

Supporting Information

for

**Molecularly-Defined Macrocycles Containing Azobenzene Main-Chain Oligomers:
Modular Stepwise Synthesis, Chain-Length and Topology-Dependent Properties**

*Xi Jiang,^a Jinjie Lu,^a Feng Zhou,^a Zhengbiao Zhang,^a Xiangqiang Pan,^a Wei Zhang,^a Yong Wang,^b
Nianchen Zhou,^{*a} Xiulin Zhu^{*a}*

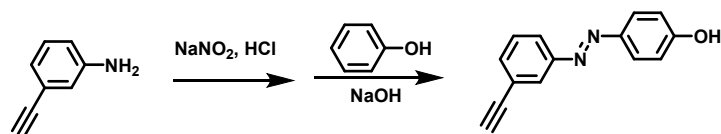
^a State and Local Joint Engineering Laboratory for Novel Functional Polymeric Materials; Jiangsu Key Laboratory of Advanced Functional Polymer Design and Application; Suzhou Key Laboratory of Macromolecular Design and Precision Synthesis; College of Chemistry, Chemical Engineering and Materials Science, Soochow University, Suzhou 215123, China.

^b Key Laboratory of Organic Synthesis of Jiangsu Province College of Chemistry, Chemical Engineering and Materials Science, Soochow University, Suzhou 215123, P. R. China.

*Corresponding authors. E-mail: xlzhu@suda.edu.cn; nczhou@suda.edu.cn

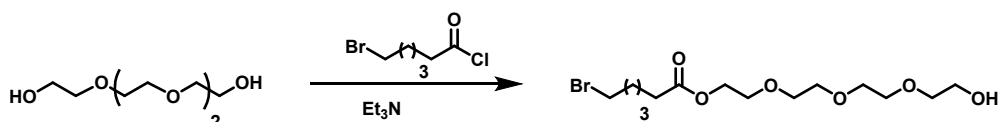
Synthetic procedures

Synthesis of HAAzo



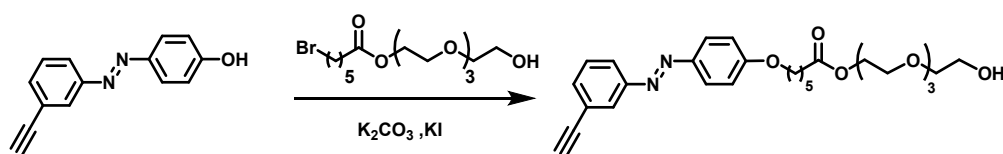
¹H NMR (CDCl₃), δ (TMS, ppm): 8.04-7.97 (s, 1H, ArH), 7.94-7.83 (m, 3H, ArH), 7.62-7.52 (d, 1H, ArH), 7.50-7.42 (m, 1H, ArH), 7.00-6.91 (d, 2H, ArH), 5.36-5.27 (s, 1H, ArOH), 3.17-3.10 (s, 1H, ArC≡CH).

Synthesis of Br-TEG-OH



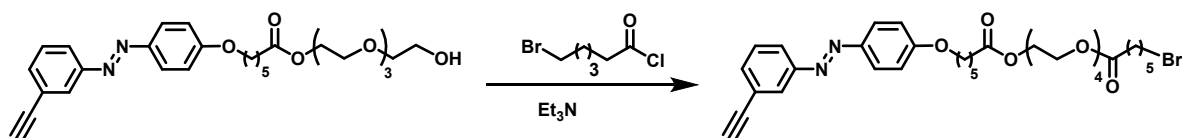
¹H NMR (300 MHz, CDCl₃) δ 4.27-4.20 (m, 2H, -COOCH₂CH₂-), 3.73 (dd, *J* = 8.8, 4.4 Hz, 4H, -CH₂CH₂O-), 3.67 (s, 8H, -OCH₂CH₂O-), 3.64-3.59 (m, 2H, -CH₂OH), 3.54 (t, *J* = 6.6 Hz, 1H, -BrCH₂-), 3.41 (t, *J* = 6.7 Hz, 1H, -BrCH₂-), 2.36 (t, *J* = 7.4 Hz, 3H, -CH₂COO-, -OH), 1.94-1.76 (m, 2H, -CH₂CH₂-), 1.73-1.60 (m, 2H, -CH₂CH₂-), 1.54-1.41 (m, 2H, -CH₂CH₂-).

Synthesis of Azo-TEG-OH



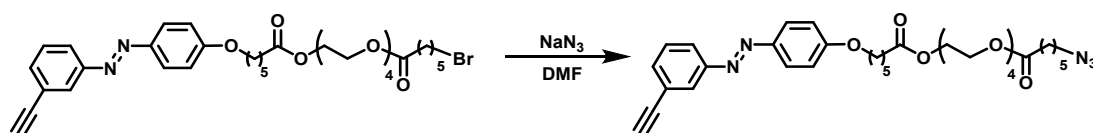
¹H NMR (300 MHz, CDCl₃) δ 8.00 (s, 1H, phenyl), 7.89 (d, *J* = 13.9, 8.5 Hz, 3H, phenyl), 7.55 (d, *J* = 7.6 Hz, 1H, phenyl), 7.45 (t, *J* = 7.8 Hz, 1H, phenyl), 7.00 (d, *J* = 9.0 Hz, 2H, phenyl), 4.28-4.22 (m, 2H, -OCOCH₂-), 4.05 (t, *J* = 6.4 Hz, 2H, -OCH₂CH₂-), 3.72 (dd, *J* = 5.1, 3.8 Hz, 4H, -OCH₂CH₂O-), 3.66 (s, 9H, -OCH₂CH₂O-), 3.63-3.57 (m, 2H, -CH₂OH), 3.13 (s, 1H, -C≡CH), 2.40 (t, *J* = 7.4 Hz, 2H, -CH₂COO-), 1.90-1.78 (m, 2H, -CH₂CH₂-), 1.71 (dt, *J* = 12.7, 6.5 Hz, 2H, -CH₂CH₂-), 1.62-1.46 (m, 2H), 1.43 (s, 2H, -CH₂CH₂-).

Synthesis of (Azo-TEG)₁-Br



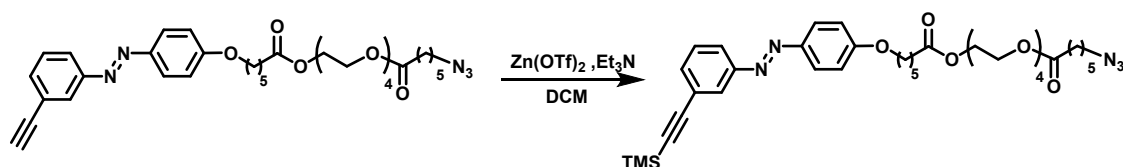
¹H NMR (300 MHz, CDCl₃) δ 8.10-7.77 (m, 4H, phenyl), 7.49 (dt, *J* = 15.5, 7.7 Hz, 2H, phenyl), 7.00 (d, *J* = 9.0 Hz, 2H, phenyl), 4.24 (dd, *J* = 9.3, 4.3 Hz, 4H, -OCOCH₂-), 4.05 (t, *J* = 6.4 Hz, 2H, -OCH₂CH₂-), 3.73-3.67 (m, 4H, -OCOCH₂-), 3.65 (s, 8H, -OCH₂CH₂O-), 3.63-3.57 (m, 2H, -CH₂Br), 3.13 (s, 1H, -C≡CH), 2.37 (dt, *J* = 12.9, 7.4 Hz, 4H, -CH₂COO-), 1.91-1.77 (m, 4H, -CH₂CH₂-), 1.68 (ddd, *J* = 21.8, 15.0, 7.2 Hz, 4H, -CH₂CH₂-), 1.60-1.42 (m, 4H, -CH₂CH₂-).

Synthesis of alkynyl-Azo-TEG-N₃



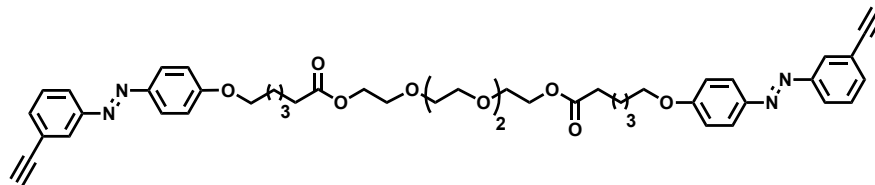
¹H NMR (300 MHz, CDCl₃) δ 8.10-7.77 (m, 4H, phenyl), 7.49 (dt, *J* = 15.5, 7.7 Hz, 2H, phenyl), 6.99 (d, *J* = 8.9 Hz, 2H, phenyl), 4.23 (dd, *J* = 9.3, 4.3 Hz, 4H, -OCOCH₂-), 4.05 (t, *J* = 6.4 Hz, 2H, -OCH₂CH₂-), 3.70 (dt, *J* = 8.3, 3.5 Hz, 4H, -OCOCH₂-), 3.65 (d, *J* = 1.1 Hz, 8H, -OCH₂CH₂O-), 3.27 (t, *J* = 6.8 Hz, 2H, -CH₂N₃), 3.13 (s, 1H, -C≡CH), 2.53-2.28 (m, 4H, -CH₂COO-), 1.92-1.31 (m, 12H, -CH₂CH₂-).

Synthesis of TMS-Azo-TEG-N₃



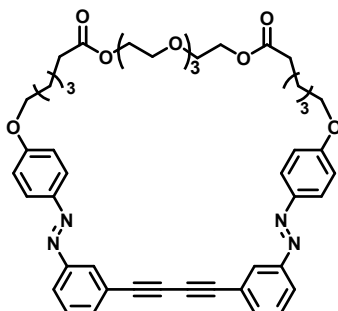
¹H NMR (300 MHz, CDCl₃) δ 8.03-7.75 (m, 4H, phenyl), 7.58-7.38 (m, 2H, phenyl), 6.99 (d, *J* = 8.1 Hz, 2H, phenyl), 4.23 (s, 4H, -OCOCH₂-), 4.05 (s, 2H, -OCH₂CH₂-), 3.69 (s, 4H, -OCOCH₂-), 3.65 (s, 8H, -OCH₂CH₂O-), 3.27 (s, 2H, -CH₂TMS), 2.46-2.29 (m, 4H, -CH₂COO-), 1.92-1.31 (m, 12H, -CH₂CH₂-), 0.27 (s, 9H, -SiH₃).

Synthesis of *l*-(Azo₂-TEG₁) (n = 1)



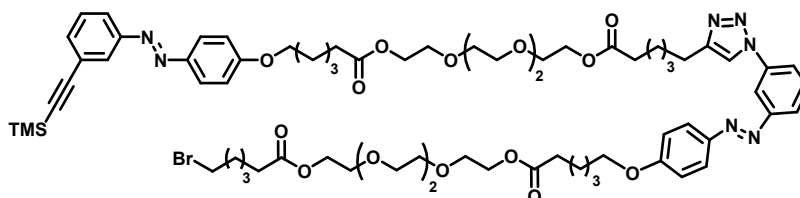
¹H NMR (300 MHz, CDCl₃) δ 8.00 (s, 2H, phenyl), 7.95-7.82 (m, 6H, phenyl), 7.49 (dt, *J* = 15.5, 7.7 Hz, 4H, phenyl), 6.99 (d, *J* = 8.9 Hz, 4H, phenyl), 4.28-4.20 (m, 4H, -OCOCH₂CH₂-), 4.04 (t, *J* = 6.4 Hz, 4H, -OCH₂CH₂-), 3.73-3.61 (m, 12H, -CH₂CH₂-), 3.13 (s, 2H, -C≡CH), 2.39 (t, *J* = 7.4 Hz, 4H, -OCOCH₂CH₂-), 1.90-1.45 (m, 12H, -CH₂CH₂-).

Synthesis of *c*-(Azo₂-TEG₁) (n = 1)



¹H NMR (300 MHz, CDCl₃) δ 8.00 (s, 2H, phenyl), 7.95-7.82 (m, 6H, phenyl), 7.49 (dt, *J* = 15.5, 7.7 Hz, 4H, phenyl), 6.99 (d, *J* = 8.9 Hz, 4H, phenyl), 4.28-4.20 (m, 4H, -OCOCH₂CH₂-), 4.04 (t, *J* = 6.4 Hz, 4H, -OCH₂CH₂-), 3.73-3.61 (m, 12H, -CH₂CH₂-), 2.39 (t, *J* = 7.4 Hz, 4H, -OCOCH₂CH₂-), 1.90-1.45 (m, 12H, -CH₂CH₂-).

