

Supporting Information

New α -Glucosidase Inhibitors from a Marine Sponge-derived Fungus

Aspergillus sp. OUCMDZ-1583

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List of Supporting Information

Bioassay Protocols	S5
The ITS sequences data of <i>Aspergillus</i> sp. OUCMDZ-1583	S7
Theory and Calculation Details	S8
Table S1. ¹ H (500 MHz) and ¹³ C (125MHz) NMR data for compounds 19–23	S9
Figure S1. The ¹ H-NMR spectrum of compound 1 in CDCl ₃	S10
Figure S2. The DEPTQ spectrum of compound 1 in CDCl ₃	S11
Figure S3. The HMQC spectrum of compound 1 in CDCl ₃	S12
Figure S4. The HMBC spectrum of compound 1 in CDCl ₃	S13
Figure S5. The ¹ H- ¹ H COSY spectrum of compound 1 in CDCl ₃	S14
Figure S6. The NOE difference spectrum of compound 1 in CDCl ₃	S15
Figure S7. The ¹ H-NMR spectrum of compound 2 in CDCl ₃	S16
Figure S8. The ¹³ C-NMR spectrum of compound 2 in CDCl ₃	S17
Figure S9. The DEPT spectrum of compound 2 in CDCl ₃	S18
Figure S10. The HMQC spectrum of compound 2 in CDCl ₃	S19
Figure S11. The ¹ H- ¹ H COSY spectrum of compound 2 in CDCl ₃	S20
Figure S12. The HMBC spectrum of compound 2 in CDCl ₃	S21
Figure S13. The NOE difference spectrum of compound 2 in CDCl ₃	S22
Figure S14. The ¹ H-NMR spectrum of compound 3 and 4 in CDCl ₃	S23
Figure S15. The ¹³ C-NMR spectrum of compound 3 and 4 in CDCl ₃	S24
Figure S16. The DEPT spectrum of compound 3 and 4 in CDCl ₃	S25

Figure S17. The HMQC spectrum of compound 3 and 4 in CDCl ₃	S26
Figure S18. The ¹ H- ¹ H COSY spectrum of compound 3 and 4 in CDCl ₃	S27
Figure S19. The HMBC spectrum of compound 3 and 4 in CDCl ₃	S28
Figure S20. The NOE difference spectrum of compound 3 and 4 in CDCl ₃	S29
Figure S21. The ¹ H-NMR spectrum of compound 5 in CDCl ₃	S30
Figure S22. The ¹³ C-NMR spectrum of compound 5 in CDCl ₃	S31
Figure S23. The DEPT spectrum of compound 5 in CDCl ₃	S32
Figure S24. The HMQC spectrum of compound 5 in CDCl ₃	S33
Figure S25. The ¹ H- ¹ H COSY spectrum of compound 5 in CDCl ₃	S34
Figure S26. The HMBC spectrum of compound 5 in CDCl ₃	S35
Figure S27. The NOE difference spectrum of compound 5 in CDCl ₃ (I).....	S36
Figure S28. The NOE difference spectrum of compound 5 in CDCl ₃ (II).....	S37
Figure S29. The ¹ H-NMR spectrum of compound 6 in DMSO- <i>d</i> ₆	S38
Figure S30. The ¹³ C-NMR spectrum of compound 6 in DMSO- <i>d</i> ₆	S39
Figure S31. The DEPT spectrum of compound 6 in DMSO- <i>d</i> ₆	S40
Figure S32. The HMQC spectrum of compound 6 in DMSO- <i>d</i> ₆	S41
Figure S33. The ¹ H- ¹ H COSY spectrum of compound 6 in DMSO- <i>d</i> ₆	S42
Figure S34. The HMBC spectrum of compound 6 in DMSO- <i>d</i> ₆	S43
Figure S35. The NOE difference spectrum of compound 6 in DMSO- <i>d</i> ₆ (I).....	S44
Figure S36. The NOE difference spectrum of compound 6 in DMSO- <i>d</i> ₆ (II).....	S45
Figure S37. The ¹ H-NMR spectrum of compound 7 and 8 in CDCl ₃	S46
Figure S38. The ¹³ C-NMR spectrum of compound 7 and 8 in CDCl ₃	S47
Figure S39. The DEPT spectrum of compound 7 and 8 in CDCl ₃	S48
Figure S40. The HMQC spectrum of compound 7 and 8 in CDCl ₃	S49
Figure S41. The ¹ H- ¹ H COSY spectrum of compound 7 and 8 in CDCl ₃	S50
Figure S42. The HMBC spectrum of compound 7 and 8 in CDCl ₃	S51
Figure S43. The NOE difference spectrum of compound 7 and 8 in CDCl ₃	S52
Figure S44. The ¹ H-NMR spectrum of compound 9 in DMSO- <i>d</i> ₆	S53
Figure S45. The ¹³ C-NMR spectrum of compound 9 in DMSO- <i>d</i> ₆	S54
Figure S46. The DEPT spectrum of compound 9 in DMSO- <i>d</i> ₆	S55
Figure S47. The HMQC spectrum of compound 9 in DMSO- <i>d</i> ₆	S56
Figure S48. The ¹ H- ¹ H COSY spectrum of compound 9 in DMSO- <i>d</i> ₆	S57
Figure S49. The HMBC spectrum of compound 9 in DMSO- <i>d</i> ₆	S58
Figure S50. The ¹ H-NMR spectrum of compound 10 in DMSO- <i>d</i> ₆	S59
Figure S51. The ¹³ C-NMR spectrum of compound 10 in DMSO- <i>d</i> ₆	S60

