# Organocatalytic enantioselective Mukaiyama-Mannich reaction of fluorinated enol silyl ethers and cyclic *N*-sulfonyl ketimines

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**General**: Reactions were monitored by thin layer chromatography using UV light to visualize the course of reaction. Purification of reaction products was carried out by flash chromatography on silica gel. Chemical yields referred to pure isolated substances. Infrared (IR) spectra were obtained using Nicolet Fourier Transform Infrared (FT-IR) Spectrometer. The  $[\alpha]_D$  was recorded using PolAAr 3005 High Accuracy Polarimeter. <sup>1</sup>H, <sup>19</sup>F, <sup>13</sup>C NMR spectra were obtained using a Bruker DPX-400 and 300 MHz spectrometer. Chemical shifts are reported in ppm from CDCl<sub>3</sub>, acetone-*d*<sub>6</sub> with the solvent resonance or (CH<sub>3</sub>)<sub>4</sub>Si as the internal standard. The following abbreviations were used to designate chemical shift multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, h = heptet, m = multiplet, br = broad. Coupling constants (*J*) are reported in Hertz.

Anhydrous toluene and THF was prepared by distillation over sodium-benzophenone ketyl prior to use. Anhydrous  $CH_2Cl_2$  and  $CH_3CN$  were prepared by first distillation over  $P_2O_5$  and then from  $CaH_2$ . Anhydrous EtOAc were prepared by first distillation over activated  $CaSO_4$  and MS 5Å prior to use. The cyclic *N*-sulfonyl ketimines **7**<sup>1</sup> were prepared according to the corresponding literature report. The difluoroenoxysilanes **1**, monofluorinated enol silyl ethers **4** and nonfluorinated analogue **8** were prepared according to literature reports.<sup>2</sup>

#### List of abbreviation:

Entry	Chemical name	Abbreviation
1	Petroleum ether	PE
2	Hexafluoroisopropyl alcohol	HFIP

<sup>&</sup>lt;sup>1</sup> (*a*) H. Wang, T. Jiang and M.-H. Xu, *J. Am. Chem. Soc.*, 2013, **135**, 971; (*b*) S. Zhang, L. Li, Y. Hu, Z. Zha, Z. Wang and T.-P. Loh, *Org. Lett.*, 2015, **17**, 1050.

<sup>&</sup>lt;sup>2</sup> (a) H. Amii, T. Kobayashi, Y. Hatamoto and K. Uneyama, Chem. Commun. 1999, 1323; (b) G. K. S. Prakash, J. Hu and G. A. Olah,

*J. Fluorine Chem.* 2001, **112**, 357; (*c*) É. Bélanger, K. Cantin, O. Messe, M. Tremblay and J.-F. Paquin, *J. Am. Chem. Soc.*, 2007, **129**, 1034; (*d*) J. Eames, G. S. Coumbarides, M. J. Suggate and N. Weerasooriya, *Eur. J. Org. Chem.* 2003, 634.

#### Part I. General procedure for the asymmetric Mukaiyama-Mannich reaction.



The Mukaiyama-Mannich reaction of difluoroenoxysilanes.

To a 5.0 mL vial were added hydroquinine derived bifunctional urea catalyst C3 (14.5 mg, 0.025 mmol) and cyclic *N*-sulfonyl ketimine 1 (0.25 mmol), followed by 1.25 mL of anhydrous toluene. The resulting mixture was stirred at 45 °C for about 10 min and then difluoroenoxysilanes 2 (0.50 mmol) was added. The resulting mixture was stirred at 45 °C for 4 days when 1 was almost full consumed by TLC analysis. Then the reaction mixture was directly subjected to flash column chromatography to afford the corresponding product 3, using indicated eluent.

Column chromatography (PE/EtOAc = 5:1 to 3:1) afforded product **3a** in 90% yield as white colloidal solid.<sup>3</sup> HPLC analysis (Chiralpak AD-H, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C; t<sub>r</sub> (minor) = 27.706 min, t<sub>r</sub> (major) = 15.889 min) gave the isomeric composition of the product: 93% ee,  $[\alpha]^{28}_{D}$  = -97.8 (c = 0.99, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.09-8.07 (m, 2H), 7.96-7.94 (m, 1H), 7.86-7.84 (m, 1H), 7.77-7.71 (m, 2H), 7.70-7.64 (m, 1H), 7.51-7.47 (m, 2H), 6.03 (s, 1H), 4.42-4.37 (m, 2H), 1.34 (t, *J* = 7.2 Hz, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -99.70 (d, *J* = 297 Hz, 1F), -105.70 (d, *J* = 297 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.73 (dd, *J* = 32 Hz, *J* = 29 Hz), 165.56 (dd, *J* = 7.3 Hz, *J* = 2.0 Hz), 136.53, 135.19, 133.57, 131.77, 131.10, 130.34 (dd, *J* = 3.7 Hz, *J* = 2.1 Hz), 130.26 (d, *J* = 1.4 Hz), 128.79, 126.98 (d, *J* = 5.6 Hz), 121.87, 115.37 (t, *J* = 268 Hz), 68.51 (dd, *J* = 26 Hz, *J* = 24 Hz), 64.36, 13.78.

<sup>&</sup>lt;sup>3</sup> J.-S.Yu and J. Zhou, Org. Biomol. Chem., 2015, 13, 10968.



Column chromatography (PE/EtOAc = 4:1) afforded product **3b** in 93% yield as white colloidal solid; HPLC analysis (Chiralpak AS-H,  $^{i}$ PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C;  $t_r$  (minor) = 24.952 min,  $t_r$  (major) = 15.804 min) gave the isomeric composition of the product: 94% ee,  $\left[\alpha\right]^{28}$  = -110.9 (c = 0.77, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.09-8.07 (m, 2H), 7.96-7.93 (m, 1H), 7.87-7.85 (m, 1H), 7.76-7.64 (m, 3H), 7.51-7.47 (m, 2H), 6.05 (s, 1H), 3.93 (s, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -99.71 (d, J = 298 Hz, 1F), -106.06 (d, J = 298 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.73 (dd, J = 32 Hz, J = 29 Hz), 166.16 (dd, J = 7.4 Hz, J = 2.0 Hz), 136.41, 135.27, 133.63, 131.82, 130.95 (t, J = 3.6 Hz), 130.36 (dd, J = 3.7 Hz, J = 2.1 Hz), 129.98 (d, J = 1.7 Hz), 128.80, 127.03 (d, J = 5.7 Hz), 121.86, 115.34(t, J = 268 Hz), 68.40 (dd, J = 26 Hz, J = 24 Hz), 54.67. IR (KBr): 3279, 1958, 1753, 1698, 1598, 1598, 1698, 1598, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 1698, 16988, 16988, 16988, 16988, 16988, 16988, 16988, 1698, 16988, 16988, 161451, 1321, 1259, 1180, 1097, 1032, 864, 763, 713, 576 cm<sup>-1</sup>. MS (EI): 381 (M<sup>+</sup>, 0.2), 322 (12), 274 (1), 226 (34), 166 (8), 105 (100), 77 (35), 51 (7). HRMS (EI): Exact mass calcd for C<sub>17</sub>H<sub>13</sub>NO<sub>5</sub>F<sub>2</sub>S [M]<sup>+</sup>: 381.0483, Found: 381.0485.

Column chromatography (PE/EtOAc = 4:1 to 2:1) afforded product 3c in 89% yield as white colloidal solid;<sup>3</sup> HPLC analysis (Chiralpak AD-H, <sup>i</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C;  $t_r$  (minor) = 24.750 min,  $t_r$  (major) = 17.218 min) gave the isomeric composition of the product: 92% ee,  $\left[\alpha\right]_{D}^{28} = -124.4$  (c = 1.05, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.09-8.07 (m, 2H), 7.73-7.72 (m, 2H), 7.67-7.64 (m, 1H), 7.52-7.47 (m, 3H), 6.01 (s, 1H), 4.43-4.36 (m, 2H), 2.52 (s, 3H), 1.34 (t, J = 7.2 Hz, 3H); <sup>19</sup>F NMR  $(376 \text{ MHz}, \text{CDCl}_3)$ :  $\delta$  -99.67 (d, J = 298 Hz, 1F), -105.87 (d, J = 298 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.72 (dd, J = 32 Hz, J = 29 Hz), 165.59 (dd, J = 7.4 Hz, J = 2.1 Hz), 144.83, 135.14, 133.83, 132.67, 131.07 (t, J = 3.4 Hz), 130.52 (d, J = 1.7 Hz), 130.30 (dd, J = 3.7 Hz, J = 2.1 Hz), 128.75, 127.03 (d, J = 5.5 Hz), 121.52, 115.36 (t, J = 268 Hz), 68.27 (dd, J = 25 Hz, J = 24 Hz), 64.23, 21.81, 13.76.

Column chromatography (PE/EtOAc = 4:1 to 2:1) afforded product **3d** in 87%EtO<sub>2</sub>C yield as pale yellow colloidal solid;<sup>3</sup> HPLC analysis (Chiralpak AD-H, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C;  $t_r$  (minor) = 35.989 min,  $t_r$ 

(major) = 19.864 min) gave the isomeric composition of the product: 94% ee,  $\left[\alpha\right]^{28}_{D}$  = -115.0 (c = 0.52, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.09-8.07 (m, 2H), 7.75 (d, J = 8.8 Hz, 1H), 7.68-7.64 (m, 1H), 7.49 (t, J = 8.0 Hz, 2H), 7.37-7.36 (m, 1H), 7.22-7.19 (m, 1H), 6.01 (s, 1H), 4.45-4.35 (m, 2H), 3.92 (s, 3H), 1.35 (t, J = 7.2 Hz, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -99.81 (d, J = 297 Hz, 1F), -106.05 (d, J = 297 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.80 (dd, J = 32 Hz, J = 29 Hz), 165.58 (dd, J = 7.4 Hz, J = 2.1 Hz), 163.81, 135.26, 132.81 (d, J = 1.7 Hz), 131.18 (t, J = 3.5 Hz), 130.41 (dd, J = 3.7 Hz, J = 2.0 Hz), 128.87, 128.53, 123.22, 118.30, 115.44 (t, J = 269 Hz), 111.41 (d, J = 5.8 Hz), 68.18 (dd, J = 25 Hz, J = 24 Hz), 64.39, 56.10, 13.90.



 $F_{3}CO + F_{3}CO + F_{NHO} + F_{NHO} + F_{NHO} + F_{3}CO + F_{NHO} + F_{3}CO + F_{NHO} + F_{3}CO + F_{NHO} + F_{N$ 25/75, 1.0 mL/min, 230 nm, 30 °C;  $t_r$  (minor) = 13.312 min,  $t_r$  (major) =

15.575 min) gave the isomeric composition of the product: 87% ee,  $[\alpha]_{D}^{28} = -82.4$  (c = 0.46, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.09-8.07 (m, 2H), 7.91 (d, *J* = 8.4 Hz, 1H), 7.78 (s, 1H), 7.70-7.66 (m, 1H), 7.58-7.56 (m, 1H), 7.53-7.49 (m, 2H), 6.15 (s, 1H), 4.50-4.35 (m, 2H), 1.35 (t, J = 7.2 Hz, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -57.88 (s, 3F, CF<sub>3</sub>), -99.70 (d, J = 297 Hz, 1F), -105.31 (d, J = 297 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.42 (dd, J = 32 Hz, J = 29 Hz), 164.97 (d, J = 7.1Hz), 152.69, 135.39, 134.82, 132.97 (d, J = 1.5 Hz), 130.94 (t, J = 3.6 Hz), 130.39 (dd, J = 3.6 Hz, J = 2.0 Hz, 128.88, 124.32, 123.77, 120.14 (g, J = 259 Hz), 119.17, 119.12, 115.26 (t, J = 269 Hz), 68.13 (dd, J = 26 Hz, J = 24 Hz), 64.78, 13.73. IR (KBr): 3275, 1752, 1701, 1598, 1475, 1259, 1185, 1099, 855, 711, 594 cm<sup>-1</sup>. MS (EI): 479 (M<sup>+</sup>, 0.7), 406 (6), 324 (10), 296 (5), 156 (7), 105 (100), 77 (29), 51 (4). HRMS (EI): Exact mass calcd for  $C_{19}H_{14}F_5NO_6S [M]^+$ : 479.0462, Found: 479.0455.