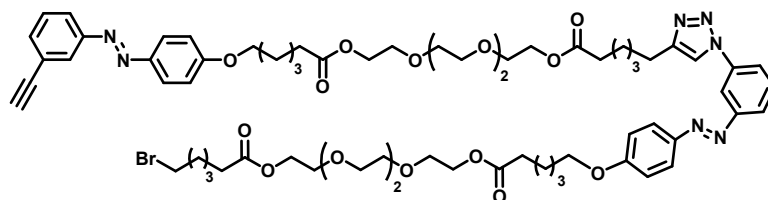
Synthesis of TMS-(Azo-TEG)₂-Br for general procedure for “CuAAC” reaction



¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 1H, triazole ring), 8.12-6.68 (m, 16H, pheny), 4.43 (t, *J* = 7.1 Hz,

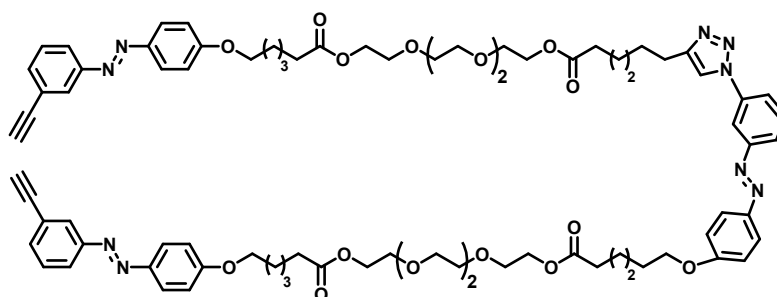
2H, triazole-CH₂CH₂-), 4.28-4.18 (m, 8H, -OCOCH₂-), 4.04 (dd, *J* = 10.5, 6.2 Hz, 4H, -OCH₂CH₂-), 3.70 (dd, *J* = 9.4, 4.7 Hz, 8H, -OCOCH₂-), 3.65 (d, *J* = 1.4 Hz, 16H, -OCH₂CH₂O-), 3.40 (s, 2H, -CH₂Br), 2.35 (s, 8H, -CH₂COO-), 1.93-1.44 (m, 24H, -CH₂CH₂-), 0.27 (s, 9H, -SiH₃).

Synthesis of (Azo-TEG)₂-Br for alkyne deprotection



¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 1H, triazole ring), 8.12-6.68 (m, 16H, pheny), 4.43 (t, *J* = 7.1 Hz, 2H, triazole-CH₂CH₂-), 4.28-4.18 (m, 8H, -OCOCH₂-), 4.04 (dd, *J* = 10.5, 6.2 Hz, 4H, -OCH₂CH₂-), 3.70 (dd, *J* = 9.4, 4.7 Hz, 8H, -OCOCH₂-), 3.65 (d, *J* = 1.4 Hz, 16H, -OCH₂CH₂O-), 3.40 (s, 2H, -CH₂Br), 3.13 (s, 1H, -C≡CH), 2.35 (s, 8H, -CH₂COO-), 1.93-1.44 (m, 24H, -CH₂CH₂-).

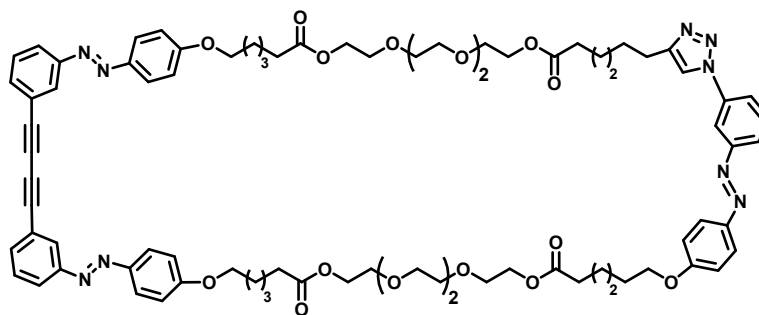
Synthesis of *l*-(Azo₃-TEG₂) (*n* = 2)



The *l*-(Azo₃-TEG₂) was prepared using the similar procedures as *l*-(Azo₂-TEG₁). A solution of (Azo-TEG)₂-Br (1.339 g, 1 mmol), HAAzo (0.888 g, 4 mmol), potassium carbonate (0.552 g, 1 mmol), a catalytic amount of potassium iodide, and 50 mL of DMF was prepared in a 100mL round bottom flask under vigorous stirring. The solution was stirred at 80 °C for 4h. After cooling to room temperature, deionized water (4×200 mL) was added and the mixture was extracted with ethyl acetate (200 mL). The organic layer obtained was dried with anhydrous MgSO₄ overnight, filtered, and evaporated in a reduced pressure. The final crude product was purified by column chromatography (silica gel, ethyl acetate) to

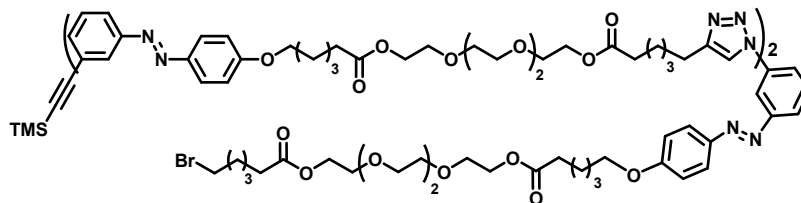
yield the *l*-(Azo₃-TEG₂) as yellow liquid (1.33 g, 90.9%). ¹H NMR (300 MHz, CDCl₃) δ 8.27 (s, 1H, triazole ring), 8.07-6.81 (m, 24H, pheny), 4.42 (t, *J* = 7.1 Hz, 2H, triazole-CH₂CH₂-), 4.29-4.15 (m, 8H, -OCOCH₂-), 4.03 (t, *J* = 6.3 Hz, 6H, -OCH₂CH₂-), 3.75-3.66 (m, 8H, -OCOCH₂-), 3.66-3.53 (m, 16H, -OCH₂CH₂O-), 3.13 (s, 2H, -C≡CH), 2.58-2.21 (m, 8H, -CH₂COO-), 2.12-1.19 (m, 24H, -CH₂CH₂-).

Synthesis of *c*-(Azo₃-TEG₂) (*n* = 2)



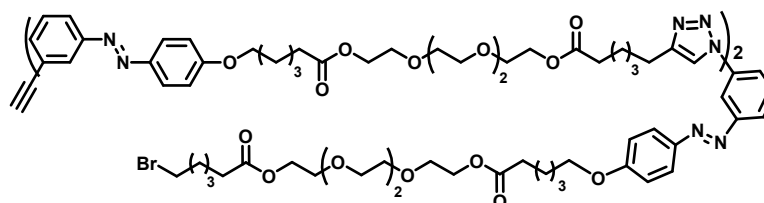
To a 1000 mL three-necked round-bottomed flask was added acetone (800 mL), triethylamine 50 mL, CuI (0.955 g, 50 mmol), and TMEDA (1.5 mL, 10 mmol), and the solution was stirred for 1 h. The *l*-(Azo₃-TEG₂) (0.148 g, 0.1 mmol) in 20 mL of acetone was added to CuI/TMEDA mixture at room temperature *via* syring pump at rate of 0.4 mL/h. After the addition of polymer solution was completed, the reaction was allowed to proceed for another period of 24 h. The reaction solution was concentrated in a reduced pressure, deionized water (3×500 mL) was added and the mixture was extracted with ethyl acetate (400 mL) to remove the copper catalyst residues. The organic layer obtained was dried with anhydrous MgSO₄ overnight, filtered, and evaporated in a reduced pressure. The final crude product was purified by column chromatography (silica gel, ethyl acetate) to yield *c*-(Azo₃-TEG₂) as yellow liquid (0.082 g, 80%). ¹H NMR (300 MHz, CDCl₃) δ 8.27 (s, 1H, triazole ring), 8.07-6.81 (m, 24H, pheny), 4.42 (t, *J* = 7.1 Hz, 2H, triazole-CH₂CH₂-), 4.29-4.15 (m, 8H, -OCOCH₂-), 4.03 (t, *J* = 6.3 Hz, 6H, -OCH₂CH₂-), 3.75-3.66 (m, 8H, -OCOCH₂-), 3.66-3.53 (m, 16H, -OCH₂CH₂O-), 2.58-2.21 (m, 8H, -CH₂COO-), 2.12-1.19 (m, 24H, -CH₂CH₂-).

Synthesis of TMS-(Azo-TEG)₃-Br



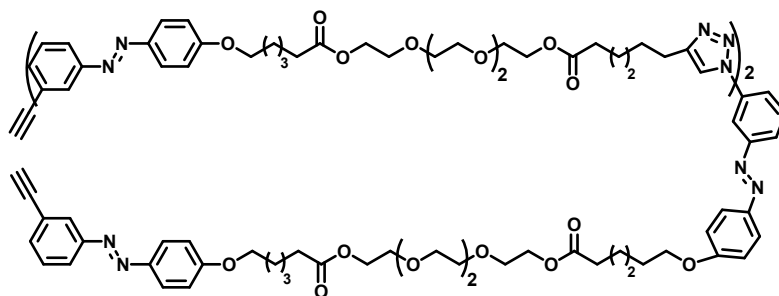
The TMS-(Azo-TEG)₃-Br was prepared from (Azo-TEG)₂-Br (3.3g, 2.46 mmol) and TMS-Azo-TEG-N₃ (2.13 g, 2.95 mmol) according to the general procedure for click coupling reaction. The product was purified by column chromatography (silica gel, THF/petroleum ether = 1/1) to yield TMS-(Azo-TEG)₃-Br as yellow liquid (4.77 g, 94%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 2H, triazole ring), 8.09-6.76 (m, 32H, pheny), 4.41 (dd, *J* = 15.5, 8.5 Hz, 4H, triazole-CH₂CH₂-), 4.23 (s, 14H, -OCOCH₂-), 4.04 (d, *J* = 4.2 Hz, 6H, -OCH₂CH₂-), 3.75-3.67 (m, 12H, -OCOCH₂-), 3.64 (d, *J* = 3.8 Hz, 32H, -OCH₂CH₂O-), 3.40 (t, *J* = 6.7 Hz, 2H, -CH₂Br), 2.37 (dt, *J* = 14.8, 7.3 Hz, 12H, -CH₂COO-), 1.93-1.44 (m, 36H, -CH₂CH₂-), 0.27 (s, 9H, -SiH₃).

Synthesis of (Azo-TEG)₃-Br



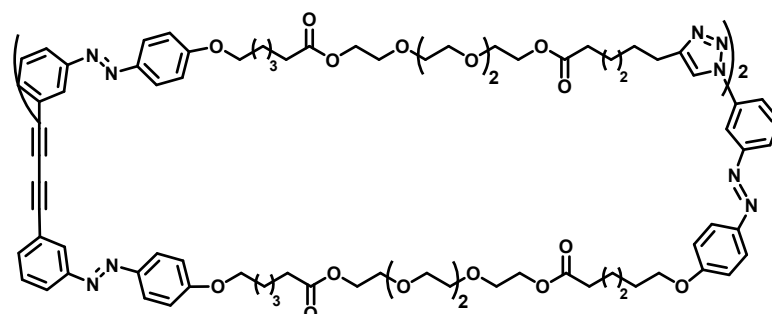
The (Azo-TEG)₃-Br was prepared from the TMS-(Azo-TEG)₃-Br (4.77 g, 2.3 mmol) according to the general procedure for alkyne deprotection, yielding 4.44 g of a viscous yellow oil (97%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 2H, triazole ring), 8.09-6.76 (m, 32H, pheny), 4.41 (dd, *J* = 15.5, 8.5 Hz, 4H, triazole-CH₂CH₂-), 4.23 (s, 14H, -OCOCH₂-), 4.04 (d, *J* = 4.2 Hz, 6H, -OCH₂CH₂-), 3.75-3.67 (m, 12H, -OCOCH₂-), 3.64 (d, *J* = 3.8 Hz, 32H, -OCH₂CH₂O-), 3.40 (t, *J* = 6.7 Hz, 2H, -CH₂Br), 3.13 (s, 1H, -C≡CH), 2.37 (dt, *J* = 14.8, 7.3 Hz, 12H, -CH₂COO-), 1.93-1.44 (m, 36H, -CH₂CH₂-).

