

# Highly Promising Discrimination of Various Catecholamine by Ratiometric Fluorescence under Intermolecular Self-Association of Two Sensing Elements

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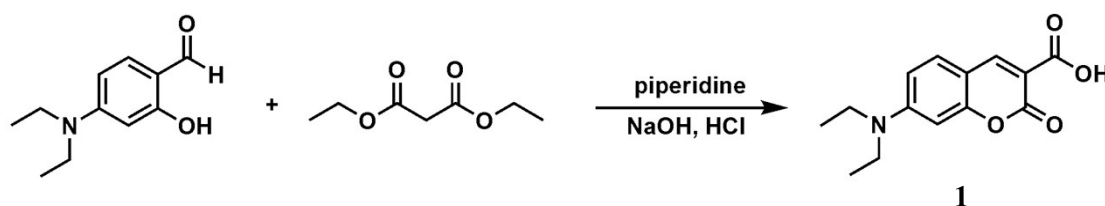
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## Supplementary data

(16 pages)

### 1. Synthesis of crown ether/coumarin based sensor

#### 1.1 Preparation of 7-diethylamino-2-oxo-2H-chromen-3-carboxylic acid (**1**)



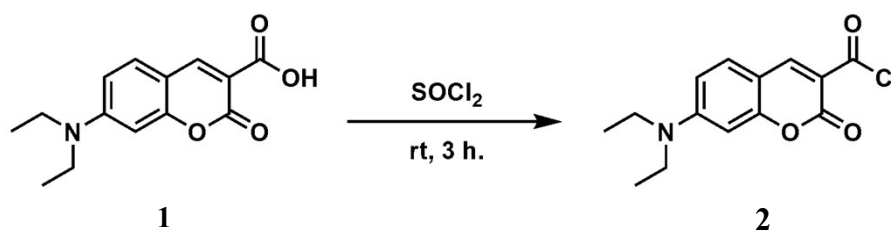
Scheme S1 Synthesis of compound **1**

Into a two-neck round bottom flask equipped with a magnetic bar, 4-diethylaminosalicylaldehyde (0.386 g, 2 mmol), diethylmalonate (0.61 mL, 4 mmol), and piperidine (0.2 mL) were dissolved in absolute ethanol (6 mL). The reaction mixture was stirred and refluxed for 6 hours under nitrogen atmosphere. Then 10% NaOH (6 mL) solution was added to the reaction and refluxed for 15 min. The solution was cooled to room temperature and acidified to pH 2 using concentrated hydrochloric acid at 0°C affording a precipitate deposit which was filtered out, washed with water, then recrystallized with ethanol to give an orange crystal (**1**, 74%). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): δ (in ppm) = 12.37 (s, 1H, COOH), 8.67 (s, 1H, ArH), 7.46 (d, *J* = 9.2 Hz, 1H, ArH), 6.71 (dd, *J* = 4.27 Hz, 1H, ArH), 6.53 (d, *J* = 2.0 Hz, 1H, ArH), 3.49 (q, *J* = 7.07 Hz, 4H, CH<sub>2</sub>CH<sub>3</sub>), 1.26 (t, *J* = 7.2 Hz, 6H, CH<sub>2</sub>CH<sub>3</sub>). <sup>13</sup>C-NMR (400 MHz, CDCl<sub>3</sub>): δ (in ppm) = 165.52, 164.42, 158.07, 153.78, 150.27, 131.94, 110.91, 108.59, 105.70, 96.89, 45.34, 12.39.

MALDI-TOF mass: Anal. Calcd for  $[C_{14}H_{15}NO_4]^+$   $m/z = 261.10$

Found  $m/z = 261.32$

1.2 Preparation of 7-diethylamino-2-oxo-2H-chromen-3-carboxylic chloride (**2**)

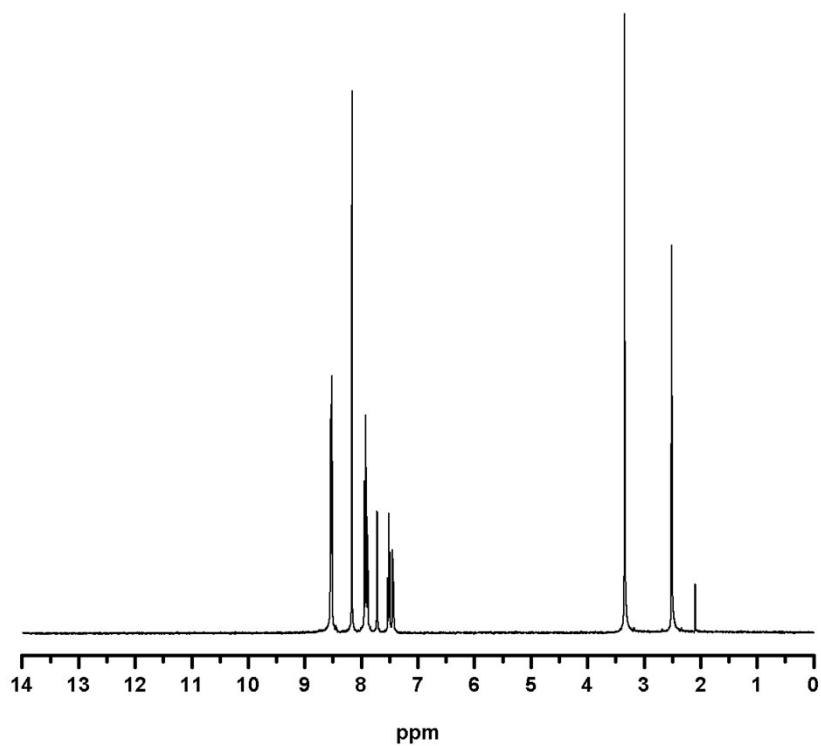


**Scheme S2** Synthesis of compound **2**

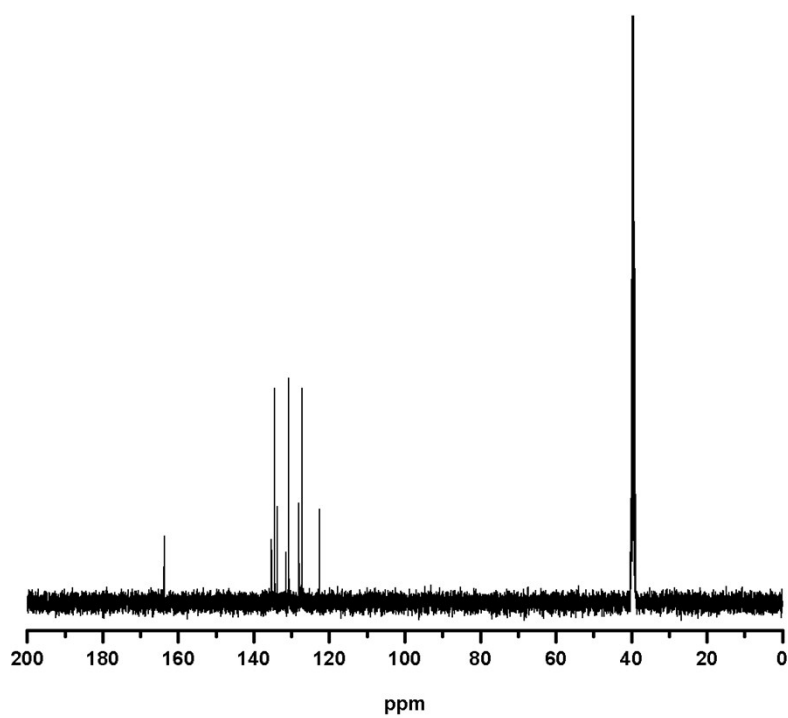
Into a two-neck round bottom flask equipped with a magnetic bar, **1** (0.187 g, 0.7 mmol) was added to dry thionyl chloride (3 mL) and the suspension was stirred at room temperature for 3 hours under nitrogen atmosphere. The precipitate was washed with dichloromethane under vacuum to give a yellow solid **2** in a quantitative yield.

