

Supporting Information

Table of Contents

1. General information	S2-3
2. General procedure for the asymmetric addition of 1 to 2 catalyzed by H	S4
3. Characterization of Adducts	S5-S18
4. Synthesis and characterization of 4 and 5 .	S19-S20
5. Copies of NMR Spectra	S21-S40

1. General information

General procedures and methods

Experiments involving moisture and/or air sensitive components were performed under a positive pressure of nitrogen in oven-dried glassware equipped with a rubber septum inlet. Dried solvents and liquid reagents were transferred by oven-dried syringes or hypodermic syringe cooled to ambient temperature in a desiccator. Reaction mixtures were stirred in 10 mL sample vial with Teflon-coated magnetic stirring bars unless otherwise stated. Moisture in non-volatile reagents/compounds was removed in high *vacuo* by means of an oil pump and subsequent purging with nitrogen. Solvents were removed in *vacuo* under ~30 mmHg and heated with a water bath at 30–35 °C using Changcheng rotary evaporator with Changcheng aspirator. The condenser was cooled with running water at 0 °C.

All experiments were monitored by analytical thin layer chromatography (TLC). TLC was performed on pre-coated plates. After elution, plate was visualized under UV illumination at 254 nm for UV active material. Further visualization was achieved by staining KMnO₄, ceric molybdate, or anisaldehyde solution. For those using the aqueous stains, the TLC plates were heated on a hot plate.

Columns for flash chromatography (FC) contained silica gel 200–300 mesh. Columns were packed as slurry of silica gel in petroleum ether and equilibrated solution using the appropriate solvent system. The elution was assisted by applying pressure of about 2 atm with an air pump.

Instrumentations

Proton nuclear magnetic resonance (¹H NMR) and carbon NMR (¹³C NMR) were recorded in CDCl₃ otherwise stated. ¹H (300 MHz) and ¹³C (75 MHz) were performed on Bruker AVANCE-III (300 MHz) spectrometers. Chemical shifts are reported in parts per million (ppm), using the residual solvent signal as an internal standard: CDCl₃ (¹H NMR: δ 7.26, singlet; ¹³C NMR: δ 77.0, triplet). Multiplicities were given as: *s* (singlet), *d* (doublet), *t* (triplet), *q* (quartet), *quintet*, *m* (multiplets), *dd* (doublet of doublets), *dt* (doublet of triplets), and *br* (broad). Coupling constants (*J*) were recorded in Hertz (Hz). The number of proton atoms (*n*) for a given resonance was indicated by *n*H. The number of carbon atoms (*n*) for a given resonance was indicated by *n*C. HRMS was reported in units of mass of charge ratio

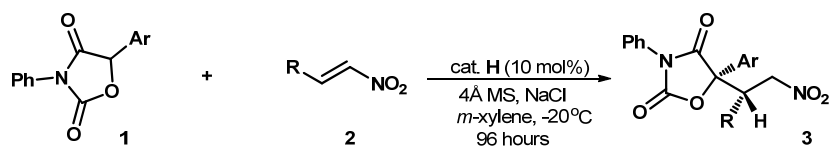
(m/z). Optical rotations were recorded on a polarimeter with a sodium lamp of wavelength 589 nm and reported as follows; $[\alpha]_{\lambda}^{T^{\circ}C}$ (c = g/100 mL, solvent). Melting points were determined on a melting point apparatus.

Enantiomeric excesses were determined by chiral High Performance Liquid Chromatography (HPLC) analysis. UV detection was monitored at 254 nm. HPLC samples were dissolved in HPLC grade isopropanol (IPA) unless otherwise stated.

Materials

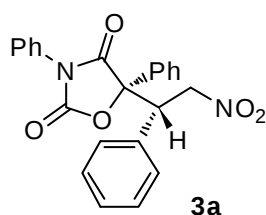
All commercial reagents were purchased with the highest purity grade. They were used without further purification unless specified. All solvents used, mainly petroleum ether (PE) and ethyl acetate (EtOAc) were distilled. Anhydrous DCM and MeCN were freshly distilled from CaH₂ and stored under N₂ atmosphere. THF, Et₂O, *m*-xylene and toluene were freshly distilled from sodium/benzophenone before use. Anhydrous methanol and ethanol were distilled from Mg. All compounds synthesized were stored in a -20 °C freezer and light-sensitive compounds were protected with aluminium foil.

2. General procedure for the asymmetric addition of **1** to **2** catalyzed by **H**

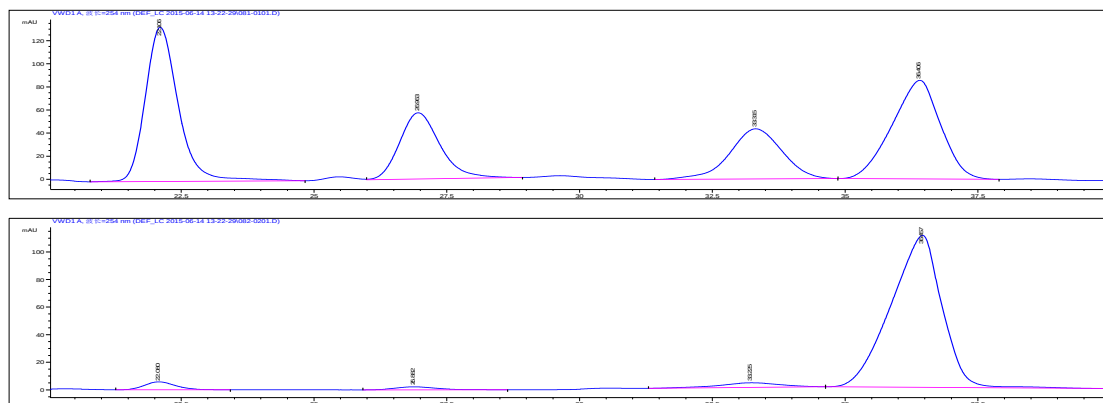


Diphenyloxazolidine-2,4-dione **1** (0.1 mmol, 1.0 equiv), nitroolefin **2** (0.12 mmol, 1.2 equiv), **H** (0.01 mmol, 0.1 equiv), NaCl (0.1 equiv) and 4Å molecular sieves (50 mg) were dissolved in *m*-xylene (1.0 mL). The reaction mixture was stirred at -20 °C for 72–96 hours and monitored by TLC. Upon complete consumption of **1**, the reaction mixture was concentrated under reduced pressure. The recovered crude material was subsequently purified by flash column chromatography on *silica gel* with PE/EtOAc mixture (20:1–5:1 ratio, the crude material was completely dissolved in CH₂Cl₂/PE before loaded on *silica gel*). After removing the solvent in *vacuo*, adducts **3** were obtained.

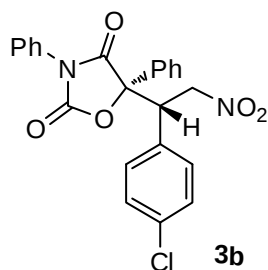
3. Characterization of adducts



White solid; Mp 152.3–153.1 °C; 94% ee; dr = 19:1; 98% yield; $[\alpha]_{\text{D}}^{22}$ +22.8 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.78 (d, *J* = 6.8, 2H), 7.52–7.33 (m, 8H), 7.27–7.12 (m, 3H), 6.59–6.44 (m, 2H), 4.91 (dd, *J* = 13.4, 11.4, 1H), 4.49 (dd, *J* = 11.4, 4.0, 1H), 4.35 (dd, *J* = 13.5, 4.0, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 170.3, 152.5, 132.3, 131.6, 130.1, 129.8, 129.6 (two peaks), 129.4 (three peaks), 129.1, 125.7, 124.9, 87.9, 74.1, 51.3; HRMS (ESI) *m/z* 403.1297 (*M*+H⁺), Calcd for C₂₃H₁₉N₂O₅ 403.1294; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; *t_R* = 22.1 min (minor peak), 26.9 min, 33.2 min, 36.4 min (major peak).

