

Electronic Supplementary Information

Efficient synthesis of 2-nitroimidazole derivatives and the bio-reductive clinical candidate Evofosfamide (TH-302).

Liam J. O'Connor,^{a,b} Cindy Cazares-Körner,^{a,b} Jaideep Saha,^a Charles N. G. Evans,^{a,b} Michael R. L. Stratford,^b Ester M. Hammond^b and Stuart J. Conway^{*a}

^aDepartment of Chemistry, Chemistry Research Laboratory, University of Oxford, Mansfield Road, Oxford, OX1 3TA, UK.

^bCancer Research UK/MRC Gray Institute for Radiation Oncology and Biology, Department of Oncology, University of Oxford, Old Road Campus Research Building, Oxford, OX3 7DQ, UK.

*To whom correspondence should be addressed: stuart.conway@chem.ox.ac.uk, telephone: +44 (0)1865 285 109, fax: +44 (0)1865 285 002.

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General experimental procedures

¹H NMR spectra were recorded on Bruker AVC500, AVX500, DRX500 (500 MHz) or Bruker AV600 (600 MHz) using deuteriochloroform (unless indicated otherwise) as a reference for the internal deuterium lock. The chemical shift data for each signal are given as δ_{H} in units of parts per million (ppm) relative to tetramethylsilane (TMS) where δ_{H} (TMS) = 0.00 ppm. The multiplicity of each signal is indicated by s (singlet); d (doublet); t (triplet); q (quartet); dd (doublet of doublets); or m (multiplet). The number of protons (n) for a given resonance signal is indicated by nH. Coupling constants (*J*) are quoted in Hz and are recorded to the nearest 0.1 Hz. Identical proton coupling constants (*J*) are averaged in each spectrum and reported to the nearest 0.1 Hz. The coupling constants are determined by analysis using Bruker TopSpin software. Spectra were assigned using COSY, HSQC and HMBC experiments as necessary.

¹³C NMR spectra were recorded on Bruker DRX500, AVC500, AVX600 or AV600 (126 MHz) spectrometers in the stated solvents, with broadband proton decoupling and an internal deuterium lock. The chemical shift data for each signal are given as δ_{C} in units of parts per million (ppm) relative to tetramethylsilane (TMS) where δ_{C} (TMS) = 0.00 ppm. The shift values of resonances are quoted to 1 decimal place. The multiplicity of each signal is singlet unless indicated by: d (doublet). Where appropriate, coupling constants (*J*) are quoted to the nearest 0.1 Hz, and were determined using Bruker TopSpin software.

³¹P NMR spectra were recorded on a Bruker AVB400 (162 MHz) or AVX500 (202 MHz) in the stated solvents as a reference for the internal deuterium lock with broadband proton decoupling. The chemical shift data for each signal are given as δ_{P} in units of parts per million (ppm). Signals are quoted as proton decoupled singlets.

Mass spectra were acquired on a VG platform spectrometer and an Agilent 6120 spectrometer (low resolution). Electro-spray ionisation spectra were obtained on Micromass LCT Premier and Bruker MicroTOF spectrometers, operating in positive or negative mode as

indicated, from solutions of MeOH or MeCN. m/z values are reported in Daltons and followed by their percentage abundance in parentheses.

Melting points were determined using a Leica Galen III hot stage microscope and are uncorrected.

Infrared spectra were obtained from neat samples, either as solids or liquids using a diamond ATR module. The spectra were recorded on a Bruker Tensor 27 spectrometer. Absorption maxima are recorded in wavenumbers (cm^{-1}), and reported as s (strong), m (medium), w (weak) or br (broad).

Analytical thin layer chromatography (TLC) was carried out on normal phase Merck silica gel 60 F₂₅₄ aluminium-supported chromatography sheets. Visualisation was by absorption of UV light (λ_{max} 254 and 365 nm).

Flash column chromatography was performed manually using VWR Prolabo silica gel 60 (240-400 mesh) under a positive pressure of nitrogen.

Semi-preparative HPLC was carried out on a Waters system (2695 pump/autosampler, 2996 diode array detector and ZQ2000 mass spectrometer). The column was a Phenomenex Luna C18(2) 10 μm column, 250 x 10 mm, eluents water (A) and methanol (B), with a gradient from 50 – 70% B over 5 min, flow rate 5 ml/min.

In vacuo refers to removal of solvent on a Buchi® rotary evaporator under reduced pressure in a water bath at 40 °C.

Petroleum ether refers to the fraction in the boiling point range 40–60 °C.

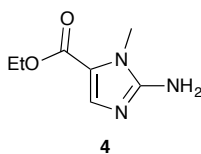
Chemicals were purchased from Sigma Aldrich UK and Alfa Aesar UK, and were used as supplied unless otherwise stated.

Compound purity for compounds was determined by elemental analysis or HPLC. Elemental analysis was obtained at the Elemental Analysis Service, London Metropolitan University, London. Elemental analysis was carried out in duplicate; average values are

reported in Supporting Information. For all tested compounds, experimentally determined hydrogen, carbon, and nitrogen composition was within 0.4% of the expected value, implying a purity of >95%.

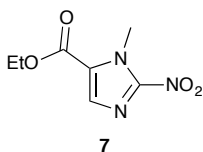
HPLC was carried out on a Waters system (2695 pump/autosampler, 2996 diode array detector and ZQ2000 mass spectrometer). The column was a Hichrom RPB, 5 μ m, 100 x 3.2 mm, and the eluents were 10 mM formic acid (A), and acetonitrile (B), with a gradient of 35 – 95% B in 5 min, flow rate 0.5 mL/min.

Ethyl 2-amino-1-methyl-1*H*-imidazole-5-carboxylate (**4**)



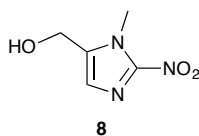
To a suspension of sarcosine ethyl ester hydrochloride (**5**, 2 g, 0.013 mol, 1 eq.) in ethyl formate (45 mL) and THF (45 mL), was added NaH (60% dispersion in mineral oil, 5 g, 0.13 mol, 10 eq.) slowly at ambient temperature, and allowed to stir for 3 h. After this time a yellow suspension had formed, and the mixture was concentrated and dried *in vacuo*. The resulting solid was triturated with hexane (2 × 75 mL), the hexane decanted, and the remaining solid dried *in vacuo*. EtOH (40 mL) and concentrated aqueous HCl (8 mL) were added to the solid, and the suspension heated under reflux for 2 h. The reaction mixture was filtered while hot, and the resulting colourless solid washed with boiling EtOH (2 × 30 mL). The filtrate was concentrated *in vacuo* to leave an aqueous solution, which was diluted with EtOH (70 mL) and distilled water (30 mL). The pH of the solution was adjusted to 3, using an aqueous 2 M solution of NaOH, and cyanamide (1.09 g, 0.026 mol, 2 eq.) was added. The resulting solution was heated under reflux for 1.5 h. After this time the solution was cooled to ambient temperature, and concentrated *in vacuo* to approximately $\frac{1}{8}$ of the original volume. The pH was then adjusted to 8-9 using solid K_2CO_3 , resulting in the formation of a precipitate. The solid was removed by filtration, washed with aqueous K_2CO_3 solution (1 M, 1 × 20 mL), H_2O (2 × 10 mL), and dried *in vacuo*, to afford **4** as a pale yellow solid (1.10 g, 50%): (found C, 35.2; H, 2.8; N 24.4. $C_7H_{11}N_3O_2$ requires C, 35.1; H, 3.0; N, 24.6); R_f 0.42 (CH_2Cl_2 - MeOH, 9:1); mp 130-133 °C (from H_2O); ν_{max} (solid)/ cm^{-1} 3126 (w), 1648 (s), 1246 (m), 1167 (s); δ_H ($CDCl_3$, 500 MHz) 7.44 (1H, s, CH), 4.41 (2H, s, NH_2), 4.27 (2H, q, J 7.1, CH_2CH_3), 3.68 (3H, s, CH_3), 1.34 (3H, t, J 7.1, CH_2CH_3); δ_C ($CDCl_3$, 126 MHz) 160.7, 152.1, 135.5, 119.1, 59.9, 30.6, 14.5; HRMS m/z (ESI⁺) [found (M+H)⁺ 170.0919, $C_7H_{11}N_3O_2$ requires M⁺ 170.0924]; m/z (ES⁺) 170.1 ([M+H]⁺, 100 %). These data are in good agreement with the literature values.¹

Ethyl 1-methyl-2-nitro-1H-imidazole-5-carboxylate (**7**)



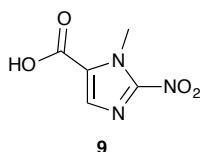
To an aqueous solution of sodium nitrite (5 mL, 3.95 g, 57.24 mmol, 10 eq.), was added aminoimidazole **4** (0.97 g, 5.72 mmol) in acetic acid (10 mL) in a drop wise manner. The solution was stirred at room temperature for 4 h, after this time no more nitrogen gas was evolved. The organic mixture was extracted with CH₂Cl₂ (1 × 20 mL), washed with brine (1 × 20 mL) and a saturated aqueous solution of Na₂SO₃ (1 × 20 mL), dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo*, the residue adsorbed onto Celite®, and the product purified by silica gel column chromatography, eluting with CH₂Cl₂, to afford **7** (826 mg, 72 %) as a yellow solid: (found C, 42.3; H, 4.4; N, 21.0. C₇H₉N₃O₄ requires C, 42.2; H, 4.6; N, 21.1); *R*_f 0.85 (CH₂Cl₂ - MeOH, 9:1); mp 56-58 °C (from CH₂Cl₂) [lit.² mp 65-66 °C]; *v*_{max} (solid)/cm⁻¹ 2983, 1723, 1519, 1281, 1234; δ_H (CDCl₃, 500 MHz) 7.76 (1H, s, *CH*), 4.40 (2H, q, *J* 7.3, CH₂CH₃), 4.35 (3H, s, CH₃), 1.41 (3H, t, *J* 7.3, CH₂CH₃); δ_C (CDCl₃, 126 MHz) 159.1, 134.6, 126.3, 61.8, 35.4, 14.1; HRMS *m/z* (ESI⁻) [found (M+Na)⁺ 222.0480, C₇H₉N₃NaO₄ requires M⁺ 222.0485]; *m/z* (ES⁺) 222.1 ([M+Na]⁺, 100 %). These data are in good agreement with the literature values.²

(1-Methyl-2-nitro-1*H*-imidazol-5-yl)methanol (8**)**



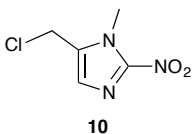
To a solution of nitroimidazole **7** (230 mg, 1.16 mmol, 1 eq.) in anhydrous THF (6 mL) and MeOH (0.5 mL) at 0 °C, was added sodium borohydride (131 mg, 3.47 mmol, 3 eq.) portion wise. The reaction mixture was stirred at 0 °C for 45 min, and then at ambient temperature for 1 h. The reaction mixture was cooled to 0 °C, quenched by addition of ice and the pH adjusted to 7 using 1 M aqueous HCl. The aqueous mixture was saturated with solid NaCl, and the organic components extracted with EtOAc (5 × 15 mL). The organic layers were combined, washed with saturated aqueous NaHCO₃, dried over MgSO₄ and filtered. The filtrate was concentrated *in vacuo*, the residue adsorbed onto Celite®, and the product purified by silica gel column chromatography, eluting with petroleum ether and EtOAc (gradient; 50-100% EtOAc) to afford **8** as pale yellow crystals (109 mg, 66%): (found C, 38.2; H, 4.5; N 26.6. C₅H₇N₃O₃ requires C, 38.2; H, 4.5; N, 26.7); *R*_f 0.5 (CH₂Cl₂ - MeOH, 9:1); mp 141-143 °C (from EtOAc) [lit.² mp 142-144 °C]; ν_{max} (solid)/cm⁻¹ 3228 (br), 1490 (s), 1396 (s), 1038 (s); δ_{H} (DMSO-D₆, 400 MHz) 7.12 (1H, s, *CH*), 5.50 (1H, t, *J* 5.4, *OH*), 4.54 (2H, d, *J* 5.4, *CH*₂), 3.92 (3H, s, *CH*₃); δ_{C} (DMSO-D₆, 126 MHz) 146.7, 138.6, 126.5, 53.0, 34.1; HRMS *m/z* (ES⁺) [found (M-H)⁺ 156.0412, C₅H₆N₃O₃ requires M⁺ 156.0414]; *m/z* (ESI⁻) 156.04 ([M-H]⁻, 100%). These data are in good agreement with the literature values.²

1-Methyl-2-nitro-1*H*-imidazole-5-carboxylic acid (**9**)



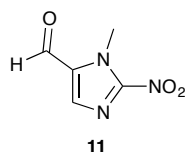
To nitroimidazole **7** (100 mg, 0.50 mmol, 1 eq.), was added a 0.75 M aqueous solution of NaOH (2 mL), and the resulting suspension was stirred for 2 h at room temperature, after which time a homogenous solution had formed. The reaction solution was acidified to pH 1 using conc. HCl, resulting in the formation of an off-white precipitate. The organic components were extracted with EtOAc (5 × 15 mL), the organic layers combined, dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo*, to afford **9** as an off-white solid (82 mg, 95 %): (found C, 35.2; H, 2.8; N 24.4. C₅H₅N₃O₄ requires C, 35.1; H, 3.0; N, 24.6); *R*_f 0.15 (AcOH-CH₂Cl₂-MeOH, 0.1:9:1); mp 155-157 °C (from CHCl₃) [lit.² mp 161-163 °C]; ν_{max} (solid) cm⁻¹: 2924 (br), 1712 (s), 1497 (s), 1362 (s), 1236 (s); δ_{H} (MeOD-D₄, 500 MHz) 7.73 (1H, s, *CH*), 4.33 (3H, s, *CH*₃); δ_{C} (MeOD-D₄, 126 MHz) 161.6, 149.0, 134.8, 128.5, 35.8; HRMS *m/z* (ESI⁻) [found; (M-H)⁻ 170.0207, C₅H₄N₃O₄ requires M⁻ 170.0207]; *m/z* (ESI⁻) 170.02 ([M-H]⁻, 68%). These data are in good agreement with the literature values.²

5-(Chloromethyl)-1-methyl-2-nitro-1*H*-imidazole (**10**)