Synthesis of *l*-(Azo₄-TEG₃) (n = 3)



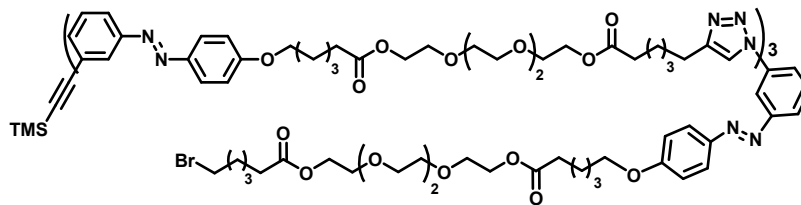
The *l*-(Azo₄-TEG₃) was prepared from (Azo-TEG)₃-Br (1.99 g, 1 mmol) using the similar procedures as *l*-(Azo₂-TEG₁). The yield *l*-(Azo₄-TEG₃) as yellow liquid (2.06 g, 97%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (t, *J* = 1.6 Hz, 2H, triazole ring), 8.09-6.62 (m, 32H, pheny), 4.41 (t, *J* = 7.0 Hz, 4H, triazole-CH₂CH₂-), 4.23 (dt, *J* = 6.1, 3.4 Hz, 12H, -OCOCH₂-), 4.03 (t, *J* = 6.3 Hz, 8H, -OCH₂CH₂-), 3.73-3.66 (m, 12H, -OCOCH₂-), 3.64 (dd, *J* = 5.1, 1.9 Hz, 24H, -OCH₂CH₂O-), 3.13 (s, 2H, -C≡CH), 2.52-2.27 (m, 12H, -CH₂COO-), 2.07-1.26 (m, 24H, -CH₂CH₂-).

Synthesis of *c*-(Azo₄-TEG₃) (n = 3)



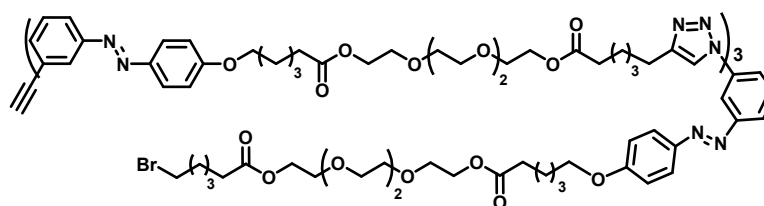
The *c*-(Azo₄-TEG₃) was prepared from *l*-(Azo₄-TEG₃) (158 mg, 0.07 mmol) according to the cyclization procedure above, using a 300 eq CuI. The yield *l*-(Azo₄-TEG₃) was purified by preparative GPC as yellow liquid (137 mg, 80%). ¹H NMR (300 MHz, CDCl₃) δ 8.25 (s, 2H, triazole ring), 8.10-6.63 (m, 32H, pheny), 4.52-4.29 (m, 4H, triazole-CH₂CH₂-), 4.31-4.12 (m, 12H, -OCOCH₂-), 4.02 (dd, *J* = 7.5, 5.0 Hz, 8H, -OCH₂CH₂-), 3.69 (dt, *J* = 9.0, 3.4 Hz, 12H, -OCOCH₂-), 3.65-3.54 (m, 24H, -OCH₂CH₂O-), 2.58-2.22 (m, 12H, -CH₂COO-), 2.06-1.08 (m, 24H, -CH₂CH₂-).

Synthesis of TMS-(Azo-TEG)₄-Br



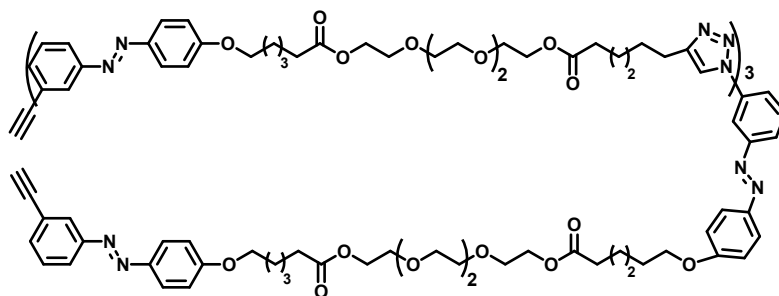
The TMS-(Azo-TEG)₄-Br was prepared from (Azo-TEG)₃-Br (2.4 g, 1.2 mmol) and TMS-Azo-TEG-N₃ (1.04 g, 1.44 mmol) according to the “CuAAC” reaction above. The product was purified by column chromatography (silica gel, THF/petroleum ether = 2/1) to yield TMS-(Azo-TEG)₄-Br as yellow liquid (2.93 g, 90%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 3H, triazole ring), 8.03-6.43 (m, 32H, pheny), 4.42 (t, *J* = 7.0 Hz, 6H, triazole-CH₂CH₂-), 4.23 (td, *J* = 7.9, 3.9 Hz, 16H, -OCOCH₂-), 4.11-3.95 (m, 8H, -OCH₂CH₂-), 3.72-3.65 (m, 16H, -OCOCH₂-), 3.64 (dd, *J* = 5.2, 1.8 Hz, 32H, -OCH₂CH₂O-), 3.57-3.32 (m, 2H, -CH₂Br), 2.48-2.28 (m, 16H, -CH₂COO-), 2.12-1.30 (m, 48H, -CH₂CH₂-), 0.27 (s, 9H, -SiH₃).

Synthesis of (Azo-TEG)₄-Br



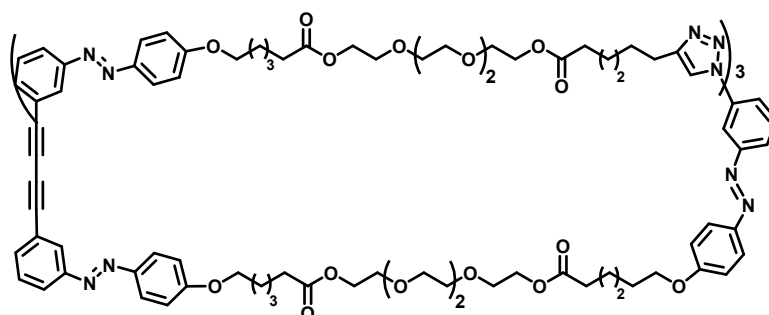
The (Azo-TEG)₄-Br was prepared from TMS-(Azo-TEG)₄-Br (2.93 g, 1 mmol) according to the alkyne deprotection above, yielding 2.56 g of a yellow liquid (97%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (d, *J* = 1.4 Hz, 3H, triazole ring), 8.10-6.71 (m, 32H, pheny), 4.52-4.32 (m, 6H, triazole-CH₂CH₂-), 4.23 (td, *J* = 8.1, 4.0 Hz, 16H, -OCOCH₂-), 4.04 (dd, *J* = 10.4, 6.2 Hz, 8H, -OCH₂CH₂-), 3.69 (dd, *J* = 9.7, 4.6 Hz, 16H, -OCOCH₂-), 3.64 (dd, *J* = 5.1, 1.9 Hz, 32H, -OCH₂CH₂O-), 3.60-3.35 (m, 2H, -CH₂Br), 3.12 (d, *J* = 10.8 Hz, 2H, -C≡CH), 2.37 (dt, *J* = 14.9, 5.8 Hz, 16H, -CH₂COO-), 2.05-1.24 (m, 48H, -CH₂CH₂-).

Synthesis of *l*-(Azo₅-TEG₄) (n = 4)



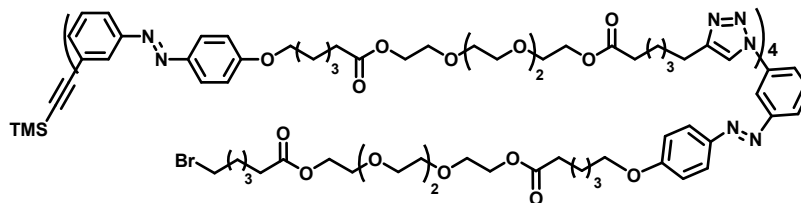
The *l*-(Azo₅-TEG₄) was prepared from (Azo-TEG)₄-Br (1.32 g, 0.5 mmol) using the similar procedures as *l*-(Azo₂-TEG₁). The product was purified by column chromatography (silica gel, THF/petroleum ether = 2/1) to yield *l*-(Azo₅-TEG₄) as yellow liquid (1.28 g, 90%). ¹H NMR (300 MHz, CDCl₃) δ 8.27 (t, *J* = 1.6 Hz, 3H, triazole ring), 8.06-6.69 (m, 40H, pheny), 4.50-4.32 (m, 6H, triazole-CH₂CH₂-), 4.28-4.14 (m, 16H, -OCOCH₂-), 4.03 (t, *J* = 6.3 Hz, 10H, -OCH₂CH₂-), 3.69 (td, *J* = 5.0, 2.6 Hz, 16H, -OCOCH₂-), 3.64 (dd, *J* = 5.0, 2.0 Hz, 32H, -OCH₂CH₂O-), 3.11 (d, *J* = 10.6 Hz, 2H, -C≡CH), 2.54-2.22 (m, 16H, -CH₂COO-), 2.06-1.23 (m, 48H, -CH₂CH₂-).

Synthesis of *c*-(Azo₅-TEG₄) (n = 4)



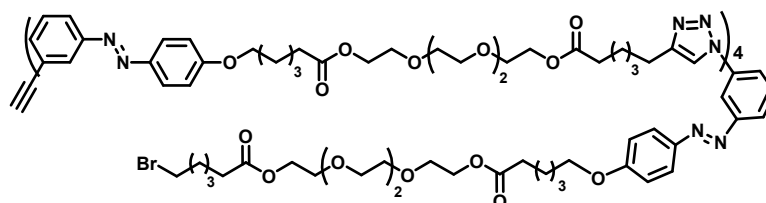
The *c*-(Azo₅-TEG₄) was prepared from *l*-(Azo₅-TEG₄) (139 mg, 0.05 mmol) according to the general procedure for cyclization, using a 300 eq CuI. The yield *c*-(Azo₅-TEG₄) was purified by preparative GPC as yellow liquid (121 mg, 75%). ¹H NMR (300 MHz, CDCl₃) δ 8.26 (s, 3H, triazole ring), 8.09-6.82 (m, 40H, pheny), 4.39 (dd, *J* = 14.7, 7.6 Hz, 6H, triazole-CH₂CH₂-), 4.22 (td, *J* = 7.9, 4.2 Hz, 16H, -OCOCH₂-), 4.10-3.94 (m, 10H, -OCH₂CH₂-), 3.68 (dd, *J* = 9.4, 4.3 Hz, 16H, -OCOCH₂-), 3.65-3.45 (m, 32H, -OCH₂CH₂O-), 2.36 (ddd, *J* = 17.4, 8.4, 5.4 Hz, 16H, -CH₂COO-), 2.05-1.20 (m, 48H, -CH₂CH₂-).

Synthesis of TMS-(Azo-TEG)₅-Br



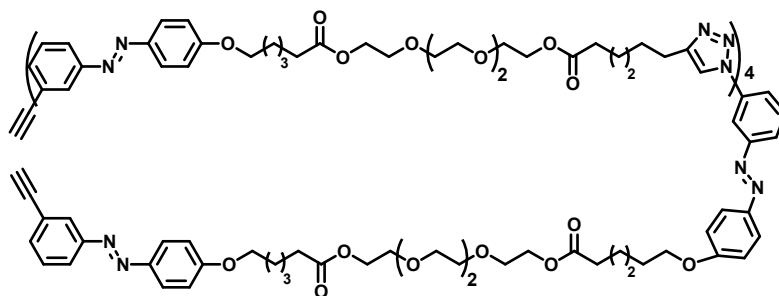
The TMS-(Azo-TEG)₅-Br was prepared from (Azo-TEG)₄-Br (1.95 g, 0.738 mmol) and TMS-Azo-TEG-N₃ (0.64 g, 0.885 mmol) according to the CuAAC procedure above. The product was purified by column chromatography (silica gel, THF/petroleum ether = 3/1) to yield TMS-(Azo-TEG)₅-Br as yellow liquid (2.23 g, 90%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 4H, triazole ring), 8.13-6.79 (m, 40H, pheny), 4.42 (t, *J* = 6.8 Hz, 8H, triazole-CH₂CH₂-), 4.31-4.15 (m, 20H, -OCOCH₂-), 4.13-3.93 (m, 10H, -OCH₂CH₂-), 3.69 (dd, *J* = 9.7, 4.8 Hz, 20H, -OCOCH₂-), 3.64 (dd, *J* = 5.3, 1.8 Hz, 40H, -OCH₂CH₂O-), 3.46 (dt, *J* = 38.7, 6.7 Hz, 2H, -CH₂Br), 2.44-2.24 (m, 20H, -CH₂COO-), 2.04-1.19 (m, 60H, -CH₂CH₂-), 0.29-0.20 (m, 9H, -SiH₃).