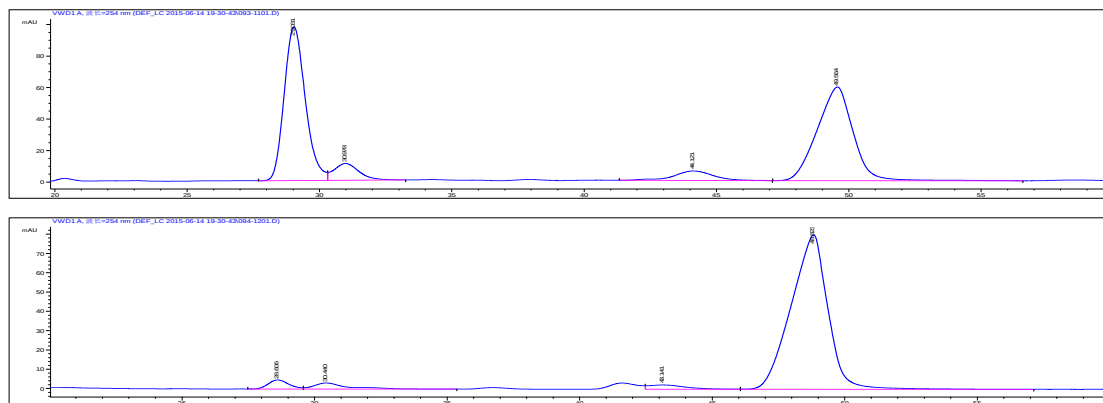


Entry	Retention Time	Area	Height	%Area
1	22.08	231	5.7	2.893
2	26.882	111.9	2.1	1.401
3	33.225	270.4	3.3	3.387
4	36.457	7371.9	110.3	92.319

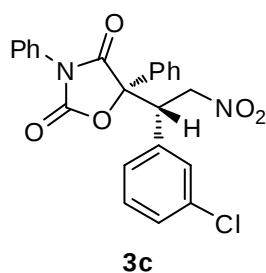


White solid; Mp 123.2–124.1 °C; 94% ee; dr = 19:1; 92% yield; $[\alpha]_{\text{D}}^{22}$ +20.5 (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.85 (dd, *J* = 8.0, 1.5, 2H), 7.61–7.50 (m, 3H), 7.50–7.39 (m, 4H), 7.36–7.30 (m, 3H), 6.77–6.45 (m, 2H), 4.94 (dd, *J* = 13.5, 11.5, 1H), 4.56 (dd, *J* = 11.4, 4.0, 1H), 4.42 (dd, *J* = 13.6, 4.0, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 170.2, 152.4, 135.9, 132.1, 130.7, 130.3, 130.2, 129.7 (two peaks), 129.6, 129.4, 125.5, 124.8, 87.7, 74.0, 50.8; HRMS (ESI) *m/z* 437.0902 (*M*+H⁺), Calcd for C₂₃H₁₈ClN₂O₅ 437.0904; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250

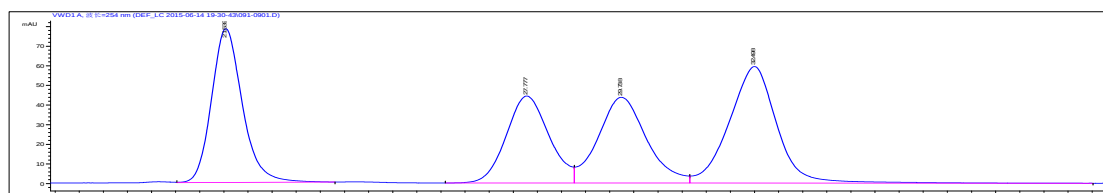
mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 28.6 min (minor peak), 30.4 min, 43.1 min, 48.8 min (major peak).

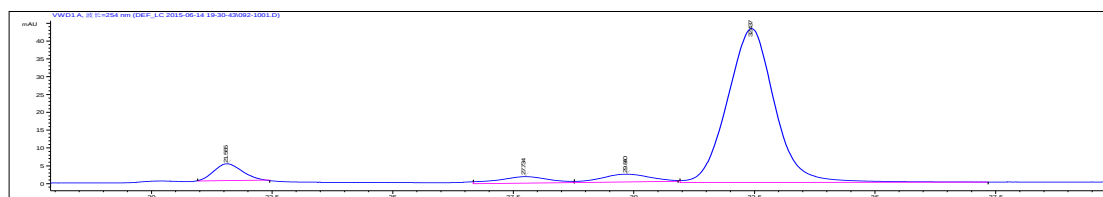


Entry	Retention Time	Area	Height	%Area
1	28.605	247.4	4.6	2.946
2	30.44	266.7	3.1	3.177
3	43.141	206.7	2.2	2.462
4	48.823	7676.2	80	91.415

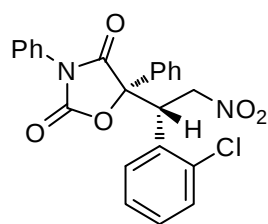


White solid; Mp 135.8–136.5 °C; 95% ee; dr = 10:1; 96% yield; $[\alpha]_D^{22} +21.3$ (c 1.00, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.85 (dd, J = 7.9, 1.4, 2H), 7.65–7.31 (m, 10H), 6.73 (dd, J = 6.4, 3.2, 2H), 4.94 (dd, J = 13.6, 11.4, 1H), 4.65–4.31 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 170.1, 152.3, 135.4, 133.8, 132.0, 129.9, 129.8, 129.7, 129.5 (two peaks), 129.4, 127.1, 125.5, 124.9, 87.6, 73.9, 50.9; HRMS (ESI) m/z 437.0901 ($\text{M}+\text{H}^+$), Calcd for $\text{C}_{23}\text{H}_{18}\text{ClN}_2\text{O}_5$ 437.0904; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 21.3 min (minor peak), 27.4 min, 29.2 min, 32.4 min (major peak).





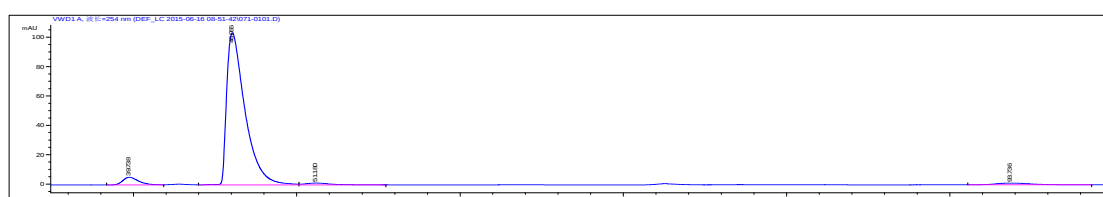
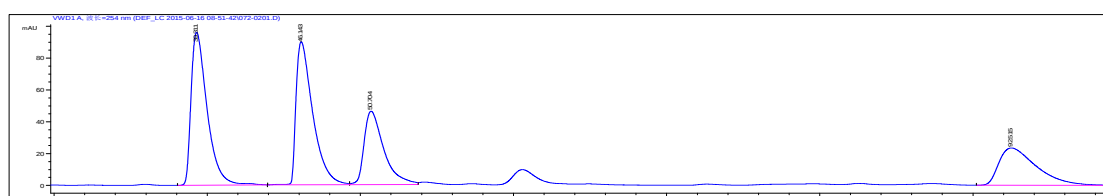
Entry	Retention Time	Area	Height	%Area
1	21.323	214	5.8	2.621
2	27.414	90.4	1.9	1.107
3	29.199	167	3.1	2.045
4	32.402	7694.8	133.8	94.227

**3d**

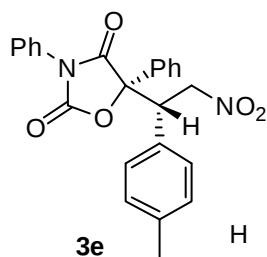
White solid; Mp 106.7–107.2 °C; 92% ee; dr = >19:1; 92% yield;

$[\alpha]_D^{22} +26.3$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.81 (d, *J* = 7.0, 2H), 7.62–7.18 (m, 10H), 6.62 (dd, *J* = 6.5, 2.6, 2H), 5.35 (dd, *J* = 11.3, 3.9, 1H), 4.85 (dd, *J* = 13.6, 11.5, 1H), 4.45 (dd, *J* = 13.8, 3.9, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 169.4, 152.6, 136.4, 132.6, 131.2, 130.6, 130.3,