$^1H$ -NMR (400 MHz,  $CDCl_3$ ):  $\delta$  (in ppm) = 8.68 (s, 1H, ArH), 7.49 (d,  $J = 4.4$  Hz, 1H, ArH), 6.76 (m, 1H, ArH), 6.59 (s, 1H, CCH), 3.50 (q,  $J = 6.9$  Hz, 4H,  $CH_2CH_3$ ), 1.28 (t,  $J = 3.4$  Hz, 6H,  $CH_2CH_3$ )

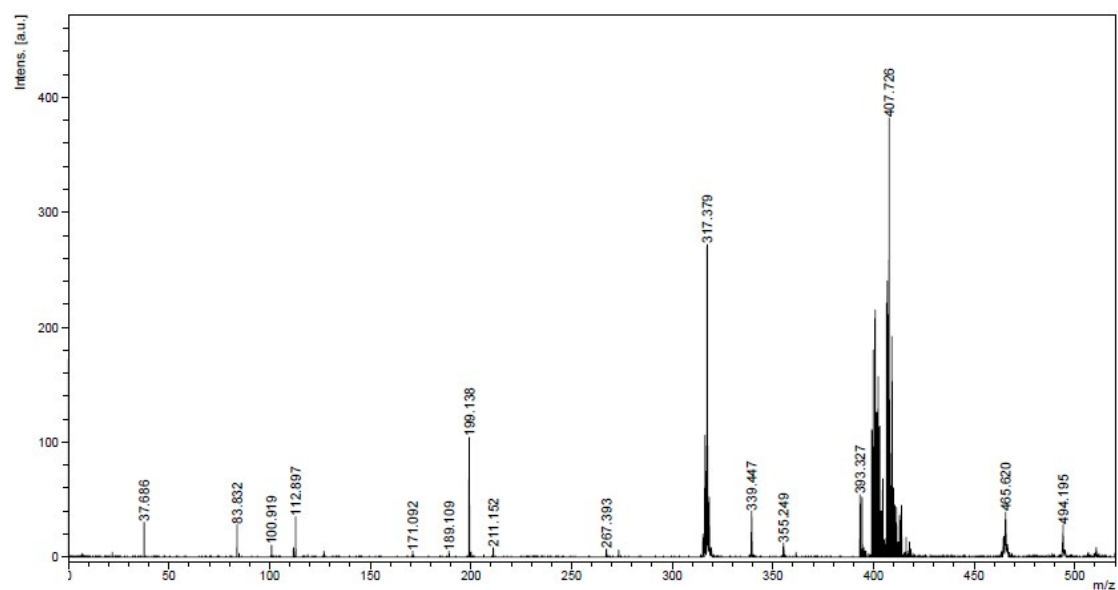
2. Characterization of the compounds



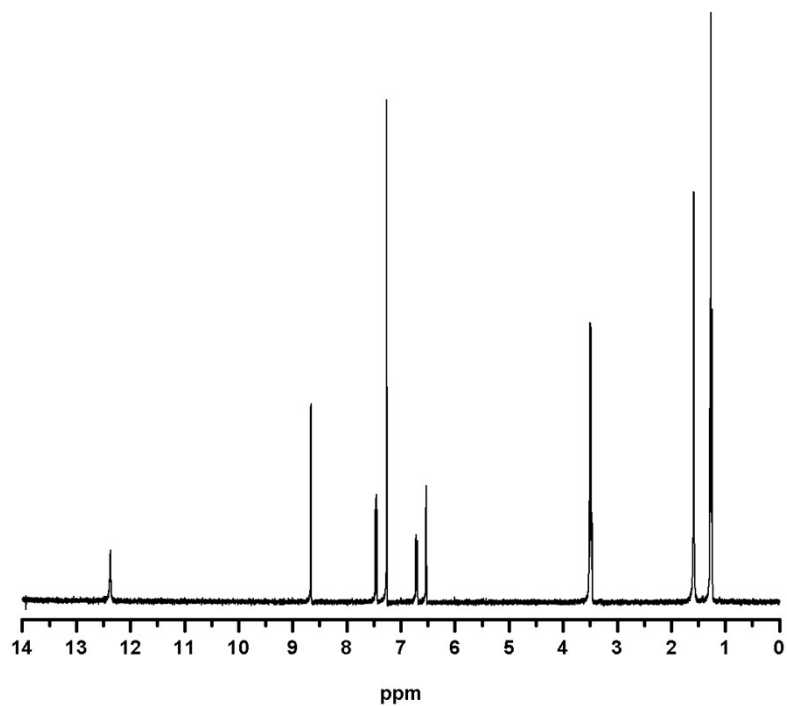
**Figure S1** The  $^1\text{H}$ -NMR spectrum of sensor **NB** in  $\text{DMSO-}d_6$  at 400 MHz



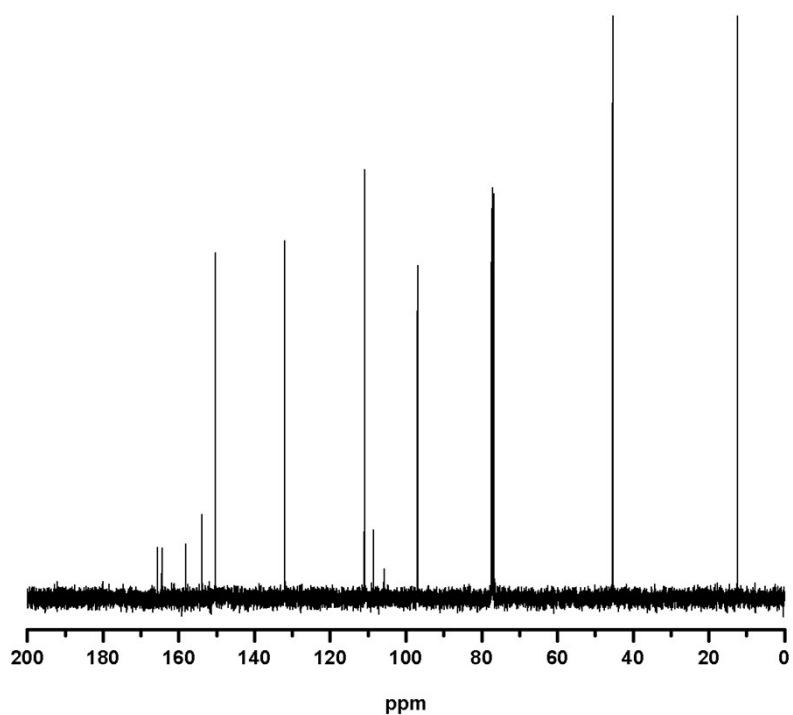
**Figure S2** The  $^{13}\text{C}$ -NMR spectrum of sensor **NB** in  $\text{DMSO-}d_6$  at 400 MHz



