

## Supporting Information

### Thioester-anchored organosilatrane: Synthetic investigations and applications in Cu<sup>2+</sup> ion binding and fabrication of hybrid silica nanoparticles

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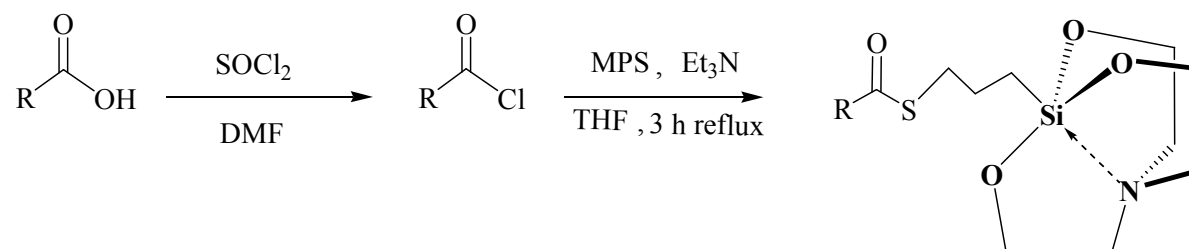
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### Optimization of reaction conditions

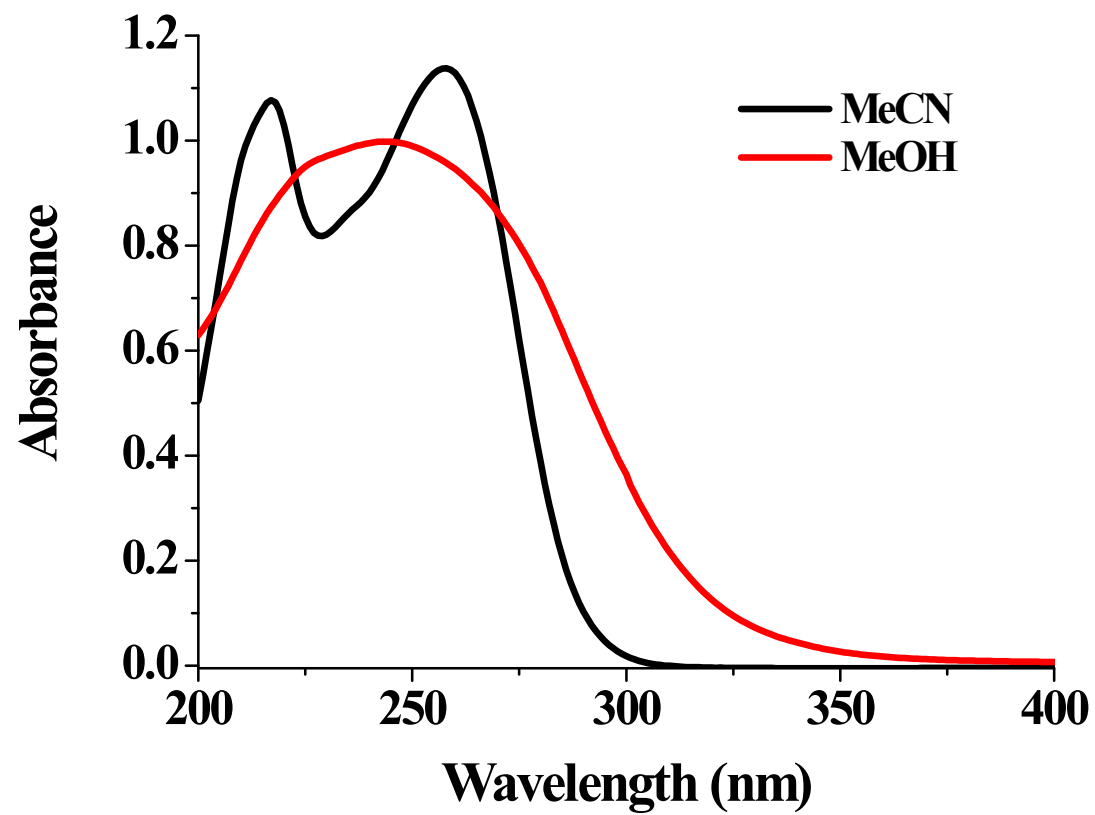
To start with, the thioester formation from free carboxylic acid **3a** and MPS was examined. As expected, the reaction did not proceed at all (Table 1, entry 1); indicating that direct thioesterification is not possible under simple azeotropic reflux conditions. The catalytic activity of conc. H<sub>2</sub>SO<sub>4</sub> was also tested for the reaction and only 30% product yield was achieved after 8 h (entry 2). In literature, there are different ways for activating carboxyl component of carboxylic acids in the form of acyl halides, acyl azides, acylimidazoles, anhydrides etc [1]. The commonly used method for the synthesis of thioesters involves the conversion of carboxylic acids into more reactive acyl chlorides, their isolation, and then reaction with thiols. Although with this method, the reaction could be accomplished in a short time period (5 h), but the moisture sensitivity of acyl chlorides, handling of thionyl chloride and formation of toxic HCl gas were the main disadvantages. Besides this, in the coupling of the acid chloride with MPS, an additional base was required to soak up the by-product HCl (Scheme 1).



### Scheme 1: Synthesis of ThE-OS derivatives using acid chlorides

These unavoidable limitations associated with this method enforced us to use some other coupling reagents. As CDI is fairly cheap, readily available and capable of generating clean reactions with innocuous by-products, we have therefore opted for the CDI mediated coupling of acids with MPS. Initially with carboxylic acid:CDI in 1:1 ratio, only 50% yield was obtained. The reaction was further attempted with acid and CDI in different ratios (Table 1, entries 5-7). Using 1:1.2 ratio of acid and CDI respectively, yield of the product improved considerably from 50 to 76%. No further improvement was observed by changing the ratios (entries 6 and 7). The solvent screening investigations revealed that THF is a better choice amongst all tried solvents (entries 8-10). Altogether, we can say that the CDI mediated coupling of heterocyclic carboxylic acids with MPS serves as an attractive route to integrate aromatic thioesters with silatranyl unit.

1. S. Iimura, K. Manabe and S. Kobayashi, *Chem. Commun.*, **2002**, 94–95.



**Fig. S1** UV-Vis plot of **3a** (0.1 mM) in acetonitrile and methanol

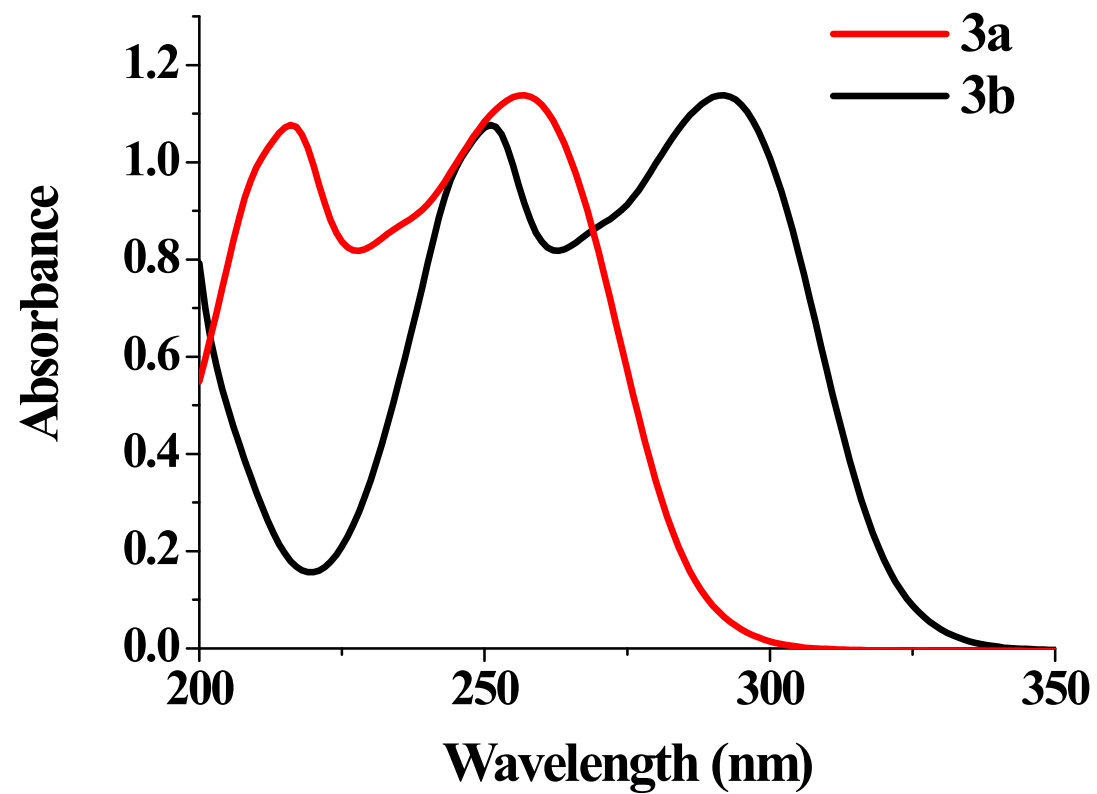


Fig. S2 UV-Vis plot of **3a** and **3b** at 0.1 mM concentration in acetonitrile

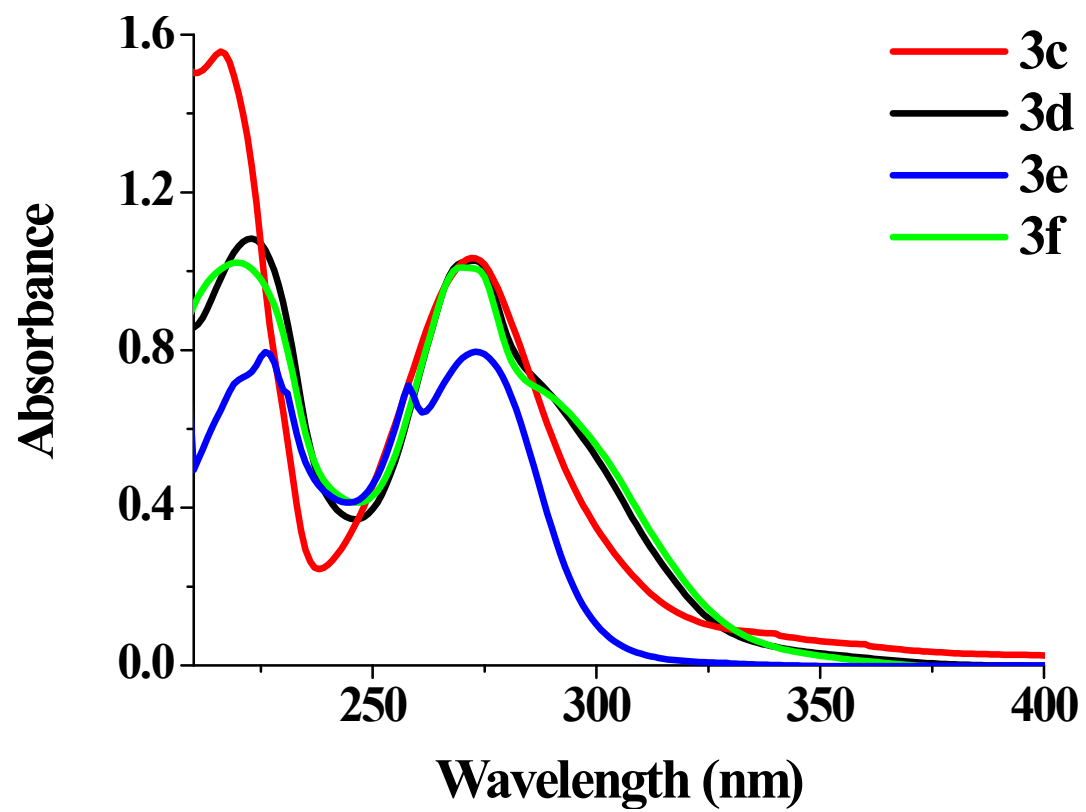


Fig. S3 UV-Vis plot of 3c, 3d, 3e and 3f at 0.1 mM concentration in acetonitrile

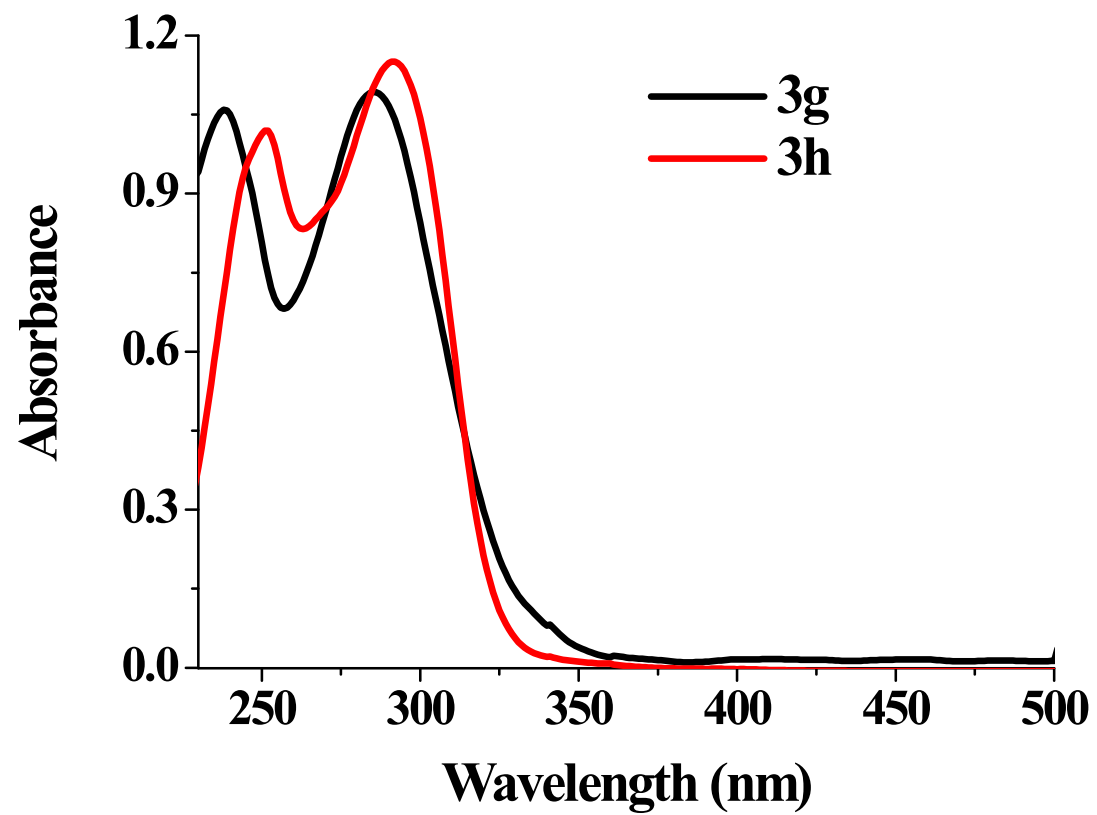
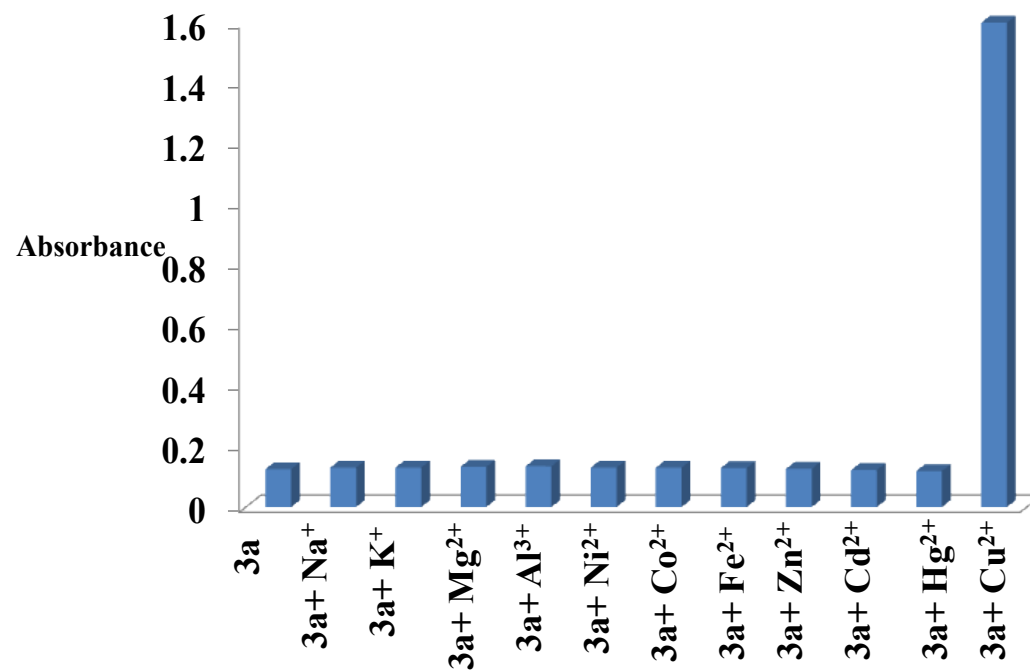
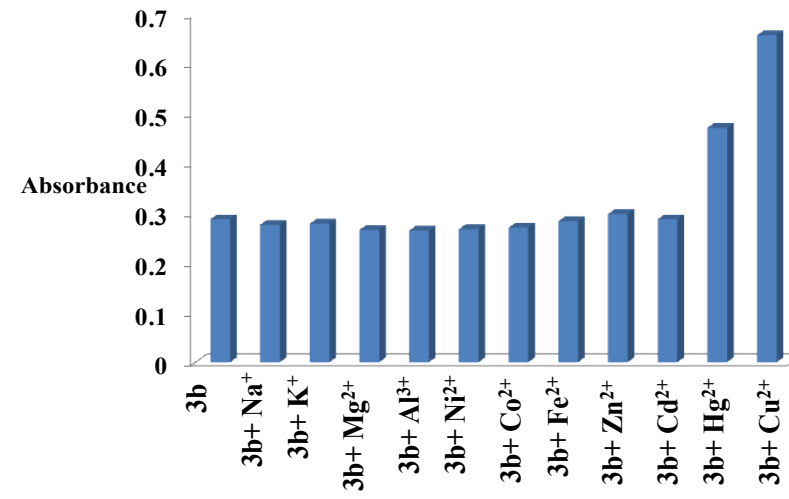


