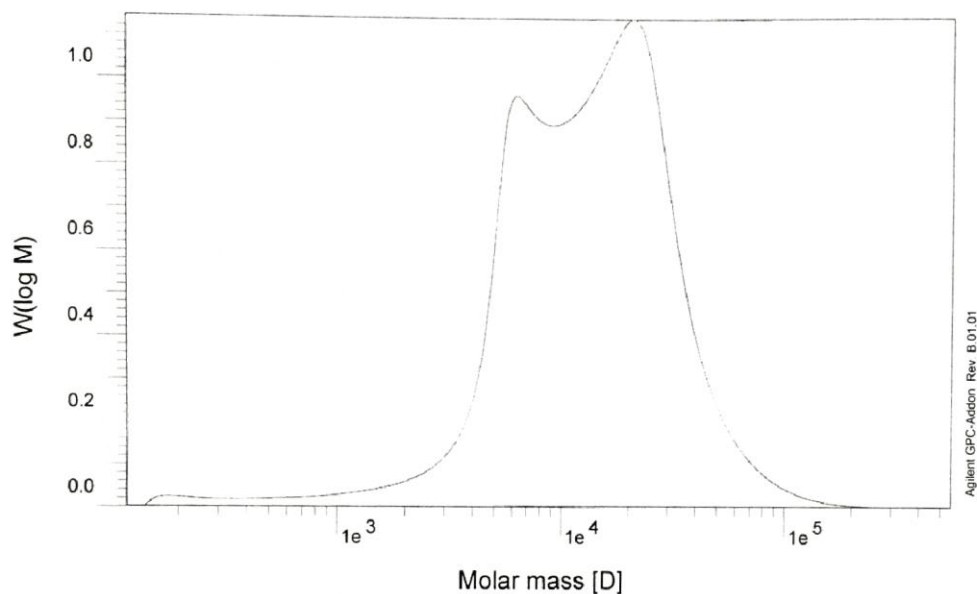
Email: hufq@zju.edu.cn

Tel: +86-571-88208441

Fax: +86-571-88208439

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 Eluent :
 Concentration : 1.000 g/l
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 Operator : CBL

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 Acquisition interval : 0.430 sec



rid1A

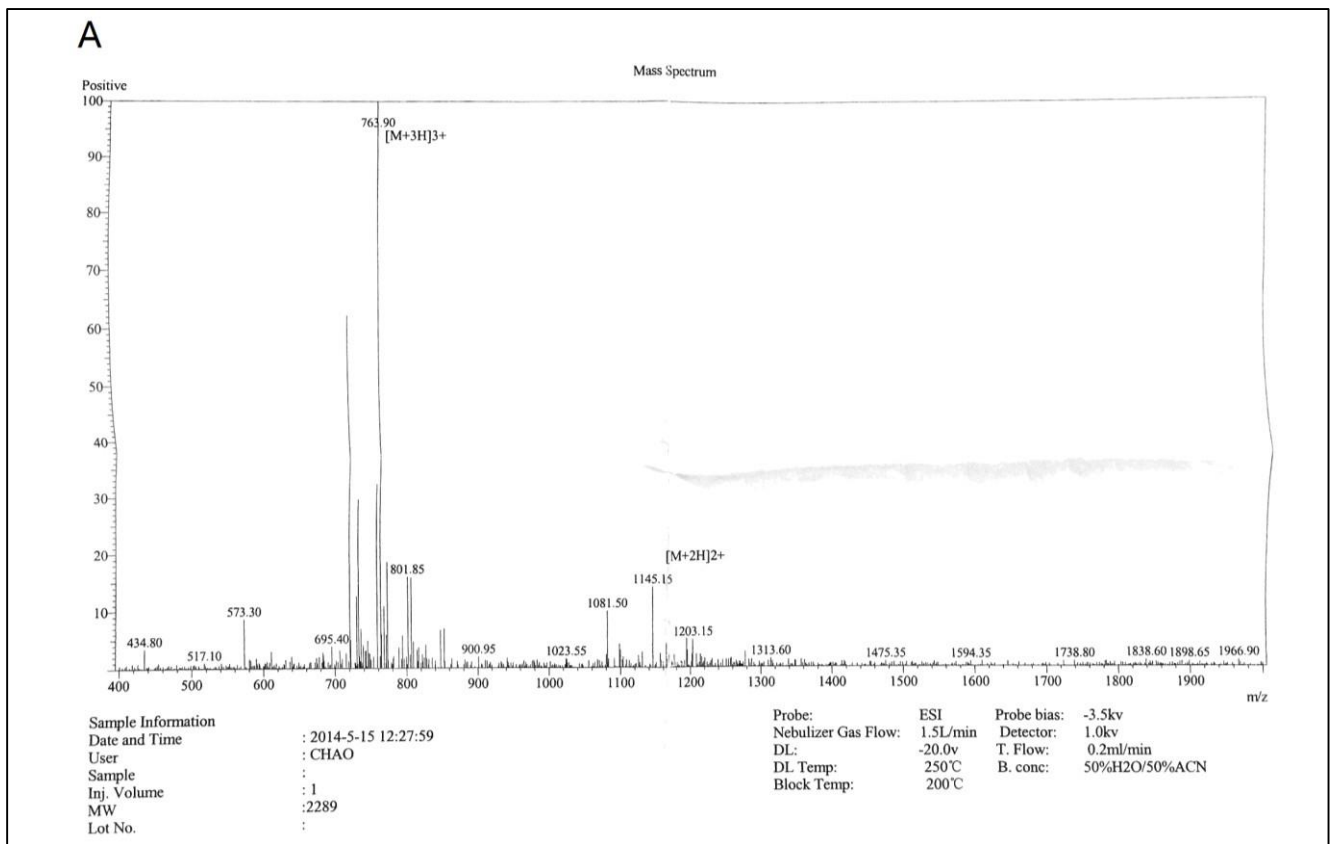
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Figure S1. Report of Molecular weight (Mw) measurement of Chitosan Oligosaccharide by GPC. The chitosan (Mw 18.1kDa) used in this research was obtained by enzymatic degradation. The reaction time of hydrolysis was controlled by molecular weight measurement. The final reaction mixture was then filtered by