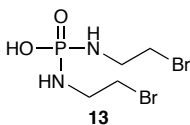
To a solution of nitroimidazole **8** (200 mg, 1.27 mmol, 1 eq.) in dry CH₂Cl₂ (8 mL) and distilled pyridine (0.30 mL, 3.63 mmol, 2.9 eq.), was added SOCl₂ (0.28 mL, 3.90 mmol, 3 eq.) in dry CH₂Cl₂ (2 mL) drop wise at 0 °C. The reaction solution was stirred at 0 °C for 1 h, followed by 1 h at room temperature. The excess SOCl₂ was quenched by addition of ice, and saturated aqueous NaHCO₃ solution (5 mL) was added. The organic mixture was extracted with CH₂Cl₂ (2 × 15 mL), the organic layers combined, washed with brine (30 mL), dried over MgSO₄, and filtered. The filtrate was concentrated *in vacuo*, the residue adsorbed onto Celite[®], and the product purified by silica gel column chromatography, eluting with petroleum ether and EtOAc (gradient; 50-100% EtOAc) to afford **10** as pale yellow crystals (147 mg, 66%): (found C, 34.3; H, 3.5; N 23.8. C₅H₆ClN₃O₂ requires C, 34.2; H, 3.4; N, 23.9); *R*_f 0.4 (petroleum ether - EtOAc, 1:1); mp 87-90 °C (from EtOAc) [lit.³ mp 94-96 °C]; ν_{\max} (solid) cm⁻¹: 1483 (s), 1360 (s); δ_{H} (CDCl₃, 500 MHz) 7.17 (1H, s, CH), 4.63 (2H, s, CH₂), 4.06 (3H, s, CH₃); δ_{C} (CDCl₃, 126 MHz) 146.3, 132.9, 128.5, 34.2, 33.9; HRMS *m/z* (ESI⁺) [found (M+Na)⁺ 198.0041, C₅H₆ClN₃NaO₂ requires M⁺ 198.0041]; *m/z* (ESI⁺) 198.1 ([M+Na]⁺, 100%). These data are in good agreement with the literature values.³

1-Methyl-2-nitro-1*H*-imidazole-5-carbaldehyde (**11**)



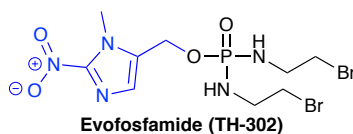
To a solution of nitroimidazole **8** (50 mg, 0.32 mmol, 1 eq.) in acetone (0.3 mL) and CH₂Cl₂ (2.7 mL), was added manganese(IV) dioxide (157 mg, 1.8 mmol, 6 eq.), and the reaction solution was stirred at ambient temperature for 3 d, under an argon atmosphere. The reaction suspension was filtered through Celite[®], and washed with CH₂Cl₂ (10 mL). The elutant was adsorbed onto Celite[®], and the product purified using silica gel column chromatography eluting with acetone and petroleum ether (1:4), to afford the carbaldehyde **11** as a colourless solid (25 mg, 52%): *R_f* 0.72 (acetone - petroleum ether, 1:1); mp 113-115 °C (from acetone) [lit.² mp 114-115 °C]; *v*_{max} (solid)/cm⁻¹ 1681 (s), 1490 (s), 1327 (s); *δ*_H (CDCl₃, 500 MHz) 9.93 (1H, s, CHO), 7.82 (1H, s, CH), 4.36 (3H, s, CH₃); *δ*_C (CDCl₃, 126 MHz) 180.7, 148.6, 139.8, 132.7, 35.9; HRMS *m/z* (ESI⁺) [found (M+MeOH+Na)⁺ 210.0484, C₆H₉N₃NaO₄ requires M⁺ 210.0485]; *m/z* (ES⁺) 210.1 ([M+MeOH+Na]⁺, 100 %). These data are in good agreement with the literature values.²

***N,N'*-bis(2-Bromoethyl)phosphorodiamidic acid (Br-IPM, 13)**



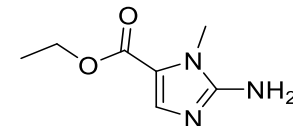
Following the procedure of Duan *et al.*⁴. To a vigorously stirring suspension of 2-bromoethylamine hydrobromide (12.6 g, 62 mmol, 1 eq.) in CH₂Cl₂ (110 mL), was added POCl₃ (2.84 mL, 31 mmol, 0.5 eq.) drop wise, while maintaining the temperature between -10 °C and -15 °C. A solution of triethylamine (17.5 mL, 124 mmol, 2 eq.) in CH₂Cl₂ (30 mL) was then added, drop wise, over a period of 30 min, while maintaining the temperature between -10 °C and -15 °C. The reaction mixture was then filtered, and the filtrate was concentrated *in vacuo* to a volume of approximately 30 mL. The residue was filtered, the combined solids were washed with cold CH₂Cl₂, and the filtrate concentrated and dried *in vacuo*. The residue was dissolved in THF (7 mL), H₂O added (10 mL), and the mixture stirred at room temperature for 2 h. The THF was removed under a flow of N₂ gas, and the remaining aqueous solution cooled to 4 °C overnight. The resulting solid was filtered, and washed with cold H₂O (5 mL) and diethyl ether (5 mL) to afford **13** as a colourless solid that was contaminated with its triethylamine salt: mp 109-112 °C (from H₂O) [lit. mp 134-135 °C]; ν_{\max} (solid) cm⁻¹: 3305 (w), 1249 (s), 1126 (s); δ_{H} (DMSO-D₆, 600 MHz) 3.43 (t, 4H, *J* 7.2, CH₂Br), 3.08 (dt, 4.37H, *J* 12.2, 7.2, NCH₂/CH₃CH₂N), 1.19 (0.7H, t, *J* 7.3, CH₃CH₂N); δ_{C} (DMSO-D₆, 126 MHz) 45.7, 43.0, 34.0 (d, *J* 4.4, CH₂Br), 8.6; δ_{P} (DMSO-D₆, 162 MHz) 11.9. The ¹H NMR data are in good agreement with the literature values.⁵

(1-Methyl-2-nitro-1*H*-imidazol-5-yl)-*N,N*-bis(2-bromoethyl) phosphordiamidate (TH-302)



To a suspension of Br-IPM (78 mg, 0.25 mmol, 1 eq.), was added nitroimidazole alcohol **8** (80 mg, 0.50 mmol, 2 eq.), and triphenylphosphine (133 mg, 0.50 mmol, 2 eq.) in THF (7 mL), and DIAD (100 μ L, 101 mg, 0.50 mmol, 2 eq.) at 0 °C. The reaction mixture was stirred at room temperature for 3 h, the solvent removed under a flow of N₂ gas, and the resulting residue dried *in vacuo*. The residue was then purified by semi-preparative HPLC on a Phenomenex Luna (C18(2), 10 μ m, 250 \times 10 mm) column, eluting with H₂O and methanol (50 – 70% methanol over 10 min, then 1 min wash with methanol, 5 mL/min flow rate) to afford **TH-302** as a yellow gum: ν_{max} (solid) cm⁻¹: 3212 (br), 1489 (m), 1350 (m), 1105 (m), 1004 (s); δ_{H} (DMSO-D₆, 400 MHz) 7.25 (1H, s, CH), 5.10–4.90 (2H, m, NHCH₂CH₂Br), 4.98 (2H, d, *J* 7.8, CH₂O), 3.94 (3H, s, CH₃), 3.42 (4H, t, *J* 7.0, NHCH₂CH₂Br), 3.11 (4H, dt, *J* 9.8, 7.2, NHCH₂CH₂Br); δ_{C} (DMSO-D₆, 126 MHz) 146.1, 134.2 (d, *J* 7.5, OCH₂CN), 128.2, 55.6 (d, *J* 4.6, CH₂O), 42.7, 34.2 (d, *J* 26.4, CH₂Br), 34.1; δ_{P} (DMSO-D₆, 202 MHz) 15.4; HRMS *m/z* (ESI⁻) [found; (M-H)⁻ 447.9216, C₉H₁₆⁷⁹Br⁸¹BrN₅O₄P requires (M-H)⁻ 447.9213]; *m/z* (ESI⁺) 448.0 ([M-H]⁺, 60%, [C₉H₁₅⁷⁹Br⁸¹BrN₅O₄P]⁺), 493.9 ([M+formate]⁺, 100%, [C₁₀H₁₇⁷⁹Br⁸¹BrN₅O₆P]⁺). These data are in good agreement with the literature values.⁴

Ethyl 2-amino-1-methyl-1*H*-imidazole-5-carboxylate (**4**)

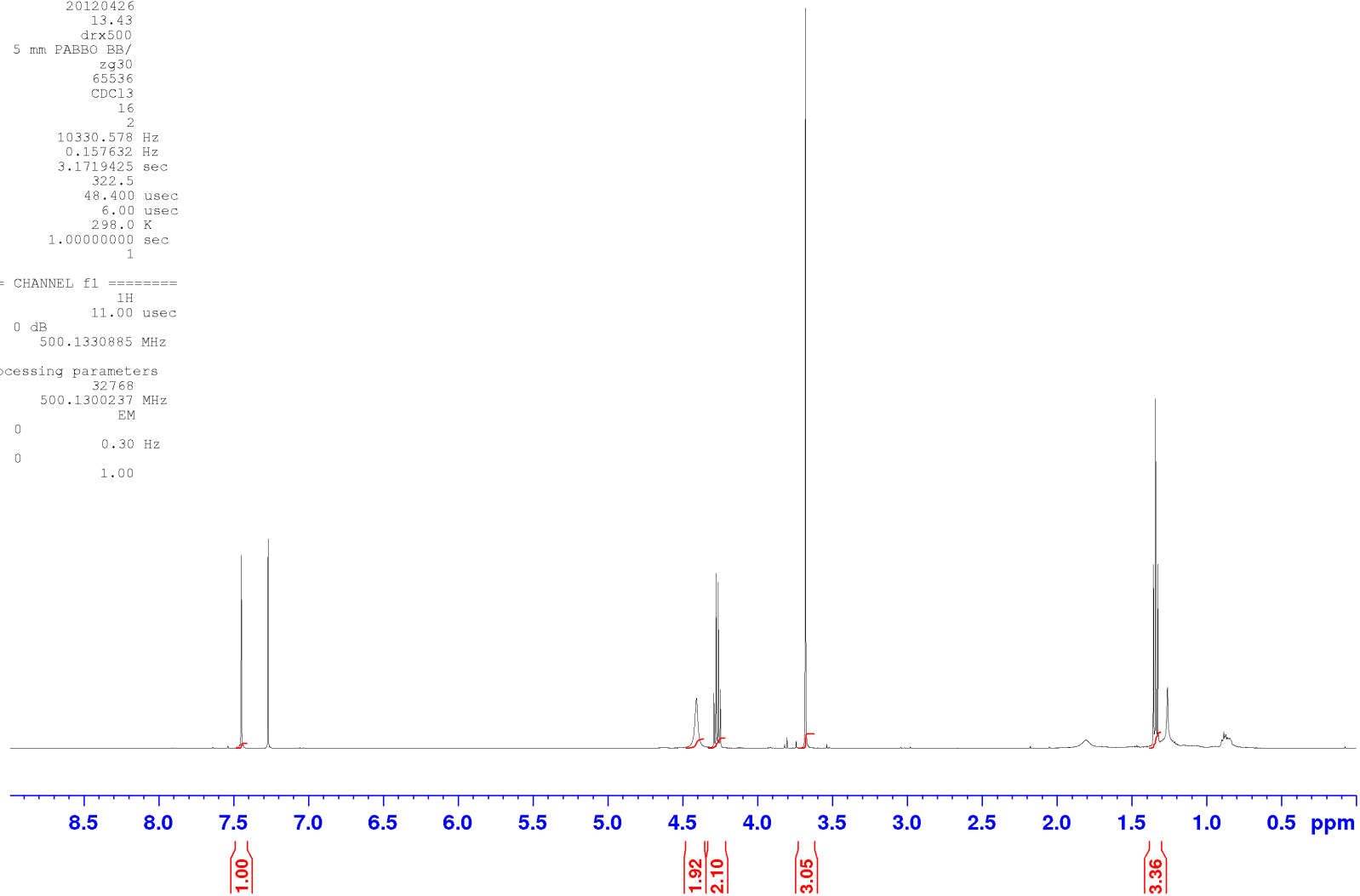


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PROCNO 1

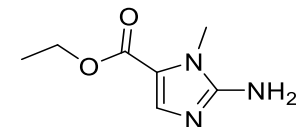
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D1 1.00000000 sec
TD0 1

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PL1 0 dB
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F2 - Processing parameters
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Ethyl 2-amino-1-methyl-1*H*-imidazole-5-carboxylate (4)



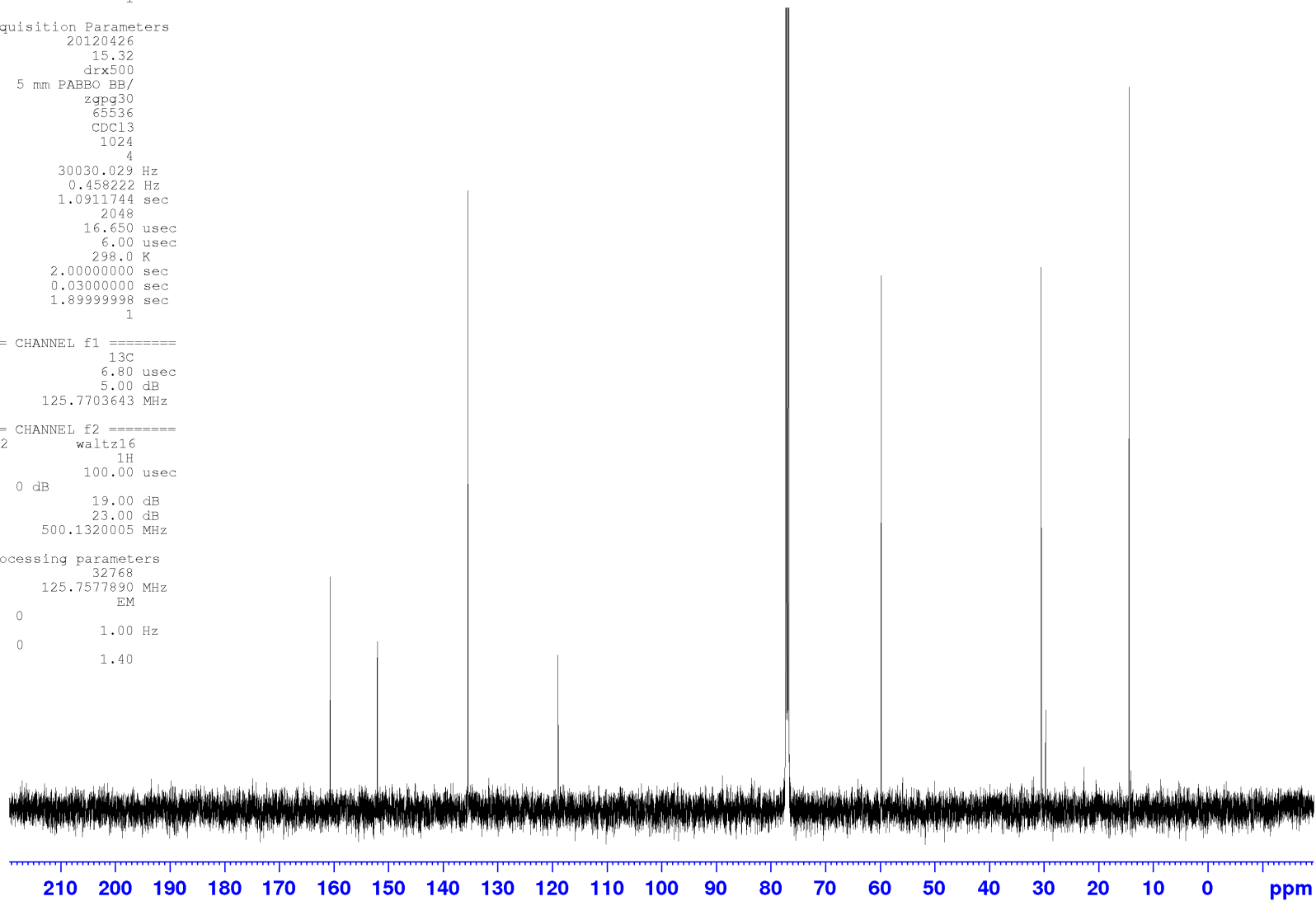
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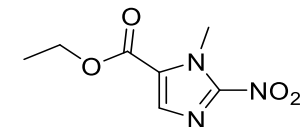
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PL13 23.00 dB
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GB 0
PC 1.40