Fig. S4 UV-Vis plot of **3g** and **3h** at 0.1 mM concentration in acetonitrile

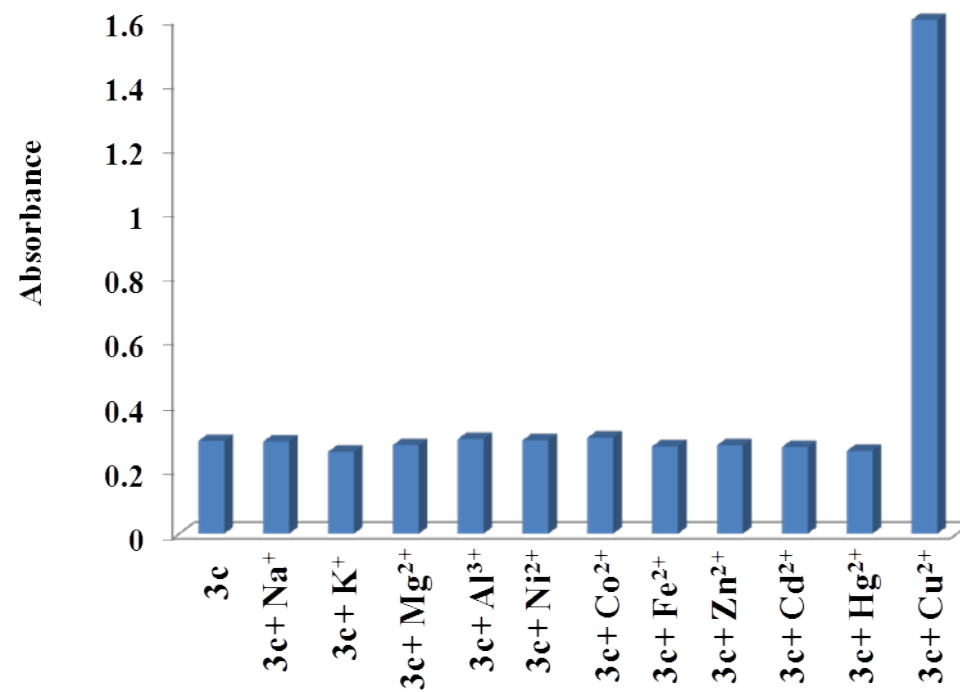


**Fig. S5** Chemosensing activity of probe **3a** towards different cationic species (20 equiv.) in acetonitrile

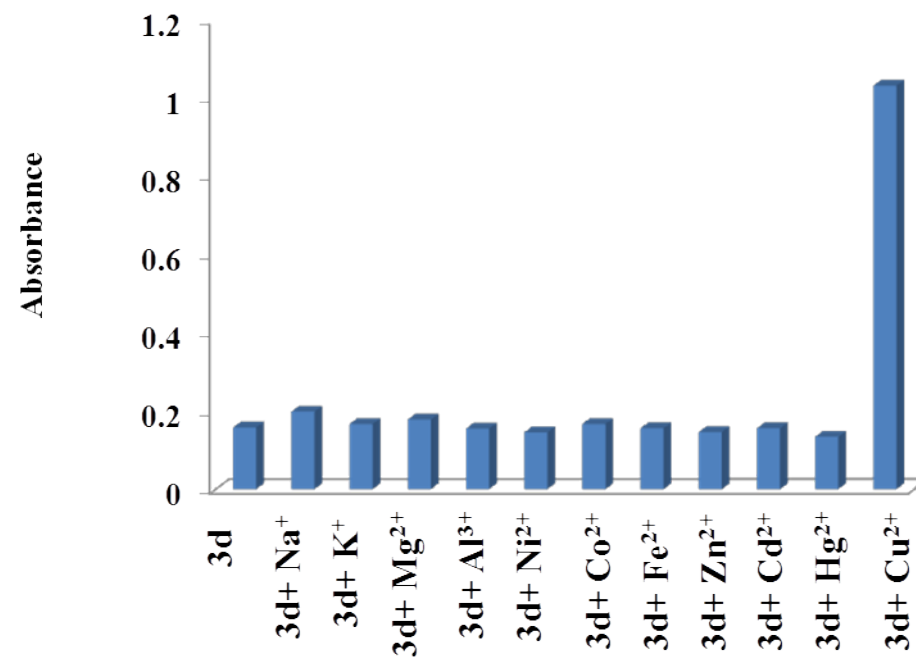




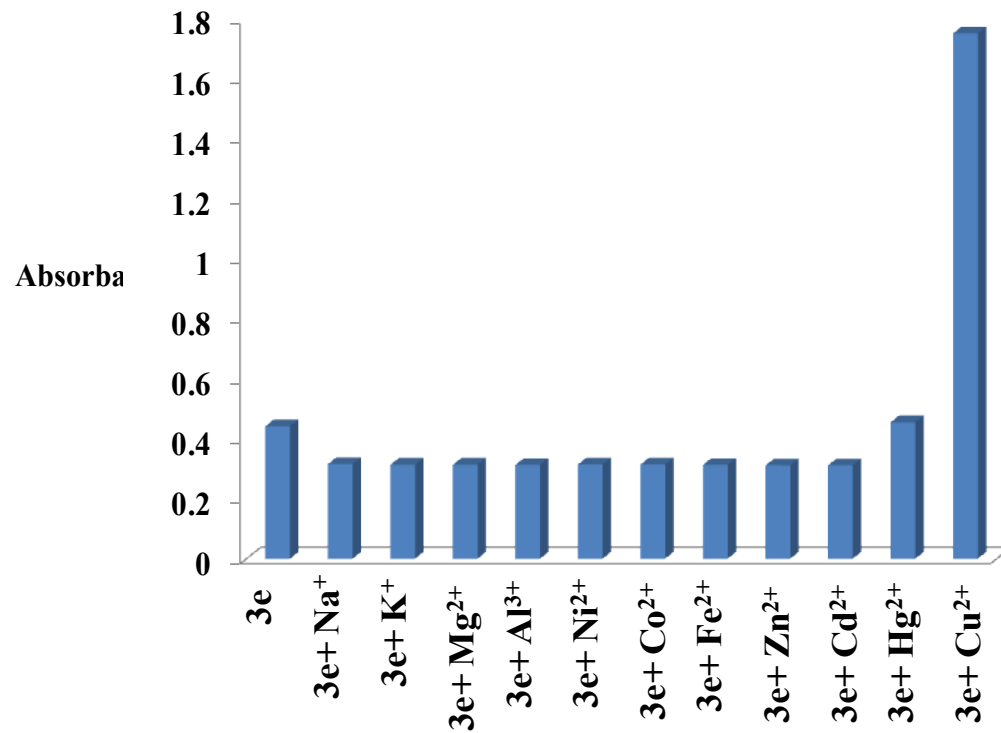
**Fig. S6** Chemosensing activity of probe **3b** towards different cationic species (20 equiv.) in acetonitrile



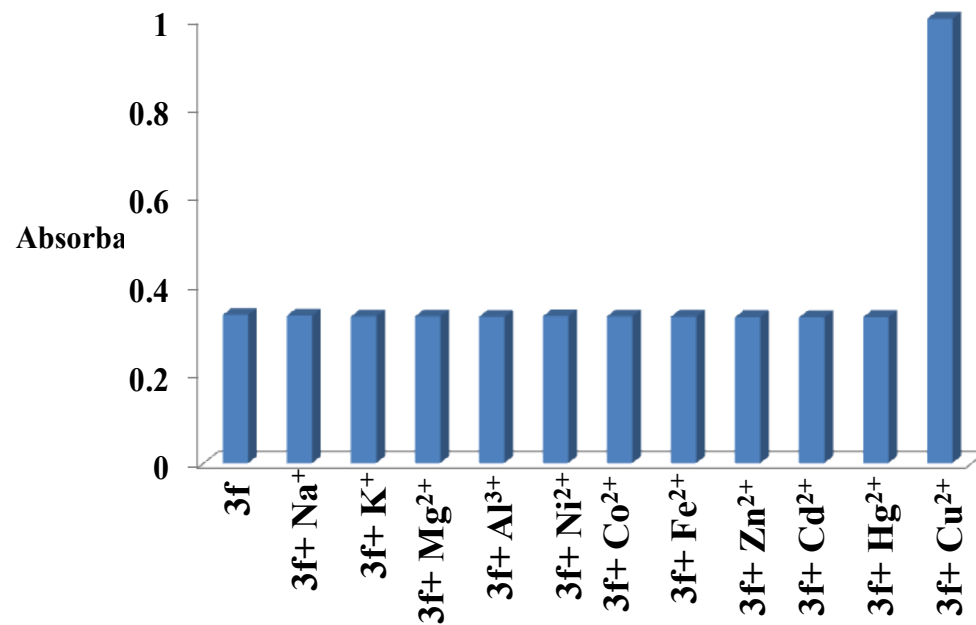
**Fig. S7** Chemosensing activity of probe **3c** towards different cationic species (20 equiv.) in acetonitrile



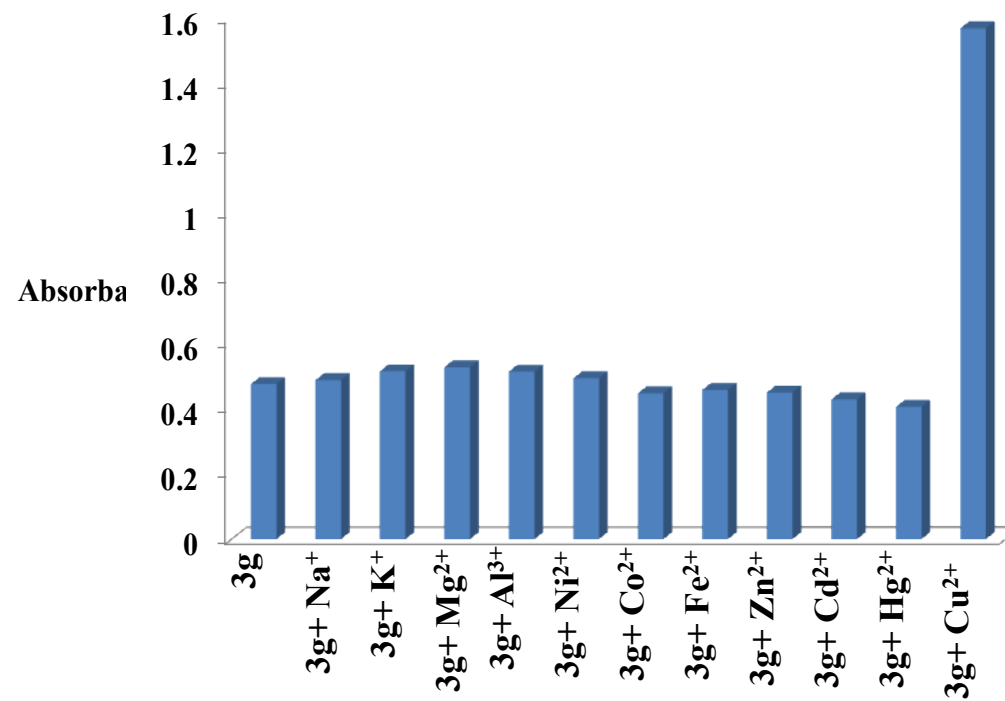
**Fig. S8** Chemosensing activity of probe **3d** towards different cationic species (20 equiv.) in acetonitrile



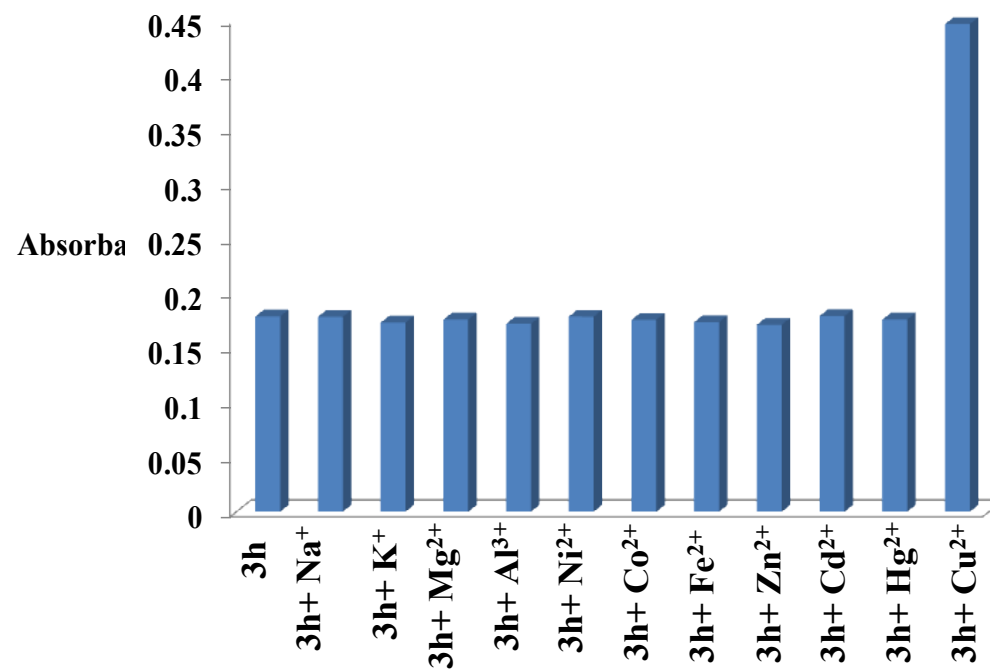
**Fig. S9** Chemosensing activity of probe **3e** towards different cationic species (20 equiv.) in acetonitrile



**Fig. S10** Chemosensing activity of probe **3g** towards different cationic species (20 equiv.) in acetonitrile



**Fig. S11** Chemosensing activity of probe **3g** towards different cationic species (20 equiv.) in acetonitrile



**Fig. S12** Chemosensing activity of probe **3h** towards different cationic species (20 equiv.) in acetonitrile

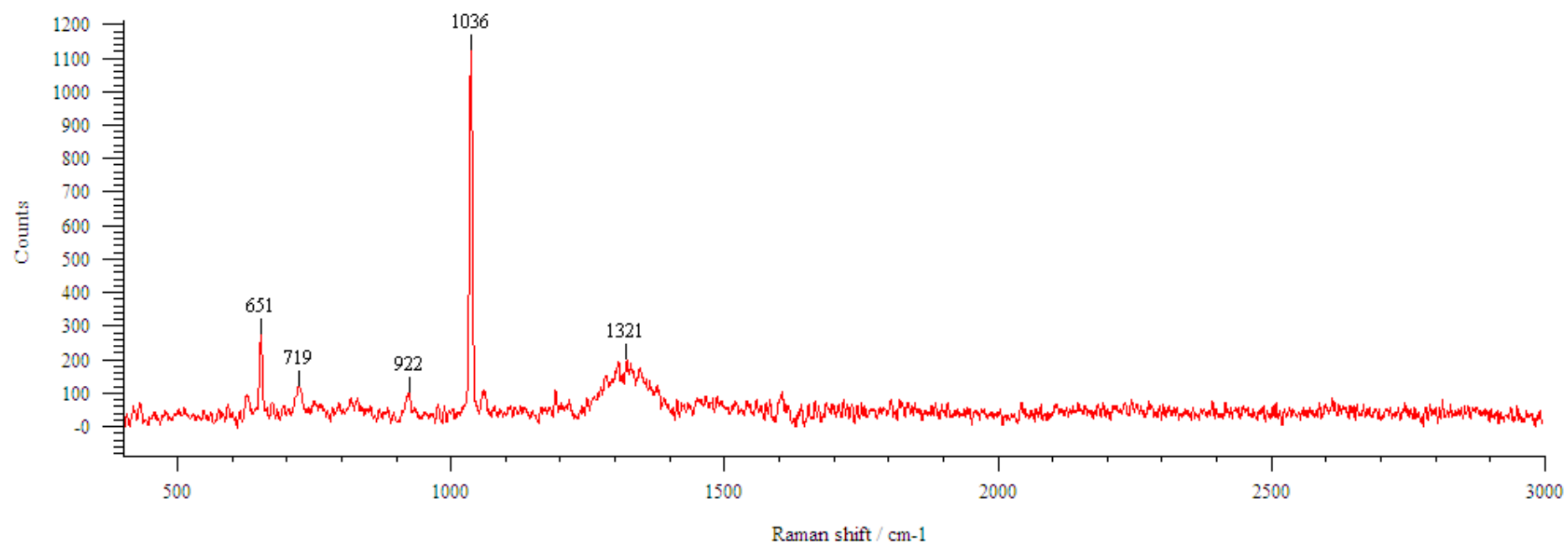
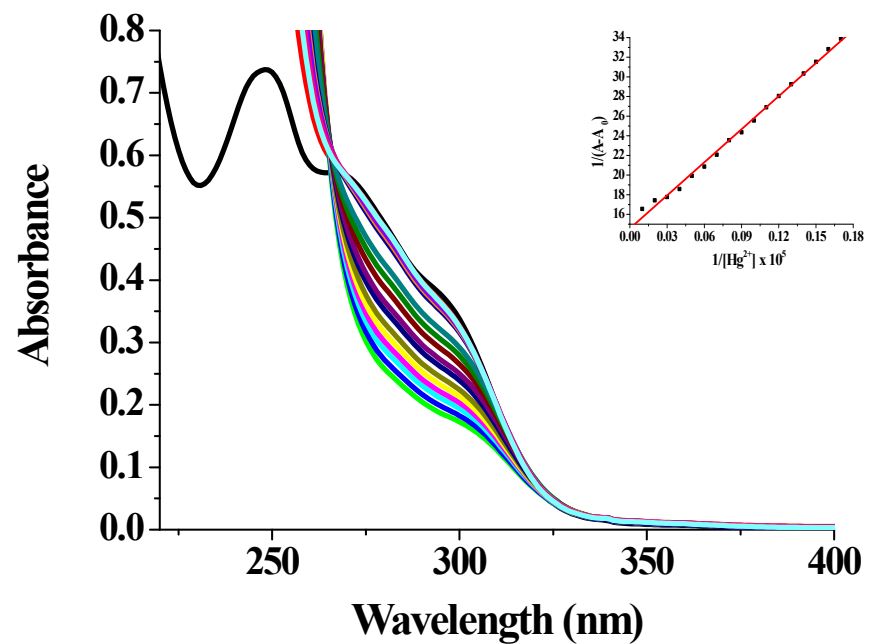


Fig. S13 Raman spectra of 3h·Cu<sup>2+</sup> complex





**Fig. S14** Family of UV–Vis spectra taken in the course of the titration of **3b** (1  $\mu\text{M}$  in  $\text{CH}_3\text{CN}$ ) with  $\text{Hg}^{2+}$  ions (0-5 equiv. (0-5 equiv.i.e. 0, 0.3, 0.6, 0.9, 1.2, 1.4, 1.5, 1.8, 2.1, 2.4, 2.7, 3, 4, 5 in the increasing order).). Inset shows the B-H plot for 1:1 complexation between **3b** and  $\text{Hg}^{2+}$  at 266 nm ( $R = .9979$ ) and  $K_a = 3.95 \times 10^5 \text{ M}^{-1}$