filter with 0.45 μm pore size, and then ultra-filtered by various molecular weight cut off ultrafiltration membrane (Millipore Labscale TFF system, Millipore Co., USA). The CSO was obtained by lyophilization. The molecular weight of final CSO was determined by gel permeation chromatography (GPC) with TSK-gel column (G3000SW, 7.5mm ID \times 30 cm) at 25 $^{\circ}\text{C}$. The mobile phase is acetate buffer solution (pH 6.0) with a flow rate of 0.8ml/min. Master samples of polysaccharide with different molecular weight ($M_w = 5.9, 11.8, 22.8, 47.3, 112, 212$ kDa) were dissolved in acetate buffer solution (pH 6.0), and their final concentration was 1mg/ml. Calibration was performed by means of polysaccharide samples using the integral molecular weight distribution method. Figure S1 was the scan file of the original report.



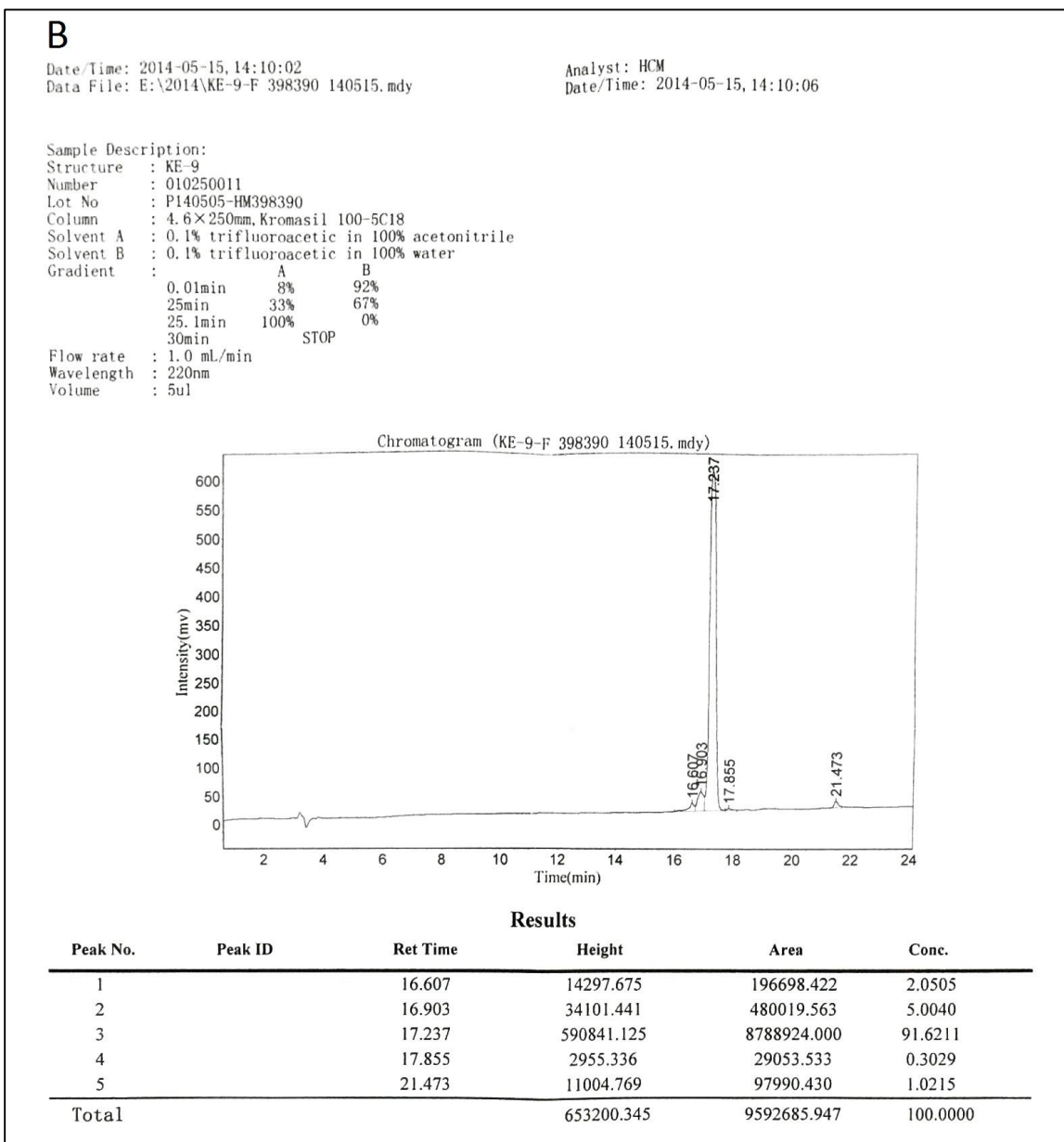
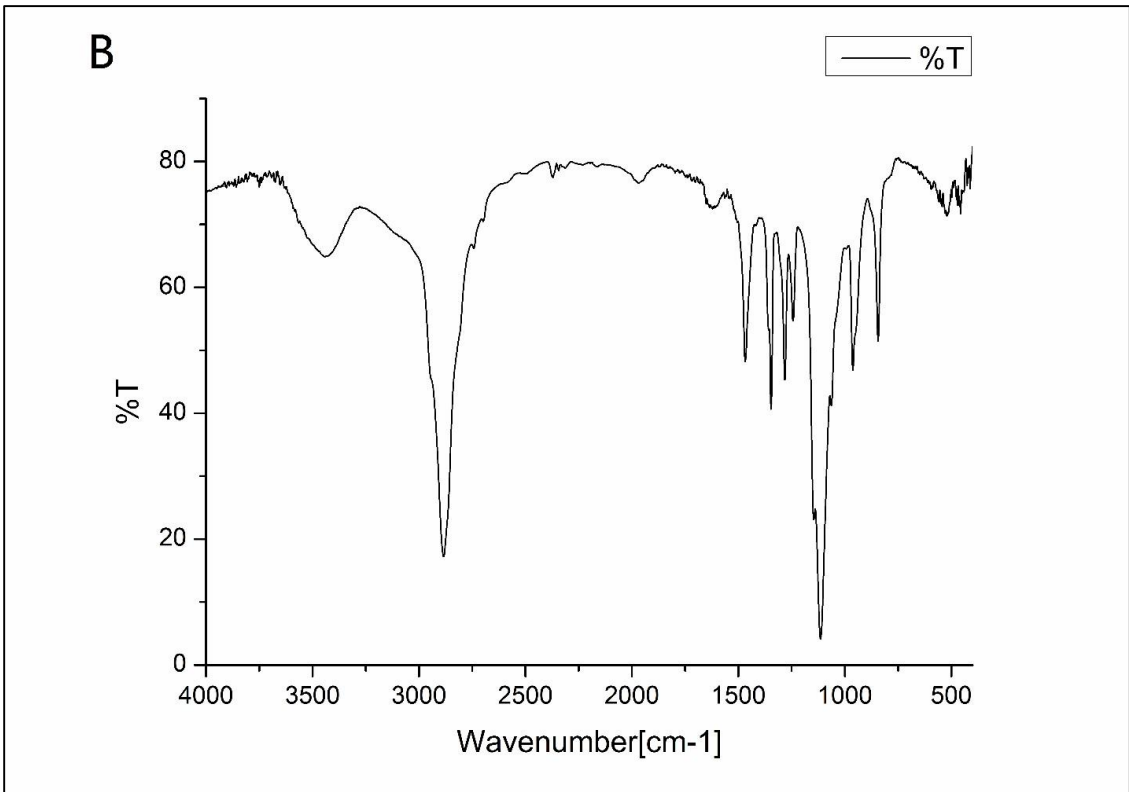
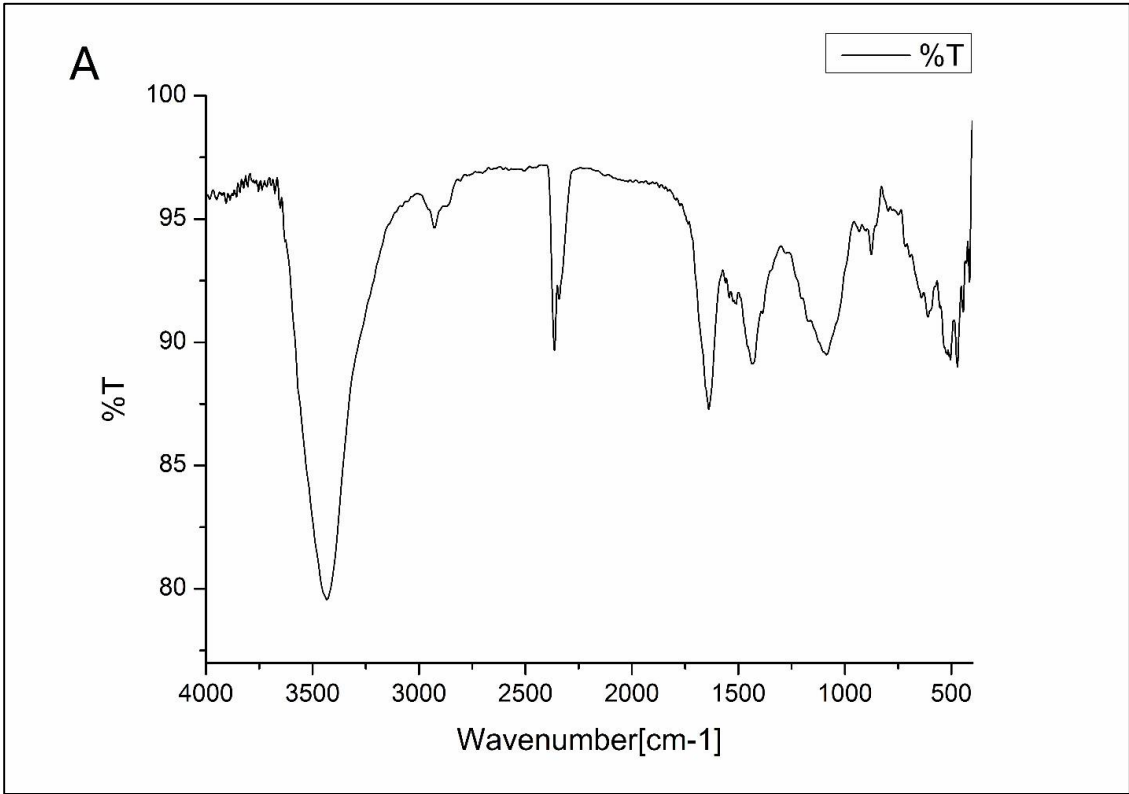


Figure S2. (A)Molecular weight measurement by mass spectrum and (B) Purity Detection by HPLC of glu modified BKT-140. BKT-140 peptide (4F-benzoyl-Arg-Arg-Nal-Cys-Tyr-Cit-Lys-D-Lys-Pro-Tyr-Arg-Cit-Cys-Arg-NH₂) was synthesized by Hangzhou Dgpeptides Co., Ltd, China. In order not to interfere the function, we need to conjugate micelles and the peptide via a site distant from the pharmacophore. Therefore, CSOSA was conjugated with BKT-140 via the Glu added on D-Lys⁸. The molecular weight was measured by mass spectrum. Both Figure S2 (A) & (B) were the scan files of the original reports.



