

Supporting Information:

A systematic understanding of gelation self-assembly: Solvophobically assisted supramolecular gelation via conformational reorientation across amide functionality on a hydrophobically modulated dipeptide based ambidextrous gelator, N-n-acyl-(L)Val-X(OBn), (X = 1,ω-amino acid)

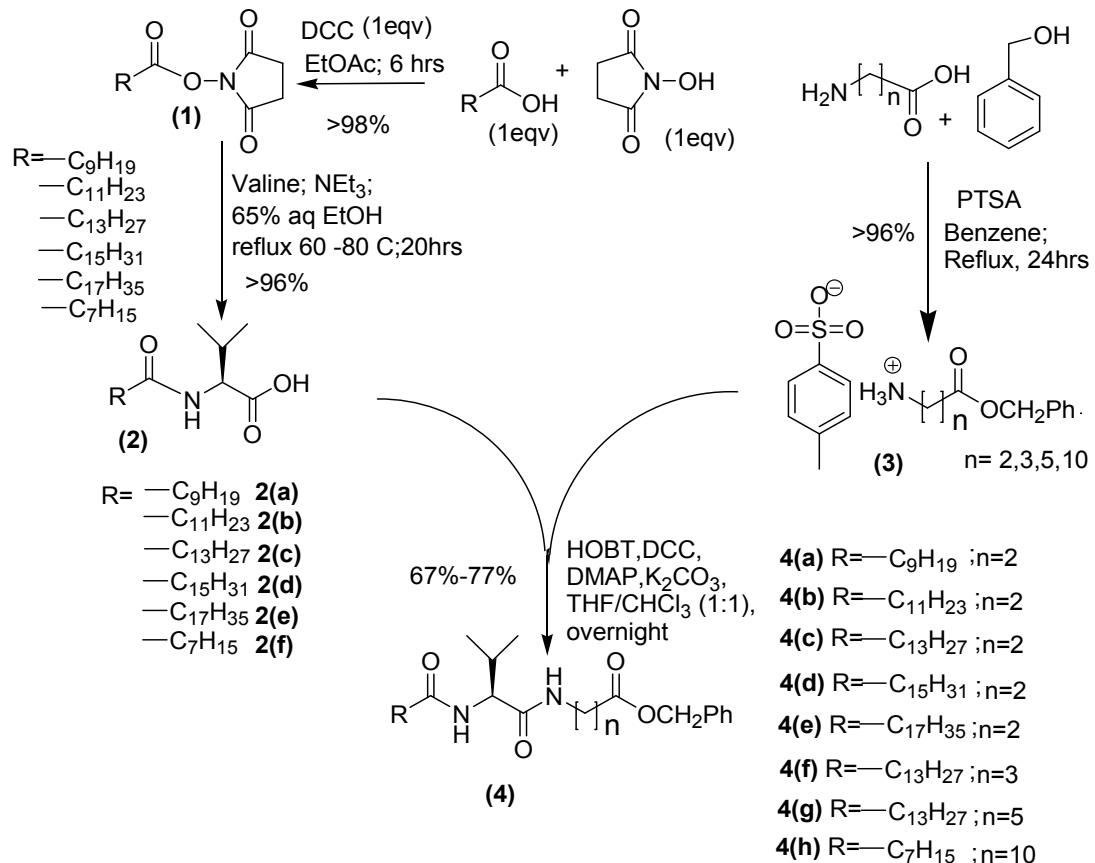
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Index

Scheme of Synthesis.....	S-ii
Synthetic procedure	S-iii-vi
¹HNMR spectrum	S-vi-ix
¹³C-NMR spectrum	S-x-xiii
Table: Gelation properties	S-xiv
Rheology	S-xv-xvi
Thermal stability	S-xvi-xvii
X-ray diffraction study	S-xviii-xxi
Solvent effect	S-xxii-xxiv
CD	S-xxv-xxvi
FTIR	S-xxvii-xxx

Scheme:



Synthetic procedure:

Synthesis of NHS ester of Fatty acids (1): To a solution of Fatty acid (1eqv) in 10 ml Ethyl acetate, N-hydroxysuccinimide (1eqv) was added followed by the addition of DCC (Dicyclohexylcarbodiimide) (1eqv) in 5 ml Ethyl acetate and the reaction stirred for 6-7 hrs. Completion of the reaction was confirmed by TLC. The reaction mixture was then filtered and the filtrate was concentrated under reduced pressure. The obtained residue was recrystallized from isopropanol and used for the next step without any further purification. (~98% yield).

Synthesis of N-Acyl-valine (2): To a solution of NHS ester of fatty acids (8.4 mmole, 1 eqv), (L)-Valine (1.2 eqv) in 40 ml of 65% aq Ethanol; Triethylamine (2.5 eqv) was added and put to reflux (60-80°C) for 20 hrs. Then the reaction mixture was concentrated under reduced pressure and taken up in 25ml Ethyl acetate and washed sequentially with ice cold 1 (N) HCl (30 ml × 2), saturated brine solution (25 ml × 2). Organic layer was collected and dried over anhyd. Na₂SO₄ and evaporated under reduced pressure to give the desired product (~8.06mmol; ~96% yield; m.p= ~95-98°C). The compounds were used for the next step without any further purification.

2a: ¹H NMR (300 MHz, CDCl₃) -δ:: 0.83-0.89 (t, CH₃ (CH₂)₈CONH-, 3H), 0.89-0.98 (q, (CH₃)₂CH-CH(COOH)NH-, 7H), 1.1-1.3 (bs, CH₃ (CH₂)₆CH₂CH₂CONH-, 12H), 1.56-1.7 (t, CH₃ (CH₂)₁₂CH₂CH₂CONH-, 2H), 2.2-2.35 (t, CH₃ (CH₂)₆CH₂CH₂CONH-, 2H), 4.55-4.64 (q, -NHCH(COOH)CH (CH₃)₂, 1H), 5.89-5.99 (d, -NH-, 1H).

2b: ¹H NMR (300 MHz, CDCl₃) -δ:: 0.88-0.92 (t, CH₃ (CH₂)₁₀CONH-, 3H), 0.89-1.0 (q, (CH₃)₂CH-CH(COOH)NH-, 7H), 1.2-1.44 (bs, CH₃ (CH₂)₈CH₂CH₂CONH-, 16H), 1.56-1.7 (t, CH₃ (CH₂)₁₂CH₂CH₂CONH-, 2H), 2.21-2.35 (t, CH₃ (CH₂)₈CH₂CH₂CONH-, 2H), 4.55-4.61 (q, -NHCH(COOH)CH (CH₃)₂, 1H), 5.89-5.99 (d, -NH-, 1H).

2c: ¹H NMR (300 MHz, CDCl₃) -δ:: 0.86-0.1 (t, CH₃ (CH₂)₁₂CONH-, 3H), 0.89-1.0 (q, (CH₃)₂CH-CH(COOH)NH-, 7H), 1.2-1.45 (bs, CH₃(CH₂)₁₀CH₂CH₂CONH-, 20H), 1.56-1.7 (t, CH₃ (CH₂)₁₂CH₂CH₂CONH-, 2H), 2.23-2.35 (t, CH₃ (CH₂)₁₀CH₂CH₂CONH-, 2H), 4.55-4.621 (q, -NHCH(COOH)CH (CH₃)₂, 1H), 5.89-5.99 (d, -NH-, 1H).

2d: ¹H NMR (300 MHz, CDCl₃) -δ:: 0.89-0.1 (t, CH₃ (CH₂)₁₄CONH-, 3H), 0.89-1.0 (q, (CH₃)₂CH-CH(COOH)NH-, 7H), 1.25-1.44 (bs, CH₃(CH₂)₁₂CH₂CH₂CONH-, 24H), 1.56-1.7 (t, CH₃ (CH₂)₁₂CH₂CH₂CONH-, 2H), 2.22-2.35 (t, CH₃ (CH₂)₁₂CH₂CH₂CONH-, 2H), 4.55-4.61 (q, -NHCH(COOH)CH (CH₃)₂, 1H), 5.89-5.99 (d, -NH-, 1H).

2e: ¹H NMR (300 MHz, CDCl₃) -δ:: 0.91-1.10 (t, CH₃ (CH₂)₁₆CONH-, 3H), 0.89-1.0 (q, (CH₃)₂CH-CH(COOH)NH-, 7H), 1.2-1.46 (bs, CH₃(CH₂)₁₄CH₂CH₂CONH-, 28H), 1.56-1.7 (t, CH₃ (CH₂)₁₂CH₂CH₂CONH-, 2H), 2.23-2.35 (t, CH₃ (CH₂)₁₄CH₂CH₂CONH-, 2H), 4.55-4.611 (q, -NHCH(COOH)CH (CH₃)₂, 1H), 5.89-5.99 (d, -NH-, 1H).

Synthesis of p-toluenesulfonates salt of 1-ω-Aminoacid benzylester (3): In 100ml of benzene a mixture of l,ω-Aminoacid(1 eqv), p-toluenesulfonic acid (PTSA).H₂O (1.1 eqv) and benzyl alcohol (2 eqv) was taken. After refluxing in Dean-Stark apparatus for 24hrs, 40 ml of diethyl ether was added to it with immediate precipitation of PTSA-salt of 1-ω-Aminoacid benzyl ester. The salt was then filtered, washed

with 10ml of dry diethyl ether and dried under reduced pressure. This compound was used for the next step without any further purification.(yield~96 %)

Synthesis of (4): To a solution of N-Acyl-(L)-valine (1 eqv) in 5ml of THF, Hydroxybenzotriazole (HOBt) (1.5 eqv) was added and kept on stirring in ice water bath at 0°C. After 10mins of stirring, DMAP (1 eqv) and a solution of DCC (1.93 eqv) in 5 ml of CHCl₃ was added to it and kept stirring. Finally, after 15mins, K₂CO₃ (1.47 eqv) and PTSA-salt of 1- ω -Amino acid benzyl ester (1.2 eqv) was added and kept on stirring overnight at room temperture. The reaction mixture was then concentrated under reduced pressure and taken up in 30 ml CHCl₃ and washed sequentially with ice cold 1 (N) HCl (25 ml \times 2), saturated brine (25 ml \times 2). The organic layer was collected and dried over anhyd Na₂SO₄ and then purified by silicagel (60-120 mesh) column chromatography in EtOAc/Petroleum ether solvent system of required ratios (~40% -60%) to get the desired product.

4a. Yield% = 77%. ¹H NMR (300 MHz, CDCl₃) : δ_H :: 0.875-0.918 (t, -NHCOCH₂CH₂(CH₂)₆CH₃ and >CHCH(CH₃)₂, 9H), 1.27 (bs, -NHCOCH₂CH₂(CH₂)₆CH₃, 12H), 1.58-1.65 (m, -NHCOCH₂CH₂(CH₂)₆CH₃, >CHCH(CH₃)₂, 3H), 2.11-2.24 (t, -NHCOCH₂CH₂(CH₂)₆CH₃, 2H), 2.59-2.62 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.5-3.6 (m, CONHCH₂CH₂COOCH₂Ph, 2H), 4.17-4.2 (t, >CHCH(CH₃)₂, 1H), 5.09-5.1 (s, -CONHCH₂CH₂COOCH₂Ph, 2H), 6.11-6.13 (d, -NHCOCH₂CH₂(CH₂)₆CH₃, 1H), 6.52 (s, -CONHCH₂CH₂COOCH₂Ph, 1H), 7.26-7.35 (bs, aromatic, 5H). ¹³C NMR (75 MHz, CDCl₃) : δ_C : 173.42, 172.56, 171.41, 135.58, 128.62, 128.4, 128.2, 77.46, 77.04, 76.61, 66.54, 58.35, 36.73, 36.67, 35.00, 33.96, 31.84, 31.19, 29.43, 29.32, 29.24, 25.74, 22.64, 19.12, 18.24, 14.07. ESI-MS (m/z) C₂₅H₄₀N₂O₄ (EXACT MASS= 432.3) 433.3(100%, [M + H]⁺), 455.3(49%, [M + Na]⁺).