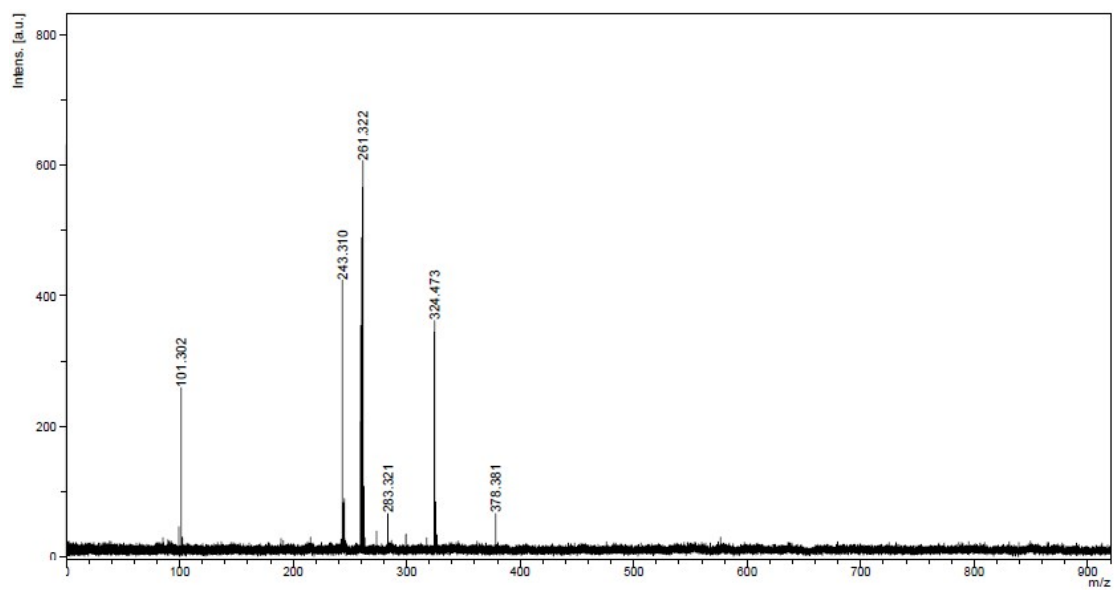
**Figure S3** MALDI-TOF mass spectrum of sensor **NB** shown at 317.379 m/z



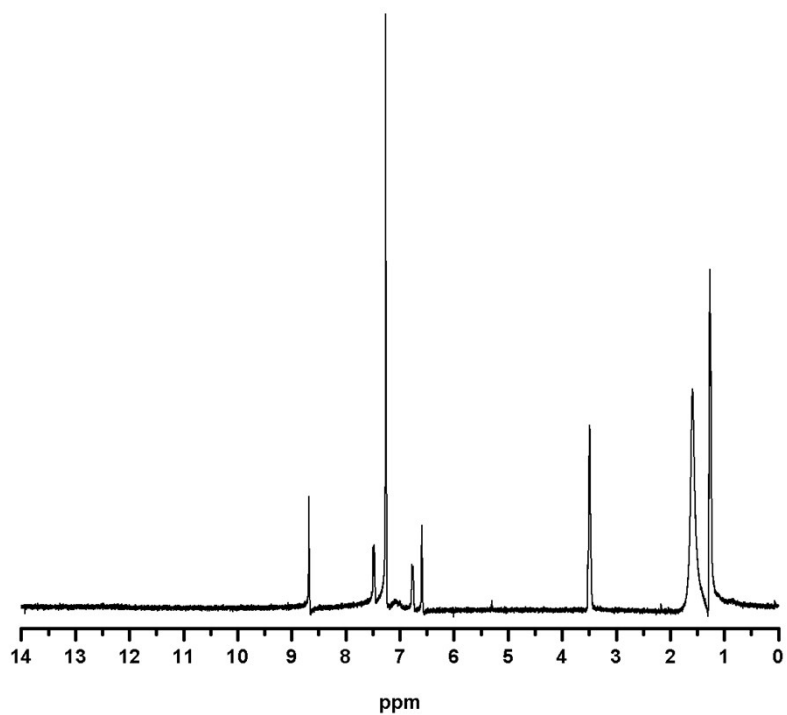
**Figure S4** The  $^1\text{H}$ -NMR spectrum of **1** in  $\text{CDCl}_3$  at 400 MHz



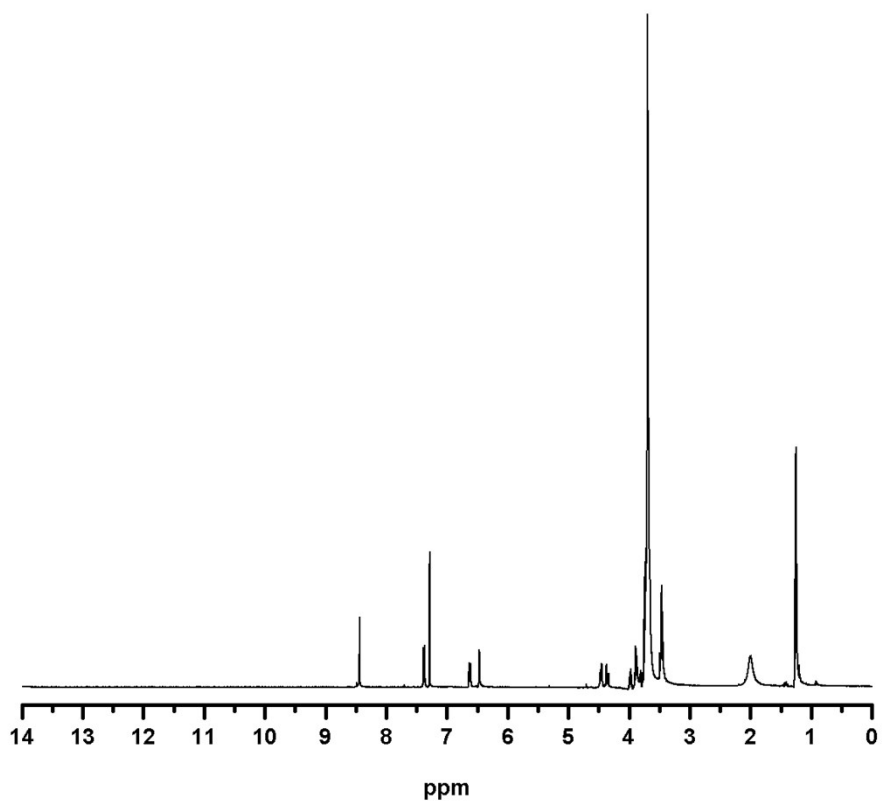
**Figure S5** The  $^{13}\text{C}$ -NMR spectrum of **1** in  $\text{CDCl}_3$  at 400 MHz



