

***Supplementary Information***

**The photochemical thiol-ene reaction as a versatile method  
for the synthesis of glutathione S-conjugates targeting the  
bacterial potassium efflux system Kef**

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**General experimental.** All reagents were purchased from Sigma Aldrich or Alfa Aesar and were used without further purification. The UV light source was a Philips HB175 Facial Solarium (UVA, 365 nm, P = 4 × 15 W). Reverse phase column chromatography was carried out on Fluka Ltd silica gel 100 C18-reversed phase, under a positive pressure of compressed air. Analytical TLC analysis was performed using Merck 60 RP-18 F<sub>254</sub>S aluminium-supported thin layer chromatography sheets and visualized using ninhydrin. <sup>1</sup>H, <sup>13</sup>C and <sup>19</sup>F NMR spectra were recorded on a Bruker Avance 400 (400 MHz, 100 MHz) or Bruker Avance III (500 MHz, 125 MHz). <sup>1</sup>H and <sup>13</sup>C spectra were assigned using 2D NMR experiments including COSY, HSQC and HMBC. Mass spectra were recorded on either a Micromass LCT Premier spectrometer (low resolution) or Bruker MicroTOF spectrometer (high resolution) using electrospray ionization (positive or negative mode, from solutions of methanol, acetonitrile or water). *m/z* values are reported in Dalton and followed by their percentage abundance in parentheses. Melting points were determined using a Kofler hot stage microscope and are uncorrected. Infrared Spectra were obtained either as a thin film of solid or solution in CHCl<sub>3</sub>. The spectra were recorded on a Bruker Tensor 27 spectrometer. Absorption maxima are reported in wavenumbers (cm<sup>-1</sup>). Microanalyses were performed by the Elemental Analysis Service, London Metropolitan University. Specific optical rotations were measured using Perkin Elmer Model 241 and 341 polarimeters, in cells with a path length of 1 dm. The light source was maintained at 589 nm. The concentration (*c*) is expressed in g/100 mL (equivalent to g/0.1 dm<sup>3</sup>). Specific rotations are denoted [α]<sub>D</sub><sup>T</sup> and are given in implied units of 10<sup>-1</sup> deg cm<sup>2</sup> g<sup>-1</sup> (T = ambient temperature in °C).

**General procedure for the synthesis of GS-X.** The desired alkene (0.34 mmol, 1 eq), L-glutathione (420 mg, 1.36 mmol, 4 eq) and 2,2-dimethoxyphenyl acetophenone (17 mg, 0.07 mmol, 0.2 eq) were stirred at RT in THF/H<sub>2</sub>O (1:2, 3 mL) in the presence of light (365 nm, 4 × 15 W) for 5 h. After this time the reaction was filtered, and the solid washed with ethanol and water. The crude solid was further purified by crystallization from boiling H<sub>2</sub>O and ethanol (×2) unless otherwise stated. [Note: where no precipitate formed, the reaction solution was washed with dichloromethane (×2) and the aqueous layer lyophilized. In this case, the crude material was purified by RP C-18 silica gel column chromatography].

**S-Hexylglutathione (5a)** was isolated as a colourless solid (130 mg, 97%): R<sub>f</sub> 0.1 (MeOH/H<sub>2</sub>O 50:50); [α]<sub>D</sub><sup>20</sup> -23.8 (c 0.5, 2 M NaOH); m.p. 235 °C (dec.) (EtOH/H<sub>2</sub>O); ν<sub>max</sub> (thin film)/cm<sup>-1</sup>; 3371 (m), 3343 (m), 2954 (m), 1673 (s), 1646 (s), 1514 (s), 1432 (m); <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 4.42 (dd, J = 9.2, 4.6 Hz, 1H), 3.65 (d, J<sub>AB</sub> = 17.0 Hz, 1H), 3.58 (d, J<sub>BA</sub> = 17.0 Hz, 1H), 3.11 (dd, J = 6.9, 5.9 Hz, 1H), 2.94 (dd, J = 14.9, 4.6 Hz, 1H), 2.70 (dd, J = 14.9, 9.2 Hz, 1H), 2.44 (td, J = 7.2, 1.9 Hz, 2H), 2.28-2.18 (m, 2H), 1.84-1.62 (m, 2H), 1.42 (qn, J = 7.2 Hz, 2H), 1.26-1.17 (m, 2H), 1.16-1.09 (m, 4H), 0.71 (t, J = 6.9 Hz, 3H); <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 182.5, 176.3, 176.2, 172.1, 55.5, 53.0, 43.3, 32.9, 32.2, 31.5, 30.8, 30.5, 28.5, 27.5, 21.8, 13.3; HRMS m/z (ES<sup>-</sup>) [Found: (M-H)<sup>-</sup> 390.1697. C<sub>16</sub>H<sub>28</sub>N<sub>3</sub>O<sub>6</sub>S requires M<sup>-</sup>, 390.1704.]; m/z (ES<sup>-</sup>) 390.1 ([M-H]<sup>-</sup>, 100%); Anal. Calcd. for C<sub>16</sub>H<sub>29</sub>N<sub>3</sub>O<sub>6</sub>S: C, 49.0; H, 7.4; N, 10.7. Found: C, 49.0; H, 7.5; N, 10.7. The data are in agreement with the available literature values.<sup>1</sup>

**S-Cyclohexylglutathione (5b)** was isolated as a colourless solid (79 mg, 59 %): R<sub>f</sub> 0.36 (MeOH:H<sub>2</sub>O 50:50); [α]<sub>D</sub><sup>20</sup> -21.5 (c 0.5, 2 M NaOH); ν<sub>max</sub> (thin film)/cm<sup>-1</sup>; 3353 (w), 2927 (w), 1675 (s), 1647 (s), 1515 (s), 1312 (m), 1231 (m); <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 4.42 (dd, J = 9.0, 4.2 Hz, 1H), 3.66 (d, J<sub>AB</sub> = 17.2 Hz, 1H), 3.59 (d, J<sub>BA</sub> = 17.2 Hz, 1H), 3.12 (dd, J = 7.4, 6.0 Hz, 1H), 2.98 (dd, J = 14.1, 4.9 Hz, 1H), 2.73 (dd, J = 14.1, 9.0 Hz, 1H), 2.71-2.63 (m, 1H), 2.35-2.24 (m, 2H), 1.91-1.70 (m, 4H), 1.67-1.60 (m, 2H), 1.54-1.47 (m, 1H), 1.27-1.09 (m, 5H); <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 182.5, 176.3, 176.2, 172.0, 55.5, 53.4, 43.4, 43.3, 33.1, 33.0, 32.3, 30.9, 30.8, 25.5, 25.4, 25.2; HRMS m/z (ES<sup>-</sup>) [Found: (M-H)<sup>-</sup> 388.1547. C<sub>16</sub>H<sub>26</sub>N<sub>3</sub>O<sub>6</sub>S requires M<sup>-</sup>, 388.1548.]; m/z (ES<sup>-</sup>) 388.1 ([M-H]<sup>-</sup>, 100%); Anal. Calcd. for C<sub>16</sub>H<sub>27</sub>N<sub>3</sub>O<sub>6</sub>S: C, 49.3; H, 6.9; N, 10.7. Found: C, 49.4; H, 6.9; N, 10.7.

**S-Decylglutathione (5c)** was isolated as a colourless solid (69 mg, 45 %): R<sub>f</sub> 0.1 (MeOH/H<sub>2</sub>O 50:50); [α]<sub>D</sub><sup>20</sup> -25.0 (c 0.5, 2 M NaOH); m.p. 225 °C (dec.) (EtOH/H<sub>2</sub>O); ν<sub>max</sub> (PTFE card)/cm<sup>-1</sup>; 3277 (m), 2923 (m), 2853 (m), 1582 (s), 1401 (w); <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 4.37 (dd, J = 8.9, 5.0 Hz,

1H), 3.61 (d,  $J_{AB} = 17.2$  Hz, 1H), 3.53 (d,  $J_{BA} = 17.2$  Hz, 1H), 3.11 (dd,  $J = 6.5$ , 6.5 Hz, 1H), 2.91-2.82 (m, 1H), 2.68-2.59 (m, 1H), 2.40 (t,  $J = 6.5$  Hz, 2H), 2.27-2.16 (m, 2H), 1.89-1.67 (m, 2H), 1.48-1.35 (m, 2H), 1.26-1.18 (m, 2H), 1.16-1.07 (m, 12H), 0.71 (t,  $J = 6.5$  Hz, 3H);  $^{13}\text{C}$  NMR (125 MHz, D<sub>2</sub>O/NaOD, pH 12):  $\delta$  182.4, 176.2, 176.0, 172.0, 55.5, 53.2, 43.4, 33.1, 32.2, 31.8, 31.5, 30.9, 29.1, 29.0, 28.95, 28.9, 28.7, 28.2, 22.3, 13.7; HRMS *m/z* (ES<sup>-</sup>) [Found: (M-H)<sup>-</sup> 446.2343. C<sub>20</sub>H<sub>36</sub>N<sub>3</sub>O<sub>6</sub>S requires M<sup>-</sup>, 446.2330.]; *m/z* (ES<sup>-</sup>) 446 ([M-H]<sup>-</sup>, 100%); Anal. Calcd. for C<sub>20</sub>H<sub>37</sub>N<sub>3</sub>O<sub>6</sub>S: C, 53.7; H, 8.3; N, 9.4. Found: C, 53.6; H, 8.4; N, 9.4.

**S-Dodecylglutathione (5d)** was isolated as a colourless solid (111 mg, 69%): R<sub>f</sub> 0.1 (MeOH/H<sub>2</sub>O 50:50);  $[\alpha]_D^{20} -6.8$  (c 0.5, 2 M NaOH); m.p. 240 °C (dec.) (EtOH/H<sub>2</sub>O);  $\nu_{\text{max}}$  (PTFE card)/cm<sup>-1</sup>; 3279 (m), 2980 (w), 2920 (w), 2850 (w), 1576 (w), 1556 (w), 1310 (s);  $^1\text{H}$  NMR (400 MHz, D<sub>2</sub>O /NaOD, pH 12):  $\delta$  4.31 (dd,  $J = 8.3$ , 5.8 Hz, 1H), 3.61 (d,  $J_{AB} = 17.1$  Hz, 1H), 3.44 (d,  $J_{BA} = 17.1$  Hz, 1H), 3.03 (dd,  $J = 6.8$ , 6.8 Hz, 1H), 2.77 (dd,  $J = 13.5$ , 5.8 Hz, 1H), 2.67-2.50 (m, 1H), 2.33 (q,  $J = 7.0$  Hz, 2H), 2.19-2.08 (m, 2H), 1.75-1.56 (m, 2H), 1.38-1.27 (m, 2H), 1.19-1.10 (m, 2H), 1.08-0.97 (m, 16H), 0.62 (t,  $J = 7.0$  Hz, 3H);  $^{13}\text{C}$  NMR (125 MHz, D<sub>2</sub>O/NaOD, pH 12):  $\delta$  182.3, 176.2, 175.7, 171.9, 55.6, 53.2, 43.5, 33.2, 32.0, 31.9, 30.9, 29.7, 29.7, 29.6, 29.4, 29.3, 29.3, 28.7, 22.6, 13.9; HRMS *m/z* (ES<sup>-</sup>) [Found: (M-H)<sup>-</sup> 474.2642. C<sub>22</sub>H<sub>40</sub>N<sub>3</sub>O<sub>7</sub>S requires M, 474.2643.]; *m/z* (ES<sup>-</sup>) 474 ([M-H]<sup>-</sup>, 100%).

**S-Hexadecylglutathione (5e)** was isolated as a colourless solid (123 mg, 70%): R<sub>f</sub> 0.1 (MeOH/H<sub>2</sub>O 50:50);  $[\alpha]_D^{20} -10.0$  (c 0.5, 2 M NaOH); m.p. 200 °C (dec.) (EtOH/H<sub>2</sub>O);  $\nu_{\text{max}}$  (PTFE card)/cm<sup>-1</sup>; 3542 (m), 2960 (w), 2920 (w), 2851 (w), 1494 (w), 1309 (s);  $^1\text{H}$  NMR (400 MHz, NaOD/D<sub>2</sub>O, pH 12):  $\delta$  4.39 (dd,  $J = 7.0$ , 7.0 Hz, 1H), 3.69 (d,  $J_{AB} = 17.3$  Hz, 1H), 3.52 (d,  $J_{BA} = 17.3$  Hz, 1H), 3.11 (dd,  $J = 6.4$ , 6.4 Hz, 1H), 2.91-2.80 (m, 1H), 2.75-2.57 (m, 1H), 2.38 (t,  $J = 7.0$  Hz, 2H), 2.27-2.16 (m, 2H), 1.83-1.63 (m, 2H), 1.48-1.35 (m, 2H), 1.26-1.18 (m, 2H), 1.16-1.07 (m, 24H), 0.71 ( $J = 6.7$  Hz, 3H);  $^{13}\text{C}$  NMR (125 MHz, D<sub>2</sub>O/NaOD, pH 12):  $\delta$  182.6, 176.6, 176.0, 171.9, 56.03, 53.2, 43.80, 33.3, 32.2, 32.1, 31.9, 30.9, 29.9, 29.8, 29.5, 29.4, 28.8, 22.6, 13.9; HRMS *m/z* (ES<sup>-</sup>) [Found: (M-H)<sup>-</sup> 530.3296. C<sub>26</sub>H<sub>48</sub>N<sub>3</sub>O<sub>6</sub>S requires M<sup>-</sup>, 530.3264.]; *m/z* (ES<sup>-</sup>) 530 ([M-H]<sup>-</sup>, 100%).

**S-3, 3-Dimethylbutylglutathione (5g)** was isolated as a colourless solid (88 mg, 66%): R<sub>f</sub> 0.27 (MeOH/H<sub>2</sub>O 50:50);  $[\alpha]_D^{20} -30.0$  (c 0.5, 2 M NaOH); m.p. 235 °C (dec.) (EtOH/H<sub>2</sub>O);  $\nu_{\text{max}}$  (thin film)/cm<sup>-1</sup>; 3375 (m), 3348 (m), 2955 (m), 1673 (s), 1645 (s), 1513 (s), 1433 (m), 1248 (m);  $^1\text{H}$  NMR (400 MHz, D<sub>2</sub>O/NaOD, pH 12):  $\delta$  4.44 (dd,  $J = 9.1$ , 4.7 Hz, 1H), 3.67 (d,  $J_{AB} = 17.1$  Hz, 1H), 3.61 (d,  $J_{BA} = 17.1$  Hz, 1H), 3.13 (dd,  $J = 6.6$ , 6.6 Hz, 1H), 2.97 (dd,  $J = 14.2$ , 4.7 Hz, 1H), 2.73 (dd,  $J = 14.2$ , 9.1 Hz, 1H), 2.45-2.40 (m, 2H), 2.20-

2.31 (m, 2H), 1.85-1.63 (m, 2H), 1.39-1.24 (m, 2H), 0.76 (s, 9H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{D}_2\text{O}/\text{NaOD}$ , pH 12):  $\delta$  182.5, 176.3, 176.2, 172.1, 55.5, 52.9, 43.3, 43.1, 32.7, 32.2, 30.8, 29.8, 28.2, 27.2; HRMS  $m/z$  ( $\text{ES}^-$ ) [Found; ( $\text{M}-\text{H}$ ) $^-$  390.1695.  $\text{C}_{16}\text{H}_{28}\text{N}_3\text{O}_6\text{S}$  requires  $\text{M}^-$ , 390.1704.];  $m/z$  ( $\text{ES}^-$ ) 390.1 ([ $\text{M}-\text{H}$ ] $^-$ , 100%); Anal. Calcd. for  $\text{C}_{16}\text{H}_{29}\text{N}_3\text{O}_6\text{S}$ : C, 49.1; H, 7.5; N, 10.7. Found C, 49.1; H, 7.4; N, 10.7.

**S-Propyl-3-hydroxyl glutathione (5h)** was purified by RP C-18 silica gel column chromatography ( $\text{H}_2\text{O}$ ) furnishing a hygroscopic colourless solid (105 mg, 85%):  $R_f$  0.7 ( $\text{H}_2\text{O}$ );  $[\alpha]_D^{20} -47.8$  ( $c$  0.5,  $\text{H}_2\text{O}$ );  $\nu_{\text{max}}$  (thin film)/cm $^{-1}$ ; 3613 (w), 3447 (w), 2365 (m), 1644 (s), 1525 (s), 1407 (m);  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  4.44 (dd,  $J$  = 8.6, 4.9 Hz, 1H), 3.84 (s, 2H), 3.69 (dd,  $J$  = 6.4, 6.4 Hz, 1H), 3.52 (t,  $J$  = 6.3 Hz, 2H), 2.92 (dd,  $J$  = 14.2, 4.9 Hz, 1H), 2.74 (dd,  $J$  = 14.2, 8.7 Hz, 1H), 2.51 (t,  $J$  = 7.3 Hz, 2H), 2.45-2.33 (m, 2H), 2.03 (dd,  $J$  = 7.6, 6.4 Hz, 2H), 1.72-1.63 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  175.1, 173.9, 173.8, 173.1, 60.4, 54.0, 53.4, 41.8, 33.0, 31.4, 31.3, 28.3, 26.3; HRMS  $m/z$  ( $\text{ES}^-$ ) [Found; ( $\text{M}-\text{H}$ ) $^-$  364.1174.  $\text{C}_{13}\text{H}_{22}\text{N}_3\text{O}_7\text{S}$  requires  $\text{M}^-$ , 364.1184.];  $m/z$  ( $\text{ES}^-$ ) 364.1 ([ $\text{M}-\text{H}$ ] $^-$ , 100%); Anal. Calcd. for  $\text{C}_{13}\text{H}_{23}\text{N}_3\text{O}_7\text{S}$ : C, 42.7; H, 6.3; N, 11.5. Found: C, 42.8; H, 6.3; N, 11.6. The data are in agreement with the available literature values.<sup>2</sup>

**S-Propyl-3-amino glutathione (5i)** was purified by RP C-18 silica gel column chromatography ( $\text{H}_2\text{O}$ ) furnishing a hygroscopic colourless solid (101 mg, 80%):  $R_f$  0.7 ( $\text{H}_2\text{O}$ );  $[\alpha]_D^{20} -27.0$  ( $c$  0.5, 2 M NaOH);  $\nu_{\text{max}}$  (thin film)/cm $^{-1}$ ; 2969 (w), 1738 (s), 1366 (s), 1217 (s);  $^1\text{H}$  NMR (500 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  4.50 (dd,  $J$  = 8.7, 5.0 Hz, 1H), 3.70-3.66 (m, 3H), 3.03-2.97 (m, 3H), 2.80 (dd,  $J$  = 14.1, 8.7 Hz, 1H), 2.64-2.53 (m, 2H), 2.50-2.40 (m, 2H), 2.10-2.04 (m, 2H), 1.91-1.82 (m, 2H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  176.1, 174.8, 174.1, 171.9, 54.0, 52.9, 43.3, 38.2, 32.6, 31.4, 28.1, 26.4, 26.1; HRMS  $m/z$  ( $\text{ES}^+$ ) [Found; ( $\text{M}+\text{H}$ ) $^+$  365.1481.  $\text{C}_{13}\text{H}_{25}\text{N}_4\text{O}_6\text{S}$  requires  $\text{M}^+$ , 365.1489.];  $m/z$  ( $\text{ES}^+$ ) 365.1 ([ $\text{M}+\text{H}$ ] $^+$ , 100%).