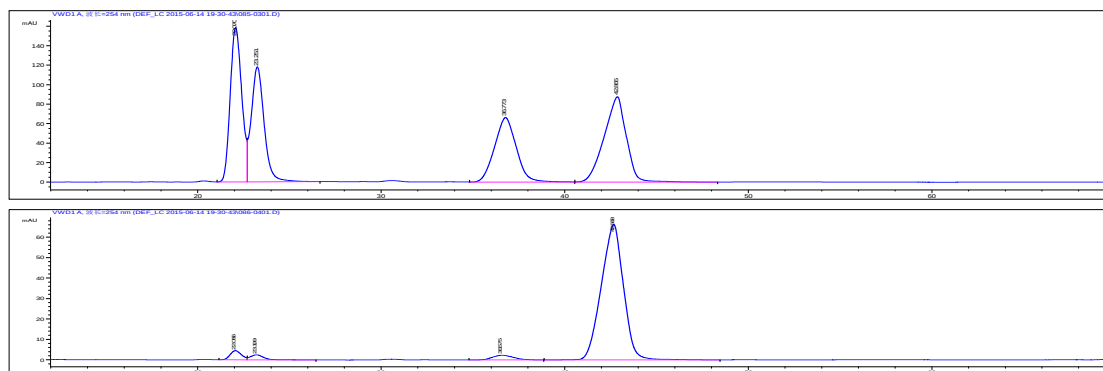
130.0 (two peaks), 129.3 (two peaks), 128.0 (two peaks), 125.6, 124.9, 87.8, 74.5, 46.1; HRMS (ESI) *m/z* 437.0897 (M+H⁺), Calcd for C₂₃H₁₈ClN₂O₅ 437.0904; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/05; flow rate 1.0 mL/min; 25 °C; 254 nm; *t*_R = 39.7 min (minor peak), 46.1 min (major peak), 51.2 min, 93.7 min.



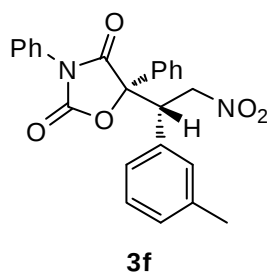
Entry	Retention Time	Area	Height	%Area
1	39.738	347.3	5.2	3.977
2	46.053	8076	103.5	92.481
3	51.174	131.1	1.2	1.502
4	93.74	178.3	1.2	2.041



White solid; Mp 146.5–147.7 °C; 94% ee; dr = 18:1; 88% yield; $[\alpha]_D^{22} +28.4$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.74 (dd, *J* = 17.9, 11.2, 2H), 7.54–7.35 (m, 3H), 7.32–7.08 (m, 7H), 6.67–6.39 (m, 2H), 4.86 (dd, *J* = 13.3, 11.4, 1H), 4.53–4.21 (m, 2H), 2.24 (d, *J* = 6.1, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.4, 152.6, 139.6, 132.4, 130.1, 130.0 (two peaks), 129.6, 129.3, 129.2 (two peaks), 128.5, 125.7, 124.9, 88.1, 74.2, 51.1, 21.1; HRMS (ESI) *m/z* 417.1446 (M+H⁺), Calcd for C₂₄H₂₁N₂O₅ 417.1450; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; *t_R* = 22.1 min (minor peak), 23.2min, 36.5 min, 42.7 min (major peak).

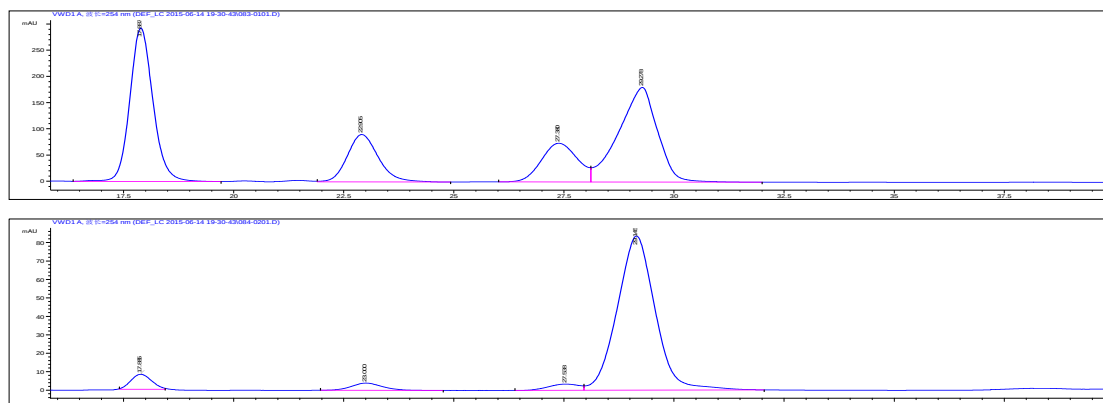


Entry	Retention Time	Area	Height	%Area
1	22.055	195	4.5	3.212
2	23.199	128	2.5	2.109
3	36.575	182.4	2.3	3.005
4	42.668	5565.6	66.2	91.674

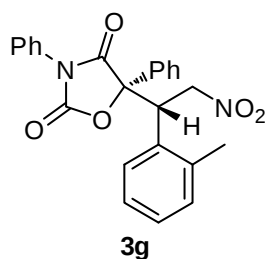


White solid; Mp 139.8–140.2 °C; 90% ee; dr = 19:1; 96% yield; $[\alpha]_D^{22} +18.4$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.98–7.68 (m, 1H), 7.65–7.39 (m, 1H), 7.37–7.13 (m, 2H), 6.69–6.46 (m, 1H), 4.98 (dd, *J* = 13.3, 11.3, 0H), 4.65–4.33 (m, 1H), 2.35 (d, *J* = 10.2, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 170.3, 152.6, 139.2, 132.4, 131.6, 130.3, 130.1, 130.0, 129.6, 129.5, 129.4 (two peaks), 126.3, 125.7, 125.6, 125.0, 87.8, 74.1, 51.6, 21.4; HRMS (ESI) *m/z* 439.1266 (M+Na⁺), Calcd for C₂₄H₂₀N₂O₅Na 439.1270; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 3 nm; *t_R* = 17.9 min (minor

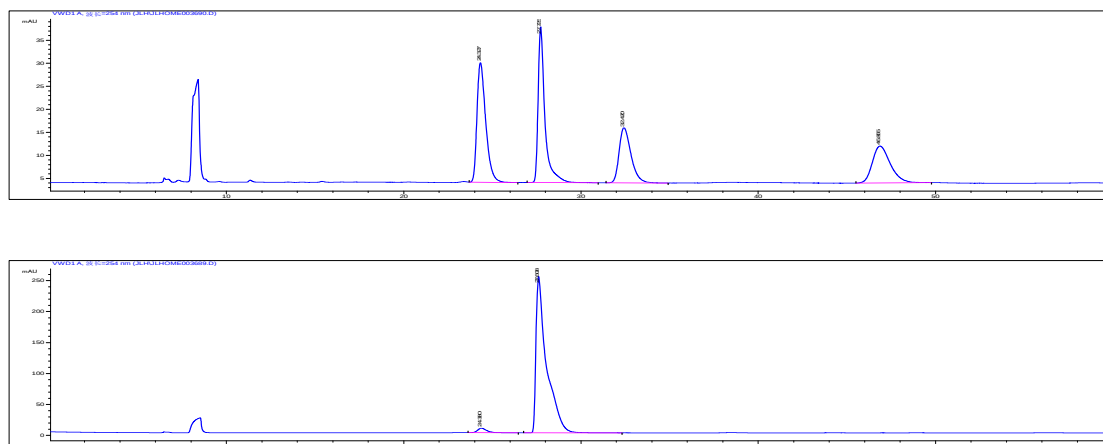
peak), 23.0 min, 27.5 min, 29.1 min (minor peak).