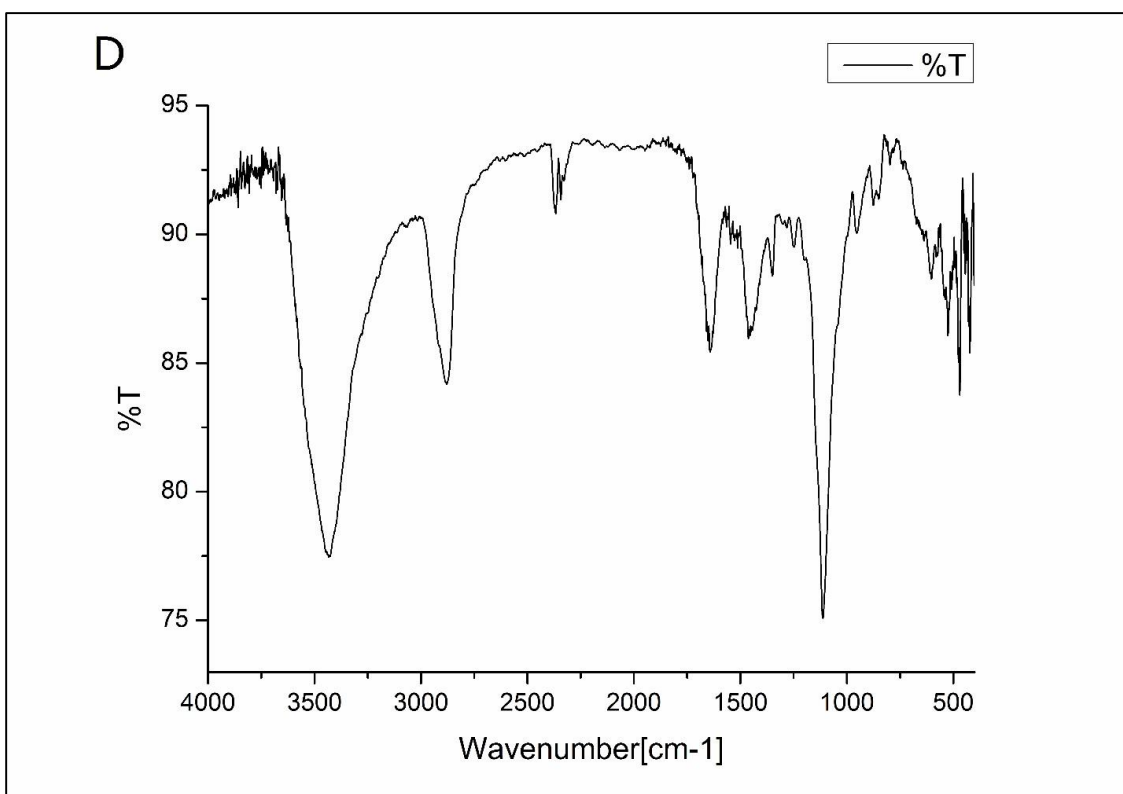
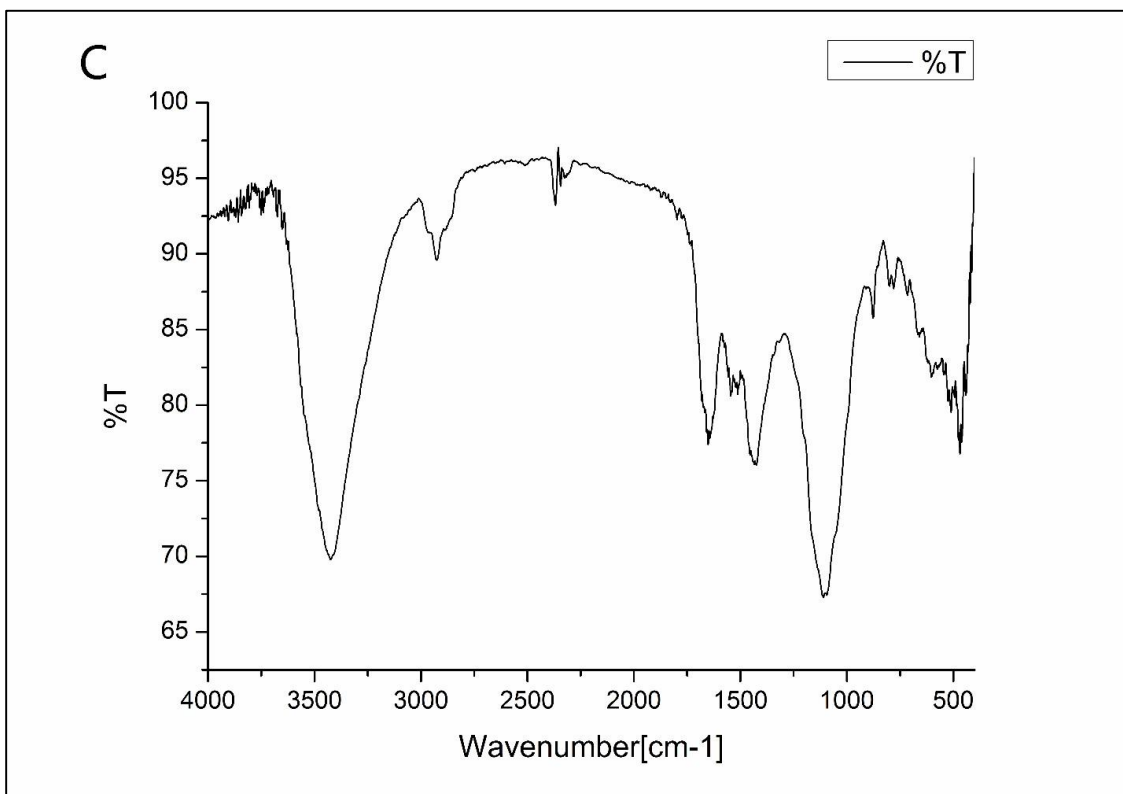
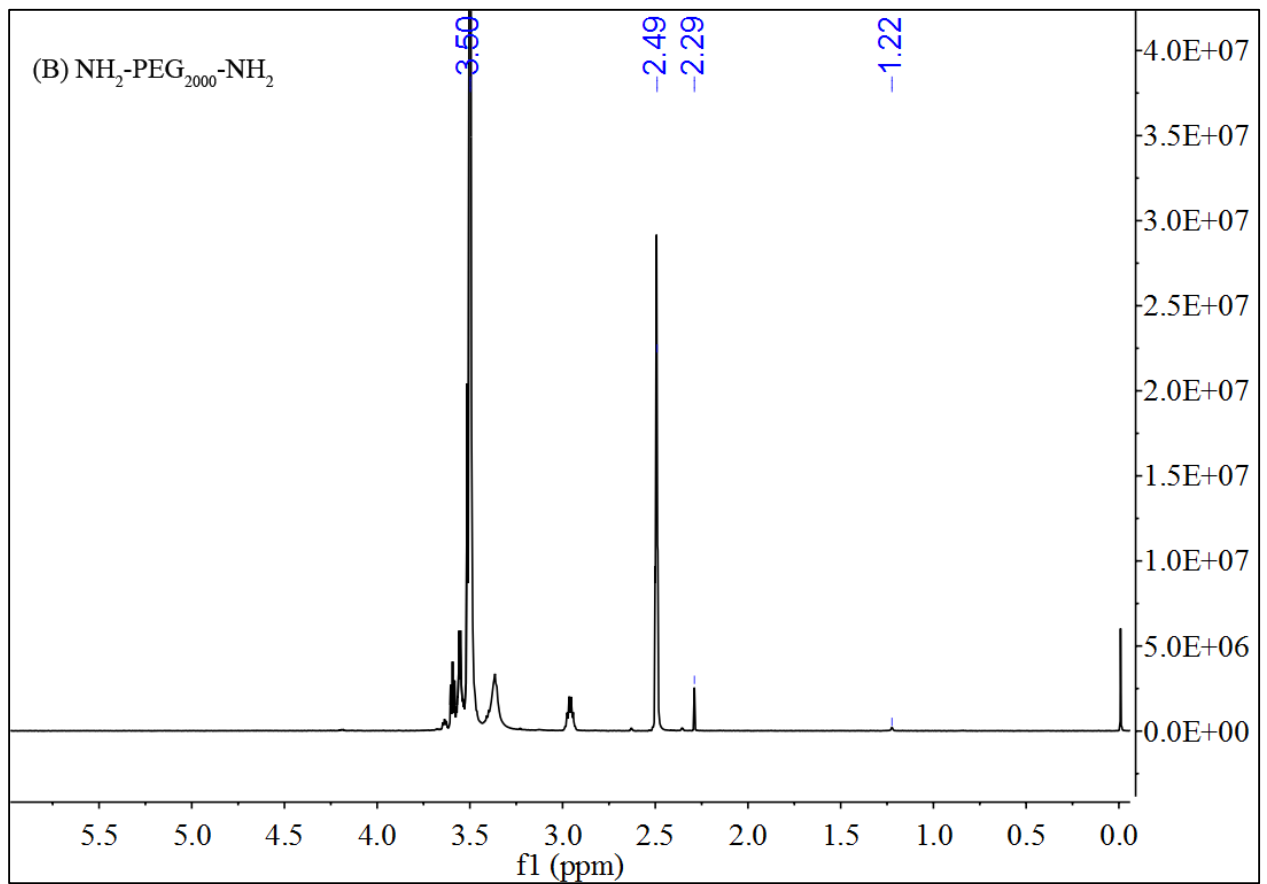