Figure S52. The DEPT spectrum of compound 10 in DMSO- <i>d</i> ₆	S61
Figure S53. The HMQC spectrum of compound 10 in DMSO- <i>d</i> ₆	S62
Figure S54. The ¹ H- ¹ H COSY spectrum of compound 10 in DMSO- <i>d</i> ₆	S63
Figure S55. The HMBC spectrum of compound 10 in DMSO- <i>d</i> ₆	S64
Figure S56. The ¹ H-NMR spectrum of compound 11 in DMSO- <i>d</i> ₆	S65
Figure S57. The ¹³ C-NMR spectrum of compound 11 in DMSO- <i>d</i> ₆	S66
Figure S58. The DEPT spectrum of compound 11 in DMSO- <i>d</i> ₆	S67
Figure S59. The HMQC spectrum of compound 11 in DMSO- <i>d</i> ₆	S68
Figure S60. The ¹ H- ¹ H COSY spectrum of compound 11 in DMSO- <i>d</i> ₆	S69
Figure S61. The HMBC spectrum of compound 11 in DMSO- <i>d</i> ₆	S70
Figure S62. The ¹ H-NMR spectrum of compound 12 in DMSO- <i>d</i> ₆	S71
Figure S63. The ¹³ C-NMR spectrum of compound 12 in DMSO- <i>d</i> ₆	S72
Figure S64. The DEPT spectrum of compound 12 in DMSO- <i>d</i> ₆	S73
Figure S65. The HMQC spectrum of compound 12 in DMSO- <i>d</i> ₆	S74
Figure S66. The ¹ H- ¹ H COSY spectrum of compound 12 in DMSO- <i>d</i> ₆	S75
Figure S67. The HMBC spectrum of compound 12 in DMSO- <i>d</i> ₆	S76
Figure S68. The NOESY spectrum of compound 12 in DMSO- <i>d</i>	S77
Figure S69. The ¹ H-NMR spectrum of compound 13 in DMSO- <i>d</i> ₆	S78
Figure S70. The ¹³ C-NMR spectrum of compound 13 in DMSO- <i>d</i> ₆	S79
Figure S71. The DEPT spectrum of compound 13 in DMSO- <i>d</i> ₆	S80
Figure S72. The HMQC spectrum of compound 13 in DMSO- <i>d</i> ₆	S81
Figure S73. The ¹ H- ¹ H COSY spectrum of compound 13 in DMSO- <i>d</i> ₆	S82
Figure S74. The HMBC spectrum of compound 13 in DMSO- <i>d</i> ₆	S83
Figure S75. The NOESY spectrum of compound 13 in DMSO- <i>d</i>	S84
Figure S76. The ¹ H-NMR spectrum of compound 14 in DMSO- <i>d</i> ₆	S85
Figure S77. The ¹³ C-NMR spectrum of compound 14 in DMSO- <i>d</i> ₆	S86
Figure S78. The DEPT spectrum of compound 14 in DMSO- <i>d</i> ₆	S87
Figure S79. The HMQC spectrum of compound 14 in DMSO- <i>d</i> ₆	S88
Figure S80. The ¹ H- ¹ H COSY spectrum of compound 14 in DMSO- <i>d</i> ₆	S89
Figure S81. The HMBC spectrum of compound 14 in DMSO- <i>d</i> ₆	S90
Figure S82. The ¹ H-NMR spectrum of compound 15 in DMSO- <i>d</i> ₆	S91
Figure S83. The ¹³ C-NMR spectrum of compound 15 in DMSO- <i>d</i> ₆	S92
Figure S84. The DEPT spectrum of compound 15 in DMSO- <i>d</i> ₆	S93
Figure S85. The HMQC spectrum of compound 15 in DMSO- <i>d</i> ₆	S94
Figure S86. The ¹ H- ¹ H COSY spectrum of compound 15 in DMSO- <i>d</i> ₆	S95