**S-(6,6-Dimethylbicyclo[3.1.1]heptan-2-yl)methyl glutathione (5k)** was purified by RP C-18 silica gel column chromatography ( $\text{H}_2\text{O}$ ) furnishing a colourless solid (1 mg, 0.006%):  $R_f$  0.25 (MeOH: $\text{H}_2\text{O}$  50:50);  $\nu_{\text{max}}$  (thin film)/cm $^{-1}$ ; 3592 (w), 1738 (s), 1641 (m), 1437 (m), 1370 (s), 1229 (s), 1217 (s);  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  4.43 (dd,  $J$  = 8.9, 5.0 Hz, 1H), 3.77 (s, 2H), 3.65 (dd,  $J$  = 6.3, 6.3 Hz, 1H), 2.92 (dd,  $J$  = 14.4, 5.0 Hz, 1H), 2.72 (dd,  $J$  = 14.4, 8.9 Hz, 1H), 2.62 (m, 2H), 2.46-2.34 (m, 2H), 2.26-2.19 (m, 1H), 2.11-1.92 (m, 3H), 1.92-1.72 (m, 4H), 1.42-1.29 (m, 1H), 1.05 (s, 3H), 0.85 (s, 3H); HRMS  $m/z$  ( $\text{ES}^+$ ) [Found; ( $\text{M}+\text{Na}$ ) $^+$  466.1966.  $\text{C}_{20}\text{H}_{33}\text{N}_3\text{O}_6\text{SNa}$  requires  $\text{M}^+$ , 466.1982.];  $m/z$  ( $\text{ES}^+$ ) 466.1 ([ $\text{M}+\text{Na}$ ] $^+$ , 100%).

**2-(4-Nitrophenyl)-S-ethyl glutathione (6a)** was isolated as a pale yellow solid (20 mg, 13%);  $R_f$  0.4 ( $H_2O/MeOH$  50:50);  $[\alpha]_D^{20} -18.1$  ( $c$  0.5,  $H_2O$ ); m.p. 200 °C (dec.) ( $EtOH/H_2O$ ) (lit. 201-202 °C);  $\nu_{max}$  (thin film)/cm<sup>-1</sup>; 3546 (w), 1646 (s), 1518 (s), 1346 (s); <sup>1</sup>H NMR (400 MHz,  $D_2O$ ):  $\delta$  8.04 (d,  $J$  = 8.5 Hz, 2H), 7.34 (d,  $J$  = 8.5 Hz, 2H), 4.36 (dd,  $J$  = 8.7, 5.2 Hz, 1H), 3.80 (s, 2H), 3.66 (dd,  $J$  = 6.4, 6.4 Hz, 1H), 2.94-2.75 (m, 5H), 2.68 (dd,  $J$  = 14.0, 8.7 Hz, 1H), 2.42-2.31 (m, 2H), 2.06-1.96 (m, 2H); HRMS  $m/z$  (ES<sup>-</sup>) [Found; ( $M-H$ )<sup>-</sup> 455.1246.  $C_{18}H_{23}N_4O_8S$  requires  $M^-$ , 455.1242.];  $m/z$  (ES<sup>-</sup>) 455.1 ([ $M-H$ ]<sup>-</sup>, 100%). The data are in agreement with the available literature values.<sup>1</sup>

**2-(4-Fluorophenyl)-S-ethyl glutathione (6c)** was isolated as a colourless solid (70 mg, 48%);  $R_f$  0.36 ( $MeOH:H_2O$  50:50);  $[\alpha]_D^{20} -19.2$  ( $c$  0.5 in 2 M NaOH); m.p. 220 °C (dec.) ( $EtOH/H_2O$ );  $\nu_{max}$  (thin film)/cm<sup>-1</sup>; 3344 (w), 2945 (w), 1672 (m), 1645 (s), 1509 (s), 1432 (s), 1352 (w), 1312 (w), 1222 (m); <sup>1</sup>H NMR (500 MHz,  $D_2O/NaOD$ , pH 12):  $\delta$  7.18-7.11 (m, 2H), 6.98-6.90 (m, 2H), 4.39 (dd,  $J$  = 9.2, 4.8 Hz, 1H), 3.64 (d,  $J_{AB}$  = 17.3 Hz, 1H), 3.62 (d,  $J_{BA}$  = 17.3 Hz, 1H), 3.10 (dd,  $J$  = 7.3, 6.1 Hz, 1H), 2.90 (dd,  $J$  = 14.1, 4.8 Hz, 1H), 2.80-2.72 (m, 4H), 2.67 (dd,  $J$  = 14.1, 9.2 Hz, 1H), 2.22 (ddd,  $J$  = 10.0, 7.0, 2.5 Hz, 2H), 1.82-1.61 (m, 2H); <sup>13</sup>C NMR (125 MHz,  $D_2O/NaOD$ , pH 12):  $\delta$  182.5, 176.3, 176.2, 172.0, 161.5 (d,  $J$  = 235 Hz), 136.1 (d,  $J$  = 2.9 Hz), 130.2 (d,  $J$  = 8.5 Hz), 115.1 (d,  $J$  = 21.5), 55.6, 52.9, 43.3, 33.9, 33.0, 32.9, 32.2, 30.8; <sup>19</sup>F NMR (125 MHz,  $D_2O/NaOD$ , pH 12);  $\delta$  -117.0; HRMS  $m/z$  (ES<sup>-</sup>) [Found; ( $M-H$ )<sup>-</sup> 428.1296.  $C_{18}H_{23}FN_3O_6SNa$  requires  $M^-$ , 428.1297.];  $m/z$  (ES<sup>-</sup>) 428.1 ([ $M-H$ ]<sup>-</sup>, 100%); Anal. Calcd. for  $C_{18}H_{24}FN_3O_6S$ : C, 50.3; H, 5.6; N, 9.8. Found: C, 50.4; H, 5.6; N, 9.8.

**2-(4-Fluorophenyl)-S-propyl glutathione (6d)** was isolated as a colourless crystalline solid (135 mg, 90%);  $R_f$  0.28 ( $MeOH:H_2O$  50:50); m.p. 170-173 °C ( $EtOH/H_2O$ );  $[\alpha]_D^{20} -6.2$  ( $c$  0.5 in 2 M NaOH);  $\nu_{max}$  (thin film)/cm<sup>-1</sup>; 3368 (w), 3344 (w), 2941 (w), 2289 (m), 1672 (m), 1645 (s), 1509 (s), 1433 (m), 1349 (w), 1217 (s); <sup>1</sup>H NMR (500 MHz,  $D_2O/NaOD$ , pH 12):  $\delta$  7.12 (d,  $J$  = 5.8 Hz, 1H), 7.10 (dd,  $J$  = 6.7, 2.1 Hz, 1H), 6.95 (dd,  $J$  = 6.7, 2.1 Hz, 1H), 6.93 (dd,  $J$  = 6.7, 2.1 Hz, 1H), 4.40 (dd,  $J$  = 9.0, 4.9 Hz, 1H), 3.67 (d,  $J_{AB}$  = 17.2 Hz, 1H), 3.59 (d,  $J_{BA}$  = 17.2 Hz, 1H), 3.20 (dd,  $J$  = 6.2, 6.2 Hz, 1H), 2.93 (dd,  $J$  = 14.4, 4.9 Hz, 1H), 2.71 (dd,  $J$  = 14.4, 9.0 Hz, 1H), 2.55 (t,  $J$  = 7.4 Hz, 2H), 2.41 (td,  $J$  = 7.2, 2.4 Hz, 2H), 2.27 (dd,  $J$  = 7.9, 7.9 Hz, 2H), 1.89-1.68 (m, 4H); <sup>13</sup>C NMR (125 MHz,  $D_2O/NaOD$ , pH 12):  $\delta$  181.8, 176.5, 176.4, 172.3, 161.5 (d,  $J$  = 240 Hz), 137.9, 130.4 (d,  $J$  = 8.0 Hz), 115.4 (d,  $J$  = 21 Hz), 55.7, 53.4, 43.7, 33.4, 33.2, 32.5, 31.1, 30.8, 30.6; <sup>19</sup>F NMR (100 MHz,  $D_2O/NaOD$ , pH 12);  $\delta$  -118.1; HRMS  $m/z$  (ES<sup>+</sup>) [Found; ( $M+Na$ )<sup>+</sup> 466.1420.  $C_{19}H_{26}FN_3NaO_6SNa$  requires  $M^+$ , 466.1419.];  $m/z$  (ES<sup>-</sup>) 442.1 ([ $M-H$ ]<sup>-</sup>, 100%); Anal. Calcd. for  $C_{19}H_{26}FN_3O_6S$ : C, 51.5; H, 5.9; N, 9.5. Found: C, 51.5; H, 5.9; N, 9.4.

**2-(Phenylsulfonyl)-S-ethyl glutathione (6e)** was purified by RP C-18 silica gel column chromatography (H<sub>2</sub>O) furnishing a hygroscopic colourless solid (73 mg, 45%): R<sub>f</sub> 0.54 (MeOH:H<sub>2</sub>O 50:50); [α]<sub>D</sub><sup>20</sup> -15.0 (c 0.5, H<sub>2</sub>O); ν<sub>max</sub> (thin film)/cm<sup>-1</sup>; 2229 (m), 2180 (m), 1741 (s), 1367 (m), 1216 (m); <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O): δ 7.82 (dd, J = 8.7, 1.5 Hz, 2H), 7.69 (tt, J = 7.4, 1.5 Hz, 1H), 7.64 (tt, J = 7.4, 1.5 Hz, 2H), 4.36 (dd, J = 8.3, 5.3 Hz, 1H), 3.81 (s, 2H), 3.61 (dd, J = 6.4, 6.4 Hz, 1H), 3.53 (dd, J = 9.4, 6.1 Hz, 2H), 2.22 (dd, J = 14.1, 5.3 Hz, 1H), 2.75-2.66 (m, 3H), 2.37 (ddd, J = 7.6, 7.6, 3.2 Hz, 2H), 2.09-2.01 (m, 2H); <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O): δ 174.7, 173.7, 173.6, 172.2, 136.5, 135.0, 129.7, 127.8, 54.9, 53.8, 52.7, 41.6, 32.7, 31.2, 25.9, 24.0; HRMS m/z (ES<sup>-</sup>) [Found; (M-H)<sup>-</sup> 474.1020. C<sub>18</sub>H<sub>24</sub>N<sub>3</sub>O<sub>8</sub>S<sub>2</sub> requires M<sup>-</sup>, 474.1010.]; m/z (ES<sup>-</sup>) 474.1 ([M-H]<sup>-</sup>, 100%); Anal. Calcd. for C<sub>18</sub>H<sub>25</sub>N<sub>3</sub>O<sub>8</sub>S<sub>2</sub>: C, 45.5; H, 5.3; N, 8.8. Found: C, 45.5; H, 5.2; N, 8.8. The data are in good agreement with the available literature data.<sup>3</sup>

**2-(4-Methoxyphenyl)-S-ethyl glutathione (6g)** was isolated as a colourless solid (125 mg, 84%): R<sub>f</sub> 0.45 (MeOH:H<sub>2</sub>O 50:50); [α]<sub>D</sub><sup>20</sup> -22.6 (c 0.5, 2 M NaOH); ν<sub>max</sub> (thin film)/cm<sup>-1</sup>; 3349 (w), 1644 (m), 1512 (s), 1245 (m); <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 7.12 (d, J = 8.3 Hz, 2H), 6.84 (d, J = 8.3 Hz, 2H), 4.39 (dd, J = 9.1, 4.9 Hz, 1H), 3.69 (s, 3H), 3.63 (d, J<sub>AB</sub> = 17.2 Hz, 1H), 3.58 (d, J<sub>BA</sub> = 17.2 Hz, 1H), 3.10 (dd, J = 7.1, 5.9 Hz, 1H), 2.90 (dd, J = 14.2, 4.9 Hz, 1H), 2.77-2.72 (m, 4H), 2.67 (dd, J = 14.2, 9.1 Hz, 1H), 2.27-2.19 (m, 2H), 1.82-1.62 (m, 2H); <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 182.4, 176.3, 176.2, 157.3, 133.1, 129.9, 114.0, 55.5, 55.4, 52.9, 43.3, 33.9, 33.1, 32.9, 32.2, 30.8; HRMS m/z (ES<sup>-</sup>) [Found; (M-H)<sup>-</sup> 440.1503. C<sub>19</sub>H<sub>26</sub>N<sub>3</sub>O<sub>7</sub>S requires M<sup>-</sup>, 440.1497.]; m/z (ES<sup>-</sup>) 440.1 ([M-H]<sup>-</sup>, 100%); Anal. Calcd. for C<sub>19</sub>H<sub>27</sub>N<sub>3</sub>O<sub>7</sub>S: C, 51.7; H, 6.2; N, 9.5. Found: C, 51.7; H, 6.1; N, 9.6.

**2-(Phenyl)-S-ethyl glutathione (6f)** was isolated as a colourless solid (75 mg, 53%); R<sub>f</sub> 0.3 (MeOH/H<sub>2</sub>O 50:50); [α]<sub>D</sub><sup>20</sup> -21.0 (c 0.5, 2 M NaOH); m.p. 210 °C (dec.) (EtOH/H<sub>2</sub>O) (lit. 199-200 °C); ν<sub>max</sub> (thin film)/cm<sup>-1</sup>; 3346 (m), 2951 (w), 1672 (s), 1645 (s), 1512 (s), 1432 (m); <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 7.25-7.17 (m, 2H), 7.16-7.09 (m, 3H), 4.36 (dd, J = 9.0, 4.8 Hz, 1H), 3.62 (d, J<sub>AB</sub> = 17.2 Hz, 1H), 3.54 (d, J<sub>BA</sub> = 17.2 Hz, 1H), 3.06 (dd, J = 6.2, 6.2 Hz, 1H), 2.88 (dd, J = 14.2, 4.8 Hz, 1H), 2.78-2.70 (m, 4H), 2.64 (dd, J = 14.2, 9.0 Hz, 1H), 2.18 (dd, J = 7.3, 7.3 Hz, 2H), 1.79-1.57 (m, 2H); <sup>13</sup>C NMR (125 MHz, D<sub>2</sub>O/NaOD, pH 12): δ 182.5, 176.3, 176.2, 172.0, 140.4, 128.7, 126.5, 55.5, 53.6, 43.4, 34.8, 32.9, 32.2, 30.8; HRMS m/z (ES<sup>-</sup>) [Found; (M-H)<sup>-</sup> 410.1392. C<sub>18</sub>H<sub>24</sub>N<sub>3</sub>O<sub>6</sub>S requires M<sup>-</sup>, 410.1391.]; m/z (ES<sup>-</sup>) 410.1 ([M-H]<sup>-</sup>, 100%); Anal. Calcd. for C<sub>18</sub>H<sub>25</sub>N<sub>3</sub>O<sub>6</sub>S: C, 52.5; H, 6.1; N, 10.2. Found: C, 52.6; H, 6.1; N, 10.3. The data are in good agreement with the available literature data.<sup>1</sup>

- (1) Vince, R.; Vince, R.; Daluge, S.; Daluge, S.; Wadd, W. B.; Wadd, W. B. *J. Med. Chem.* **1971**, *14*, 402–404.
- (2) Jin, L.; Baillie, T. A. *Chem. Res. Toxicol.* **1997**, *10*, 318–327.
- (3) Horner, L.; Lindel, H. *Phosphorous and Sulfur and the Related Elements* **1983**, *15*, 1–8.

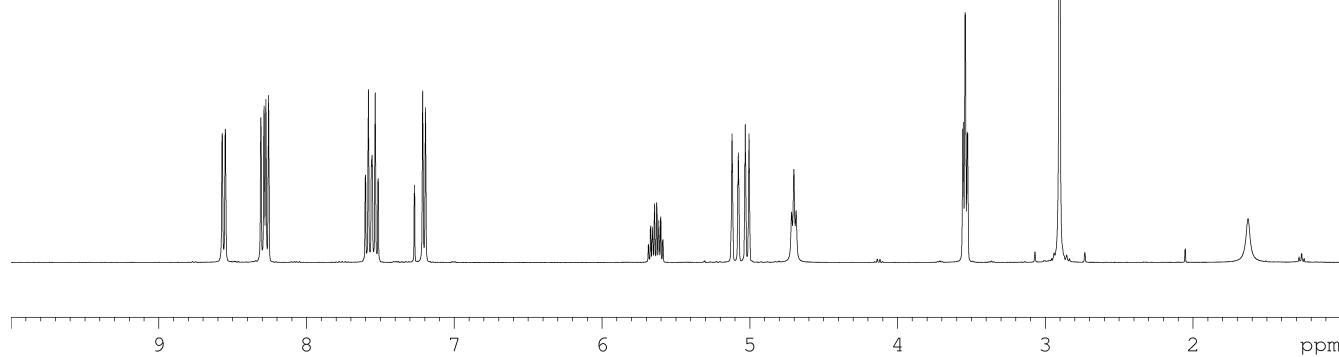
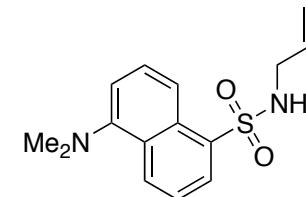
*N*-Allyl-5-(dimethylamino)naphthalene sulfonamide (**2**) –  $^1\text{H}$  NMR

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*N*-Allyl-5-(dimethylamino)naphthalene sulfonamide (**2**) –  $^{13}\text{C}$  NMR