<sup>*i*</sup>PrOH/hexane = 25/75, 1.0 mL/min, 230 nm, 30 °C;  $t_r$  (minor) = 22.250 min,  $t_r$ 

(major) = 26.887 min) gave the isomeric composition of the product: 92% ee,  $[\alpha]^{28}_{D}$  = -98.6 (c = 0.65, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.09-8.07 (m, 2H), 7.88-7.84 (m, 1H), 7.69-7.61 (m, 2H), 7.53-7.49 (m, 2H), 7.45-7.40 (m, 1H), 6.10 (s, 1H), 4.42 (q, J = 6.8 Hz, 2H), 1.36 (t, J = 6.8

Hz, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -99.73 (d, *J* = 297 Hz, 1F), -102.62 (s, 1F), -105.53 (d, *J* = 297 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.48 (dd, *J* = 32 Hz, *J* = 29 Hz), 165.40 (d, *J* = 255 Hz), 165.06 (d, *J* = 7.2 Hz), 135.33, 133.35 (dd, *J* = 9.9 Hz, *J* = 1.8 Hz), 132.66 (d, *J* = 2.9 Hz), 130.97 (t, *J* = 2.8 Hz), 130.36 (dd, *J* = 3.7 Hz, *J* = 2.1 Hz), 128.85, 124.07 (d, *J* = 9.9 Hz), 119.83 (d, *J* = 24 Hz), 115.27 (t, *J* = 269 Hz), 114.415 (dd, *J* = 26 Hz, *J* = 5.7 Hz), 68.04 (t, *J* = 26 Hz), 64.69, 13.79. IR (KBr): 3276, 1751, 1700, 1596, 1481, 1326, 1258, 1183, 1098, 820, 725 cm<sup>-1</sup>. MS (EI): 340 (7), 258 (11), 249 (4), 195 (27), 156 (6), 105 (95), 103 (100), 77 (33). HRMS (EI): Exact mass calcd for C<sub>18</sub>H<sub>14</sub>NO<sub>5</sub>F<sub>3</sub>S [M]<sup>+</sup>: 413.0545, Found: 413.0554.

Column chromatography (PE/EtOAc = 4:1) gave **3g** in 74% yield as pale white colloidal solid,<sup>3</sup> HPLC analysis (Chiralpak AS-H, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C; t<sub>r</sub> (minor) = 23.453 min, t<sub>r</sub> (major) = 12.626 min) gave the isomeric composition of the product: 92% ee,  $[\alpha]^{28}_{D}$  = -108.8 (c = 0.60, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.09-8.07 (m, 2H), 7.92-7.91 (m, 1H), 7.81-7.79 (m, 1H), 7.71-7.66 (m, 2H), 7.53-7.49 (m, 2H), 6.08 (s, 1H), 4.42 (q, *J* = 7.2 Hz, 2H), 1.36 (t, *J* = 7.2 Hz, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -99.52 (d, *J* = 298 Hz, 1F), -105.41 (d, *J* = 298 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.45 (dd, *J* = 32 Hz, *J* = 29 Hz), 165.02 (dd, *J* = 7.2 Hz, *J* = 2.0 Hz), 140.15, 135.33, 135.11, 132.30, 132.25 (d, *J* = 1.8 Hz), 130.95 (t, *J* = 3.6 Hz), 130.37 (dd, *J* = 3.6 Hz, *J* = 2.1 Hz), 128.86, 127.16 (d, *J* = 5.9 Hz), 123.02, 115.27 (t, *J* = 269 Hz), 68.11 (dd, *J* = 26 Hz, *J* = 24 Hz), 64.69, 13.80.

 $F_{3}C$   $F_{0}C$   $F_{0}C$  F

isomeric composition of **3h**: 84% ee,  $[\alpha]^{28}{}_{D}$  = -66.2 (c = 0.90, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.21-8.20 (m, 1H), 8.09-8.07 (m, 2H), 8.01 (s, 2H), 7.70-7.66 (m, 1H), 7.53-7.49 (m, 2H), 6.20 (s, 1H), 4.51-4.36 (m, 2H), 1.35 (t, *J* = 7.2 Hz, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -62.82 (s, 3F, *CF*<sub>3</sub>), -99.41 (d, *J* = 299 Hz, 1F), -105.09 (d, *J* = 299 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.37 (dd, *J* = 31 Hz, *J* = 29 Hz), 164.90 (dd, *J* = 7.2 Hz, *J* = 1.9 Hz), 139.85, 135.70 (q, *J* = 33 Hz), 135.42, 131.45 (d, *J* = 1.7 Hz), 130.89 (t, *J* = 3.6 Hz), 130.39 (dd, *J* = 3.6 Hz, *J* = 2.3 Hz), 129.13 (q, *J* = 3.5 Hz), 128.90, 124.41 (q, J = 3.9 Hz), 122.89, 122.79 (q, J = 272 Hz), 115.28 (t, J = 269 Hz), 68.41 (dd, J = 26 Hz, J = 24 Hz), 64.81, 13.75. IR (KBr): 3277, 1752, 1702, 1371, 1329, 1178, 1031, 850, 715 cm<sup>-1</sup>. MS (EI): 463 (M<sup>+</sup>, 0.6), 390 (5), 308 (7), 234 (4), 156 (11), 105 (100), 77 (29), 43 (13). HRMS (EI): Exact mass calcd for C<sub>19</sub>H<sub>14</sub>NO<sub>5</sub>F<sub>5</sub>S [M]<sup>+</sup>: 463.0513, Found: 463.0511.

Column chromatography (PE/EtOAc = 4:1 to 3:1) afforded product **3i** in 98% yield as pale white colloidal solid; HPLC analysis (Chiralpak AD-H, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C; t<sub>r</sub>(minor) = 13.557 min, t<sub>r</sub>(major) = 16.901 min) gave the isomeric composition of the product: 90% ee,  $[\alpha]^{28}_{D}$  = -89.8 (c = 0.86, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.09-8.07 (m, 2H), 7.86-7.84 (m, 1H), 7.70-7.64 (m, 3H), 7.52-7.49 (m, 2H), 6.11 (s, 1H), 4.43-4.38 (m, 2H), 1.35 (t, *J* = 7.2 Hz, 3H); <sup>19</sup>F NMR (282 MHz, CDCl<sub>3</sub>):  $\delta$  -99.30 (d, *J* = 298 Hz, 1F,), -105.44 (d, *J* = 298 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  187.54 (dd, *J* = 30 Hz, *J* = 29 Hz), 165.17 (dd, *J* = 7.4 Hz, *J* = 2.1 Hz), 135.31, 134.64, 134.58. 132.91 (d, *J* = 1.7 Hz), 132.60, 131.03 (t, *J* = 3.6 Hz), 130.37 (dd, *J* = 3.8 Hz, *J* = 2.1 Hz), 129.61, 128.86, 125.28 (d, *J* = 5.8 Hz), 115.31 (t, *J* = 269 Hz), 67.54 (dd, *J* = 26 Hz, *J* = 24 Hz), 64.63, 13.81. IR (KBr): 3305, 1739, 1698, 1455, 1262, 1182, 1095, 843, 778, 589 cm<sup>-1</sup>. MS (EI): 429 (M<sup>+</sup>, 0.3), 356 (9), 274 (16), 246 (6), 156 (4), 105 (100), 77 (30), 51 (5). HRMS (EI): Exact mass calcd for C<sub>18</sub>H<sub>14</sub>NO<sub>5</sub>F<sub>2</sub>S<sup>35</sup>Cl [M]<sup>+</sup>: 429.0249, Found: 429.0251.

Column chromatography (PE/EtOAc = 5:1 to 3:1) afforded product **3j** in 75% yield as pale white foamy solid;<sup>3</sup> HPLC analysis (Chiralpak AD-H,  $^{i}$ PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C; t<sub>r</sub> (minor) = 39.292 min, t<sub>r</sub> (major) = 20.875 min) gave the isomeric composition of the product: 92% ee,  $[\alpha]^{28}_{D}$  = -101.5 (c = 1.00, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.09-8.07 (m, 2H), 7.96-7.93 (m, 1H), 7.87-7.85 (m, 1H), 7.75-7.72 (m, 2H), 6.98-6.94 (m, 2H), 6.00 (s, 1H), 4.41-4.34 (m, 2H), 3.89 (s, 3H), 1.34 (t, J = 7.2 Hz, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -99.29 (d, J = 296 Hz, 1F), -105.08 (dd, J = 296 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  185.86 (dd, J = 31 Hz, J = 29 Hz), 165.68 (dd, J = 7.4 Hz, J = 2.0 Hz), 165.23, 136.57, 133.51, 133.07 (dd, J = 3.9 Hz, J = 1.9 Hz), 131.70, 130.44 (d, J = 1.8 Hz), 127.03 (d, J = 5.7 Hz), 123.95 (dd, J = 3.8 Hz, J = 2.8 Hz), 121.87, 115.74 (t, J = 268 Hz), 114.19, 68.54 (dd, J = 26 Hz, J = 24 Hz), 64.24, 55.63, 13.81.

Column chromatography (PE/EtOAc = 4:1 to 3:1) gave **3k** in 74% yield as pale white colloidal solid;<sup>3</sup> HPLC analysis (Chiralpak AD-H, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C; t<sub>r</sub> (minor) = 24.700 min, t<sub>r</sub> (major) = 15.211 min) gave the isomeric composition of the product: 93% ee,  $[\alpha]^{28}_{D}$  = -100.8 (c = 1.08, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.04-8.01 (m, 2H), 7.96-7.93 (m, 1H), 7.87-7.85 (m, 1H), 7.78-7.73 (m, 2H), 7.49-7.46 (m, 2H), 6.04 (s, 1H), 4.43-4.37 (m, 2H), 1.34 (t, *J* = 7.2 Hz, 3H); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>):  $\delta$  -99.87 (d, *J* = 297 Hz, 1F), -105.86 (d, *J* = 297 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  186.73 (dd, *J* = 32 Hz, *J* = 29 Hz), 165.40 (dd, *J* = 7.3 Hz, *J* = 2.0 Hz), 142.01, 136.51, 133.61, 131.84, 131.72 (dd, *J* = 3.9 Hz, *J* = 2.1 Hz), 130.08 (d, *J* = 1.7 Hz), 129.47 (t, *J* = 3.0 Hz), 129.23, 126.95 (d, *J* = 5.4 Hz), 121.90, 115.25 (t, *J* = 267 Hz), 68.42 (dd, *J* = 26 Hz, *J* = 24 Hz), 64.46, 13.78.

#### The Mukaiyama-Mannich reaction of monofluorinated enol silyl ethers



To a 5.0 mL vial were added hydroquinine derived bifunctional urea catalyst C3 (14.5 mg, 0.025 mmol) and cyclic *N*-sulfonyl ketimine 1 (0.25 mmol), followed by 1.25 mL of anhydrous toluene. The resulting mixture was stirred at 35 °C for about 10 min and then mono-fluorinated enol silyl ether 4 (0.375 mmol) was added. The resulting mixture was stirred at 35 °C till full conversion of 1 by TLC analysis. The reaction mixturing was directly subjected to flash column chromatography to afford the corresponding product 5, using indicated eluent.

Column chromatography (PE/EtOAc = 5:1 to 3:1) gave **5a** in 99% yield as white solid (m.p. = 146-148 °C), dr value was determined by <sup>1</sup>H NMR or <sup>19</sup>F NMR of crude mixture. HPLC analysis (Chiralpak AD-H, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C; t<sub>r</sub> (minor) = 18.152 min, t<sub>r</sub> (major) = 9.277 min) gave the isomeric composition of the product: 90% ee,  $[\alpha]^{28}_{D}$  = +19.4 (c = 0.72, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.92-7.89 (m, 1H), 7.78-7.77 (m, 2H), 7.69-7.63 (m, 3H), 7.44-7.40 (m, 2H), 6.21 (s, 1H), 4.42-4.28 (m, 2H), 3.63 (dd, *J* = 18 Hz, *J* = 15.6 Hz, 1H), 3.37 (dd, *J* = 24.4 Hz, *J* = 16.4 Hz, 1H), 1.31 (t. J = 7.2 Hz, 3H); <sup>19</sup>F NMR (282 MHz, CDCl<sub>3</sub>);  $\delta$  -158.71 (s. 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  196.18 (d, J = 18 Hz), 167.63, 149.59 (d, J = 2 Hz), 136.55, 135.72, 134.45, 133.12, 131.74, 131.12, 128.59, 127.33 (d, J = 5 Hz), 126.14, 125.31, 121.44, 95.74 (d, J = 201 Hz), 69.72 (d, J = 27 Hz), 64.22, 36.84 (d, J = 24 Hz), 13.83. IR (KBr): 3289, 3001, 1724, 1605, 1467, 1360, 1314, 1174, 1024, 888, 739, 577 cm<sup>-1</sup>. MS (EI): 389 (M<sup>+</sup>, 0.5), 316 (100), 296 (6), 150 (80), 130 (14), 103 (52). HRMS (EI): Exact mass calcd for C<sub>19</sub>H<sub>16</sub>NO<sub>5</sub>FS [M]<sup>+</sup>: 389.0733, Found: 389.0735.