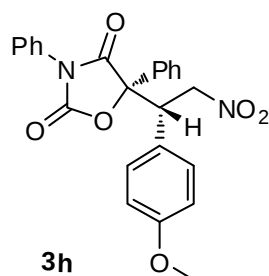
Entry	Retention Time	Area	Height	%Area
1	17.885	268.8	8.2	4.753
2	23.001	197	3.9	3.484
3	27.534	168.4	3.5	2.978
4	29.145	5020.1	83.7	88.785



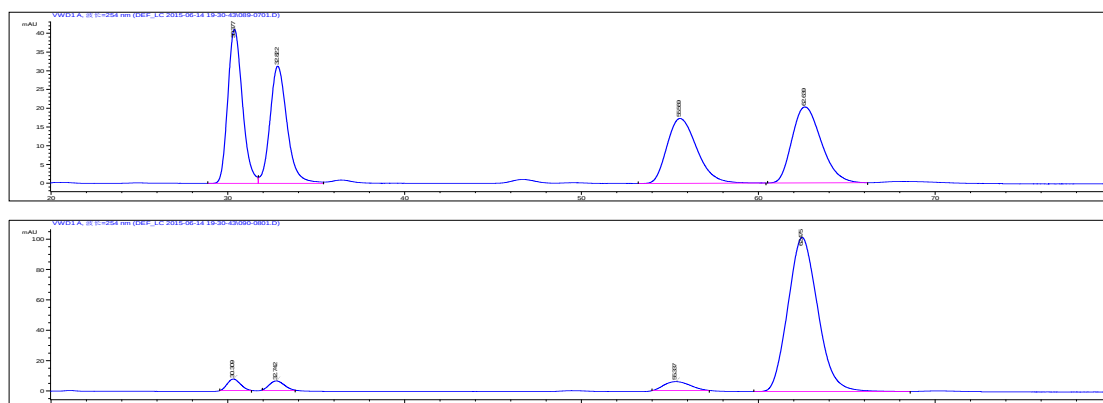
White solid; Mp 205.9–206.1 °C; 96% ee; dr = >19:1; 95% yield; $[\alpha]_D^{22} +25.6$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.96–7.84 (m, 2H), 7.65–7.48 (m, 5H), 7.38–7.26 (m, 5H), 6.66–6.55 (m, 2H), 5.04–4.88 (m, 2H), 4.48 (q, *J* = 10.3, 1H), 2.59 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.4, 152.7, 138.9, 132.9, 131.9, 131.0, 130.1, 129.9, 129.6, 129.4, 129.3, 127.0, 126.4, 125.9, 125.6, 124.9, 88.3, 75.0, 46.3, 20.0; HRMS (ESI) *m/z* 417.1447 (*M*+H⁺), Calcd for C₂₄H₂₁N₂O₅ 417.1450; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/05; flow rate 1.0 mL/min; 25 °C; 254 nm; *t_R* = 24.3 min (minor peak), 27.6 min (major peak).



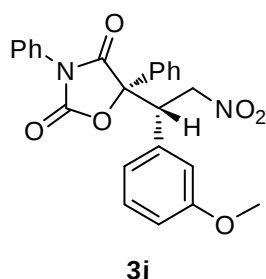
Entry	Retention Time	Area	Height	%Area
1	24.38	254.2	7.1	2.366
2	27.608	10489.2	252.2	97.634



White solid; Mp 102.3–103.7 °C; 91% ee; dr = 16:1; 96% yield; $[\alpha]_D^{22} +21.2$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.95–7.82 (m, 2H), 7.63–7.27 (m, 8H), 7.08–6.84 (m, 2H), 6.69 (dd, *J* = 6.7, 2.9, 2H), 4.93 (dd, *J* = 13.1, 11.6, 1H), 4.61–4.33 (m, 2H), 3.89–3.74 (m, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.5, 160.5, 152.7, 132.4, 130.5, 130.1, 130.0, 129.6, 129.4, 129.3, 125.7, 124.9, 123.3, 114.8, 88.2, 74.4, 55.4, 50.8; HRMS (ESI) *m/z* 433.1404 (M+H⁺), Calcd for C₂₄H₂₁N₂O₆ 433.1400; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; *t*_R = 30.3 min (minor peak), 32.7 min, 55.3 min, 62.4 min (major peak).

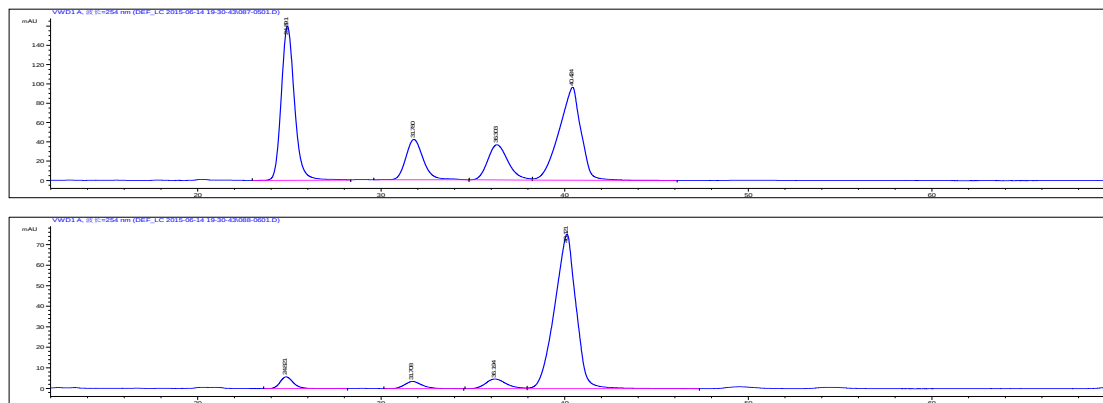


Entry	Retention Time	Area	Height	%Area
1	30.31	378.4	7.5	2.867
2	32.74	367.1	6.3	2.782
3	55.338	583.9	5.8	4.425
4	62.475	11867.2	101.7	89.926

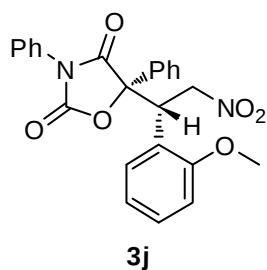


White solid; Mp 106.6–108.1 °C; 91% ee; dr = 14:1; 98% yield; $[\alpha]_D^{22} +27.4$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.86 (dd, *J* = 8.1, 1.4, 2H), 7.62–7.17 (m, 7H), 7.14–6.82 (m, 3H), 6.67 (dt, *J* = 8.7, 3.6, 2H), 4.97 (dd, *J* = 13.4, 11.3, 1H), 4.68–4.28 (m, 2H), 3.78 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 170.3, 160.1, 152.6, 133.1, 132.3, 130.4, 130.2, 130.0, 129.6, 129.3, 125.7, 124.9, 121.4, 115.6, 115.0, 114.4, 129.7, 87.9, 74.2,

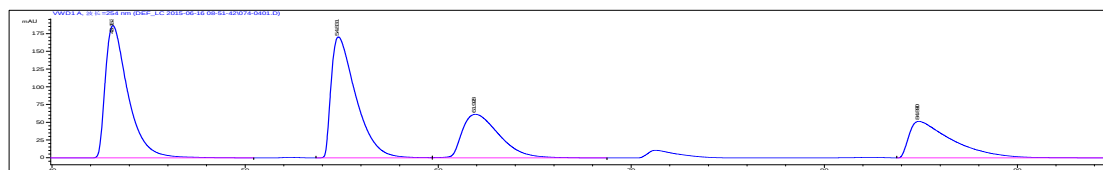
55.4, 51.4; HRMS (ESI) m/z 433.1411 ($M+H^+$), Calcd for $C_{24}H_{21}N_2O_6$ 433.1400; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 24.9 min (minor peak), 31.8 min, 36.3 min, 40.4 min (major peak).