Synthesis of (Azo-TEG)₅-Br



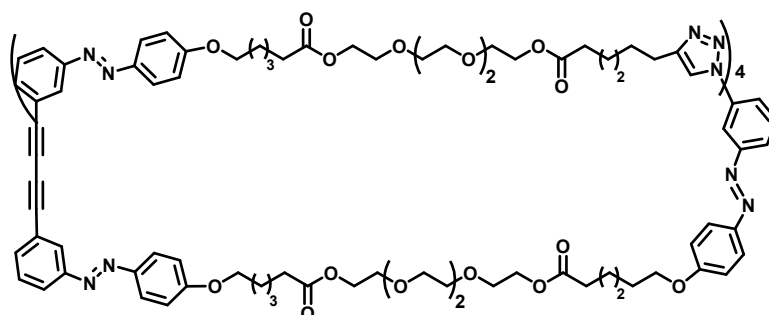
The (Azo-TEG)₅-Br was prepared from TMS-(Azo-TEG)₅-Br (2.23 g, 6.6 mmol) according to the alkyne deprotection above, yielding 2.06 g of a viscous yellow oil (94%). ¹H NMR (300 MHz, CDCl₃) δ 8.27 (s, 4H, triazole ring), 8.03-6.87 (m, 40H, pheny), 4.49-4.34 (m, 8H, triazole-CH₂CH₂-), 4.31-4.16 (m, 20H, -OCOCH₂-), 4.10-3.98 (m, 10H, -OCH₂CH₂-), 3.69 (dd, *J* = 9.7, 4.7 Hz, 20H, -OCOCH₂-), 3.64 (dd, *J* = 5.3, 1.8 Hz, 40H, -OCH₂CH₂O-), 3.40 (t, *J* = 6.7 Hz, 2H, -CH₂Br), 3.11 (d, *J* = 10.5 Hz, 1H, -C≡CH), 2.47-2.26 (m, 20H, -CH₂COO-), 2.06-1.31 (m, 60H, -CH₂CH₂-).

Synthesis of *l*-(Azo₆-TEG₅) (n = 5)



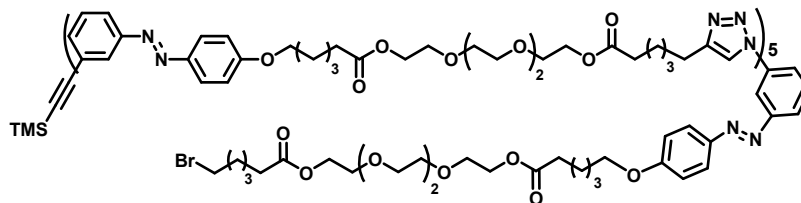
The *l*-(Azo₆-TEG₅) was prepared from (Azo-TEG)₅-Br (548 mg, 0.16 mmol) using the similar procedures of preparing the *l*-(Azo₂-TEG₁). The product was purified by column chromatography (silica gel, THF/petroleum ether = 3/1) to yield *l*-(Azo₆-TEG₅) as yellow liquid (453 mg, 86%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 4H, triazole ring), 8.07-6.60 (m, 48H, pheny), 4.41 (t, *J* = 7.0 Hz, 8H, triazole-CH₂CH₂-), 4.22 (dd, *J* = 8.7, 3.9 Hz, 20H, -OCOCH₂-), 4.03 (t, *J* = 6.3 Hz, 12H, -OCH₂CH₂-), 3.68 (dd, *J* = 8.6, 4.0 Hz, 20H, -OCOCH₂-), 3.64 (d, *J* = 3.7 Hz, 40H, -OCH₂CH₂O-), 3.14 (s, 2H, -C≡CH), 2.47-2.26 (m, 20H, -CH₂COO-), 2.04-1.27 (m, 60H, -CH₂CH₂-).

Synthesis of *c*-(Azo₆-TEG₅) (n = 5)



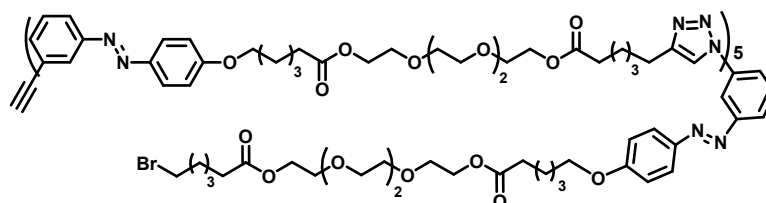
The *c*-(Azo₆-TEG₅) was prepared from *l*-(Azo₆-TEG₅) (112 mg, 0.03 mmol) according to similar cyclization procedure above, using a 400 eq CuI. The yield *c*-(Azo₆-TEG₅) was purified by preparative GPC as yellow liquid (100 mg, 75%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 4H, triazole ring), 8.07-6.60 (m, 48H, pheny), 4.41 (t, *J* = 7.0 Hz, 8H, triazole-CH₂CH₂-), 4.22 (dd, *J* = 8.7, 3.9 Hz, 20H, -OCOCH₂-), 4.03 (t, *J* = 6.3 Hz, 12H, -OCH₂CH₂-), 3.68 (dd, *J* = 8.6, 4.0 Hz, 20H, -OCOCH₂-), 3.64 (d, *J* = 3.7 Hz, 40H, -OCH₂CH₂O-), 2.47-2.26 (m, 20H, -CH₂COO-), 2.04-1.27 (m, 60H, -CH₂CH₂-)

Synthesis of TMS-(Azo-TEG)₆-Br



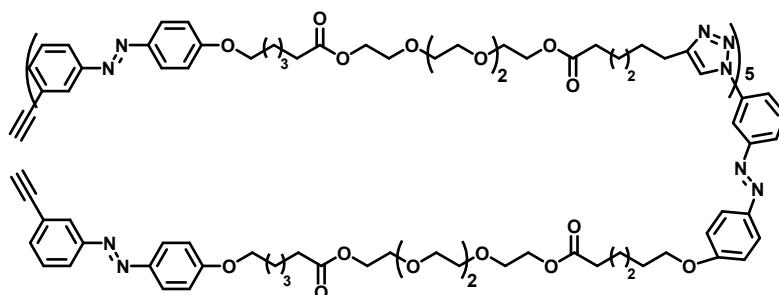
The TMS-(Azo-TEG)₆-Br was prepared from (Azo-TEG)₅-Br (2.4 g, 0.738 mmol) and TMS-Azo-TEG-N₃ (0.64 g, 0.885 mmol) according to the click coupling reaction above. The product was purified by column chromatography (silica gel, THF/petroleum ether = 3/1) to yield TMS-(Azo-TEG)₆-Br as yellow liquid (2.51 g, 85%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 5H, triazole ring), 8.13-6.79 (m, 56H, pheny), 4.50-4.31 (m, 10H, triazole-CH₂CH₂-), 4.30-4.17 (m, 24H, -OCOCH₂-), 4.04 (dd, *J* = 8.2, 4.3 Hz, 12H, -OCH₂CH₂-), 3.69 (dd, *J* = 9.6, 4.7 Hz, 24H, -OCOCH₂-), 3.64 (dd, *J* = 5.2, 1.8 Hz, 48H, -OCH₂CH₂O-), 3.46 (dt, *J* = 38.7, 6.7 Hz, 2H, -CH₂Br), 2.38 (ddd, *J* = 16.6, 11.2, 5.8 Hz, 24H, -CH₂COO-), 2.03-1.11 (m, 72H, -CH₂CH₂-), 0.35-0.20 (m, 9H, -SiH₃).

Synthesis of (Azo-TEG)₆-Br



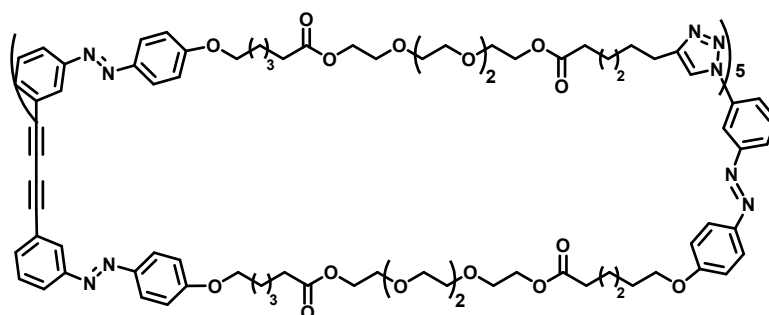
The (Azo-TEG)₆-Br was prepared from TMS-(Azo-TEG)₆-Br (2.51 g, 6.3 mmol) according to the general procedure for alkyne deprotection, yielding 2.35 g of a viscous yellow oil (95%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 5H, triazole ring), 8.13-6.79 (m, 56H, pheny), 4.50-4.31 (m, 10H, triazole-CH₂CH₂-), 4.30-4.17 (m, 24H, -OCOCH₂-), 4.04 (dd, *J* = 8.2, 4.3 Hz, 12H, -OCH₂CH₂-), 3.69 (dd, *J* = 9.6, 4.7 Hz, 24H, -OCOCH₂-), 3.64 (dd, *J* = 5.2, 1.8 Hz, 48H, -OCH₂CH₂O-), 3.46 (dt, *J* = 38.7, 6.7 Hz, 2H, -CH₂Br), 3.12 (d, *J* = 10.6 Hz, 2H, -C≡CH), 2.38 (ddd, *J* = 16.6, 11.2, 5.8 Hz, 24H, -CH₂COO-), 2.03-1.11 (m, 72H, -CH₂CH₂-).

Synthesis of *l*-(Azo₇-TEG₆) (n = 6)



The *l*-(Azo₇-TEG₆) was prepared from (Azo-TEG)₆-Br (408 mg, 0.1 mmol) using the similar procedures as *l*-(Azo₂-TEG₁). The product was purified by column chromatography (silica gel, THF/petroleum ether = 3/1) to yield *l*-(Azo₇-TEG₆) as yellow liquid (351 mg, 86%). ¹H NMR (300 MHz, CDCl₃) δ 8.28 (t, *J* = 1.6 Hz, 5H, triazole ring), 8.04-6.87 (m, 56H, pheny), 4.47-4.34 (m, 10H, triazole-CH₂CH₂-), 4.23 (dt, *J* = 9.5, 3.2 Hz, 24H, -OCOCH₂-), 4.03 (t, *J* = 6.4 Hz, 12H, -OCH₂CH₂-), 3.72-3.66 (m, 24H, -OCOCH₂-), 3.64 (dd, *J* = 5.1, 2.0 Hz, 48H, -OCH₂CH₂O-), 3.12 (d, *J* = 10.6 Hz, 2H, -C≡CH), 2.42-2.30 (m, 24H, -OCOCH₂-), 2.06-1.22 (m, 72H, -CH₂CH₂-).

Synthesis of *c*-(Azo₇-TEG₆) (n = 6)



The *c*-(Azo₇-TEG₆) was prepared from *l*-(Azo₇-TEG₆) (82 mg, 0.02 mmol) according to the similar procedure as cyclization above, using a 400 eq CuI. The yield *c*-(Azo₇-TEG₆) was purified by preparative GPC as yellow liquid (73 mg, 70%). ¹H NMR (300 MHz, CDCl₃) δ 8.27 (s, 5H, triazole ring), 8.07-6.60 (m, 56H, pheny), 4.40 (dd, *J* = 8.8, 5.2 Hz, 10H, triazole-CH₂CH₂-), 4.21 (dd, *J* = 6.2, 3.2 Hz, 24H, -OCOCH₂-), 4.02 (t, *J* = 6.3 Hz, 12H, -OCH₂CH₂-), 3.79-3.65 (m, 24H, -OCOCH₂-), 3.63 (dd, *J* = 4.5, 1.8

Hz, 48H, -OCH₂CH₂O-), 2.41-2.30 (m, 24H, -OCOCH₂-), 2.04-1.27 (m, 72H, -CH₂CH₂-).

The generated *c*-(Azo_{n+1}-TEG_n) (when n was larger than 3) was needed to purify by the preparative-GPC to move small amounts of linear precursor. The corresponding GPC traces before and after preparative-GPC can be seen from Figure S2.

Additional Figures:

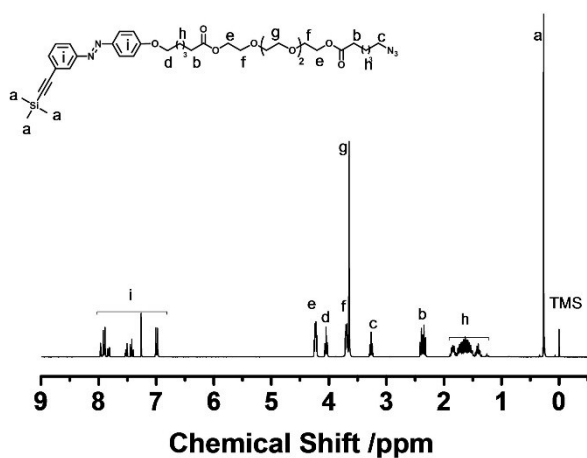


Figure S1. ¹H NMR spectrum of TMS-Azo-TEG-N₃ in CDCl₃.