EtO<sub>2</sub>

Column chromatography (PE/EtOAc = 5:1 to 3:1) afforded product **5b** in 78% yield as white solid (m.p. = 68-70 °C), dr value was determined by  ${}^{1}$ H NMR or <sup>19</sup>F NMR of crude reaction mixture. HPLC analysis (Chiralpak AD-H, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C; t<sub>r</sub> (minor) = 19.698 min, t<sub>r</sub> (major) = 10.810 min) gave the isomeric composition of the product: 92% ee,  $\left[\alpha\right]_{D}^{28} = +14.7$  (c = 0.49, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.79-7.77 (m, 1H), 7.69-7.63 (m, 2H), 7.44-7.40 (m, 2H), 7.30-7.29 (m, 1H), 7.16-7.13 (m, 1H), 6.11 (s, 1H), 4.41-4.33 (m, 2H), 3.86 (s, 3H), 3.58 (t, J = 17.6 Hz, 1H), 3.36 (dd, J = 24.8 Hz, J = 18 Hz, 1H), 1.34 (t, J = 7.2 Hz, 3H); <sup>19</sup>F NMR (282 MHz, CDCl<sub>3</sub>):  $\delta$  -158.71 (s, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  196.01 (d, J = 18 Hz), 167.53, 163.34, 149.60 (d, J = 3 Hz), 136.50, 134.57, 134.16, 128.57, 127.87, 126.15, 125.23, 122.69, 117.75, 111.80 (d, J = 5 Hz), 95.97 (d, J = 201 Hz), 69.31 (d, J = 26 Hz), 64.19, 55.87, 36.87 (d, J = 24 Hz), 13.88. IR (KBr): 3415, 1729, 1602, 1485, 1306, 1259, 1169, 1021 cm<sup>-1</sup>. MS (EI): 346 (54), 309 (5),

195 (24), 150 (36), 133 (41), 103 (100). HRMS (EI): Exact mass calcd for C<sub>20</sub>H<sub>18</sub>NO<sub>6</sub>FS [M]<sup>+</sup>: 419.0839, Found: 419.0836.

> Column chromatography (PE/EtOAc = 5:1 to 3:1) afforded product **5c** in 88% yield as white solid (m.p. = 76-78  $^{\circ}$ C), dr value was determined by <sup>1</sup>H NMR or

<sup>19</sup>F NMR of crude reaction mixture. HPLC analysis (Chiralpak AD-H, 5c <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C;  $t_r$  (minor) = 19.178 min,  $t_r$  (major) = 9.397 min) gave the isomeric composition of the product: 90% ee,  $[\alpha]^{28}_{D} = +18.5$  (c = 0.61, CHCl<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.91-7.89 (m, 1H), 7.80-7.78 (m, 1H), 7.71-7.66 (m, 2H), 7.57 (s, 1H), 7.48-7.45 (m, 1H), 7.31-7.29 (m, 1H), 6.24 (s, 1H), 4.41-4.27 (m, 2H), 3.58 (t, *J* = 17.6 Hz, 1H), 3.32 (dd, *J* = 24.8 Hz, *J* = 18 Hz, 1H), 2.39 (s, 3H), 1.30 (t, *J* = 7.2 Hz, 3H); <sup>19</sup>F NMR (282 MHz, CDCl<sub>3</sub>):  $\delta$  -158.51 (s, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  196.23 (d, *J* = 18 Hz), 167.66, 147.02 (d, *J* = 3 Hz), 138.74, 137.85, 135.71, 134.55, 133.08, 131.83, 131.06, 127.30 (d, *J* = 5 Hz), 125.82, 125.12, 121.40, 95.98 (d, *J* = 200 Hz), 69.80 (d, *J* = 27 Hz), 64.14, 36.53 (d, *J* = 24 Hz), 21.03, 13.81. IR (KBr): 3264, 2932, 1728, 1369, 1312, 1253, 1176, 1067, 758, 578 cm<sup>-1</sup>. MS (EI): 330 (9), 310 (3), 195 (24), 164 (11), 103 (100), 76 (16). HRMS (EI): Exact mass calcd for C<sub>20</sub>H<sub>18</sub>NO<sub>5</sub>FS [M]<sup>+</sup>: 403.0890, Found: 403.0888.

Column chromatography (PE/EtOAc = 2:1) afforded product **9** in 24% yield as white solid.<sup>4</sup> HPLC analysis (Chiralpak IC, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C; t<sub>r</sub> (minor) = 30.409 min, t<sub>r</sub> (major) = 20.532 min) gave the isomeric composition of the product: 32% ee; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.93-7.91 (m, 2H), 7.85-7.83 (m, 1H), 7.72-7.71 (m, 2H), 7.67-7.59 (m, 2H), 7.49-7.45 (m, 2H), 6.08 (s, 1H), 4.38-4.27 (m, 2H), 4.09 (d, *J* = 17.6 Hz, 1H), 3.73 (d, *J* = 17.6 Hz, 1H), 1.31 (t, *J* = 6.8 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  195.92, 169.58, 136.87, 135.52, 134.01, 133.64, 130.80, 128.78, 128.17, 124.20, 121.88, 65.43, 63.50, 49.13, 13.94.

<sup>&</sup>lt;sup>4</sup> S. Nakamura, M. Sano, A. Toda, D. Nakane and H. Masuda, *Chem. Eur. J.*, 2015, **21**, 3929.

#### Part II. Synthetic elaborations.



The product 3i (107.0 mg, 0.25 mmol) was dissolved in CH<sub>2</sub>Cl<sub>2</sub>/HFIP (2:1, 3.0 mL), followed by the addition of *m*-chloroperoxybenzoic acid (*m*-CPBA) (276.0 mg, 1.25 mmol, 85% wt) and phosphate buffer (0.25 mL, pH = 7.6) at ambient temperature.<sup>5</sup> The mixture was stirred until full consumption of **3j** by TLC analysis (ca. 5 h), and then quenched by saturated Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> (aq), followed by extraction using  $CH_2Cl_2$  (10.0 mL  $\times$  3). The combined organic layers were washed with saturated NaHCO<sub>3</sub> and brine (10.0 mL  $\times$  3), and then dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The residue was purified by column chromatography (using PE/EtOAc = 2/1 as eluent) to afford 6 (107.0 mg, 97% yield) as pale yellow oil. HPLC analysis (Chiralpak AD-H, <sup>*i*</sup>PrOH/hexane = 40/60, 1.0 mL/min, 230 nm, 30 °C;  $t_r$  (minor) = 20.938 min,  $t_r$  (major) = 13.596 min) gave the isomeric composition of the product: 91% ee,  $[\alpha]^{25}_{D} = -54.8$  (c = 1.0, CHCl<sub>3</sub>); IR (KBr): 3264, 2966, 1793, 1748, 1505, 1455, 1324, 1251, 1108, 1030, 861, 761, 573 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 8.03-8.01 (m, 1H), 7.84-7.82 (m, 1H), 7.78-7.71 (m, 2H), 7.08-7.04 (m, 2H), 6.90-6.85 (m, 2H), 6.09 (s, 1H), 4.49-4.36 (m, 2H), 3.78 (s, 3H), 1.37 (t, J = 7.2 Hz, 3H); <sup>19</sup>F NMR (282 MHz, CDCl<sub>3</sub>):  $\delta$  -107.88 (d, J = 266 Hz, 1F), -109.81 (d, J = 266 Hz, 1F); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.91 (d, J = 3 Hz), 160.09 (t, J = 32 Hz), 158.04, 142.98, 135.90, 133.77, 131.95, 127.23 (t, J = 3 Hz), 121.80, 121.59, 114.63, 112.95 (t, J = 264 Hz), 69.12 (dd, J = 27 Hz, J = 26Hz), 65.01, 55.58, 13.86. MS (EI): 441 (M<sup>+</sup>, 5), 368 (3), 262 (2), 124 (53), 105 (55), 85 (72), 83 (100), 43 (22); HRMS (EI): Exact mass calcd for  $C_{19}H_{17}NO_7SF_2$  [M]<sup>+</sup>: 441.0694, Found: 441.0692.

<sup>&</sup>lt;sup>5</sup> S. Kobayashi, H. Tanaka, H. Amii and K. Uneyama, *Tetrahedron* 2003, **59**, 1547.



To a 25.0 mL Schlenk tube was added product **3a** (100 mg, 0.253 mmol), followed by 3.0 mL anhydrous EtOH under N2 atmosphere. The mixture was cooled to -20 °C, and then 0.6 mL CH<sub>3</sub>NH<sub>2</sub> in ethanol (27-30% wt) was added dropwise at -20 °C. The mixture was stirred at room temperature until full consumption of 3a by TLC analysis (about 5 h). Then the reaction mixture was concentrated under reduced pressure, and the residue was purified by column chromatography (PE/ethyl acetate = 1/1) to afford product 7 as a single isomer and white solid in 63% yield (60.7 mg). M. p. 240-242 °C. HPLC analysis (Chiralpak AD-H, <sup>*i*</sup>PrOH/hexane = 40/60, 0.8 mL/min, 230 nm,  $t_r$  (minor) = 10.973 min,  $t_r$  (major) = 8.275 min) gave the isomeric composition of the product: 92% ee,  $[\alpha]^{25}_{D}$  = +8.4 (c = 0.55, acetone); <sup>1</sup>H NMR (400 MHz, acetone-*d*<sub>6</sub>):  $\delta$  7.80-7.76 (m, 1H), 7.69-7.64 (m, 2H), 7.61-7.58 (m, 2H), 7.54-7.50 (m, 1H), 7.44-7.36 (m, 3H), 7.04 (s, br, 1H), 2.73 (s, 4H); <sup>19</sup>F NMR (282 MHz, acetone- $d_6$ ):  $\delta$  -109.41 (d, J = 230 Hz, 1F), -128.38 (d, J = 230 Hz, 1F); <sup>13</sup>C NMR (100 MHz, acetone-*d*<sub>6</sub>): δ 169.30 (d, J = 6 Hz), 138.69, 133.88, 133.75, 132.17, 132.04, 130.61, 129.40 (d, J = 2.5 Hz), 129.29, 128.74 (d, J = 7 Hz), 121.72, 118.51 (dd, J = 273 Hz, J = 120.40253 Hz), 92.66 (dd, J = 29 Hz, J = 24 Hz), 67.60 (dd, J = 29 Hz, J = 19 Hz), 27.31; IR (KBr): 3375, 1737, 1452, 1419, 1364, 1302, 1243, 1158, 1113, 1063, 768, 754, 526 cm<sup>-1</sup>; HRMS (ESI): Exact mass calcd for  $C_{17}H_{18}F_2N_3O_4S[M+NH_4]^+$ : 398.0981, Found: 398.0981.

## Part III. X-ray crystal data of 5a.<sup>6</sup>



Data intensity of C<sub>19</sub>H<sub>16</sub>FNO<sub>5</sub>S (**5a**) was collected using a Bruker SMART APEX II (Mo radiation) at 296 K in a nitrogen stream. Data collection and reduction were done by using the Bruker ApexII software package. The structures were solved by direct methods and refined by full-matrix least-squares on  $F^2$  with anisotropic displacement parameters for non-H atoms using SHELX-97. Hydrogen atoms were added at their geometrically idea positions and refined isotropically. Crystal data for **5a**: C<sub>19</sub>H<sub>16</sub>FNO<sub>5</sub>S, T = 296(2) K, Orthorhombic, P2(1)2(1)2(1) a = 10.342(2) Å, b = 12.672(3) Å, c = 13.549(3) Å, alpha = 90 deg, beta = 90 deg, gamma = 90 deg.  $V = 1775.6(7) Å^3$ . Z = 4,  $d_{calc} = 1.457$  mg/m<sup>3</sup>. Total number of reflections 20679 [R(int) = 0.0282], R<sub>1</sub> = 0.0294, wR<sub>2</sub> = 0.0739 (all data), GOF = 1.029, and 244 parameter.