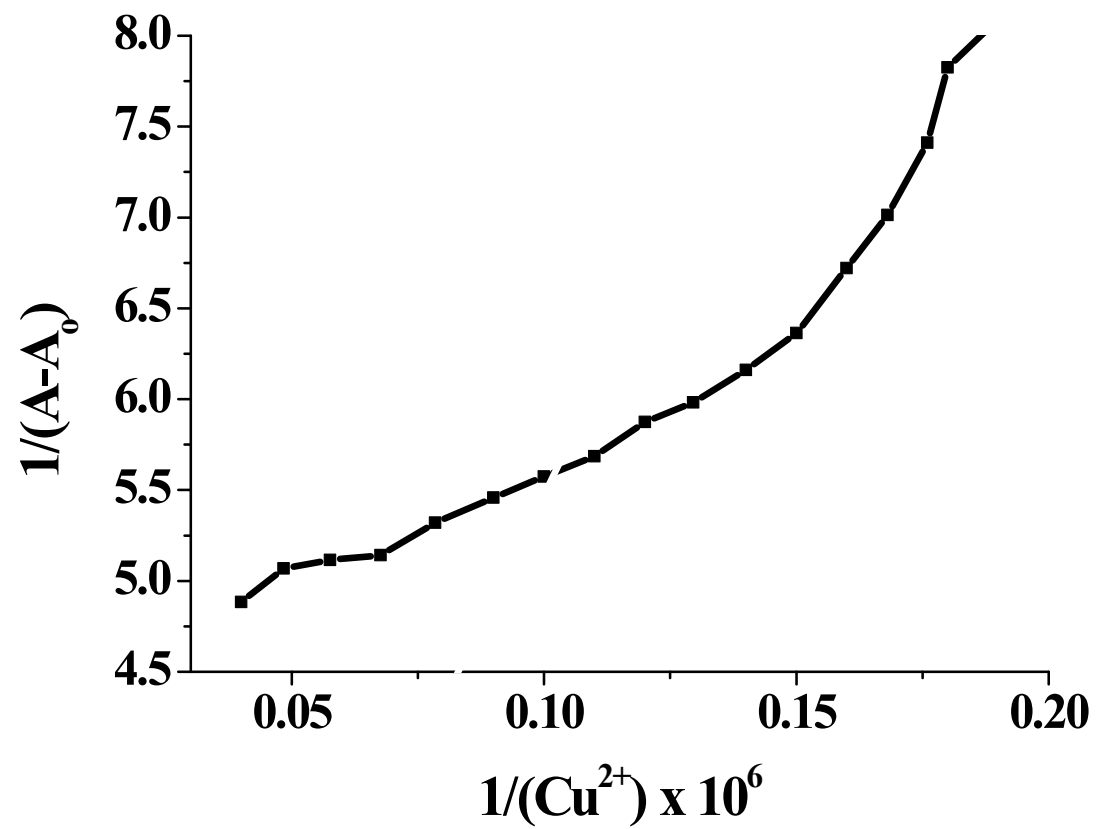
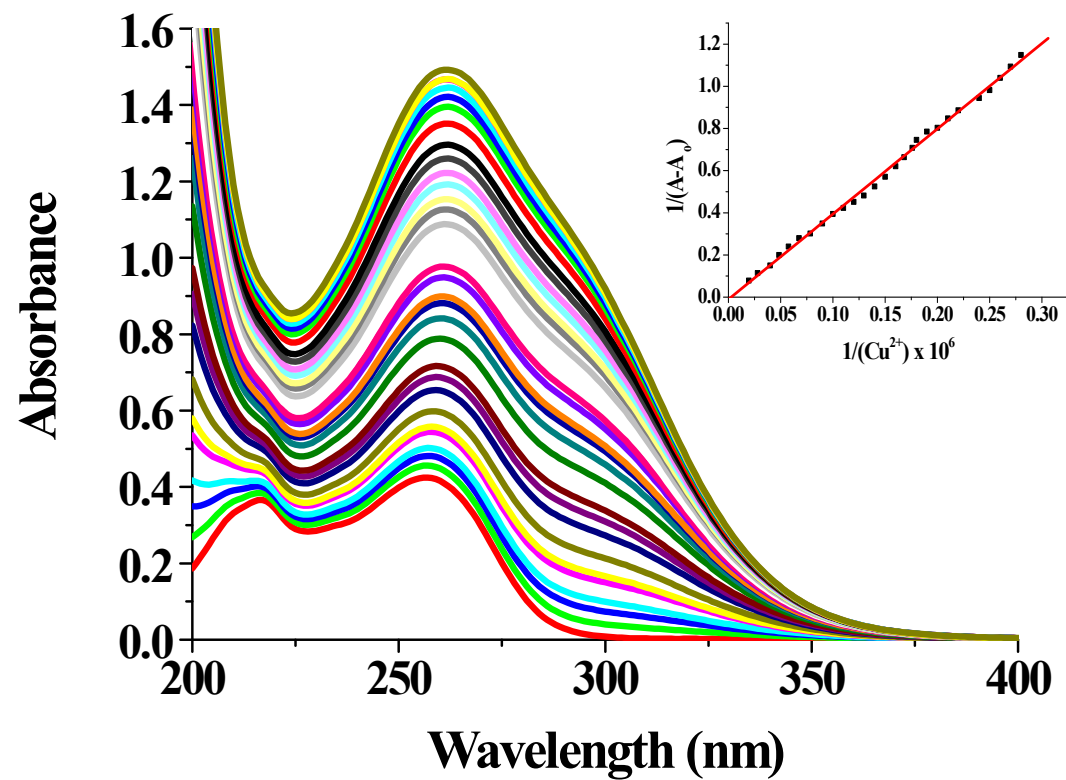
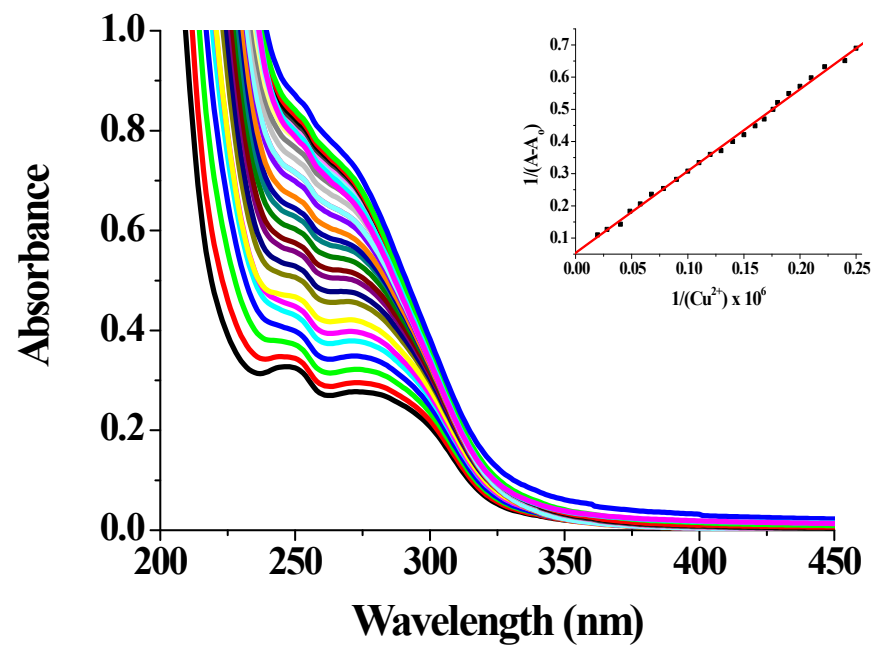


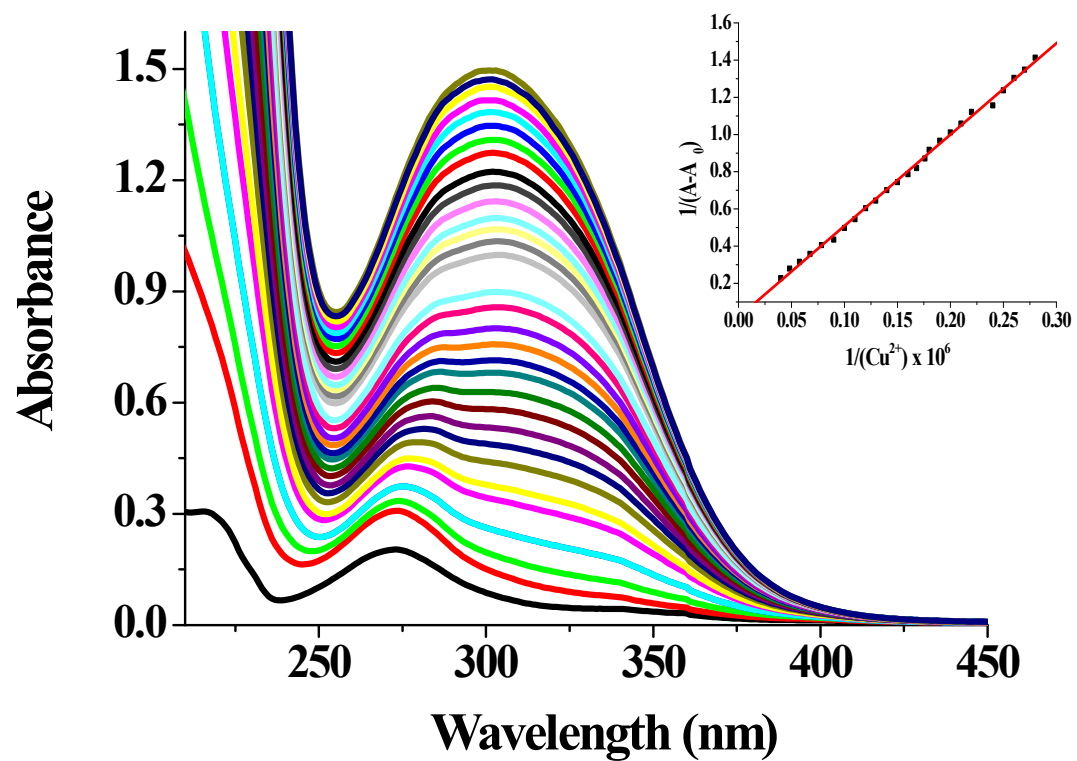
Fig. S15 B-H plot for 1:1 complexation between **3e** and  $\text{Cu}^{2+}$  at 296 nm



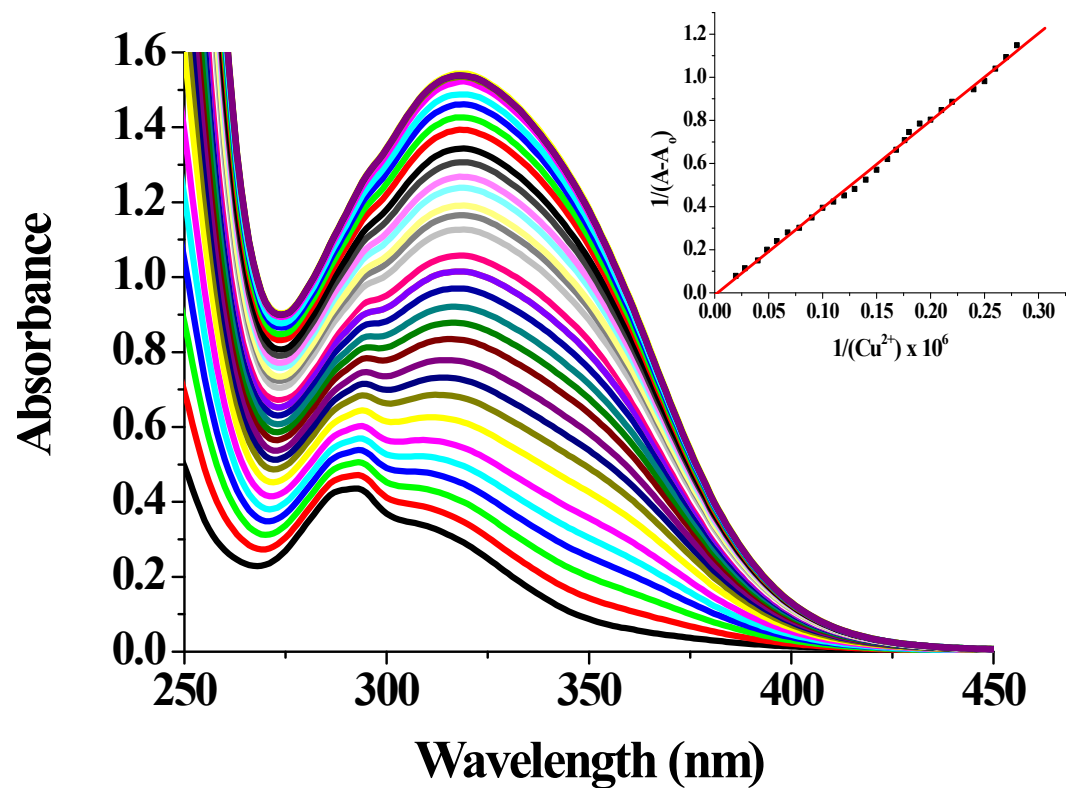
**Fig. S16** Family of UV-Vis spectra taken in the course of the titration of **3a** (1  $\mu\text{M}$  in  $\text{CH}_3\text{CN}$ ) with  $\text{Cu}^{2+}$  ions (0-10 equiv. 0, 0.4, 0.8, 1.2, 1.6, 2.0, 2.4, 2.8, 3.2, 3.6, 4.0, 4.4, 4.8, 5.2, 5.6, 6.0, 6.4, 6.8, 7.2, 7.6, 8.0, 8.4, 8.8, 9.2, 9.6, 10.0). Inset shows the B-H plot for 1:1 complexation between **3a** and  $\text{Cu}^{2+}$  at 261 nm ( $R = 0.9929$ )



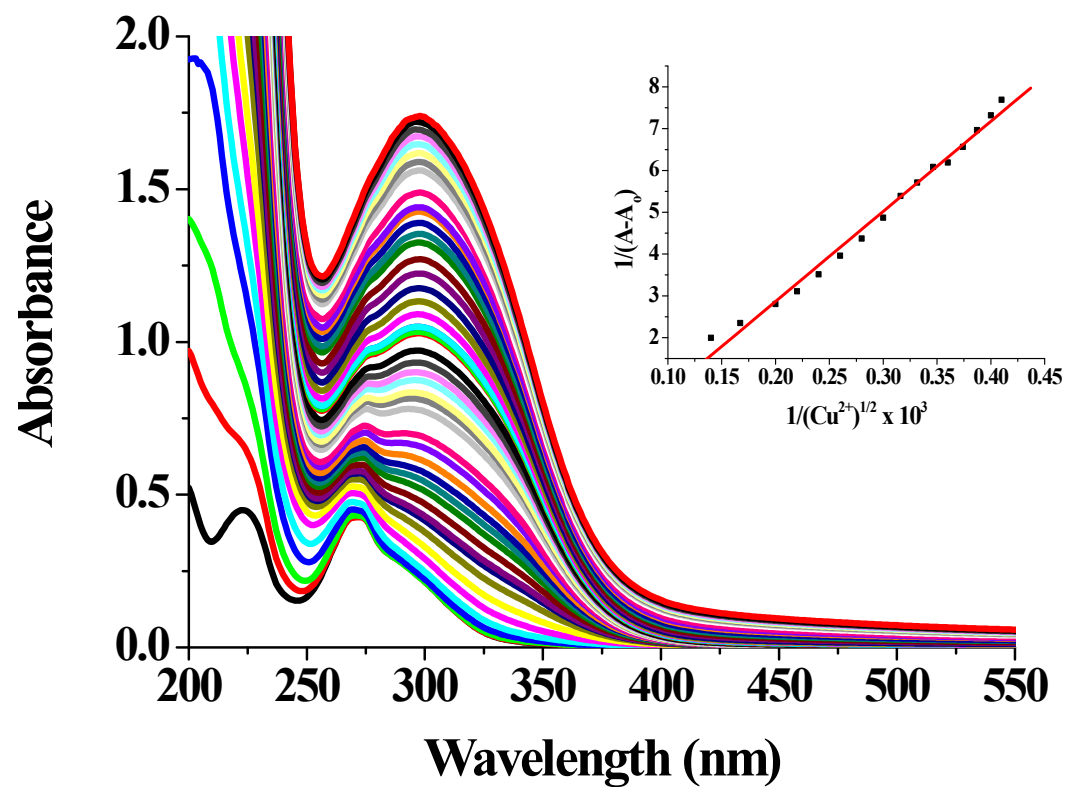
**Fig. S17** Family of UV–Vis spectra taken in the course of the titration of **3b** (1  $\mu\text{M}$  in  $\text{CH}_3\text{CN}$ ) with  $\text{Cu}^{2+}$  ions (0-4 equiv. i.e. 0, 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0). Inset shows the B-H plot for 1:1 complexation between **3b** and  $\text{Cu}^{2+}$  ion at 292nm ( $R = 0.9987$ )



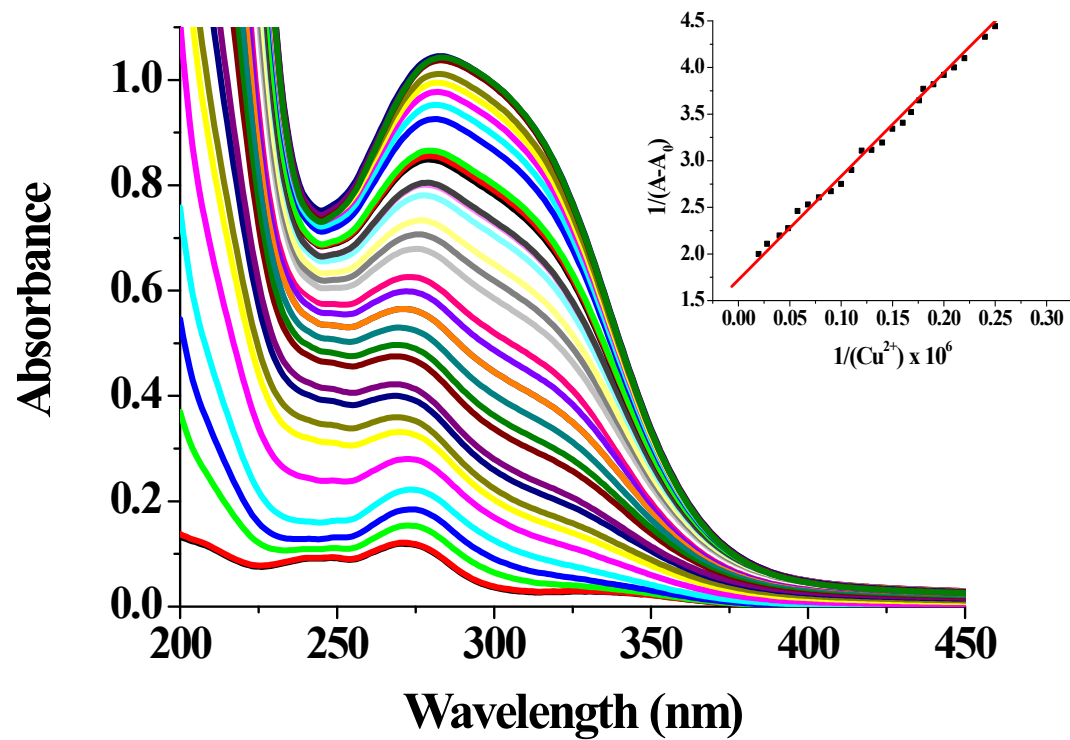
**Fig. S18** Family of UV-Vis spectra taken in the course of the titration of **3c** (1  $\mu\text{M}$  in  $\text{CH}_3\text{CN}$ ) with  $\text{Cu}^{2+}$  ions (0-4 equiv. i.e. 0, 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0). Inset shows the B-H plot for 1:1 complexation between **3c** and  $\text{Cu}^{2+}$  at 302 nm ( $R = 0.9989$ )



**Fig. S19** Family of UV-Vis spectra taken in the course of the titration of **3d** (1  $\mu\text{M}$  in  $\text{CH}_3\text{CN}$ ) with  $\text{Cu}^{2+}$  ions (0-4 equiv. i.e. 0, 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0). Inset shows the B-H plot for 1:1 complexation between **3d** and  $\text{Cu}^{2+}$  at 300 nm ( $R = 0.9987$ )

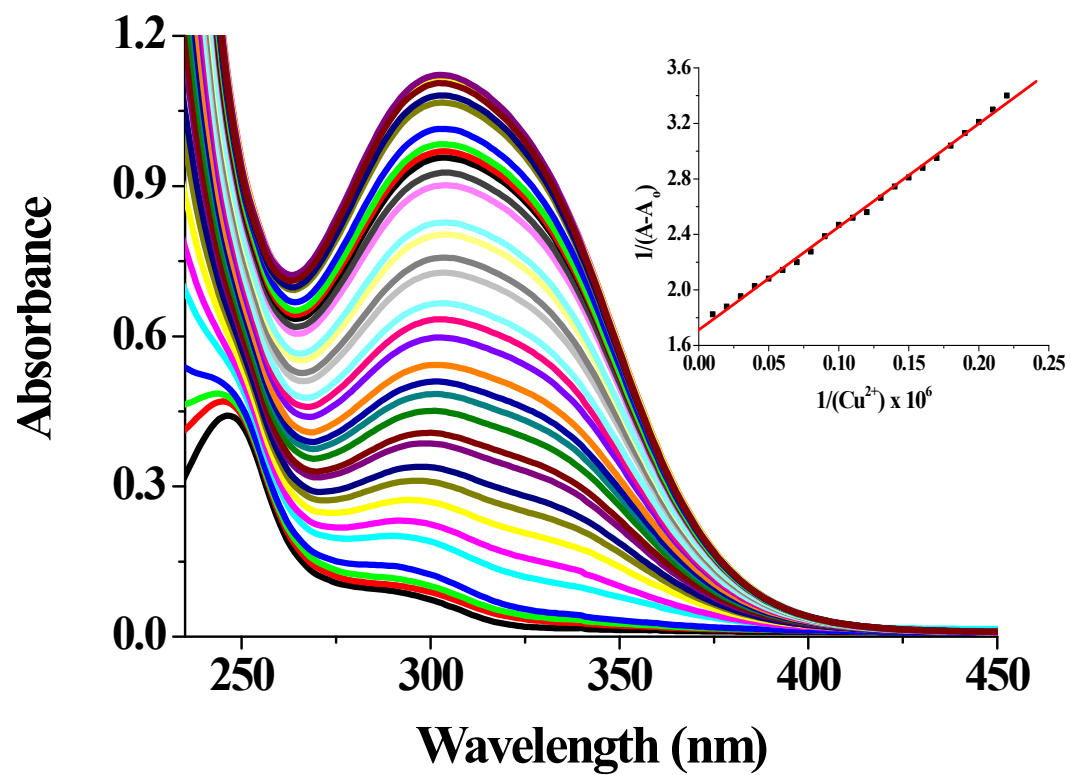


**Fig. S20** Family of UV-Vis spectra taken in the course of the titration of **3e** (1  $\mu\text{M}$  in  $\text{CH}_3\text{CN}$ ) with  $\text{Cu}^{2+}$  ions (0-10 equiv. i.e. 0, 0.3, 0.6, 0.9, 1.2, 1.5, 1.8, 2.1, 2.4, 2.7, 3.0, 3.3, 3.6, 3.9, 4.2, 4.5, 4.8, 5.1, 5.4, 5.7, 6.0, 6.3, 6.6, 6.9, 7.2, 7.5, 7.8, 8.1, 8.4, 8.7, 9, 10). Inset shows the B-H plot for 1:0.5 complexation between **3e** and  $\text{Cu}^{2+}$  at 296 nm ( $R = 0.9923$ ).