4b. Yield=73%. ¹H NMR (300 MHz, CDCl₃) : δ_H :: 0.87-0.91(t, -NHCOCH₂CH₂(CH₂)₈CH₃ and >CHCH(CH₃)₂, 9H), 1.269 (bs, -NHCOCH₂CH₂(CH₂)₈CH₃, 16H), 1.63-1.74 (m, -NHCOCH₂CH₂(CH₂)₈CH₃, >CHCH(CH₃)₂, 3H), 2.11-2.2 (t, -NHCOCH₂CH₂(CH₂)₈CH₃, 2H), 2.58-2.62 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.5-3.63 (m, CONHCH₂CH₂COOCH₂Ph, 2H), 4.17-4.22 (t, >CHCH(CH₃)₂, 1H), 5.12 (s, -CONHCH₂CH₂COOCH₂Ph, 2H), 6.10-6.13 (d, -NHCOCH₂CH₂(CH₂)₈CH₃, 1H), 6.51(s, -CONHCH₂CH₂COOCH₂Ph, 1H), 7.37 (bs, aromatic, 5H). ¹³C NMR (75 MHz, CDCl₃) : δ_C : 173.2, 171.97, 171.35, 135.58, 128.62, 128.4, 128.3, 77.46, 77.04, 76.61, 66.61, 58.28, 36.73, 34.9, 33.9, 31.8, 31.2, 29.6, 29.4, 29.32, 29.2, 25.74, 22.6, 19.12, 18.2, 14.09. ESI-MS (m/z) C₂₇H₄₄N₂O₄ (EXACT MASS= 460.33) 461.34(100%, [M + H]⁺), 483.32(34%, [M + Na]⁺).

4c. Yield=76%. ¹H NMR (300 MHz, CDCl₃) : δ_H :: 1.1-1.25 (t, -NHCOCH₂CH₂(CH₂)₁₀CH₃ and >CHCH(CH₃)₂, 9H), 1.57 (bs, -NHCOCH₂CH₂(CH₂)₁₀CH₃, 20H), 1.87-2.03 (m, -NHCOCH₂CH₂(CH₂)₁₀CH₃, >CHCH(CH₃)₂, 3H), 2.3-2.4 (t, -NHCOCH₂CH₂(CH₂)₆CH₃, 2H), 2.79-2.81 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.49 (s, CONHCH₂CH₂COOCH₂Ph, 2H), 4.19-4.32 (t, >CHCH(CH₃)₂, 1H), 5.12 (s, -CONHCH₂CH₂COOCH₂Ph, 2H), 6.12-6.13 (t, -NHCOCH₂CH₂(CH₂)₁₀CH₃, 1H), 6.63-6.651(t, -CONHCH₂CH₂COOCH₂Ph, 1H), 7.29-7.37 (m, aromatic, 5H). ¹³C NMR (75 MHz, CDCl₃) : δ_C : 173.27, 172.93, 171.407, 135.6, 128.62, 128.4, 128.28, 77.46, 77.041, 76.61, 66.59, 58.256, 36.7, 34.976, 33.99, 33.69, 31.9, 31.2, 29.63, 29.49, 29.33, 29.28, 25.75, 25.54, 25.544, 24.865, 24.77, 22.67, 19.13, 18.22, 14.09. ESI-MS (m/z) C₂₉H₄₈N₂O₄ (EXACT MASS= 488.36) 489.369(100%, [M + H]⁺), 511.36(51%, [M + Na]⁺).

4d. Yield=72%. ¹H NMR (300 MHz, CDCl₃) : δ_H :: 0.875-0.918 (m, -NHCOCH₂CH₂(CH₂)₁₂CH₃ and >CHCH(CH₃)₂, 9H), 1.27 (bs, -NHCOCH₂CH₂(CH₂)₁₂CH₃, 24H), 1.58-1.65 (m, -NHCOCH₂CH₂(CH₂)₁₂CH₃, >CHCH(CH₃)₂, 3H), 2.11-2.24 (t, -NHCOCH₂CH₂(CH₂)₁₂CH₃, 2H), 2.59-2.62 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.5-3.63 (m, CONHCH₂CH₂COOCH₂Ph, 2H), 4.17-4.22 (t, >CHCH(CH₃)₂, 1H), 5.12 (s, -CONHCH₂CH₂COOCH₂Ph, 2H), 6.10-6.13 (d, -NHCOCH₂CH₂(CH₂)₁₂CH₃, 1H), 6.51(s, -CONHCH₂CH₂COOCH₂Ph, 1H), 7.37 (bs, aromatic, 5H). ¹³C NMR (75 MHz, CDCl₃) : δ_C : 173.2, 171.97, 171.35, 135.58, 128.62, 128.4, 128.3, 77.46, 77.04, 76.61, 66.61, 58.28, 36.73, 34.9, 33.9, 31.8, 31.2, 29.6, 29.4, 29.32, 29.2, 25.74, 22.6, 19.12, 18.2, 14.09. ESI-MS (m/z) C₃₁H₅₆N₂O₄ (EXACT MASS= 520.46) 521.46(100%, [M + H]⁺), 543.46(49%, [M + Na]⁺).

$(CH_2)_{12}CH_3 > CHCH$ ($CH_3)_2$, 3H), 2.11-2.24 (t, -NHCOCH₂CH₂(CH₂)₁₂CH₃, 2H), 2.59-2.62 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.5-3.6 (m, CONHCH₂CH₂COOCH₂Ph, 2H), 4.17-4.2 (t, >CHCH(CH₃)₂, 1H), 5.09-5.1 (s, -CONHCH₂CH₂COOCH₂Ph, 2H), 6.11-6.13 (d, -NHCOCH₂CH₂(CH₂)₁₂CH₃, 1H), 6.52 (s, -CONHCH₂CH₂COOCH₂Ph, 1H), 7.26-7.35 (bs, aromatic, 5H). ¹³C NMR(75 MHz, CDCl₃) : δ_C : 173.6, 172.4, 171.8, 135.64, 128.6, 128.34, 128.18, 77.47, 77.05, 76.62, 66.53, 57.53, 41.35, 36.53, 35.02, 33.9, 33.47, 31.9, 29.67, 29.64, 29.61, 29.5, 29.33, 29.26, 25.64, 24.81, 22.82, 22.66, 22.20, 14.08. ESI-MS (m/z) C₃₁H₅₂N₂O₄ (EXACT MASS=516.37) 517.379 (100%, [M + H]⁺), 539.37 (20%, [M + Na]⁺).

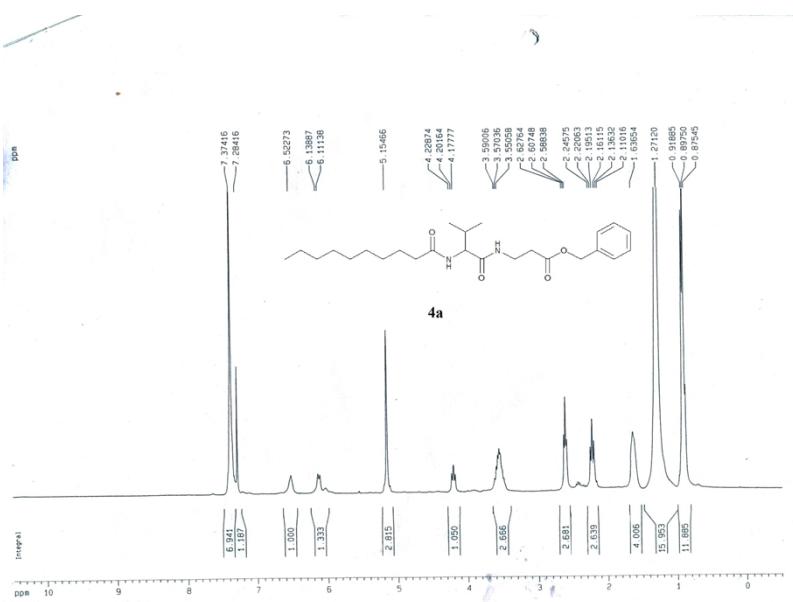
4e. Yield=69%. ¹HNMR (300 MHz, CDCl₃) : δ :: 0.85-0.89(m, -NHCOCH₂CH₂(CH₂)₁₄CH₃ and >CHCH(CH₃)₂, 9H), 1.24 (bs, -NHCOCH₂CH₂(CH₂)₁₄CH₃, 28H), 1.6 (s, -NHCOCH₂CH₂(CH₂)₁₄CH₃, >CHCH(CH₃)₂, 3H), 2.14-2.19 (t, -NHCOCH₂CH₂(CH₂)₁₄CH₃, 2H), 2.56-2.60 (t, -CONHCH₂CH₂COOCH₂Ph, 2H), 3.5-3.54 (t, CONHCH₂CH₂COOCH₂Ph, 2H), 4.35-4.42 mt, >CHCH(CH₃)₂, 1H), 5.13 (s, -CONHCH₂CH₂COOCH₂Ph, 2H), 6.11-6.13 (d, -NHCOCH₂CH₂(CH₂)₁₄CH₃, 1H), 6.62 (s, -CONHCH₂CH₂COOCH₂Ph, 1H), 7.35 (bs, aromatic, 5H). ¹³C NMR(75 MHz, CDCl₃) : δ_C : 173.24, 172.56, 135.62, 128.62, 128.4, 128.3, 77.44, 77.21, 76.6, 66.68, 58.35, 36.75, 34.97, 34.8, 34.13, 33.39, 31.9, 31.17, 29.68, 29.65, 29.61, 29.48, 29.34, 29.26, 25.73, 25.68, 22.68, 19.1, 18.2, 14.1. ESI-MS (m/z) C₃₃H₅₆N₂O₄ (EXACT MASS=544.42) 545.429(100%, [M + H]⁺), 567.4(49%, [M + Na]⁺).

4f. Yield=69%. ¹HNMR (300 MHz, CDCl₃) : δ :: 0.85-0.91 (m, -NHCOCH₂CH₂(CH₂)₁₀CH₃ and >CHCH(CH₃)₂, 9H), 1.4 (bs, -NHCOCH₂CH₂(CH₂)₁₀CH₃, 20H), 1.58-1.65 (m, -NHCOCH₂CH₂(CH₂)₁₀CH₃, >CHCH(CH₃)₂, 3H), 1.93-1.94 (t, -CONHCH₂CH₂CH₂COOCH₂Ph, 2H), 2.08-2.17 (t, -NHCOCH₂CH₂(CH₂)₁₀CH₃, 2H), 2.55-2.59 (t, -CONHCH₂CH₂CH₂COOCH₂Ph, 2H), 3.44-3.59 (m, -CONHCH₂CH₂CH₂COOCH₂Ph, 2H), 4.37-4.39 (t, >CHCH(CH₃)₂, 1H), 5.132 (bs, -CONHCH₂CH₂CH₂COOCH₂Ph, 2H), 5.89 (s, -NHCOCH₂CH₂(CH₂)₁₀CH₃, 1H), 6.58 (s, -CONHCH₂CH₂CH₂COOCH₂Ph, 1H), 7.35 (bs, aromatic, 5H). ¹³C NMR(75 MHz, CDCl₃) : δ_C : 173.45, 173.26, 171.39, 135.75, 128.54, 128.32, 128.22, 77.45, 77.23, 77.03, 76.61, 66.44, 58.36, 38.92, 36.78, 36.70, 31.92, 31.76, 31.62, 30.95, 29.49, 29.35, 25.76, 24.61, 24.47, 22.68, 19.26, 14.11. ESI-MS (m/z) C₂₇H₄₄N₂O₄ (EXACT MASS= 502.38) 503.389 (100%, [M + H]⁺), 525.38 (68%, [M + Na]⁺), 541.3 (28%, [M + K]⁺).