Table 1. Crystal data and structure refinement for z.

Identification code	Z	
Empirical formula	$C_{19}H_{16}FNO_5S$	
Formula weight	389.39	
Temperature	296(2) K	
Wavelength	0.71073 Å	
Crystal system, space group	Orthorhombic, P2(1	)2(1)2(1)
Unit cell dimensions	a = 10.342(2)  Å	alpha = 90 deg.
	b = 12.672(3) Å	beta = 90 deg.
	c = 13.549(3)  Å	gamma = 90 deg

<sup>&</sup>lt;sup>6</sup> Supplementary crystallographic data have been deposited at the Cambridge Crystallographic Data Center. (CCDC 1439365).

Volume	1775.6(7) Å <sup>3</sup>
Z, Calculated density	4, 1.457 Mg/m <sup>3</sup>
Absorption coefficient	0.224 mm <sup>-1</sup>
F(000)	808
Crystal size	0.46 x 0.32 x 0.26 mm
Theta range for data collection	2.20 to 24.99 deg.
Limiting indices	-12<=h<=12, -15<=k<=15, -16<=l<=16
Reflections collected / unique	20679 / 3128 [R(int) = 0.0282]
Completeness to theta $= 24.99$	100.0 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.9441 and 0.9040
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	3128 / 0 / 244
Goodness-of-fit on F <sup>2</sup>	1.029
Final R indices [I>2sigma(I)]	$R_1 = 0.0273, wR_2 = 0.0720$
R indices (all data)	$R_1 = 0.0294, wR_2 = 0.0739$
Absolute structure parameter	0.00(6)
Largest diff. peak and hole	$0.254 \text{ and } -0.220 \text{ e.A}^{-3}$

Table 2. Atomic coordinates (  $x \ 10^4$ ) and equivalent isotropic displacement parameters (A<sup>2</sup>  $x \ 10^3$ ) for z. U(eq) is defined as one third of the trace of the orthogonalized Uij tensor.

	Х	У	Z	U(eq)
S(1)	9204(1)	9407(1)	763(1)	42(1)
O(1)	9824(2)	8747(1)	53(1)	66(1)
O(2)	9627(2)	9312(1)	1762(1)	65(1)
O(3)	7761(1)	11865(1)	1769(1)	39(1)
O(4)	8190(1)	12998(1)	537(1)	43(1)
O(5)	10200(1)	11982(1)	-1093(1)	51(1)
N(1)	9273(2)	10638(1)	390(1)	38(1)
F(1)	8059(1)	10538(1)	-1465(1)	46(1)
C(1)	7517(2)	9349(1)	707(1)	36(1)
C(2)	6766(2)	8474(2)	927(1)	46(1)
C(3)	5449(2)	8572(2)	856(2)	50(1)
C(4)	4902(2)	9527(2)	584(2)	50(1)
C(5)	5661(2)	10397(2)	370(1)	41(1)
C(6)	6994(2)	10304(1)	425(1)	33(1)
C(7)	8010(2)	11137(1)	181(1)	31(1)

C(8)	7932(2)	11467(1)	-916(1)	33(1)
C(9)	6708(2)	12050(2)	-1235(1)	38(1)
C(10)	7187(2)	12932(1)	-1889(1)	37(1)
C(11)	6457(2)	13641(2)	-2432(2)	49(1)
C(12)	7103(3)	14390(2)	-2993(2)	58(1)
C(13)	8438(3)	14440(2)	-3012(2)	56(1)
C(14)	9176(2)	13747(2)	-2466(1)	46(1)
C(15)	8524(2)	12994(1)	-1902(1)	37(1)
C(16)	9068(2)	12168(1)	-1280(1)	36(1)
C(17)	7970(2)	12124(1)	840(1)	31(1)
C(18)	7855(2)	12716(2)	2494(1)	45(1)
C(19)	7865(2)	12216(2)	3494(2)	56(1)

 Table 3.
 Bond lengths [Å] and angles [deg] for z.

S(1)-O(1)	1.4275(17)
S(1)-O(2)	1.4282(16)
S(1)-N(1)	1.6408(16)
S(1)-C(1)	1.7484(19)
O(3)-C(17)	1.318(2)
O(3)-C(18)	1.462(2)
O(4)-C(17)	1.203(2)
O(5)-C(16)	1.220(2)
N(1)-C(7)	1.478(2)
N(1)-H(1A)	0.8600
F(1)-C(8)	1.399(2)
C(1)-C(6)	1.379(3)
C(1)-C(2)	1.386(3)
C(2)-C(3)	1.371(3)
C(2)-H(2A)	0.9300
C(3)-C(4)	1.386(3)
C(3)-H(3A)	0.9300
C(4)-C(5)	1.384(3)
C(4)-H(4A)	0.9300
C(5)-C(6)	1.386(3)
C(5)-H(5A)	0.9300
C(6)-C(7)	1.526(2)
C(7)-C(17)	1.537(2)
C(7)-C(8)	1.546(2)
C(8)-C(9)	1.528(3)
C(8)-C(16)	1.553(2)
C(9)-C(10)	1.510(3)
C(9)-H(9A)	0.9700

0.9700
1.384(3)
1.384(3)
1.388(3)
0.9300
1.383(4)
0.9300
1.379(3)
0.9300
1.396(3)
0.9300
1.457(3)
1.495(3)
0.9700
0.9700
0.9600
0.9600
0.9600
116.87(11)
109.30(10)
111.03(10)
113.23(10)
110.08(10)
94.02(9)
116.61(14)
115.31(12)
122.3
122.3
122.82(18)
111.46(14)
125.71(15)
118.00(18)
121.0
121.0
120.16(19)
119.9
119.9
121.3(2)
119.3
119.3
119.04(19)
120.5

C(6)-C(5)-H(5A)	120.5
C(1)-C(6)-C(5)	118.63(16)
C(1)-C(6)-C(7)	113.42(16)
C(5)-C(6)-C(7)	127.93(17)
N(1)-C(7)-C(6)	105.69(13)
N(1)-C(7)-C(17)	105.12(13)
C(6)-C(7)-C(17)	114.73(14)
N(1)-C(7)-C(8)	110.29(14)
C(6)-C(7)-C(8)	111.05(14)
C(17)-C(7)-C(8)	109.67(14)
F(1)-C(8)-C(9)	109.53(14)
F(1)-C(8)-C(7)	106.14(13)
C(9)-C(8)-C(7)	116.54(14)
F(1)-C(8)-C(16)	103.98(13)
C(9)-C(8)-C(16)	105.11(14)
C(7)-C(8)-C(16)	114.82(15)
C(10)-C(9)-C(8)	104.58(15)
C(10)-C(9)-H(9A)	110.8
C(8)-C(9)-H(9A)	110.8
C(10)-C(9)-H(9B)	110.8
C(8)-C(9)-H(9B)	110.8
H(9A)-C(9)-H(9B)	108.9
C(15)-C(10)-C(11)	120.07(18)
C(15)-C(10)-C(9)	112.17(16)
C(11)-C(10)-C(9)	127.76(18)
C(10)-C(11)-C(12)	118.2(2)
C(10)-C(11)-H(11A)	120.9
C(12)-C(11)-H(11A)	120.9
C(13)-C(12)-C(11)	121.5(2)
C(13)-C(12)-H(12A)	119.3
C(11)-C(12)-H(12A)	119.3
C(14)-C(13)-C(12)	120.9(2)
C(14)-C(13)-H(13A)	119.6
C(12)-C(13)-H(13A)	119.6
C(13)-C(14)-C(15)	117.5(2)
C(13)-C(14)-H(14A)	121.2
C(15)-C(14)-H(14A)	121.2
C(10)-C(15)-C(14)	121.87(19)
C(10)-C(15)-C(16)	109.73(16)
C(14)-C(15)-C(16)	128.38(19)
O(5)-C(16)-C(15)	128.96(17)
O(5)-C(16)-C(8)	123.35(16)
C(15)-C(16)-C(8)	107.58(15)

O(4)-C(17)-O(3)	125.85(16)
O(4)-C(17)-C(7)	123.08(16)
O(3)-C(17)-C(7)	110.89(14)
O(3)-C(18)-C(19)	107.29(17)
O(3)-C(18)-H(18A)	110.3
C(19)-C(18)-H(18A)	110.3
O(3)-C(18)-H(18B)	110.3
C(19)-C(18)-H(18B)	110.3
H(18A)-C(18)-H(18B)	108.5
C(18)-C(19)-H(19B)	109.5
C(18)-C(19)-H(19C)	109.5
H(19B)-C(19)-H(19C)	109.5
C(18)-C(19)-H(19D)	109.5
H(19B)-C(19)-H(19D)	109.5
H(19C)-C(19)-H(19D)	109.5

Symmetry transformations used to generate equivalent atoms: Table 4. Anisotropic displacement parameters ( $Å^2 \times 10^3$ ) for z. The anisotropic displacement factor exponent takes the form: -2 pi<sup>2</sup> [  $h^2 a^{*2} U11 + ... + 2 h k a^* b^* U12$  ]

	U11	U22	U33	U23	U13	U12
S(1)	46(1)	35(1)	44(1)	4(1)	-4(1)	8(1)
O(1)	68(1)	46(1)	84(1)	-9(1)	11(1)	18(1)
O(2)	68(1)	70(1)	57(1)	24(1)	-21(1)	-3(1)
O(3)	53(1)	33(1)	30(1)	-1(1)	0(1)	-3(1)
O(4)	56(1)	32(1)	41(1)	2(1)	5(1)	-8(1)
O(5)	33(1)	59(1)	61(1)	14(1)	7(1)	2(1)
N(1)	33(1)	36(1)	45(1)	4(1)	-1(1)	2(1)
F(1)	62(1)	37(1)	39(1)	-9(1)	3(1)	-3(1)
C(1)	48(1)	32(1)	28(1)	-1(1)	-1(1)	0(1)
C(2)	70(1)	32(1)	38(1)	4(1)	2(1)	-4(1)
C(3)	62(1)	41(1)	48(1)	0(1)	9(1)	-18(1)
C(4)	44(1)	53(1)	52(1)	-3(1)	7(1)	-12(1)
C(5)	40(1)	37(1)	46(1)	0(1)	6(1)	-2(1)
C(6)	41(1)	30(1)	28(1)	-2(1)	3(1)	-4(1)
C(7)	31(1)	30(1)	33(1)	1(1)	0(1)	0(1)
C(8)	37(1)	30(1)	32(1)	-2(1)	1(1)	0(1)
C(9)	36(1)	43(1)	35(1)	4(1)	-3(1)	-3(1)
C(10)	44(1)	37(1)	29(1)	-2(1)	-2(1)	0(1)
C(11)	54(1)	48(1)	45(1)	5(1)	-3(1)	6(1)
C(12)	84(2)	45(1)	46(1)	10(1)	0(1)	13(1)
C(13)	84(2)	40(1)	44(1)	8(1)	14(1)	-5(1)

C(14)	58(1)	41(1)	39(1)	0(1)	9(1)	-6(1)
C(15)	45(1)	35(1)	30(1)	-2(1)	4(1)	-2(1)
C(16)	37(1)	36(1)	34(1)	-3(1)	6(1)	-1(1)
C(17)	26(1)	33(1)	35(1)	0(1)	-2(1)	-1(1)
C(18)	56(1)	42(1)	37(1)	-9(1)	-3(1)	-3(1)
C(19)	61(1)	71(1)	35(1)	-4(1)	-2(1)	-3(1)

Table 5. Hydrogen coordinates (  $x \ 10^4$ ) and isotropic displacement parameters (Å<sup>2</sup>  $x \ 10^3$ ) for z.