Ethyl 2-nitro-1-methyl-1*H*-imidazole-5-carboxylate (**7**)

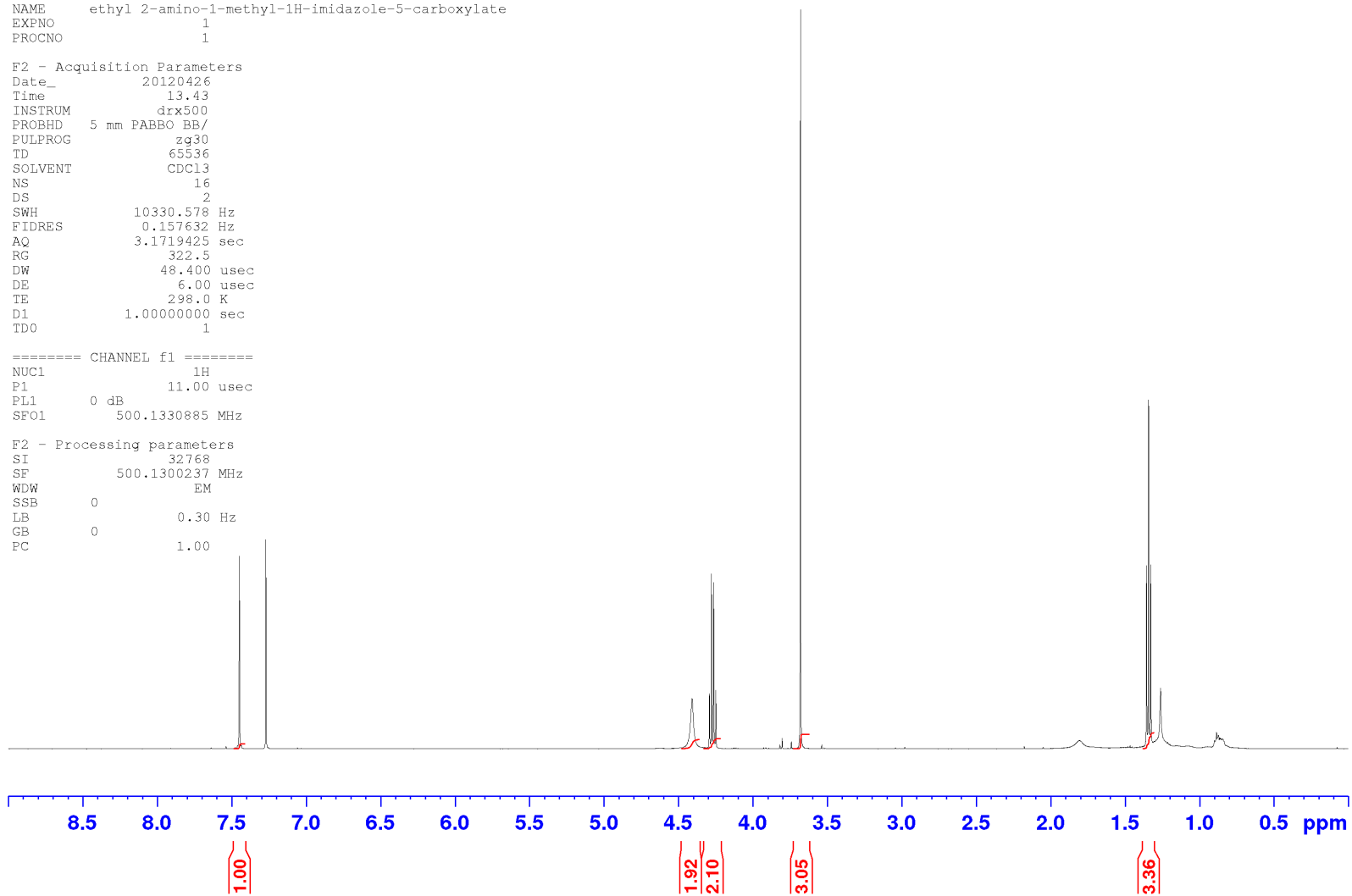


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PROCNO 1

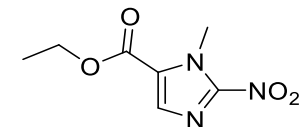
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D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
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P1 11.00 usec
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SF01 500.1330885 MHz

F2 - Processing parameters
SI 32768
SF 500.1300237 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Ethyl 2-nitro-1-methyl-1*H*-imidazole-5-carboxylate (7)



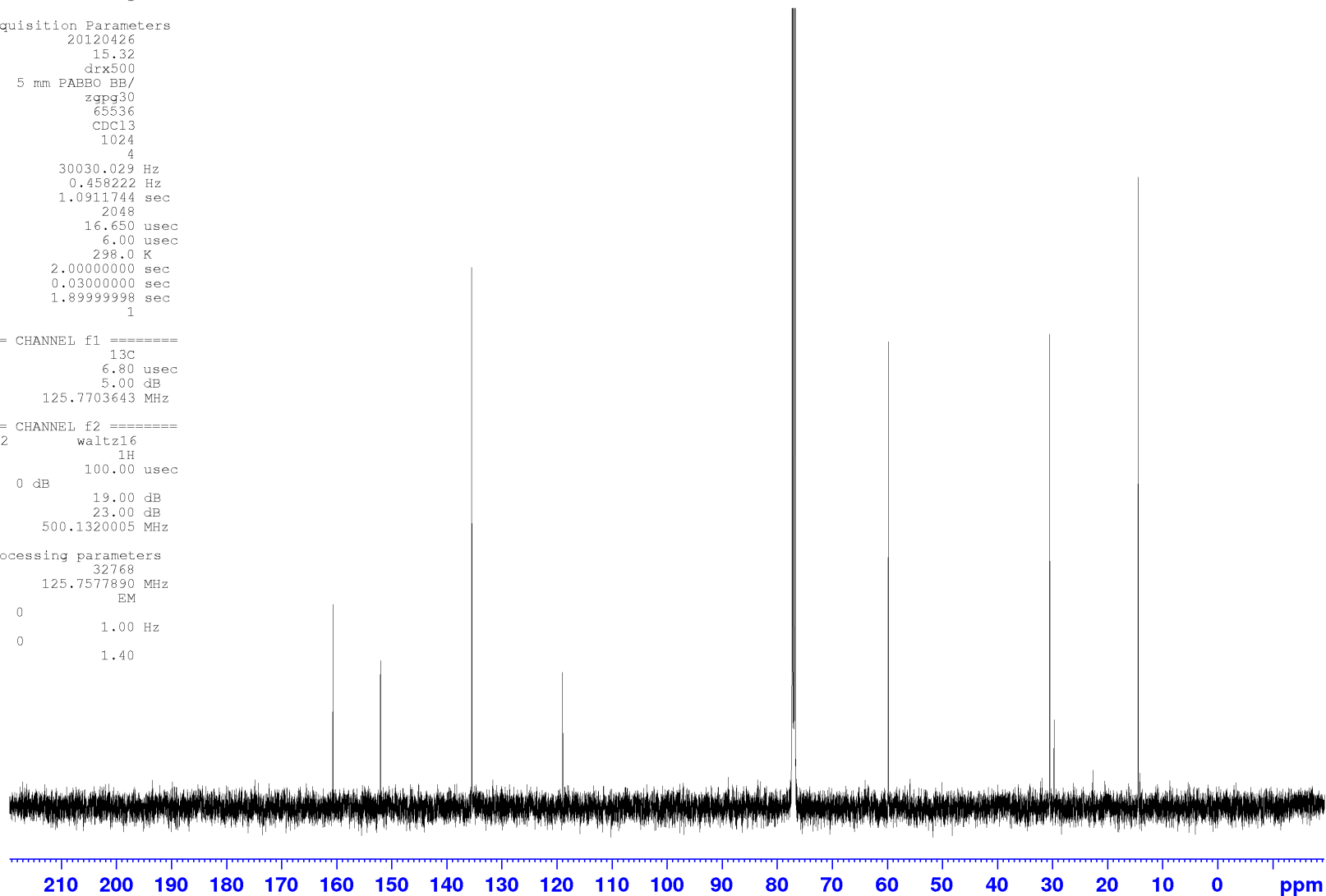
Current Data Parameters
NAME ethyl 2-amino-1-methyl-1*H*-imidazole-5-carboxylate
EXPNO 5
PROCNO 1

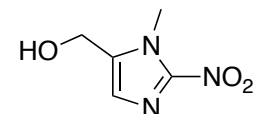
F2 - Acquisition Parameters
Date_ 20120426
Time 15.32
INSTRUM drx500
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 4
SWH 30030.029 Hz
FIDRES 0.458222 Hz
AQ 1.0911744 sec
RG 2048
DW 16.650 usec
DE 6.00 usec
TE 298.0 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 6.80 usec
PL1 5.00 dB
SFO1 125.7703643 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0 dB
PL12 19.00 dB
PL13 23.00 dB
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





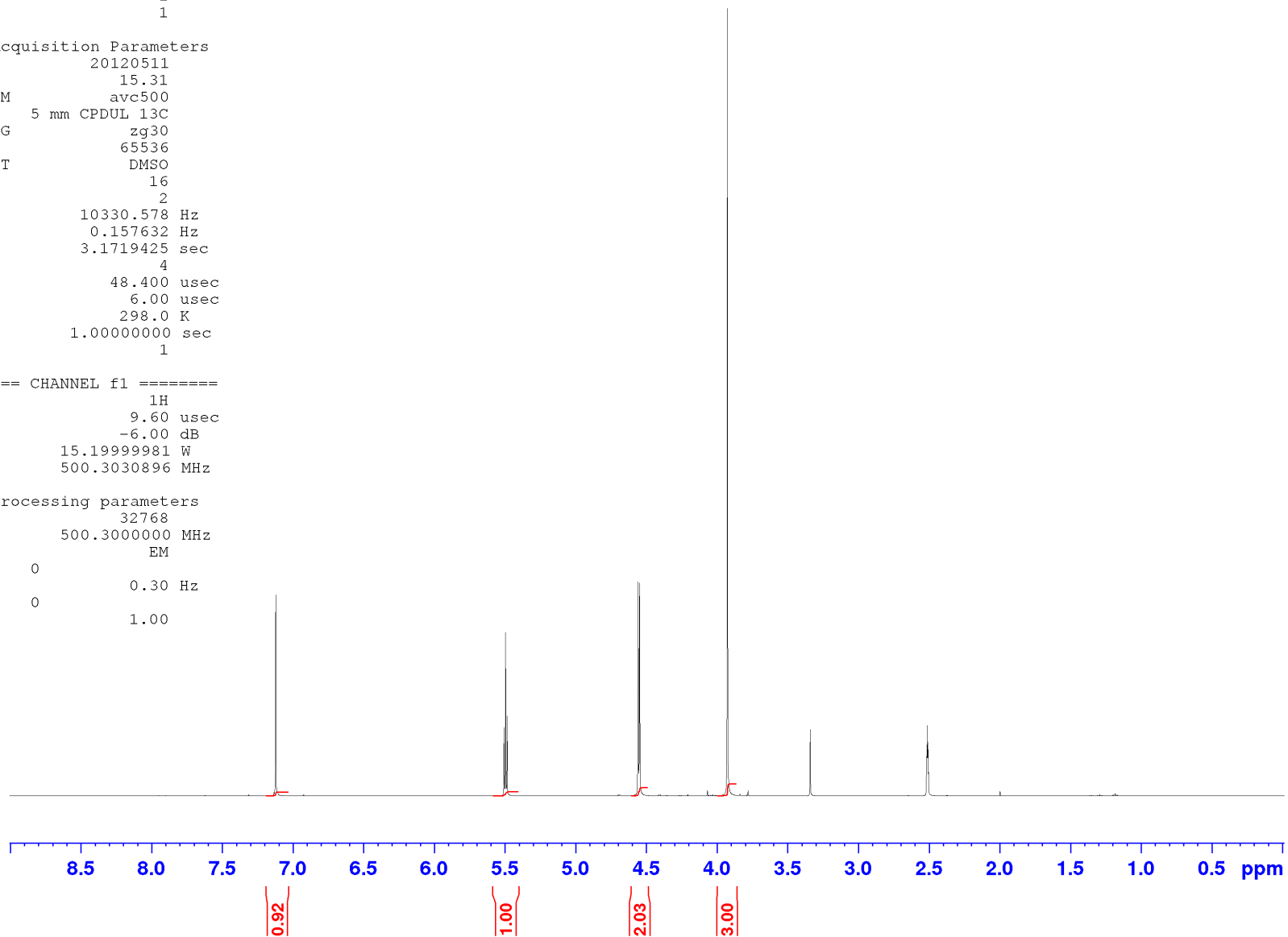
(1-Methyl-2-nitro-1*H*-imidazol-5-yl)methanol (**8**)