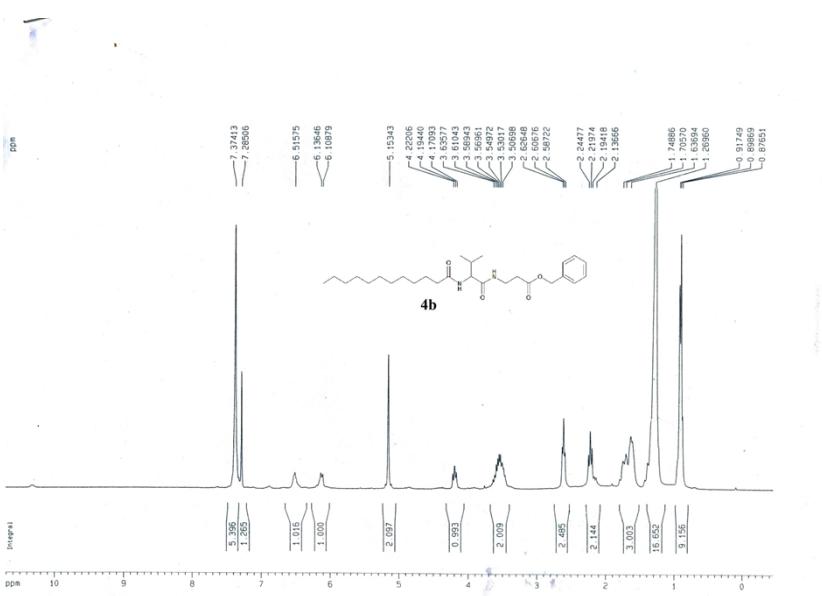
4g. Yield=69%. ¹HNMR (300 MHz, CDCl₃) : δ :: 0.84-0.85 (m, -NHCOCH₂CH₂(CH₂)₁₀CH₃ and >CHCH(CH₃)₂, 9H), 1.25 (bs, -NHCOCH₂CH₂(CH₂)₁₀CH₃, -CONHCH₂CH₂(CH₂)₂CH₂COOCH₂Ph, 24H), 1.6 (s, -NHCOCH₂CH₂(CH₂)₁₀CH₃, >CHCH(CH₃)₂, 3H), 1.780-1.788 (d, -CONHCH₂CH₂(CH₂)₂CH₂COOCH₂Ph, 2H), 2.08-2.2 (t, -NHCOCH₂CH₂(CH₂)₁₀CH₃, 2H), 2.56-2.60 (t, -CONHCH₂CH₂(CH₂)₂CH₂COOCH₂Ph, 2H), 3.44-3.58 (m, -CONHCH₂CH₂(CH₂)₂CH₂COOCH₂Ph, 2H), 4.17-4.22 (t, >CHCH(CH₃)₂, 1H), 5.09-5.13 (s, -CONH(CH₂)₅COOCH₂Ph, 2H), 6.01-6.04 (d, -NHCOCH₂CH₂(CH₂)₁₀CH₃, 1H), 6.35 (s, -CONH(CH₂)₅COOCH₂Ph, 1H), 7.35 (bs, aromatic, 5H). ¹³C NMR(75 MHz, CDCl₃) : δ_C : 173.27, 172.93, 171.407, 135.58, 128.62, 128.4, 128.28, 77.46, 77.041, 76.61, 66.59, 58.256, 40.2, 36.73, 36.67, 35.00, 33.1, 31.84, 31.19, 29.63, 29.49, 29.33, 29.28, 25.74, 25.54, 22.64, 18.212, 17.124, 14.102. ESI-MS (m/z) C₂₇H₄₄N₂O₄ (EXACT MASS= 530.41) 531.419(100%, [M + H]⁺), 553.421 (62%, [M + Na]⁺).

4h. Yield=69%. ¹HNMR (300 MHz, CDCl₃) : δ :: 0.846-0.934 (m, -NHCOCH₂CH₂(CH₂)₄CH₃ and >CHCH(CH₃)₂, 9H), 1.233-1.280 (m, -NHCOCH₂CH₂(CH₂)₄CH₃, -CONH(CH₂)₂(CH₂)₇CH₂COOCH₂Ph, 22H), 1.476-1.495 (d, -NHCOCH₂CH₂(CH₂)₄CH₃, -CONHCH₂CH₂(CH₂)₆CH₂CH₂COOCH₂Ph, 4H), 1.970-2.0 (m, >CHCH(CH₃)₂, 1H), 2.130-2.243 (m, -CONHCH₂CH₂(CH₂)₇CH₂COOCH₂Ph, 2H), 2.3-2.36 (m, -NHCOCH₂CH₂(CH₂)₄CH₃, 2H), 3.2-3.266 (m, -CONHCH₂(CH₂)₉COOCH₂Ph, 2H), 4.084-4.25

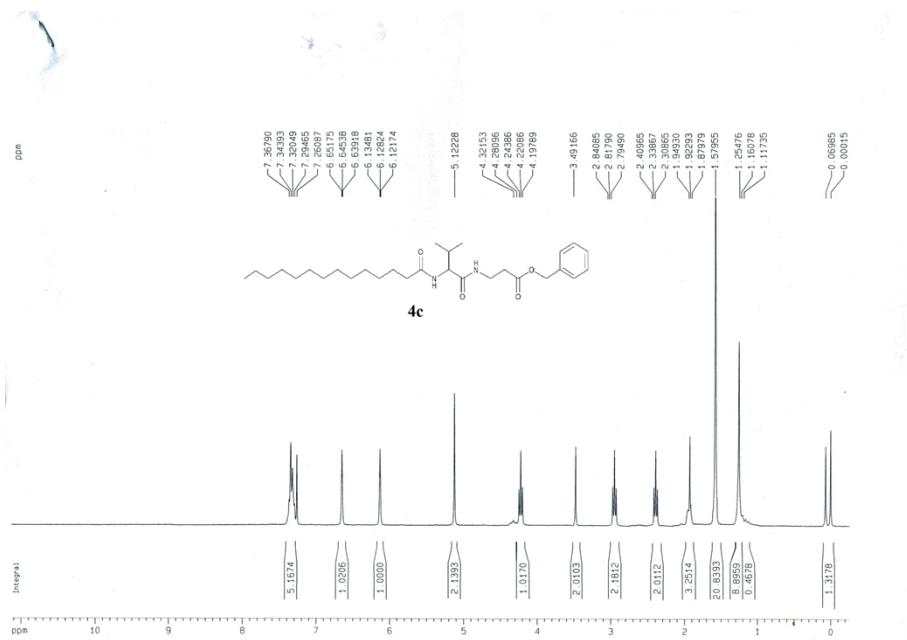
(m, >CHCH (CH₃)₂, 1H), 5.11 (s, -CONH(CH₂)₁₀COOCH₂Ph, 2H), 6.625-6.719 (m, -NHCO(CH₂)₆CH₃), -CONH(CH₂)₁₀COOCH₂Ph, 2H), 7.35 (s, aromatic, 5H). ¹³C NMR(75 MHz, CDCl₃) :δ_C : 173.42, 172.56, 171.41, 135.58, 128.62, 128.4, 128.2, 77.46, 77.041, 76.61, 66.54, 58.35, 40.3, 36.73, 36.67, 35.00, 33.1, 31.84, 31.19, 29.7, 29.7, 29.43, 29.32, 29.24, 28.7 25.74, 22.64, 17.4, 17.1, 14.1.ESI-MS (m/z) C₂₇H₄₄N₂O₄ (EXACT MASS= 516.37) 517.379(100%, [M + H]⁺), 539.39(58%, [M + Na]⁺), 555.329(18%, [M + K]⁺).



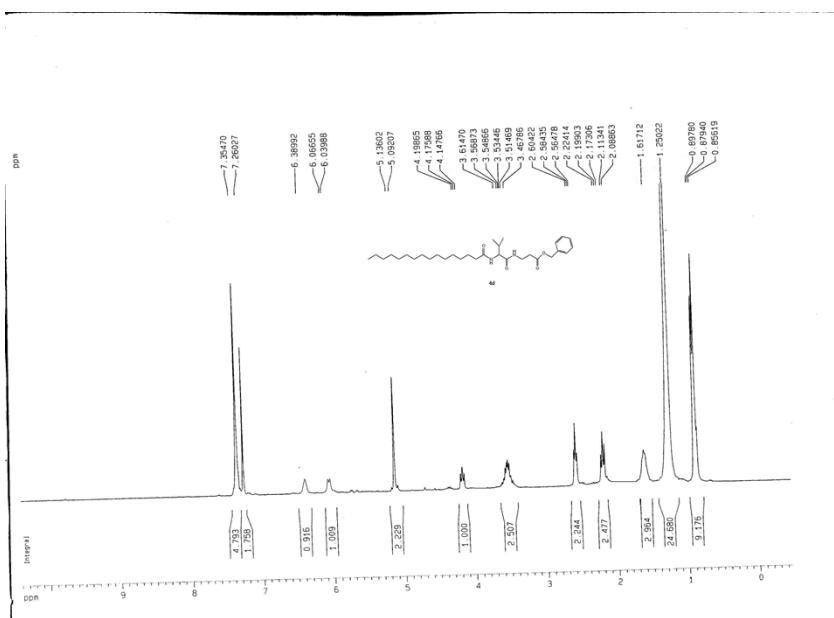
¹H NMR spectra of gelator **4a**.



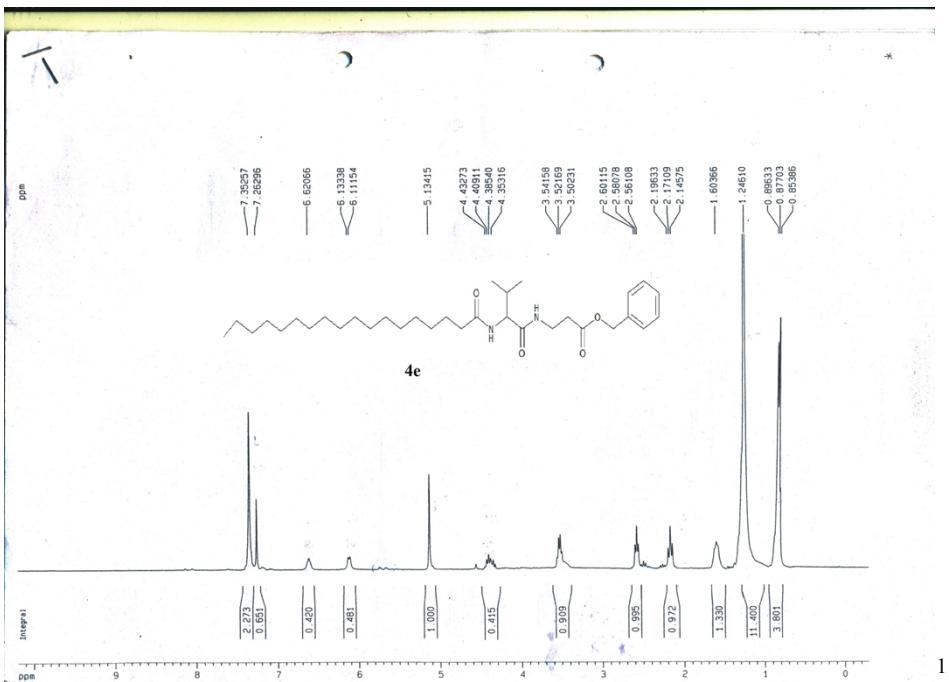
¹H NMR spectra of gelator **4b**.



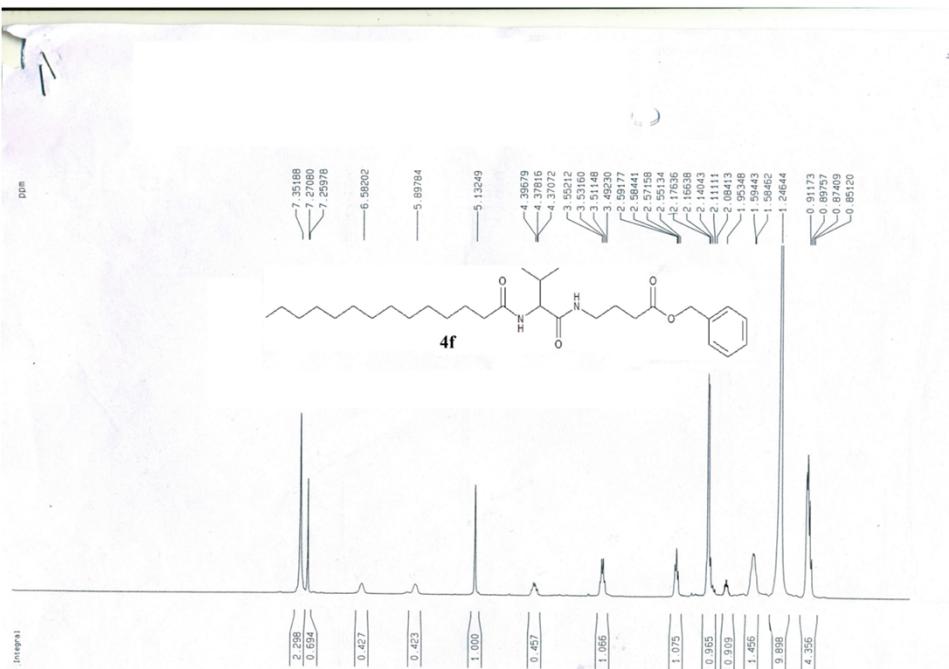
¹HNMR spectra of gelator **4c**.