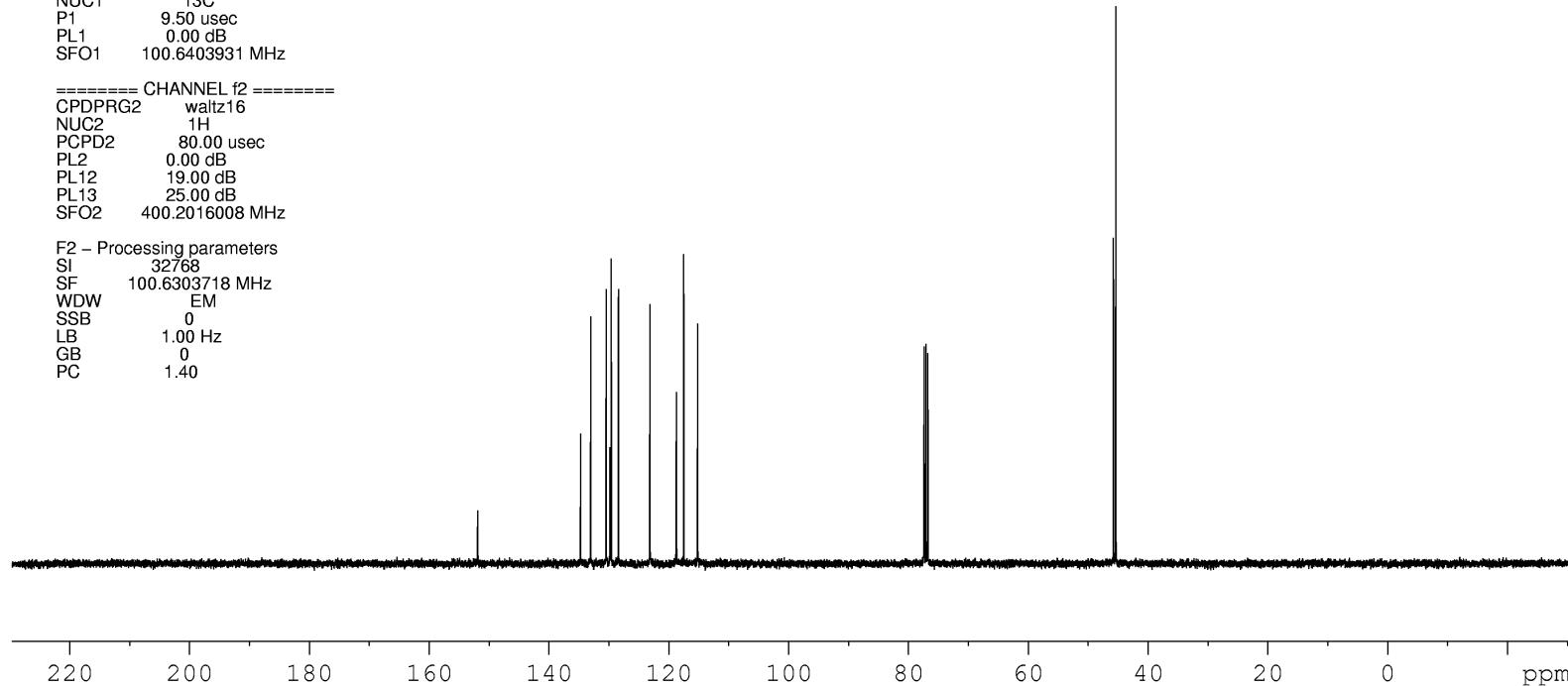
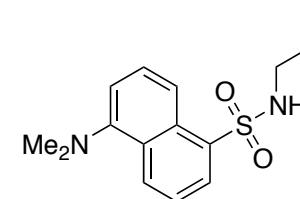
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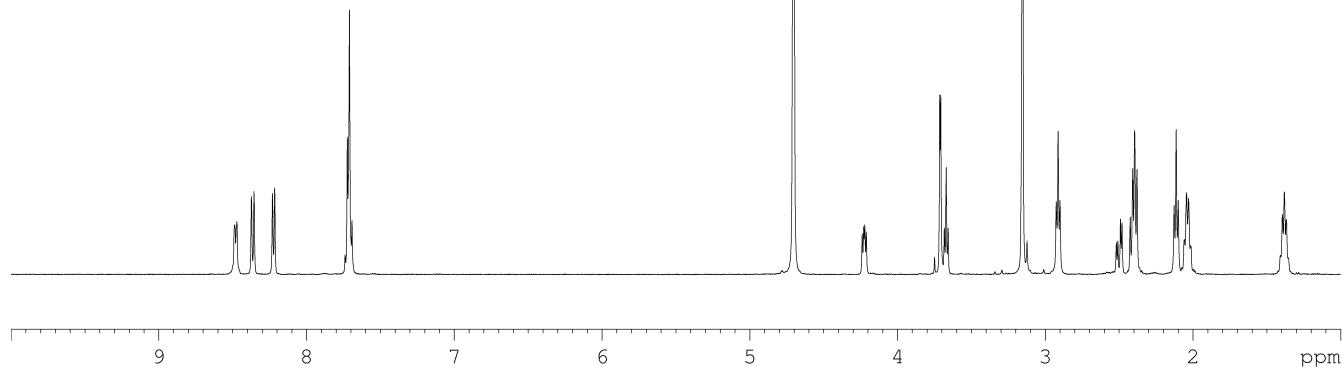
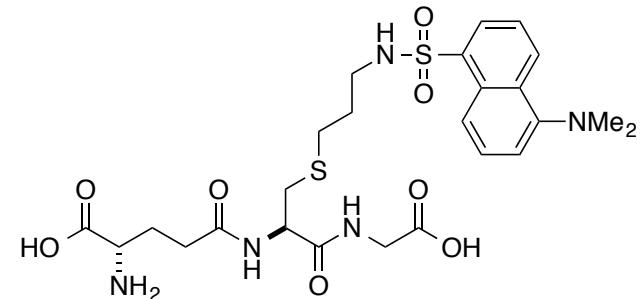
S-(N-Propyl-5-(dimethylamino)naphthalene-1-sulfonamide glutathione (**3**) –  $^1\text{H}$  NMR

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S-(N-Propyl-5-(dimethylamino)naphthalene-1-sulfonamide glutathione (**3**) –  $^{13}\text{C}$  NMR

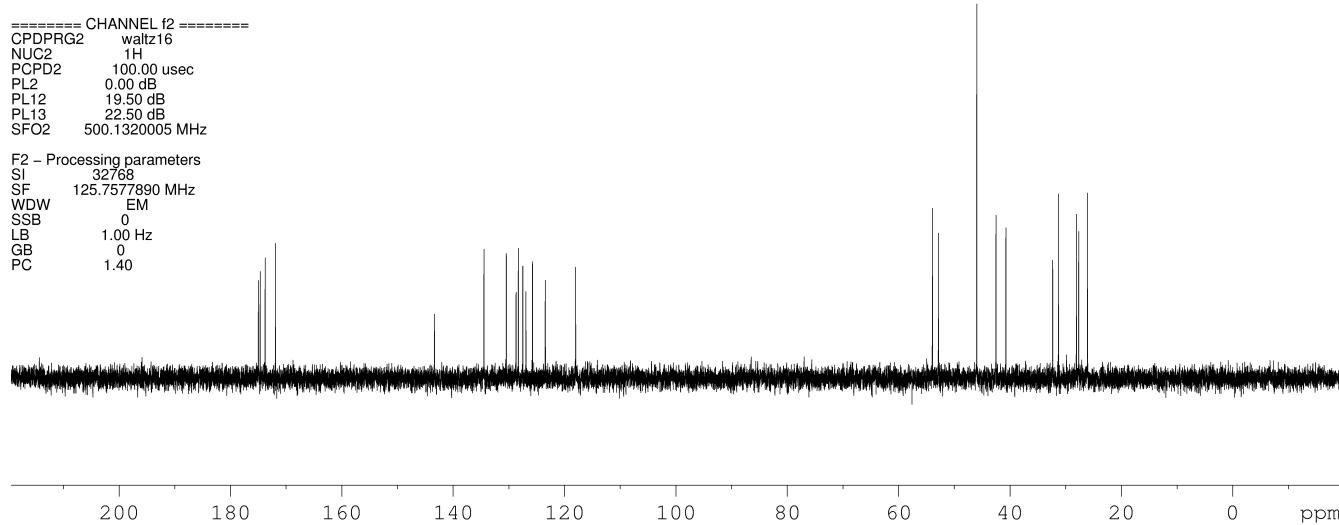
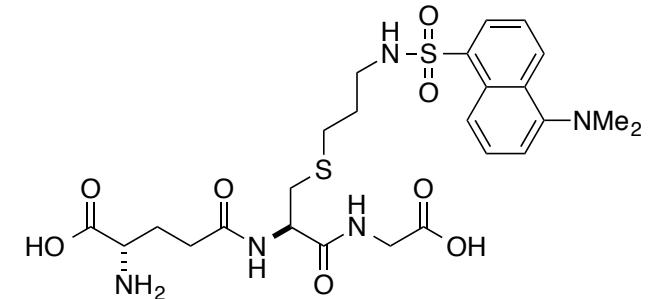
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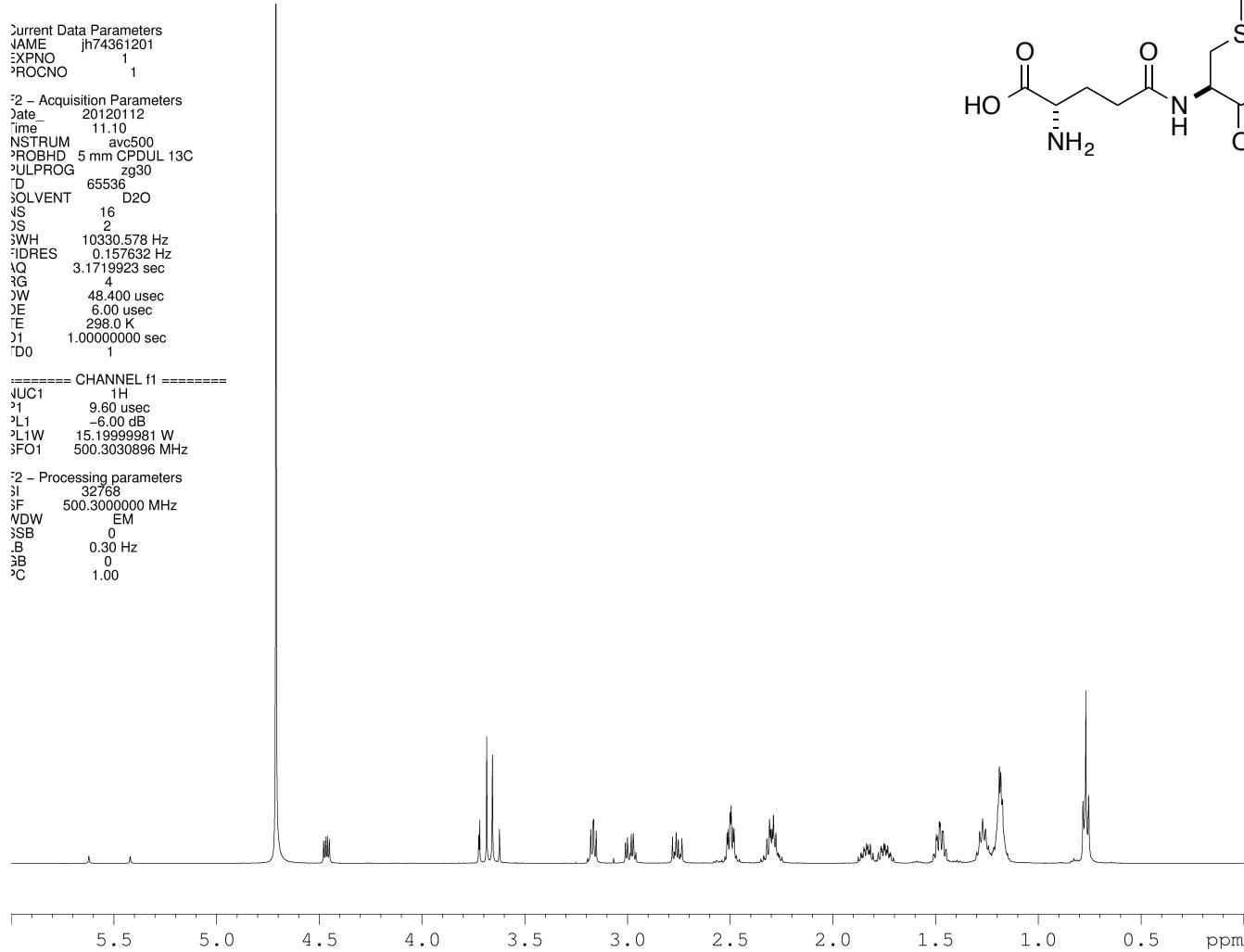
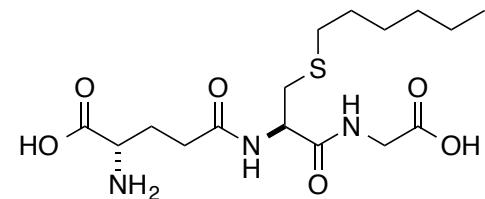
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F2 – Processing parameters  
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### S-Hexylglutathione (**5a**) – $^1\text{H}$ NMR

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### S-Hexylglutathione (**5a**) – $^{13}\text{C}$ NMR

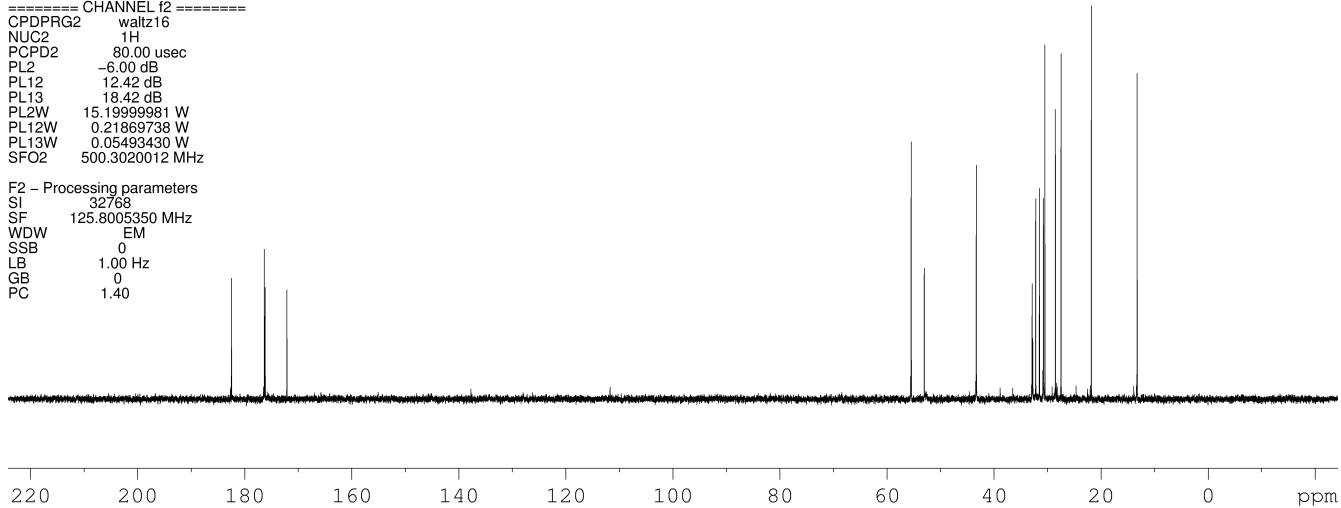
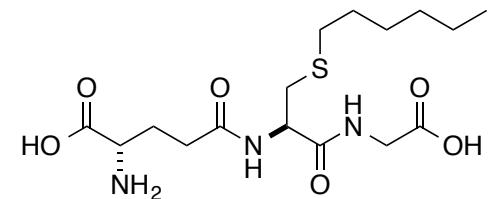
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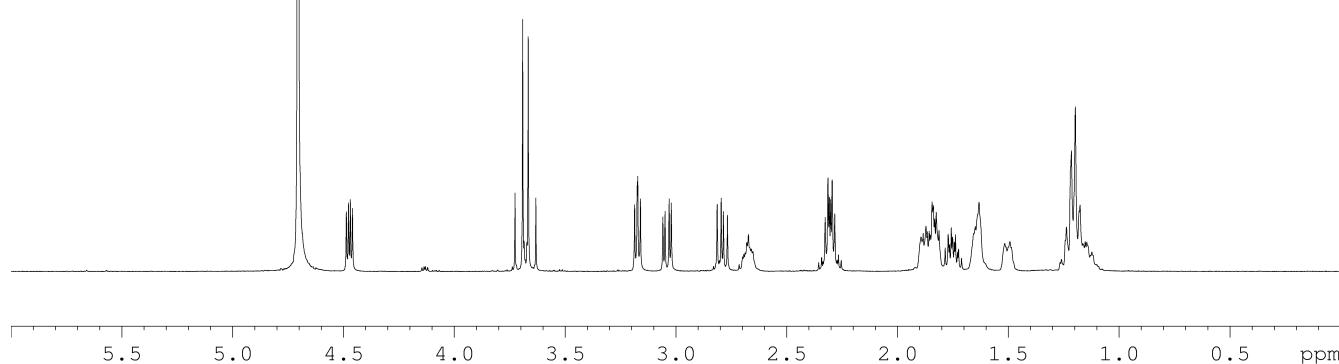
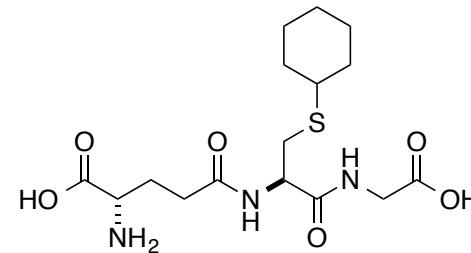
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 TE 298.0 K  
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### S-Cyclohexylglutathione (**5b**) – $^{13}\text{C}$ NMR

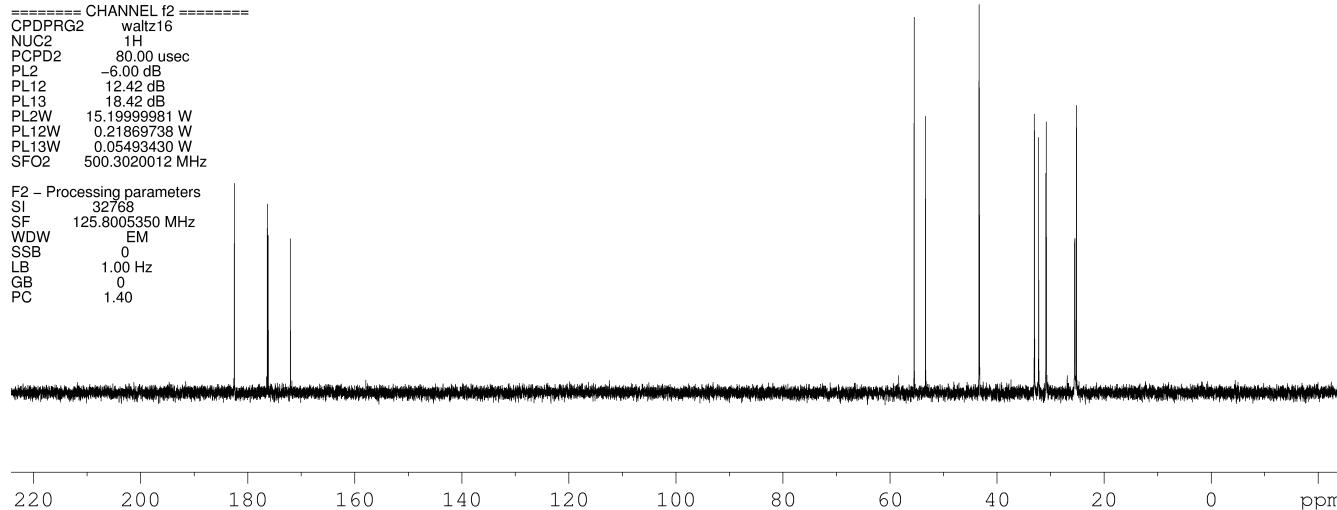
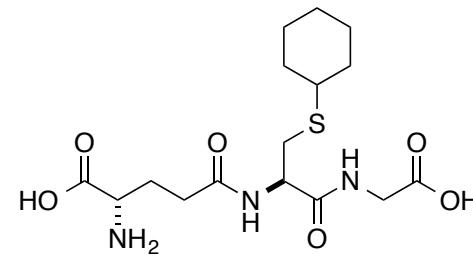
Current Data Parameters  
 NAME jh81302102  
 EXPNO 4  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120221  
 Time 9.41  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1731  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 1820  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.0300000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