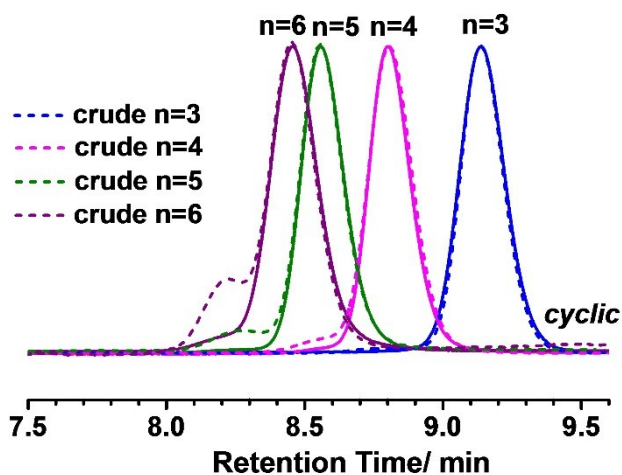


Figure S2. The GPC trace of *c*-(Azo_{n+1}-TEG_n) before and after pre-GPC using THF as the eluent (n = 3-6).

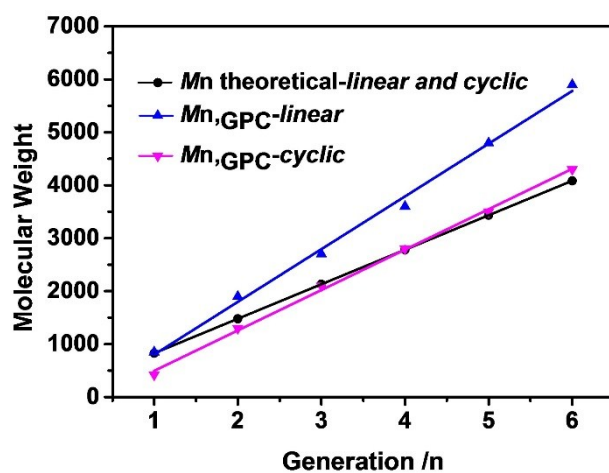


Figure S3. The dependence of the theoretical and experimental (determined by GPC) molecular weights of l -(Azo_{n+1}-TEG_n) and c -(Azo_{n+1}-TEG_n) on generation (n).

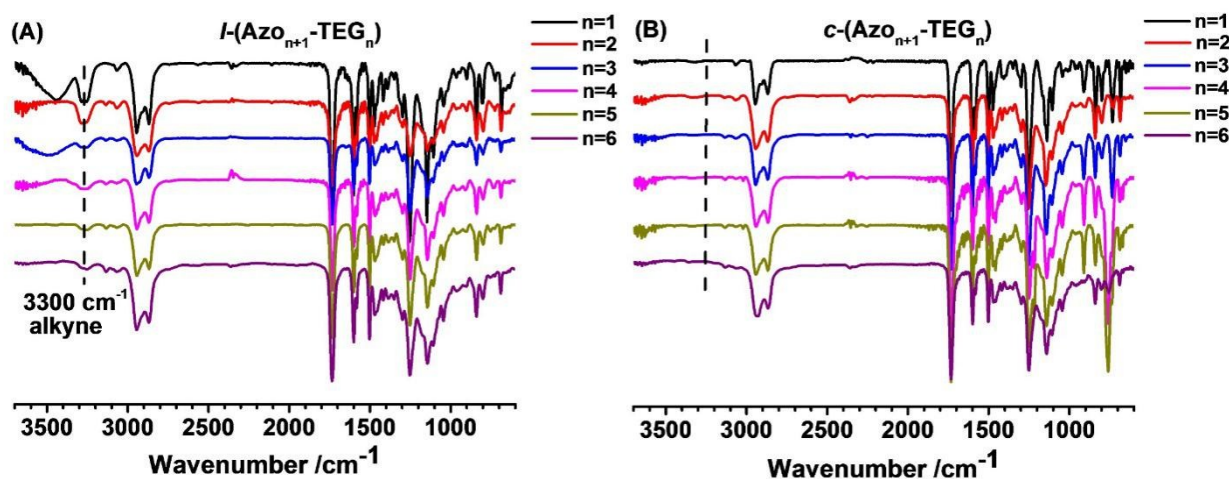


Figure S4. The FT-IR spectra of l -(Azo_{n+1}-TEG_n) (A) and c -(Azo_{n+1}-TEG_n) (B) ($n = 1-6$).

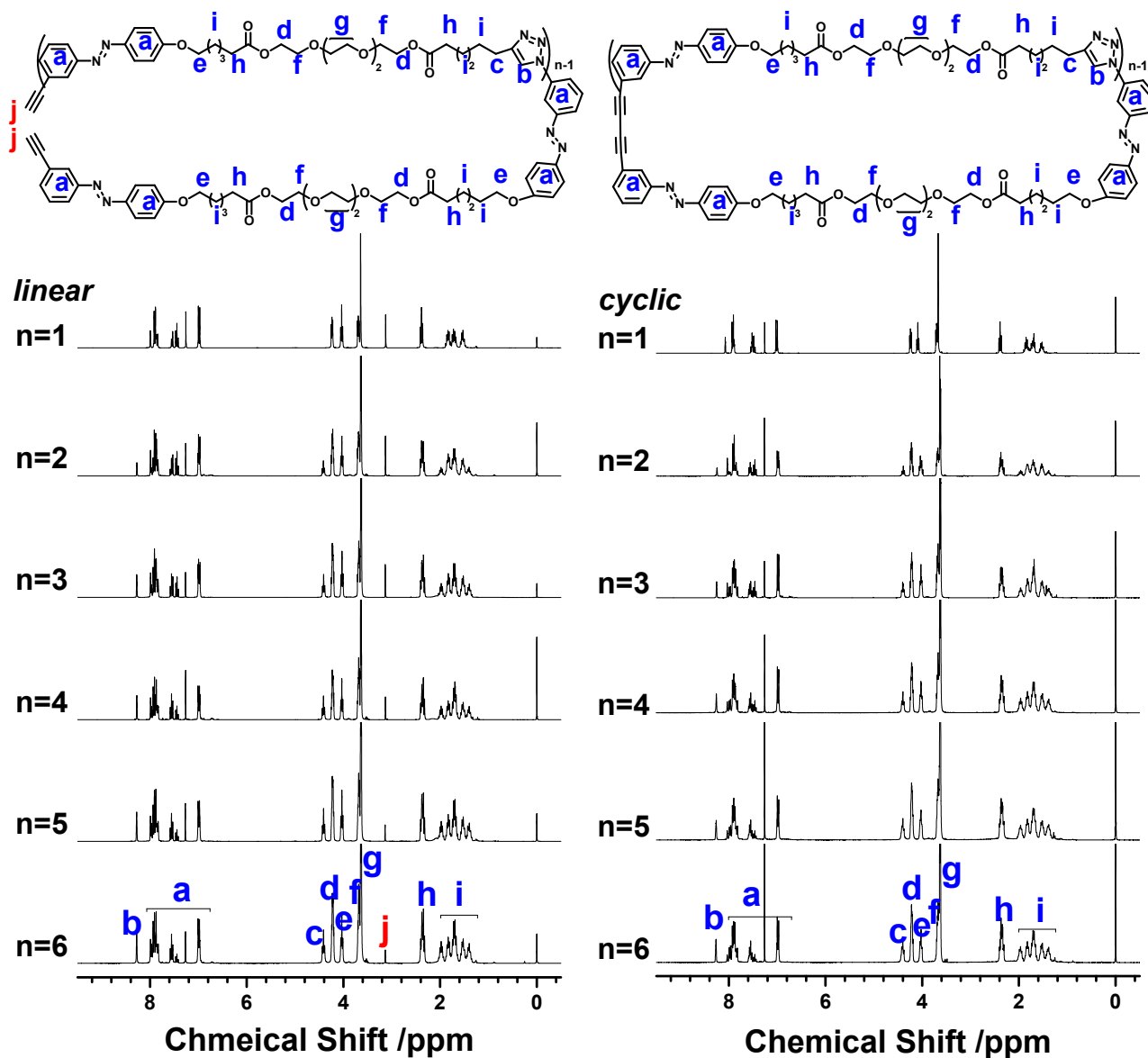


Figure S5. ^1H NMR spectra of the $l\text{-(Azo}_{n+1}\text{-TEG}_n)$ and $c\text{-(Azo}_{n+1}\text{-TEG}_n)$ in CDCl_3 ($n=1-6$).

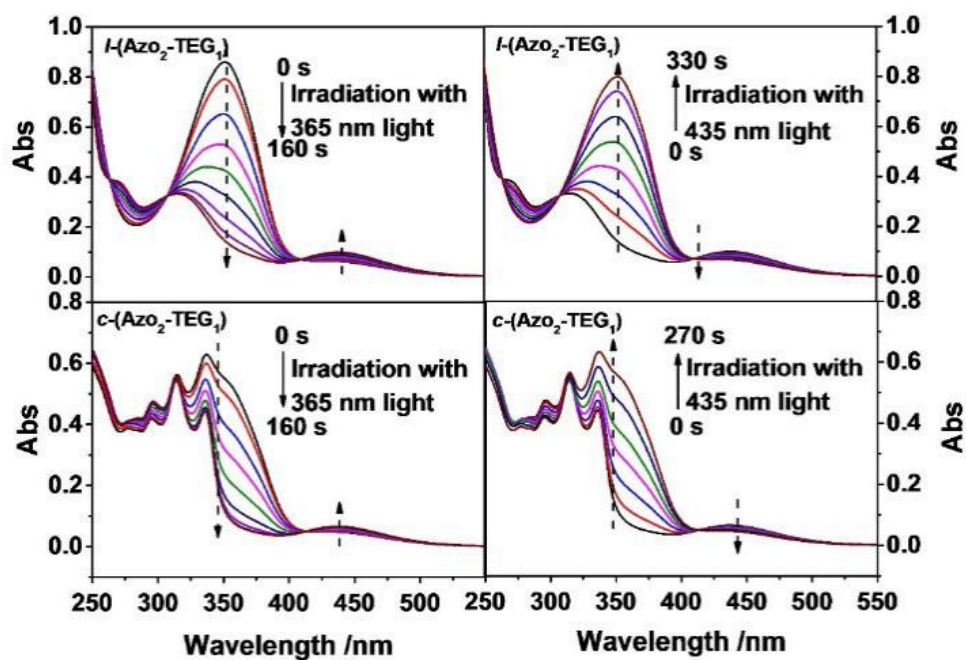


Figure S6. The UV-*vis* absorption spectra of *l*-(Azo₂-TEG₁) and *c*-(Azo₂-TEG₁) in DCM under irradiation with 365 nm UV light and 435 nm visible light at different time intervals until the photo-stationary were achieved, respectively. The concentration of solution is 1.5×10^{-3} mg/mL for both linear and cyclic polymers.

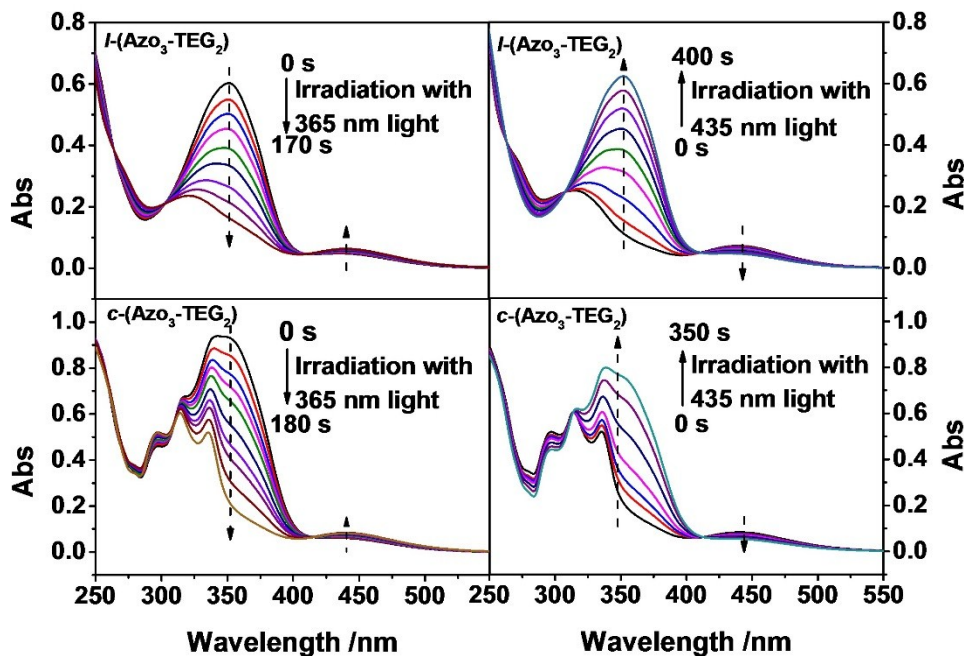


Figure S7. The UV-*vis* absorption spectra of *l*-(Azo₃-TEG₂) and *c*-(Azo₃-TEG₂) in DCM under irradiation with 365 nm UV light and 435 nm visible light at different time intervals until the photo-stationary were achieved, respectively. The concentration of solution is 1.78×10^{-3} mg/mL for both linear and cyclic polymers.