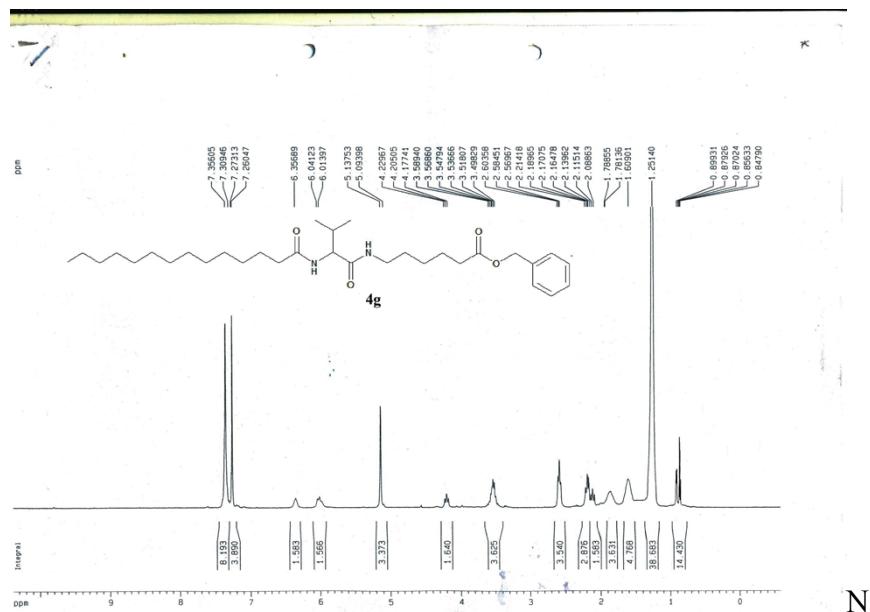


¹H NMR spectra of gelator 4d.

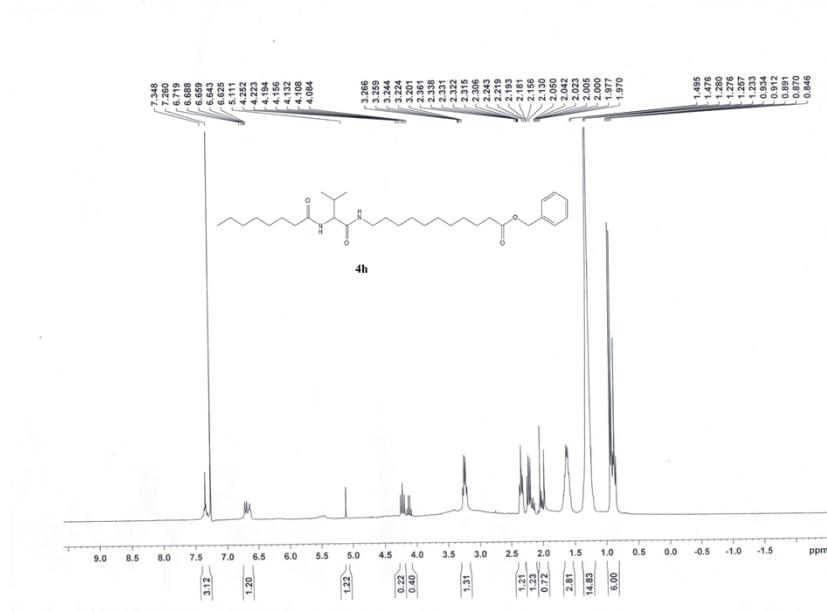


HNMR spectra of gelator **4e**.





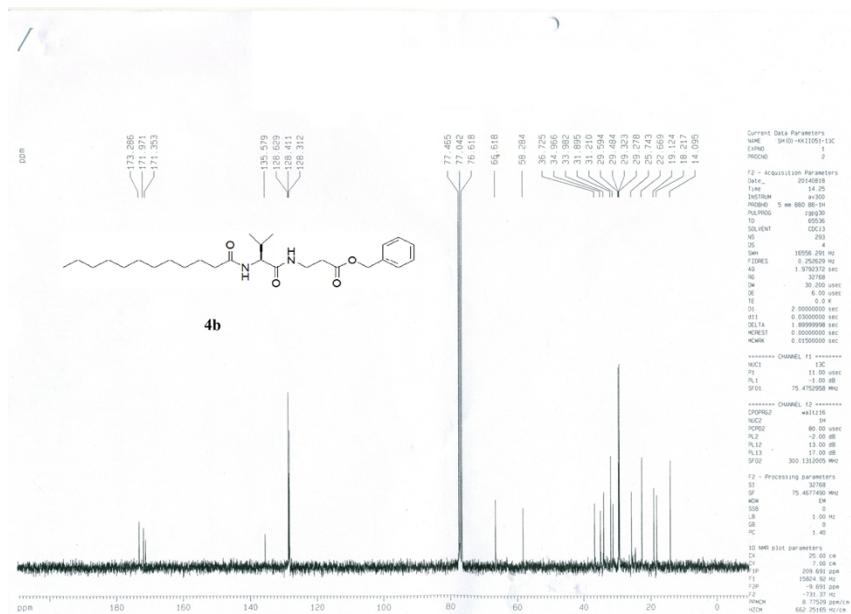
MR spectra of gelator **4g**.



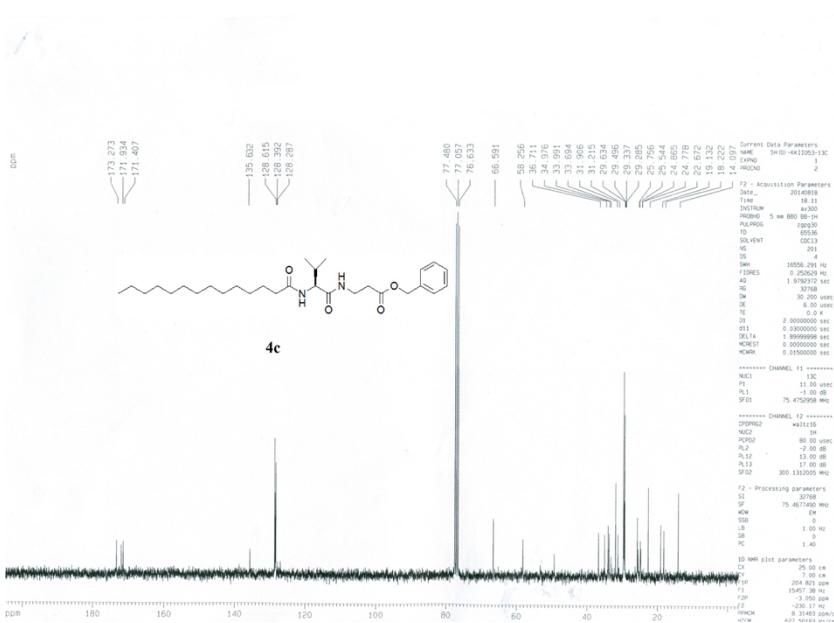
NMR spectra of gelator **4h**.



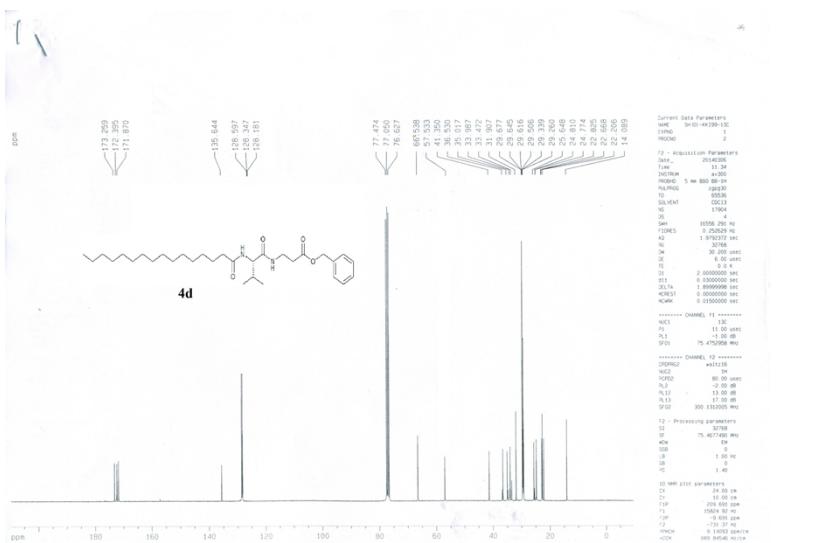
¹³C NMR spectra of gelator **4a**.



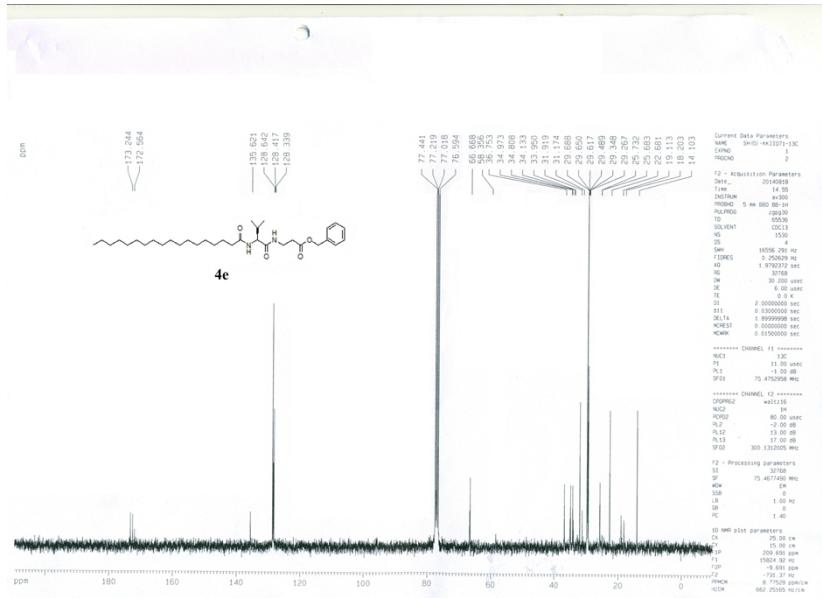
¹³C NMR spectra of gelator **4b**.



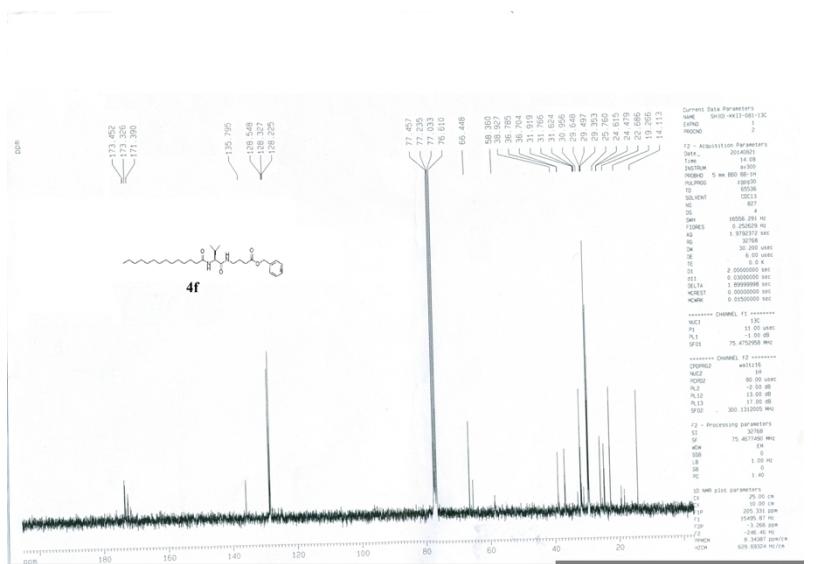
¹³C NMR spectra of gelator 4c.