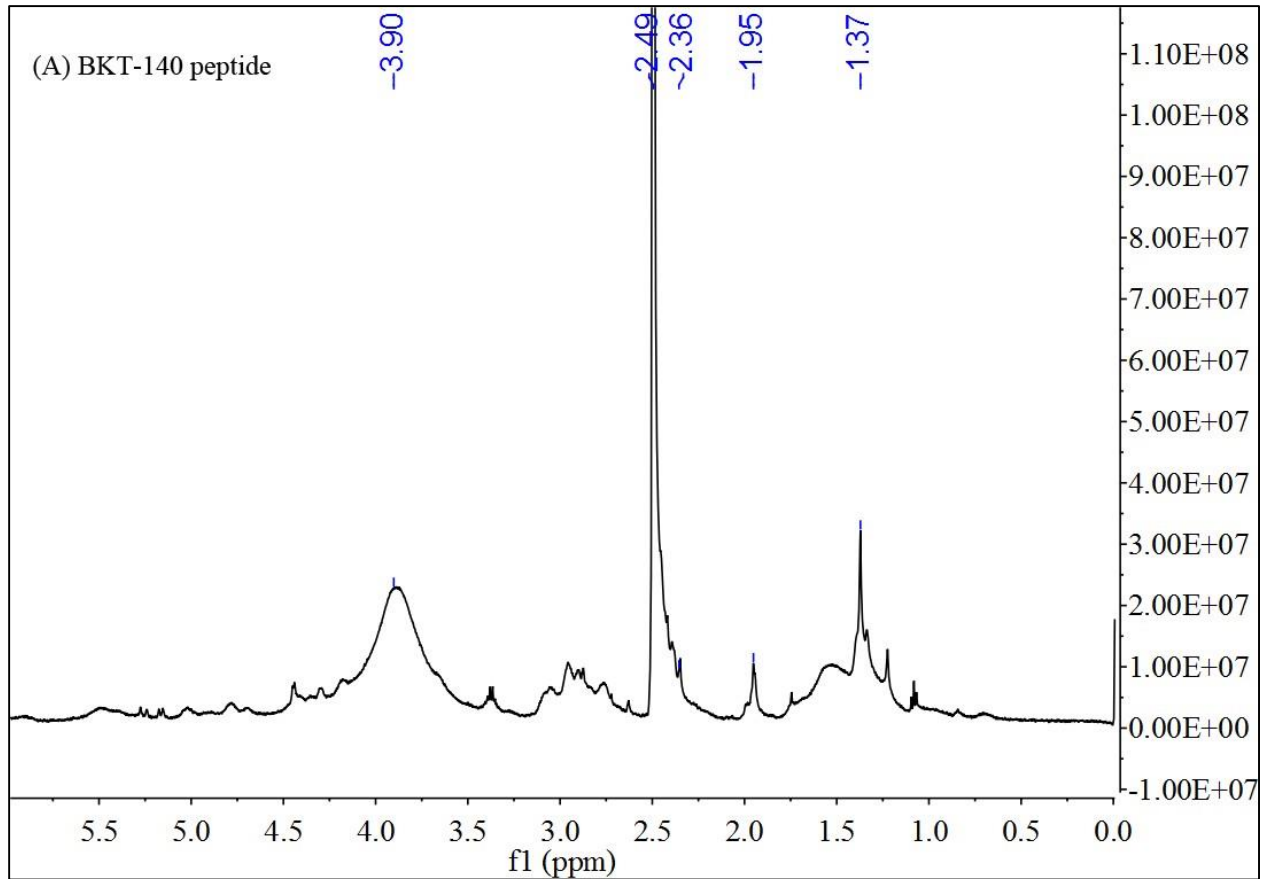
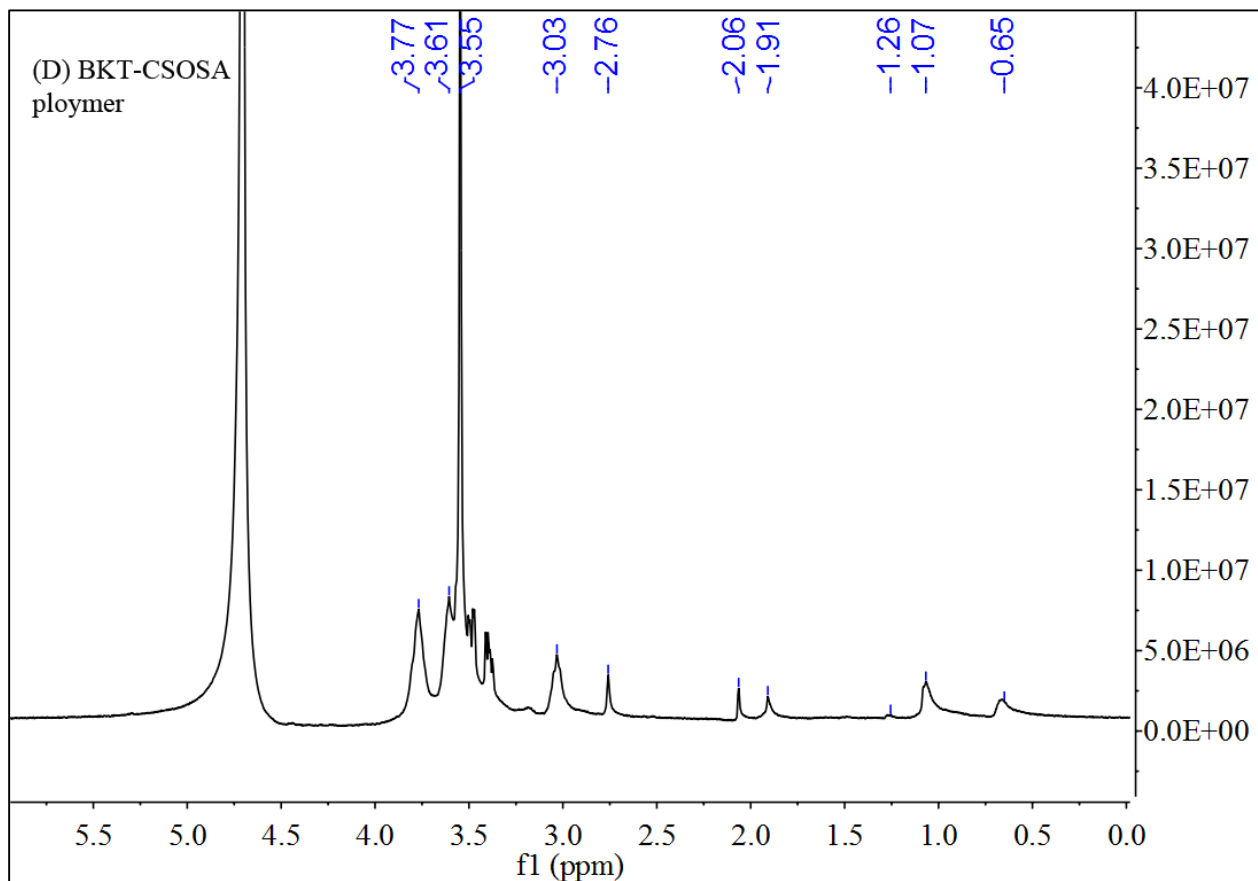
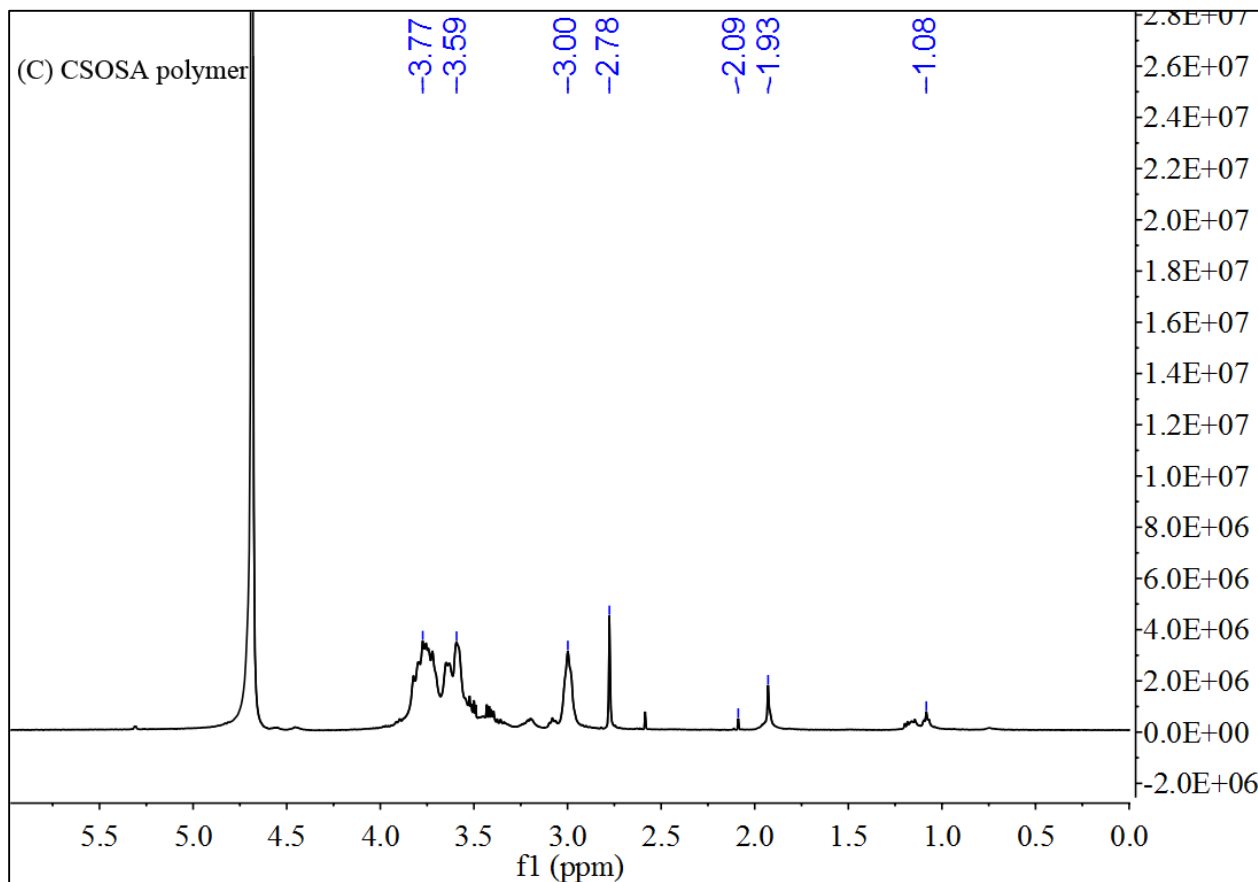


Figure S3. IR data of (A) BKT-140 peptide, (B) $\text{NH}_2\text{-PEG}_{2000}\text{-NH}_2$, (C) BKT-140 conjugated PEG_{2000} (BKT- $\text{PEG}_{2000}\text{-NH}_2$) and (D) BKT-140 peptide mixed with $\text{NH}_2\text{-PEG}_{2000}\text{-NH}_2$.