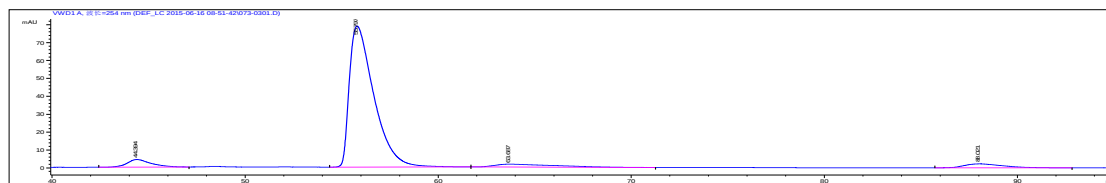


Entry	Retention Time	Area	Height	%Area
1	24.822	280.2	5.7	4.032
2	31.708	223.8	3.4	3.221
3	36.194	356.8	4.6	5.135
4	40.122	6088.6	75.2	87.612

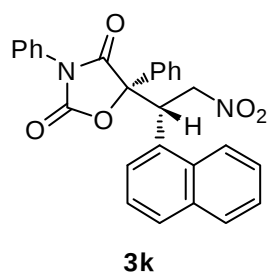


White solid; Mp 117.5–119.2 °C; 91% ee; dr = 8:1; 95% yield; $[\alpha]_D^{22} +31.7$ (c 1.00, $CHCl_3$); 1H NMR (300 MHz, $CDCl_3$) δ 7.89 (dd, J = 8.1, 1.3, 2H), 7.60–7.27 (m, 8H), 6.96 (m, 2H), 6.65 (dd, J = 6.7, 2.9, 2H), 5.39–4.93 (m, 2H), 4.50 (dd, J = 13.5, 4.3, 1H), 3.84 (d, J = 23.4, 2H); ^{13}C NMR (75 MHz, $CDCl_3$) δ 169.9, 158.1, 152.9, 132.9, 130.7, 130.2, 130.0, 129.4, 129.3, 129.2, 128.9, 125.9, 125.1, 121.2, 120.8, 112.1, 74.3, 56.2; HRMS (ESI) m/z 433.1403 ($M+H^+$), Calcd for $C_{24}H_{21}N_2O_6$ 433.1400; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/05; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 44.4 min (minor peak), 55.8 min (major peak), 63.7 min, 88.0 min.

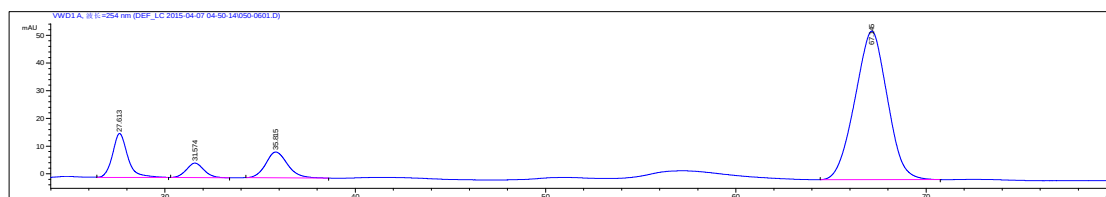
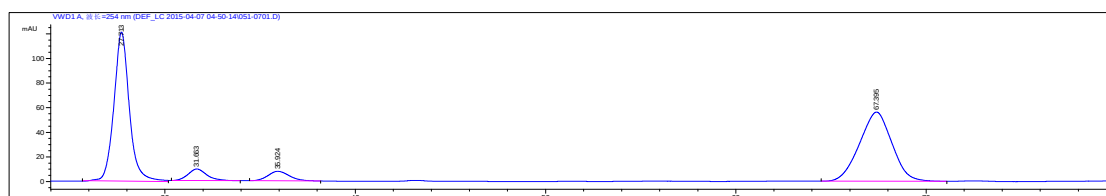




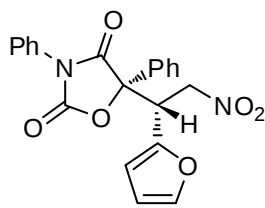
Entry	Retention Time	Area	Height	%Area
1	44.398	353.8	4.2	4.458
2	55.795	6949.3	78.8	87.552
3	63.684	335	1.6	4.221
4	88.011	299.2	2.2	3.769



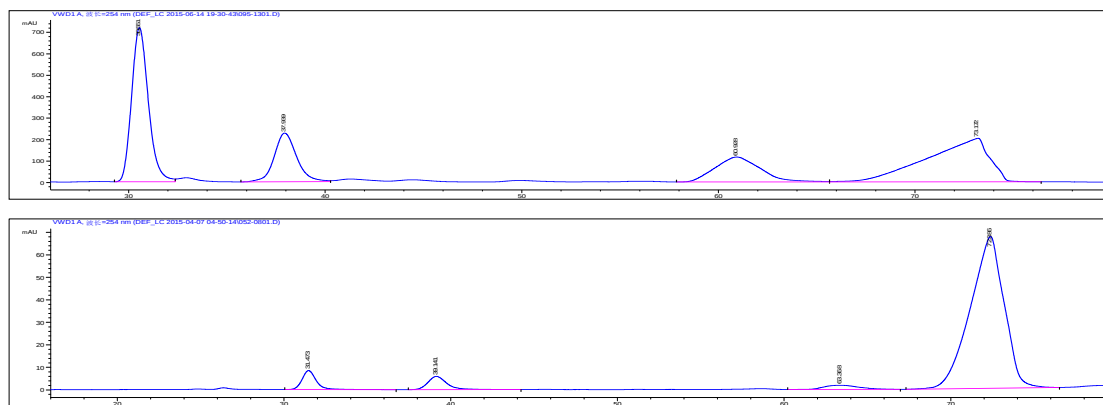
White solid; Mp 130.8–132.3 °C; 86% ee; dr = 7:1; 95% yield; $[\alpha]_D^{22} +21.1$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 8.04–7.70 (m, 6H), 7.65–6.95 (m, 9H), 6.35 (d, *J* = 7.8, 2H), 5.27–4.96 (m, 1H), 4.86–4.38 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 170.4, 152.6, 133.5, 133.2, 132.4, 130.2, 129.7, 129.4, 129.3, 129.1, 129.0, 128.3, 127.6, 127.2, 127.0, 125.7, 125.5, 125.0, 88.2, 74.3, 51.6; HRMS (ESI) *m/z* 453.1441 (M+H⁺), Calcd for C₂₇H₂₁N₂O₅; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; *t*_R = 30.4 min (minor peak), 37.8 min, 41.6 min, 69.3 min (major peak).



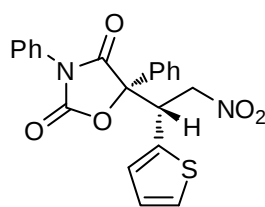
Entry	Retention Time	Area	Height	%Area
1	30.466	534.3	8.9	6.126
2	37.881	628	8.2	7.200
3	41.688	472.9	6.8	5.422
4	69.346	7087.1	46.3	81.252

**3l**

White solid; Mp 152.9–154.1 °C; 94% ee; dr = 16:1; 98% yield; $[\alpha]_D^{22} +25.8$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.89 (m, 7H), 7.71–7.44 (m, 7H), 7.21–6.99 (m, 3H), 6.35 (d, *J* = 7.8, 2H), 5.12 (dd, *J* = 13.5, 11.5, 1H), 4.86–4.38 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 170.5, 152.7, 133.6, 133.2, 132.5, 130.3, 129.8, 129.2, 128.4, 127.7, 127.3, 127.1, 125.6, 125.1, 88.3, 74.4, 51.7; HRMS (ESI) *m/z* 415.0916 (M+Na⁺), Calcd for C₂₁H₁₆N₂O₆Na 415.0906; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; *t_R* = 31.9 min (minor peak), 39.1 min, 63.4 min, 72.4 min (major peak).

