

Electronic Supplementary Information

A comparison of two classic Pb^{2+} -dependent RNA-cleaving DNazymes

Runjhun Saran and Juewen Liu*

Department of Chemistry, Waterloo Institute for Nanotechnology

Waterloo, Ontario, Canada N2L 3G1

Email: liujw@uwaterloo.ca

Table S1. DNA sequences used in the paper.

A=Adenine; C=Cytosine; T=Thymine; G=Guanine; 2AP=2-AminoPurine; HX=Hypoxanthine

DNA	SEQUENCE (5' - 3')
Substrate	
WT	GTCACGAGTCACTAT rAG GAAGATGGCGAAA
rAA	GTCACGAGTCACTAT rAA GAAGATGGCGAAA
rA2AP	GTCACGAGTCACTAT rA2AP GAAGATGGCGAAA
rAHX	GTCACGAGTCACTAT rAHX GAAGATGGCGAAA
GR5 DNAzyme	
WT	TTTCGCCATCT-- GAAGTAGCGCCGCCG TATAGTGACTCGTGAC
A6C	TTTCGCCATCT-- GAAGTCGCGCCGCCG TATAGTGACTCGTGAC
A6T	TTTCGCCATCT-- GAAGTTGCGCCGCCG TATAGTGACTCGTGAC
A6G	TTTCGCCATCT-- GAAGTGGCGCCGCCG TATAGTGACTCGTGAC
G7A	TTTCGCCATCT-- GAAGTACGCGCCGCCG TATAGTGACTCGTGAC
G7C	TTTCGCCATCT-- GAAGTACCGCGCCGCCG TATAGTGACTCGTGAC
G7T	TTTCGCCATCT-- GAAGTATCGCGCCGCCG TATAGTGACTCGTGAC
C8A	TTTCGCCATCT-- GAAGTAGAGCCGCCG TATAGTGACTCGTGAC
C5G	TTTCGCCATCT-- GAAGTAGGGCCGCCG TATAGTGACTCGTGAC
C8T	TTTCGCCATCT-- GAAGTAGTGCCGCCG TATAGTGACTCGTGAC
G9A	TTTCGCCATCT-- GAAGTAGCACCGCCG TATAGTGACTCGTGAC
G9C	TTTCGCCATCT-- GAAGTAGCCCGCCG TATAGTGACTCGTGAC
G9T	TTTCGCCATCT-- GAAGTAGCTCCGCCG TATAGTGACTCGTGAC
C14A	TTTCGCCATCT-- GAAGTAGCGCCGCAG TATAGTGACTCGTGAC
C14G	TTTCGCCATCT-- GAAGTAGCGCCGCGG TATAGTGACTCGTGAC
C14T	TTTCGCCATCT-- GAAGTAGCGCCGCTG TATAGTGACTCGTGAC
G15A	TTTCGCCATCT-- GAAGTAGCGCCGCCA TATAGTGACTCGTGAC
G15C	TTTCGCCATCT-- GAAGTAGCGCCGCCC TATAGTGACTCGTGAC
G15T	TTTCGCCATCT-- GAAGTAGCGCCGCCCT TATAGTGACTCGTGAC
1=T	TTTCGCCATCT T -- GAAGTAGCGCCGCCG TATAGTGACTCGTGAC
1=TC	TTTCGCCATCT TC -- GAAGTAGCGCCGCCG TATAGTGACTCGTGAC
2=AAG	TTTCGCCATCT-- AAGTAGCGCCGCCG TATAGTGACTCGTGAC
2=AG	TTTCGCCATCT-- AGTAGCGCCGCCG TATAGTGACTCGTGAC
2=G	TTTCGCCATCT-- GTAGCGCCGCCG TATAGTGACTCGTGAC
2=none	TTTCGCCATCT-- TAGCGCCGCCG TATAGTGACTCGTGAC
3=A	TTTCGCCATCT-- GAAGAAGCGCCGCCG TATAGTGACTCGTGAC
3=C	TTTCGCCATCT-- GAAGCAGCGCCGCCG TATAGTGACTCGTGAC
3=G	TTTCGCCATCT-- GAAGGAGCGCCGCCG TATAGTGACTCGTGAC
17E DNAzyme	
WT	TTTCGCCATCTTTCTCCGAGCCGGT CGAA ATAGTGACTCGTGAC
A6C	TTTCGCCATCTTTCTCCG CG CCGGT CGAA ATAGTGACTCGTGAC
A6G	TTTCGCCATCTTTCTCCG GG CCGGT CGAA ATAGTGACTCGTGAC
A6T	TTTCGCCATCTTTCTCCG TG CCGGT CGAA ATAGTGACTCGTGAC
G7A	TTTCGCCATCTTTCTCCG AAC GGT CGAA ATAGTGACTCGTGAC
G7C	TTTCGCCATCTTTCTCCG ACC GGT CGAA ATAGTGACTCGTGAC
G7T	TTTCGCCATCTTTCTCCG ATC GGT CGAA ATAGTGACTCGTGAC