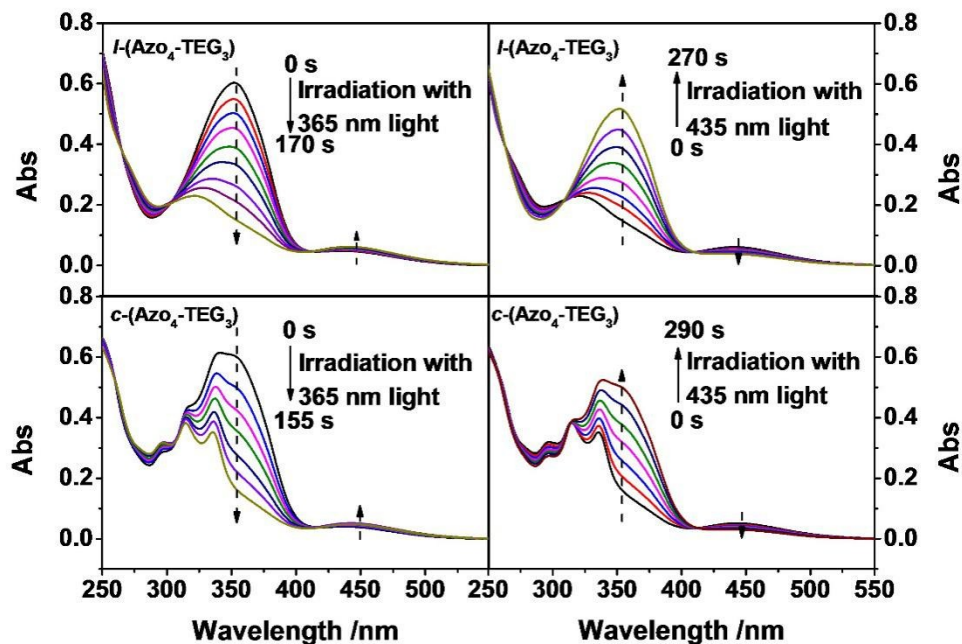


Figure S8. The UV-*vis* absorption spectra of *l*-(Azo₄-TEG₃) and *c*-(Azo₄-TEG₃) in DCM under irradiation with 365 nm UV light and 435 nm visible light at different time intervals until the photo-stationary state were achieved, respectively. The concentration of solution is 1.92×10^{-3} mg/mL for both linear and cyclic polymers.

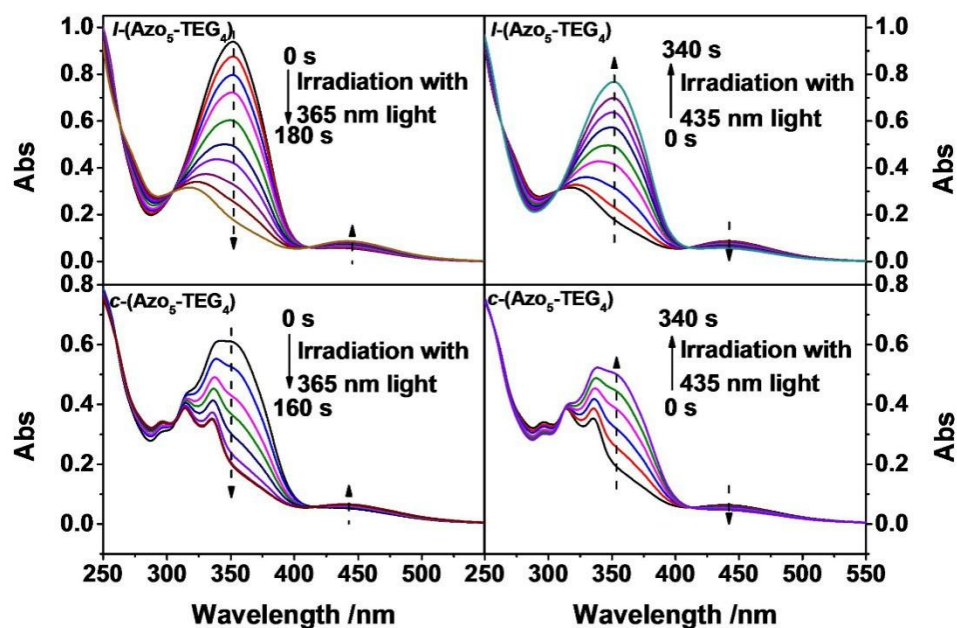


Figure S9. The UV-*vis* absorption spectra of *l*-(Azo₅-TEG₄) and *c*-(Azo₅-TEG₄) in DCM under irradiation with 365 nm UV light and 435 nm visible light at different time intervals until the photo-stationary state were achieved, respectively. The concentration of solution is 2.01×10^{-3} mg/mL for both linear and cyclic polymers.

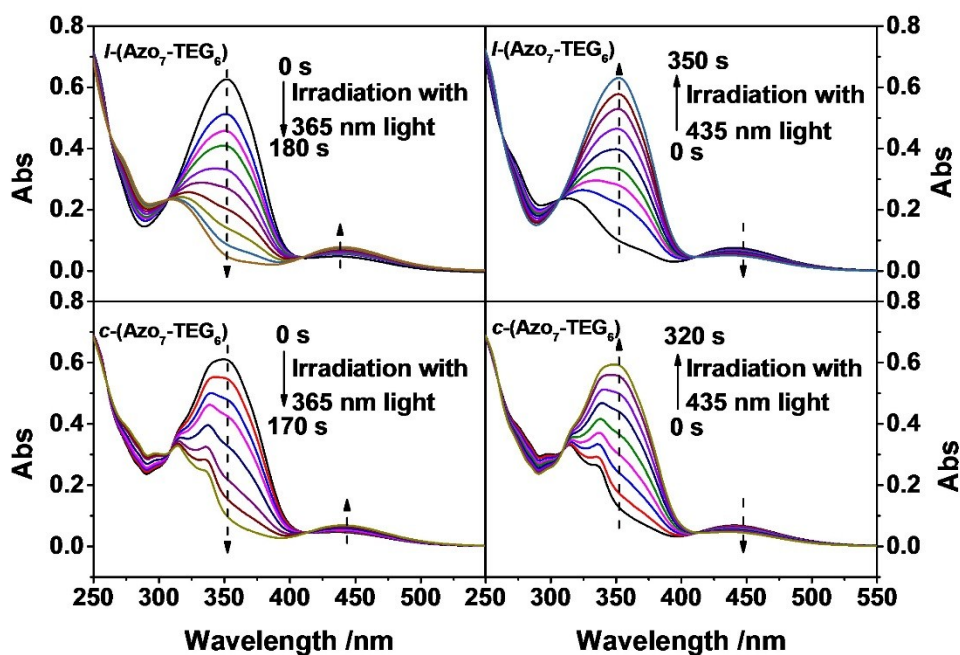


Figure S10. The UV-*vis* absorption spectra of *l*-(Azo₇-TEG₆) and *c*-(Azo₇-TEG₆) in DCM under irradiation with 365 nm UV light and 435 nm visible light at different time intervals until the photo-stationary were achieved, respectively. The concentration of solution is 2.46×10^{-3} mg/mL for both linear and cyclic polymers.

According to DFT and TDDFT calculations,¹ the HOMOs and LUMOs of *c*-(Azo_{n+1}-TEG_n) (n = 1-5) all predominately delocalize on the azobenzene (azo) and alkynyl (yne) group moieties.

As shown in Figure S11(A), the lowest energy band for the *c*-(Azo₂-TEG₁) is found around 472 nm mainly originated from HOMO-4→LUMO transitions, which can be assigned to $\pi^*_{\text{azo}} \rightarrow \pi^*_{\text{azo}}$ and $\pi^*_{\text{azo}} \rightarrow \pi^*_{\text{yne}}$ due to their orbital characters of corresponding starting and arriving states coming from the azobenzene moieties connected to 1, 3-diyne. For the *c*-(Azo₃-TEG₂) (Figures S11(B)) around 476 nm mainly originates from HOMO-7→LUMO transitions, which can also be assigned to $\pi^*_{\text{azo}} \rightarrow \pi^*_{\text{azo}}$ and $\pi^*_{\text{azo}} \rightarrow \pi^*_{\text{yne}}$ due to their orbital characters. However, for larger ring, the lowest energy band experimentally found for *c*-(Azo₄-TEG₃) (Figures S11(C)), *c*-(Azo₅-TEG₄) (Figures S11(D)), and *c*-(Azo₆-TEG₅) (Figures S11(E)) around 478 nm transitions are mainly assigned to $\pi^*_{\text{azo}} \rightarrow \pi^*_{\text{azo}}$, coming from the third azobenzene group moiety, which is totally different with those small ring complexes (*c*-(Azo₂-TEG₁) and *c*-(Azo₃-TEG₂)).

According to the calculations, in the UV-*vis* spectra, the absorption at 350 nm for complex $c\text{-(Azo}_2\text{-TEG}_1)$ may come from the HOMO-2 \rightarrow LUMO+1 transition ($\pi\rightarrow\pi^*$). The characteristic azobenzene and alkynyl of HOMO-2 and LUMO+1 indicate that the HOMO-2 \rightarrow LUMO+1 absorption transition possesses an intramolecular charge transfer (ICT) character. Similarly, the absorptions at about 370 nm of $c\text{-(Azo}_{n+1}\text{-TEG}_n)$ ($n = 2\text{-}5$) come from $\pi_{\text{azo (or yne)}}\rightarrow\pi^*_{\text{azo (or yne)}}$ character, probably with some mixing of a charge transfer character from the azobenzene and alkynyl ring.

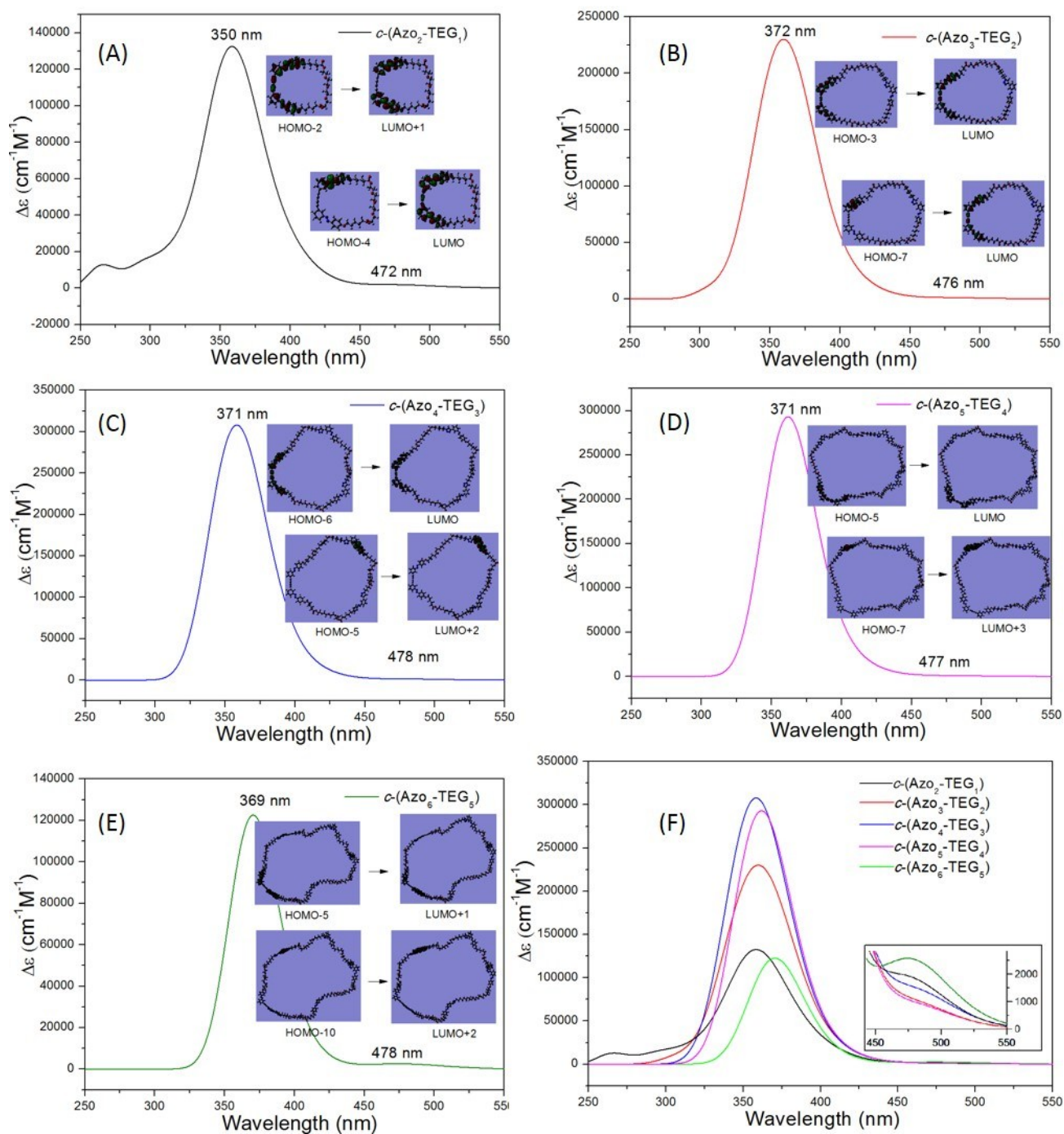


Figure S11. Calculated UV-*vis* spectra of the *c*-(Azo_{n+1}-TEG_n) (n = 1-5) at the level of B3LYP/6-31G*.