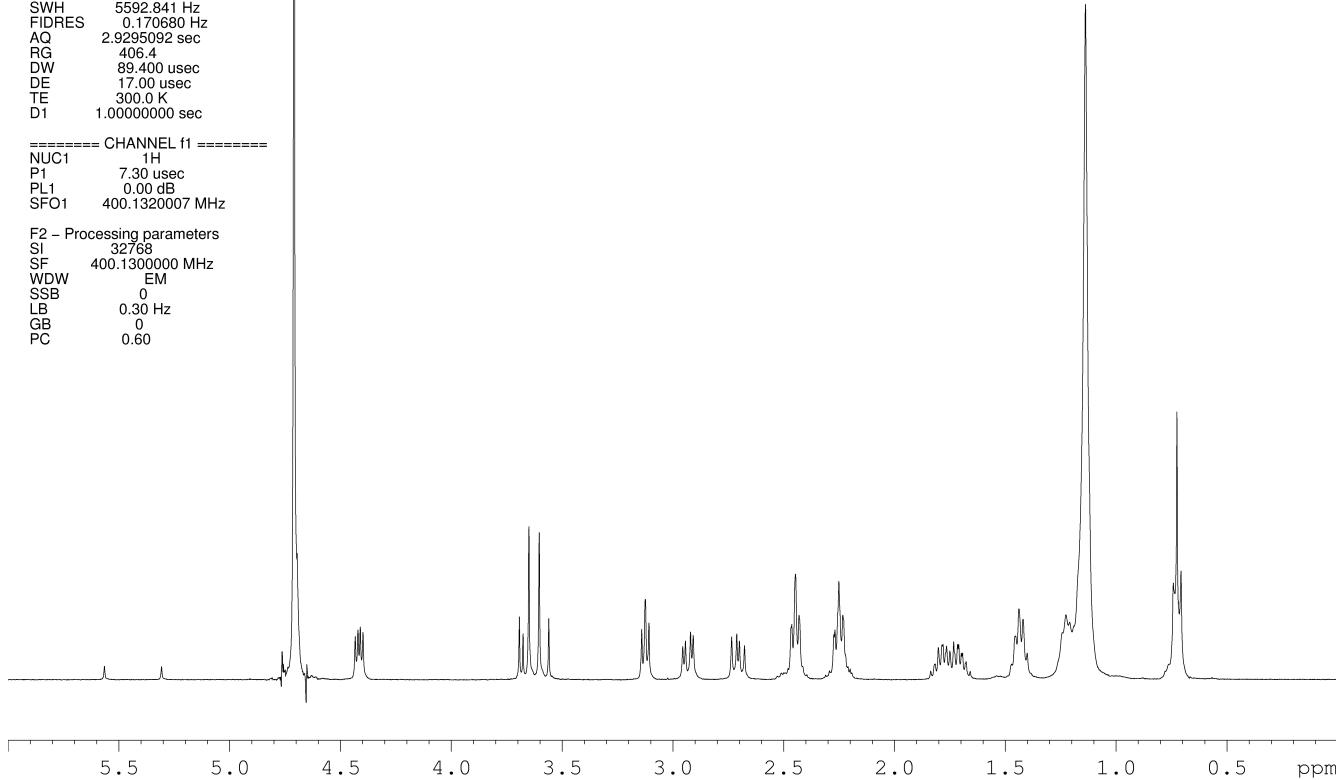
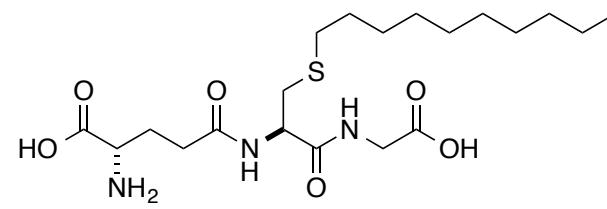
===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.1999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



### S-Decylglutathione (**5c**) – $^1\text{H}$ NMR

Current Data Parameters  
 NAME Dec20-2011-5  
 EXPNO 1  
 PROCN0 1  
 F2 – Acquisition Parameters  
 Date 20111220  
 Time 11.11  
 INSTRUM dpx400  
 PROBHD 5 mm Dual 1H/1H  
 PULPROG zg60  
 TD 32768  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 5592.841 Hz  
 FIDRES 0.170680 Hz  
 AQ 2.9295092 sec  
 RG 406.4  
 DW 89.400 usec  
 DE 17.00 usec  
 TE 300.0 K  
 D1 1.0000000 sec  
 ===== CHANNEL f1 =====  
 NUC1  $^1\text{H}$   
 P1 7.30 usec  
 PL1 0.00 dB  
 SFO1 400.1320007 MHz  
 F2 – Processing parameters  
 SI 32768  
 SF 400.1300000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 0.60



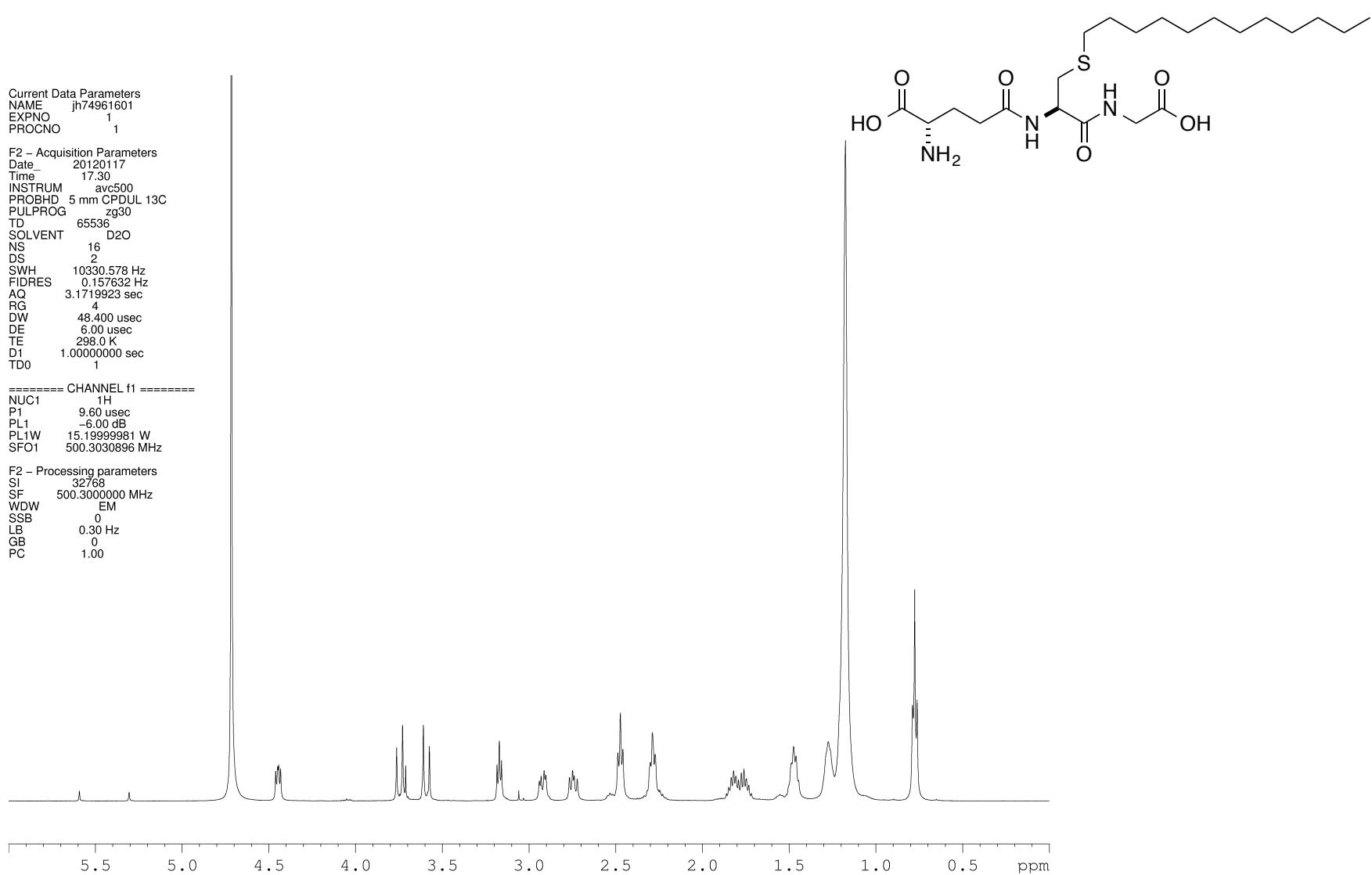
### S-Dodecylglutathione (**5d**) – $^1\text{H}$ NMR

Current Data Parameters  
 NAME jh74961601  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120117  
 Time 17:30  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zg30  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 10330.578 Hz  
 FIDRES 0.157632 Hz  
 AQ 3.1719923 sec  
 RG 4  
 DW 48.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 1.0000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.60 usec  
 PL1 -6.00 dB  
 PL1W 15.1999981 W  
 SFO1 500.3030896 MHz

F2 – Processing parameters  
 SI 32768  
 SF 500.3000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



### S-Dodecylglutathione (**5d**) – $^{13}\text{C}$ NMR

Current Data Parameters  
NAME jh81302102  
EXPNO 4  
PROCNO 1

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F2 - Acquisition Parameters
Date_   20120221
Time    9.41
INSTRUM avc500
PROBHD  5 mm CPDUL 13C
PULPROG zgpg30
TD      65536
SOLVENT D2O
NS      1731
DS      2
SWH    312500.000 Hz
FIDRES 0.476837 Hz
AQ     1.048625 sec
RG      1820
DW     16.000 usec
DE     20.00 usec
TE     298.0 K
D1    2.00000000 sec
D11   0.03000000 sec
TDO     1

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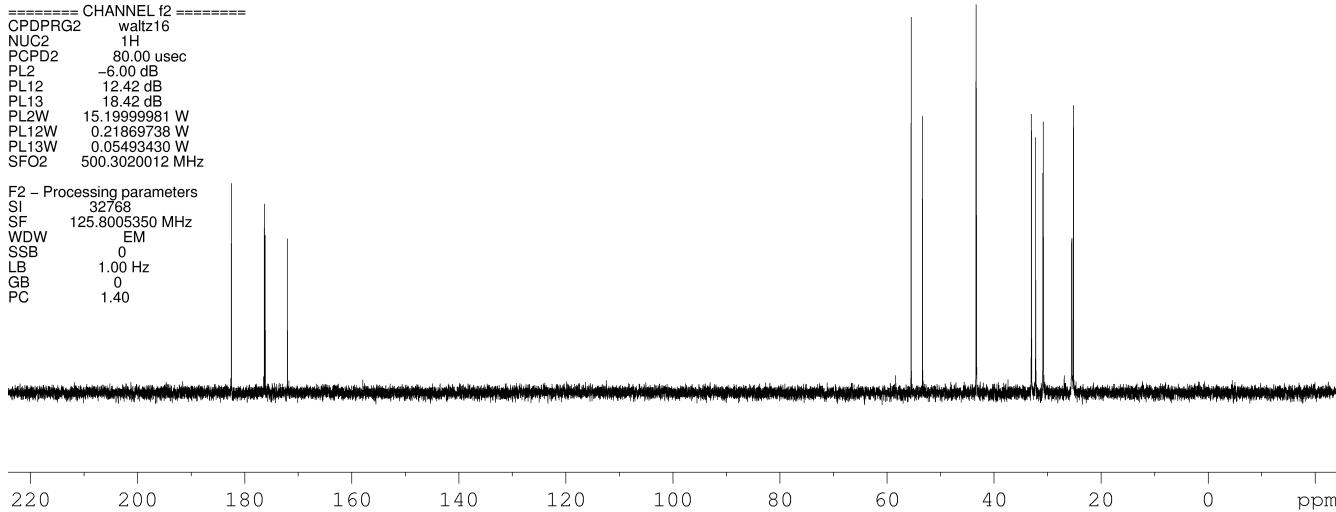
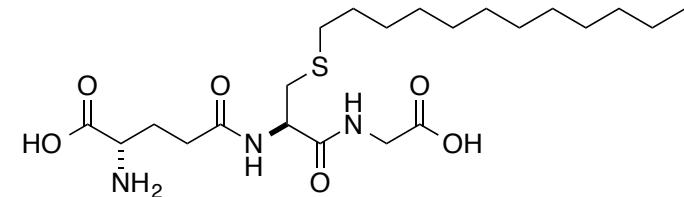
===== CHANNEL f1 =====  
NUC1 13C  
P1 10.00 usec  
PL1 -4.40 dB  
PL1W 28.1575209 W  
SFO1 125.8131151 MHz

```
===== CHANNEL f2 =====
CPDPRG2      walt16
NUC2          1H
PCPD2        80.00 usec
PL2          -6.00 dB
PL12         12.42 dB
PL13         18.42 dB
PL2W         15.19999991 W
PL12W        0.21869738 W
PL13W        0.05493430 W
SFO2         500.3020012 MHz
```

```

F2 - Processing parameters
SI      32768
SF      125.8005350 MHz
WDW     EM
SSB     0
LB      1.00 Hz
GB      0
PC      1.40

```



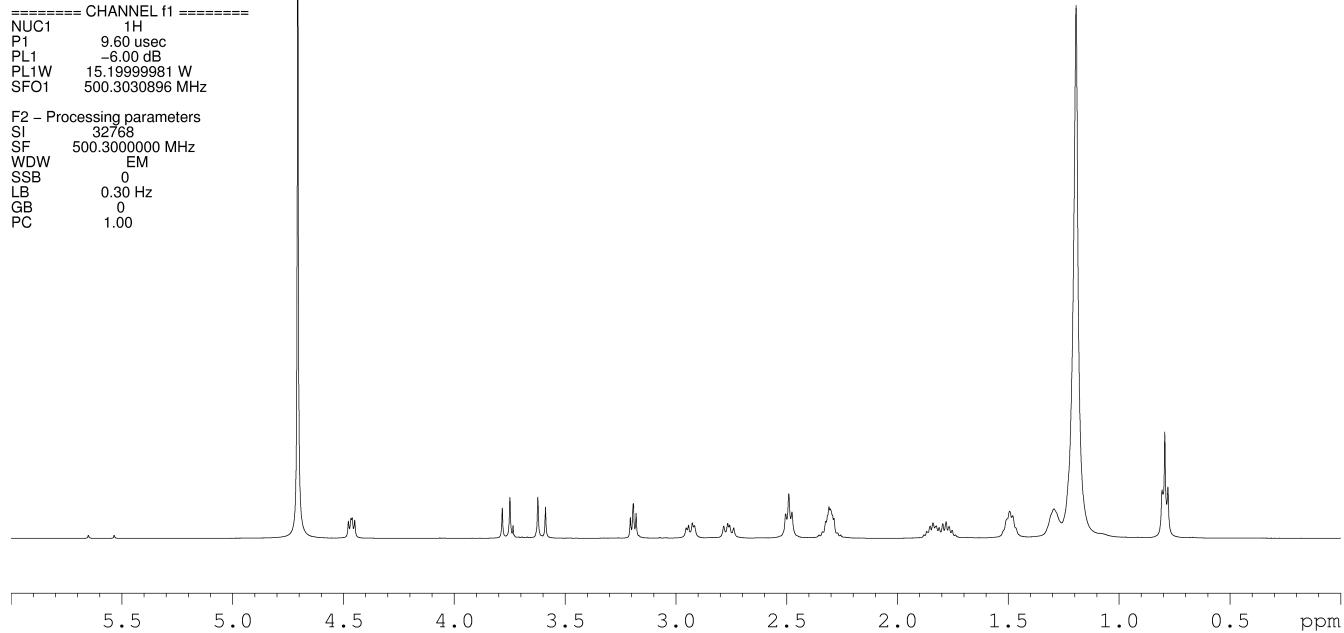
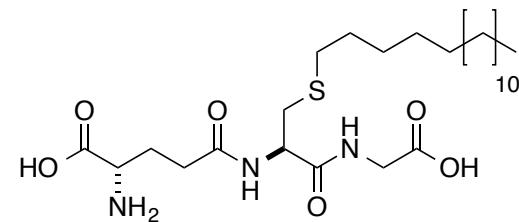
### S-Hexadecylglutathione (**5e**) – $^1\text{H}$ NMR

Current Data Parameters  
 NAME jh74951601  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120117  
 Time 14.57  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zg30  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 10330.578 Hz  
 FIDRES 0.157632 Hz  
 AQ 3.1719923 sec  
 RG 4  
 DW 48.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 1.0000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.60 usec  
 PL1 -6.00 dB  
 PL1W 15.1999981 W  
 SFO1 500.3030896 MHz

F2 – Processing parameters  
 SI 32768  
 SF 500.3000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



### S-Hexadecylglutathione (**5e**) – $^{13}\text{C}$ NMR

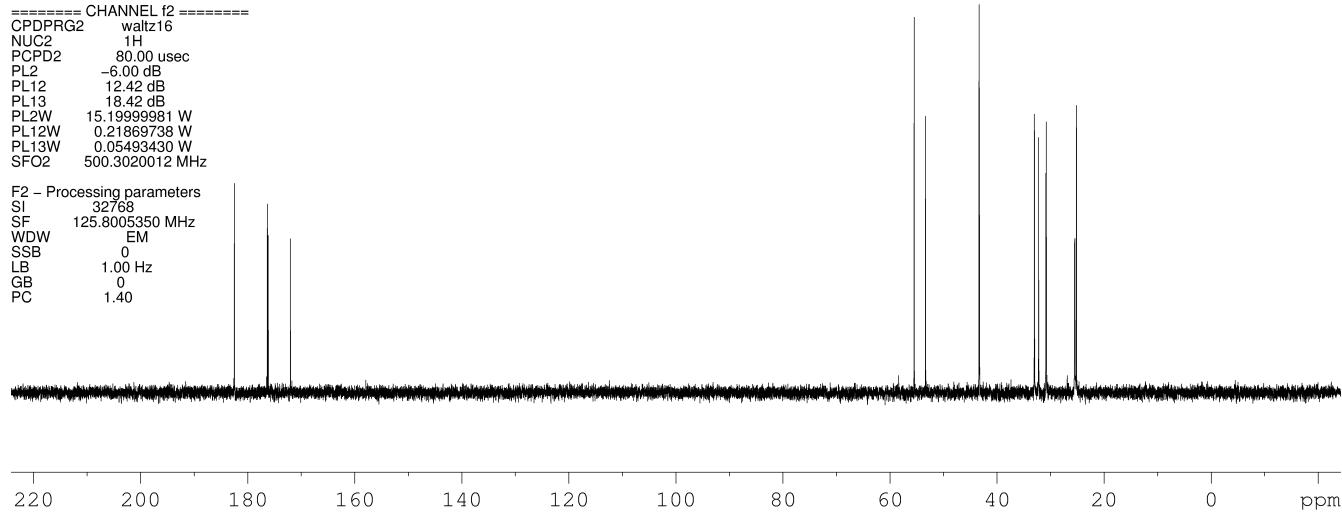
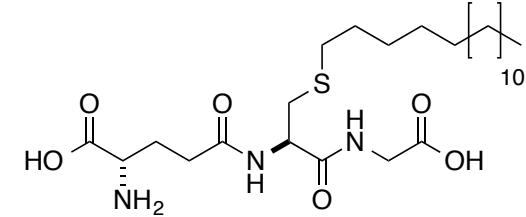
Current Data Parameters  
 NAME jh81302102  
 EXPNO 4  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120221  
 Time 9.41  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1731  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 1820  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.1999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