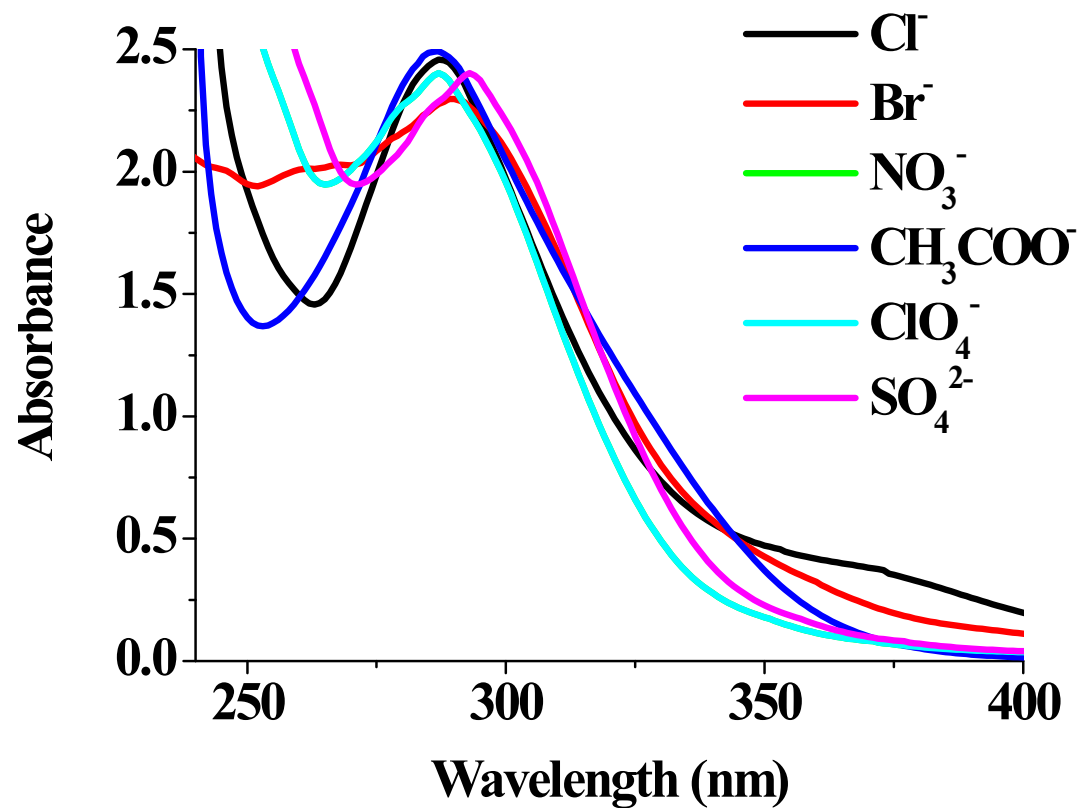


**Fig. S21** Family of UV-Vis spectra taken in the course of the titration of **3f** (1  $\mu\text{M}$  in  $\text{CH}_3\text{CN}$ ) with  $\text{Cu}^{2+}$  ions (0-10 equiv. i.e. 0, 0.4, 0.8, 1.2, 1.6, 2.0, 2.4, 2.8, 3.2, 3.6, 4.0, 4.4, 4.8, 5.2, 5.6, 6.0, 6.4, 6.8, 7.2, 7.6, 8.0, 8.4, 8.8, 9.2, 9.6, 10.0). Inset shows the B-H plot for 1:1 complexation between **3f** and  $\text{Cu}^{2+}$  at 283 nm ( $R = 0.9978$ )

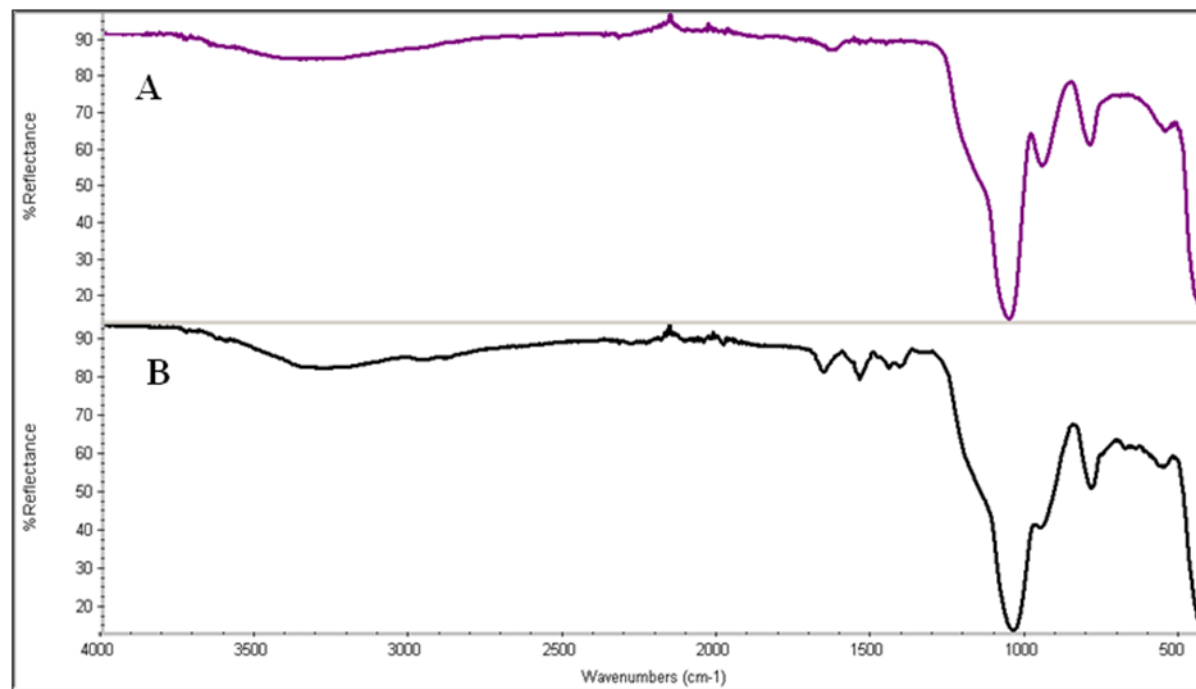




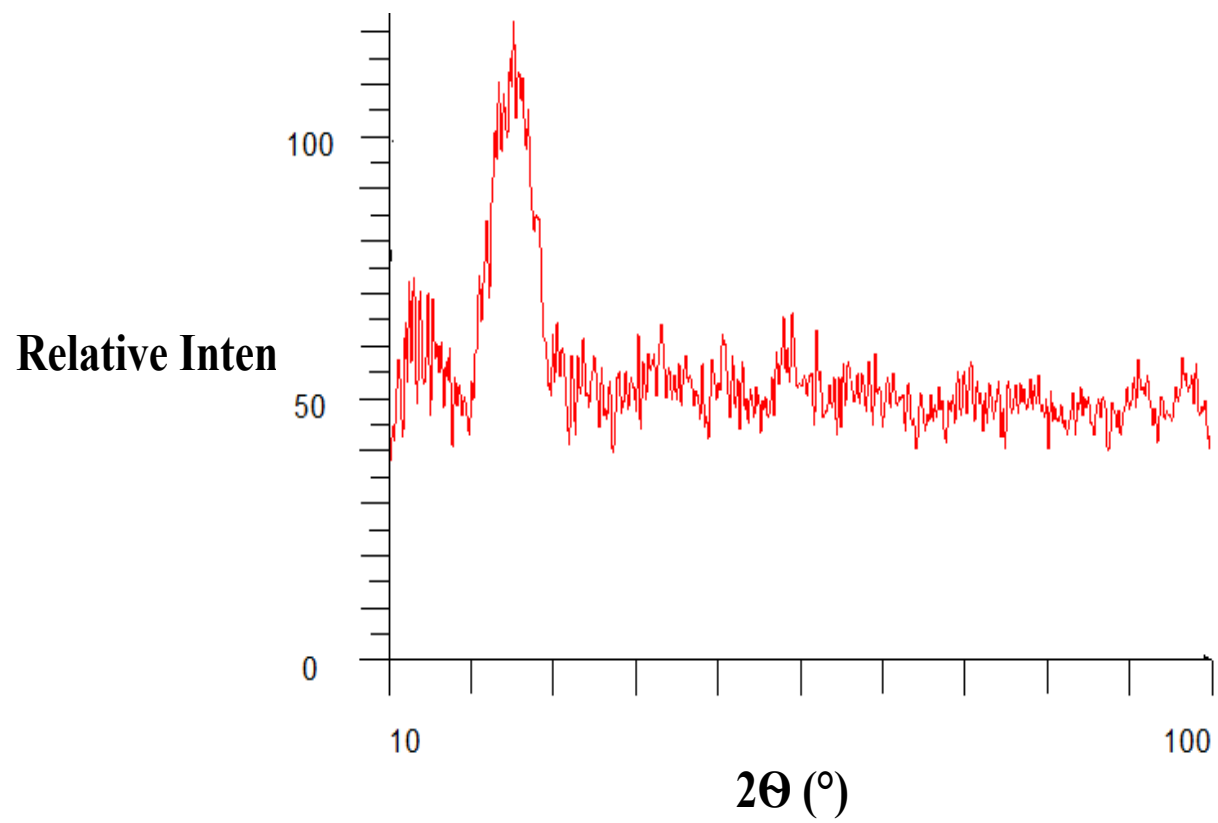
**Fig. S22** Family of UV-Vis spectra taken in the course of the titration of **3g** (1  $\mu\text{M}$  in  $\text{CH}_3\text{CN}$ ) with  $\text{Cu}^{2+}$  ions (0-4 equiv. i.e. 0, 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0). Inset shows the B-H plot for 1:1 complexation between **3g** and  $\text{Cu}^{2+}$  at 318 nm ( $R = 0.9879$ )



**Fig. S23** UV-Vis absorption spectra of **3h** in the presence of CuX<sub>2</sub> (where X = Cl<sup>-</sup>, Br<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, CH<sub>3</sub>COO<sup>-</sup>, ClO<sub>4</sub><sup>-</sup>, SO<sub>4</sub><sup>2-</sup>)



**Fig. S24** IR spectra of SiNPs (A) and H-SiNPs (B)



**Fig. S25** X-ray diffraction (XRD) graphs of SiNPs (A) and H-SiNPs (B)

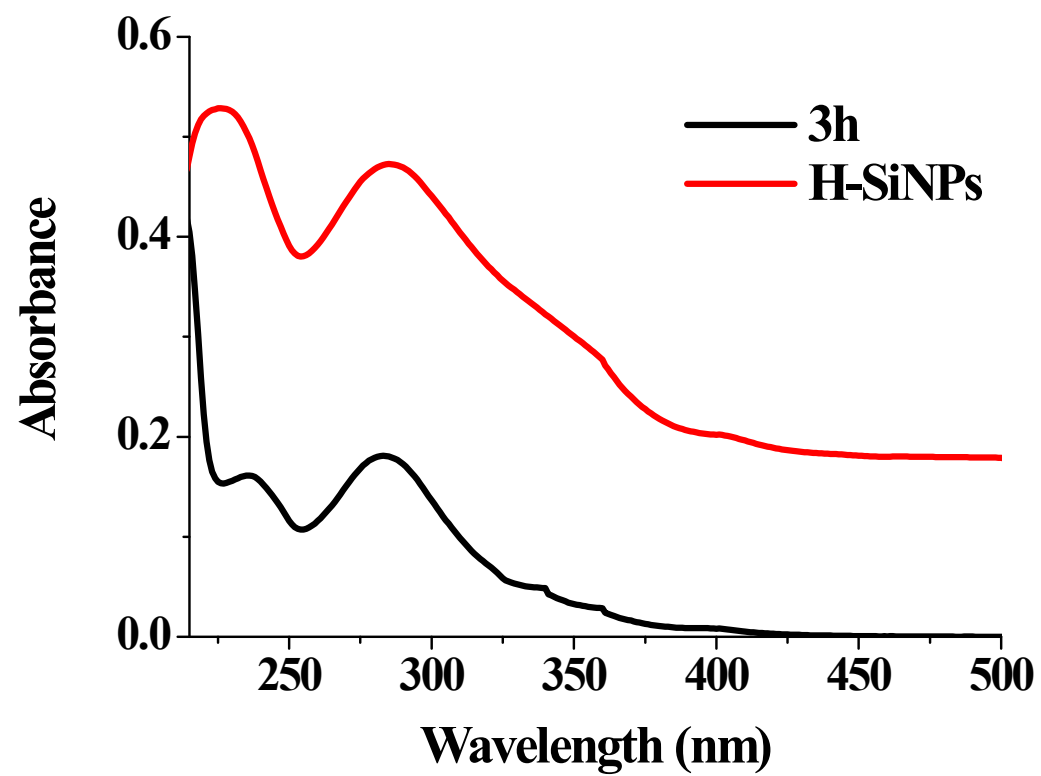
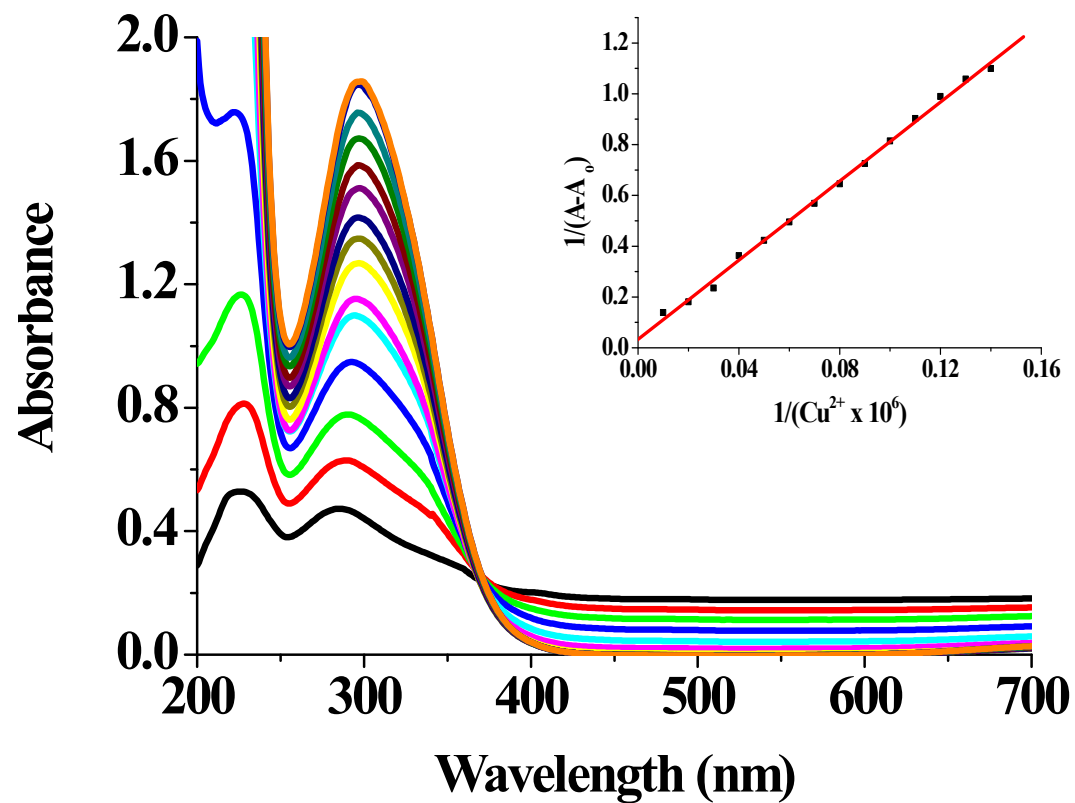
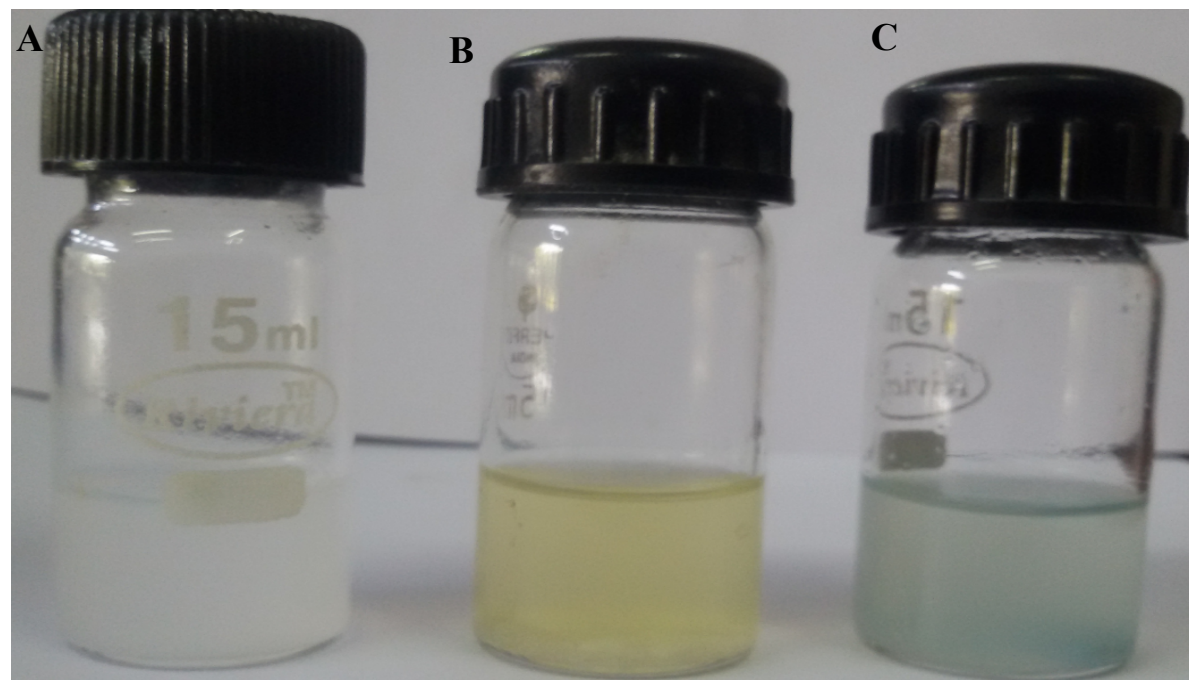


Fig. S26 UV-Vis plot of **3h** and **H-SiNPs** in acetonitrile



**Fig. S27** Family of UV-Vis spectra taken in the course of the titration of **H-SiNPs** (in CH<sub>3</sub>CN) with Cu<sup>2+</sup> ions. Inset shows the B-H plot for 1:1 complexation between **H-SiNPs** and Cu<sup>2+</sup> at 297nm (R = 0.9955)



**Fig. S28** Digital photographs of aqueous dispersions of SiNPs (A), H-SiNPs (B) and  $\text{Cu}^{2+}$ -H-SiNPs complex (C)

**Table T1** Selected crystal data and details on the structure determinations from single crystal data for compound **3e**

<b>Compound</b>	<b>3e</b>
Formula	C <sub>15</sub> H <sub>22</sub> N <sub>2</sub> O <sub>4</sub> SSi
MW [g·mol <sup>-1</sup> ]	354.49
Crystal system	Monoclinic
Space group	<i>P</i> 12 <sub>1</sub> / <i>n</i> 1
<i>a</i> [Å]	11.5580(1)
<i>b</i> [Å]	7.3752(7)
<i>c</i> [Å]	19.4783(2)
<i>α</i> [deg]	90
<i>β</i> [deg]	93.613(3)
<i>γ</i> [deg]	90
<i>V</i> [Å <sup>3</sup> ]	1657.1(3)
<i>T</i> [K]	170.15
<i>Z</i>	4