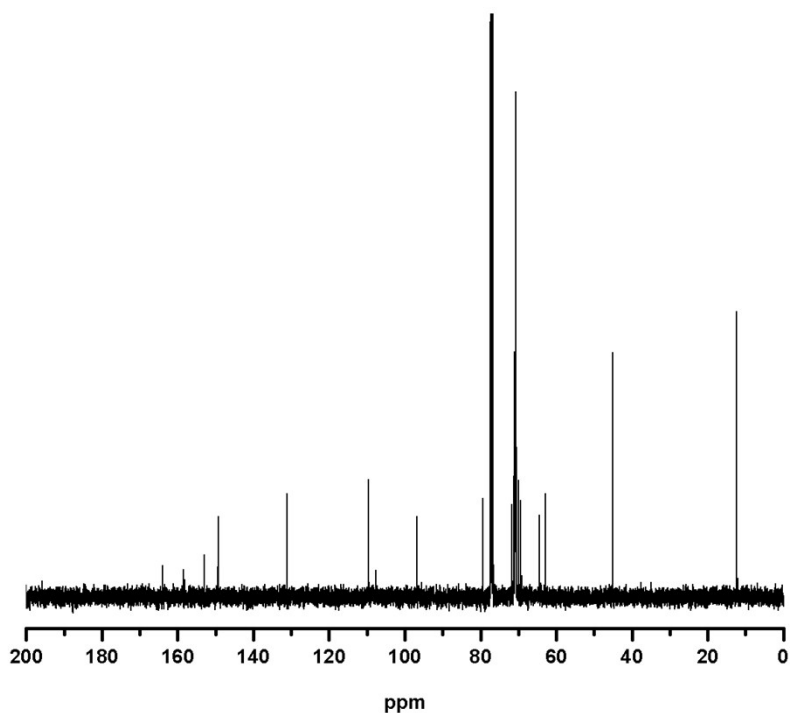
**Figure S6** MALDI-TOF mass spectrum of **1** shown at 261.322m/z



**Figure S7** The  $^1\text{H}$ -NMR spectrum of **2** in  $\text{CDCl}_3$  at 400 MHz



**Figure S8** The  $^1\text{H}$ -NMR spectrum of sensor **CC** in  $\text{CDCl}_3$  at 400 MHz



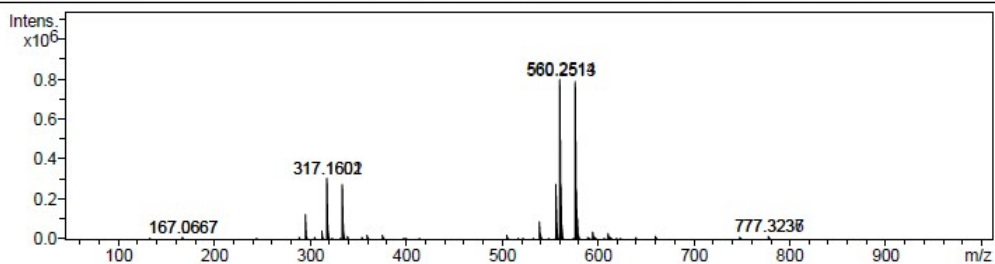
**Figure S9** The  $^{13}\text{C}$ -NMR spectrum of sensor **CC** in  $\text{CDCl}_3$  at 400 MHz

### Mass Spectrum List Report

<b>Analysis Info</b>		Acquisition Date	3/15/2013 2:22:02 PM
Analysis Name	D:\Data\Data Service\Year 2013\Small molecule\03152013\CC_pos.d	Operator	BDAL@DE
Method	tune_low.m	Instrument / Ser#	microTOF-Q II 10335
Sample Name	CC_pos		
Comment			

#### Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	1000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Waste