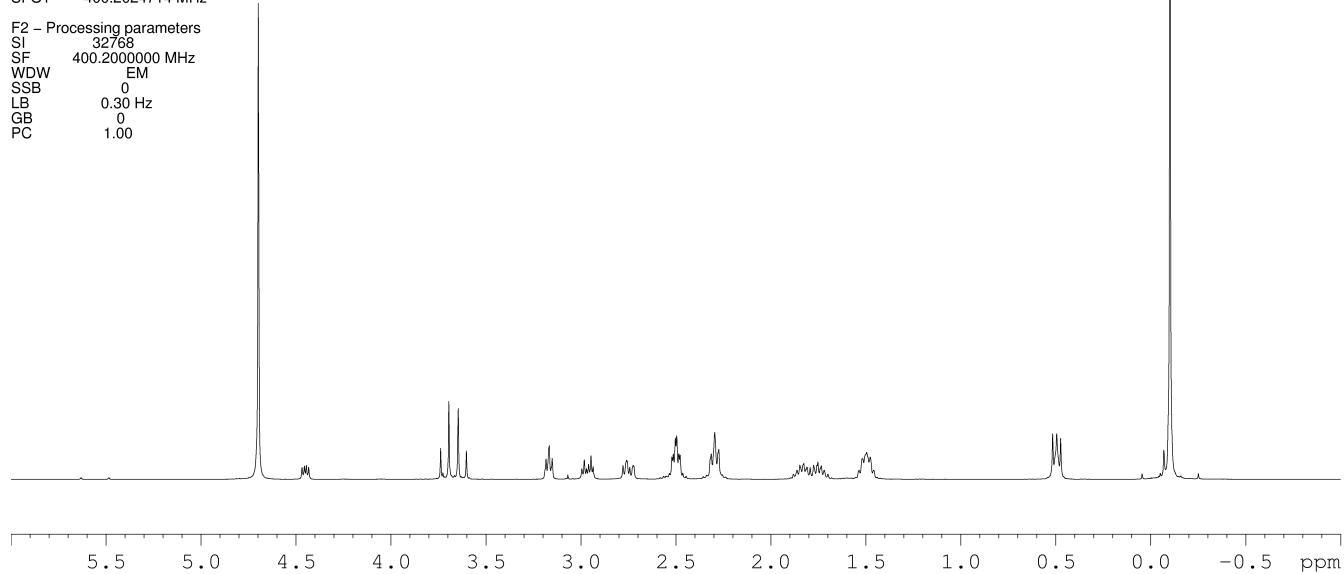
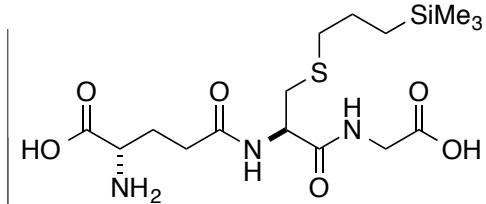
### S-Propyltrimethylsilylglutathione (**5f**) – $^1\text{H}$ NMR

Current Data Parameters  
 NAME Jan11-2012-34  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120111  
 Time 17.21  
 INSTRUM av400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zg60  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 8278.146 Hz  
 FIDRES 0.126314 Hz  
 AQ 3.9584243 sec  
 RG 35.9  
 DW 60.400 usec  
 DE 7.50 usec  
 TE 300.0 K  
 D1 1.0000000 sec

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL1 0.00 dB  
 SFO1 400.2024714 MHz

F2 – Processing parameters  
 SI 32768  
 SF 400.2000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



### S-Propyltrimethylsilylglutathione (**5f**) – $^{13}\text{C}$ NMR

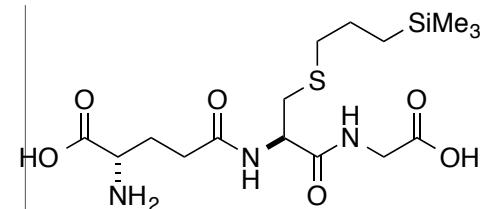
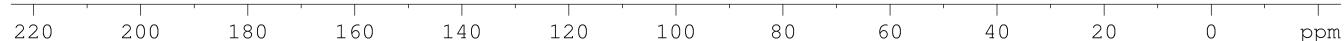
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 NAME Jan11-2012-34  
 EXPNO 3  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120111  
 Time 17:33  
 INSTRUM av400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT D2O  
 NS 256  
 DS 4  
 SWH 26178.010 Hz  
 FIDRES 0.798889 Hz  
 AQ 0.6259188 sec  
 RG 32768  
 DW 19.100 usec  
 DE 7.50 usec  
 TE 300.0 K  
 D1 1.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1  $^{13}\text{C}$   
 P1 9.50 usec  
 PL1 0.00 dB  
 SFO1 100.6403931 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2  $^1\text{H}$   
 PCPD2 80.00 usec  
 PL2 0.00 dB  
 PL12 19.00 dB  
 PL13 25.00 dB  
 SFO2 400.2016008 MHz

F2 – Processing parameters  
 SI 32768  
 SF 100.6303300 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



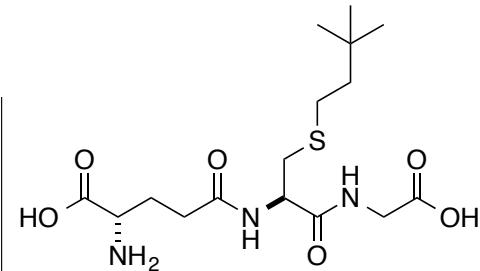
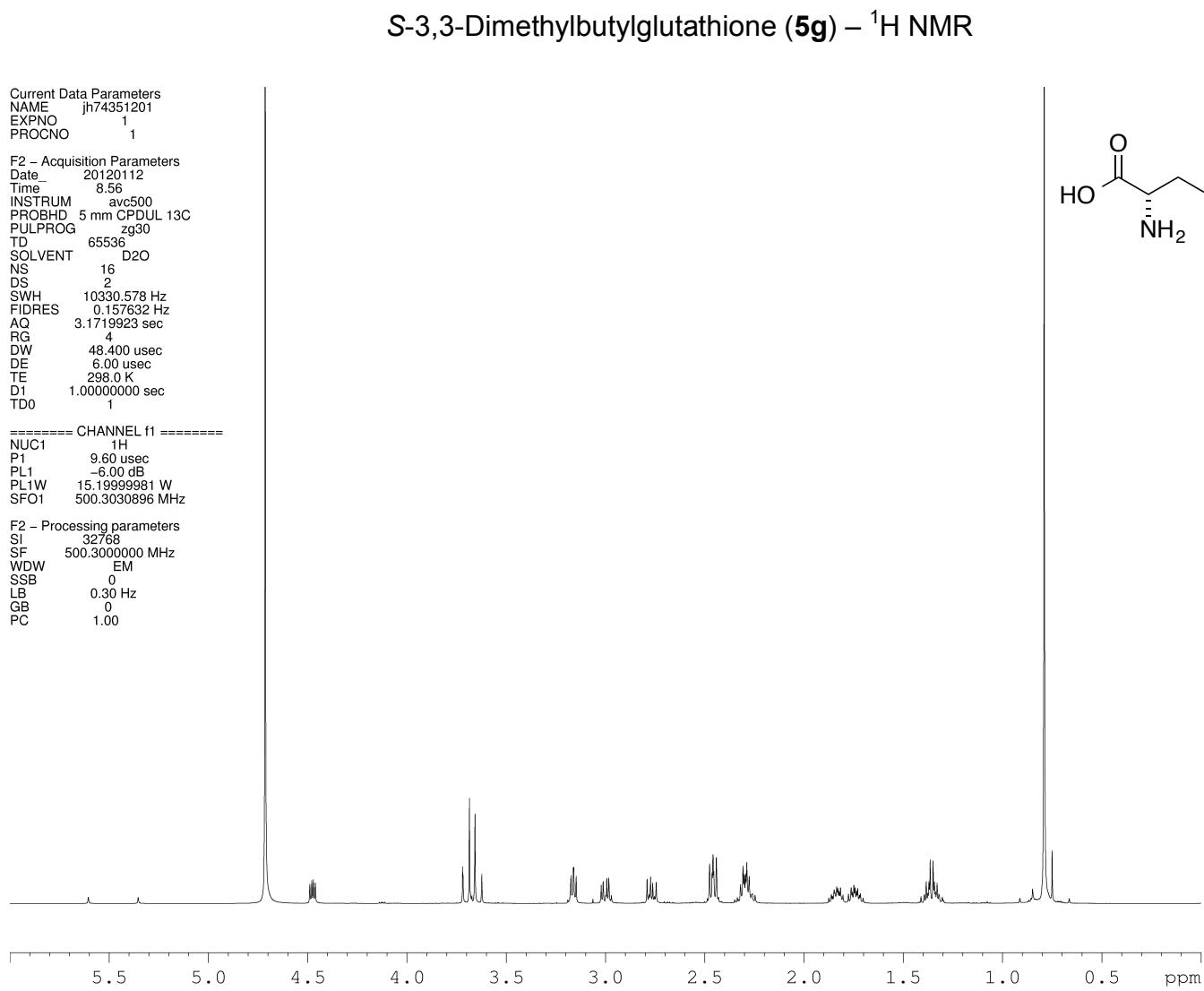
S-3,3-Dimethylbutylglutathione (**5g**) –  $^1\text{H}$  NMR

Current Data Parameters  
 NAME jh74351201  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120112  
 Time 8.56  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zg30  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 10330.578 Hz  
 FIDRES 0.157632 Hz  
 AQ 3.1719923 sec  
 RG 4  
 DW 48.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 1.0000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.60 usec  
 PL1 -6.00 dB  
 PL1W 15.1999981 W  
 SFO1 500.3030896 MHz

F2 – Processing parameters  
 SI 32768  
 SF 500.3000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



### S-3,3-Dimethylbutylglutathione (**5g**) – $^{13}\text{C}$ NMR

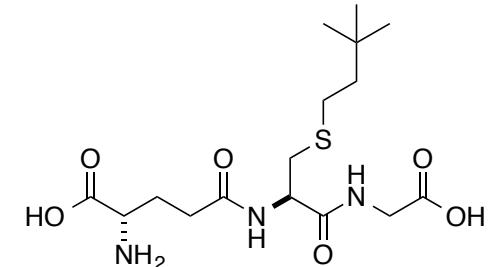
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 EXPNO 4  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120112  
 Time 12.10  
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 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1024  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 812  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPDP2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.19999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



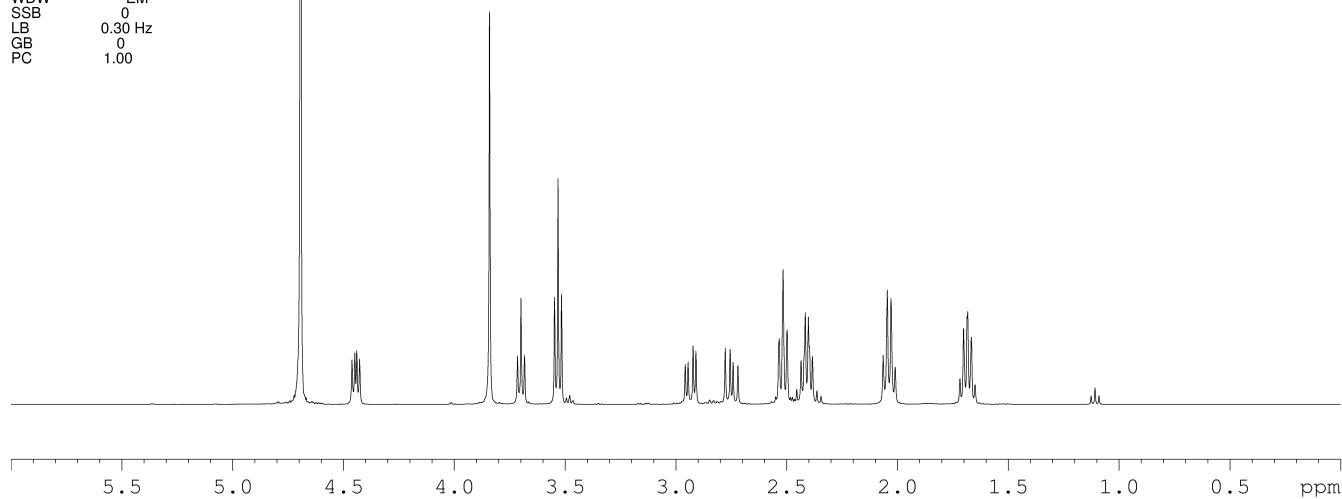
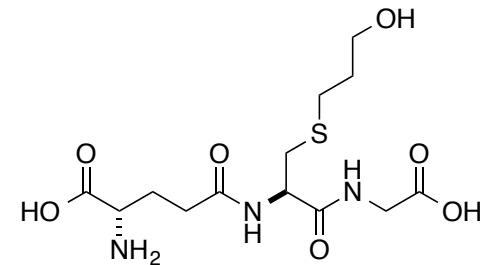
### S-Propanol glutathione (**5h**) – $^1\text{H}$ HMR

Current Data Parameters  
 NAME Dec20-2011-12  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 2011220  
 Time 10.43  
 INSTRUM av400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zg60  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 8278.146 Hz  
 FIDRES 0.126314 Hz  
 AQ 3.9584243 sec  
 RG 90.5  
 DW 60.400 usec  
 DE 7.50 usec  
 TE 300.0 K  
 D1 1.0000000 sec

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL1 0.00 dB  
 SFO1 400.2024714 MHz

F2 – Processing parameters  
 SI 32768  
 SF 400.2000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



### S-Propanol glutathione (**5h**) – $^{13}\text{C}$ HMR

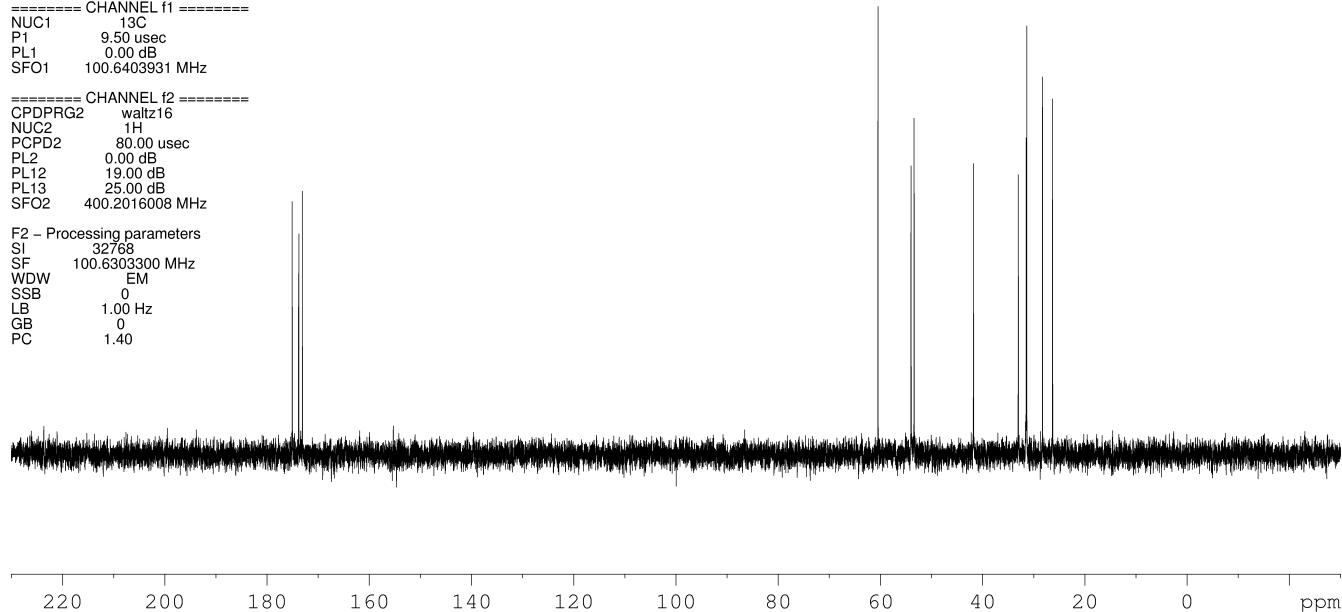
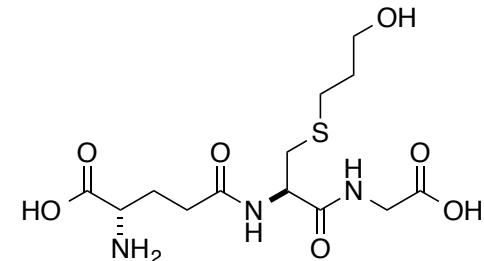
Current Data Parameters  
 NAME Dec20-2011-12  
 EXPNO 3  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20111220  
 Time 10.55  
 INSTRUM av400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT D2O  
 NS 256  
 DS 4  
 SWH 26178.010 Hz  
 FIDRES 0.798889 Hz  
 AQ 0.6259188 sec  
 RG 32768  
 DW 19.100 usec  
 DE 7.50 usec  
 TE 300.0 K  
 D1 1.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1  $^{13}\text{C}$   
 P1 9.50 usec  
 PL1 0.00 dB  
 SFO1 100.6403931 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2  $^1\text{H}$   
 PCPD2 80.00 usec  
 PL2 0.00 dB  
 PL12 19.00 dB  
 PL13 25.00 dB  
 SFO2 400.2016008 MHz

F2 – Processing parameters  
 SI 32768  
 SF 100.6303300 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