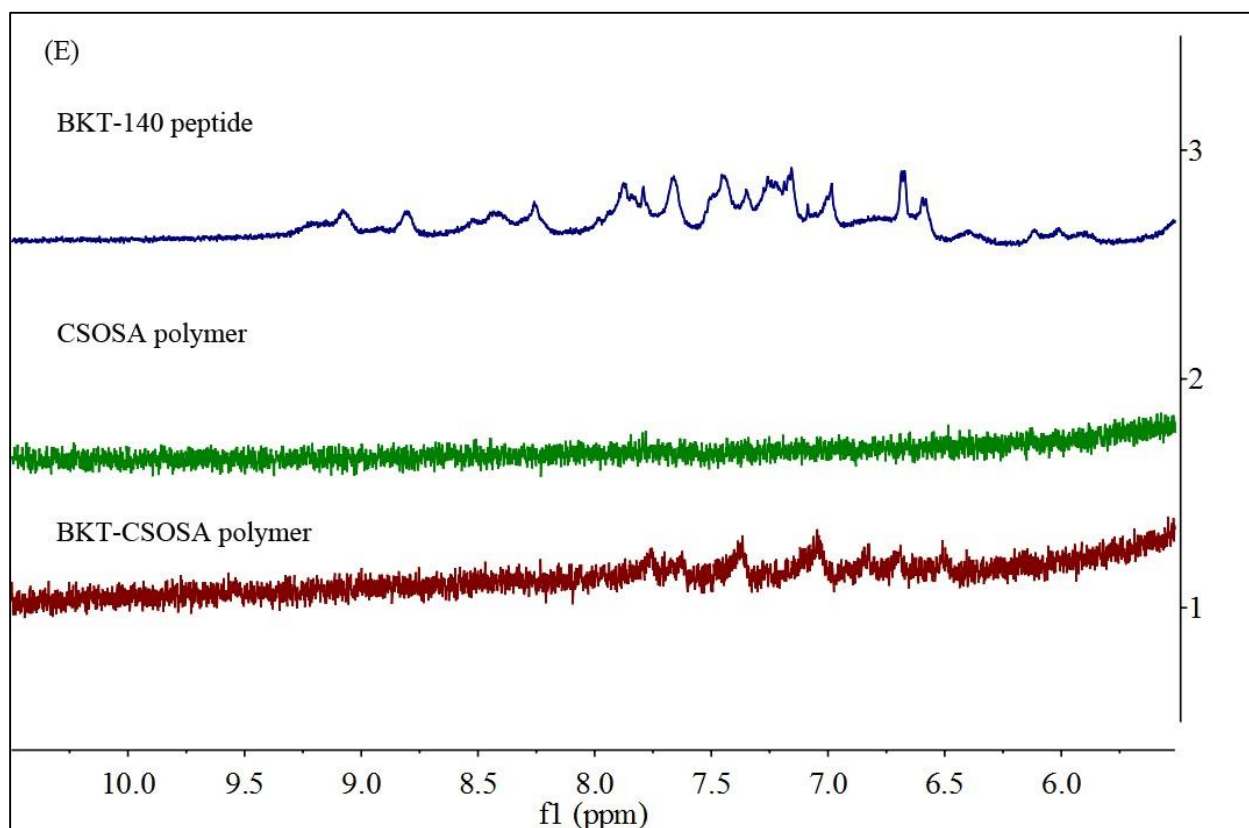


Figure S4. ^1H NMR spectrum of (A) BKT-140 peptide, (B) $\text{NH}_2\text{-PEG}_{2000}\text{-NH}_2$, (C) CSOSA polymer and (D) BKT-CSOSA. (E) shows the magnification of waves between 9.5-6.0 ppm. The chemical structures of BKT-140 peptide, $\text{NH}_2\text{-PEG}_{2000}\text{-NH}_2$, CSOSA polymer and BKT-CSOSA polymer were measured by ^1H NMR (Figure S4 (A) (B) (C) (D)). The peaks at about 1.94 ppm, 2.10 ppm and 3.76 ppm of BKT-CSOSA were attributed to $-\text{CH}_3$, $-\text{NH}_2$ and $-\text{COOH}$ of CSOSA respectively. And the peaks at about 3.58 ppm of BKT-CSOSA were attributed to $-\text{CH}_2\text{CH}_2\text{O}-$ of $\text{NH}_2\text{-PEG}_{2000}\text{-NH}_2$. The tiny but indelible waves near peaks at 6.68 ppm, 6.99 ppm and 7.78 ppm of BKT-CSOSA polymer (Figure S4 (E)) belong to BKT-140 peptide. These results indicated that BKT-140 peptide was conjugated to CSOSA.