**Figure S10** The ESI-High Resolution Mass spectrum of sensor **CC** at 560.25 m/z

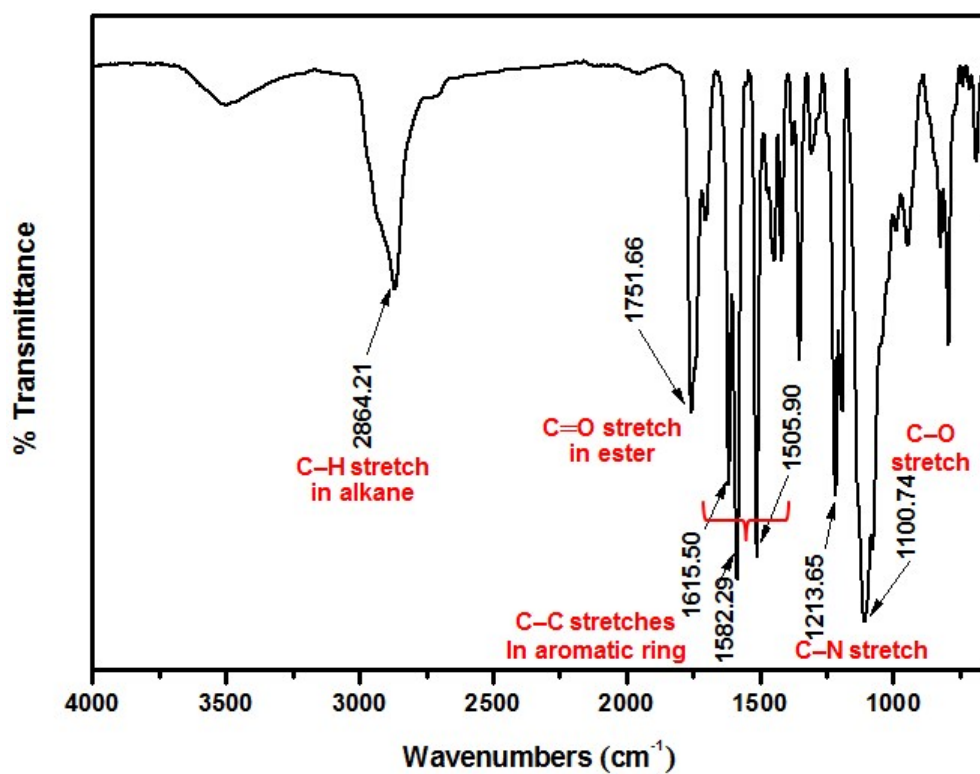
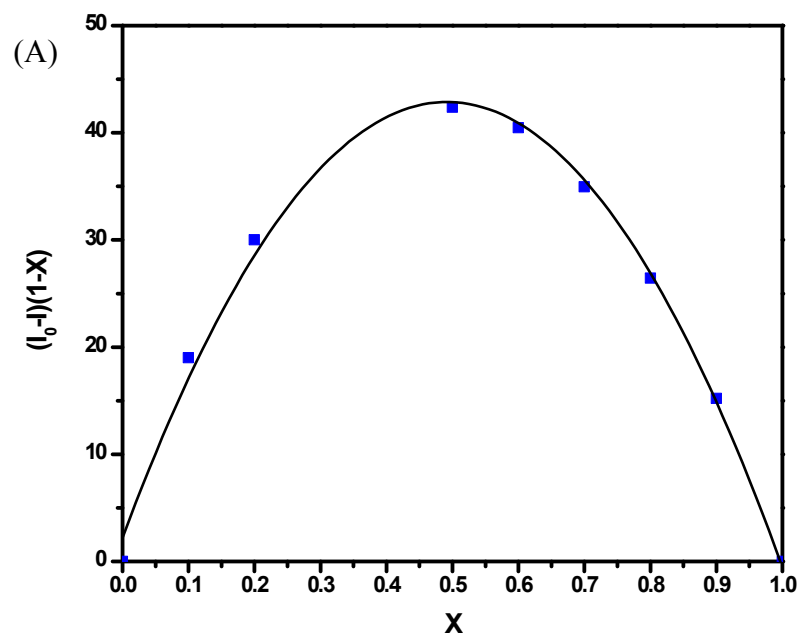
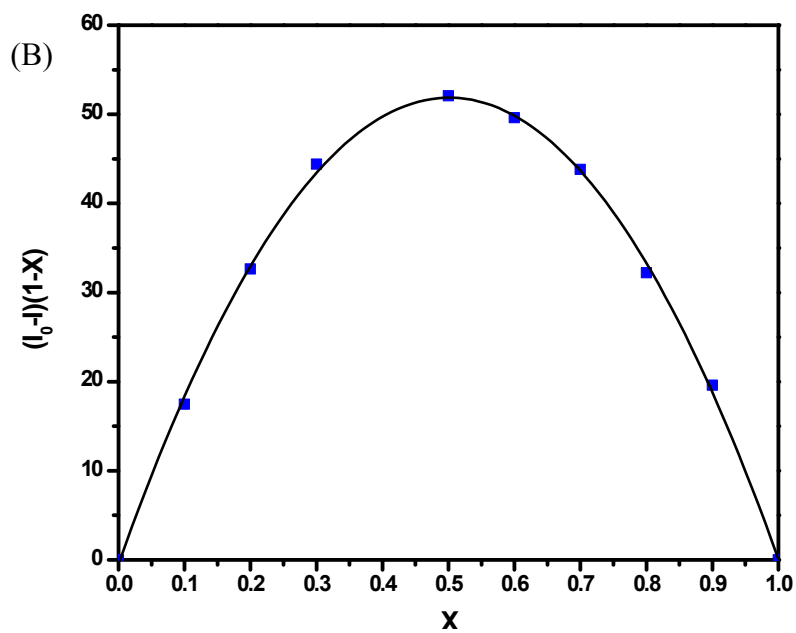


Figure S11 IR spectrum of sensor CC

### 3. Job's plots studies

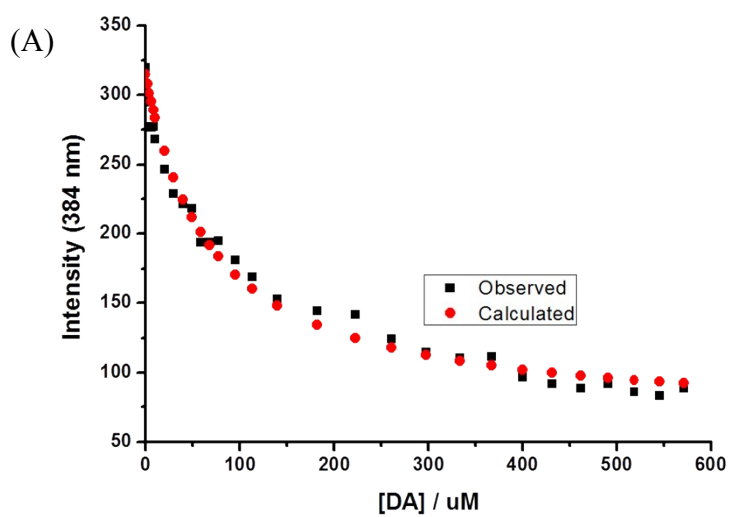


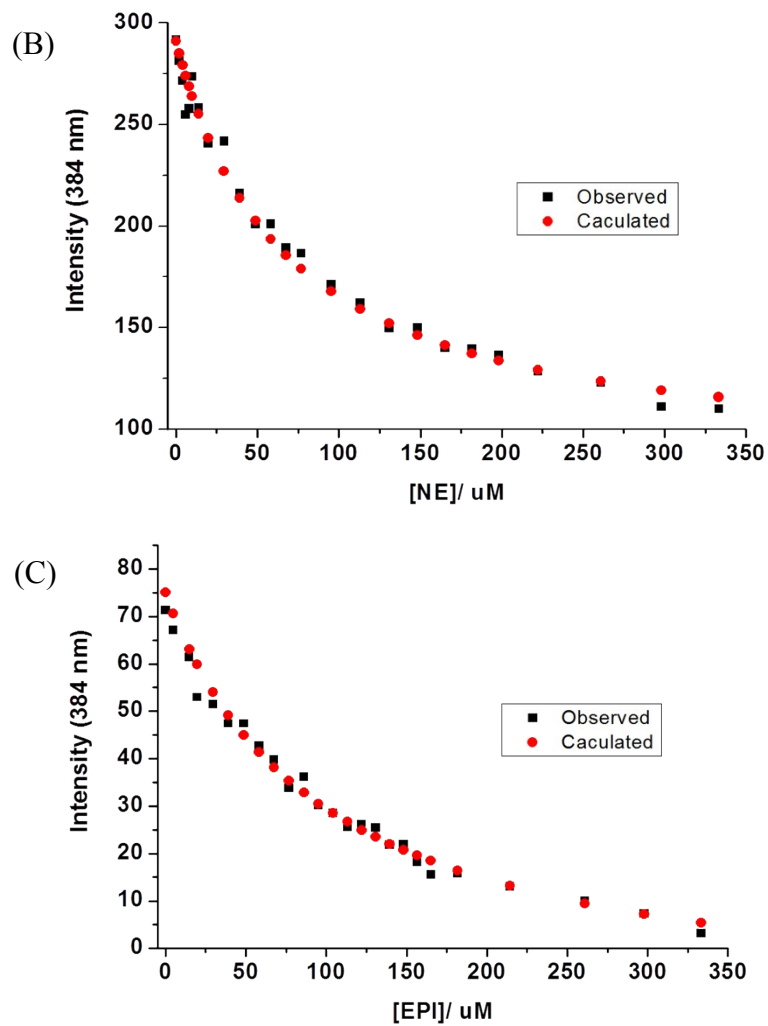




**Figure S12** Job's plots for 1:1 complex of sensor **NB** with (A) DA (B) NE at  $1 \times 10^{-5}$  M in DMSO:phosphate buffer (0.01 M, pH 7.4, 1:9, v/v)