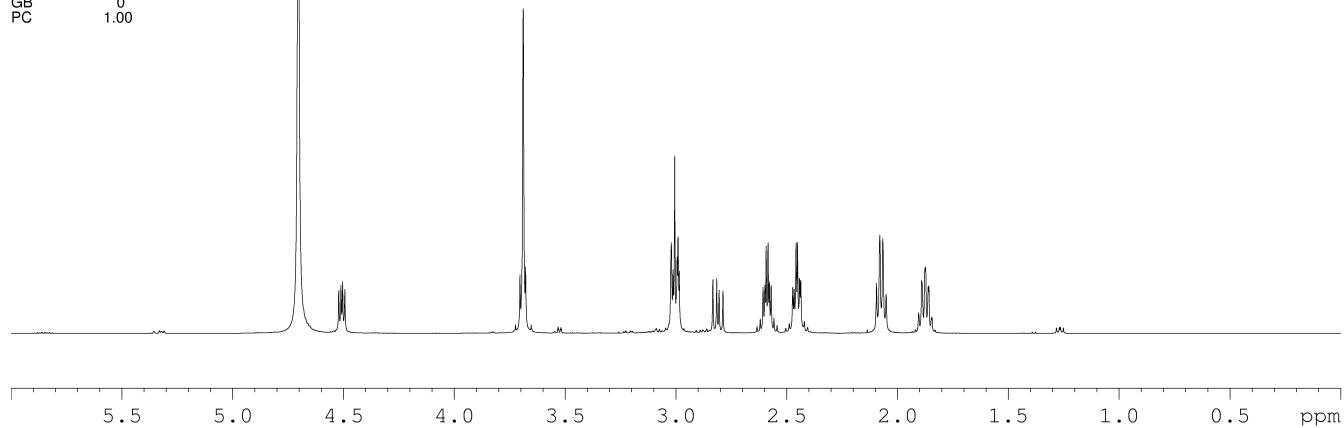
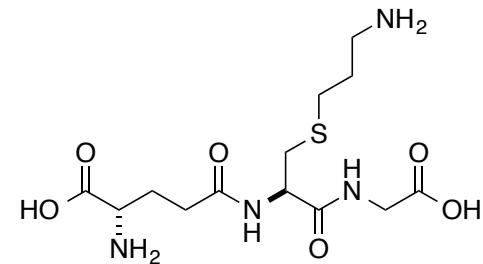
### S-Propylamino glutathione (**5i**) – $^1\text{H}$ NMR

Current Data Parameters  
 NAME jh80411502  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120216  
 Time 10.31  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zg30  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 10330.578 Hz  
 FIDRES 0.157632 Hz  
 AQ 3.1719923 sec  
 RG 4  
 DW 48.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 1.0000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.60 usec  
 PL1 -6.00 dB  
 PL1W 15.1999981 W  
 SFO1 500.3030896 MHz

F2 – Processing parameters  
 SI 32768  
 SF 500.3000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



### S-Propylamino glutathione (**5i**) – $^{13}\text{C}$ NMR

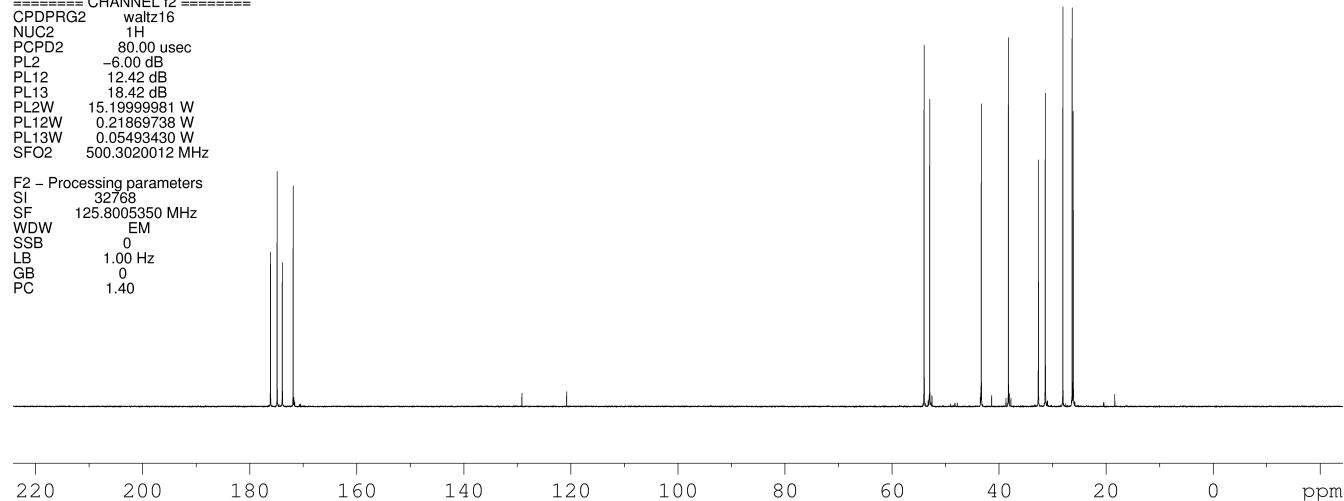
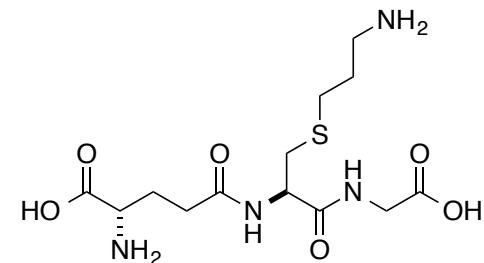
Current Data Parameters  
 NAME jh80411502  
 EXPNO 4  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120216  
 Time 11.46  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1024  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 1440  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1  $^{13}\text{C}$   
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2  $^1\text{H}$   
 PCPD2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.19999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



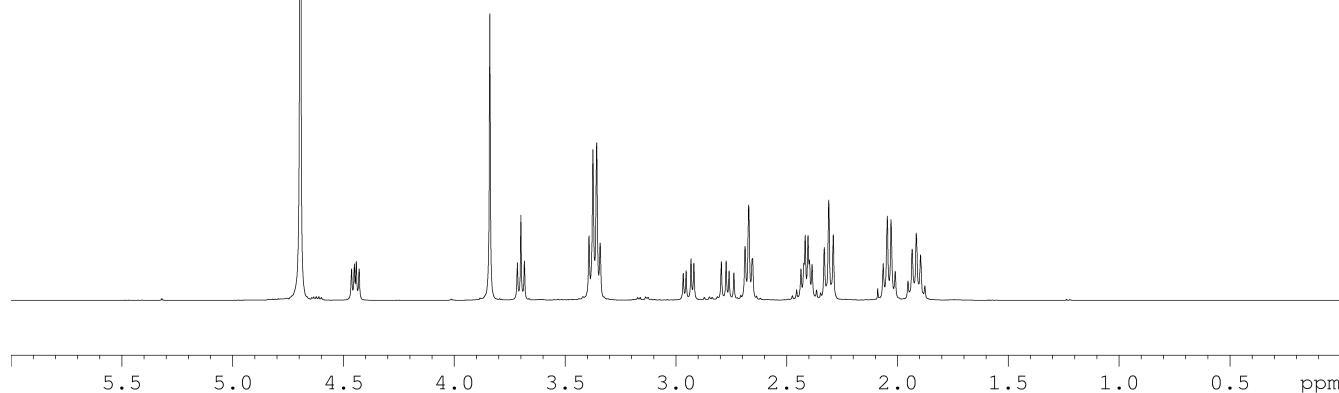
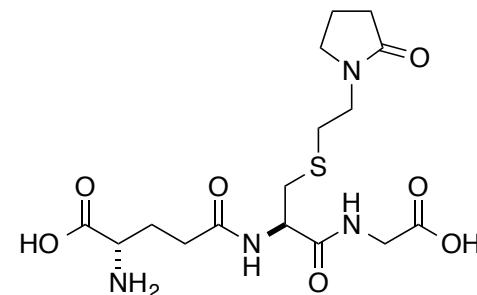
S-1-Ethyl-N-pyrrolidinone glutathione (**5j**) –  $^1\text{H}$  NMR

Current Data Parameters  
 NAME Jan23-2012-15  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120123  
 Time 14.14  
 INSTRUM av400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zg60  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 8278.146 Hz  
 FIDRES 0.126314 Hz  
 AQ 3.9584243 sec  
 RG 35.9  
 DW 60.400 usec  
 DE 7.50 usec  
 TE 300.0 K  
 D1 1.0000000 sec

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL1 0.00 dB  
 SFO1 400.2024714 MHz

F2 – Processing parameters  
 SI 32768  
 SF 400.2000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



## S-1-Ethyl-N-pyrrolidinone glutathione (**5j**) – $^{13}\text{C}$ MR

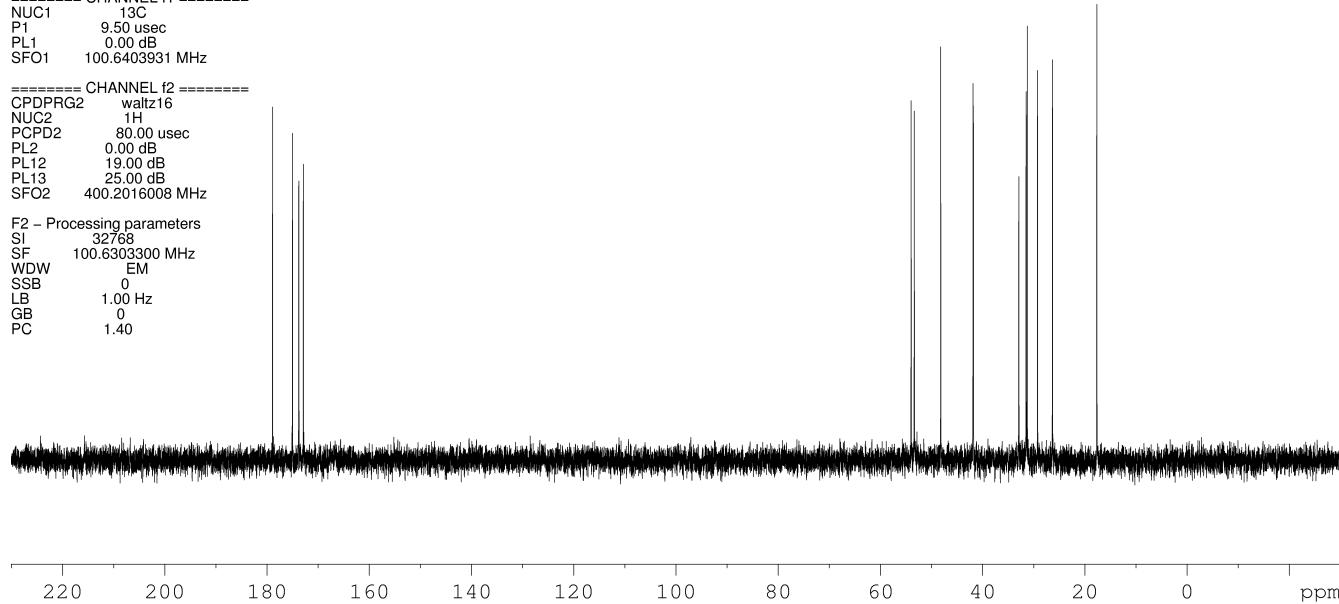
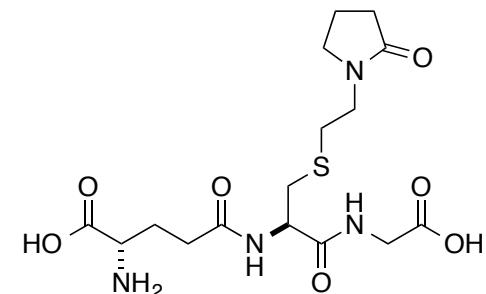
Current Data Parameters  
 NAME Jan23-2012-15  
 EXPNO 3  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120123  
 Time 14.26  
 INSTRUM av400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT D2O  
 NS 256  
 DS 4  
 SWH 26178.010 Hz  
 FIDRES 0.798889 Hz  
 AQ 0.6259188 sec  
 RG 32768  
 DW 19.100 usec  
 DE 7.50 usec  
 TE 300.0 K  
 D1 1.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 9.50 usec  
 PL1 0.00 dB  
 SFO1 100.6403931 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 0.00 dB  
 PL12 19.00 dB  
 PL13 25.00 dB  
 SFO2 400.2016008 MHz

F2 – Processing parameters  
 SI 32768  
 SF 100.6303300 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



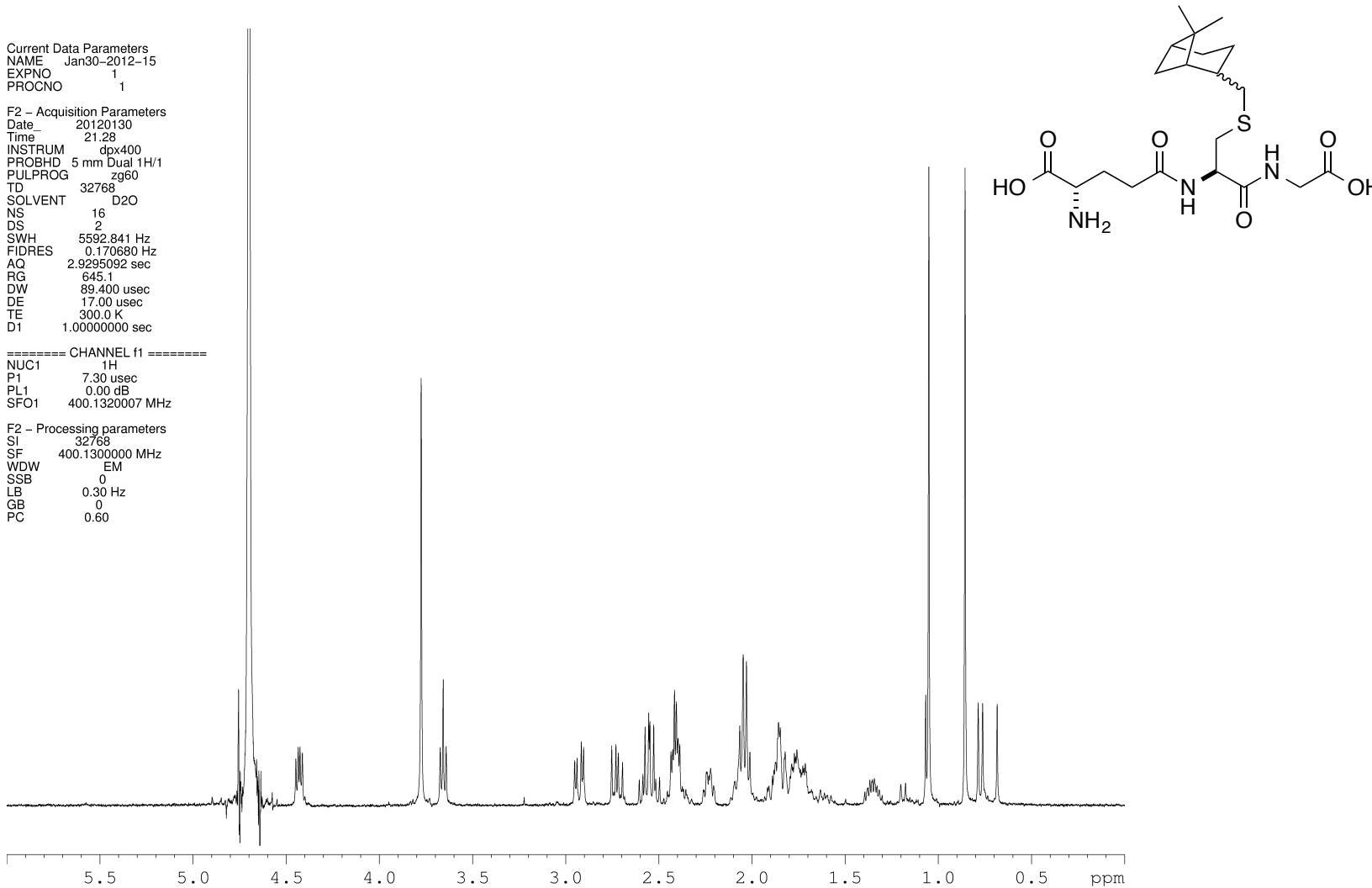
S-(6,6-dimethylbicyclo[3.1.1]heptan-2-yl)methyl glutathione (**5k**) –  
<sup>1</sup>H NMR

Current Data Parameters  
NAME Jan30–2012–15  
EXPNO 1  
PROCNO 1

F2 – Acquisition Parameters  
Date 20120130  
Time 21.28  
INSTRUM dpx400  
PROBHD 5 mm Dual 1H/1  
PULPROG zg60  
TD 32768  
SOLVENT D2O  
NS 16  
DS 2  
SWH 5592.841 Hz  
FIDRES 0.170680 Hz  
AQ 2.9295092 sec  
RG 645.1  
DW 89.400 usec  
DE 17.00 usec  
TE 300.0 K  
D1 1.0000000 sec

===== CHANNEL f1 =====  
NUC1 1H  
P1 7.30 usec  
PL1 0.00 dB  
SFO1 400.1320007 MHz

F2 – Processing parameters  
SI 32768  
SF 400.1300000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 0.60



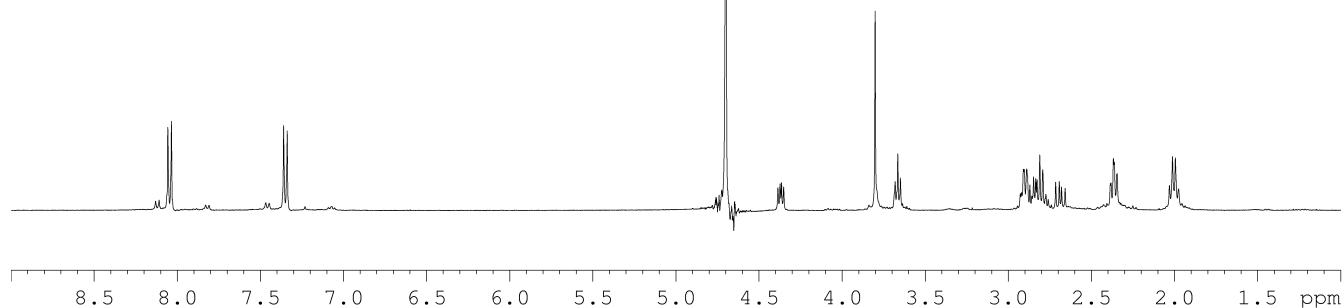
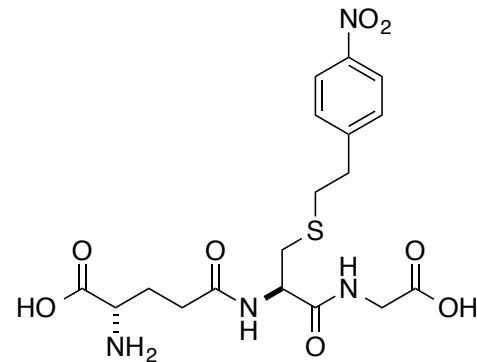
S-Ethyl-4-nitrophenyl glutathione (**6a**) –  $^1\text{H}$  NMR

Current Data Parameters  
 NAME Feb16-2012-59  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120216  
 Time 17.18  
 INSTRUM dpx400  
 PROBHD 5 mm Dual 1H/1H  
 PULPROG zg60  
 TD 32768  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 5592.841 Hz  
 FIDRES 0.170680 Hz  
 AQ 2.9295092 sec  
 RG 456.1  
 DW 89.400 usec  
 DE 17.00 usec  
 TE 300.0 K  
 D1 1.0000000 sec

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 7.30 usec  
 PL1 0.00 dB  
 SFO1 400.1320007 MHz

F2 – Processing parameters  
 SI 32768  
 SF 400.1300000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 0.60