$\rho_{\text{calc}}$ [g·cm <sup>-3</sup> ]	1.421
$\mu$ [mm <sup>-1</sup> ]	0.289
Min/max transmission	0.961/0.977
$\theta_{\text{max}}$ [deg]	28.370
Measured reflections	27042
Unique reflections	4117
Reflections [ $F_0 > 4\sigma(F_0)$ ]	2944
Parameter	208
$R_{\text{int}}$	0.0713
$R_1$ [ $F_0 > 4\sigma(F_0)$ ]	0.0409
$wR_2$ [all data]	0.0978
GOF	1.018
$\Delta\rho_{\text{max}}/\Delta\rho_{\text{min}}$ [e·Å <sup>-3</sup> ]	0.300/-0.379

**Table T2** Comparison of different sensing systems bearing ‘thio’ functionality for the detection of metal ions

<b>Sensors</b>	<b>Sensing metals</b>	<b>Binding constants (M<sup>-1</sup>)</b>	<b>Stoichiometry</b>	<b>Reference</b>
Thio-urea conjugated 1,8-naphthalimide	Fe <sup>3+</sup>	1.854 × 10 <sup>3</sup>	1:1	42 (a)
Thio-urea conjugated pyrene	Cu <sup>2+</sup>	1.09 × 10 <sup>4</sup>	1:1	42 (b)
Thio-spirolactam conjugated rhodamine	Hg <sup>2+</sup>	1.82 × 10 <sup>5</sup>	1:1	42 (c)
Thio-semicarbazide conjugated naphthalimide	Ag <sup>+</sup>	4.80 × 10 <sup>4</sup>	1:1	42 (d)
thio- β-enaminone analog	Cu <sup>2+</sup>	1.10 × 10 <sup>4</sup>	1:1	13
<b>Bis (thio-ester) framed silatrane (3h)</b>	<b>Cu<sup>2+</sup></b>	<b>4.93 × 10<sup>5</sup></b>	<b>1:1</b>	<b>Present work</b>

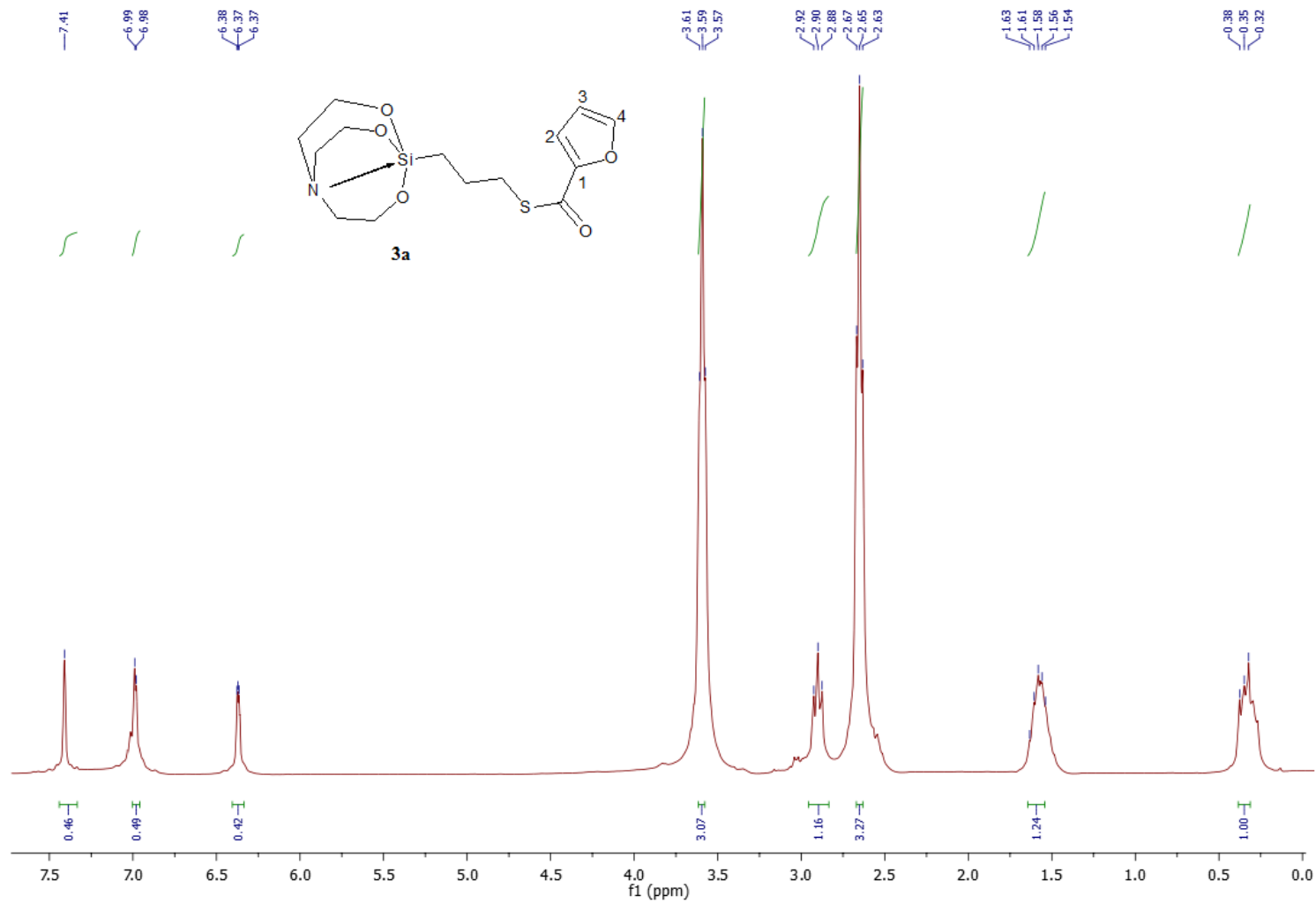


Fig. S29 <sup>1</sup>H NMR spectrum of 3a

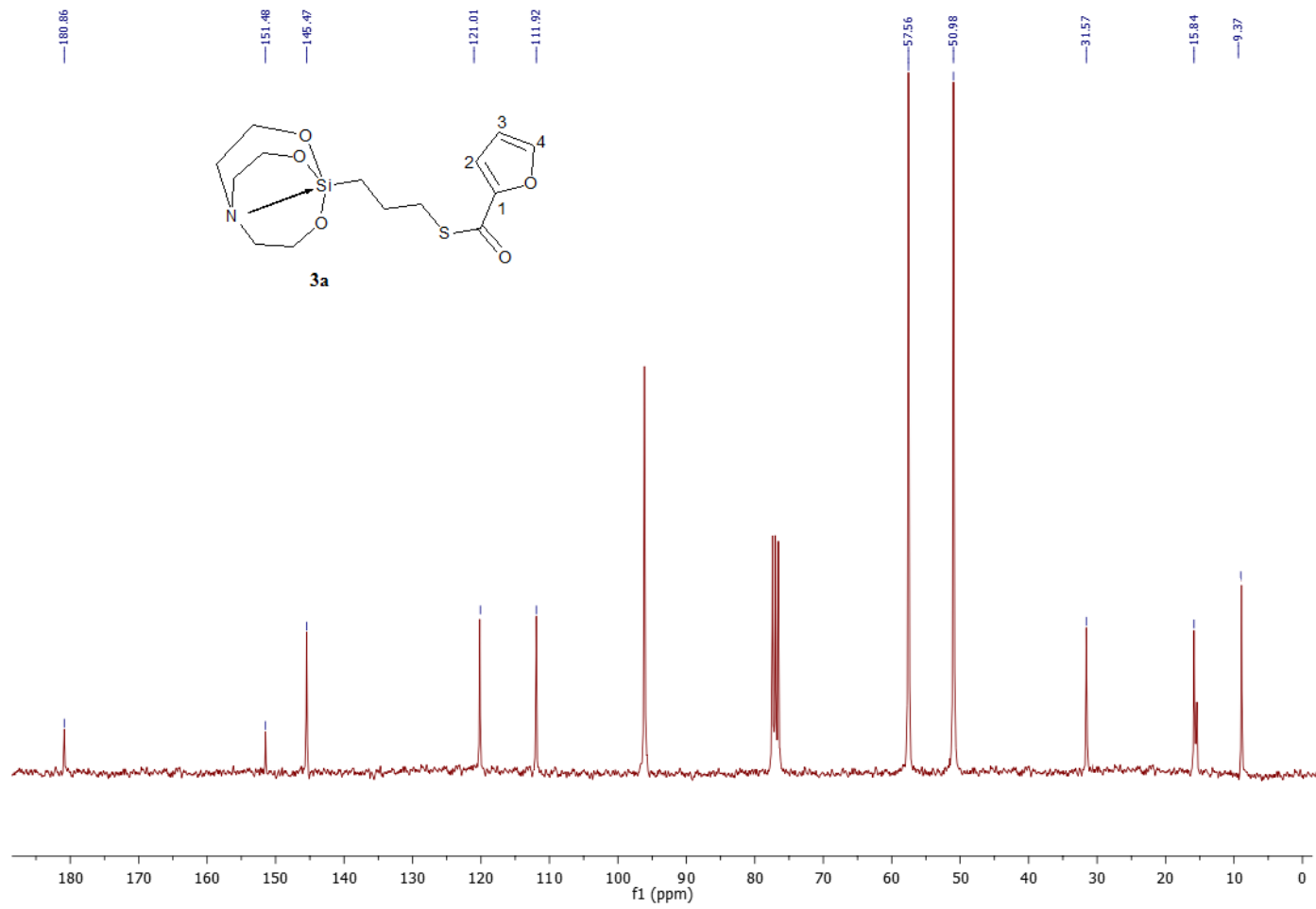


Fig. S30 <sup>13</sup>C NMR spectrum of 3a

WATERS, Q-TOF MICROMASS (LC-MS)

SUNITA S-2 10 (0.112) Cm (9:30)

SAIF/CIL,PANJAB UNIVERSITY,CHANDIGARH

TOF MS ES+  
4.25e3

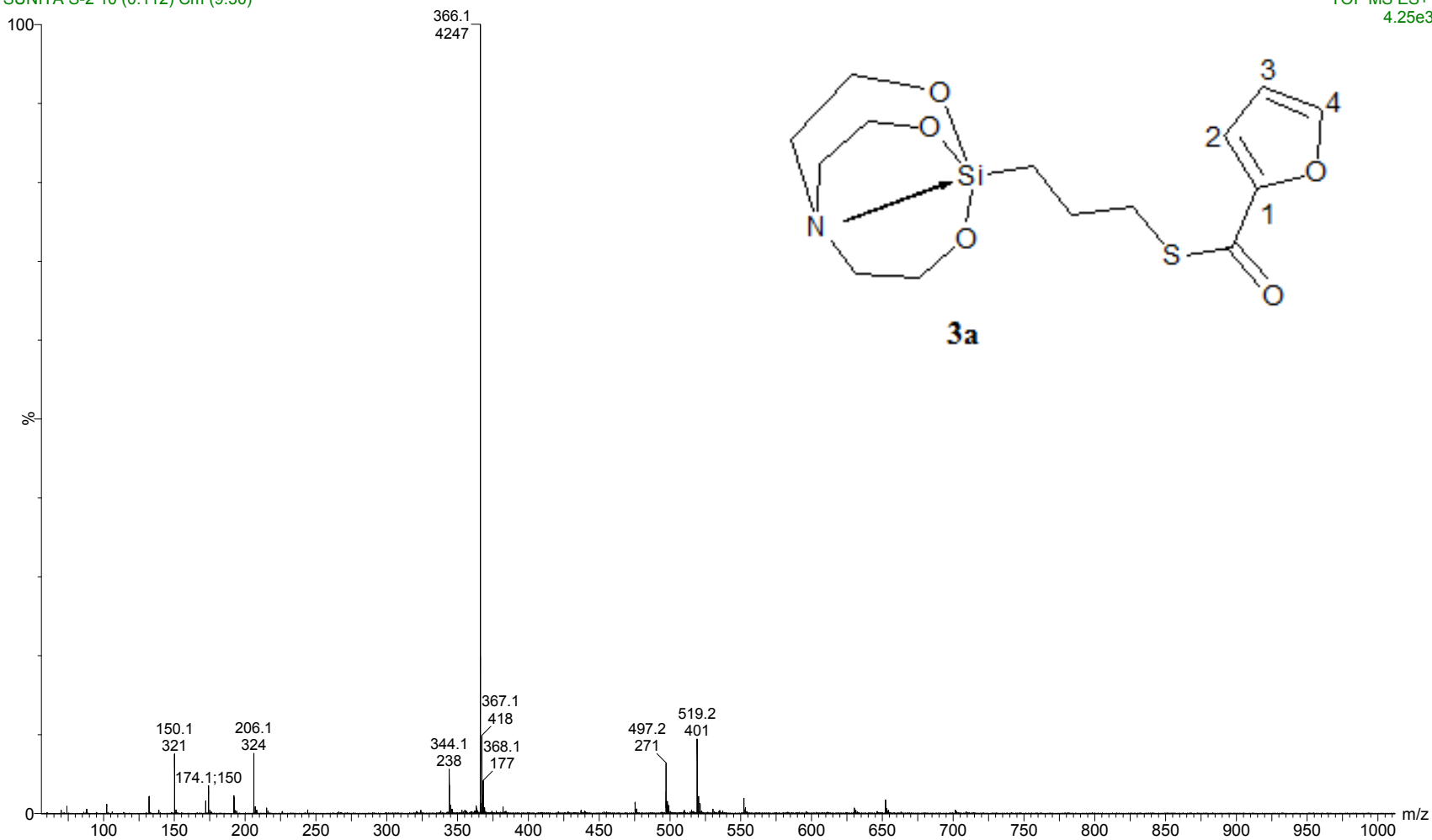


Fig. S31 Mass spectrum of 3a

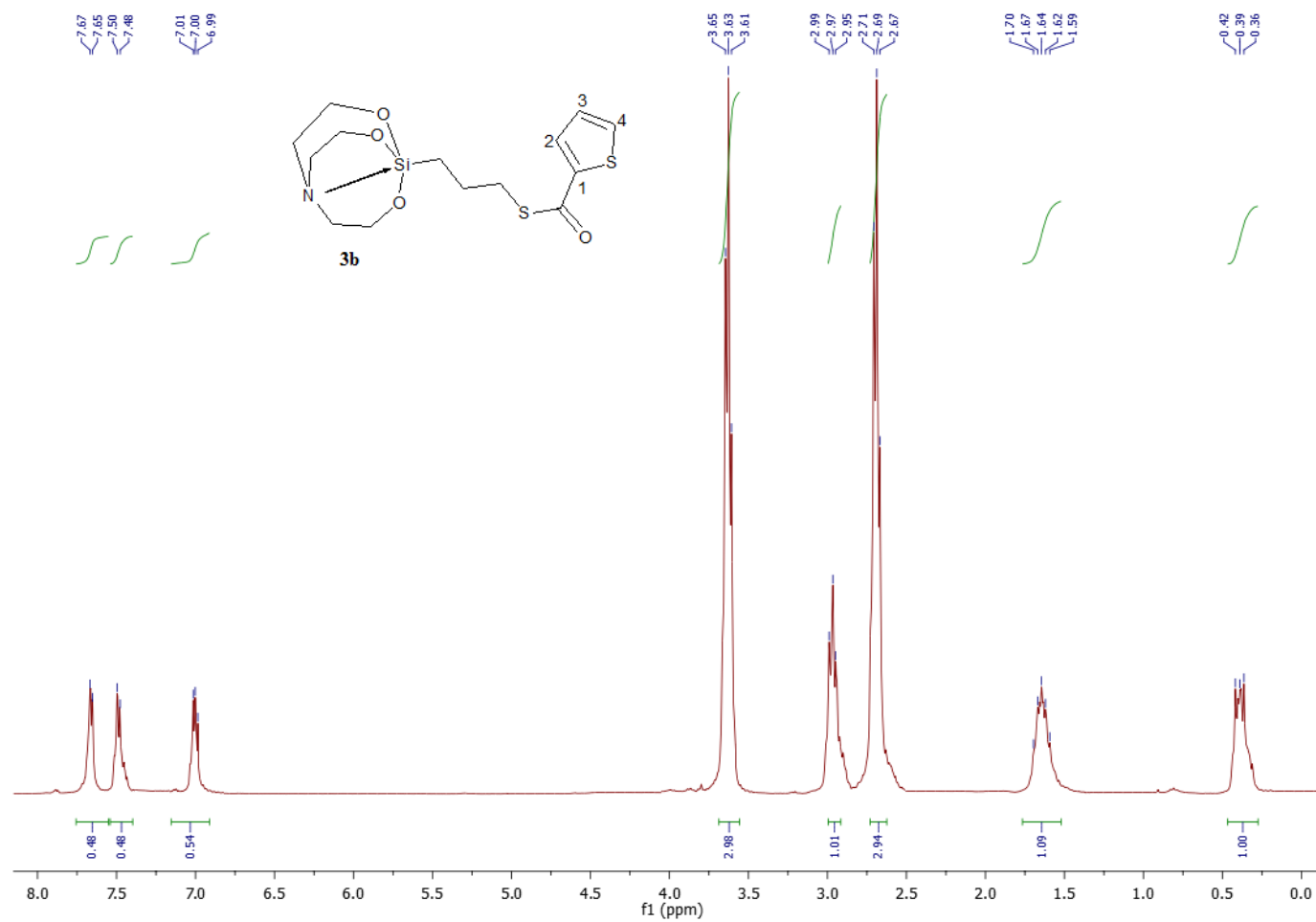
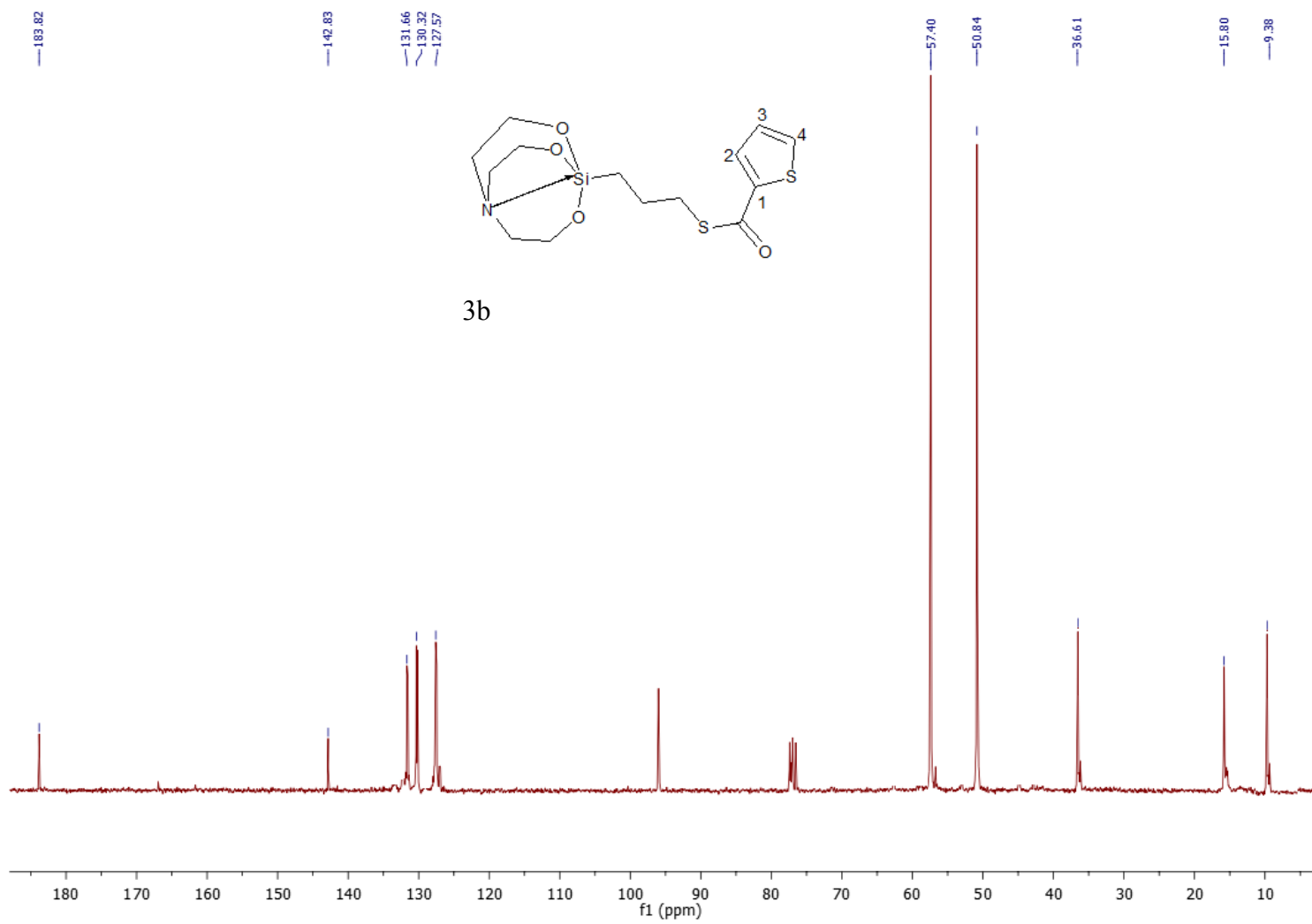
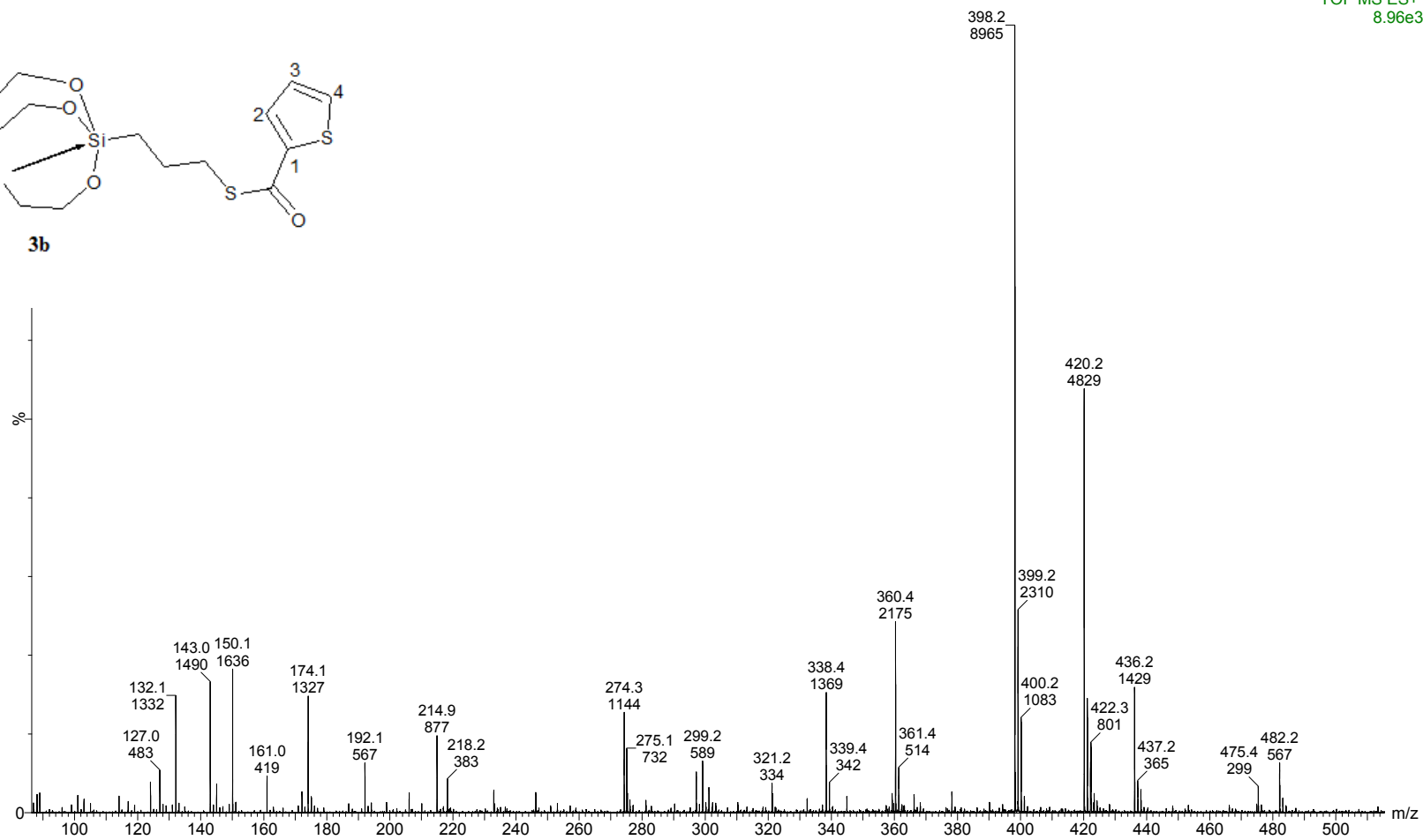
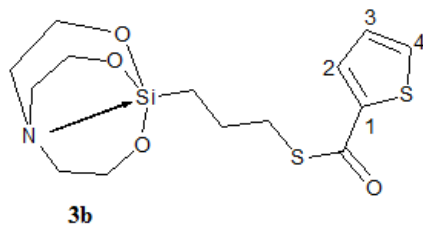


