Supporting information

A BODIPY-based fluorescent probe for the differential recognition of Hg(II) and Au(III) ions

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1. General methods

All reagents were purchased from commercial suppliers (Aldrich and Merck) and they were used without further purification. ¹H NMR and ¹³C NMR were measured on a Varian VNMRJ 400 Nuclear Magnetic Resonance Spectrometer. UV absorption spectra were obtained on Shimadzu UV-2550 Spectrophotometer. Fluorescence measurements were performed by using Varian Cary Eclipse Fluorescence spectrophotometer. Samples were contained in 10.0 mm path length quartz cuvettes (2.0 mL volume). Upon excitation at 460 nm, the emission spectra were integrated over the range 480 nm to 750 nm. The slit width was 5 nm for both excitation and emission. Melting points were determined by using an Electrothermal Melting Point Apparatus 9200. The pH was recorded by HI-8014 instrument (HANNA). All measurements were conducted at least in triplicate.

2. Synthesis section

The synthesis pathway for **BOD-ZN** was shown in Scheme S1. **BOD-1** and **BOD-AL** were synthesized according to the literature procedure^[1]. **BOD-1** was converted to its aldehyde form by using the well known Vilsmeier Haack's formylation reaction. By using hydrazine hydrate, phenyl isothiocyanate and catalytic amount of acetic acid in methanol, **BOD-ZN** was synthesized.



Scheme S1: Synthesis pathway of **BOD-ZN. (i)** DCM, RT, overnight, then Et₃N and BF₃OEt₂ (ii) POCl₃, DMF, 0°C, then DCE, 60°C, overnight, (iii) Hydrazine hydrate, phenyl isothiocyanate, methanol, reflux.

Synthesis of BOD-ZN



To a mixture of **BOD-AL** (52.1 mg, 0.15 mmol) and hydrazine hydrate (12 μ L, 0.18 mmol) in 5 ml of methanol, phenyl isothiocyanate (23 μ L, 0.18 mmol) was injected. Then, a drop of glacial acetic acid was added and resulting solution was refluxed for an hour. After cooling room temperature, the solvent was removed under reduced pressure. The resultant residue was purified by column chromatography (10:1 (Hexane:Ethyl acetate)) to afford **BOD-ZN** as purple solid (29.4 mg, 40 % yield)^[2]. Mp: 228-231 °C. ¹H NMR (400 MHz, CDCl₃) δ : 9.92 (s, 1H), 8.95 (s, 1H), 7.97 (s, 1H), 7.60 (d, J=8.0 Hz, 2H), 7.52-7.51 (m, 3H), 7.36 (t, J=8.0 Hz, 2H), 7.28-7.26 (m, 2H), 7.21 (t, J=7.2 Hz 1H), 6.09 (s, 1H), 2.75 (s, 3H), 2.59 (s, 3H), 1.50 (s, 3H), 1.40 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ : 174.7, 159.4, 153.9, 145.9, 142.5, 139.6, 138.1, 137.9, 134.5, 133.0, 130.5, 129.4, 128.7, 127.9, 125.9, 123.9, 123.1, 122.1, 14.9, 14.7, 14.1, 12.5. HRMS (EI) calc. for C₂₇H₂₆BF₂N₅S [M-H]⁺ 500.18918, found 500.18066.

3. Determination of quantum yields

Fluorescence quantum yields of **BOD-ZN** were determined by using optically matching solutions of Rhodamine 6G (Φ_F =0.95 in water) as a standard^[3]. The quantum yield was calculated according to the equation;

$$\Phi_{F(X)} = \Phi_{F(S)} (A_S F_X / A_X F_S) (n_X / n_S)^2$$

Where Φ_F is the fluorescence quantum yield, A is the absorbance at the excitation wavelength, F is the area under the corrected emission curve, and n is the refractive index of the solvents used. Subscripts S and X refer to the standard and to the unknown, respectively.

4. Cell imaging

A549 Human Lung Adenocarcinoma cell lines were grown in DMEM supplemented with 10% FBS (fetal bovine serum) in an atmosphere of 5 % CO₂ at 37 °C. The cells were plated on 12 mm cover glasses in 6-well plate and allowed to grow for 24h. Before the experiments, the cells were washed with PBS buffer, and then the cells were incubated **BOD-ZN** (10 μ M) for 20 min at 37 °C then washed with PBS three times. After incubating with Au³⁺ and Hg²⁺ (10 μ M) for 20 min at 37 °C, cells were rinsed with PBS three times, and DAPI for 10 min at 37°C then washed with PBS three times. Then, the fluorescence images were acquired through an Olympus IX71 fluorescence microscope.