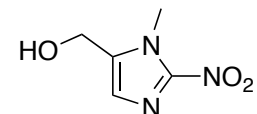
Current Data Parameters
 NAME (1-methyl-2-nitro-1*H*-imidazol-5-yl)methanol
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120511
 Time 15.31
 INSTRUM avc500
 PROBHD 5 mm CPDUL 13C
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 10330.578 Hz
 FIDRES 0.157632 Hz
 AQ 3.1719425 sec
 RG 4
 DW 48.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 9.60 usec
 PL1 -6.00 dB
 PL1W 15.19999981 W
 SFO1 500.3030896 MHz

F2 - Processing parameters
 SI 32768
 SF 500.300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





(1-Methyl-2-nitro-1*H*-imidazol-5-yl)methanol (**8**)

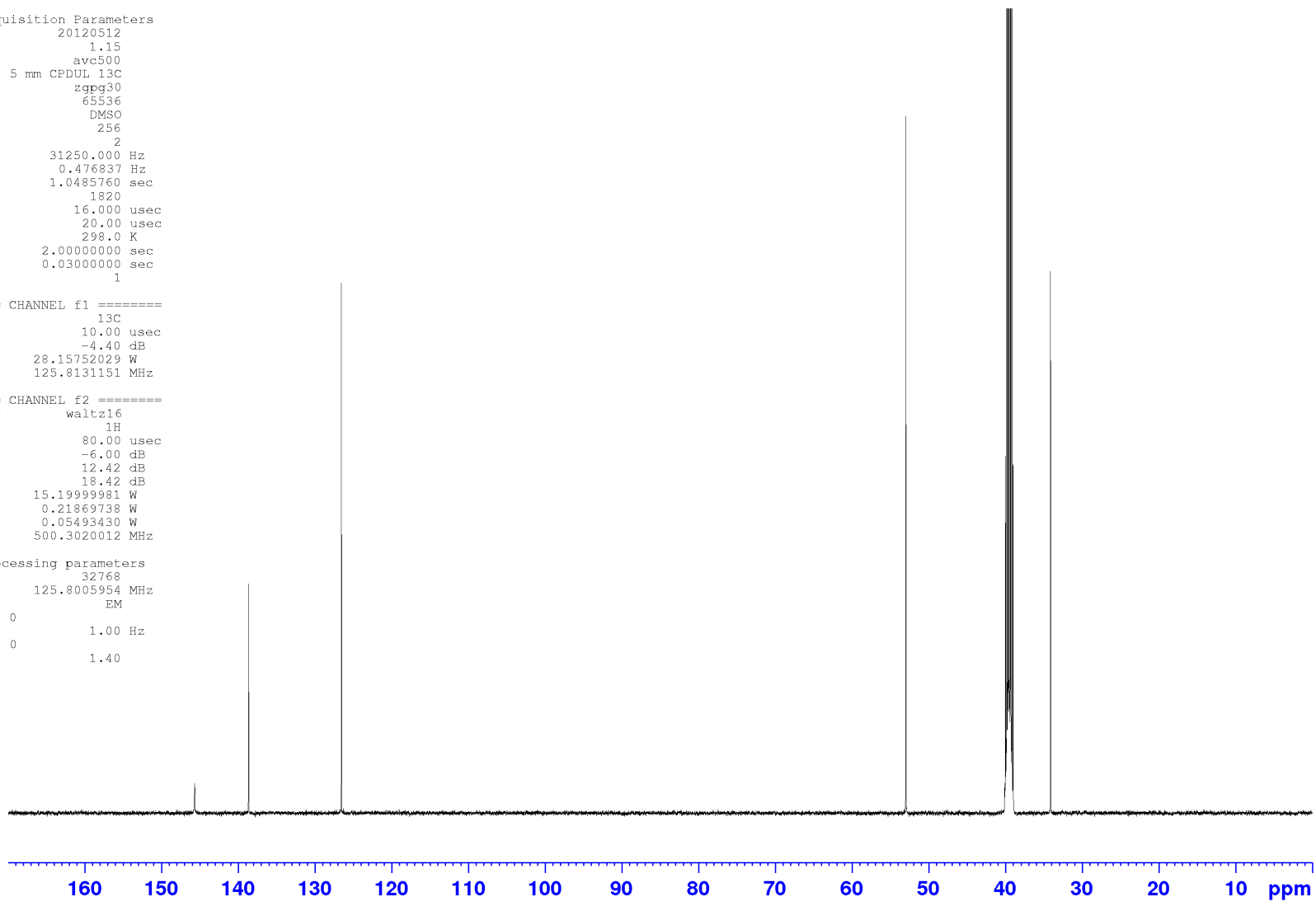
Current Data Parameters
 NAME (1-methyl-2-nitro-1*H*-imidazol-5-yl)methanol
 EXPNO 4
 PROCNO 1

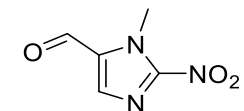
F2 - Acquisition Parameters
 Date_ 20120512
 Time 1.15
 INSTRUM avc500
 PROBHD 5 mm CPDUL 13C
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 256
 DS 2
 SWH 31250.000 Hz
 FIDRES 0.476837 Hz
 AQ 1.0485760 sec
 RG 1820
 DW 16.000 usec
 DE 20.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -4.40 dB
 PL1W 28.15752029 W
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -6.00 dB
 PL12 12.42 dB
 PL13 18.42 dB
 PL2W 15.19999981 W
 PL12W 0.21869738 W
 PL13W 0.05493430 W
 SFO2 500.3020012 MHz

F2 - Processing parameters
 SI 32768
 SF 125.8005954 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40





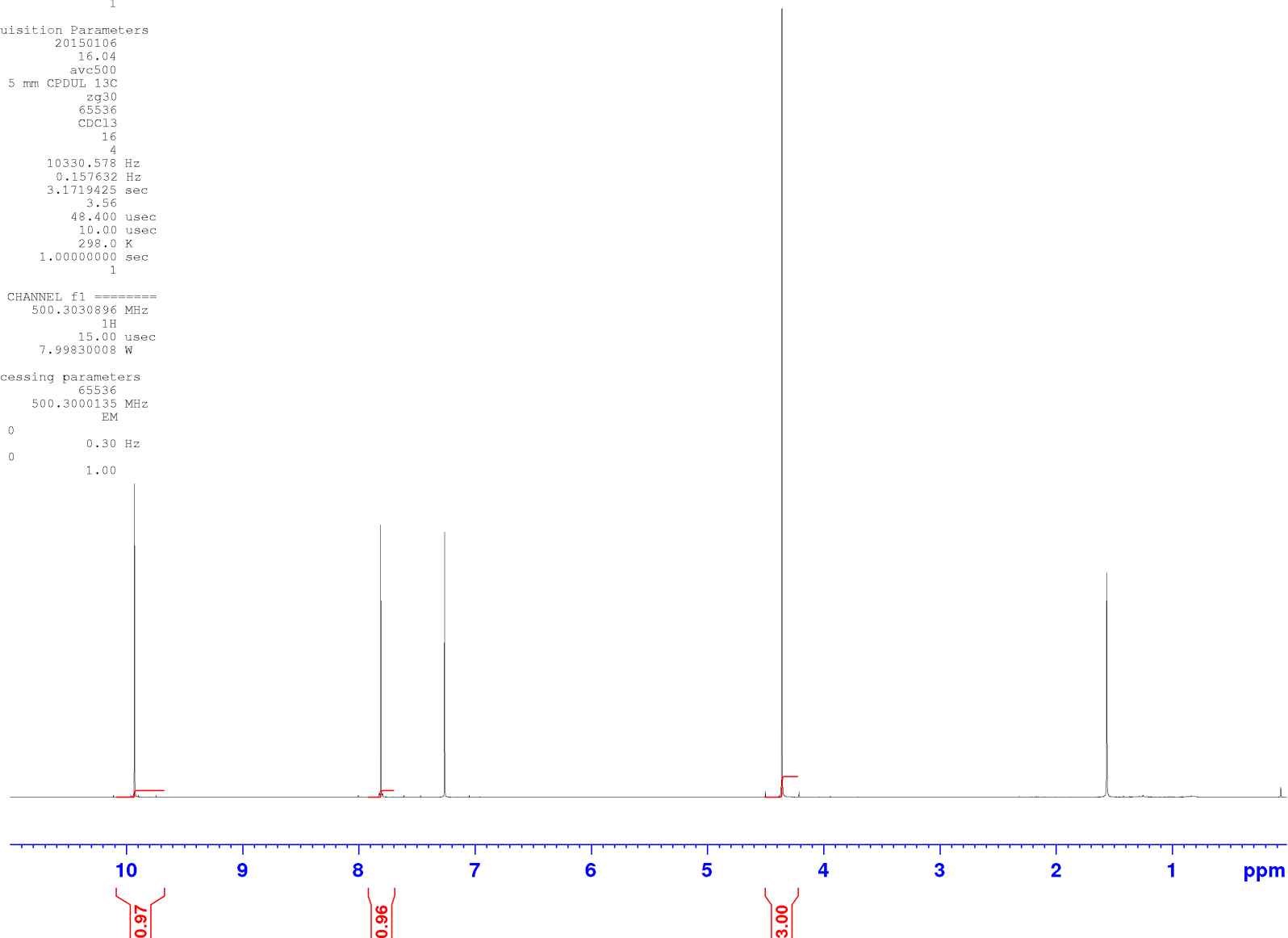
1-Methyl-2-nitro-1H-imidazole-5-carbaldehyde (11)

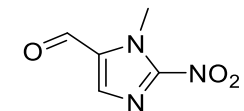
Current Data Parameters
NAME 1-methyl-2-nitro-1H-imidazole-5-carbaldehyde
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150106
Time 16.04
INSTRUM avc500
PROBHD 5 mm CPDUL 13C
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 4
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719425 sec
RG 3.56
DW 48.400 usec
DE 10.00 usec
TE 298.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 500.3030896 MHz
NUC1 1H
P1 15.00 usec
PLW1 7.99830008 W

F2 - Processing parameters
SI 65536
SF 500.3000135 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





1-Methyl-2-nitro-1*H*-imidazole-5-carbaldehyde (11)

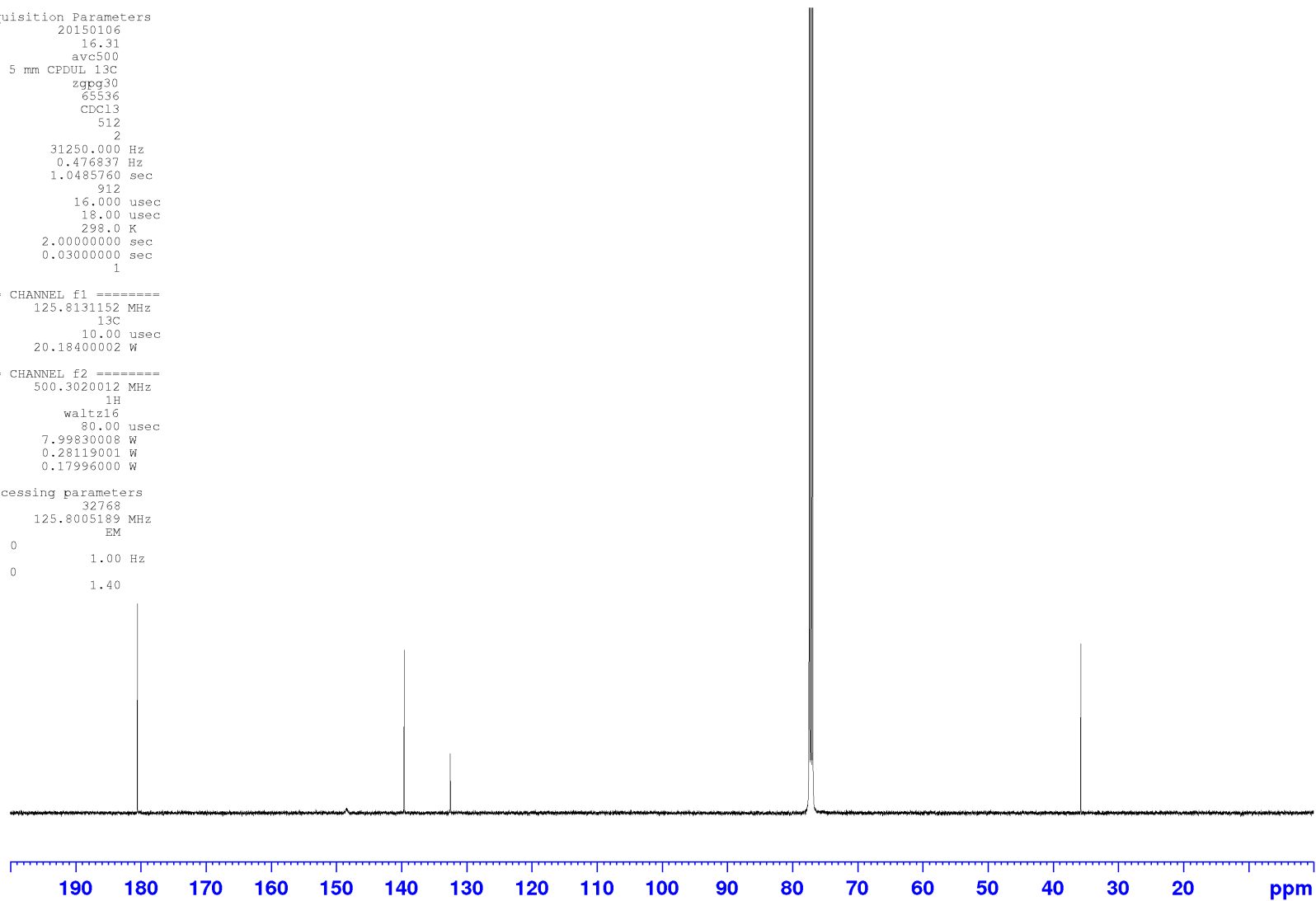
Current Data Parameters
NAME 1-methyl-2-nitro-1*H*-imidazole-5-carbaldehyde
EXPNO 2
PROCNO 1