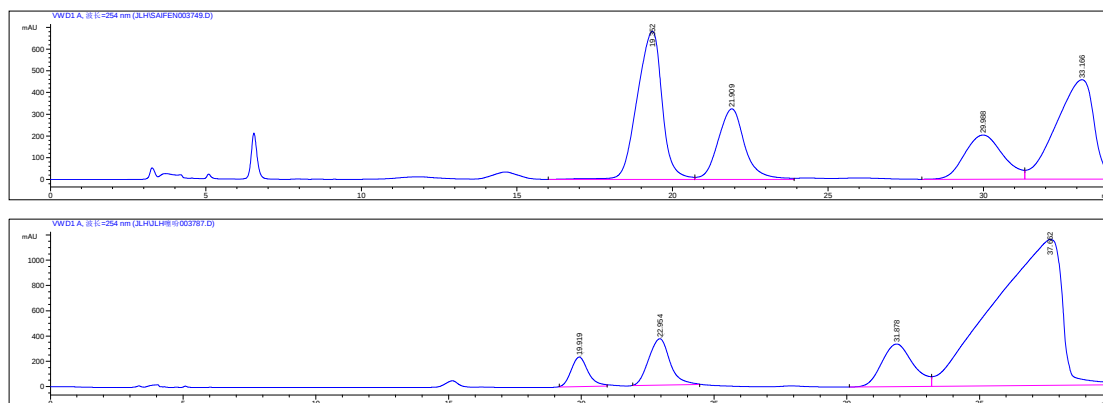


Entry	Retention Time	Area	Height	%Area
1	31.474	495.9	8.4	4.607
2	39.141	479.3	5.9	4.453
3	63.368	276.7	1.9	2.571
4	72.386	9511.3	67.4	88.369

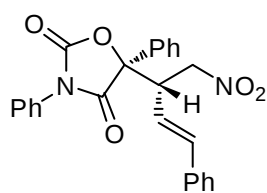
**3m**

White solid; Mp 114.5–116.2 °C; 91% ee; dr = 8:1; 98% yield; $[\alpha]_D^{22} +18.2$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.85 (d, *J* = 6.7, 2H), 7.63–7.47 (m, 3H), 7.44–7.32 (m, 4H), 7.24 (d, *J* = 3.5, 1H), 7.16–6.95 (m, 1H), 6.81 (dd, *J* = 6.1, 3.2, 2H), 4.95–4.75 (m, 2H), 4.44 (dd, *J* = 12.2, 2.3, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 170.2, 152.7, 133.2, 132.0, 130.3, 130.0, 129.7, 129.6, 129.4, 127.6, 127.3, 125.8, 124.9, 87.8, 75.6, 47.1; HRMS (ESI) *m/z* 431.0671 (M+Na⁺), Calcd for C₂₁H₁₆N₂O₅SNa 431.0678; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/05; flow rate 1.0 mL/min; 25 °C; 254 nm; *t_R* = 19.9 min (minor peak), 22.9 min, 31.9

min, 37.7 min (major peak).

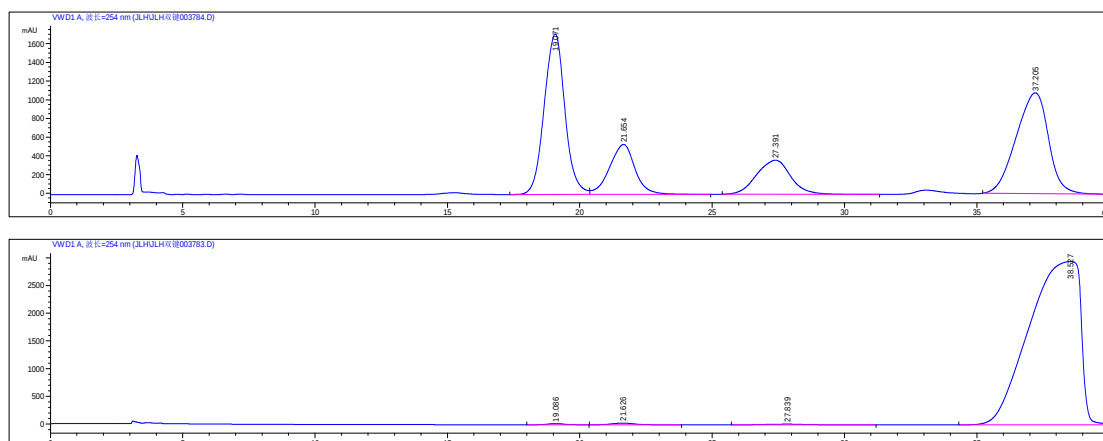


Entry	Retention Time	Area	Height	%Area
1	19.919	10332.5	233.3	3.942
2	22.954	20514.5	367.9	7.826
3	31.878	27664	337.3	10.553
4	37.662	203626.3	1155.5	77.679

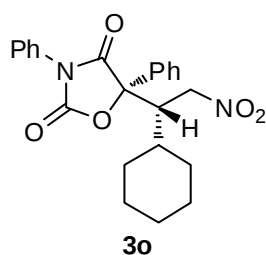


3n

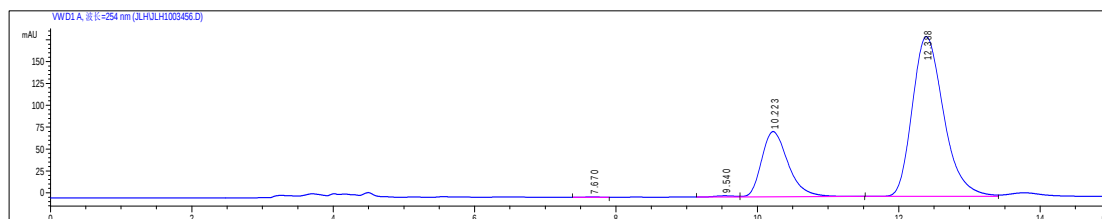
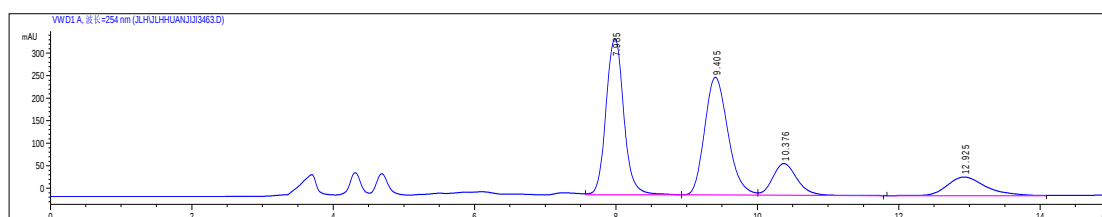
White solid; Mp 170.9–172.1 °C; 99% ee; dr = 19:1; 90% yield; $[\alpha]_D^{22} +20.6$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.91–7.71 (m, 2H), 7.64–7.45 (m, 3H), 7.39–7.29 (m, 8H), 7.07 (dd, *J* = 7.5, 2.1, 2H), 6.88 (d, *J* = 15.9, 1H), 6.12 (dd, *J* = 15.9, 9.1, 1H), 4.74–4.43 (m, 1H), 4.33–4.06 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 170.7, 152.9, 139.8, 135.1, 132.1, 130.2, 129.7, 129.4, 129.0, 128.8, 126.8, 126.1, 125.9, 124.7, 118.4, 88.2, 74.8, 49.4; HRMS (ESI) *m/z* 451.1282 (*M*+Na⁺), Calcd for C₂₅H₂₀N₂O₅Na 451.1270; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/05; flow rate 1.0 mL/min; 25 °C; 254 nm; *t*_R = 19.1 min (minor peak), 21.6 min, 27.8 min, 38.5 min (major peak).



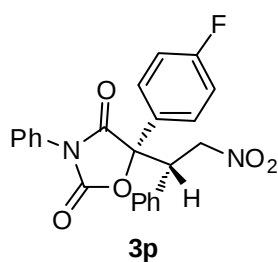
Entry	Retention Time	Area	Height	%Area
1	19.086	835	19	0.208
2	21.626	1663.8	28.3	0.415
3	27.839	1208.5	11.8	0.301
4	38.527	397356	2945.3	99.076



White solid; Mp 130.5–131.7 °C; 50% ee; dr = 15:1; 59% yield; $[\alpha]_D^{22} +23.3$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.68 (ddd, *J* = 7.5, 4.0, 2.3, 2H), 7.58–7.30 (m, 8H), 4.52 (dd, *J* = 14.2, 6.8, 1H), 4.31 (dd, *J* = 14.2, 4.8, 1H), 3.47 (t, *J* = 5.1, 1H), 1.94–1.60 (m, 6H), 1.15 (m, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 171.5, 152.8, 133.6, 130.4, 130.0, 129.5, 129.3, 125.6, 124.9, 89.9, 72.0, 49.6, 39.5, 33.0, 28.9, 26.9, 26.7, 25.8; HRMS (ESI) *m/z* 409.1759 (M+H⁺), Calcd for C₂₃H₂₅N₂O₅ 409.1763; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; *t*_R = 7.6 min, 9.5 min, 10.2 min (minor peak), 12.4 min (major peak).