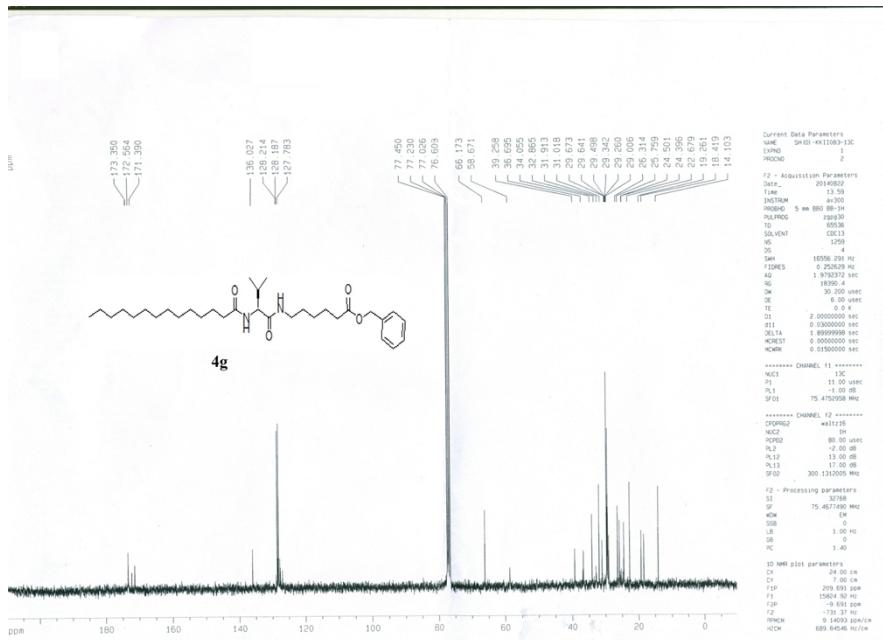
¹³C NMR spectra of gelator **4d**.



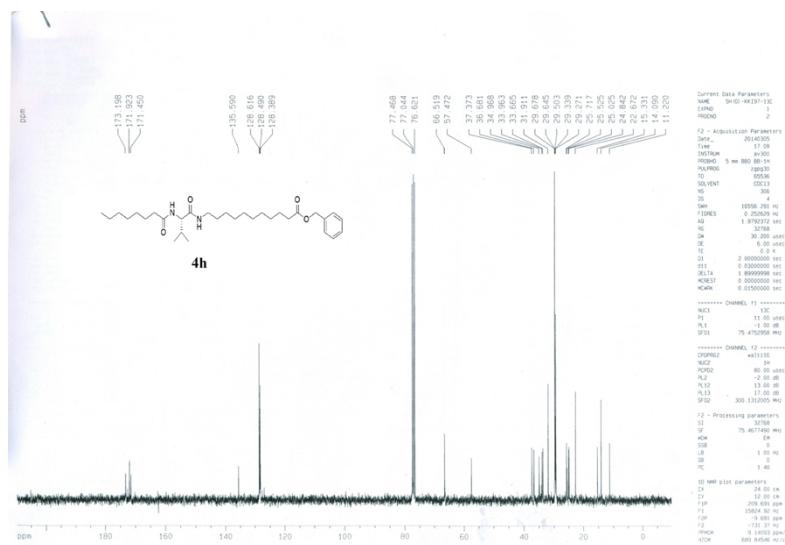
¹³C NMR spectra of gelator **4e**.



¹³C NMR spectra of gelator **4f**.



¹³C NMR spectra of gelator **4g**.



¹³C NMR spectra of gelator **4h**.

Table S1.. Gelation number and Gel to Sol phase transition temperature (T_{gel}) of gelators in different solvents

	(4a)	(4b)	(4c)	(4d)	(4e)	(4f)	(4g)	(4h)
c LogP →	5.726	6.611	7.544	7.942	9.153	8.172	9.117	8.285
SOLVENTS	GN	T _g						
Benzene	30	32.8	231	33.8	243	35.8	322	40.9
O-Xylene	32	32.6	188	33.6	196	35.6	265	41.3
Toluene	22	32.4	173	33.4	174	35.7	202	41.4
O-DCB	30	33.2	174	33.2	181	36.2	199	44.2
Anisole	21	32	66	32	73	35.5	85	39.1
PE	s	54	33.7	52	36.1	64	58.8	62
Hexane	410	33.9	398	35.1	270	37.1	240	42.1
Cyclohexane	124	33	157	33.9	200	35.6	302	42.5
THF	s	-	s	-	s	-	295	38.9
CHCl ₃	s	-	s	-	s	-	295	38.9
Dioxane	s	-	s	-	s	-	295	38.9
Acetonitrile	107	33.4	532	34.1	551	35.5	863	47.9
EtOAc	17	32.1	104	33.3	159	36.1	216	38
DMF	s	-	112	31.8	143	35.3	165	30.5
DMSO	60	32.3	215	33.4	234	36.2	258	54.3
Acetone	68	32.5	135	33.5	225	35.5	487	44.2
Nitromethane	319	33.6	729	34.9	836	36.8	2981	49.8
MeOH	74	32.1	271	33.1	310	35.8	500	51
80% aq MeOH	225	33.8	559	34.5	738	36.4	999	39.8
70% aq MeOH	114	33.6	456	34.2	695	36	886	39.5
50% aq MeOH	93	33.5	392	34	477	35.8	656	39.1
40% aq MeOH	s	-	s	-	s	-	521	37.7
80% aq EtOH	231	33.8	691	34.5	923	36.3	1377	41.2
70% aq EtOH	104	33.7	511	32	127	33.4	50	141
2-Propanol	s	-	77	33.5	89	35.2	113	35.9
Butanol	s	-	84	33.7	101	35.5	187	36.4
50% aq EG	is	-	is	-	is	-	is	-
50% aq Glycerol	is	-	is	-	is	-	is	-

O-DCB= O-Dichlorobenzene PE= Petroleum ether EG= Ethylene Glycol

Rheology:

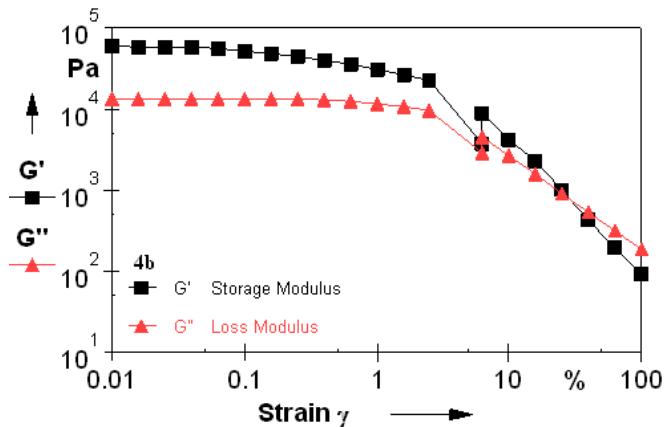


Figure S1. Strain amplitude sweep experiment (ω at 10rads/sec) for **(4b)** in acetonitrile.

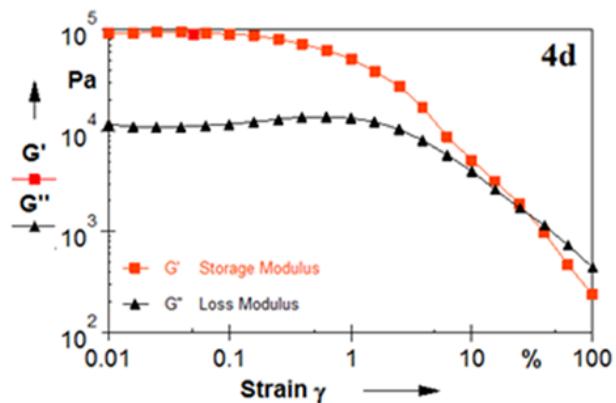


Figure S2. Strain amplitude sweep experiment (ω at 10rads/sec) for **(4d)** in acetonitrile.

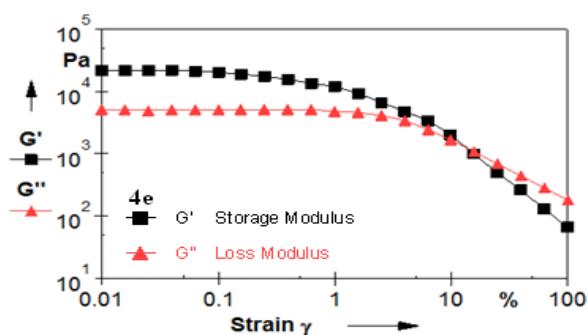


Figure S3. Strain amplitude sweep experiment (ω at 10rads/sec) for **(4e)** in acetonitrile

Continuation of Rheology....

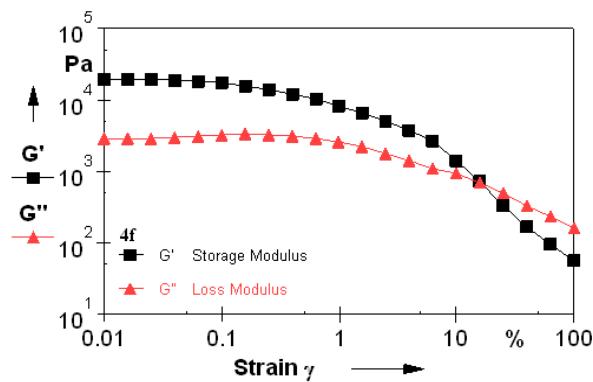


Figure S4. Strain amplitude sweep experiment (ω at 10rads/sec) for (4f) in acetonitrile.

Thermal stability

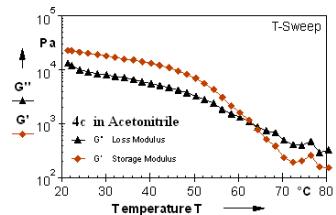


Figure S5. Temperature sweep experiment for (4c) in acetonitrile

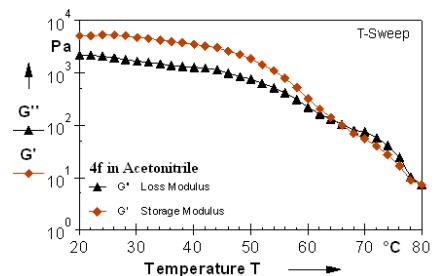


Figure S6. Temperature sweep experiment for (4f) in acetonitrile

Continuation of Thermal Stability.....

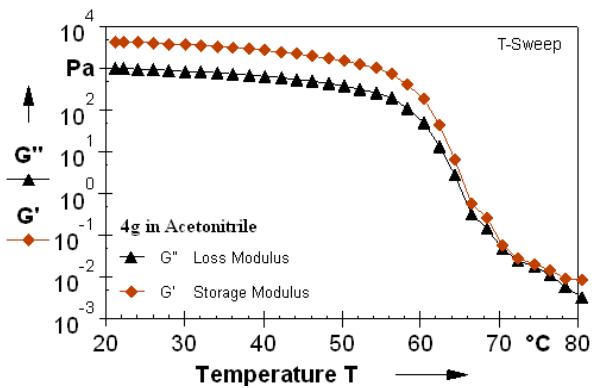


Figure S7.Temperature sweep experiment for (4g) in acetonitrile

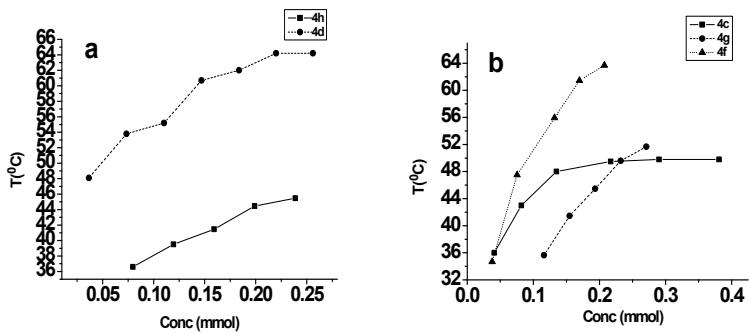


Figure S8. T_{gel} (°C) versus concentration (mmoles) plots for gelators of (a)

SET-II and (b) SET-III in Acetonitrile.

XRD :

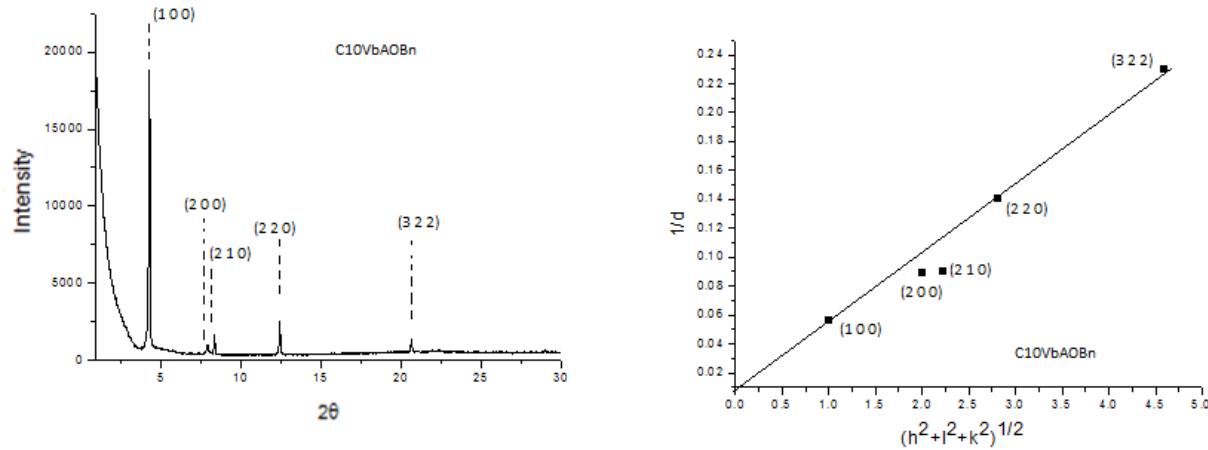


Figure S9. XRD profile of xerogels (intensity vs. 2θ) for **4a** in acetonitrile and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$.