#### 4. Non-linear regression plots





**Figure S13.** Non-linear regression plots of sensor **NB** ( $1 \times 10^{-5}$  M) toward 0-80 equiv. of (A) DA (B) NE and (C) EPI in DMSO:phosphate buffer (0.01 M, pH 7.4, 1:9, v/v) for calculation of binding constant ( $K_s$ )

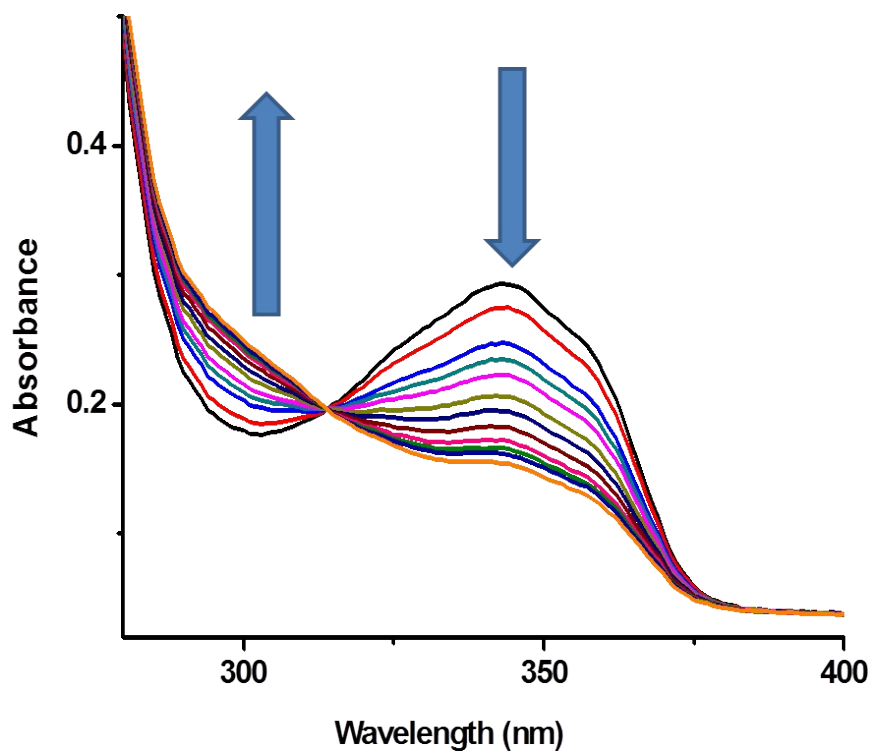
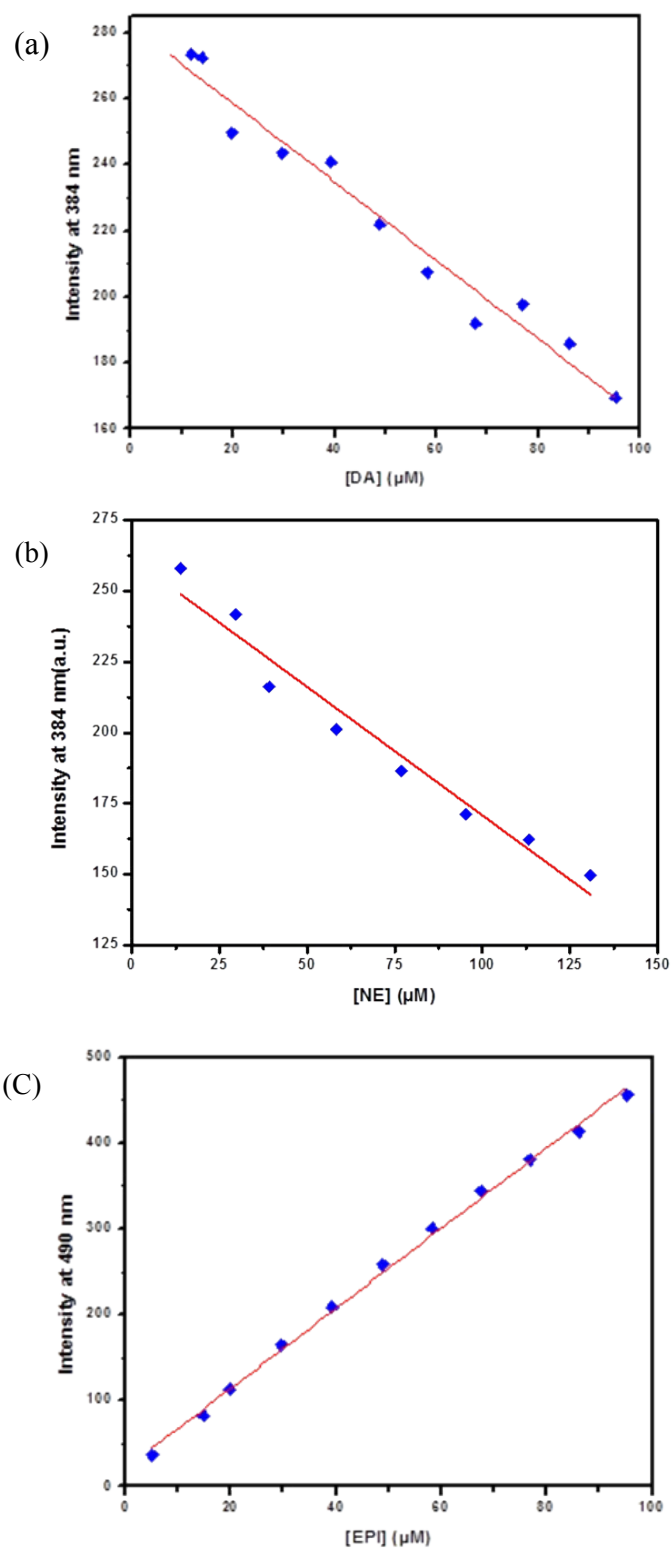