5. Determination of the association constant and stoichiometry

The association constant of [Hg²⁺] was determined by using fluorescence titration data with the help of following equation;^[4]

$$\ln[(F - F_0) / (F_{\text{max}} - F_0)] = n \ln[\text{Hg}^{2^+}] + n \ln(K_{\text{asscn}})....(1)$$

where n is the number of mercury ions associating with each molecule of **BOD-ZN**, K_{assoc} is the association constant, F_0 is the fluorescence of the free probe, F_{max} is the fluorescence intensity at saturation point, and F is the fluorescence of probe obtained with Hg²⁺ addition,



Figure S1: Plot of $\ln[(F-F_0)/(F_{max}-F)]$ against $\ln[Hg^{2+}]$; the stoichimetry of **BOD-ZN** Hg^{2+} association, obtained directly from the slope, is $1.9 \approx 2$. Following equation 1, the intercept gave an association constant of **BOD-ZN** as $4.2 \times 10^4 \text{ M}^{-2}$ (in 0.1 M potassium phosphate buffer, pH 7.0/EtOH (v/v, 1:4) (λ_{ex} : 460 nm, $\lambda_{em} = 480$ nm at 25 °C).

6. Determination of detection limits

a) For Au³⁺;

The detection limit was calculated based on the fluorescence titration^[1]. To determine the S/N ratio, the emission intensity of **BOD-ZN** (10 μ M) without Au³⁺ was measured by 10 times and the standard deviation of blank measurements was determined. Under the present conditions, a good linear relationship between the fluorescence intensity and Au³⁺ concentration could be obtained in the 0.1-1.0 μ M (R = 0.9915). The detection limit is then calculated with the equation: detection limit = 3σ bi/m, where σ bi is the standard deviation of blank measurements; m is the slope between intensity versus sample concentration. The detection limit was determined to be 128.0 nm at S/N = 3.



Figure S2: Fluorescence changes of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) upon the addition of Au³⁺ (0.01 to 0.1 equiv.) (0.1-1 μ M)

b) For Hg²⁺;

Under the present conditions, a good linear relationship between the fluorescence intensity and Hg²⁺ concentration could be obtained in the 0.1-1.0 μ M (R = 0.9931). The detection limit was measured to be 160.0 nM at S/N = 3.



Figure S3: Fluorescence changes of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) upon addition of Hg²⁺ (0.01 to 0.1 equiv.) (0.1-1 μ M)

6. References

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7. Absorption and fluorescence spectra of BOD-ZN with Au³⁺



Figure S4: a) Absorbance and **b)** fluorescence spectra of **BOD-ZN** (10 μ M) in the presence of 10.0 equiv. (100 μ M) of AuCl₃ measured in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) ($\lambda_{ex} = 460 \text{ nm}, \lambda_{em} = 480 \text{ nm}$)

8. Time-dependent fluorescence change of BOD-ZN with Au³⁺



Figure S5: Time-dependent fluorescence change of **BOD-ZN** (10 μ M) in the presence of 1.0 (**•**), 3.0 (**•**) and 5.0 (**•**) equiv. (10, 30 and 50 μ M) of AuCl₃ measured in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) ($\lambda_{ex} = 460$ nm, $\lambda_{em} = 480$ nm)

9. Fluorescence titration of BOD-ZN with Au³⁺



Figure S6: Fluorescence spectra of BOD-ZN (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) in the presence of Au³⁺ (mole equivalents = 0.01-10.0) (0.1-100 μ M) (λ_{ex} = 460 nm, λ_{em} = 480 nm) Inset: Fluorescence intensity changes of BOD-ZN vs. equivalents of Au³⁺.

10. The fluorescence responses of BOD-ZN with Hg²⁺, Au³⁺, and other metal ions



Figure S7: Fluorescence intensities of BOD-ZN (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) in the presence of 20.0 equiv. (200 μ M) of the cations interests: BOD-ZN only , Au⁺, Ba²⁺, Ca²⁺, Cd²⁺, Cr³⁺, Cu²⁺, Fe³⁺, F⁻, I⁻, Li⁺, Mg²⁺, Ni²⁺, Pb²⁺, Zn²⁺ (λ_{ex} = 460 nm, λ_{em} = 480 nm)

11. The fluorescence responses of BOD-ZN in the presence of Au³⁺ and other metal ions



Figure S8: Fluorescence intensities of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) in the presence of 20.0 equiv. (200 μ M) of the cations interest : 1, **BOD-ZN** only; 2, Au³⁺; 3, Au⁺; 4, Ba²⁺; 5, Ca²⁺; 6, Cd²⁺; 7, Cr³⁺; 8, Cu²⁺; 9, Fe³⁺; 10, K⁺; 11, Li⁺; 12, Mg²⁺; 13, Ni²⁺; 14, Zn²⁺; 15, Ag⁺; 16, F⁻ ($\lambda_{ex} = 460 \text{ nm}, \lambda_{em} = 480 \text{ nm}$)