Fig. S32  $^1\text{H}$  NMR spectrum of **3b**



**Fig. S33**  $^{13}\text{C}$  NMR spectrum of **3b**



**Fig. S34**  $^1\text{H}$  NMR spectrum of **3b**



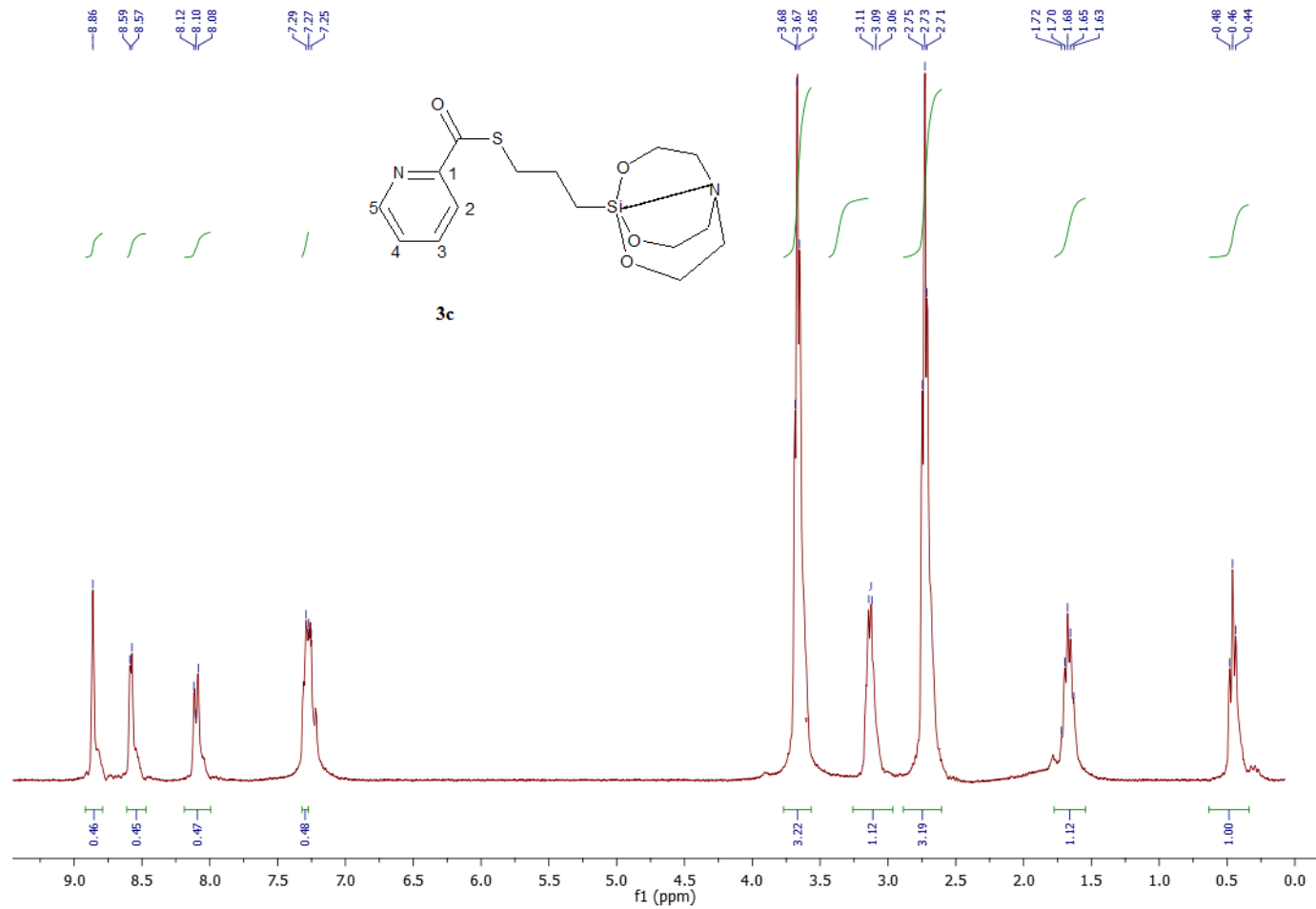


Fig. S35 <sup>1</sup>H NMR spectrum of **3c**

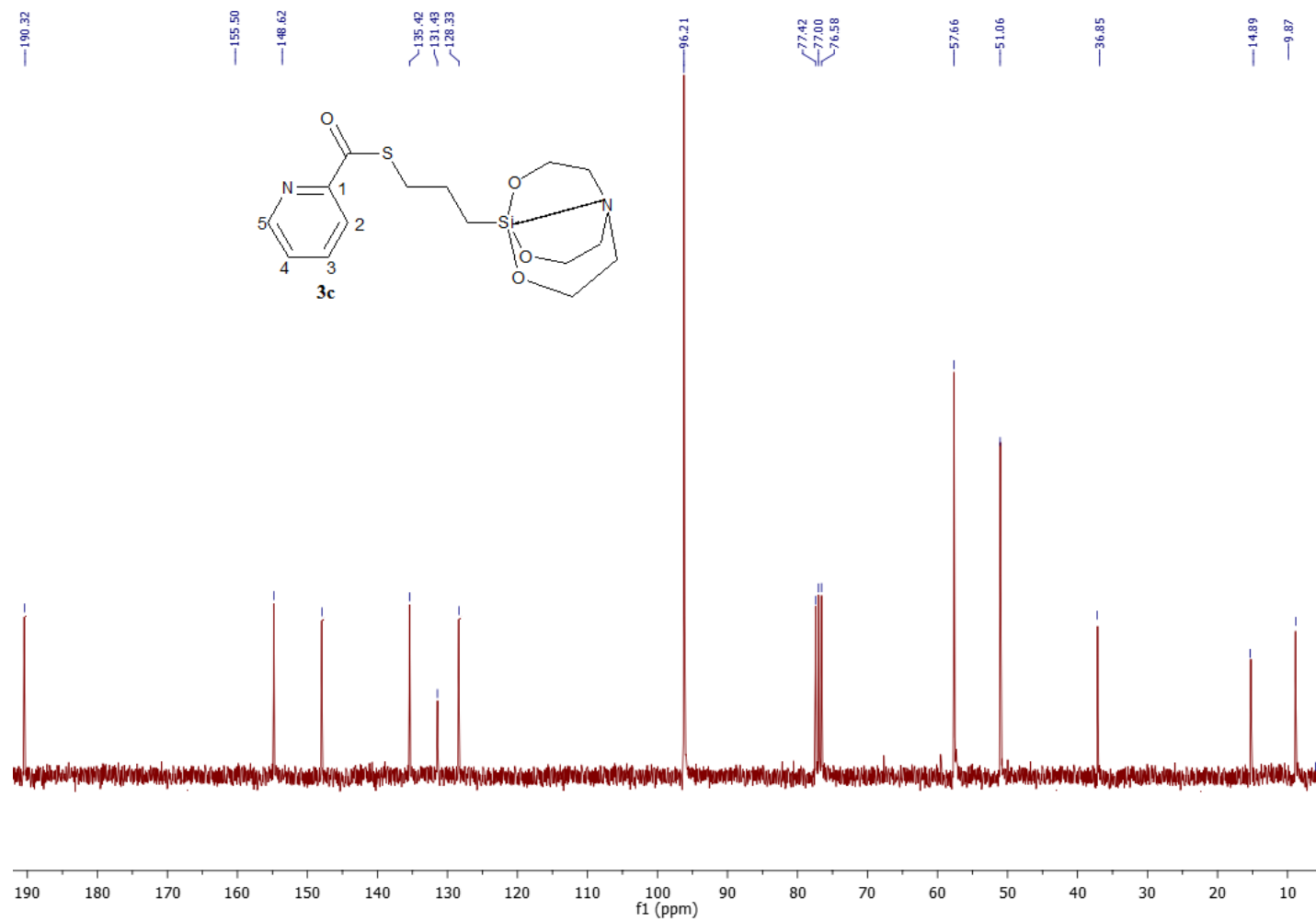


Fig. S36  $^{13}\text{C}$  NMR spectrum of **3c**

WATERS, Q-TOF MICROMASS (LC-MS)

SUNITA S-6 103 (1.154) Cm (53:118)