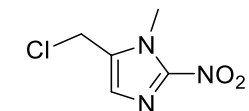
F2 - Acquisition Parameters
Date_ 20150106
Time 16.31
INSTRUM avc500
PROBHD 5 mm CPDUL 13C
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 2
SWH 31250.000 Hz
FIDRES 0.476837 Hz
AQ 1.0485760 sec
RG 912
DW 16.000 usec
DE 18.00 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 125.8131152 MHz
NUC1 13C
P1 10.00 usec
PLW1 20.18400002 W

===== CHANNEL f2 =====
SFO2 500.3020012 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 80.00 usec
PLW2 7.99830008 W
PLW12 0.28119001 W
PLW13 0.17996000 W

F2 - Processing parameters
SI 32768
SF 125.8005189 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





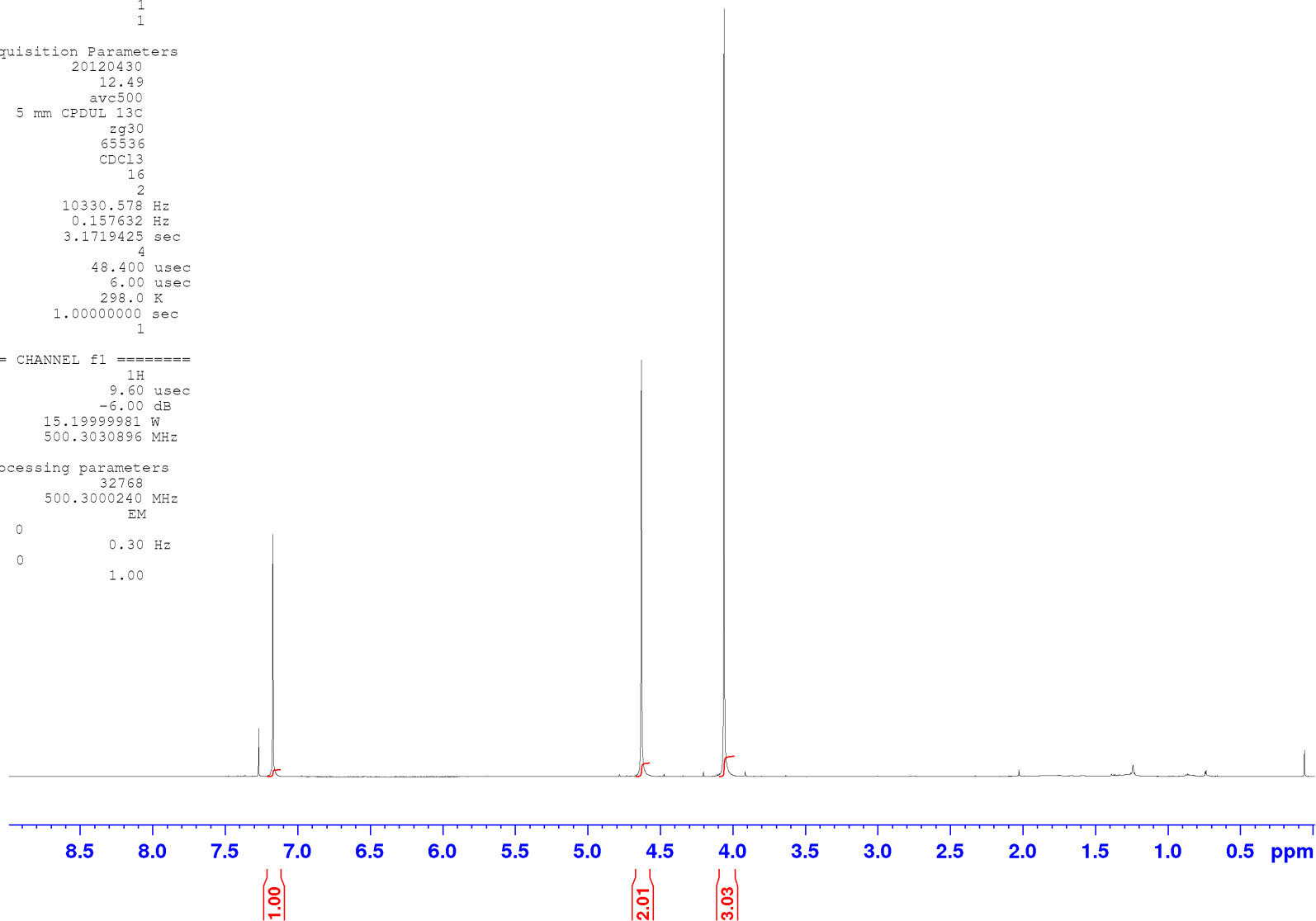
5-(Chloromethyl)-1-methyl-2-nitro-1H-imidazole (10)

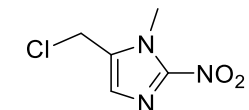
Current Data Parameters
NAME 5-(chloromethyl)-1-methyl-2-nitro-1H-imidazole
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120430
Time 12.49
INSTRUM avc500
PROBHD 5 mm CPDUL 13C
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719425 sec
RG 4
DW 48.400 usec
DE 6.00 usec
TE 298.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.60 usec
PL1 -6.00 dB
PL1W 15.19999981 W
SFO1 500.3030896 MHz

F2 - Processing parameters
SI 32768
SF 500.3000240 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





5-(Chloromethyl)-1-methyl-2-nitro-1H-imidazole (10)

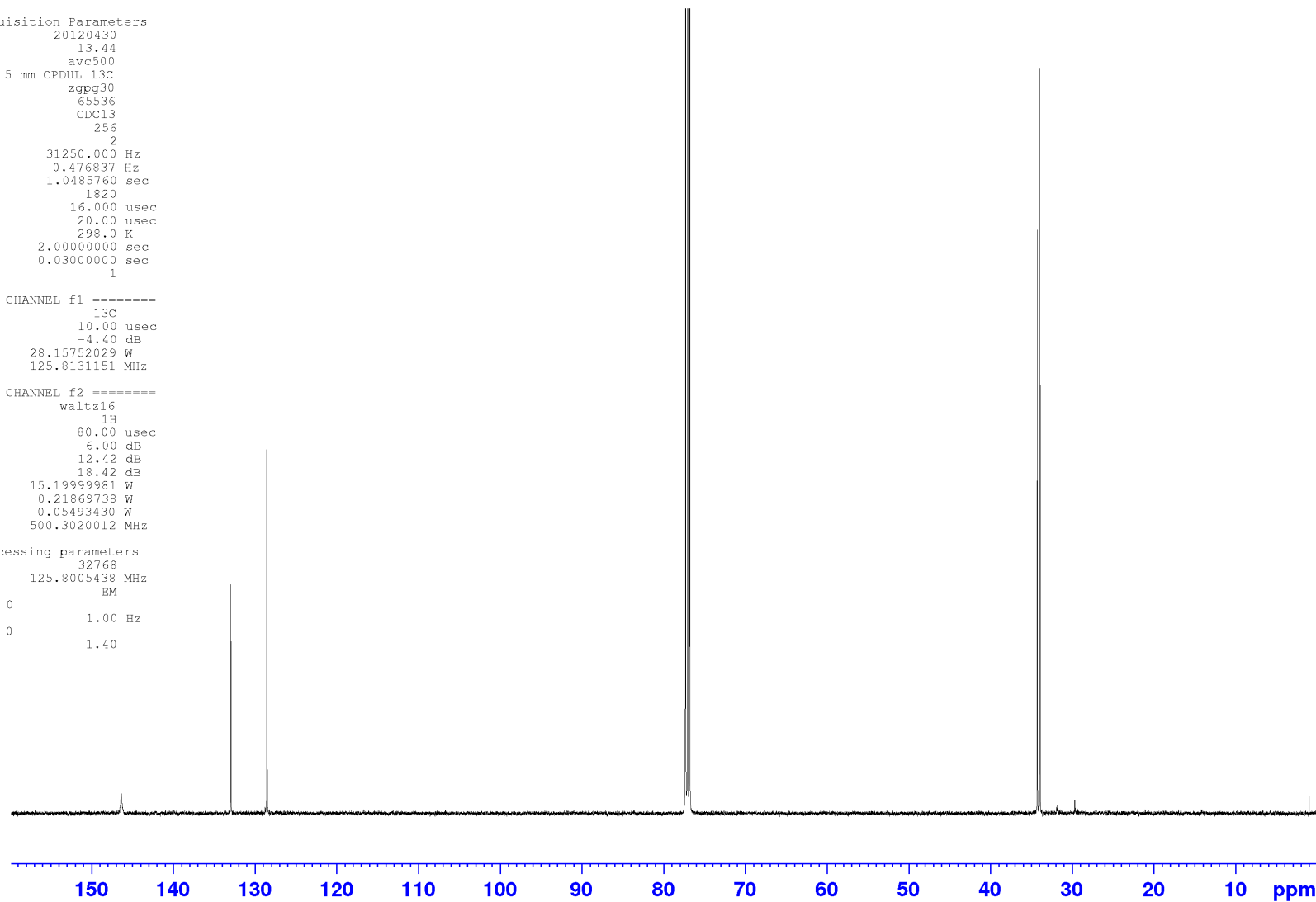
Current Data Parameters
NAME 5-(chloromethyl)-1-methyl-2-nitro-1H-imidazole
EXPNO 4
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120430
Time 13.44
INSTRUM avc500
PROBHD 5 mm CPDUL 13C
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 256
DS 2
SWH 31250.000 Hz
FIDRES 0.476837 Hz
AQ 1.0485760 sec
RG 1820
DW 16.000 usec
DE 20.00 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

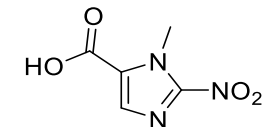
===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -4.40 dB
PL1W 28.15752029 W
SFO1 125.8131151 MHz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -6.00 dB
PL12 12.42 dB
PL13 12.42 dB
PL2W 15.19999981 W
PL12W 0.21869738 W
PL13W 0.05493430 W
SFO2 500.3020012 MHz

F2 - Processing parameters
SI 32768
SF 125.8005438 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



1-Methyl-2-nitro-1H-imidazole-5-carboxylic acid (**9**)

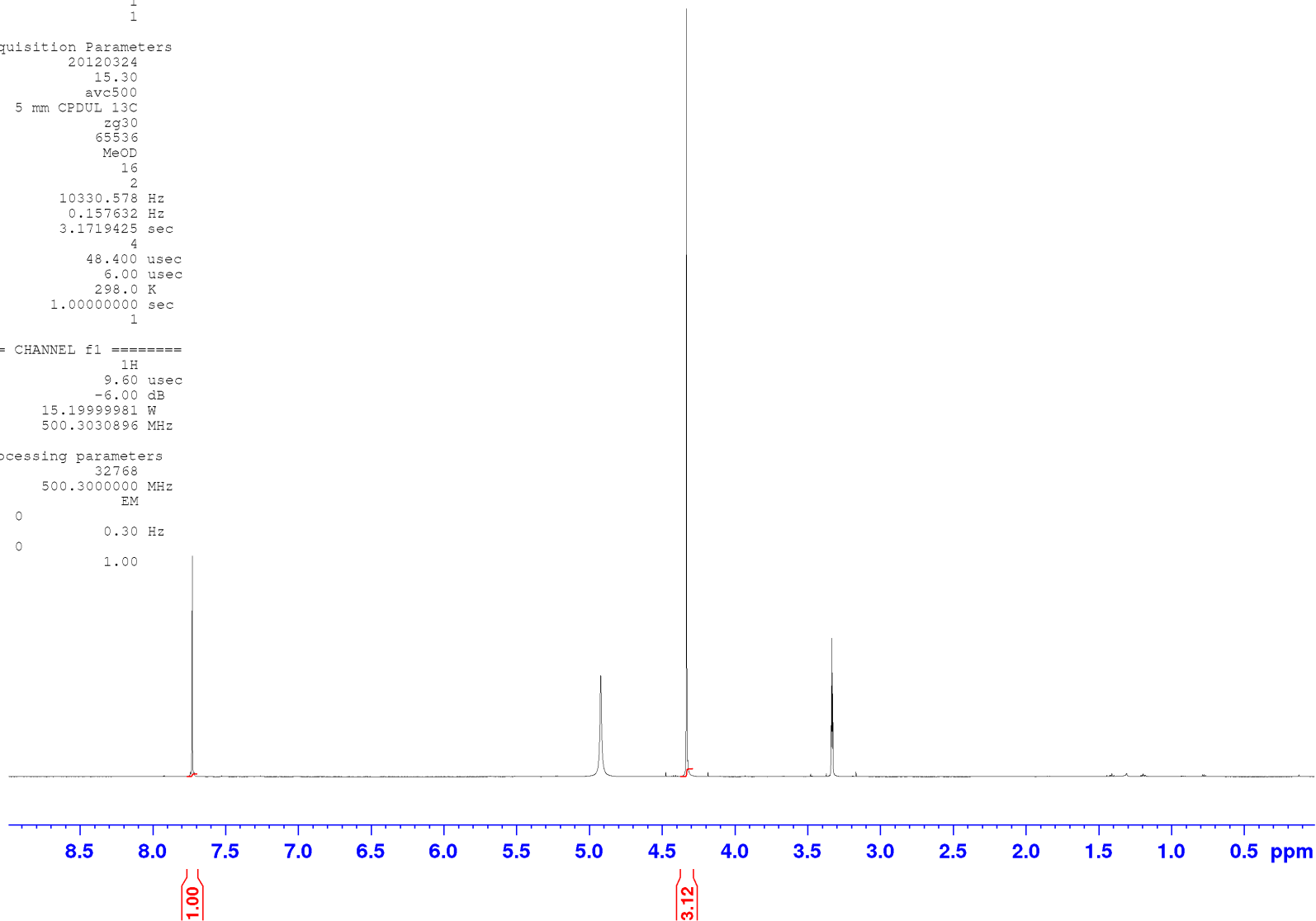


Current Data Parameters
NAME 1-methyl-2-nitro-1H-imidazole-5-carboxylic acid
EXPNO 1
PROCNO 1

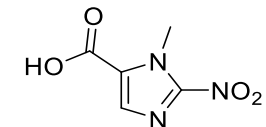
F2 - Acquisition Parameters
Date_ 20120324
Time 15.30
INSTRUM avc500
PROBHD 5 mm CPDUL 13C
PULPROG zg30
TD 65536
SOLVENT MeOD
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719425 sec
RG 4
DW 48.400 usec
DE 6.00 usec
TE 298.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.60 usec
PL1 -6.00 dB
PL1W 15.19999981 W
SFO1 500.3030896 MHz

F2 - Processing parameters
SI 32768
SF 500.3000000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