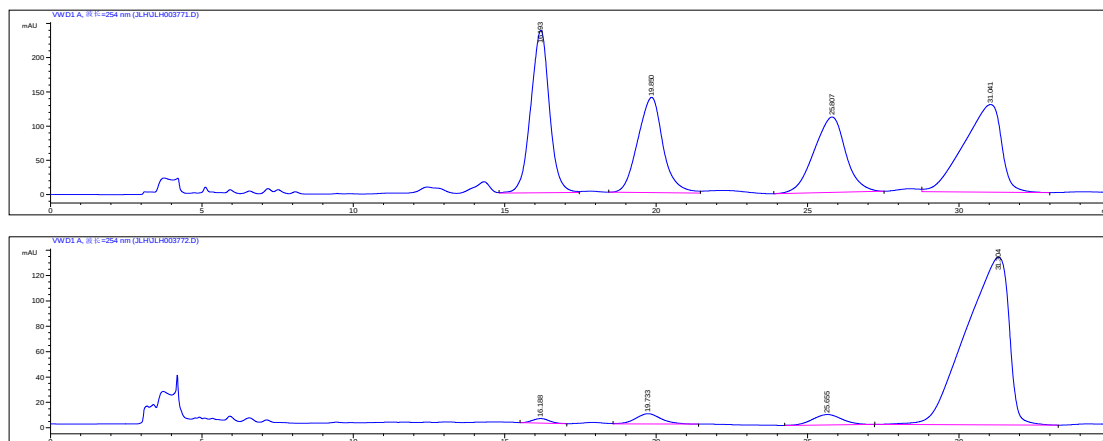


Entry	Retention Time	Area	Height	%Area
1	7.67	4.8	3.2E-1	0.063
2	9.54	24.8	1.3	0.327
3	10.223	1940.5	74.5	25.567
4	12.388	5619.8	182.6	74.043

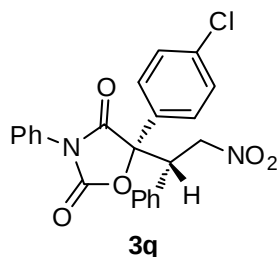


White solid; Mp 162.3–163.1 °C; 98% ee; dr = 14:1; 90% yield; $[\alpha]_D^{22} +20.5$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.89 (dd, *J* = 8.4, 5.1, 2H), 7.58–7.16 (m, 10H), 6.63 (d, *J* = 5.4, 2H),

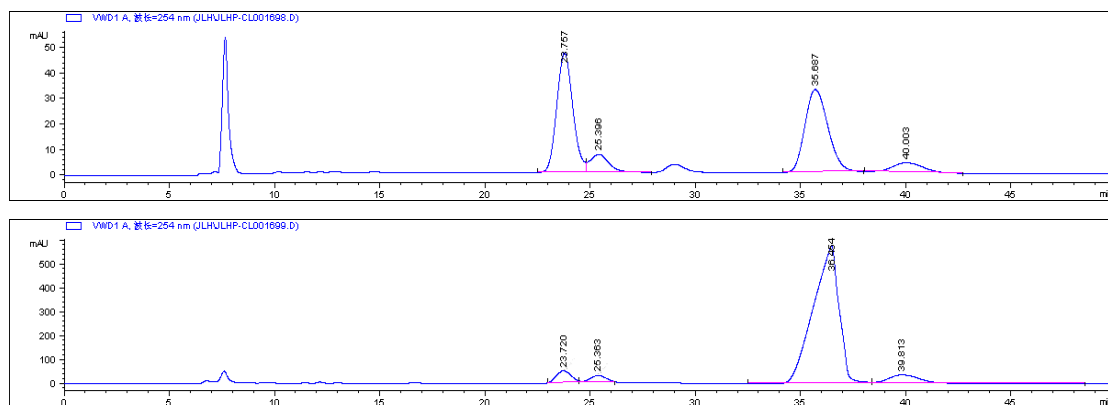
5.13–4.91 (m, 1H), 4.64–4.32 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 170.3, 165.3, 162.0, 152.4, 131.5, 129.7, 129.5, 129.3, 128.1, 127.2, 127.0, 125.7, 116.9, 116.6, 87.6, 74.1, 51.4; HRMS (ESI) m/z 421.1195 ($\text{M}+\text{H}^+$), Calcd for $\text{C}_{23}\text{H}_{18}\text{FN}_2\text{O}_5$ 421.1200; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/05; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 16.2 min (minor peak), 19.7 min, 25.6 min, 31.3 min (major peak).



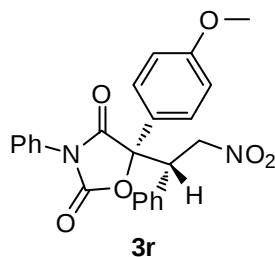
Entry	Retention Time	Area	Height	%Area
1	16.188	136.4	3.6	0.963
2	19.733	477.1	8.1	3.368
3	25.655	545.4	8.2	3.850
4	31.304	13007.1	132.1	91.819



White solid; Mp 151.8–153.1 °C; 91% ee; dr = 12:1; 89% yield; $[\alpha]_D^{22} +21.3$ (c 1.00, CHCl_3); ^1H NMR (300 MHz, CDCl_3) δ 7.74 (d, J = 8.4, 2H), 7.52–7.13 (m, 13H), 6.52 (d, J = 7.9, 2H), 4.96–4.84 (m, 1H), 4.45 (dd, J = 11.1, 3.8, 1H), 4.36 (dd, J = 13.4, 3.9, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 170.1, 152.3, 136.6, 131.4, 130.8, 130.1, 129.8, 129.7, 129.5, 129.4, 129.3, 126.4, 125.8, 87.6, 74.1, 51.4; HRMS (ESI) m/z 437.0901 ($\text{M}+\text{H}^+$), Calcd for $\text{C}_{23}\text{H}_{18}\text{ClN}_2\text{O}_5$ 437.0904; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; t_R = 23.7 min (minor peak), 25.3 min, 36.4 min (major peak), 39.8 min.

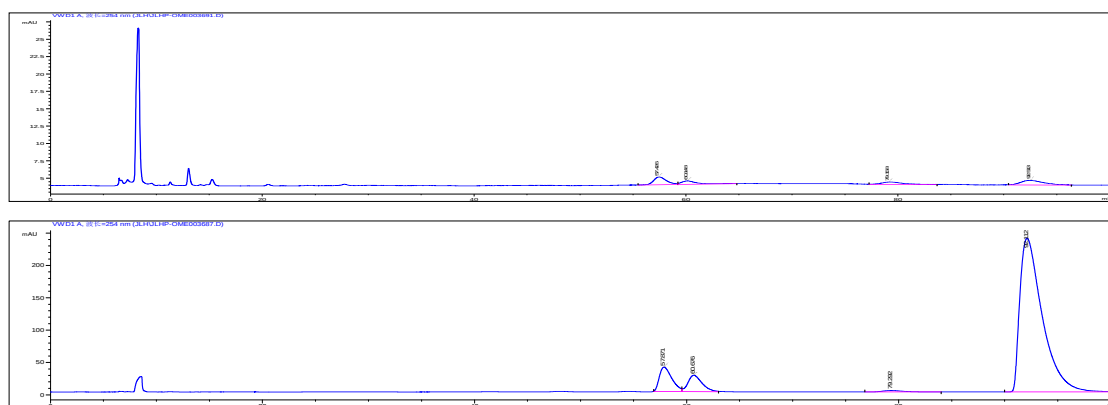


Entry	Retention Time	Area	Height	%Area
1	23.72	2445.5	50.1	4.288
2	25.363	1472.7	27.5	2.582
3	36.454	49165.5	577.9	86.198
4	39.813	3954.4	36.2	6.933

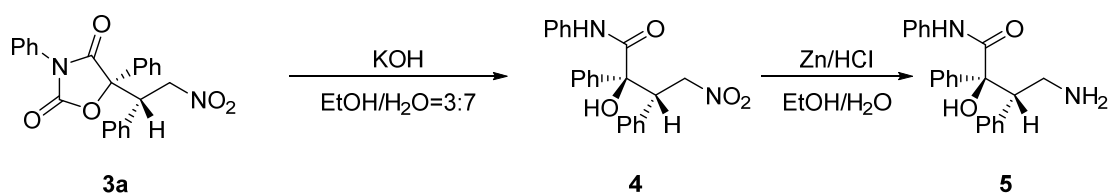


White solid; Mp 156.7–158.2 °C; 85% ee; dr = 11:1; 91% yield; $[\alpha]_D^{22} +25.3$ (c 1.00, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.76 (d, *J* = 8.9, 2H), 7.42 (ddd, *J* = 11.0, 9.7, 4.7, 7H), 7.33–7.28 (m, 2H), 7.09–6.95 (m, 2H), 6.65–6.54 (m, 2H), 4.97 (td, *J* = 14.5, 4.7, 1H), 4.67–4.31 (m, 2H), 3.86 (d, *J* = 4.3, 3H); ¹³C NMR (75 MHz, CDCl₃)