	Х	У	Z	U(eq)
H(1A)	9993	10969	322	45
H(2A)	7144	7840	1117	56
H(3A)	4921	7995	990	60
H(4A)	4007	9585	544	60
H(5A)	5282	11035	192	49
H(9A)	6134	11584	-1597	46
H(9B)	6250	12329	-668	46
H(11A)	5559	13615	-2420	59
H(12A)	6627	14869	-3365	70
H(13A)	8844	14948	-3399	67
H(14A)	10074	13780	-2473	55
H(18A)	7122	13190	2432	54
H(18B)	8642	13118	2392	54
H(19B)	7923	12755	3990	83
H(19C)	8595	11752	3548	83
H(19D)	7082	11820	3586	83

Table 6.Torsion angles [deg] for z.

O(1)-S(1)-N(1)-C(7)	116.24(14)
O(2)-S(1)-N(1)-C(7)	-113.40(14)
C(1)-S(1)-N(1)-C(7)	-0.09(13)
O(1)-S(1)-C(1)-C(6)	-114.89(15)
O(2)-S(1)-C(1)-C(6)	112.23(14)
N(1)-S(1)-C(1)-C(6)	-1.90(14)
O(1)-S(1)-C(1)-C(2)	66.09(19)
O(2)-S(1)-C(1)-C(2)	-66.79(19)
N(1)-S(1)-C(1)-C(2)	179.09(17)
C(6)-C(1)-C(2)-C(3)	0.2(3)
S(1)-C(1)-C(2)-C(3)	179.14(16)
C(1)-C(2)-C(3)-C(4)	-0.9(3)
C(2)-C(3)-C(4)-C(5)	0.6(3)

C(3)-C(4)-C(5)-C(6)
C(2)-C(1)-C(6)-C(5)
S(1)-C(1)-C(6)-C(5)
C(2)-C(1)-C(6)-C(7)
S(1)-C(1)-C(6)-C(7)
C(4)-C(5)-C(6)-C(1)
C(4)-C(5)-C(6)-C(7)
S(1)-N(1)-C(7)-C(6)
S(1)-N(1)-C(7)-C(17)
S(1)-N(1)-C(7)-C(8)
C(1)-C(6)-C(7)-N(1)
C(5)-C(6)-C(7)-N(1)
C(1)-C(6)-C(7)-C(17)
C(5)-C(6)-C(7)-C(17)
C(1)-C(6)-C(7)-C(8)
C(5)-C(6)-C(7)-C(8)
N(1)-C(7)-C(8)-F(1)
C(6)-C(7)-C(8)-F(1)
C(17)-C(7)-C(8)-F(1)
N(1)-C(7)-C(8)-C(9)
C(6)-C(7)-C(8)-C(9)
C(17)-C(7)-C(8)-C(9)
N(1)-C(7)-C(8)-C(16)
C(6)-C(7)-C(8)-C(16)
C(17)-C(7)-C(8)-C(16)
F(1)-C(8)-C(9)-C(10)
C(7)-C(8)-C(9)-C(10)
C(16)-C(8)-C(9)-C(10)
C(8)-C(9)-C(10)-C(15)
C(8)-C(9)-C(10)-C(11)
C(15)-C(10)-C(11)-C(12)
C(9)-C(10)-C(11)-C(12)
C(10)-C(11)-C(12)-C(13)
C(11)-C(12)-C(13)-C(14)
C(12)-C(13)-C(14)-C(15)
C(11)-C(10)-C(15)-C(14)
C(9)-C(10)-C(15)-C(14)
C(11)-C(10)-C(15)-C(16)
C(9)-C(10)-C(15)-C(16)
C(13)-C(14)-C(15)-C(10)
C(13)-C(14)-C(15)-C(16)
C(10)-C(15)-C(16)-O(5)
C(14)-C(15)-C(16)-O(5)

0.3(3) 0.7(3) -178.36(14) -177.67(16) 3.28(18) -0.9(3)177.14(17) 1.79(17) 123.53(13) -118.32(13) -3.20(19)178.63(17) -118.52(17)63.3(2) 116.42(16) -61.8(2) 60.91(17) -55.92(17) 176.23(13) -176.84(14)66.32(19) -61.53(19) -53.34(19)-170.17(14) 61.97(18) -102.46(16) 137.09(15) 8.72(18) -6.1(2)174.37(18) 1.1(3)-179.5(2)-0.4(3) -0.4(4)0.4(3) -1.1(3)179.40(17) -179.88(17) 0.6(2) 0.3(3) 178.88(19) -178.55(19) 2.8(3)

C(10)-C(15)-C(16)-C(8)	5.2(2)
C(14)-C(15)-C(16)-C(8)	-173.50(18)
F(1)-C(8)-C(16)-O(5)	-70.2(2)
C(9)-C(8)-C(16)-O(5)	174.75(18)
C(7)-C(8)-C(16)-O(5)	45.3(2)
F(1)-C(8)-C(16)-C(15)	106.35(15)
C(9)-C(8)-C(16)-C(15)	-8.74(18)
C(7)-C(8)-C(16)-C(15)	-138.14(15)
C(18)-O(3)-C(17)-O(4)	-2.4(3)
C(18)-O(3)-C(17)-C(7)	172.85(15)
N(1)-C(7)-C(17)-O(4)	100.68(19)
C(6)-C(7)-C(17)-O(4)	-143.66(17)
C(8)-C(7)-C(17)-O(4)	-17.9(2)
N(1)-C(7)-C(17)-O(3)	-74.72(16)
C(6)-C(7)-C(17)-O(3)	40.9(2)
C(8)-C(7)-C(17)-O(3)	166.72(14)
C(17)-O(3)-C(18)-C(19)	-168.24(17)

Symmetry transformations used to generate equivalent atoms:Table 7.Hydrogen bonds for z [Å and deg.].

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Part IV. X-ray data of 7.7



Data intensity of  $C_{17}H_{14}F_2N_2O_4S$  (7) was collected using a Bruker SMART APEX II (Mo radiation) at 296 K in a nitrogen stream. Data collection and reduction were done by using the Bruker ApexII software package. The structures were solved by direct methods and refined by full-matrix least-squares on  $F^2$  with anisotropic displacement parameters for non-H atoms using SHELX-97. Hydrogen atoms were added at their geometrically idea positions and refined isotropically. Crystal data for 7:  $C_{17}H_{14}F_2N_2O_4S$ , T = 296(2) K, Orthorhombic, P2(1)2(1)2(1). a = 7.1956(3) Å, b = 9.6833(3) Å, c = 23.2873(8) Å, alpha = 90 deg, beta = 90 deg, gamma = 90 deg. V = 1622.59(10) Å<sup>3</sup>. Z = 4,  $d_{calc} = 1.557$  mg/m<sup>3</sup>. Total number of reflections 18989/[R(int) = 0.0297], R<sub>1</sub> = 0.0337, wR<sub>2</sub> = 0.0801 (all data), GOF = 1.052, and 235 parameter.

Table 1. Crystal data and structure refinement for z.

Identification code	Z
Empirical formula	$C_{17}H_{14}F_2N_2O_4S$
Formula weight	380.36
Temperature	296(2) K
Wavelength	0.71073 Å
Crystal system, space group	Orthorhombic, $P2(1)2(1)2(1)$
Unit cell dimensions	a = 7.1956(3)  Å alpha = 90 deg.
	b = 9.6833(3) Å beta = 90 deg.
	c = 23.2873(8)  Å gamma = 90 deg.
Volume	1622.59(10) Å <sup>3</sup>
Z, Calculated density	4, 1.557 Mg/m <sup>3</sup>
Absorption coefficient	$0.249 \text{ mm}^{-1}$

<sup>&</sup>lt;sup>7</sup> Supplementary crystallographic data have been deposited at the Cambridge Crystallographic Data Center. (CCDC 1439366).

F(000)	784
Crystal size	0.41 x 0.25 x 0.17 mm
Theta range for data collection	1.75 to 25.01 deg.
Limiting indices	-8<=h<=8, -10<=k<=11, -27<=l<=27
Reflections collected / unique	18989 / 2871 [R(int) = 0.0297]
Completeness to theta $= 25.01$	100.0 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.9590 and 0.9049
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	2871 / 0 / 235
Goodness-of-fit on F <sup>2</sup>	1.052
Final R indices [I>2sigma(I)]	$R_1 = 0.0314$ , $wR_2 = 0.0781$
R indices (all data)	$R_1 = 0.0337, wR_2 = 0.0801$
Absolute structure parameter	-0.03(8)
Largest diff. peak and hole	0.258 and -0.230 e. Å <sup>-3</sup>

Table 2. Atomic coordinates (  $x \ 10^4$ ) and equivalent isotropic displacement parameters (Å<sup>2</sup>  $x \ 10^3$ ) for z. U(eq) is defined as one third of the trace of the orthogonalized Uij tensor.

	Х	У	Z	U(eq)
S(1)	1952(1)	5451(1)	304(1)	34(1)
F(1)	3297(2)	5852(2)	-1405(1)	71(1)
F(2)	3229(3)	7570(2)	-819(1)	94(1)
O(1)	3828(2)	5964(2)	353(1)	48(1)
O(2)	989(3)	5282(2)	831(1)	54(1)
O(3)	719(3)	8924(2)	-1391(1)	50(1)
O(4)	-2518(2)	5587(2)	-856(1)	64(1)
N(1)	738(3)	6368(2)	-143(1)	42(1)
N(2)	-828(3)	6802(2)	-1517(1)	33(1)
C(1)	1914(3)	3997(2)	-140(1)	34(1)
C(2)	2386(3)	2658(3)	12(1)	48(1)
C(3)	2251(4)	1669(3)	-411(2)	62(1)
C(4)	1703(4)	2017(3)	-958(2)	62(1)
C(5)	1233(4)	3354(3)	-1100(1)	50(1)
C(6)	1331(3)	4360(2)	-682(1)	34(1)
C(7)	815(3)	5879(2)	-733(1)	31(1)
C(8)	-1079(3)	6062(2)	-1030(1)	37(1)
C(9)	930(3)	7543(2)	-1556(1)	32(1)

C(10)	2142(3)	6717(3)	-1122(1)	40(1)
C(11)	1790(3)	7551(2)	-2149(1)	33(1)
C(12)	3041(4)	8583(3)	-2291(1)	47(1)
C(13)	3953(4)	8542(3)	-2816(1)	64(1)
C(14)	3626(5)	7477(4)	-3194(1)	68(1)
C(15)	2393(4)	6461(4)	-3052(1)	64(1)
C(16)	1481(4)	6480(3)	-2534(1)	48(1)
C(17)	-2431(4)	7242(3)	-1859(1)	47(1)

Table 3. Bond lengths [Å] and angles [deg] for z.

S(1)-O(2)	1.4176(18)	
S(1)-O(1)	1.4431(18)	
S(1)-N(1)	1.6247(19)	
S(1)-C(1)	1.748(2)	
F(1)-C(10)	1.352(3)	
F(2)-C(10)	1.338(3)	
O(3)-C(9)	1.399(3)	
O(3)-H(3B)	0.8200	
O(4)-C(8)	1.203(3)	
N(1)-C(7)	1.453(3)	
N(1)-H(1A)	0.8600	
N(2)-C(8)	1.353(3)	
N(2)-C(9)	1.457(3)	
N(2)-C(17)	1.466(3)	
C(1)-C(6)	1.374(3)	
C(1)-C(2)	1.387(3)	
C(2)-C(3)	1.377(4)	
C(2)-H(2A)	0.9300	
C(3)-C(4)	1.375(4)	
C(3)-H(3A)	0.9300	
C(4)-C(5)	1.378(4)	
C(4)-H(4A)	0.9300	
C(5)-C(6)	1.379(3)	
C(5)-H(5A)	0.9300	
C(6)-C(7)	1.522(3)	
C(7)-C(8)	1.539(3)	
C(7)-C(10)	1.546(3)	
C(9)-C(11)	1.514(3)	
C(9)-C(10)	1.556(3)	
C(11)-C(12)	1.384(3)	
C(11)-C(16)	1.388(3)	