12. Effect of pH



Figure S9: Effect of pH on the fluorescence intensity of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (v/v, 1:4) in the absence and presence of Au³⁺ (10.0 equiv., 100 μ M) ($\lambda_{ex} = 460$ nm, $\lambda_{em} = 480$ nm)

13. Effect of fraction of water



Figure S10: Effect of fraction of water on the fluorescence intensity of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) in the absence and presence of Au³⁺ (10.0 equiv., 100 μ M) (λ_{ex} = 460 nm, λ_{em} = 480 nm)





Figure S11: a) Absorbance and **b)** fluorescence spectra of **BOD-ZN** (10 μ M) in the presence of 10.0 equiv. (100 μ M) of HgCl₂ measured in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4)

15. Time-dependent fluorescence change of BOD-ZN with Hg²⁺



Figure S12: Time-dependent fluorescence change of **BOD-ZN** (10 μ M) measured in 0.1 M Phosphate Buffer/EtOH (pH = 7.0, v/v, 1:4) in the presence of 10 equiv. (100 μ M) of HgCl₂ ($\lambda_{ex} = 460 \text{ nm}, \lambda_{em} = 480 \text{ nm}$)

16. Fluorescence titration of BOD-ZN with Hg²⁺



Figure S13: Fluorescence spectra of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) in the presence of Hg²⁺ (mole equivalents = 0.01-10.0) (0.1-100 μ M) (λ_{ex} = 460 nm, λ_{em} = 480 nm) Inset: Fluorescence intensity changes of **BOD-ZN** vs. equivalents of Hg²⁺

17. The fluorescence responses of BOD-ZN in the presence of Hg²⁺ and other metal ions



Figure S14: Fluorescence intensities of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) in the presence of 20.0 equiv. (200 μ M) of the cations interest : 1, **BOD-ZN** only; 2, Hg²⁺; 3, Au⁺; 4, Ba²⁺; 5, Ca²⁺; 6, Cd³⁺; 7, Cr²⁺; 8, Cu²⁺; 9, Fe²⁺; 10, I⁻; 11, Mg²⁺; 12, Ni²⁺; 13, Pb²⁺; 14, Zn²⁺; 15, Ag⁺; 16, F⁻($\lambda_{ex} = 460 \text{ nm}, \lambda_{em} = 480 \text{ nm}$)

18. The fluorescence intensity changes of BOD-ZN in the presence of Hg²⁺ ions and Na₂S



Figure S15: Fluorescence intensity changes of **BOD-ZN** (10 μ M) in EtOH at pH = 7.0 at $\lambda_{max} = 542$ nm after addition of 10 equiv. Hg²⁺, 10 equiv. Hg²⁺ + 10 equiv. Na₂S, 10 equiv. Hg²⁺ + 10 equiv. Na₂S + 10 equiv. Hg²⁺, respectively. ($\lambda_{ex} = 460$ nm, $\lambda_{em} = 480$ nm)

19. Effect of pH



Figure S16: Effect of pH on the fluorescence intensity of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (v/v, 1:4) in the absence and presence of Hg²⁺ (10.0 equiv., 100 μ M) ($\lambda_{ex} = 460 \text{ nm}, \lambda_{em} = 480 \text{ nm}$)

20. Effect of fraction of water



Figure S17: Effect of fraction of water on the fluorescence intensity of **BOD-ZN** (10 μ M) in 0.1 M phosphate buffer/EtOH (pH = 7.0, v/v, 1:4) in the absence and presence of Hg²⁺ (10.0 equiv., 100 μ M) (λ_{ex} = 460 nm, λ_{em} = 480 nm)

21. Selectivity of BOD-ZN in the presence of both Hg²⁺ and Au³⁺ ions



Figure S18: Selectivity of **BOD-ZN** (10 μ M) in the presence of both Hg²⁺ and Au³⁺ (10 equiv., 100 μ M) in 0.1 M phosphate buffer/EtOH (pH 7.0, v/v, 1:4) ($\lambda_{ex} = 460$ nm, $\lambda_{em} = 480$ nm)

22. Photograph image of the hydrolysis reaction of BOD-ZN mediated by Au(III)



Figure S19: a) Day light photograph image of BOD-ZN + Au(III) (left), BOD-ZN (right),
(b) fluorescence image of BOD-ZN + Au(III) (left), BOD-ZN (right)

23. Photograph image of the coordination of BOD-ZN mediated by Hg(II) ions



Figure S20: a) Day light photograph image of **BOD-ZN** + Hg(II) (left), **BOD-ZN** (right), (b) fluorescence image of **BOD-ZN** + Hg(II) (left), **BOD-ZN** (right)

24. ¹H and ¹³C NMR spectra





Counts vs. Mass-to-Charge(m/z)

25. HRMS spectra of BOD-ZN in the absence and presence of Hg²⁺ ion