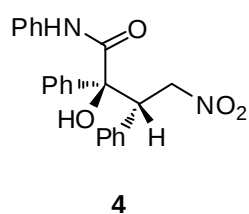
δ 170.6, 160.9, 152.7, 131.8, 129.9, 129.5, 129.4, 129.3, 129.1, 127.1, 126.3, 125.7, 124.0, 114.9, 87.9, 74.3, 55.5, 51.4; HRMS (ESI) *m/z* 455.1225 (M+Na⁺), Calcd for C₂₄H₂₀N₂O₆Na 455.1219; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm) + CHIRALPAK ID-3 (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; *t*_R = 57.8 min (minor peak), 60.6 min, 79.3 min, 92.1 min (major peak).



Entry	Retention Time	Area	Height	%Area
1	57.871	3100.7	37.8	7.590
2	60.676	2340	25.4	5.728
3	79.292	312.7	2.4	0.765
4	92.112	35097.4	237.6	85.916

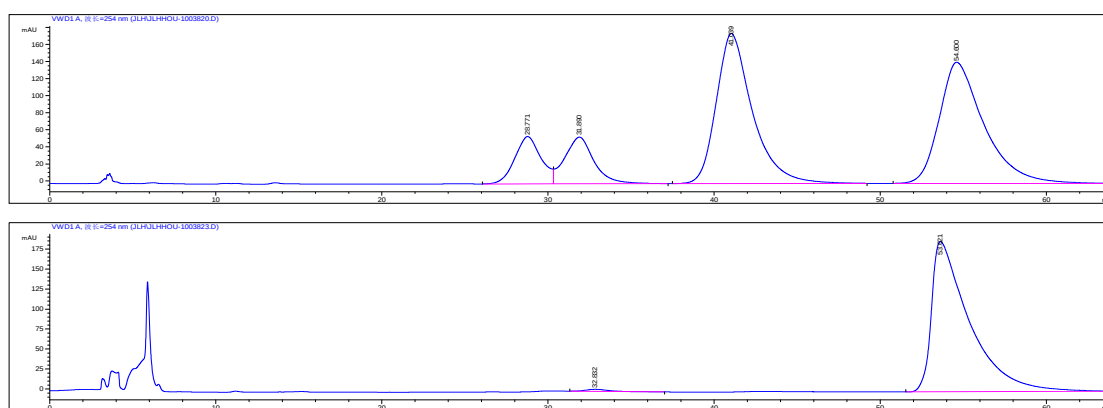
4. Synthesis and characterization of **4**

A mixture of **3a** (0.169 g, 0.42 mmol) and KOH (0.094 g, 1.68 mmol) in a mixture of EtOH/H₂O (3:7) (10 mL) was stirred at room temperature for 1 h. The solvent was removed under vacuum, and the residue was dissolved in CH₂Cl₂ (10 mL) and washed with a 5% aqueous solution of HCl until neutral. The organic layer was dried (Na₂SO₄), and the solvent removed under vacuum. The residue was purified by column chromatography *silica gel* to give **4** as a white solid.



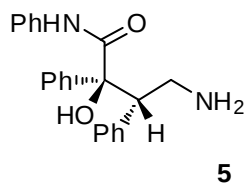
White solid; Mp 152.8–154.1 °C; >99% ee; dr > 19:1; 85% yield; $[\alpha]_{\text{D}}^{22} +28.8$ (c 1.00, CHCl₃); ¹H NMR (300 MHz, Acetone) δ 8.97 (s, 1H), 7.97 (dd, *J* = 10.2, 8.8, 2H), 7.76–6.44 (m, 13H), 5.70 (s, 1H), 5.12–4.80 (m, 2H), 4.57 (dd, *J* = 12.4, 2.9, 1H); ¹³C NMR (75 MHz,

Acetone) δ 169.8, 140.3, 137.9, 135.7, 130.1, 128.6, 128.5, 128.2, 128.0, 127.8, 125.8, 124.0, 120.0, 80.6, 76.7, 50.5; HRMS (ESI) *m/z* 377.1502 (M+H⁺), Calcd for C₂₂H₂₁N₂O₄ 377.1501; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; *t_R* = 28.8 min, 31.9 min, 41.0 min (minor peak), 54.6 min (major peak).

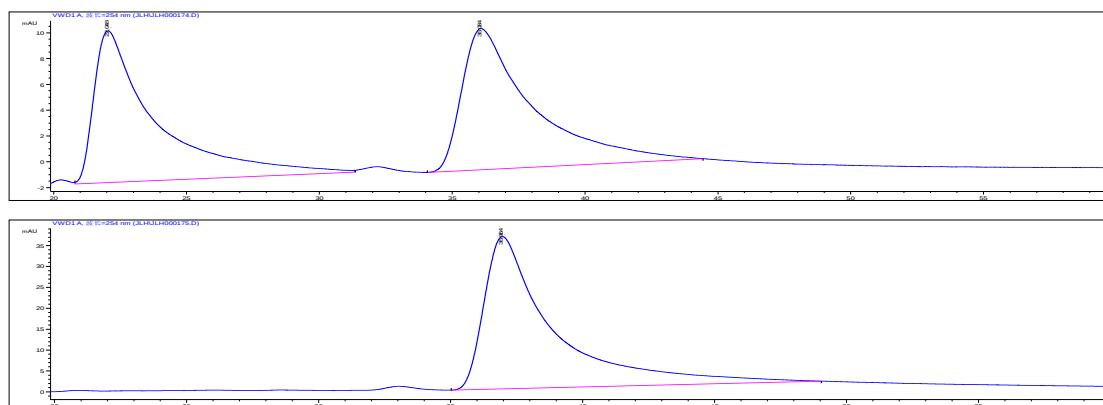


Entry	Retention Time	Area	Height	%Area
1	32.832	242.4	2.6	0.805
2	53.621	29879.6	187.7	99.195

To a solution of **3a** (68 mg, 0.18 mmol) in EtOH/H₂O (1.7 and 0.2 mL) was added Zn dust (130 mg, 2.02 mmol) and *conc.* HCl (0.34 mL). The reaction mixture was refluxed for 4 hours. Reaction quenched with sat. NH₄Cl (aq) (10 mL) and extracted with CH₂Cl₂ (3 x 10 mL), dried (magnesium sulfate), filtered and concentrated *in vacuo* to afford the compound **5** as a colorless oil.



Colorless oil; >99% ee; dr > 19:1; 78% yield; $[\alpha]_D^{22} +30.2$ (*c* 1.00, CHCl₃); ¹H NMR (300 MHz, DMSO) δ 9.37 (d, *J* = 42.8, 1H), 8.12–6.80 (m, 15H), 4.04 (d, *J* = 9.3, 2H), 3.22–2.95 (m, 2H), 2.70 (d, *J* = 12.8, 1H); ¹³C NMR (75 MHz, DMSO) δ 171.3, 142.9, 138.5, 130.8, 128.8, 128.5, 128.3, 127.9, 127.4, 126.5, 124.0, 120.3, 82.0, 50.8, 41.4; HRMS (ESI) *m/z* 347.1712 (M+H⁺), Calcd for C₂₂H₂₁N₂O₄ 347.1715; The ee was determined by HPLC analysis. CHIRALPAK IA (4.6 mm i.d. x 250 mm); Hexane/2-propanol = 80/10; flow rate 1.0 mL/min; 25 °C; 254 nm; *t*_R = 22.1 min (minor peak), 36.1 min (major peak).



Entry	Retention Time	Area	Height	%Area
1	36.964	6657.3	36.4	100.000

5. Copies of NMR Spectra