C(12)-C(13)	1.388(4)
C(12)-H(12A)	0.9300
C(13)-C(14)	1.377(5)
C(13)-H(13A)	0.9300
C(14)-C(15)	1.366(5)
C(14)-H(14A)	0.9300
C(15)-C(16)	1.374(4)
C(15)-H(15A)	0.9300
C(16)-H(16A)	0.9300
C(17)-H(17A)	0.9600
C(17)-H(17B)	0.9600
C(17)-H(17C)	0.9600
O(2)-S(1)-O(1)	115.41(11)
O(2)-S(1)-N(1)	110.81(11)
O(1)-S(1)-N(1)	111.41(11)
O(2)-S(1)-C(1)	114.34(11)
O(1)-S(1)-C(1)	109.79(10)
N(1)-S(1)-C(1)	92.93(10)
C(9)-O(3)-H(3B)	109.5
C(7)-N(1)-S(1)	114.06(15)
C(7)-N(1)-H(1A)	123.0
S(1)-N(1)-H(1A)	123.0
C(8)-N(2)-C(9)	115.40(18)
C(8)-N(2)-C(17)	120.30(19)
C(9)-N(2)-C(17)	120.42(17)
C(6)-C(1)-C(2)	123.2(2)
C(6)-C(1)-S(1)	110.05(16)
C(2)-C(1)-S(1)	126.71(19)
C(3)-C(2)-C(1)	116.7(3)
C(3)-C(2)-H(2A)	121.6
C(1)-C(2)-H(2A)	121.6
C(4)-C(3)-C(2)	120.8(2)
C(4)-C(3)-H(3A)	119.6
C(2)-C(3)-H(3A)	119.6
C(3)-C(4)-C(5)	121.5(2)
C(3)-C(4)-H(4A)	119.2
C(5)-C(4)-H(4A)	119.2
C(4)-C(5)-C(6)	118.8(3)
C(4)-C(5)-H(5A)	120.6
C(6)-C(5)-H(5A)	120.6
C(1)-C(6)-C(5)	118.9(2)
C(1)-C(6)-C(7)	113.14(18)

C(5)-C(6)-C(7)	128.0(2)
N(1)-C(7)-C(6)	104.53(17)
N(1)-C(7)-C(8)	110.69(18)
C(6)-C(7)-C(8)	111.29(18)
N(1)-C(7)-C(10)	113.94(18)
C(6)-C(7)-C(10)	113.72(18)
C(8)-C(7)-C(10)	102.87(17)
O(4)-C(8)-N(2)	126.8(2)
O(4)-C(8)-C(7)	124.56(19)
N(2)-C(8)-C(7)	108.65(19)
O(3)-C(9)-N(2)	111.05(18)
O(3)-C(9)-C(11)	106.82(17)
N(2)-C(9)-C(11)	114.49(17)
O(3)-C(9)-C(10)	111.98(19)
N(2)-C(9)-C(10)	101.10(16)
C(11)-C(9)-C(10)	111.49(18)
F(2)-C(10)-F(1)	106.3(2)
F(2)-C(10)-C(7)	112.11(18)
F(1)-C(10)-C(7)	109.93(19)
F(2)-C(10)-C(9)	110.6(2)
F(1)-C(10)-C(9)	110.21(17)
C(7)-C(10)-C(9)	107.72(18)
C(12)-C(11)-C(16)	119.3(2)
C(12)-C(11)-C(9)	119.1(2)
C(16)-C(11)-C(9)	121.3(2)
C(11)-C(12)-C(13)	119.8(3)
C(11)-C(12)-H(12A)	120.1
C(13)-C(12)-H(12A)	120.1
C(14)-C(13)-C(12)	120.3(3)
C(14)-C(13)-H(13A)	119.8
C(12)-C(13)-H(13A)	119.8
C(15)-C(14)-C(13)	119.7(3)
C(15)-C(14)-H(14A)	120.2
C(13)-C(14)-H(14A)	120.2
C(14)-C(15)-C(16)	120.9(3)
C(14)-C(15)-H(15A)	119.5
C(16)-C(15)-H(15A)	119.5
C(15)-C(16)-C(11)	120.0(3)
C(15)-C(16)-H(16A)	120.0
C(11)-C(16)-H(16A)	120.0
N(2)-C(17)-H(17A)	109.5
N(2)-C(17)-H(17B)	109.5
H(17A)-C(17)-H(17B)	109.5

N(2)-C(17)-H(17C)	109.5
H(17A)-C(17)-H(17C)	109.5
H(17B)-C(17)-H(17C)	109.5

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters (Å<sup>2</sup> x 10<sup>3</sup>) for z. The anisotropic displacement factor exponent takes the form: -2 pi<sup>2</sup> [  $h^2 a^{*2} U11 + ... + 2 h k a^* b^* U12$  ]

	U11	U22	U33	U23	U13	U12	
<b>S</b> (1)	43(1)	30(1)	30(1)	1(1)	-6(1)	2(1)	
F(1)	40(1)	98(1)	77(1)	45(1)	25(1)	29(1)	
F(2)	106(2)	112(2)	65(1)	36(1)	-48(1)	-79(1)	
O(1)	51(1)	42(1)	52(1)	-4(1)	-14(1)	-9(1)	
O(2)	69(1)	61(1)	32(1)	7(1)	-1(1)	5(1)	
O(3)	63(1)	32(1)	57(1)	-12(1)	25(1)	-6(1)	
O(4)	35(1)	89(2)	68(1)	33(1)	3(1)	-12(1)	
N(1)	64(1)	33(1)	28(1)	-4(1)	-4(1)	17(1)	
N(2)	32(1)	34(1)	34(1)	3(1)	-2(1)	-3(1)	
C(1)	25(1)	29(1)	47(1)	-1(1)	-2(1)	-2(1)	
C(2)	36(1)	33(1)	77(2)	4(1)	-10(1)	3(1)	
C(3)	38(1)	29(1)	118(3)	-10(2)	-8(2)	4(1)	
C(4)	52(2)	42(1)	91(2)	-31(2)	6(2)	-4(1)	
C(5)	47(2)	50(1)	53(1)	-18(1)	4(1)	-9(1)	
C(6)	28(1)	32(1)	42(1)	-5(1)	3(1)	-4(1)	
C(7)	34(1)	33(1)	26(1)	0(1)	0(1)	-2(1)	
C(8)	34(1)	41(1)	36(1)	4(1)	-1(1)	-3(1)	
C(9)	36(1)	28(1)	31(1)	-3(1)	3(1)	-3(1)	
C(10)	36(1)	48(1)	36(1)	4(1)	-5(1)	-14(1)	
C(11)	35(1)	34(1)	31(1)	4(1)	1(1)	4(1)	
C(12)	51(1)	43(1)	48(1)	5(1)	11(1)	-2(1)	
C(13)	57(2)	74(2)	62(2)	22(2)	21(2)	0(2)	
C(14)	64(2)	105(3)	35(1)	5(2)	14(1)	16(2)	
C(15)	68(2)	87(2)	36(1)	-17(1)	8(1)	6(2)	
C(16)	55(2)	49(1)	40(1)	-9(1)	2(1)	1(1)	
C(17)	38(1)	52(2)	49(1)	10(1)	-8(1)	1(1)	

	Х	У	Z	U(eq)
H(3B)	164	8961	-1084	76
H(1A)	106	7080	-41	50
H(2A)	2774	2438	382	58
H(3A)	2534	755	-326	74
H(4A)	1649	1335	-1238	74
H(5A)	855	3573	-1470	60
H(12A)	3270	9301	-2035	57
H(13A)	4788	9237	-2912	77
H(14A)	4243	7450	-3545	81
H(15A)	2167	5747	-3310	77
H(16A)	657	5775	-2440	58
H(17A)	-3465	6642	-1783	70
H(17B)	-2122	7198	-2260	70
H(17C)	-2757	8173	-1759	70

Table 5. Hydrogen coordinates (  $x \ 10^4$ ) and isotropic displacement parameters (Å<sup>2</sup>  $x \ 10^3$ ) for z.

Table 6. Torsion angles [deg] for z.

-139.92(17)
90.13(18)
-22.47(18)
128.45(17)
-99.99(17)
14.02(17)
-51.4(2)
80.1(2)
-165.9(2)
0.1(4)
180.00(19)
1.1(4)
-1.4(5)
0.5(4)
-1.0(3)
179.09(18)
177.4(2)
-2.4(2)
0.7(4)
-177.5(2)
23.3(2)

S(1)-N(1)-C(7)-C(8)	143.21(17)
S(1)-N(1)-C(7)-C(10)	-101.4(2)
C(1)-C(6)-C(7)-N(1)	-12.2(2)
C(5)-C(6)-C(7)-N(1)	166.1(2)
C(1)-C(6)-C(7)-C(8)	-131.67(19)
C(5)-C(6)-C(7)-C(8)	46.6(3)
C(1)-C(6)-C(7)-C(10)	112.7(2)
C(5)-C(6)-C(7)-C(10)	-69.0(3)
C(9)-N(2)-C(8)-O(4)	165.3(2)
C(17)-N(2)-C(8)-O(4)	7.4(4)
C(9)-N(2)-C(8)-C(7)	-16.4(3)
C(17)-N(2)-C(8)-C(7)	-174.4(2)
N(1)-C(7)-C(8)-O(4)	-56.5(3)
C(6)-C(7)-C(8)-O(4)	59.3(3)
C(10)-C(7)-C(8)-O(4)	-178.6(2)
N(1)-C(7)-C(8)-N(2)	125.20(19)
C(6)-C(7)-C(8)-N(2)	-119.0(2)
C(10)-C(7)-C(8)-N(2)	3.1(2)
C(8)-N(2)-C(9)-O(3)	-97.3(2)
C(17)-N(2)-C(9)-O(3)	60.5(3)
C(8)-N(2)-C(9)-C(11)	141.59(19)
C(17)-N(2)-C(9)-C(11)	-60.5(3)
C(8)-N(2)-C(9)-C(10)	21.6(2)
C(17)-N(2)-C(9)-C(10)	179.50(19)
N(1)-C(7)-C(10)-F(2)	11.7(3)
C(6)-C(7)-C(10)-F(2)	-108.0(2)
C(8)-C(7)-C(10)-F(2)	131.6(2)
N(1)-C(7)-C(10)-F(1)	129.66(19)
C(6)-C(7)-C(10)-F(1)	10.0(3)
C(8)-C(7)-C(10)-F(1)	-110.5(2)
N(1)-C(7)-C(10)-C(9)	-110.2(2)
C(6)-C(7)-C(10)-C(9)	130.11(19)
C(8)-C(7)-C(10)-C(9)	9.6(2)
O(3)-C(9)-C(10)-F(2)	-22.4(3)
N(2)-C(9)-C(10)-F(2)	-140.7(2)
C(11)-C(9)-C(10)-F(2)	97.2(2)
O(3)-C(9)-C(10)-F(1)	-139.62(19)
N(2)-C(9)-C(10)-F(1)	102.09(19)
C(11)-C(9)-C(10)-F(1)	-20.0(2)
O(3)-C(9)-C(10)-C(7)	100.4(2)
N(2)-C(9)-C(10)-C(7)	-17.8(2)
C(11)-C(9)-C(10)-C(7)	-139.93(18)
O(3)-C(9)-C(11)-C(12)	33.8(3)

N(2)-C(9)-C(11)-C(12)	157.2(2)
C(10)-C(9)-C(11)-C(12)	-88.8(2)
O(3)-C(9)-C(11)-C(16)	-152.1(2)
N(2)-C(9)-C(11)-C(16)	-28.7(3)
C(10)-C(9)-C(11)-C(16)	85.3(3)
C(16)-C(11)-C(12)-C(13)	0.7(4)
C(9)-C(11)-C(12)-C(13)	174.9(2)
C(11)-C(12)-C(13)-C(14)	-0.4(4)
C(12)-C(13)-C(14)-C(15)	0.3(5)
C(13)-C(14)-C(15)-C(16)	-0.5(5)
C(14)-C(15)-C(16)-C(11)	0.8(4)
C(12)-C(11)-C(16)-C(15)	-0.9(4)
C(9)-C(11)-C(16)-C(15)	-175.0(2)

Symmetry transformations used to generate equivalent atoms:

Table 7. Hydrogen bonds for z [Å and deg.].