The first-order rate constant k_e of *trans*-to-*cis* photoisomerization was determined by the Formula S1:

$$\text{Ln}[(A_\infty - A_t)/(A_\infty - A_0)] = -k_e t \quad \text{Formula S1}$$

Where A_∞ , A_t , and A_0 are absorbance at about 350 nm corresponded to the π - π^* transition of *trans* isomers of azobenzene at infinite time, time t and time zero with irradiation of 365 nm UV light (0.5 mW cm⁻²) at room temperature, respectively.

The first-order rate constant k_H of *cis*-to-*trans* recovery was determined by the Formula S2:

$$\text{Ln}[(A_\infty - A_t)/(A_\infty - A_0)] = -k_H t \quad \text{Formula S2}$$

Where A_∞ , A_t , and A_0 are absorbance at about 350 nm corresponded to the π - π^* transition of *trans* isomers of azobenzene at infinite time, time t and time zero with irradiation of 435 nm visible light (0.53 mW cm⁻²) at room temperature, respectively.

The first-order kinetic curves of photo-isomerization are plotted in Figure S13

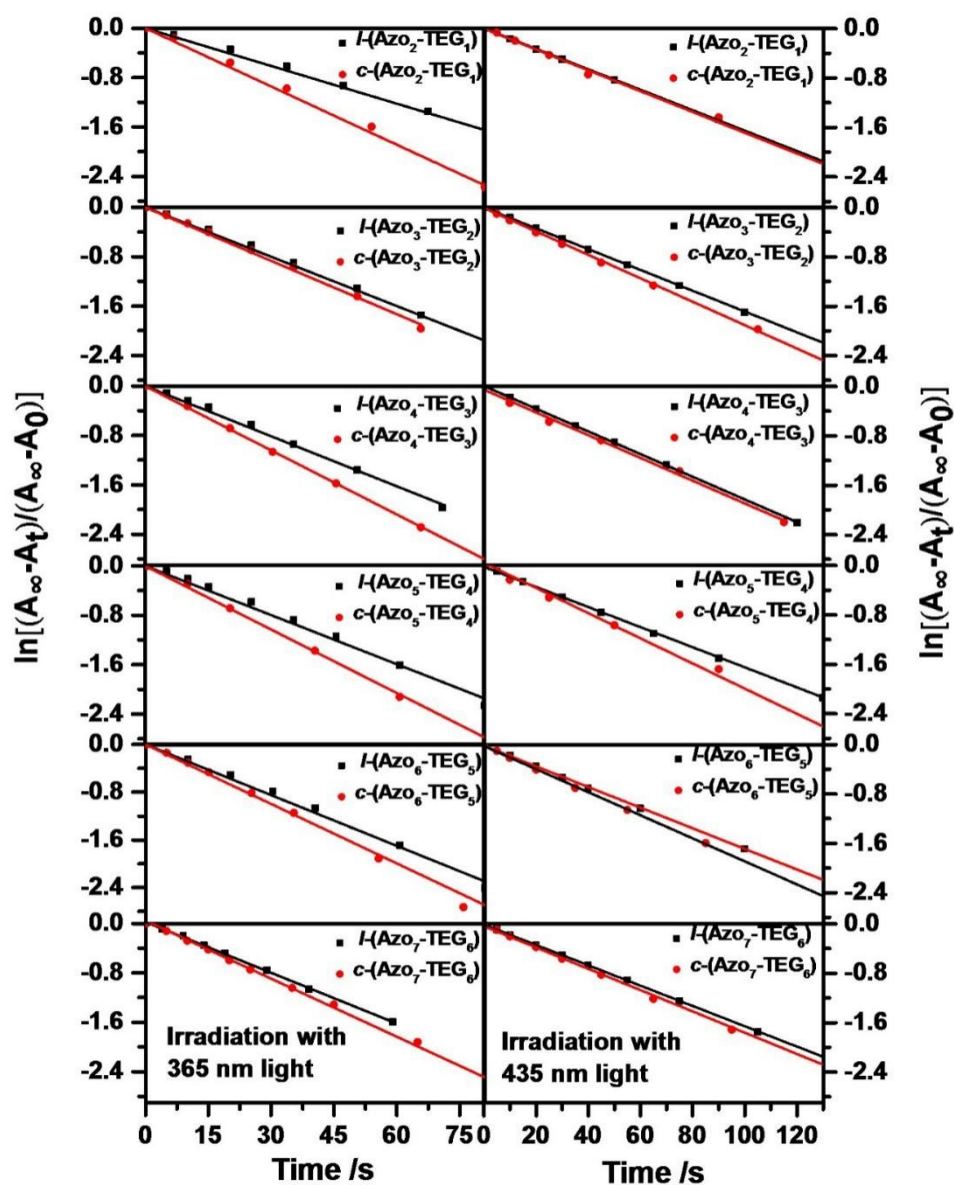


Figure S12. First-order kinetics for the photoisomerization of *trans*-to-*cis* and *cis*-to-*trans* recovery of *l*-(Azo_{*n*+1}-TEG_{*n*}) and *c*-(Azo_{*n*+1}-TEG_{*n*}) (*n* = 1-6) in DCM corresponding to Figure 3 and Figure S6-S10.

As we know, *trans* to *cis* isomerization of azobenzene moieties leads to new sets of resonances, and thus the resonances of the aromatic protons of the *cis*-azo isomers shift to higher field in ¹H NMR.² The characteristic signals of *cis*-azo isomers were confirmed from ¹H NMR spectra of *l*-(Azo_{*n*+1}-TEG_{*n*}) and *c*-(Azo_{*n*+1}-TEG_{*n*}) by contrasting their ¹H NMR spectra after UV light radiation (PSS_{uv} states). Therefore, the percentage of *cis*-azo isomers in the *l*-(Azo_{*n*+1}-TEG_{*n*}) (*F*_{*l-cis*}) and *c*-(Azo_{*n*+1}-TEG_{*n*}) (*F*_{*c-cis*}) could be calculated quantitatively according to Formula S3 and Formula S4.

$$F_{l-cis} = (1 - I_{l-trans}/I_{l-azo}) \times 100\% \quad \text{Formula S3}$$

$$F_{c-cis} = (1 - I_{c-trans}/I_{c-azo}) \times 100\% \quad \text{Formula S4}$$

Where I_{l-azo} and I_{c-azo} are the integrals of characteristic signals of *trans*-azo and *cis*-azo of l -(Azo_{n+1}-TEG_n) and c -(Azo_{n+1}-TEG_n) in the aromatic region, and I_{l-tran} and I_{c-tran} are the integrals of characteristic signals of *trans*-azo of l -(Azo_{n+1}-TEG_n) and c -(Azo_{n+1}-TEG_n) respectively.

For l -(Azo₂-TEG₁), I_{l-azo} is the integrals of characteristic signals of *trans*-azo and *cis*-azo around from 8.03 ppm to 6.65 ppm, and I_{l-tran} is the integrals of characteristic signals at 7.99 ppm, 7.95-7.81 ppm, 7.59-7.50 ppm, 7.44 ppm and 7.05-6.91 ppm respectively (Figure S13).

For c -(Azo₂-TEG₁), I_{c-azo} is the integrals of characteristic signals of *trans*-azo and *cis*-azo around from 8.19 ppm to 6.55 ppm, and $I_{c-trans}$ is the integrals of characteristic signals of *trans*-azo of c -(Azo₂-TEG₁) at 8.07 ppm, 7.96-7.87 ppm, 7.56-7.50 ppm 7.47 ppm and 7.05-6.97 ppm respectively (Figure S13).

The calculation of F_{l-cis} and F_{c-cis} of l -(Azo_{n+1}-TEG_n) and c -(Azo_{n+1}-TEG_n) ($n = 2-6$) is similar to the described method above. For l -(Azo_{n+1}-TEG_n), I_{l-azo} is the integrals of characteristic signals of *trans*-azo and *cis*-azo in the aromatic region around from 8.05 ppm to 6.57 ppm, and I_{l-tran} is the integrals of characteristic signals of *trans*-azo at 7.99 ppm, 7.96-7.80 ppm, 7.60-7.51 ppm, 7.44 ppm and 7.03-6.94 ppm respectively.

For l -(Azo_{n+1}-TEG_n), I_{c-azo} is the integrals of characteristic signals of *trans*-azo and *cis*-azo in the aromatic region around from 8.09 ppm to 6.57 ppm, and $I_{c-trans}$ is the integrals of characteristic signals at 8.08-7.95 ppm, 7.94-7.78 ppm, 7.62-7.51 ppm, 7.45 ppm and 7.06-6.88 ppm respectively.

Using l -(Azo₃-TEG₂) and c -(Azo₃-TEG₂) as typical samples, ¹H NMR spectra are as follow (Figure S14).

Unlike the l -(Azo₂-TEG₁) and c -(Azo₂-TEG₁) above, the aromatic protons of l -(Azo₃-TEG₂) and c -(Azo₃-TEG₂) appeared weak signals at higher field (6.5-6.9 ppm), which meant that a small amount of azobenzene moieties were assigned to *cis*-configuration.

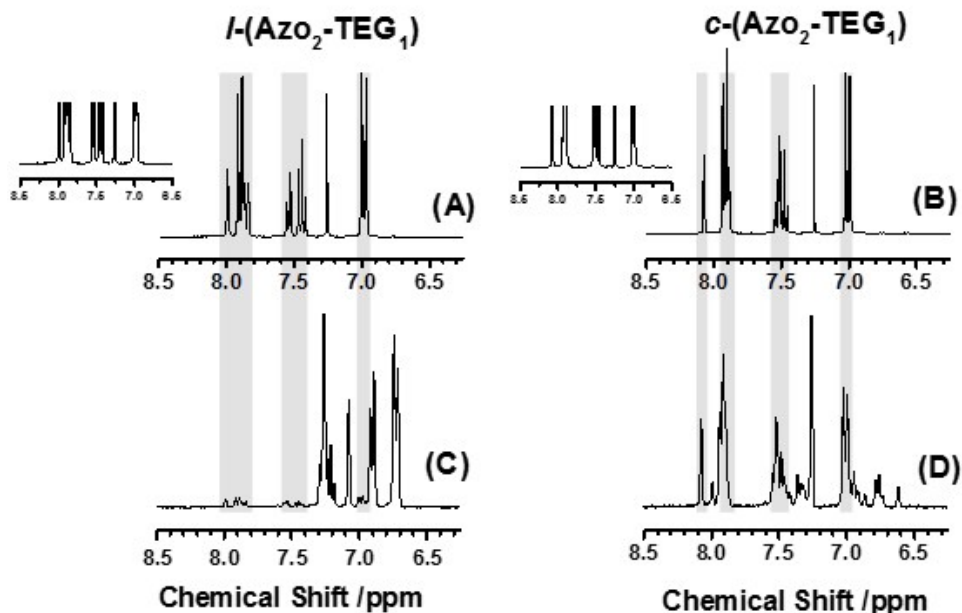


Figure S13. ^1H NMR spectra in the aromatic region of l -(Azo_2 - TEG_1) and c -(Azo_2 - TEG_1) in CDCl_3 at initial states (A), (B) and at the PSS_{UV} states (C), (D).