C13A	TTTCGCCATCTTTCTCCGAGCCGGT A GAAATAGTGACTCGTGAC
C13G	TTTCGCCATCTTTCTCCGAGCCGGT G GAAATAGTGACTCGTGAC
C13T	TTTCGCCATCTTTCTCCGAGCCGGT T GAAATAGTGACTCGTGAC
G14A	TTTCGCCATCTTTCTCCGAGCCGGT C AAATAGTGACTCGTGAC
G14C	TTTCGCCATCTTTCTCCGAGCCGGT C AAATAGTGACTCGTGAC
G14T	TTTCGCCATCTTTCTCCGAGCCGGT T AAATAGTGACTCGTGAC

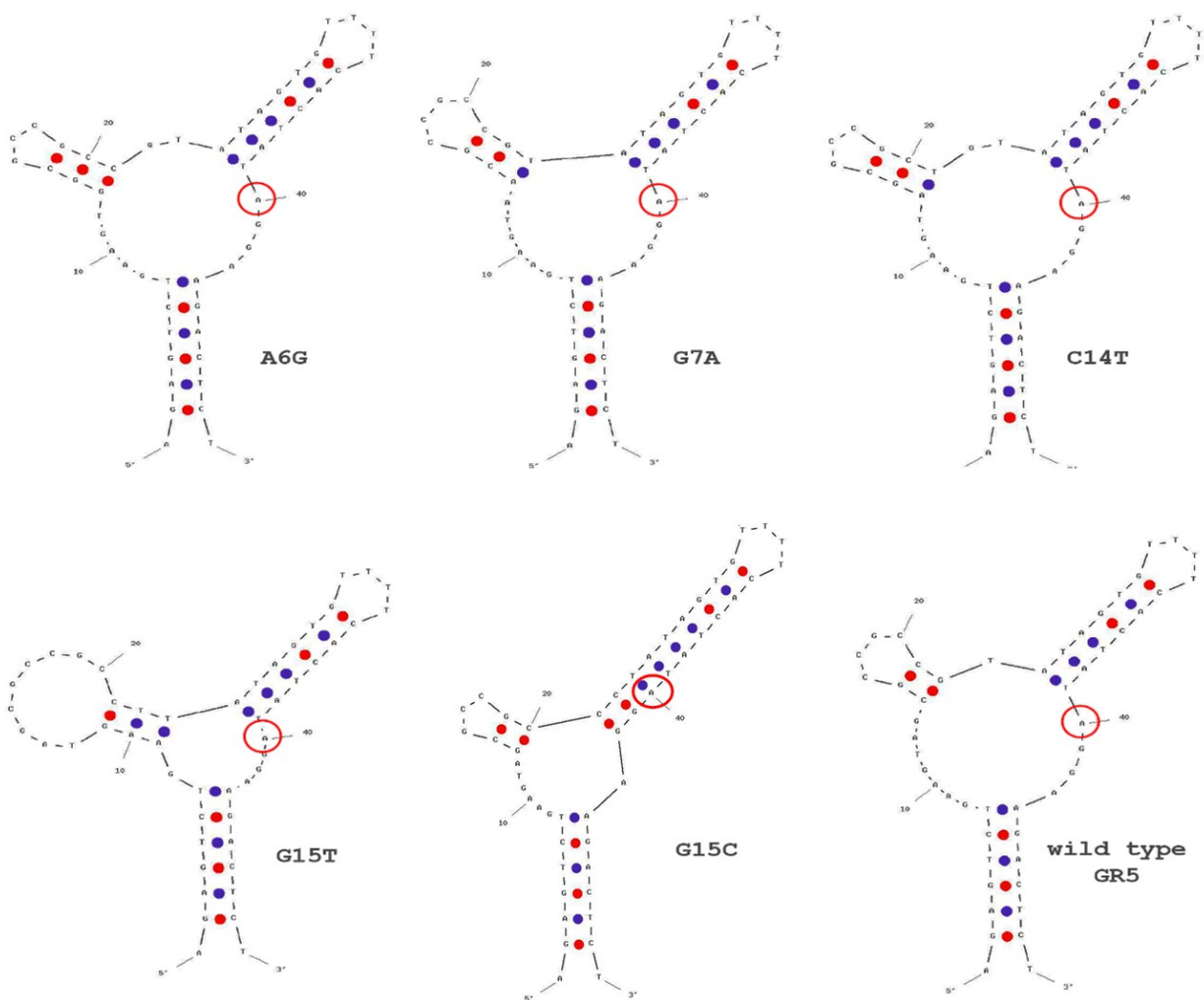


Figure S1. Mfold predicted secondary structures from a few GR5 mutants and the wild-type GR5. All the other mutants fold into a structure similar to that for the wild-type GR5 and are not listed here. The cleavage site adenine is marked by the red circle. The substrate and enzyme strands are linked by a TTTT loop. In the wild-type GR5 structure, two G-C base pairs are predicted in the enzyme loop. However, these nucleotides are highly conserved for catalysis and thus are unlikely

to be confined in such a hairpin structure. Therefore, we still used the simple loop structure in the original GR5 selection paper (Breaker, R. R.; Joyce, G. F. *Chem. Biol.* **1994**, *1*, 223) in this work. The mutants in this figure have three base pairs in the loop and their misfold (i.e. deviation from the simple loop structure) might also be a reason for their inhibited activity.