D-H...A

d(D-H) d(H...A)

<(DHA)

d(D...A)

### Part IV. <sup>1</sup>H, <sup>19</sup>F, <sup>13</sup>C NMR spectra





yjs-yo-25 yjs-yo-25 



-82 -84 -86 -88 -90 -92 -94 -96 -98 -100 -102 -104 -106 -108 -110 -112 -114 -116 -118 -120 -122 f1 (ppm)






















yjs-yo-33









yjs−yo-37















yjs-yo-36





















































yjs-yo-52 yjs-yo-52

















10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)







10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 f1 (ppm)


















-80 -82 -84 -86 -88 -90 -92 -94 -96 -98 -102 -106 -110 -114 -118 -122 -126 -130 -134 -138 f1 (ppm)







-118 -122 -126 -130 -134 f1 (ppm) -86 -90 -110 -114 . . . . . . . . . . . . . . -94 -98 . . . . . . . -102 -106 -134 -138 -142 -146 -150

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LabSolutions Analysis Report

## <Sample Information>

Sample Name Sample ID Data Filename Method Filename	yjs-yn-30 yjs-yo-25-rac-ADH-60-40-1 chl-0128.lcm	1.0-30oC.lcd	
Vial #	: 1-1	Sample Type	: Unknown
Date Acquired Date Processed	: 2015/6/22 15:14:37 : 2015/6/22 15:48:08	Acquired by Processed by	: System Administrator : System Administrator

### <Chromatogram>

mV



## <Peak Table>

Detect	or A Chann	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	15.837	2517679	92795	50.092
2	27.356	2508408	52185	49.908
Total		5026087	144980	



# Analysis Report

## <Sample Information>

Sample Name Sample ID	yjs-yo-25-asy		
Data Filename	: vis-vo-25-asv-ADH-60-40-1.	0-30oC.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/6/22 14:12:51	Acquired by	: System Administrator
Date Processed	: 2015/6/22 14:43:03	Processed by	: System Administrator

### <Chromatogram>



#### Feak Table>

Detect	or A Chann	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	15.889	14189785	501101	96.596
2	27.706	500114	11684	3.404
Total		14689899	512785	



### D:\Data\yujinsheng\yjs-yo-25-rac-ADH-60-40-1.0-30oC.lcd

### D:\Data\yujinsheng\yjs-yo-25-asy-ADH-60-40-1.0-30oC.lcd

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# Analysis Report

### <Sample Information>

Sample Name	yjs-yo-43-3		
Data Filename	yjs-yo-43-3-rac-ASH-60-40-1.0-7	0.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename			
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/6/25 11:30:16	Acquired by	: System Administrator
Date Processed	: 2015/6/26 21:43:00	Processed by	: System Administrator

### <Chromatogram>



Detect	or A Chann	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	15.503	2709441	75392	50.230
2	24.164	2684642	42183	49.770
Total		5394083	117575	



# Analysis Report

## <Sample Information>

Sample Name Sample ID	yjs-yo-41		
Data Filename	. yjs-yo-41-asy-ASH-60-40-1.0	0-30 oC.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/6/26 21:57:09	Acquired by	: System Administrator
Date Processed	: 2015/6/26 22:27:20	Processed by	: System Administrator

### <Chromatogram>



Detect	or A Chann	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	15.804	15457782	406497	96.866
2	24.952	500079	8526	3.134
Total		15957861	415023	



### D:\Data\yujinsheng\yjs-yo-41-asy-ASH-60-40-1.0-30 oC.lcd

D:\Data\yujinsheng\yjs-yo-43-3-rac-ASH-60-40-1.0-70.lcd

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# Analysis Report

# <Sample Information>

Sample Name	: yjs-yn-40		
Sample ID Data Filename	: vis-vo-33-rac-ADH60-40-1	0-30oC.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename	÷		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL	Acquired by	: Sustem Administrator
Date Processed	: 2015/6/22 22:08:13	Processed by	: System Administrator
Date Acquired Date Processed	: 2015/6/22 21:39:23 : 2015/6/22 22:08:13	Acquired by Processed by	: System Administrator : System Administrator

#### <Chromatogram>



Detect	or A Channe	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	17.331	15434406	477606	49.971
2	24.449	15452284	323224	50.029
Total		30886690	800831	



# Analysis Report

# <Sample Information>

Sample Name	: yjs-yo-33		
Sample ID			
Data Filename	: yjs-yo-33-asy-ADH-60-40-1	.0-30oC.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename	:		
√ial #	: 1-1	Sample Type	: Unknown
njection Volume	: 10 uL		
Date Acquired	: 2015/6/22 20:40:58	Acquired by	: System Administrator
Date Processed	: 2015/6/22 22:08:44	Processed by	: System Administrator
Date Flouesseu	. 2010/0/22 22.00.44	FIDLESSED Dy	. System Aurimistrate

### <Chromatogram>



#### Peak Table>

Detect	or A Chann	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	17.218	30199480	869152	96.011
2	24.750	1254561	28816	3.989
Total		31454041	897968	



### D:\Data\yujinsheng\yjs-yo-33-rac-ADH--60-40-1.0-30oC.lcd

D:\Data\yujinsheng\yjs-yo-33-asy-ADH-60-40-1.0-30oC.lcd

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# Analysis Report

# <Sample Information>

Sample Name	yjs-yn-34		
Data Filename	: yjs-yo-37rac-ADH-60-40-1.0-30	0 oC.lcd	
Method Filename	: chl-0128.lcm		
Vial #	1-1	Sample Type	: Unknown
Injection Volume	: 10 uL	Accurate hu	· Ovetern Administrator
Date Processed	: 2015/6/25 18:12:30	Processed by	: System Administrator

### <Chromatogram>



Detecto	Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.		
1	19.804	8453927	237919	49.964		
2	35.240	8466108	127806	50.036		
Total		16920035	365725			



# Analysis Report

# <Sample Information>

Sample Name Sample ID	yjs-yo-37		
Data Filename	: yjs-yo-37-asy-ADH-60-40-	1.0-30 oC.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename		Original Trans	
Vial #	: 1-1	Sample Type	: Unknown
njection volume	: 10 UL	A considered have	· Ourteau Administration
Date Acquired	: 2015/6/25 16:11:57	Acquired by	: System Administrator
Date Processed	: 2015/6/25 18:13:33	Processed by	: System Administrator

## <Chromatogram>



Detecto	or A Channe	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	19.864	23289963	627917	96.860
2	35.989	755040	12787	3.140
Total		24045003	640704	



### D:\Data\yujinsheng\yjs-yo-37-asy-ADH-60-40-1.0-30 oC.lcd

D:\Data\yujinsheng\yjs-yo-37--rac-ADH-60-40-1.0-30 oC.lcd

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# Analysis Report

## <Sample Information>

Sample Name	: yjs-yo-43-2		
Sample ID	:		
Data Filename	: yjs-yo-43-2-rac-ADH-75-25-1.0.lcd		
Method Filename	: chl-0128.lcm		
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/6/23 21:25:08	Acquired by	: System Administrator
Date Processed	: 2015/6/26 21:42:37	Processed by	: System Administrator

### <Chromatogram>



Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.	
1	13.405	11208005	474205	50.026	
2	15.767	11196186	400511	49.974	
Total		22404191	874716		



# Analysis Report

## <Sample Information>

Sample Name	yjs-yo-42		
Data Filename	: yjs-yo-42-asy-ADH-75-25-1	.0-30 oC.lcd	
Method Filename	: chl-0128.lcm		
Satch Filename	1.1	Sample Ture	: Llokoowo
niection Volume	: 10 uL	Sample Type	. Onknown
Date Acquired	: 2015/6/26 22:31:59	Acquired by	: System Administrator
Date Processed	: 2015/6/27 9:15:39	Processed by	: System Administrator

### <Chromatogram>



Detect	or A Channe	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	13.312	1750365	80354	6.390
2	15.575	25642720	856595	93.610
Total		27393085	936949	



### D:\Data\yujinsheng\yjs-yo-42-asy-ADH-75-25-1.0-30 oC.lcd

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# Analysis Report

## <Sample Information>

Sample Name	: yjs-yo-43-1		
Sample ID Data Eilename	: vie.vo.36-rec.ADH-75-25-1.0-30	oC led	
Method Filename	: chl-0128.lcm	00.100	
Batch Filename	÷		
Vial #	: 1-1	Sample Type	: Unknown
Date Acquired Date Processed	: 2015/6/25 20:09:06 : 2015/6/25 20:40:31	Acquired by Processed by	: System Administrator : System Administrator

### <Chromatogram>



Detect	or A Channe	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	22.159	10615291	285524	49.932
2	27.258	10644142	219800	50.068
Total		21259433	505324	



# Analysis Report

## <Sample Information>

Sample Name Sample ID	yjs-yo-36		
Data Filename	: yjs-yo-36-asy-ADH-75-25-1	.0-30 oC.lcd	
Method Filename	: chl-0128.lcm		
Satch Filename		Comple Trace	- Halva even
/ial #	: 1-1	Sample Type	: Unknown
Date Acquired	· 2015/6/25 19:36:24	Acquired by	· System Administrator
Date Processed	: 2015/6/25 20:39:51	Processed by	: System Administrator
Batch Filename /ial # njection Volume Date Acquired Date Processed	: 1-1 : 10 uL : 2015/6/25 19:36:24 : 2015/6/25 20:39:51	Sample Type Acquired by Processed by	: Unknown : System Administra : System Administra

### <Chromatogram>



Detect	Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.		
1	22.250	797797	23337	4.193		
2	26.887	18230562	348156	95.807		
Total		19028359	371493			



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# Analysis Report

### <Sample Information>

Sample Name Sample ID Data Filename	: yjs-yn-35 : : yjs-yo-32rac-ASH60-40-0.8-3	0oC.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename	÷		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 UL	Acquired by	· Sustem Administrator
Date Acquired	2015/6/23 10:10:16	Processed by	System Administrator
Date Frocessed	. 2010/0/20 10.10.10	FIGGESSEG Dy	. Oystern Administrator

#### <Chromatogram>



#### a cak labie

Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.	
1	12.620	25316656	866379	50.164	
2	23.313	25151586	385830	49.836	
Total		50468242	1252209		



# Analysis Report

## <Sample Information>

Sample Name	: yjs-yo-32		
Sample ID	:		
Data Filename	: yjs-yo-32-asy-ASH60-40-0.8-	30oC.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/6/23 9:13:00	Acquired by	: System Administrator
Date Processed	: 2015/6/23 10:12:37	Processed by	: System Administrator

## <Chromatogram>



#### Peak Table>

Detect	Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.		
1	12.626	37455266	1271889	95.879		
2	23.453	1609983	27173	4.121		
Total		39065249	1299062			





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# Analysis Report

## <Sample Information>

Sample Name	: yjs-yo-46-2		
Sample ID Data Eilonamo	: : via va 46 2 roo ADH 60 40 1 0	60 lod	
Method Filename	: chl-0128.lcm	-00.100	
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Date Acquired	: 2015/6/25 11:06:31	Acquired by	: System Administrator
Date Processed	: 2015/6/25 11:20:36	Processed by	: System Administrator

## <Chromatogram>



Detect	Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.		
1	7.634	9667010	673767	50.049		
2	11.504	9648148	418082	49.951		
Total		19315158	1091849			



# Analysis Report

## <Sample Information>

Sample Name	yjs-yo-48		
Data Filename	: yjs-yo-48asy-ADH-60-40-1.0.lcd		
Method Filename	: chl-0128.lcm		
Batch Filename	:		
/ial #	: 1-1	Sample Type	: Unknown
njection Volume	: 10 uL		
Date Acquired	: 2015/6/30 20:10:35	Acquired by	: System Administrator
Date Processed	: 2015/6/30 20:25:50	Processed by	: System Administrator

### <Chromatogram>



Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.	
1	7.521	901964	70145	7.934	
2	11.350	10467027	493775	92.066	
Total		11368991	563921		





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# Analysis Report

# <Sample Information>

Sample Name	yjs-yo-46-1		
Data Filename	; yjs-yo-46-1-rac-ADH-60-40-1.0-60.l	lcd	
Method Filename	: chl-0128.lcm		
Vial #	1-1	Sample Type	: Unknown
Injection Volume	: 10 uL	A	Our terre Andre Selected and
Date Acquired Date Processed	: 2015/6/30 17:26:55	Processed by	: System Administrator : System Administrator

### <Chromatogram>



Detect	Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.		
1	13.924	9771498	393230	49.985		
2	17.332	9777276	312655	50.015		
Total		19548774	705885			



# Analysis Report

## <Sample Information>

Sample Name Sample ID Data Filename Method Filename	: yjs-yo-47 : : yjs-yo-47-asy-ADH-60-40-1.0.lcd : chl-0128.lcm		
atch Filename	:		
/ial #	: 1-1	Sample Type	: Unknown
njection Volume	: 10 uL		
ate Acquired	: 2015/6/30 16:58:48	Acquired by	: System Administrator
ate Processed	: 2015/7/9 21:36:38	Processed by	: System Administrator