SAIF/CIL,PANJAB UNIVERSITY,CHANDIGARH

TOF MS ES+  
744

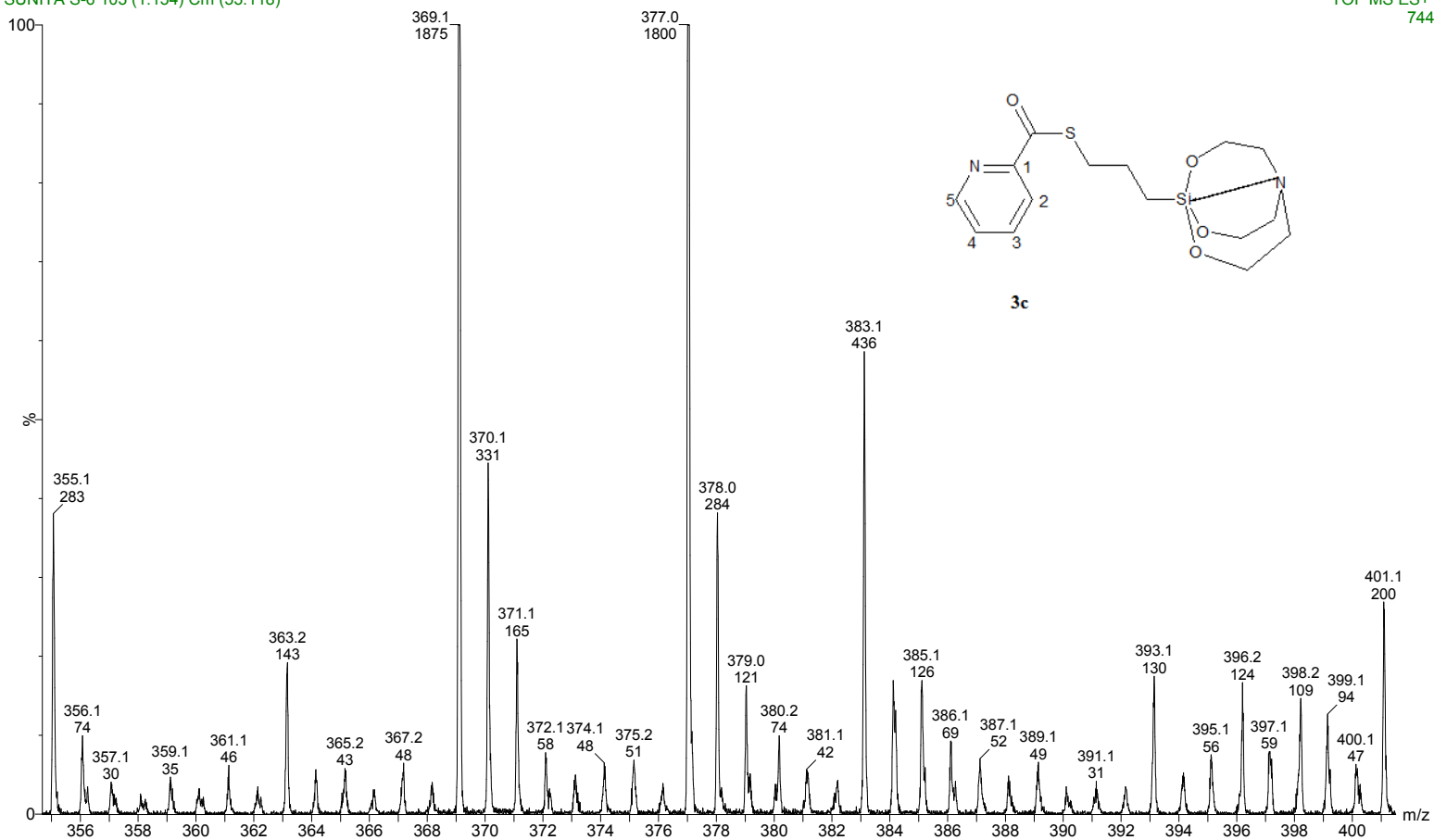


Fig. S37 Mass spectrum of 3c

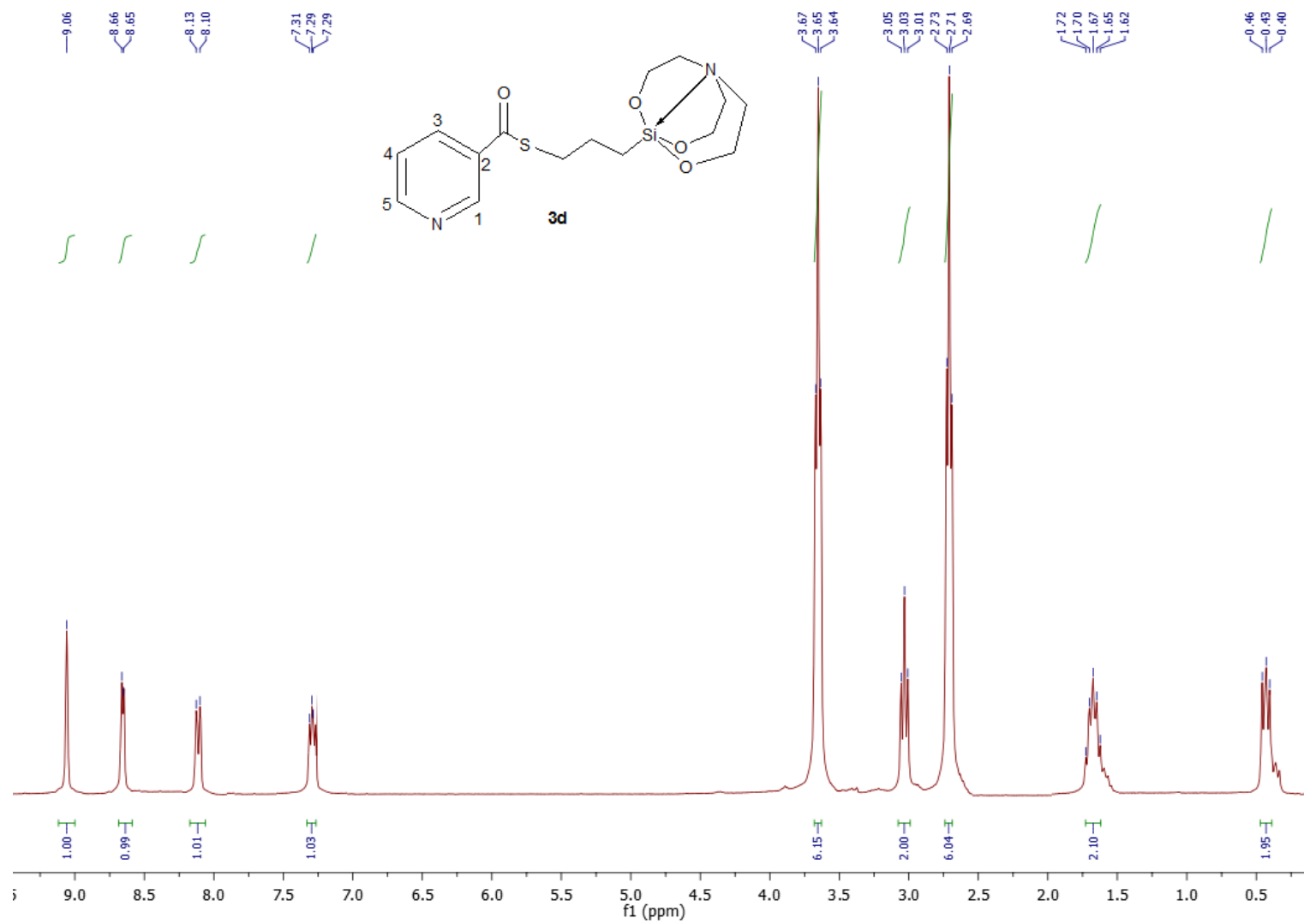


Fig. S38 <sup>1</sup>H NMR spectrum of **3d**

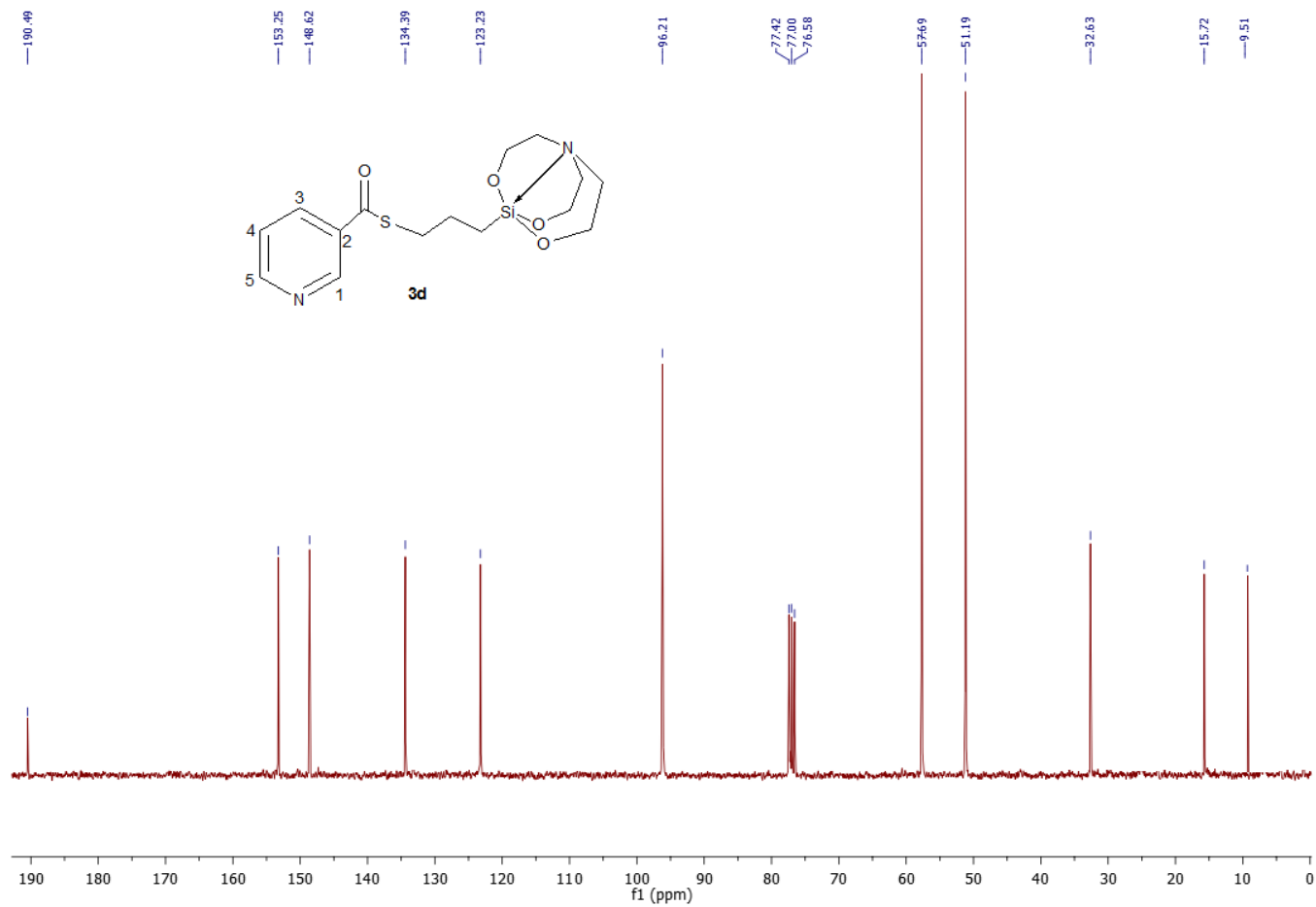


Fig. S39  $^1\text{H}$  NMR spectrum of **3d**

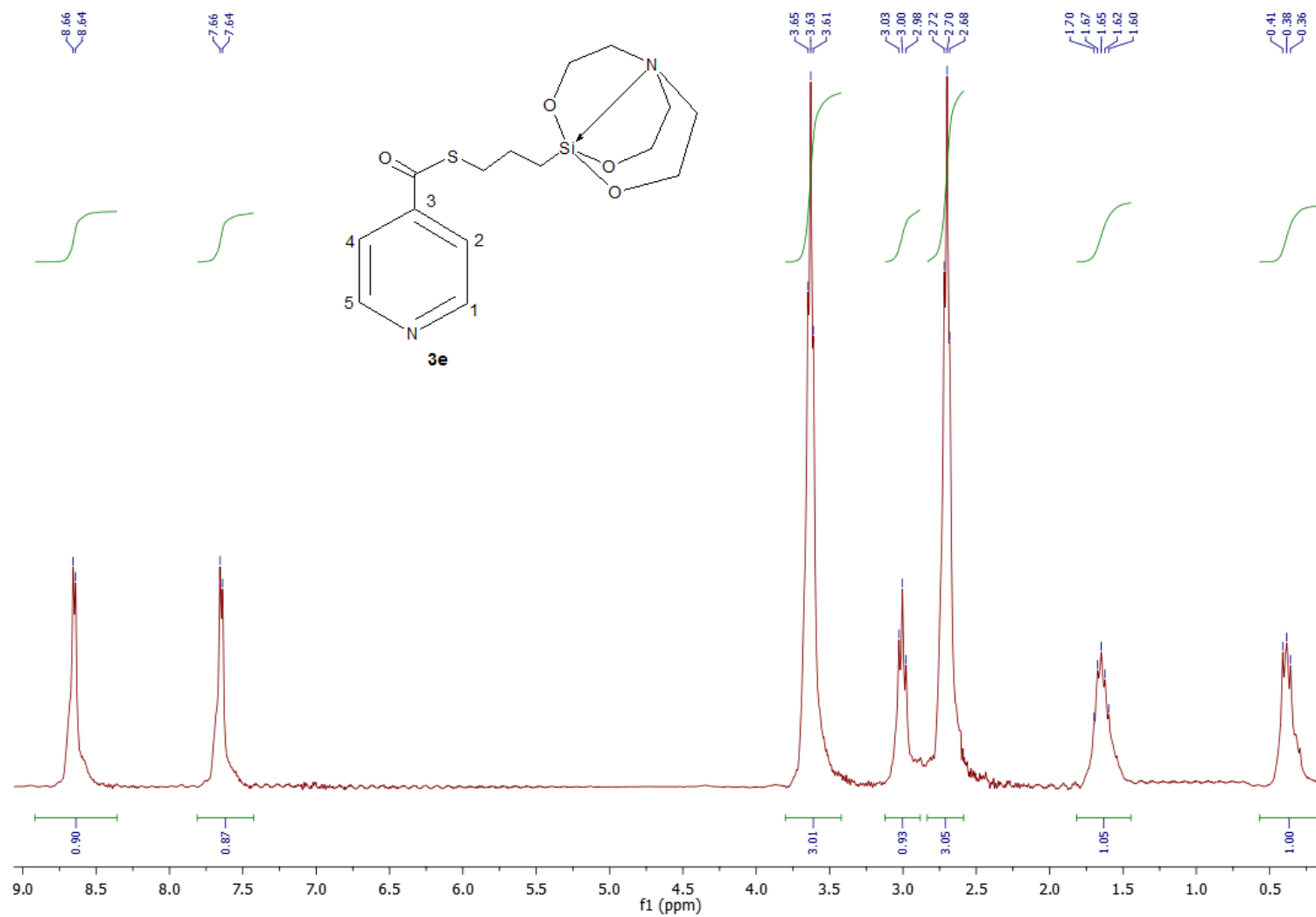


Fig. S40 <sup>1</sup>H NMR spectrum of **3e**

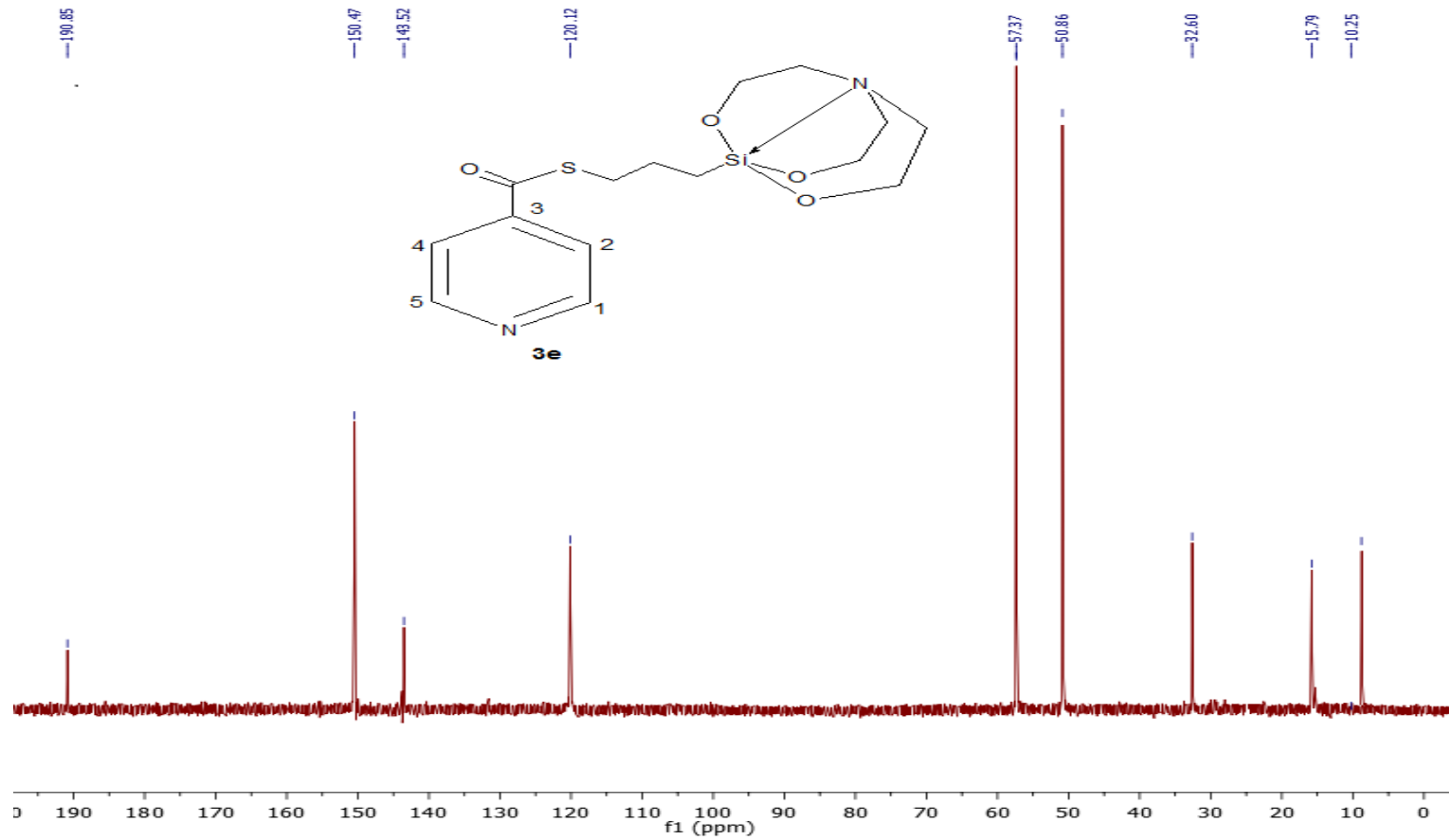
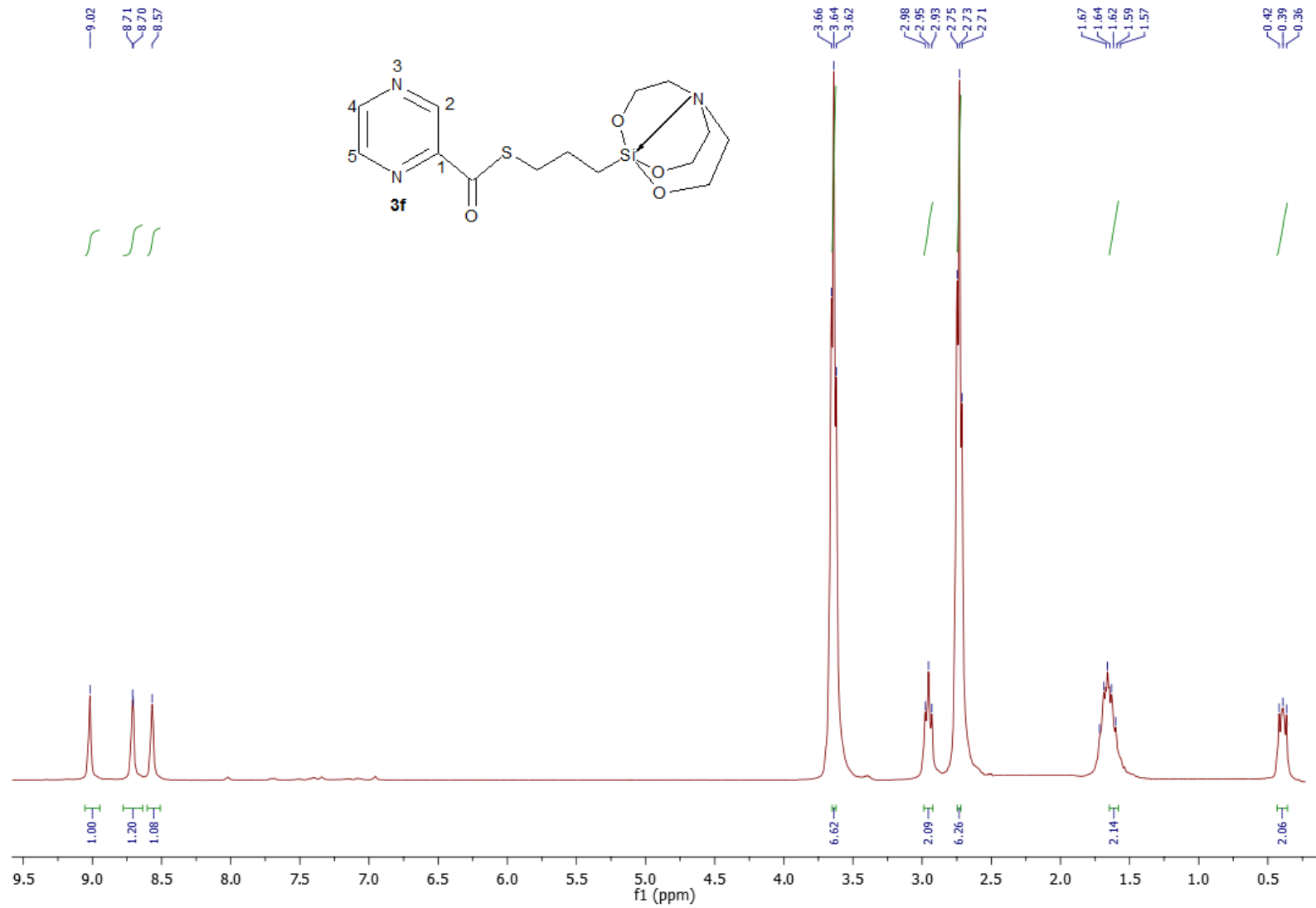


Fig. S41 <sup>13</sup>C NMR spectrum of **3e**



**Fig. S42**  $^1\text{H}$  NMR spectrum of **3f**



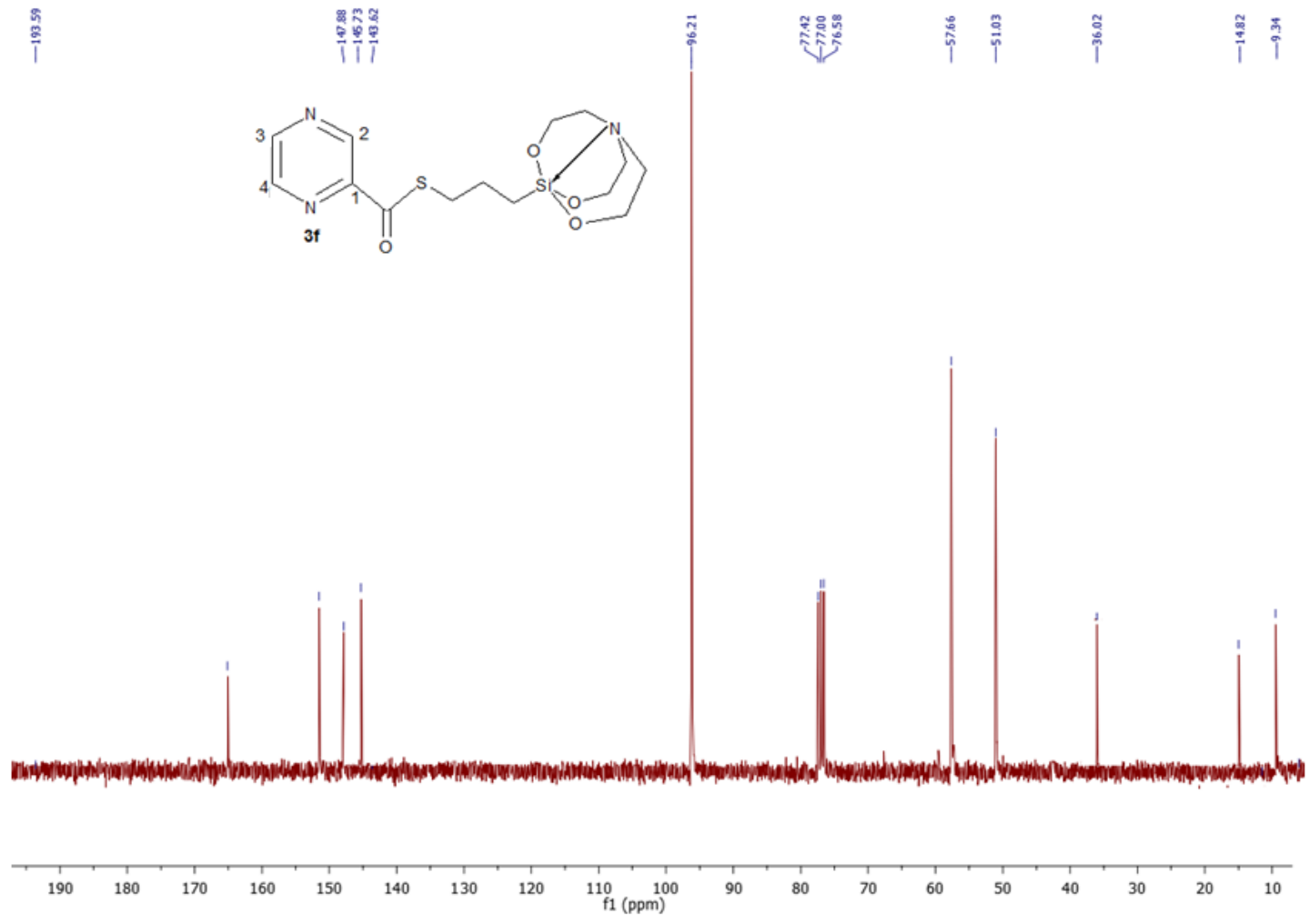


Fig. S43 <sup>13</sup>C NMR spectrum of **3f**

WATERS, Q-TOF MICROMASS (LC-MS)