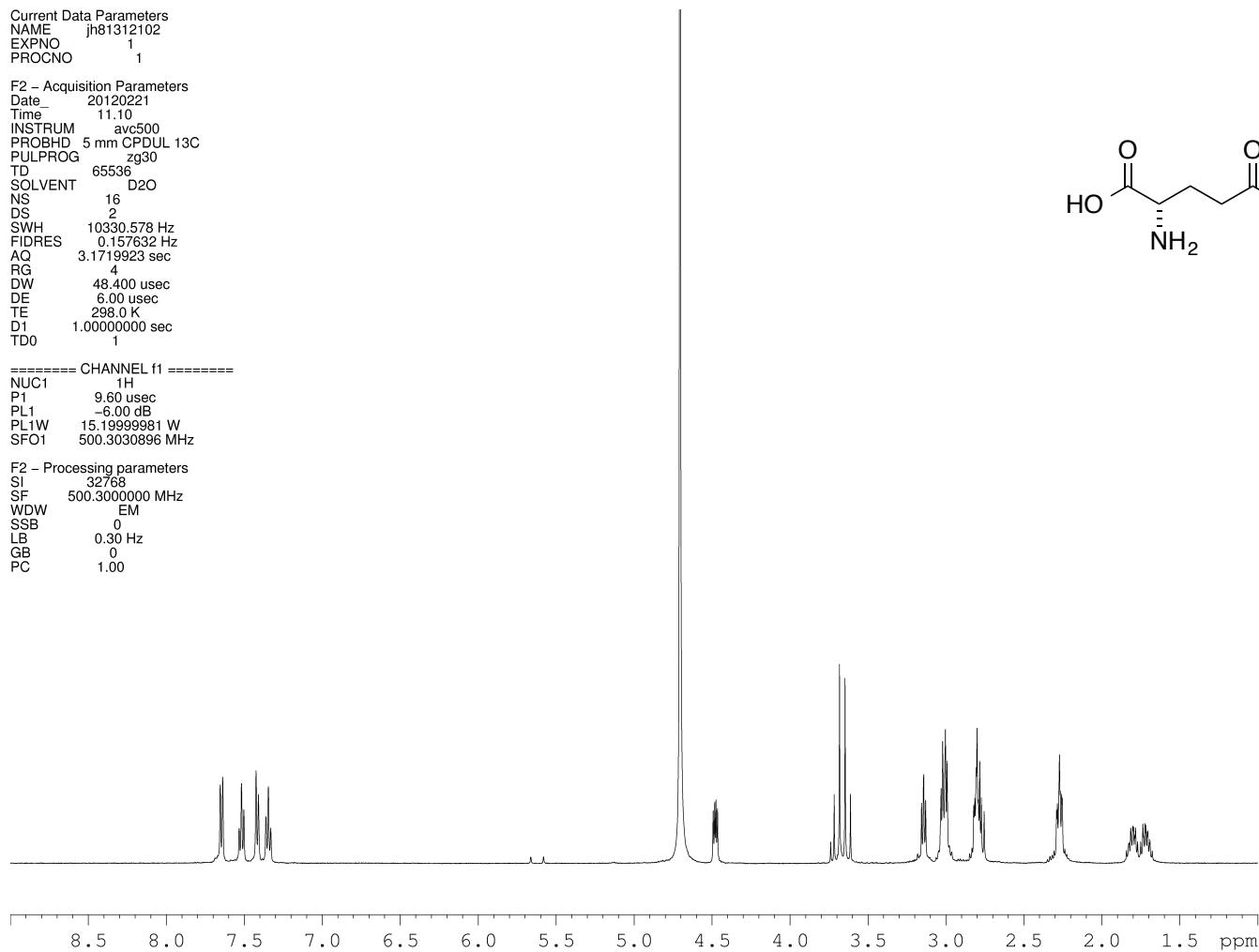
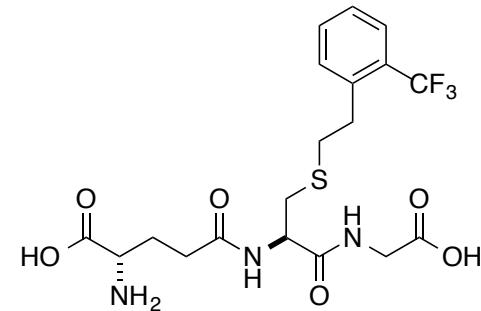
S-Ethyl-2-trifluoromethylphenyl glutathione (**6b**) –  $^1\text{H}$  NMR

Current Data Parameters  
 NAME jh81312102  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120221  
 Time 11.10  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zg30  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 10330.578 Hz  
 FIDRES 0.157632 Hz  
 AQ 3.1719923 sec  
 RG 4  
 DW 48.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 1.0000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.60 usec  
 PL1 -6.00 dB  
 PL1W 15.1999981 W  
 SFO1 500.3030896 MHz

F2 – Processing parameters  
 SI 32768  
 SF 500.3000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



S-Ethyl-2-trifluoromethylphenyl glutathione (**6b**) –  $^{13}\text{C}$  NMR

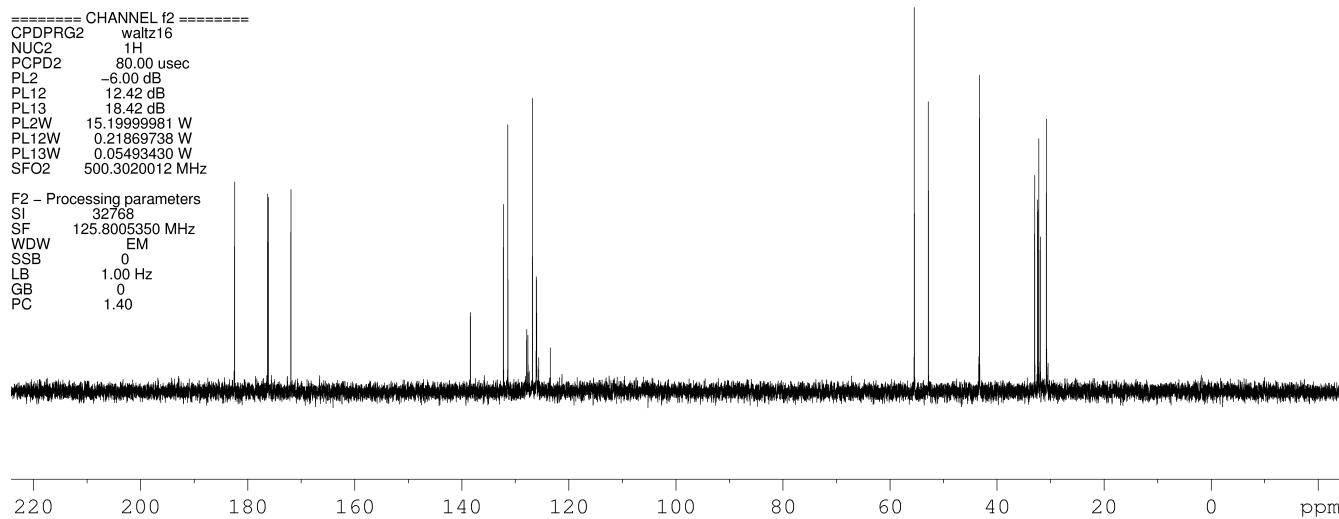
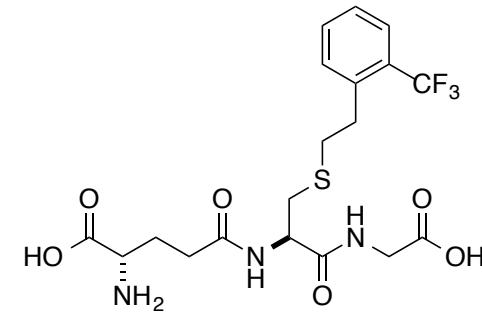
Current Data Parameters  
 NAME jh81312102  
 EXPNO 4  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120221  
 Time 12.06  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 2048  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 1820  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPDP2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.19999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



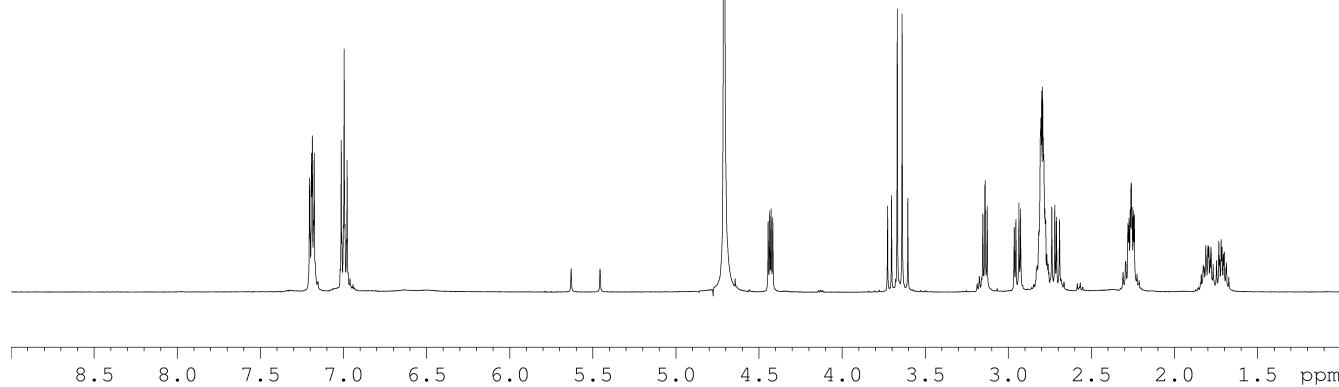
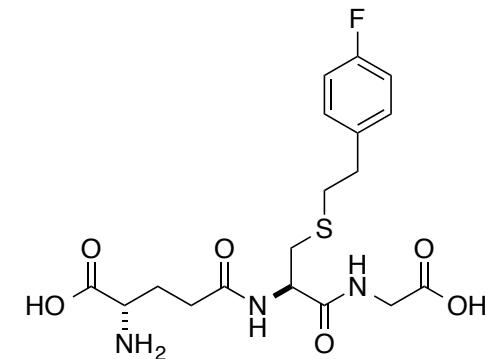
S-Ethyl-4-fluorophenyl glutathione (**6c**) –  $^1\text{H}$  NMR

Current Data Parameters  
 NAME jh77650202  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120202  
 Time 10.59  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zg30  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 10330.578 Hz  
 FIDRES 0.157632 Hz  
 AQ 3.1719923 sec  
 RG 4  
 DW 48.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 1.0000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.60 usec  
 PL1 -6.00 dB  
 PL1W 15.1999981 W  
 SFO1 500.3030896 MHz

F2 – Processing parameters  
 SI 32768  
 SF 500.3000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



### S-Ethyl-4-fluorophenyl glutathione (**6c**) – $^{13}\text{C}$ NMR

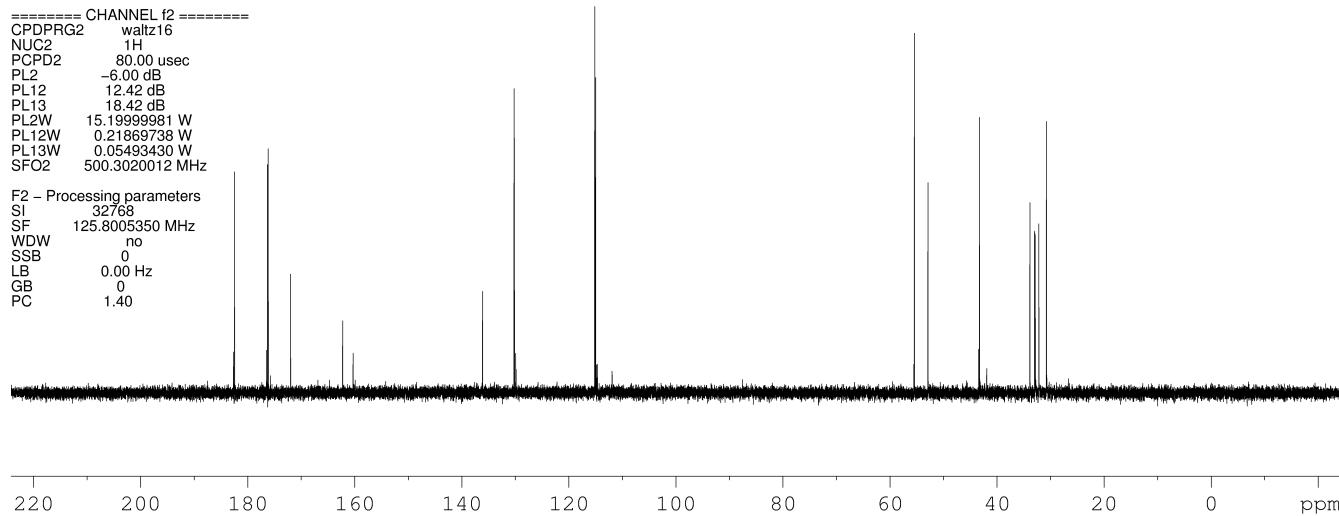
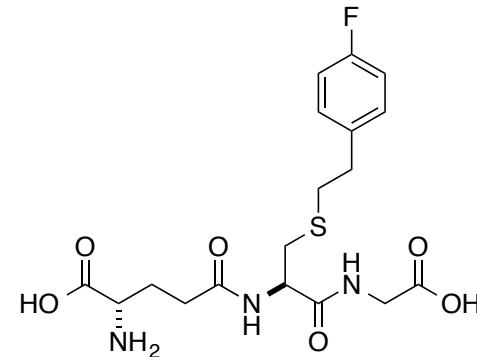
Current Data Parameters  
 NAME jh77650202  
 EXPNO 3  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120202  
 Time 11.52  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 2048  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 1440  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.19999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.40



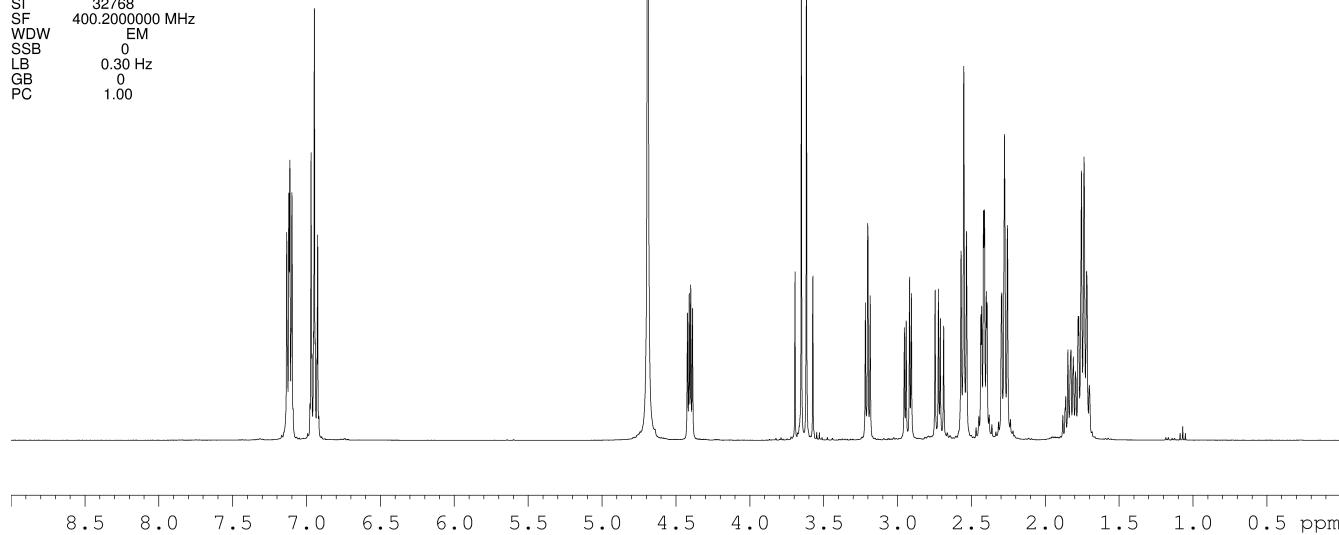
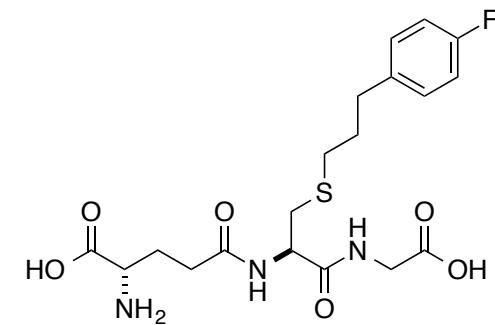
S-Propyl-4-fluorophenyl glutathione (**6d**) –  $^1\text{H}$  NMR

Current Data Parameters  
 NAME Jan30-2012-35  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120131  
 Time 2.17  
 INSTRUM av400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zg60  
 TD 65536  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 8278.146 Hz  
 FIDRES 0.126314 Hz  
 AQ 3.9584243 sec  
 RG 71.8  
 DW 60.400 usec  
 DE 7.50 usec  
 TE 300.0 K  
 D1 1.0000000 sec

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.00 usec  
 PL1 0.00 dB  
 SFO1 400.2024714 MHz

F2 – Processing parameters  
 SI 32768  
 SF 400.2000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