## <Chromatogram>



Detect	Detector A Channel 2 230nm					
Peak#	Ret. Time	Area	Height	Conc.		
1	13.557	1298862	57048	5.217		
2	16.901	23599005	776097	94.783		
Total		24897867	833145			



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# Analysis Report

### <Sample Information>

Sample Name Sample ID Data Filename Method Filename	yjs-yn-57 yjs-yo-61-rac-ADH-60-40-1.0-30oC. chl-0128.lcm	lcd	
Vial #	1-1	Sample Type	: Unknown
Date Acquired Date Processed	: 2015/7/9 21:12:20 : 2015/7/9 21:55:29	Acquired by Processed by	: System Administrator : System Administrator

### <Chromatogram>



etector A Channel 2 230nm						
Peak#	Ret. Time	Area	Height	Conc.		
1	20.882	21423891	594982	49.824		
2	38.864	21575096	294442	50.176		
Total		42998987	889424			



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# Analysis Report

## <Sample Information>

Sample Name Sample ID Data Filename Method Filename	: yjs-yo-61-asy : : yjs-yo-61-asy-ADH-60-40- : chl-0128.lcm	1.0-30oC.lcd	
Batch Filename Vial #	1-1 10 ul	Sample Type	: Unknown
Date Acquired Date Processed	: 2015/7/9 20:23:51 : 2015/7/9 21:37:26	Acquired by Processed by	: System Administrator : System Administrator

## <Chromatogram>



### <Peak Table>

Detecto	or A Channe	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	20.875	61811330	1653485	95.966
2	39.292	2598302	39762	4.034
Total		64409633	1693247	



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# Analysis Report

# <Sample Information>

Sample Name	: yjs-yn-85		
Sample ID	: 		
Method Filename	: chl-0128 lcm		
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/7/4 18:05:26	Acquired by	: System Administrator
Date Processed	. 2010/1/4 10:33:10	Processed by	. System Administrator

## <Chromatogram>



Detect	or A Channe	el 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	15.212	5859055	226137	49.623
2	24.537	5948123	136741	50.377
Total		11807178	362878	



# Analysis Report

# <Sample Information>

sach Filename : fall # :1-1 Sample Type : Unknown njection Volume : 10 uL. Jate Acquired : 2015/7/4 16:57:53 Acquired by : System Administrator Jate Acqueed : 2015/7/4 18:05:44 Processed by : System Administrator	Sample Name Sample ID Data Filename Method Filename	: yjs-yo-52 : : yjs-yo-52-asy-60-40-1.0-30oC.lcd : chl-0128.lcm		
//ail # :1-1 Sample Type <td: njection="" td="" unknown="" volume<="">   .10 Li 2015/7/4 16:57:53 Acquired by : System Administrator   .10 Li 2015/7/4 16:57:53 Acquired by : System Administrator</td:>	Satch Filename			
njection Volume : 10 uL Jate Acquired : 2015/7/4 16:57:53 Acquired by : System Administrator Jate Processed : 2015/7/4 18:06:44 Processed by : System Administrator	/ial #	: 1-1	Sample Type	: Unknown
Date Acquired : 2015/7/4 16:57:53 Acquired by : System Administrator	njection Volume	: 10 uL		
Date Processed 2015/7/4 18:06:44 Processed by System Administrator	Date Acquired	: 2015/7/4 16:57:53	Acquired by	: System Administrator
	Date Processed	2015/7/4 18:06:44	Processed by	· System Administrator

## <Chromatogram>



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Detect	or A Channe	l 2 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	15.211	7773930	297027	96.523
2	24.700	280048	7143	3.477
Total		8053979	304170	



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# Analysis Report

# <Sample Information>

Sample Name	: yjs-yo-83-1		
Sample ID	: 	Cled	
Method Filename	2 lom	C.ICu	
Batch Filename	2.1011		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/7/30 16:06:37	Acquired by	: System Administrator
Date Processed	: 2015/8/25 10:00:02	Processed by	: System Administrator

## <Chromatogram>



Detecto	r A Channel	1 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	9.249	3371490	226353	49.851
2	18.057	3391588	110938	50.149
Total		6763079	337290	



# Analysis Report

# <Sample Information>

Sample Name	yjs-yo-85		
Data Filename	: yjs-yo-85-asy-ADH-60-40-1	.0-30oC.lcd	
Method Filename	: 2.lcm		
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/7/30 15:02:32	Acquired by	: System Administrator
Date Processed	: 2015/7/30 15:25:17	Processed by	: System Administrator

### <Chromatogram>



Detector A Channel 1 230nm					
Peak#	Ret. Time	Area	Height	Conc.	
1	9.277	3879831	258240	94.844	
2	18.152	210935	7296	5.156	
Total		4090766	265536		



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2015/8/25 10:58:12 Page 1 / 1

# Analysis Report

# <Sample Information>

Sample Name	: yjs-yo-83-3		
Sample ID Data Filename	: vis-vo-90-RAC-ADH-60-40-1 0-3	NoC Icd	
Method Filename	: 2.lcm	000.00	
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Date Acquired	: 2015/8/6 10:48:00	Acquired by	: System Administrator
Date Processed	: 2015/8/25 9:59:10	Processed by	: System Administrator

### <Chromatogram>



Detect	Detector A Channel 1 230nm					
Peak#	Ret. Time	Area	Height	Conc.		
1	10.830	1502076	73446	50.709		
2	19.686	1460067	37446	49.291		
Total		2962143	110892			



# Analysis Report

# <Sample Information>

Sample Name Sample ID	: yjs-yo-90		
Data Filename	: yjs-yo-90-asy-ADH-60-40-	1.0-30oC.lcd	
Method Filename	: 2.lcm		
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 10 uL		
Date Acquired	: 2015/8/6 10:22:41	Acquired by	: System Administrator
Date Processed	: 2015/8/25 9:59:13	Processed by	: System Administrator

## <Chromatogram>



Detect	or A Channe	el 1 230nm		
Peak#	Ret. Time	Area	Height	Conc.
1	10.810	22298635	1216664	96.209
2	19.698	878665	25067	3.791
Total		23177300	1241731	



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# Analysis Report

# <Sample Information>

Sample Name	: yjs-yo-77 rac		
Sample ID Data Eilonamo	: : via va 72 2 roa ADH 60 40 1	0.20 oC lod	
Method Filename	: 2.lcm	0-50 00.100	
Batch Filename			
Vial #	: 1-1	Sample Type	: Unknown
Date Acquired Date Processed	: 2015/7/23 15:23:01 : 2015/8/6 10:25:58	Acquired by Processed by	: System Administrator : System Administrator

### <Chromatogram>



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Detector A Channel 2 205nm					
Peak#	Ret. Time	Area	Height	Conc.	
1	9.391	7230600	424102	49.913	
2	18.981	7255720	215574	50.087	
Total		14486319	639676		



# Analysis Report

# <Sample Information>

Sample Name Sample ID	yjs-yo-89		
Data Filename	: yjs-yo-89-asy-ADH-60-40-	1.0-30oC.lcd	
Method Filename	: 2.lcm		
Batch Filename	÷		
/ial #	: 1-1	Sample Type	: Unknown
njection Volume	: 10 uL		
Date Acquired	: 2015/8/6 9:59:31	Acquired by	: System Administrator
Date Processed	: 2015/8/25 9:59:36	Processed by	: System Administrator

## <Chromatogram>



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Detect	Detector A Channel 2 205nm					
Peak#	Ret. Time	Area	Height	Conc.		
1	9.397	55903166	2503747	94.935		
2	19.178	2982657	100253	5.065		
Total		58885823	2604000			



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# Analysis Report

# <Sample Information>

Sample Name	: yjs-yo-121-rac		
Data Filename	: vis-vo-125-rac-ADH-60-40-1.0-	-30oC.lcd	
Method Filename	: chl-0128.lcm		
Vial #	1-1	Sample Type	: Unknown
Injection Volume	: 10 uL	eample type	
Date Acquired Date Processed	: 2015/9/23 11:25:04 : 2015/9/23 11:51:35	Acquired by Processed by	: System Administrator : System Administrator

### <Chromatogram>



Detector	Detector A Channel 1 205nm					
Peak# F	Ret. Time	Area	Height	Conc.		
1	13.567	13431145	580226	49.409		
2	20.728	13752328	378549	50.591		
Total		27183472	958775			



# Analysis Report

# <Sample Information>

Sample Name : yjs-yo-125-asy		
Data Filename : yjs-yo-125-asy-ADH-60-40-1.0-	30oC.lcd	
Nethod Filename : chl-0128.icm		
Batch Filename :		
/ial # : 1-1	Sample Type	: Unknown
niection Volume : 10 uL		
Date Acquired : 2015/9/22 21:22:09	Acquired by	: System Administrator
Date Processed : 2015/9/22 21:52:34	Processed by	: System Administrator

## <Chromatogram>



Detect	or A Channe	el 1 205nm		
Peak#	Ret. Time	Area	Height	Conc.
1	13.596	43672025	1794141	95.297
2	20.938	2155129	63613	4.703
Total		45827154	1857754	



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# Analysis Report

# <Sample Information>

Sample Name	yjs-yo-109-rac-60-40-0.8			
Data Filename	yjs-yo-93racADH-60-40-0.8-230.lcd			
Batch Filename	. chi-0128.ichi			
Vial #	1-1	Sample Type	: Unknown	
Date Acquired Date Processed	: 10 uL : 2015/9/10 15:51:13 : 2015/9/10 16:06:33	Acquired by Processed by	: System Administrator : System Administrator	

## <Chromatogram>



Detector A Channel 1 205nm					
Peak#	Ret. Time	Area	Height	Conc.	
1	8.299	16951251	905067	49.700	
2	10.902	17155762	717750	50.300	
Total		34107013	1622817		



# Analysis Report

# <Sample Information>

Sample Name	: yjs-yo-109-asy-60-40-0.8			
Sample ID	:			
Data Filename	: yjs-yo-109-asyADH-60-40-0.8-230.lcd			
Method Filename	: chl-0128.lcm			
Vial #	: 1-1	Sample Type	: Unknown	
Date Acquired	: 2015/9/10 16:09:27	Acquired by	: System Administrator	
Date Processed	: 2015/9/10 16:25:17	Processed by	: System Administrator	

#### <Chromatogram>



#### Peak Table>

Detector A Channel 1 205nm						
Peak#	Ret. Time	Area	Height	Conc.		
1	8.275	31826783	1573605	96.144		
2	10.973	1276561	60770	3.856		
Total		33103344	1634375			



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# Analysis Report

# <Sample Information>

Sample Name	: yjs-yo-36-rac		
Data Filename	yjs-yo-36-rac-IC-60-40-1.0-50	) Pa-30 oC1.lcd	
Method Filename	: chl-0128.lcm		
Batch Filename	:		
Vial #	: 1-1	Sample Type	: Unknown
Injection Volume	: 20 uL		
Date Acquired	: 2015/11/27 9:28:27	Acquired by	: System Administrator
Date Processed	: 2015/11/27 10:10:53	Processed by	: System Administrator

## <Chromatogram>



# <Peak Table>

Detector A Channel 2 230nm						
Peak#	Ret. Time	Area	Height	Conc.		
1	20.474	6851387	152437	49.948		
2	30.265	6865775	101629	50.052		
Total		13717162	254066			

# Analysis Report

# <Sample Information>

Sample Name Sample ID Data Filename Method Filename	: yjs-yo-36-asy : : yjs-yo-36-asy-IC-60-40-1.0- : chl-0128.lcm	s-yo-36-asy s-yo-36-asy-IC-60-40-1.0-50 Pa-30 oC2.lcd nI-0128.lcm			
Batch Filename /ial # niection Volume	: : 1-1 : 20 ul	Sample Type	: Unknown		
Date Acquired Date Processed	: 2015/11/27 10:26:32 : 2015/11/27 11:02:02	Acquired by Processed by	: System Administrator : System Administrator		

### <Chromatogram>



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De	Detector A Channel 2 230nm					
Pe	eak#	Ret. Time	Area	Height	Conc.	
	1	20.532	5777663	128586	65.862	
	2	30.409	2994758	44394	34.138	
	Total		8772421	172980		





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