SUNITA S-3 10 (0.112) Cm (8:28)

SAIF/CIL, PANJAB UNIVERSITY, CHANDIGARH

TOF MS ES+  
5.58e3

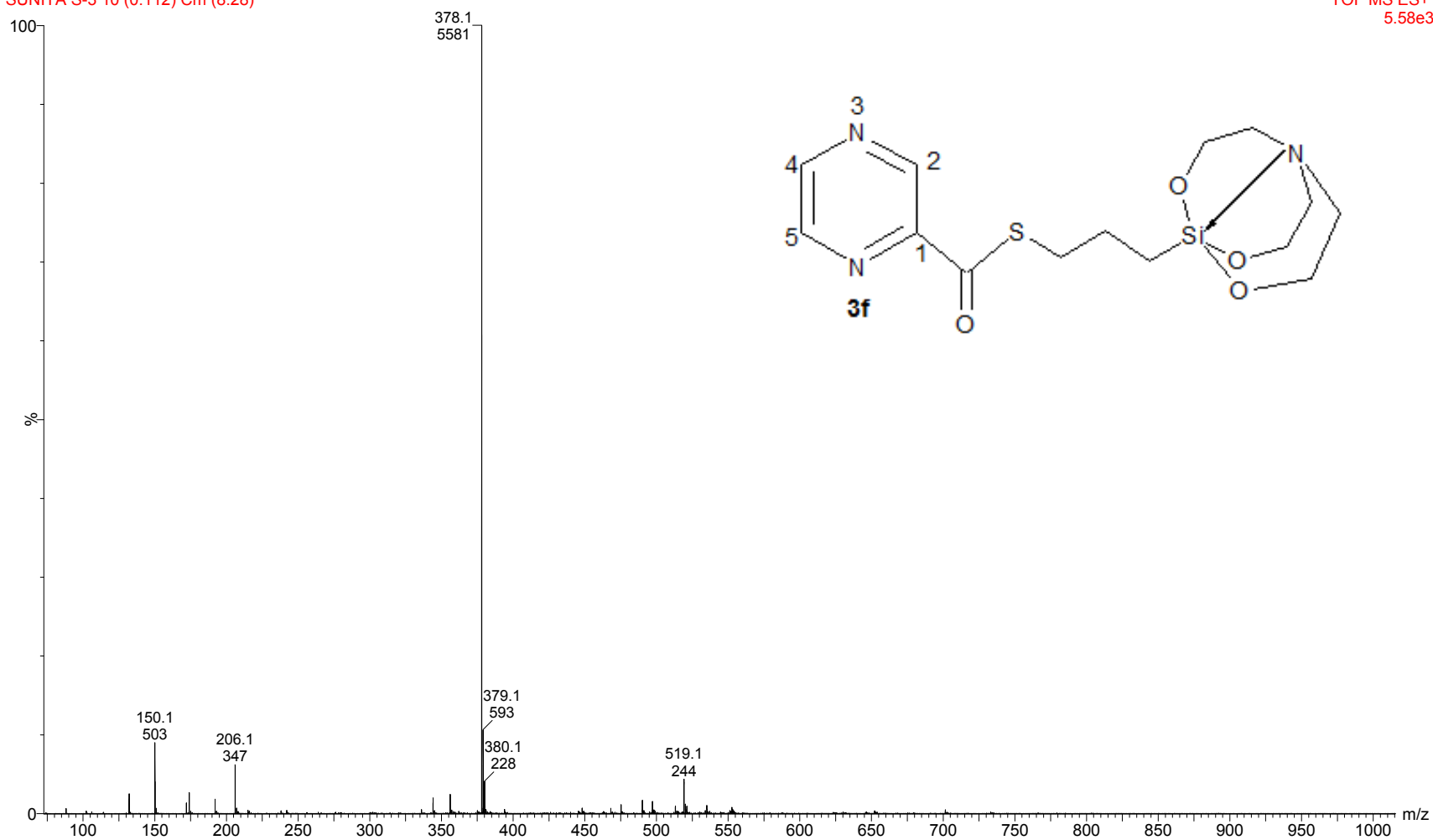


Fig. S44 Mass spectrum of 3f

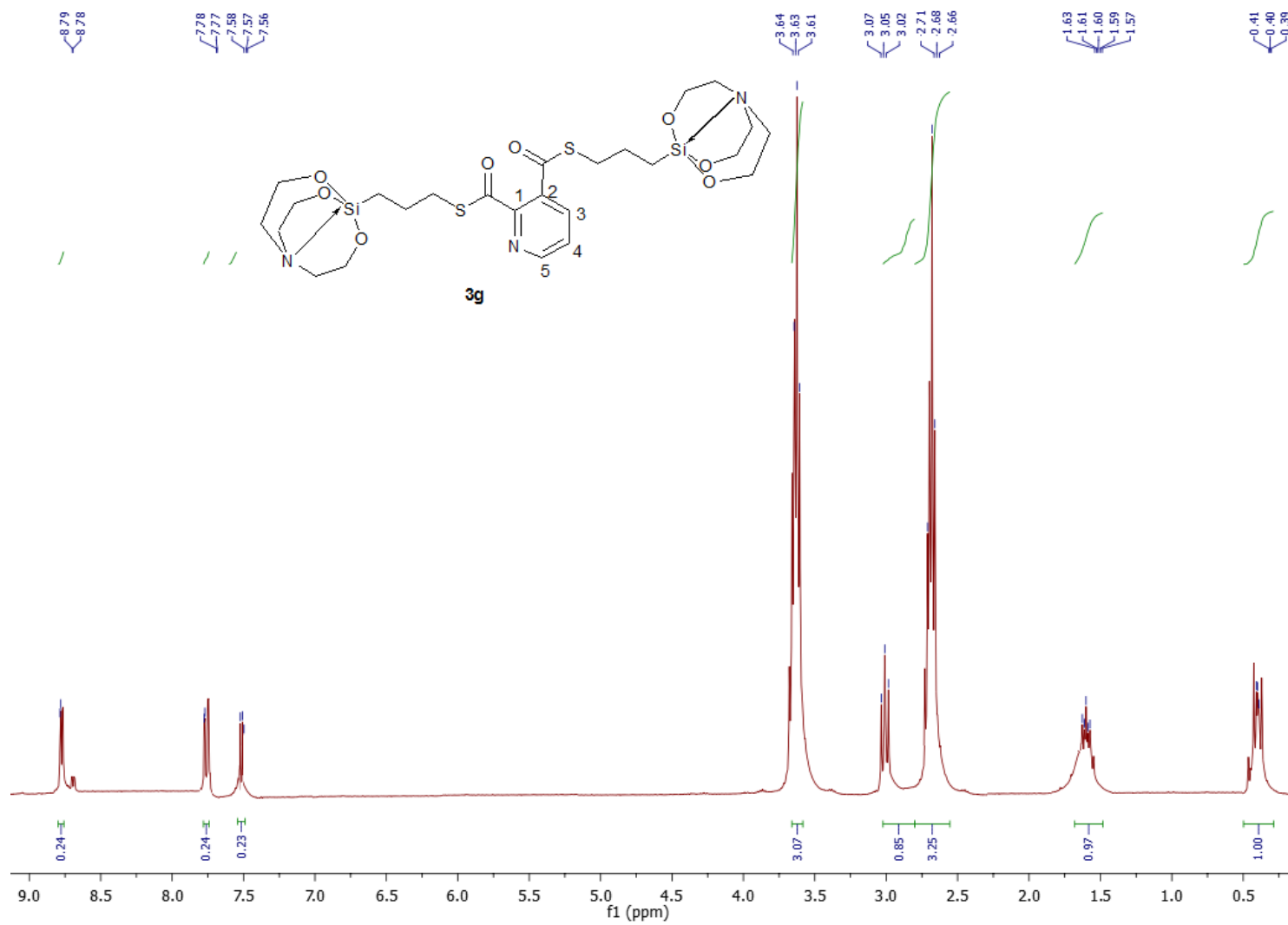


Fig. S45  $^1\text{H}$  NMR spectrum of **3g**

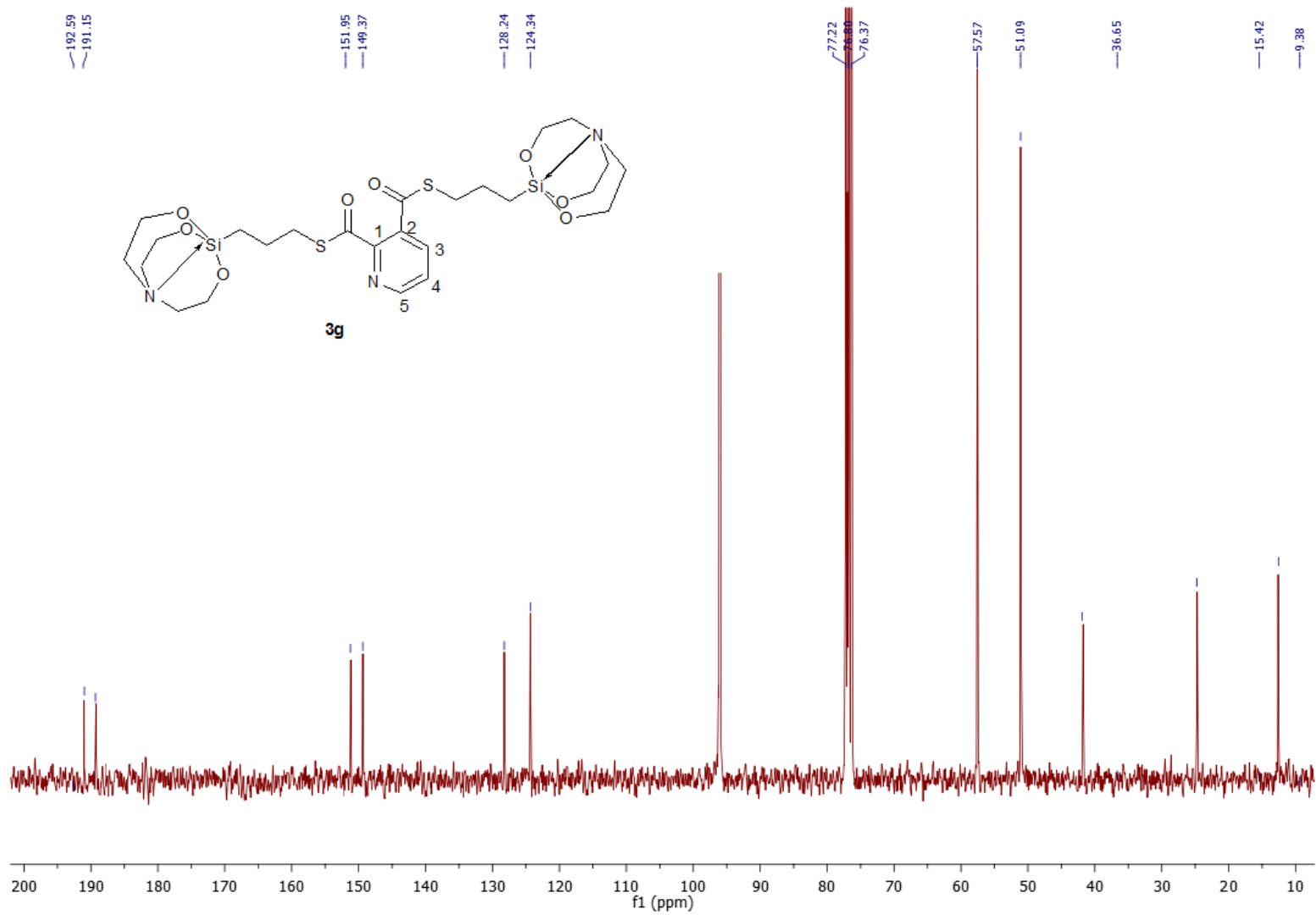
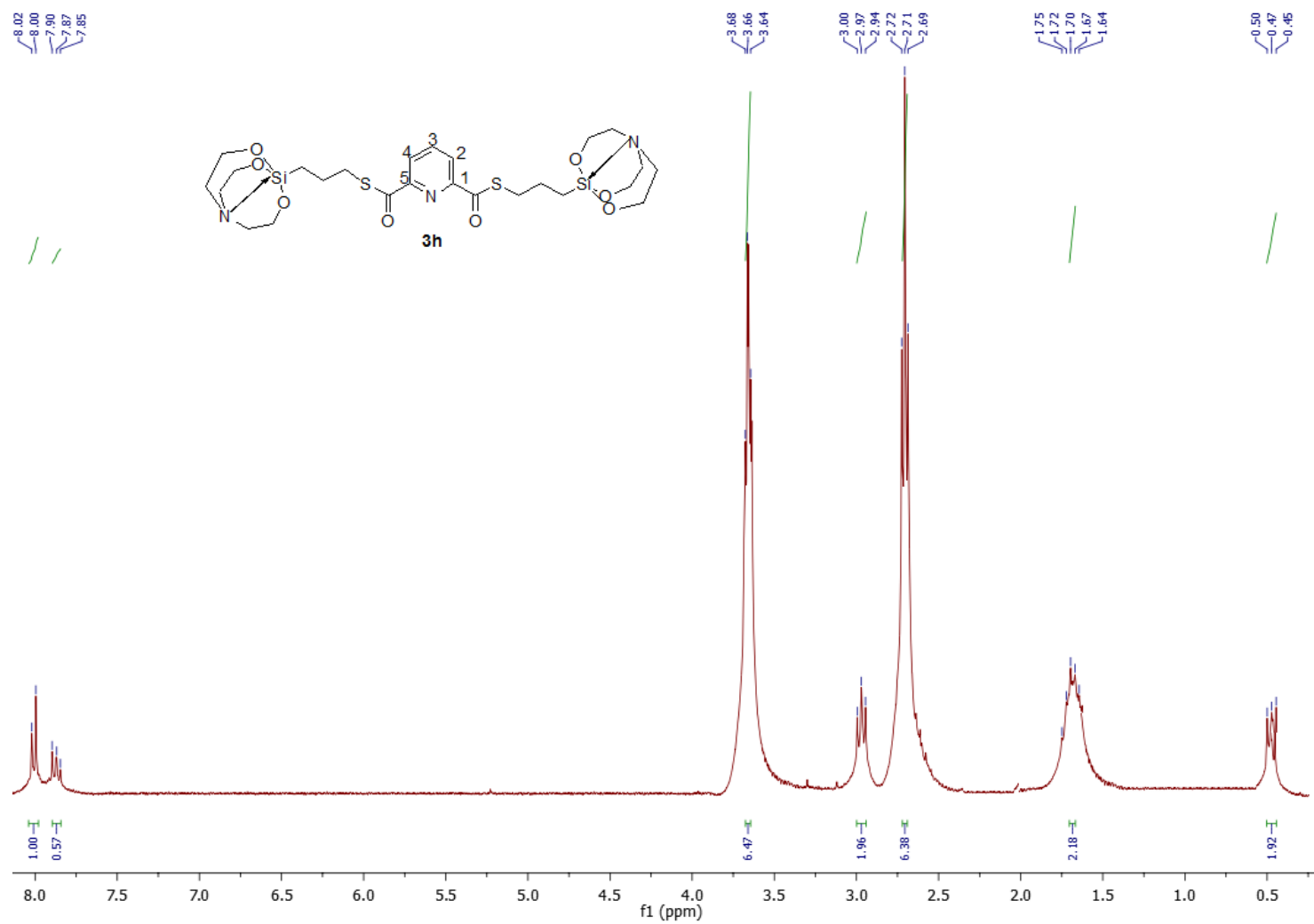


Fig. S46 <sup>13</sup>C NMR spectrum of 3g





**Fig. S48**  $^1\text{H}$  NMR spectrum of **3h**

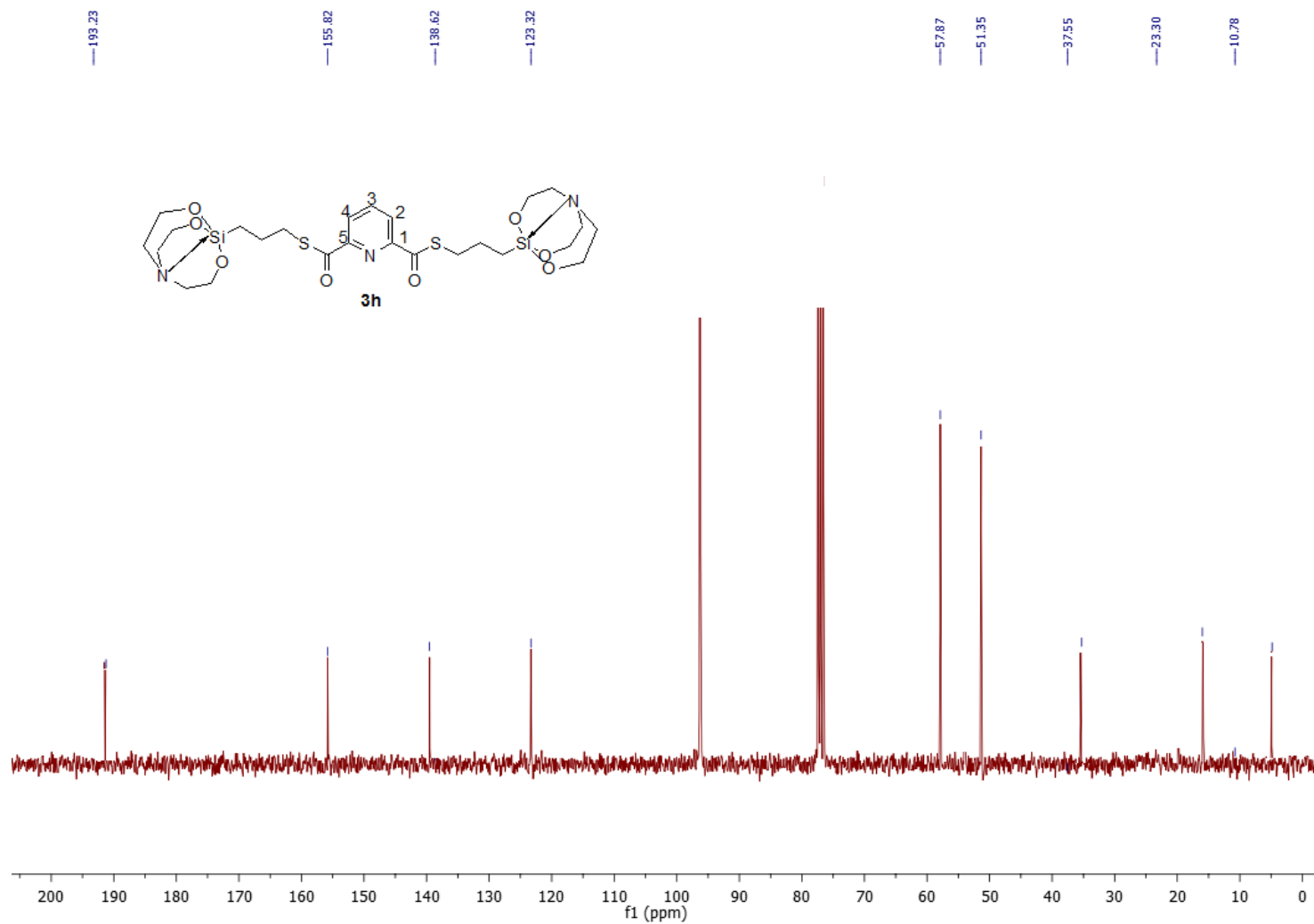


Fig. S49  $^{13}\text{C}$  NMR spectrum of **3h**