1-Methyl-2-nitro-1*H*-imidazole-5-carboxylic acid (9)



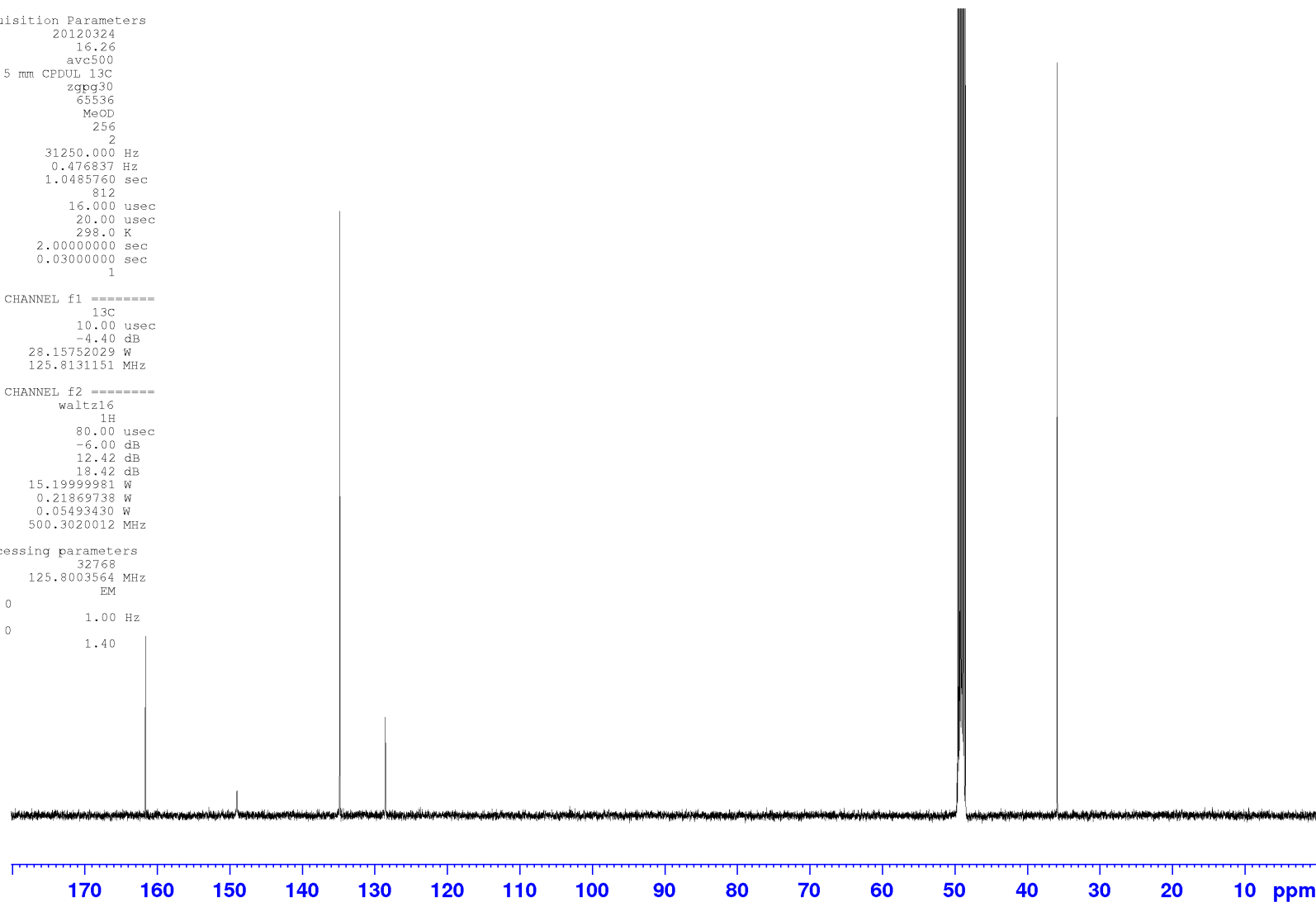
Current Data Parameters
 NAME 1-methyl-2-nitro-1H-imidazole-5-carboxylic acid
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120324
 Time 16.26
 INSTRUM avc500
 PROBHD 5 mm CPDUL 13C
 PULPROG zgpg30
 TD 65536
 SOLVENT MeOD
 NS 256
 DS 2
 SWH 31250.000 Hz
 FIDRES 0.476837 Hz
 AQ 1.0485760 sec
 RG 812
 DW 16.000 usec
 DE 20.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

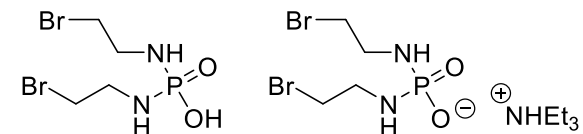
===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -4.40 dB
 PL1W 28.15752029 W
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 FCPD2 80.00 usec
 PL2 -6.00 dB
 PL12 12.42 dB
 PL13 18.42 dB
 PL2W 15.19999981 W
 PL12W 0.21869738 W
 PL13W 0.05493430 W
 SFO2 500.3020012 MHz

F2 - Processing parameters
 SI 32768
 SF 125.8003564 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



N,N'-bis(2-Bromoethyl)phosphorodiamidic acid (Br-IPM, **13**)

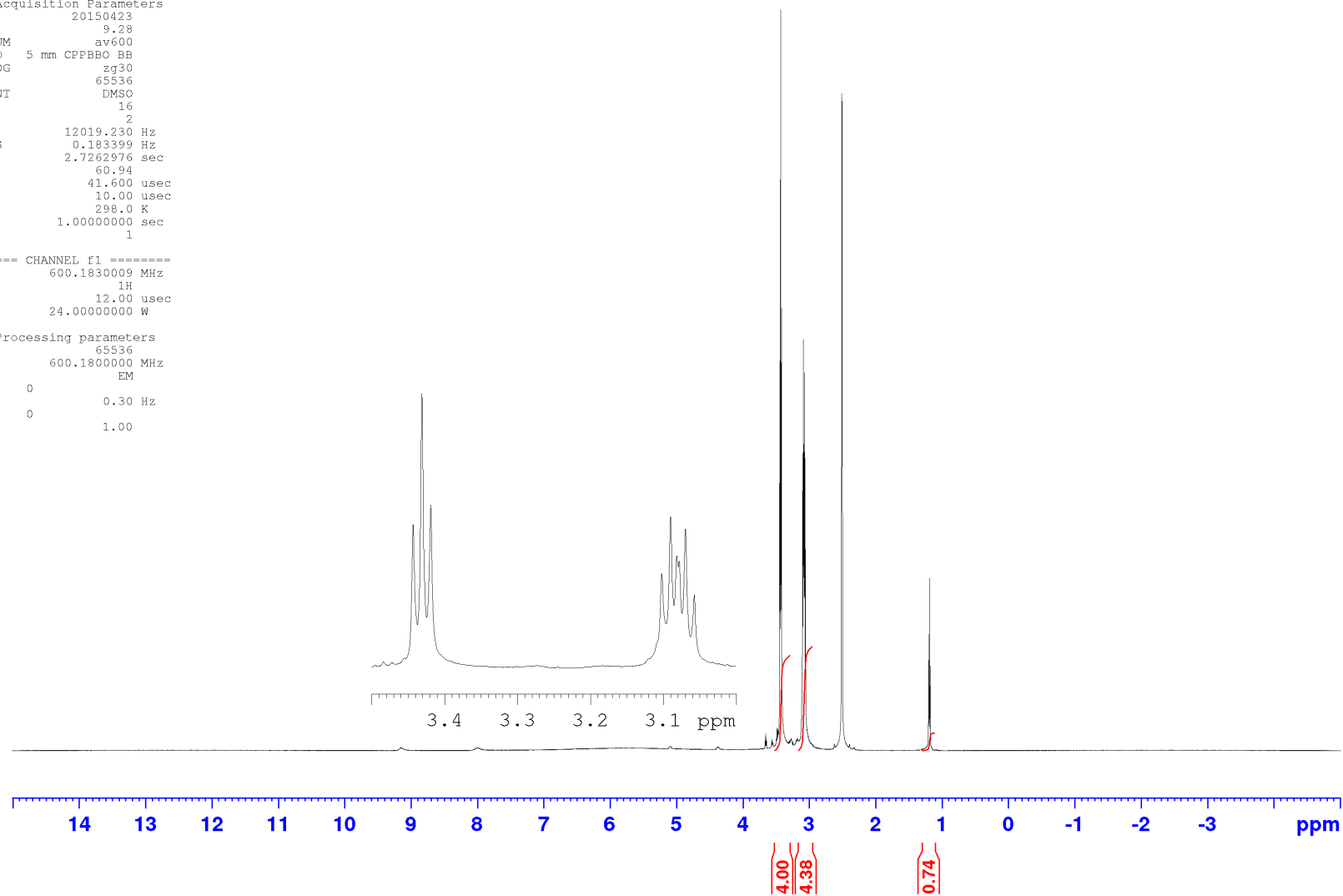


Current Data Parameters
 NAME LOC297 Service 1010192304
 EXPNO 1
 PROCNO 1

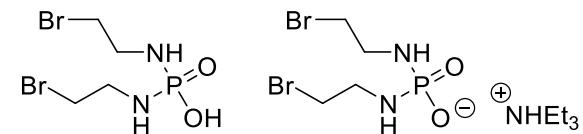
F2 - Acquisition Parameters
 Date_ 20150423
 Time_ 9.28
 INSTRUM av600
 PROBHD 5 mm CPBPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 60.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 600.1830009 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 24.00000000 W

F2 - Processing parameters
 SI 65536
 SF 600.1800000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



N,N'-bis(2-Bromoethyl)phosphorodiamidic acid (Br-IPM, **13**)



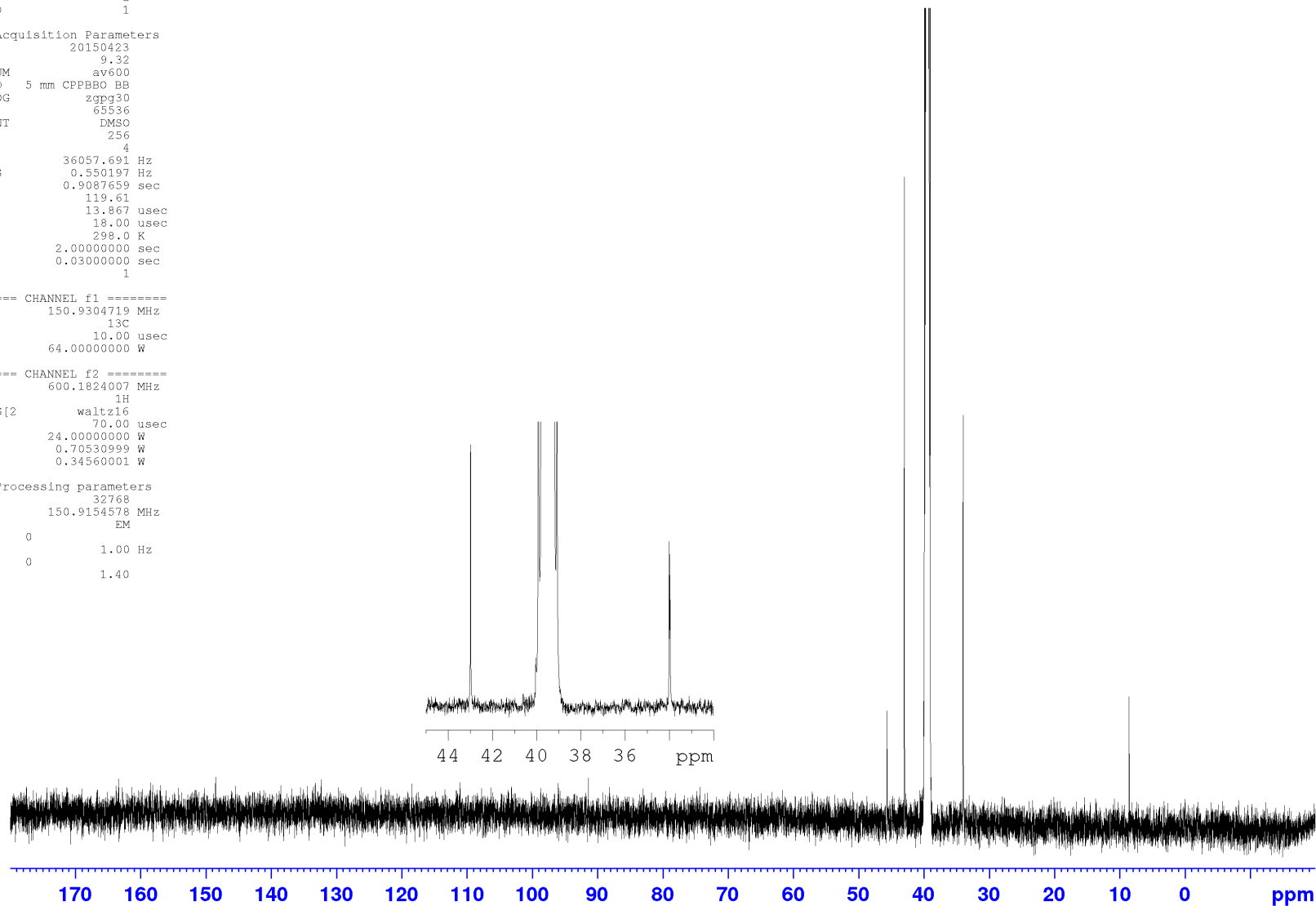
Current Data Parameters
 NAME LOC297 Service lo10192304
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150423
 Time 9.32
 INSTRUM av600
 PROBHD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 256
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 119.61
 DW 13.867 usec
 DE 18.00 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

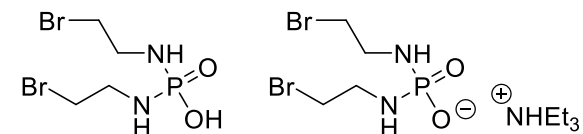
===== CHANNEL f1 =====
 SFO1 150.9304719 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 64.00000000 W

===== CHANNEL f2 =====
 SFO2 600.1824007 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 24.00000000 W
 PLW12 0.70530999 W
 PLW13 0.34560001 W

F2 - Processing parameters
 SI 32768
 SF 150.9154578 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