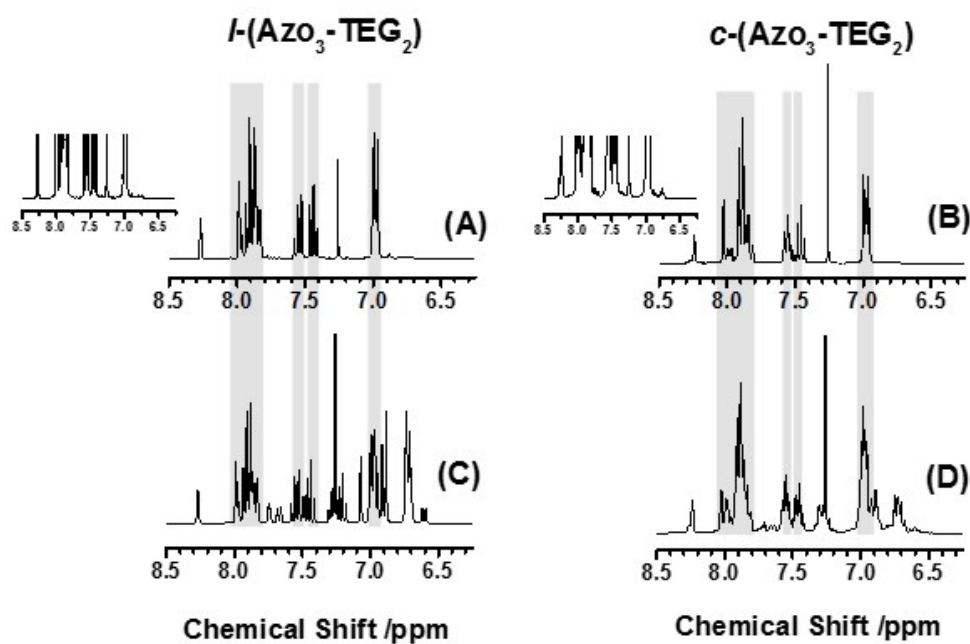


Figure S14. ^1H NMR spectra in the aromatic region of l -(Azo_3 - TEG_2) and c -(Azo_3 - TEG_2) in CDCl_3 at initial states (A), (B) and at the PSS_{UV} states (C), (D).

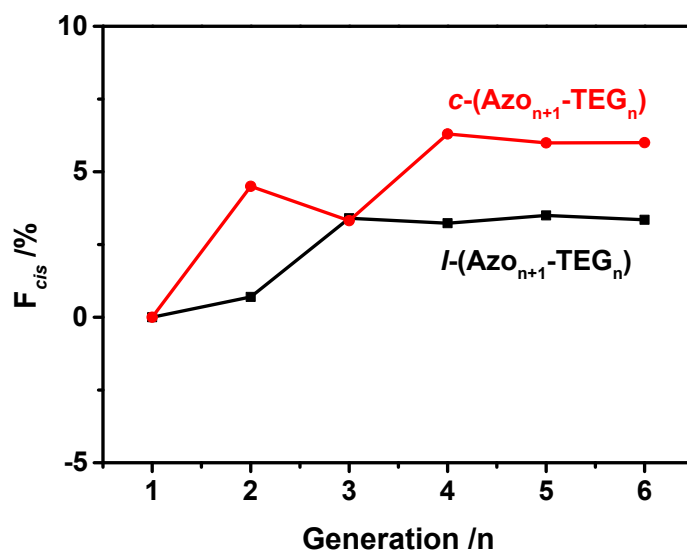


Figure S15. The evolution of *cis* isomers percentage estimated by NMR spectra with the generation (n) of $l-(Azo_{n+1}-TEG_n)$ and $c-(Azo_{n+1}-TEG_n)$ ($n=1-6$) in the initial states.

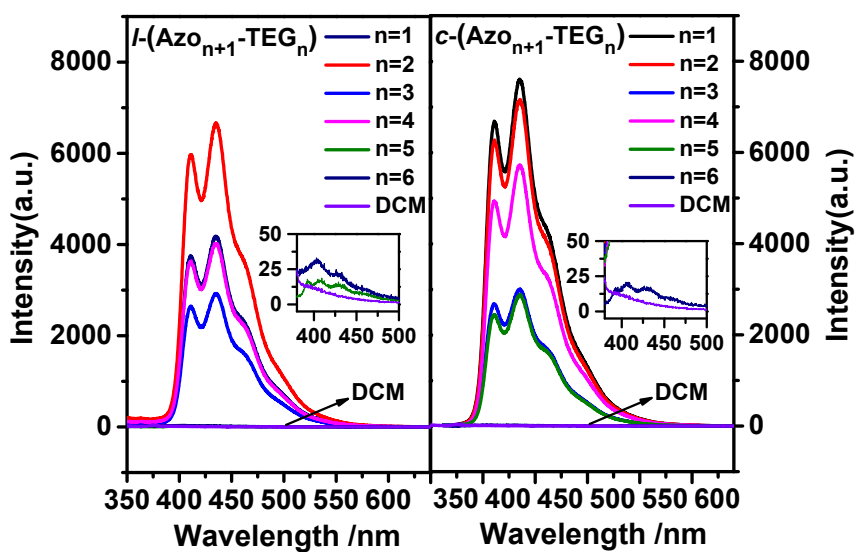


Figure S16. Fluorescence emission spectra (excitation wavelength at 350 nm) of $l-(Azo_{n+1}-TEG_n)$ and $c-(Azo_{n+1}-TEG_n)$ ($n = 1-6$) in DCM at room temperature. The concentration of solution is 3.6×10^{-8} mol/mL of azobenzene units for both linear and cyclic oligomers.

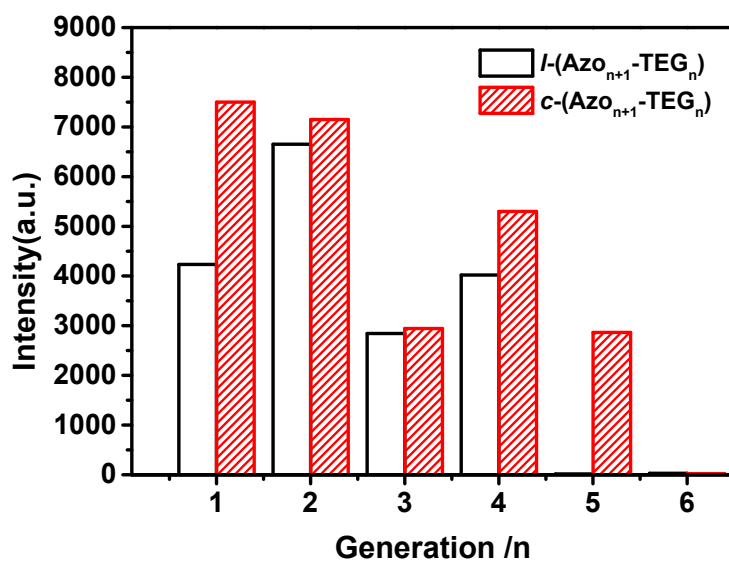


Figure S17. The evolution of fluorescence emission (excitation wavelength at 350 nm) of l -(Azo_{n+1}-TEG_n) and c -(Azo_{n+1}-TEG_n) ($n=1-6$) with generation (n) in DCM at room temperature.

Additional Tables

Table S1. Data corresponding to mass peaks in MALDI-TOF mass spectra (Figure 4) and molecular weights ($M_{n, \text{GPC}}$) determined by GPC of l -(Azo_{n+1}-TEG_n) and c -(Azo_{n+1}-TEG_n) (n = 1-6).

| Sample | Theor. [Da] | Obsed. [m/z] | $M_{n, \text{GPC}}$ [Da] |
|--|----------------|---------------------|-----------------------------|
| l -(Azo ₂ -TEG ₁) | 830.39 | 835.11 | 850 |
| c -(Azo ₂ -TEG ₁) | 828.37 | 850.52 | 420 |
| l -(Azo ₃ -TEG ₂) | 1481.72 | 1504.26 | 1900 |
| c -(Azo ₃ -TEG ₂) | 1479.70 | 1502.51 | 1300 |
| l -(Azo ₄ -TEG ₃) | 2133.04 | 2156.51 | 2700 |
| c -(Azo ₄ -TEG ₃) | 2131.03 | 2154.26 | 2100 |
| l -(Azo ₅ -TEG ₄) | 2784.37 | 2807.95 | 3600 |
| c -(Azo ₅ -TEG ₄) | 2782.35 | 2805.56 | 2800 |
| l -(Azo ₆ -TEG ₅) | 3435.70 | 3458.98 | 4800 |
| c -(Azo ₆ -TEG ₅) | 3433.68 | 3457.31 | 3500 |
| l -(Azo ₇ -TEG ₆) | 4087.03 | 4110.02 | 5900 |
| c -(Azo ₇ -TEG ₆) | 4085.01 | 4108.01 | 4300 |

Theor. calculated molecular mass (Da);

Theor. (l -(Azo_{n+1}-TEG_n)) (Da) = 830.39+ M_{mon} (651.33) \times (n-1)

Theor. (c -(Azo_{n+1}-TEG_n)) (Da) = 828.37+ M_{mon} (651.33) \times (n-1)

Obsed. (l -(Azo_{n+1}-TEG_n) and c -(Azo_{n+1}-TEG_n)): molecular mass obtained from MALDI-TOF mass spectra (m/z);

Calcd. (l -(Azo_{n+1}-TEG_n)) (Da) = 830.39+ M_{mon} (651.33) \times (n-1)+ M_{Na} (22.99);

Calcd. (c -(Azo_{n+1}-TEG_n)) (Da) = 828.37+ M_{mon} (651.33) \times (n-1)+ M_{Na} (22.99);

M_{Na} : the molecular mass of Na;

M_{mon} : the molecular mass of monomer;

n : the number of repeat units of monomer;

Table S2. Data of photoresponsive behavior of the *l*-(Azo_{n+1}-TEG_n) and *c*-(Azo_{n+1}-TEG_n) (n = 1-6)

| Sample | λ_{max} (trans) (nm) | λ_{max} (cis) (nm) | k_{e} s ⁻¹ | $k'_{\text{e}} / k_{\text{e}}$ | k_{H} s ⁻¹ | $k'_{\text{H}} / k_{\text{H}}$ |
|---|---|---|-----------------------------------|--------------------------------|-----------------------------------|--------------------------------|
| <i>l</i> -(Azo ₂ -TEG ₁) | 351.4 | 439.0 | 2.6 | 1.61 | 1.67 | 1.01 |
| <i>c</i> -(Azo ₂ -TEG ₁) | 338.5 | 439.0 | 4.2 | | 1.68 | |
| <i>l</i> -(Azo ₃ -TEG ₂) | 352.5 | 439.0 | 2.6 | 1.07 | 1.67 | 1.12 |
| <i>c</i> -(Azo ₃ -TEG ₂) | 343.0 | 439.0 | 3.0 | | 1.87 | |
| <i>l</i> -(Azo ₄ -TEG ₃) | 351.5 | 439.0 | 2.7 | 1.21 | 1.82 | 1.05 |
| <i>c</i> -(Azo ₄ -TEG ₃) | 341.3 | 439.0 | 3.3 | | 1.91 | |
| <i>l</i> -(Azo ₅ -TEG ₄) | 352.0 | 439.0 | 2.4 | 1.41 | 1.63 | 1.20 |
| <i>c</i> -(Azo ₅ -TEG ₄) | 342.2 | 439.0 | 3.4 | | 1.96 | |
| <i>l</i> -(Azo ₆ -TEG ₅) | 352.3 | 439.0 | 2.8 | 1.23 | 1.68 | 1.14 |
| <i>c</i> -(Azo ₆ -TEG ₅) | 350.6 | 439.0 | 3.5 | | 1.88 | |
| <i>l</i> -(Azo ₇ -TEG ₆) | 352.5 | 439.0 | 2.6 | 1.15 | 1.65 | 1.07 |
| <i>c</i> -(Azo ₇ -TEG ₆) | 351.3 | 439.0 | 3.1 | | 1.77 | |

The quantum yield was calculated using the following equation:

$$\Phi_s = \Phi_r [F_s A_r / F_r A_s] (n_r / n_s)^2$$

where Φ_s is the fluorescence quantum yield of the sample, F is the area of the emission peak, n is the refractive index of solution, and A is the absorbance of the solution at the exciting wavelength. The subscripts r and s denote reference and sample, respectively.³

Table S3. The quantum yields (Φ_s) of the l -(Azo_{n+1}-TEG_n) and c -(Azo_{n+1}-TEG_n) ($n = 1-6$).

| Sample | Φ_s (10 ⁻²) | Sample | Φ_s (10 ⁻²) |
|--|---------------------------------|--|---------------------------------|
| l -(Azo ₂ -TEG ₁) | 0.984 | c -(Azo ₂ -TEG ₁) | 2.055 |
| l -(Azo ₃ -TEG ₂) | 1.695 | c -(Azo ₃ -TEG ₂) | 2.007 |
| l -(Azo ₄ -TEG ₃) | 0.805 | c -(Azo ₄ -TEG ₃) | 0.837 |
| l -(Azo ₅ -TEG ₄) | 0.848 | c -(Azo ₅ -TEG ₄) | 1.498 |
| l -(Azo ₆ -TEG ₅) | 0.004 | c -(Azo ₆ -TEG ₅) | 0.917 |
| l -(Azo ₇ -TEG ₆) | 0.009 | c -(Azo ₇ -TEG ₆) | 0.005 |

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