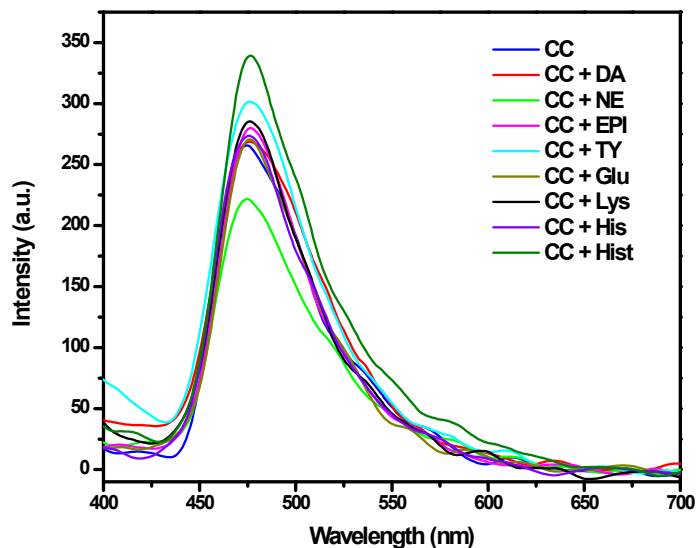
Figure S14. UV-vis spectra titration **NB** with EPI

5. Calibration curves of detection limit for DA, NE and EPI

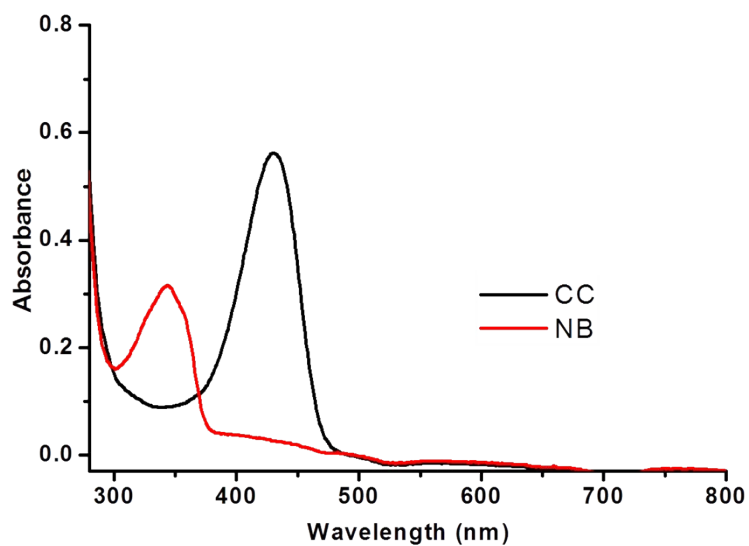


**Figure S15.** Linear plot of fluorescence intensity between sensor with a) DA b) NE c) EPI complexes and concentration of catecholamine guests

6. Complexation studies of sensor **CC** with various guests by fluorescence spectrophotometry technique

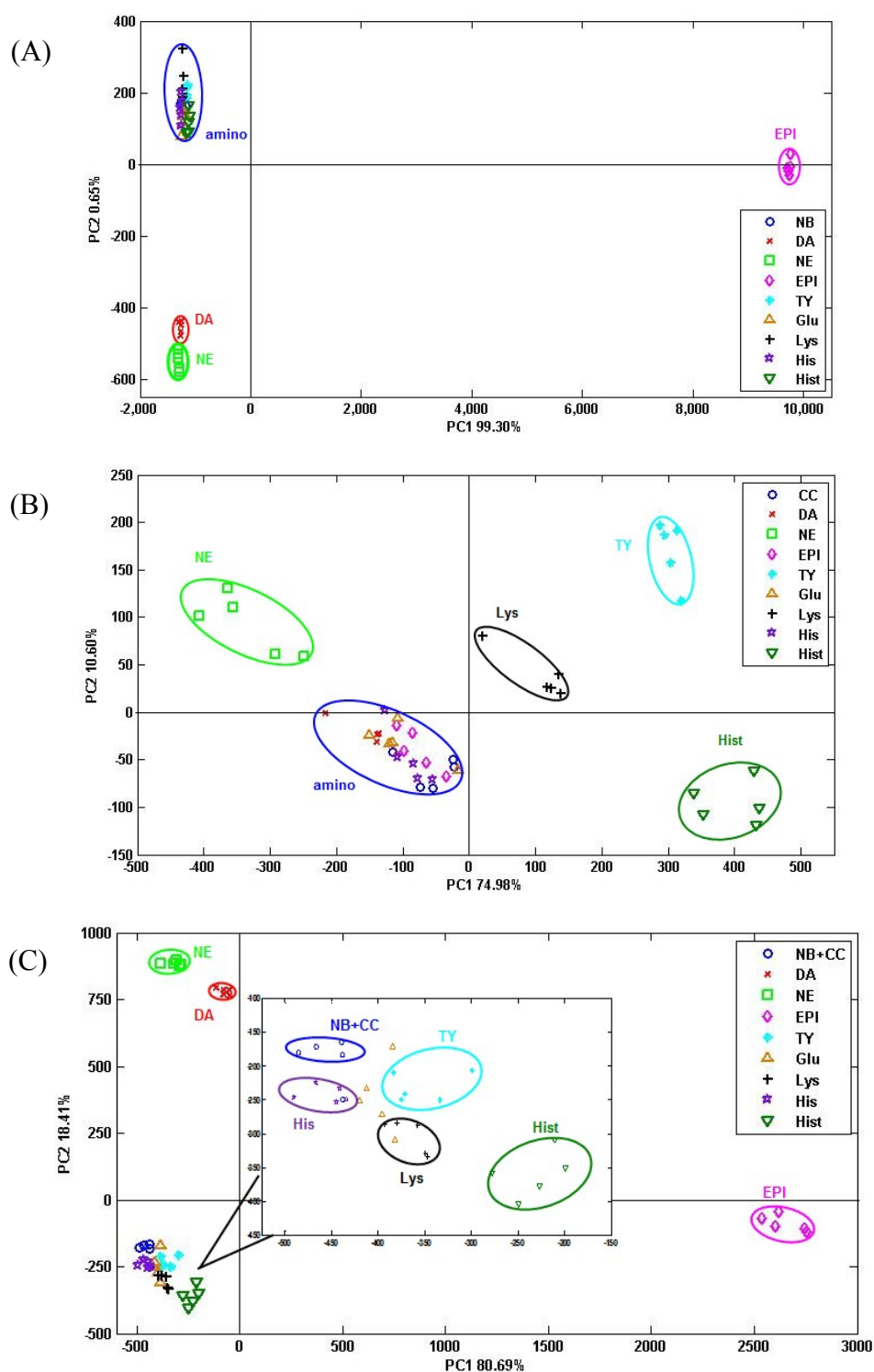


**Figure S16** Fluorescence spectra of sensor **CC** ( $5 \times 10^{-5}$  M) in DMSO:phosphate buffer (0.1 M, pH 7.4, 1:9, v/v) in the present of various guest 100 equiv. ( $\lambda_{\text{ex}} = 340$  nm)