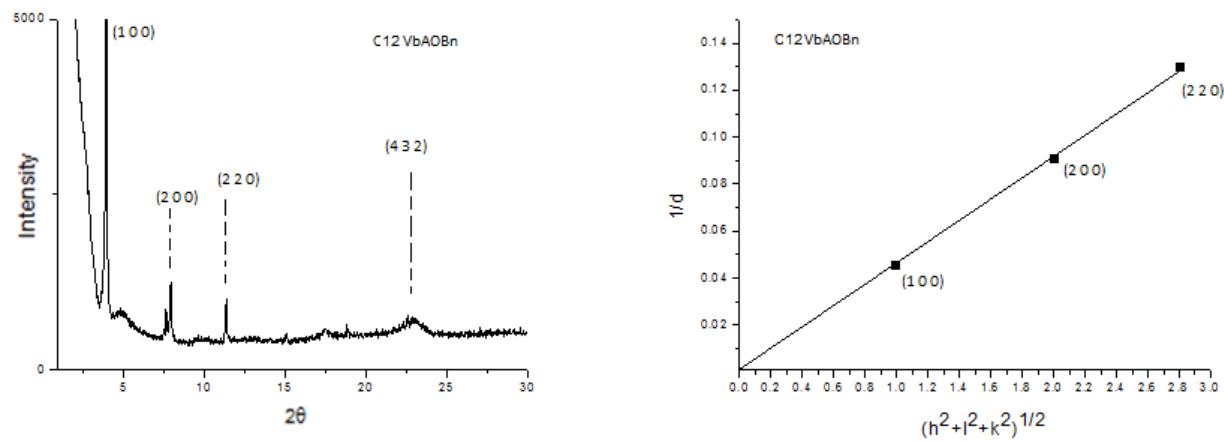
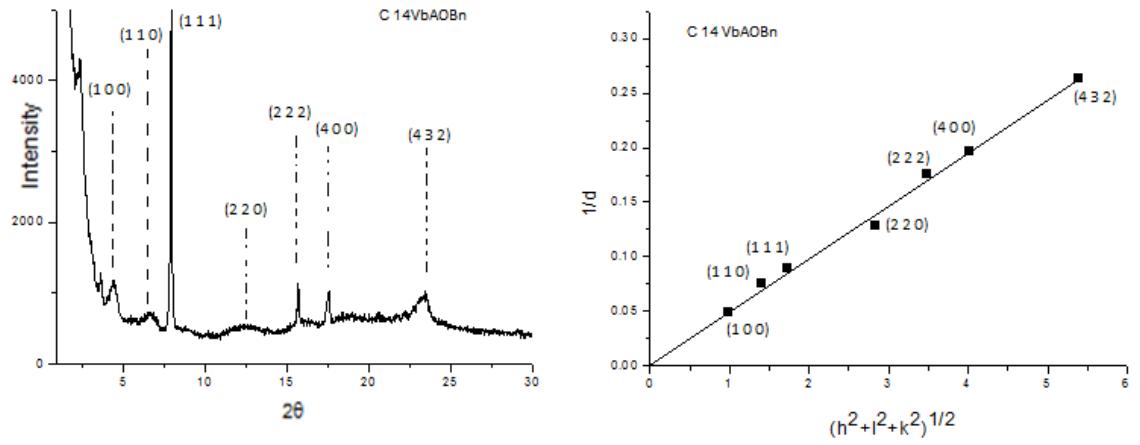


Figure S10. XRD profile of xerogels (intensity vs. 2θ) for **4b** in acetonitrile and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$.

Continuation of XRD....



Figure

S11.XRD profile of xerogels (intensity vs. 2θ) for **4c** in acetonitrile and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$.

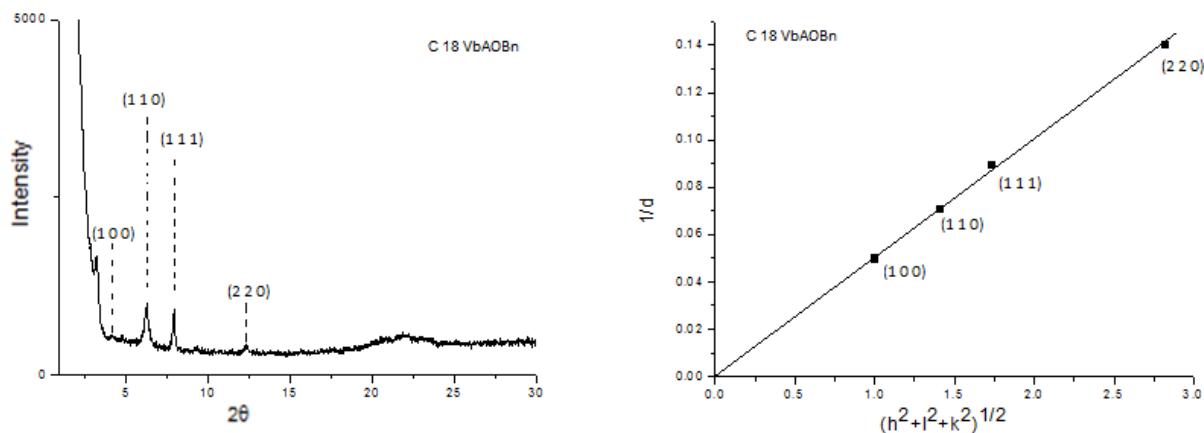


Figure S12. XRD profile of xerogels (intensity vs. 2θ) for **4e** in acetonitrile and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$.

Continuation of XRD....

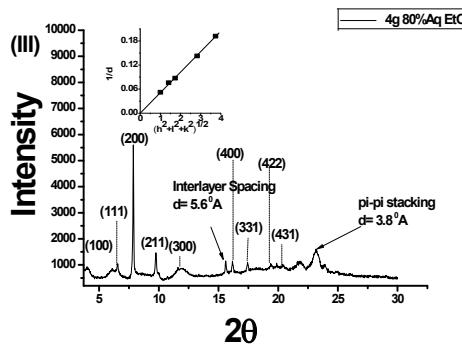


Figure S13.XRD profile of xerogels (intensity vs. 2θ) for **4g** in 80%(v/v) Ethanol. H_2O and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$ (at inset).

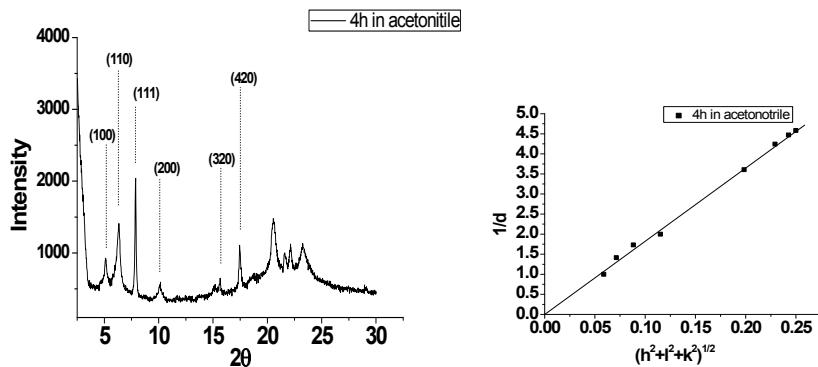


Figure S14.XRD profile of xerogels (intensity vs. 2θ) for **4h** in acetonitrile and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$.

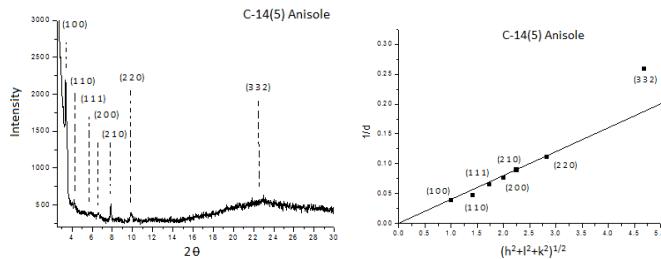


Figure S14.XRD profile of xerogels (intensity vs. 2θ) for **4g** in anisole and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$

Continuation of XRD.....

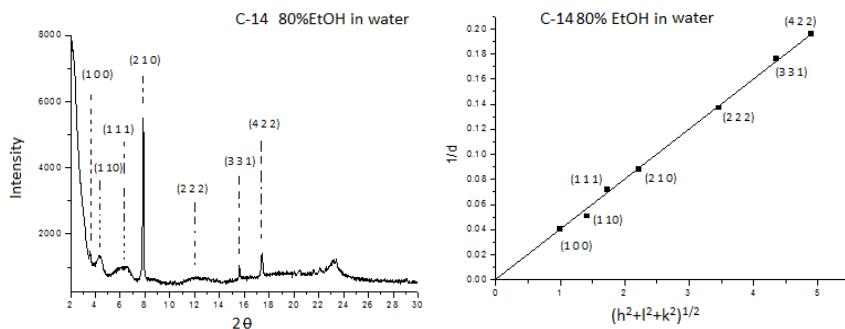


Figure S15.XRD profile of xerogels (intensity vs. 2θ) for **4c** in 80% aq. EtOH and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$.

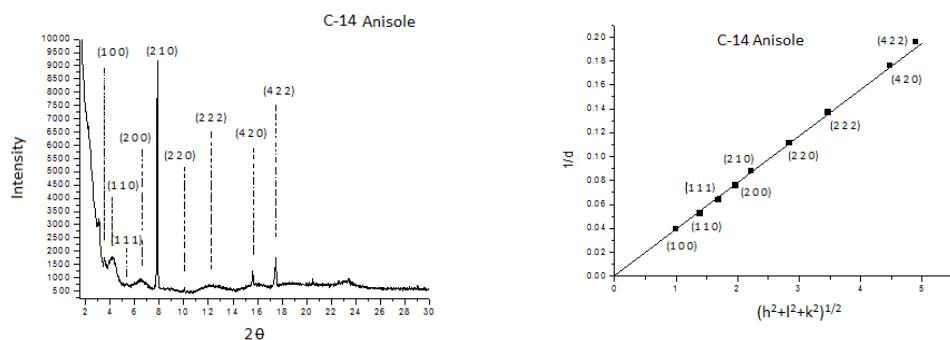


Figure S16.XRD profile of xerogels (intensity vs. 2θ) for **4c** in Anisole and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$.

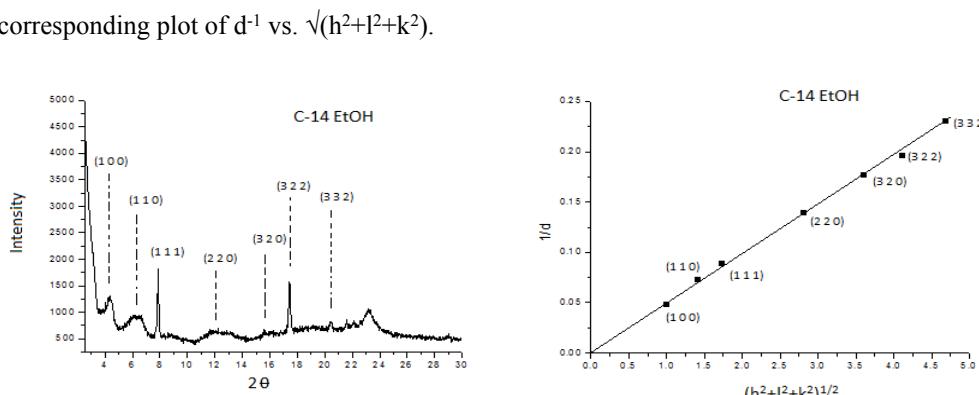
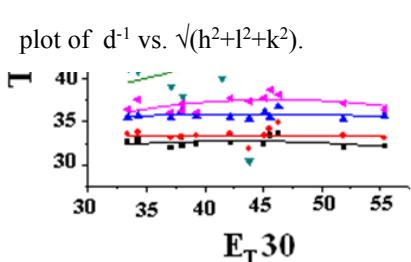


Figure S17.XRD profile of xerogels (intensity vs. 2θ) for **4c** in EtOH and its corresponding plot of d^{-1} vs. $\sqrt{(h^2+l^2+k^2)}$.