N,N'-bis(2-Bromoethyl)phosphorodiamidic acid (Br-IPM, **13**)



```

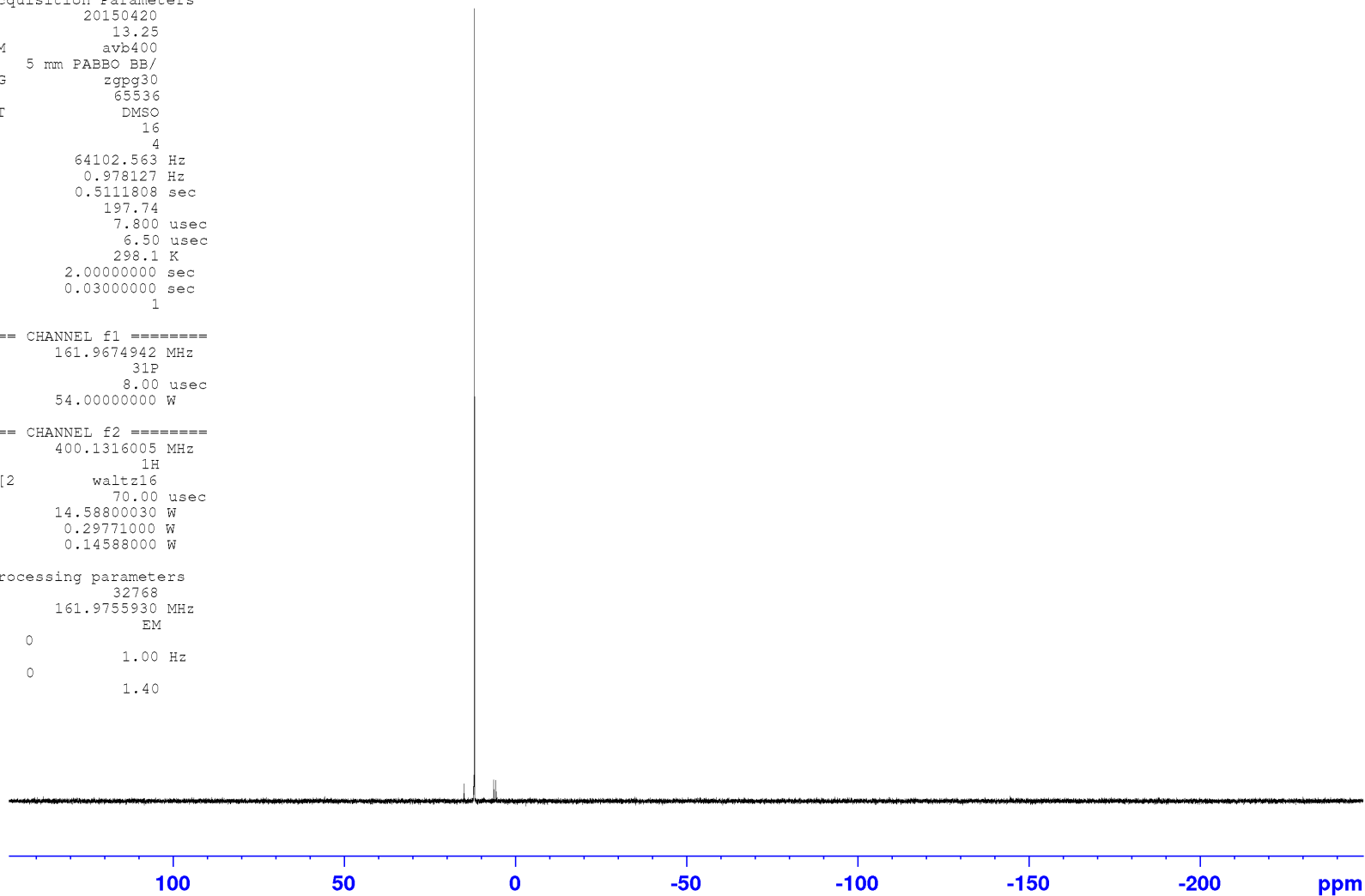
Current Data Parameters
NAME      LOC297 AVB400
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20150420
Time      13.25
INSTRUM   avb400
PROBHD    5 mm PABBO BB/
PULPROG   zgpg30
TD        65536
SOLVENT   DMSO
NS        16
DS        4
SWH       64102.563 Hz
FIDRES    0.978127 Hz
AQ        0.5111808 sec
RG        197.74
DW        7.800 usec
DE        6.50 usec
TE        298.1 K
D1        2.00000000 sec
D11       0.03000000 sec
TD0       1

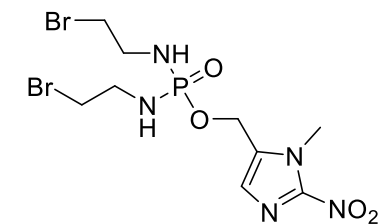
===== CHANNEL f1 =====
SFO1      161.9674942 MHz
NUC1      31P
P1        8.00 usec
PLW1      54.00000000 W

===== CHANNEL f2 =====
SFO2      400.1316005 MHz
NUC2      1H
CPDPRG[2] waltz16
PCPD2     70.00 usec
PLW2      14.58800030 W
PLW12     0.29771000 W
PLW13     0.14588000 W

F2 - Processing parameters
SI        32768
SF        161.9755930 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40
  
```



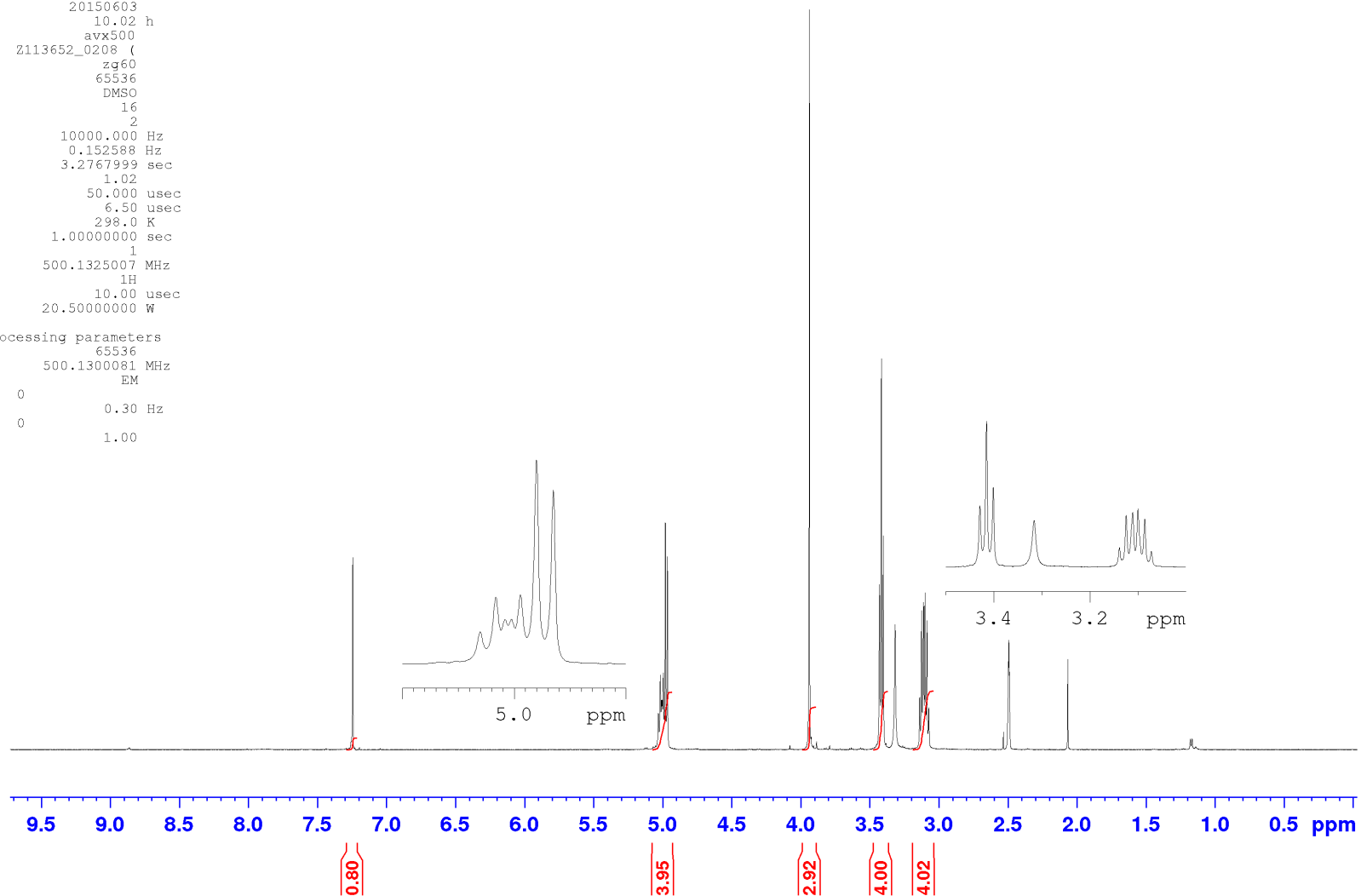
(1-Methyl-2-nitro-1*H*-imidazol-5-yl) *N,N*-bis(2-bromoethyl) phosphordiamidate (TH-302)



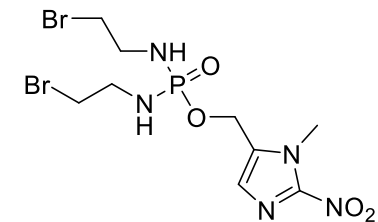
Current Data Parameters
 NAME LOC300 Service 1014870306
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150603
 Time 10.02 h
 INSTRUM avx500
 PROBHD Z113652_0208 (
 PULPROG zg60
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 1.02
 DW 50.000 usec
 DE 6.50 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1
 SF01 500.1325007 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 20.50000000 W

F2 - Processing parameters
 SI 65536
 SF 500.1300081 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



(1-Methyl-2-nitro-1*H*-imidazol-5-yl) *N,N*-bis(2-bromoethyl) phosphordiamidate (TH-302)



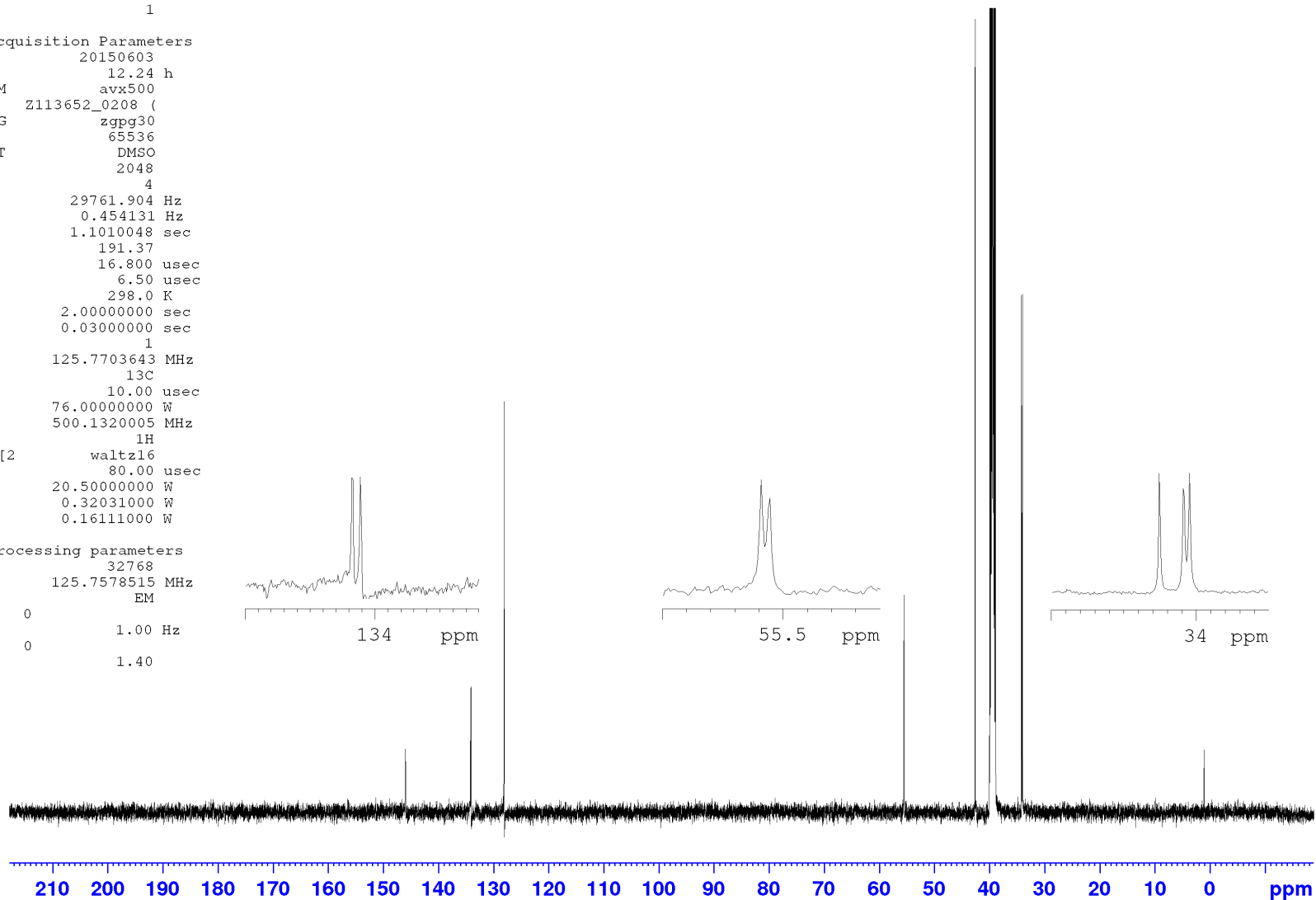
Current Data Parameters
 NAME LOC300 Service lol4870306
 EXPNO 8
 PROCNO 1

F2 - Acquisition Parameters

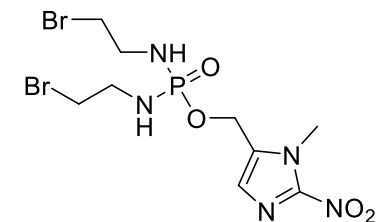
Date_ 20150603
 Time 12.24 h
 INSTRUM avx500
 PROBHD Z113652_0208 (
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 191.37
 DW 16.800 usec
 DE 6.50 usec
 TE 298.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1
 SFO1 125.7703643 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 76.00000000 W
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 20.50000000 W
 PLW12 0.32031000 W
 PLW13 0.16111000 W