### S-Propyl-4-fluorophenyl glutathione (**6d**) – $^{13}\text{C}$ NMR

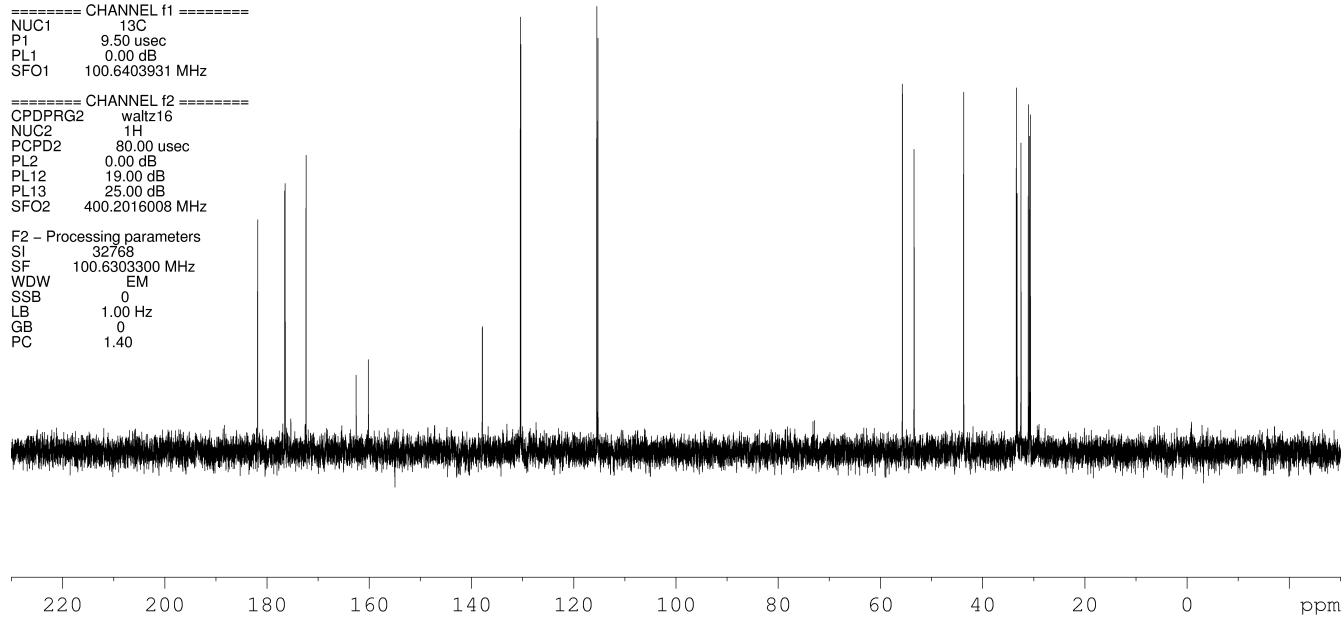
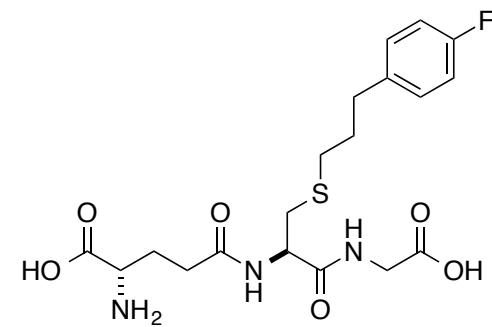
Current Data Parameters  
 NAME Jan30-2012-35  
 EXPNO 4  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120131  
 Time 2.30  
 INSTRUM av400  
 PROBHD 5 mm QNP 1H/13  
 PULPROG zgpg30  
 TD 32768  
 SOLVENT D2O  
 NS 256  
 DS 4  
 SWH 26178.010 Hz  
 FIDRES 0.798889 Hz  
 AQ 0.6259188 sec  
 RG 32768  
 DW 19.100 usec  
 DE 7.50 usec  
 TE 300.0 K  
 D1 1.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1  $^{13}\text{C}$   
 P1 9.50 usec  
 PL1 0.00 dB  
 SFO1 100.6403931 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2  $^1\text{H}$   
 PCPD2 80.00 usec  
 PL2 0.00 dB  
 PL12 19.00 dB  
 PL13 25.00 dB  
 SFO2 400.2016008 MHz

F2 – Processing parameters  
 SI 32768  
 SF 100.6303300 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



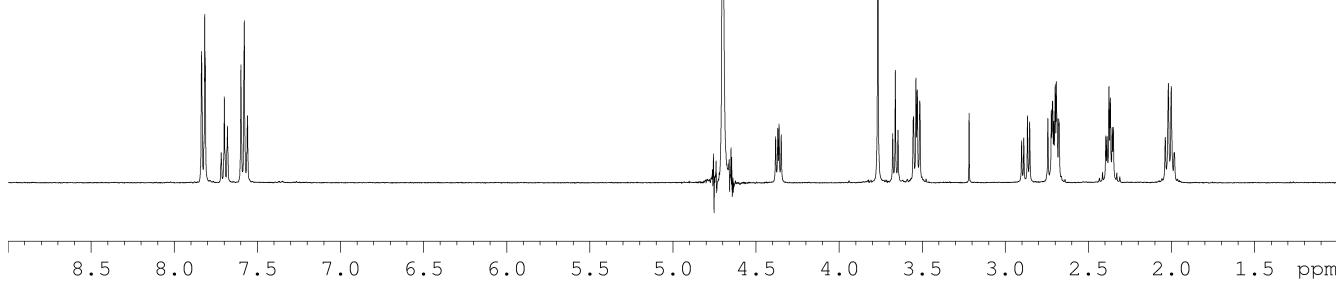
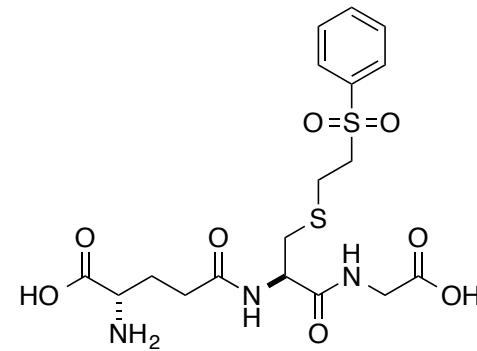
S-Phenylethylsulfone glutathione (**6e**) –  $^1\text{H}$  NMR

Current Data Parameters  
 NAME Jan31-2012-51  
 EXPNO 1  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120131  
 Time 11.14  
 INSTRUM dpx400  
 PROBHD 5 mm Dual 1H/1  
 PULPROG zg60  
 TD 32768  
 SOLVENT D2O  
 NS 16  
 DS 2  
 SWH 5592.841 Hz  
 FIDRES 0.170680 Hz  
 AQ 2.9295092 sec  
 RG 574.7  
 DW 89.400 usec  
 DE 17.00 usec  
 TE 300.0 K  
 D1 1.0000000 sec

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 7.30 usec  
 PL1 0.00 dB  
 SFO1 400.1320007 MHz

F2 – Processing parameters  
 SI 32768  
 SF 400.1300000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 0.60



S-Phenylethylsulfone glutathione (**6e**) –  $^1\text{H}$  NMR

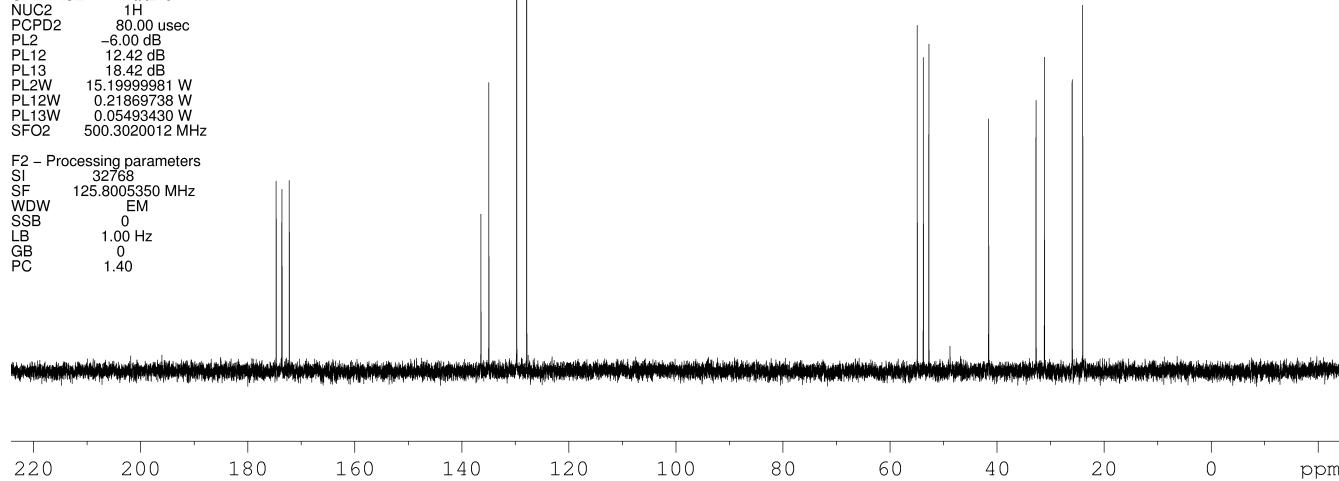
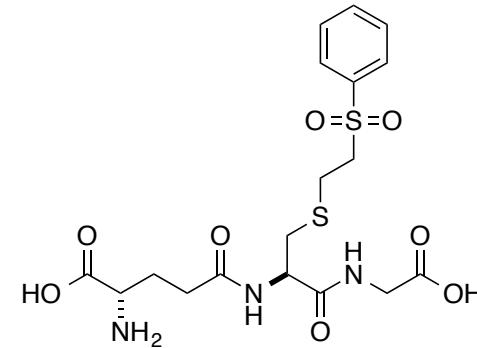
Current Data Parameters  
 NAME jh77863101  
 EXPNO 4  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120202  
 Time 14.55  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1024  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 1820  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.19999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



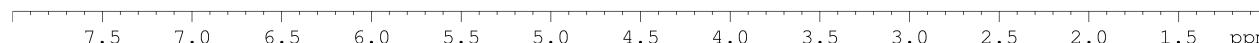
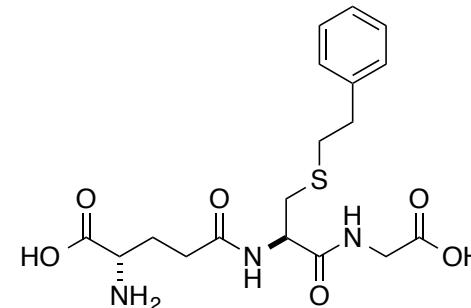
### S-Phenylethyl glutathione (**6f**) – $^1\text{H}$ NMR

Current Data Parameters  
NAME jh77640102  
EXPNO 1  
PROCNO 1

F2 – Acquisition Parameters  
Date 20120201  
Time 9.40  
INSTRUM avc500  
PROBHD 5 mm CPDUL 13C  
PULPROG zg30  
TD 65536  
SOLVENT D2O  
NS 16  
DS 2  
SWH 10330.578 Hz  
FIDRES 0.157632 Hz  
AQ 3.1719923 sec  
RG 4  
DW 48.400 usec  
DE 6.00 usec  
TE 298.0 K  
D1 1.0000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1  $^1\text{H}$   
P1 9.60 usec  
PL1 -6.00 dB  
PL1W 15.1999981 W  
SFO1 500.3030896 MHz

F2 – Processing parameters  
SI 32768  
SF 500.3000000 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



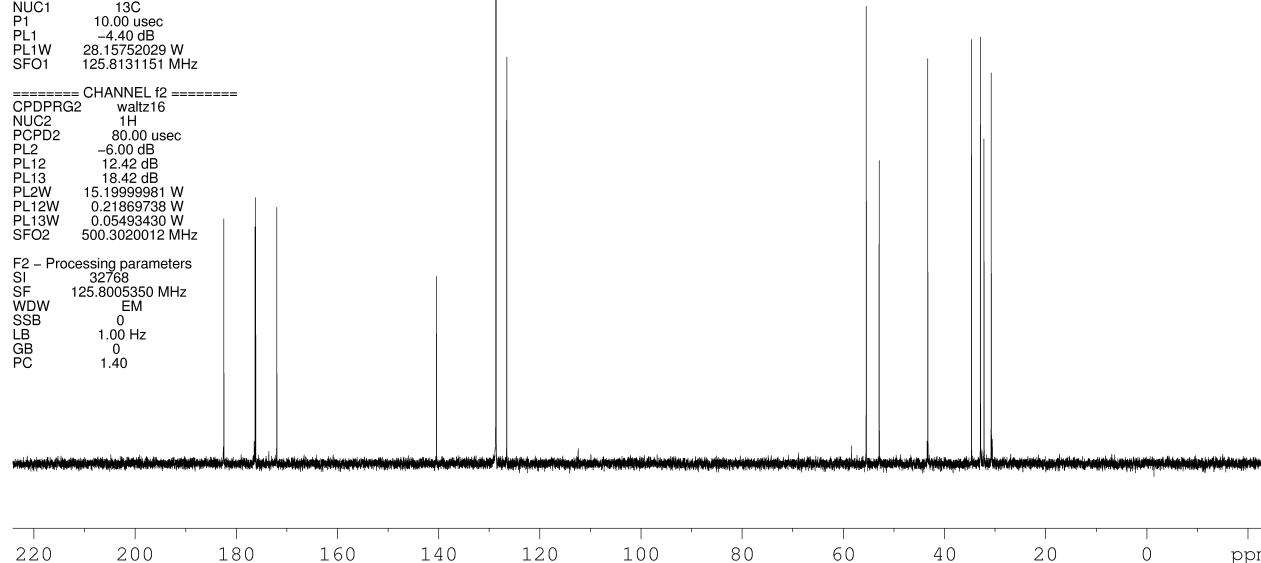
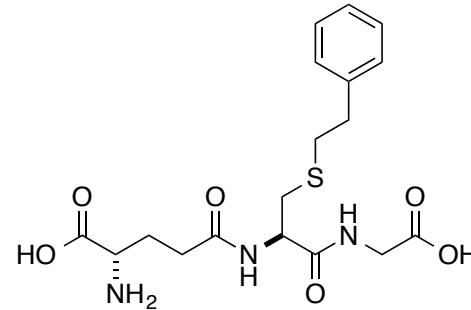
### S-Phenylethyl glutathione (**6f**) – $^{13}\text{C}$ NMR

Current Data Parameters  
 NAME jh77640102  
 EXPNO 3  
 PROCNO 1  
 F2 – Acquisition Parameters  
 Date 20120201  
 Time 10:23  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1168  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 1440  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

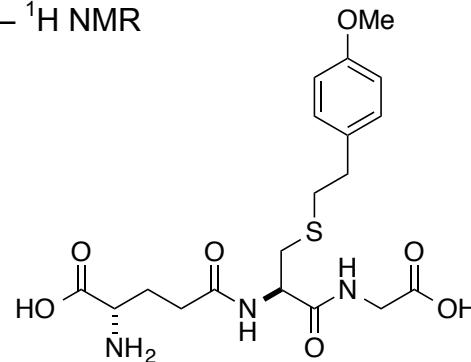
===== CHANNEL f1 =====  
 NUC1  $^{13}\text{C}$   
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2  $^1\text{H}$   
 PCPD2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.1999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

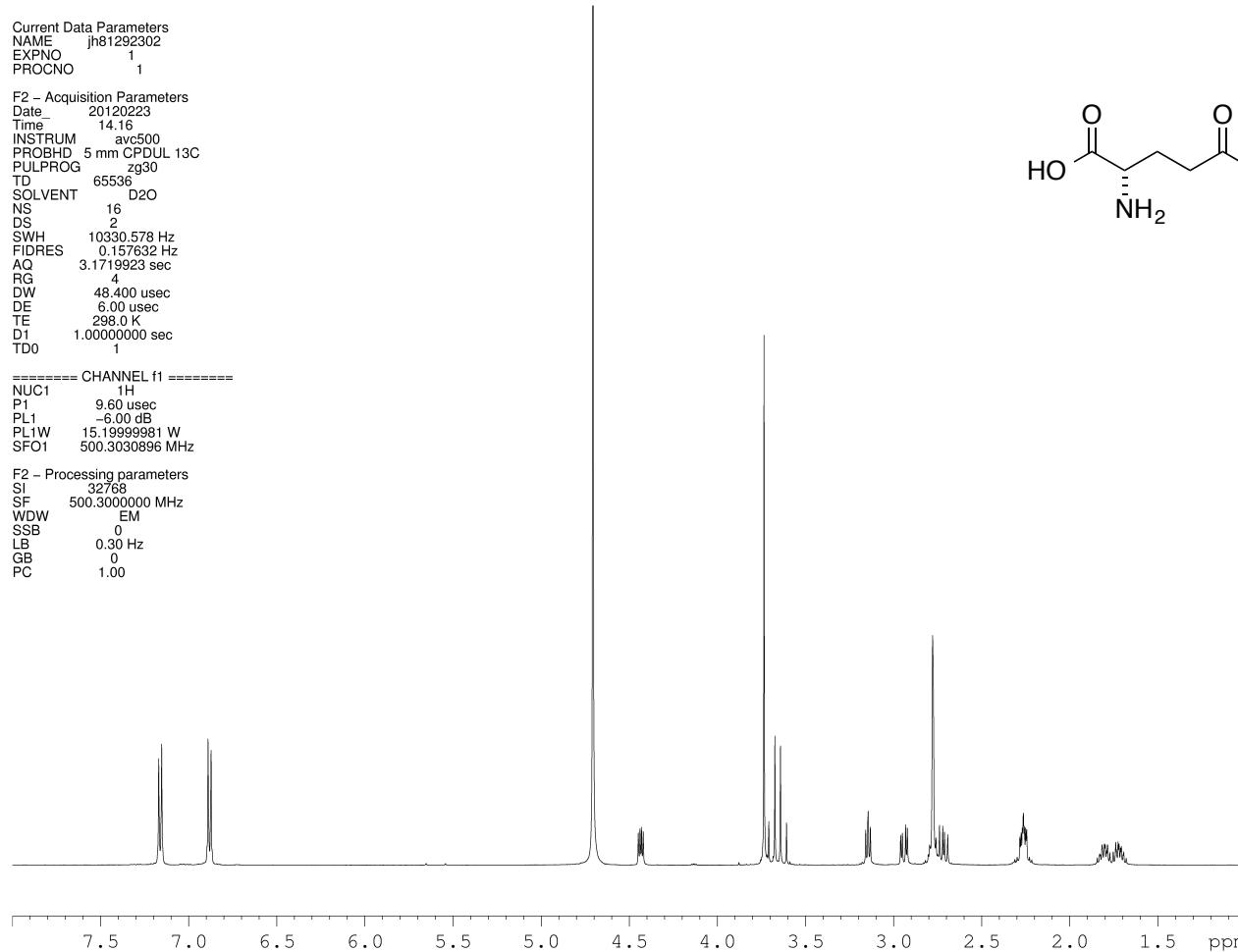
F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40



S-Ethyl-4-methoxyphenyl glutathione (**6g**) –  $^1\text{H}$  NMR



Current Data Parameters  
 NAME jh81292302  
 EXPNO 1  
 PROCNO 1  
 F2 – Acquisition Parameters  
 Date 20120223  
 Time 14.16  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zg30  
 TD 65536  
 SOLVENT D<sub>2</sub>O  
 NS 16  
 DS 2  
 SWH 10330.578 Hz  
 FIDRES 0.157632 Hz  
 AQ 3.1719923 sec  
 RG 4  
 DW 48.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 1.0000000 sec  
 TD0 1  
 ===== CHANNEL f1 ======  
 NUC1 <sup>1</sup>H  
 P1 9.60 usec  
 PL1 -6.00 dB  
 PL1W 15.1999981 W  
 SFO1 500.3030896 MHz  
 F2 – Processing parameters  
 SI 32768  
 SF 500.3000000 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00



S-Ethyl-4-methoxyphenyl glutathione (**6g**) –  $^{13}\text{C}$  NMR

Current Data Parameters  
 NAME jh81292302  
 EXPNO 4  
 PROCNO 1

F2 – Acquisition Parameters  
 Date 20120223  
 Time 14.52  
 INSTRUM avc500  
 PROBHD 5 mm CPDUL 13C  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1041  
 DS 2  
 SWH 31250.000 Hz  
 FIDRES 0.476837 Hz  
 AQ 1.0486259 sec  
 RG 1820  
 DW 16.000 usec  
 DE 20.00 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.03000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1  $^{13}\text{C}$   
 P1 10.00 usec  
 PL1 -4.40 dB  
 PL1W 28.15752029 W  
 SFO1 125.8131151 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2  $^1\text{H}$   
 PCPD2 80.00 usec  
 PL2 -6.00 dB  
 PL12 12.42 dB  
 PL13 18.42 dB  
 PL2W 15.19999981 W  
 PL12W 0.21869738 W  
 PL13W 0.05493430 W  
 SFO2 500.3020012 MHz

F2 – Processing parameters  
 SI 32768  
 SF 125.8005350 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