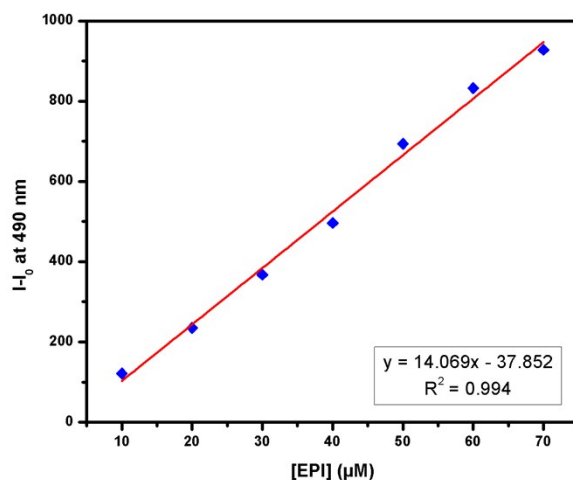
**Figure 17.** UV-vis spectra of **NB** and **CC**  $1 \times 10^{-4}$  M

7. Principle component analysis (PCA) method for analysis of complexation

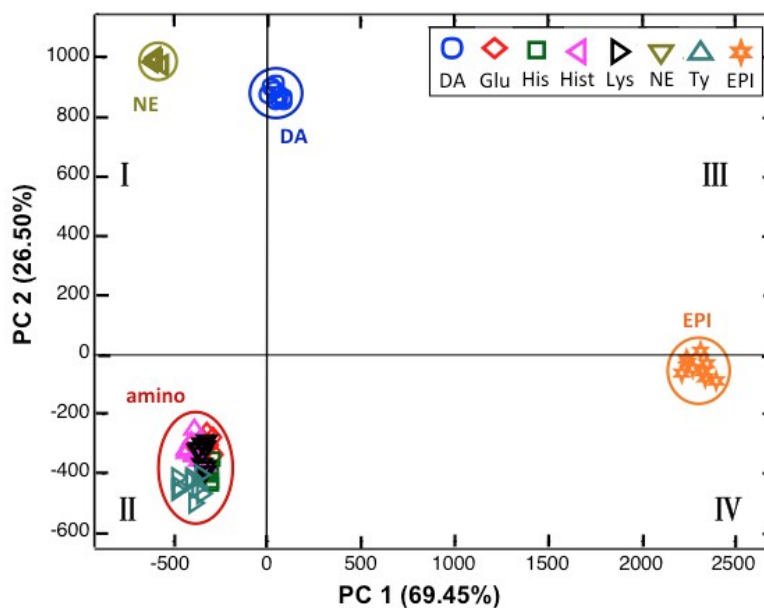


**Figure S18.** PCA score plot of (A) sensor NB (B) sensor CC and (C) the mixture sensors NB and CC upon addition of various guests (100 equiv.) in 1:9, v/v DMSO:phosphate buffer (pH 7.4). PCA score plot shows clustering for all 9 samples.

8. Complexation study of sensor **NB** with EPI in human urine sample



**Figure S19.** Calibration curve of sensor **NB** with the spiked EPI in the synthetic urine



**Figure S20.** PCA score plots of mixed **NB** and **CC** upon addition of various guests (100 equiv.) in urine samples and including fluorescence data of EPI for determining of PCA analysis

**Table S1** Composition of synthetic urine [1]

Species	Concentration (g l <sup>-1</sup> )	Concentration (mmol l <sup>-1</sup> )
CaCl <sub>2</sub> .H <sub>2</sub> O	0.65	Ca: 4.3
MgCl <sub>2</sub> .6H <sub>2</sub> O	0.651	Mg:3.2
NaCl	4.6	SO <sub>4</sub> : 16
Na <sub>2</sub> SO <sub>4</sub>	2.3	Citrate: 2.3
Na <sub>3</sub> C <sub>6</sub> H <sub>8</sub> O <sub>7</sub> .2H <sub>2</sub> O	0.65	Oxalate: 0.149
Na <sub>2</sub> C <sub>2</sub> O <sub>4</sub>	0.020	
KH <sub>2</sub> PO <sub>4</sub>	2.8	PO <sub>4</sub> : 20.5
KCl	1.6	
NH <sub>4</sub> Cl	1	NH <sub>4</sub> : 19
CO(NH <sub>2</sub> ) <sub>2</sub>	25	
C <sub>4</sub> H <sub>7</sub> N <sub>3</sub> O	1.1	
Total Na = 118 mEq		
Total K = 42 mEq		
pH = 5.8		

**Table S2** Comparison of the sensitivity of sensory system for dopamine

Sensing materials	Detection	Linear range	LOD	Ref.
MIP-Si-ITO <sup>a</sup> electrode	Electrochemistry	2x10 <sup>-6</sup> – 8x10 <sup>-4</sup> M.	2x10 <sup>-6</sup> M.	2
Au at SiO <sub>2</sub> -MIPs-GCE	Electrochemistry	4.8x10 <sup>-8</sup> -5x10 <sup>-5</sup> M.	2x10 <sup>-8</sup> M	3
C60-CNT/IL/GC electrode	Electrochemistry	0.06-25 μM	15 nM	4
Citrate-caped AuNPs	Colorimetric	2.5-20 μM	2500 nM	5
MBA-and DSP-modified AuNPs <sup>b</sup>	Colorimetric	5-180 nM	0.5 nM	6
Fe <sub>3</sub> O <sub>4</sub> NPs	Fluorescence	0.01-0.4 μM	3 μM	7
Self-assembled <b>PBA</b> and <b>CC</b> <sup>c</sup>	Fluorescence	16.7-47.4 μM	1.47 μM	8
Phosphate-modified TiO <sub>2</sub> NPs	Fluorescence	0.5-100 μM	30 nM	9
Napthalimide-boronic acid	Fluorescence	11.9-95.2 μM	7.71 μM	This work

<sup>a</sup>Si-ITO: silanized indium tin oxide electrode.

<sup>b</sup>**MBA**, 4-mercaptophenylboronic acid; **DSP**, dithiobis (succinimidyl propionate)

<sup>c</sup>**PBA**, Pyrene boronic acid; **CC**, crown-coumarin

[1] E. Tilley, J. Atwater, D. Mavrinic, Effects of storage on phosphorus recovery from urine, *Environ. Technol.* 29 (2007) 807-816.

[2] N. Gao, Z. Xu, F. Wang, S. Dong, *Electroanalysis*, 2007, **19**, 1655–1660.



- [3] D. Yu, Y. Zeng, Y. Qi, T. Zhou, G. Shi, *Biosen. Bioelectron.*, 2012, **38**, 270–277.
- [4] M. Mazloun-Ardakani, A. Khoshroo, *Electrochem. Commun.* 2014, **42**, 9–12.
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- [7] C.-H. Liu, C. J. Yu, W.-L. Tseng, *Anal. Chim. Acta*, 2012, **745**, 143-148.
- [8] A. Chaicham, S. Sahasithiwat, T. Tuntulani, B. Tomapatanaget, *Chem. Commun.*, 2013, **49**, 9287-9289
- [9] H.P. Wu, T.L. Chang, W.L. Tseng, *Langmuir*, 2007, **23** 7880-7885.