F2 - Processing parameters

SI 32768
 SF 125.7578515 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



(1-Methyl-2-nitro-1*H*-imidazol-5-yl) *N,N*-bis(2-bromoethyl) phosphordiamidate (TH-302)



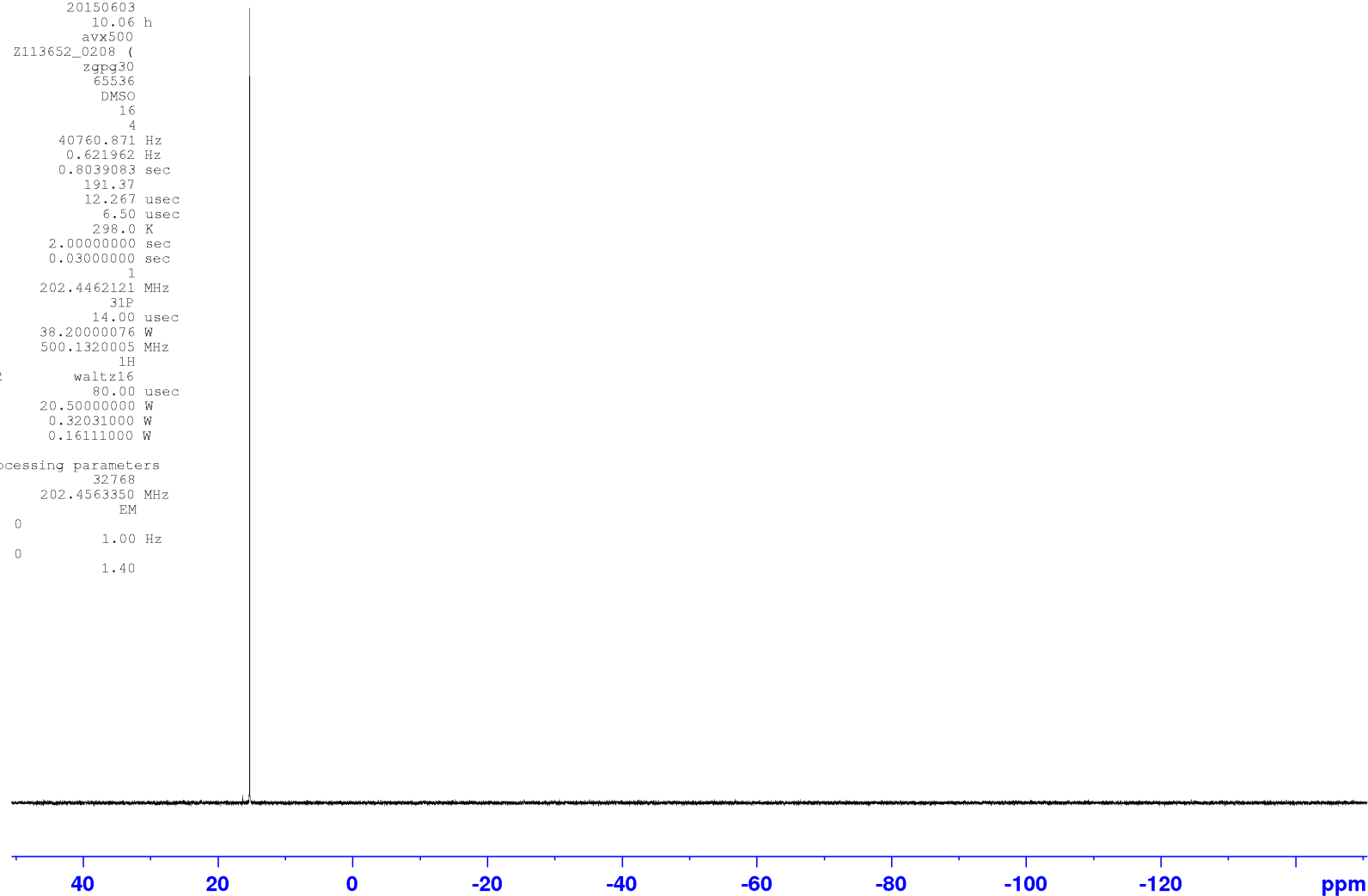
Current Data Parameters
NAME LOC300 Service 1014870306
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150603
Time 10.06 h
INSTRUM avx500
PROBHD Z113652_0208 (
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 16
DS 4
SWH 40760.871 Hz
FIDRES 0.621962 Hz
AQ 0.8039083 sec
RG 191.37
DW 12.267 usec
DE 6.50 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 202.4462121 MHz
NUC1 31P
P1 14.00 usec
PLW1 38.20000076 W
SFO2 500.1320005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 80.00 usec
PLW2 20.50000000 W
PLW12 0.32031000 W
PLW13 0.16111000 W

F2 - Processing parameters

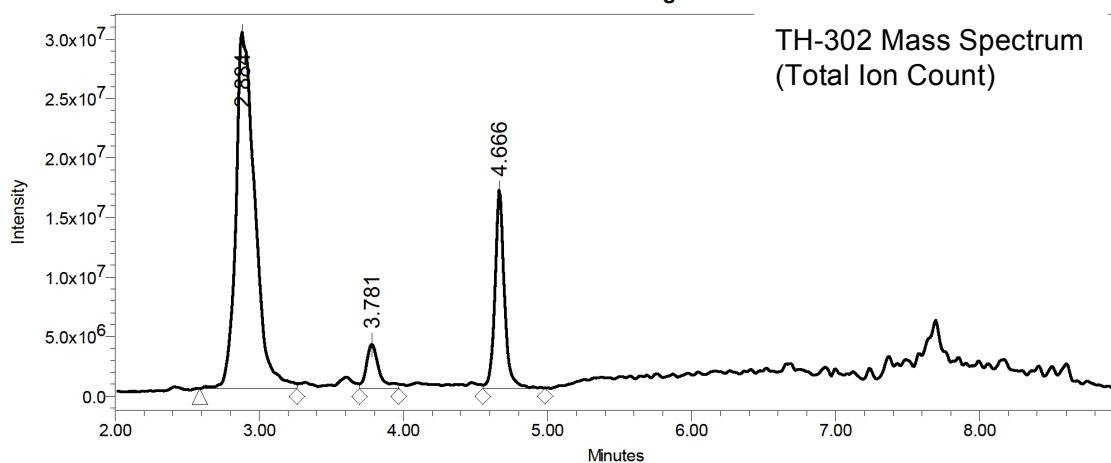
SI 32768
SF 202.4563350 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



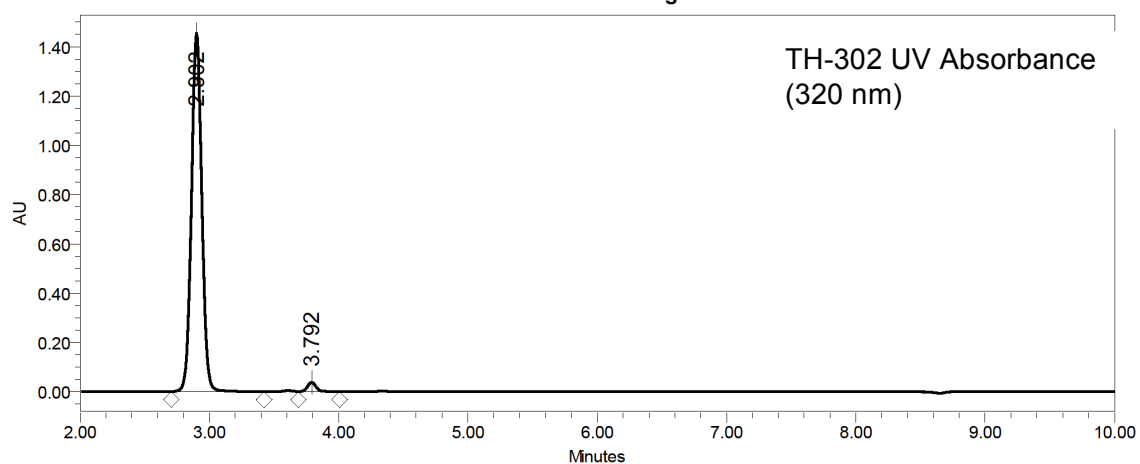
Sample Name: loc300 bulk prep
 Vial: 7
 Injection Volume: 0.50 ul
 Run Time: 10.0 Minutes
 Date Acquired: 14/05/2015 12:21:52 BST

Acq. Method Set: MS_35_95 Liam
 Processing Method: PM_TIC, PM_PDA
 Channel Name: 320NM, MS TIC
 Proc. Chnl. Descr.: ZQ F1 Scan MS TIC, Smoothed by

Auto-Scaled Chromatogram

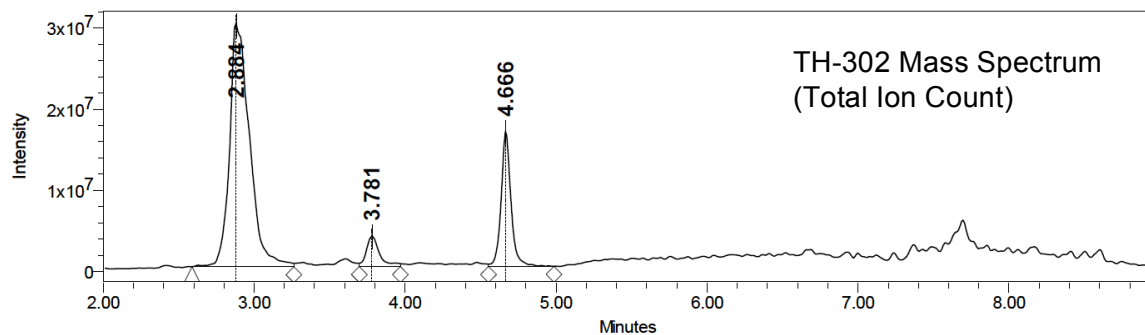
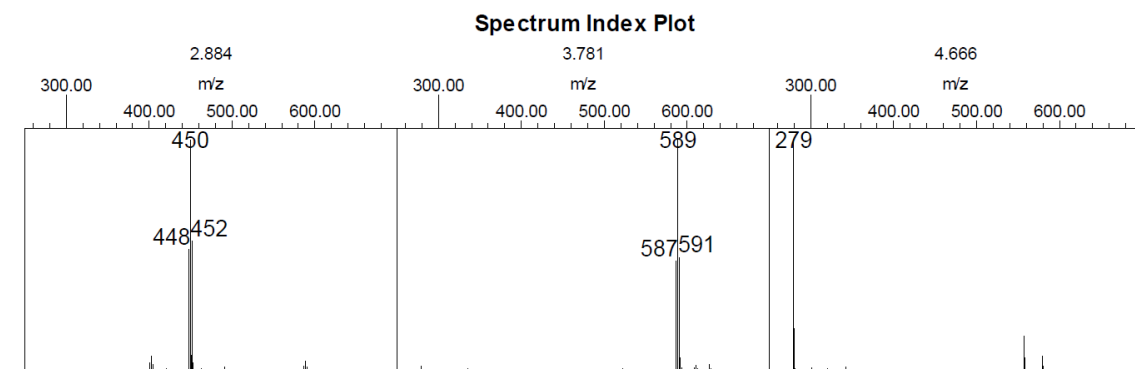


Auto-Scaled Chromatogram

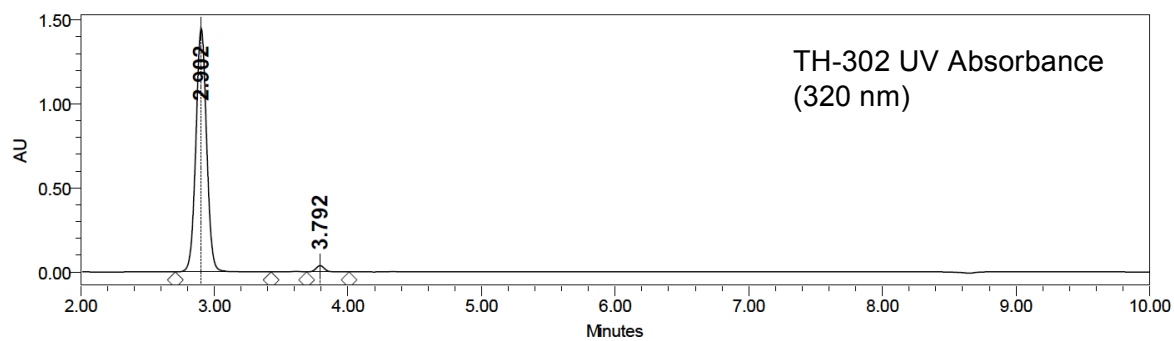
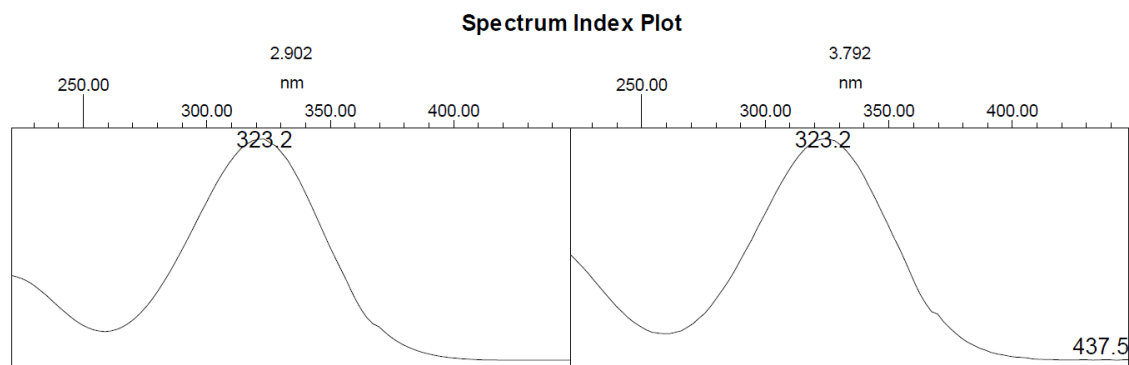


Peak Results

	Name	RT	Area	% Area	Channel Name
1		2.902	8097904	97.86	320NM
2		3.792	177107	2.14	320NM
3		2.884	275854110	73.71	MS TIC
4		3.781	22717370	6.07	MS TIC
5		4.666	75667782	20.22	MS TIC



SampleName: loc300 bulk prep; Vial: 7; Injection: 1; Date Acquired: 14/05/2015 12:21:52 BST



SampleName: loc300 bulk prep; Vial: 7; Injection: 1; Date Acquired: 14/05/2015 12:21:52 BST

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