S-xxi

Figure S18. E_{T30} versus T_g plot of gelator **4a**, **4b**, **4c**, **4d** and **4e** in different solvents.

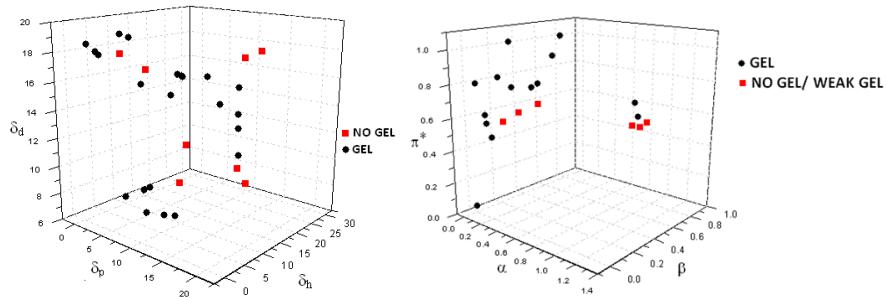


Figure S19.3D plots of Hansen and Kamlet Taft parameters at different solvent for **4a-4g**

gelators.

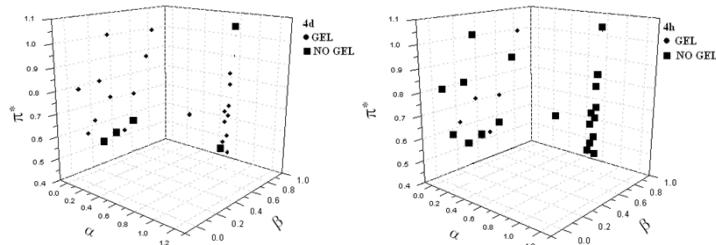


Figure S20.Comparison of 3D plots of Kamlet Taft parameters at different solvents for **4d-4h** gelators.

Continuation of Solvent effect....

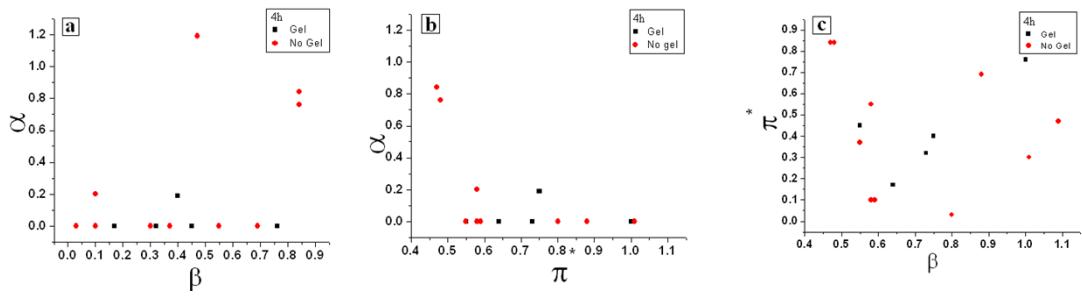


Figure S22. Plots of

Figure S21. Plots of δ_p vs δ_h , δ_d vs δ_h and δ_p vs δ_d (HSP) for gelator **4h** [gelation is found in solvents with lower δ_h values, i.e., solvents with lower H-bonding capability].

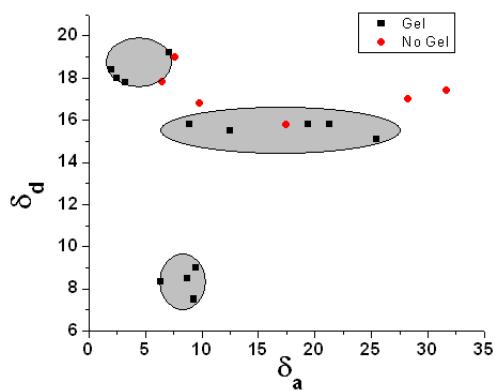


Figure S23. Plots of δ_d vs δ_h for gelators **4a-4g**.

Continuation of Solvent effect....

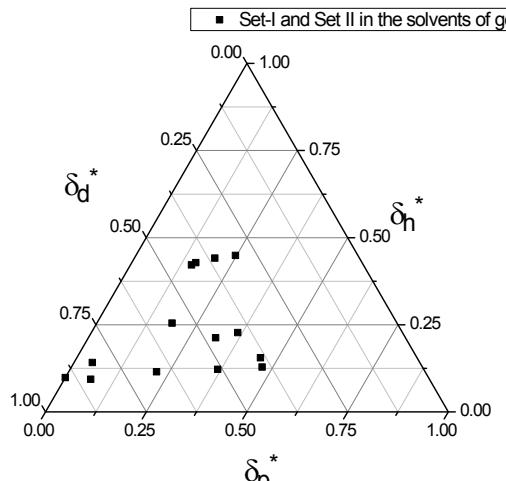


Figure S24.Teas plot of Hansen Parameters for gelators of Set-I and Set-II in solvents of gelation.

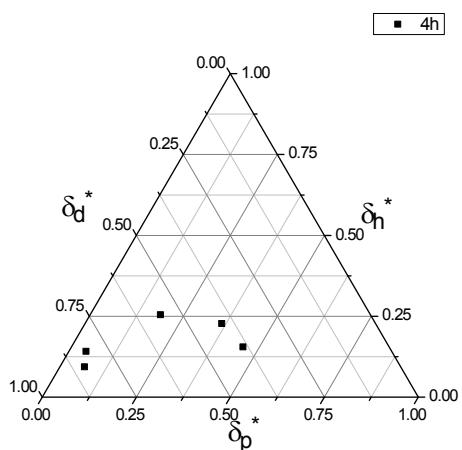


Figure S25.Teas plot of Hansen Parameters for **4h** in solvents of gelation.

CD

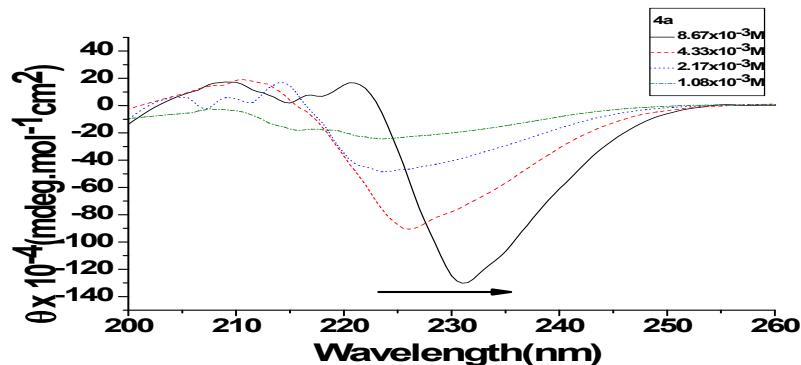


Figure S26. CD spectra of **4f** at different concentrations.

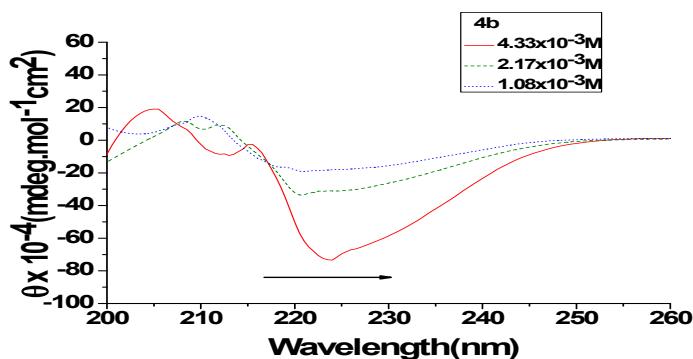


Figure S27. CD spectra of **4f** at different concentrations.

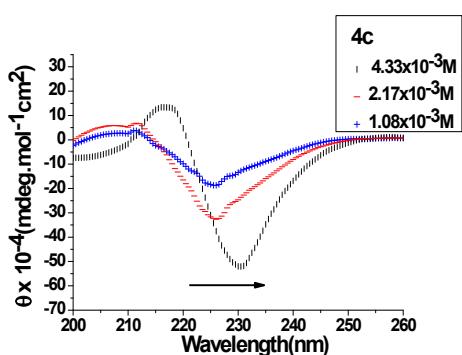


Figure S28. CD spectra of **4f** at different concentrations.

Continuation of CD.....

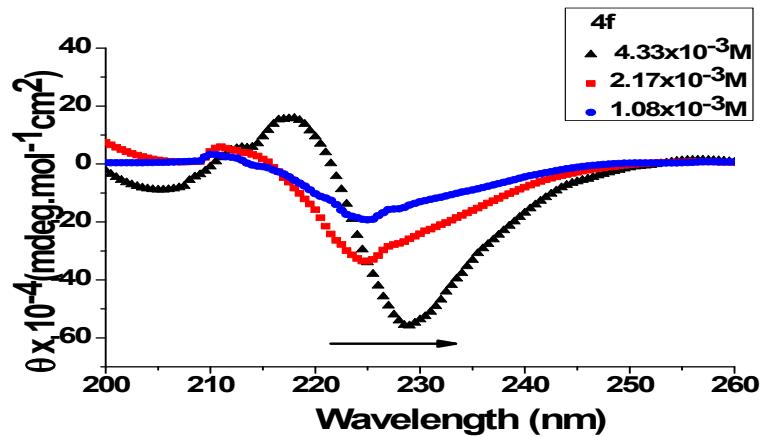


Figure S29. CD spectra of **4f** at different concentrations.

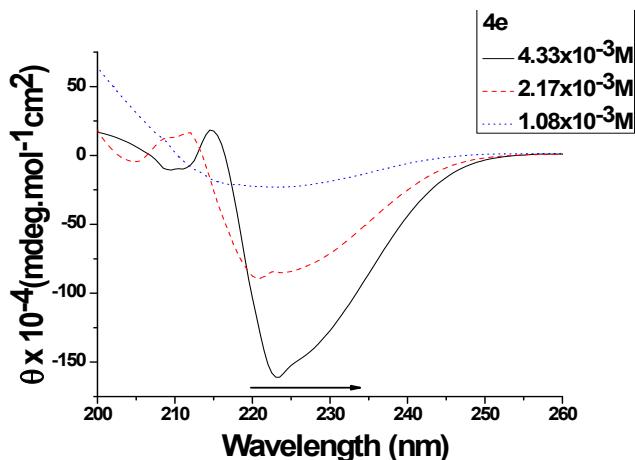


Figure S30. CD spectra of **4f** at different concentrations.

FT-IR spectra:

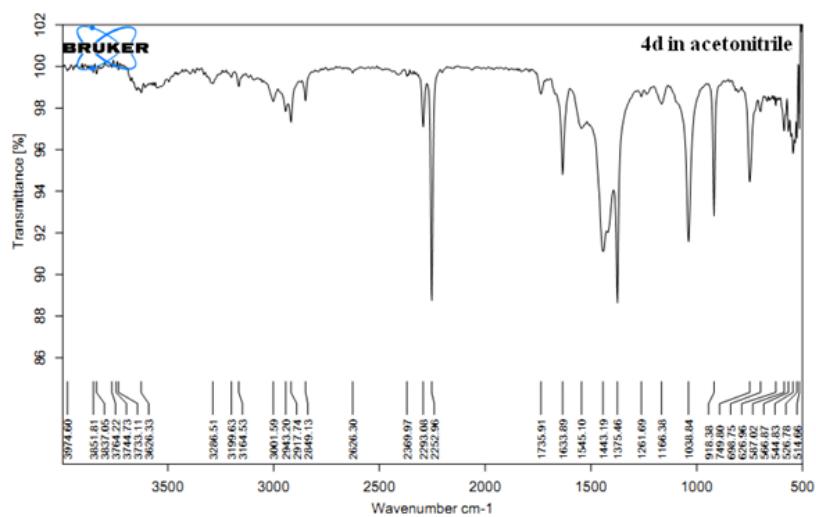


Figure S31. IR spectra of gelator **4d** in acetonitrile in gel state.

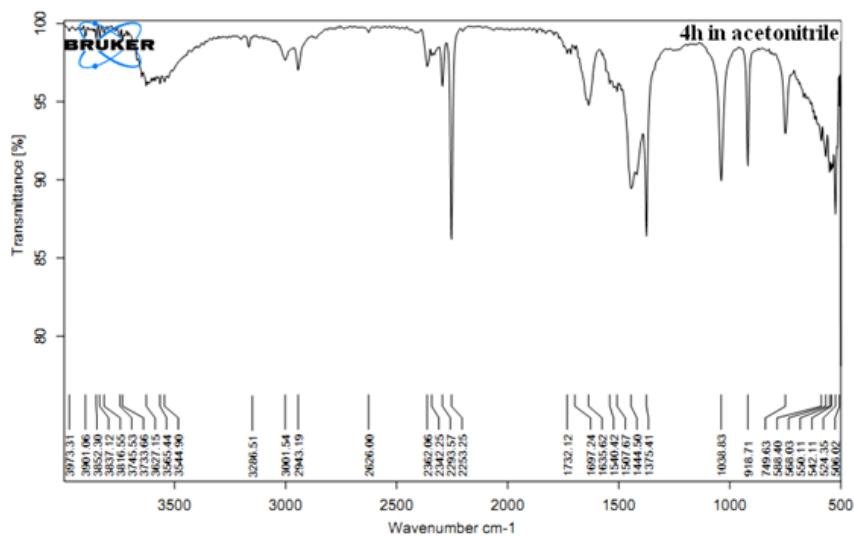


Figure S32. IR spectra of gelator **4h** in acetonitrile in gel state.

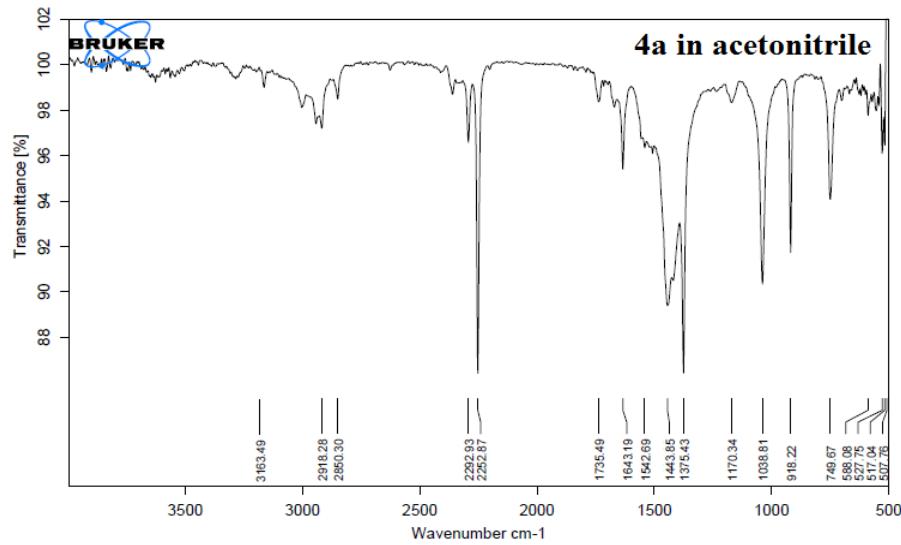


Figure S33. IR spectra of gelator **4a** in acetonitrile in gel state.

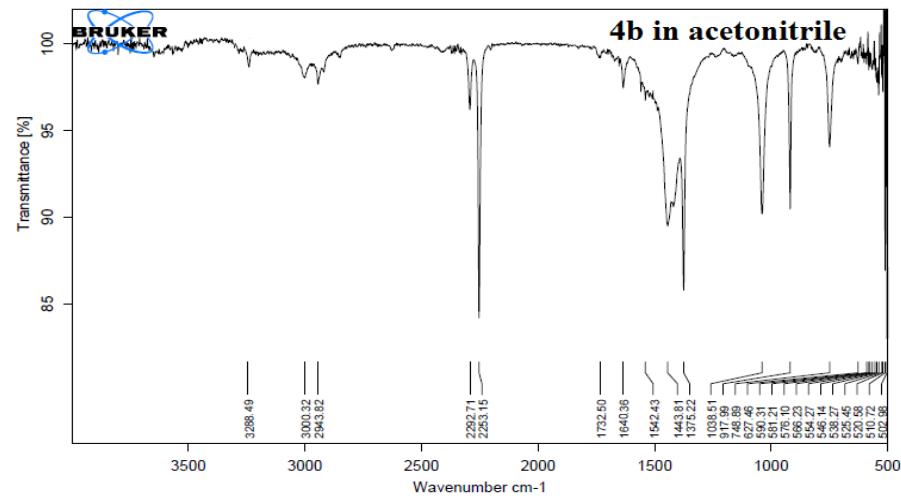


Figure S34. IR spectra of gelator **4b** in acetonitrile in gel state.

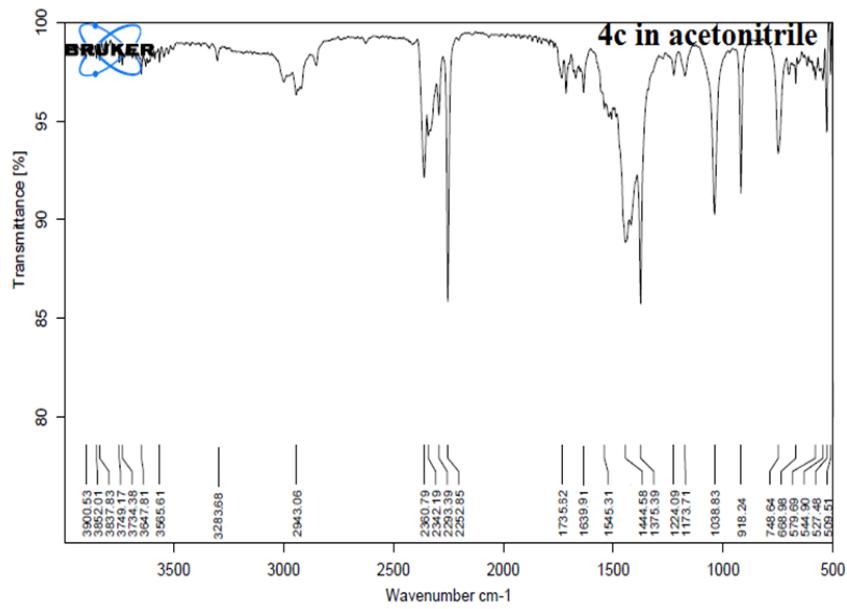


Figure S35. IR spectra of gelator **4c** in acetonitrile in gel state.

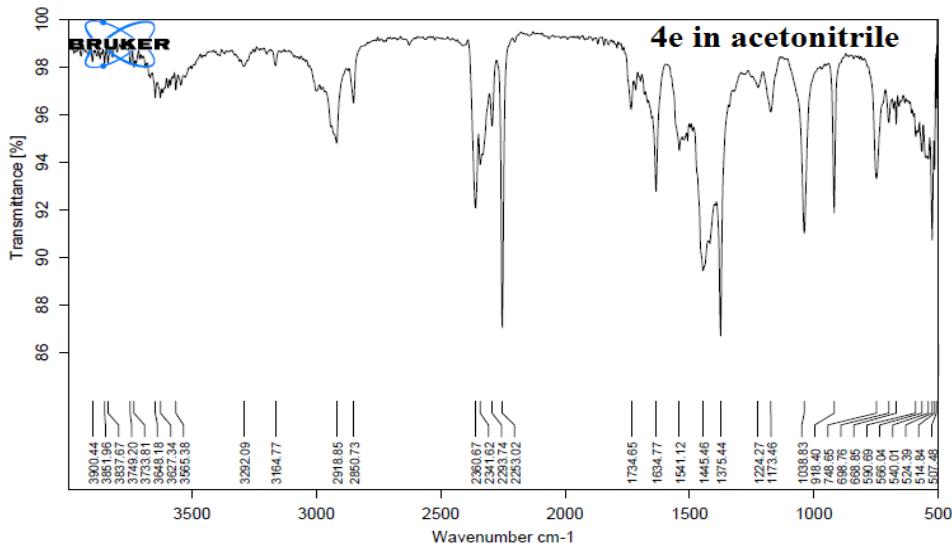


Figure S36. IR spectra of gelator **4e** in acetonitrile in gel state.

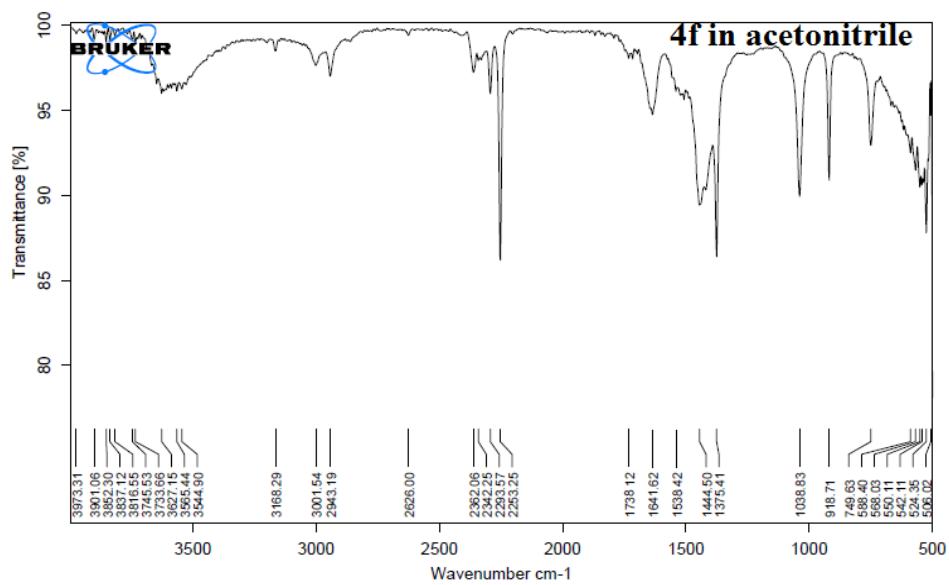


Figure S37. IR spectra of gelator **4f** in acetonitrile in gel state.

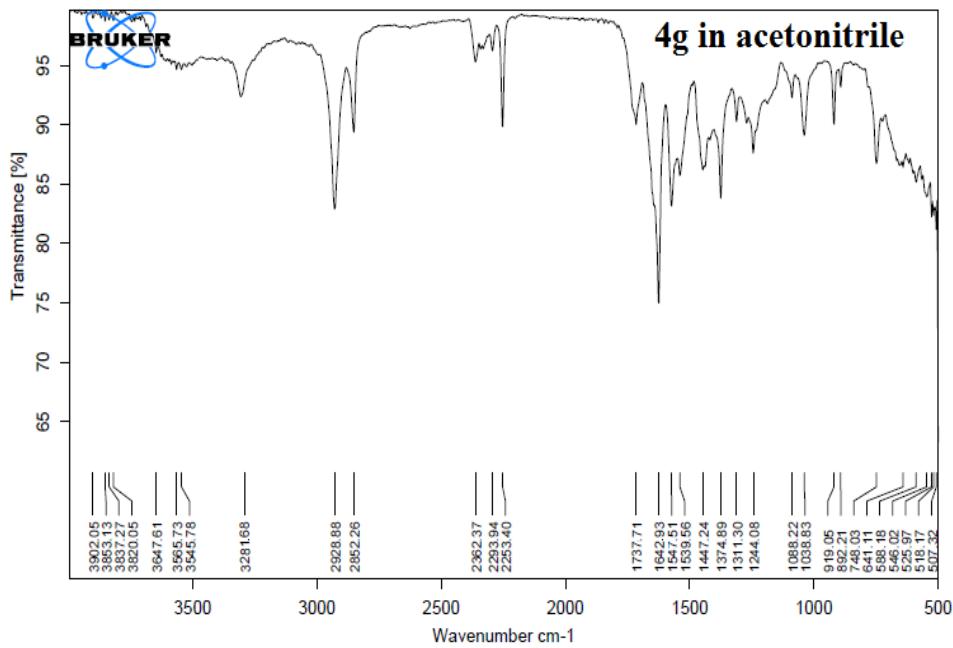


Figure S38. IR spectra of gelator **4g** in acetonitrile in gel state.