Figure S87. The HMBC spectrum of compound 15 in DMSO- <i>d</i> ₆	S96
Figure S88. The ¹ H-NMR spectrum of compound 16 in DMSO- <i>d</i> ₆	S97
Figure S89. The ¹³ C-NMR spectrum of compound 16 in DMSO- <i>d</i> ₆	S98
Figure S90. The DEPT spectrum of compound 16 in DMSO- <i>d</i> ₆	S99
Figure S91. The HMQC spectrum of compound 16 in DMSO- <i>d</i> ₆	S100
Figure S92. The ¹ H- ¹ H COSY spectrum of compound 16 in DMSO- <i>d</i> ₆	S101
Figure S93. The HMBC spectrum of compound 16 in DMSO- <i>d</i> ₆	S102
Figure S94. The ¹ H-NMR spectrum of compound 17 in DMSO- <i>d</i> ₆	S103
Figure S95. The ¹³ C-NMR spectrum of compound 17 in DMSO- <i>d</i> ₆	S104
Figure S96. The DEPT spectrum of compound 17 in DMSO- <i>d</i> ₆	S105
Figure S97. The HMQC spectrum of compound 17 in DMSO- <i>d</i> ₆	S106
Figure S98. The ¹ H- ¹ H COSY spectrum of compound 17 in DMSO- <i>d</i> ₆	S107
Figure S99. The HMBC spectrum of compound 17 in DMSO- <i>d</i> ₆	S108
Figure S100. The ¹ H-NMR spectrum of compound 18 in CDCl ₃	S109
Figure S101. The ¹³ C-NMR spectrum of compound 18 in CDCl ₃	S110
Figure S102. The DEPT spectrum of compound 18 in CDCl ₃	S111
Figure S103. The HMQC spectrum of compound 18 in CDCl ₃	S112
Figure S104. The ¹ H- ¹ H COSY spectrum of compound 18 in CDCl ₃	S113
Figure S105. The HMBC spectrum of compound 18 in CDCl ₃	S114
Figure S106. The NOESY spectrum of compound 18 in CDCl ₃	S115
Figure S107. X-ray data and structure of compound 20	S116
Figure S108. The ¹ H-NMR spectrum of compound 12a in CDCl ₃	S117
Figure S109. The DEPTQ spectrum of compound 12a in CDCl ₃	S118
Figure S110. The HMQC spectrum of compound 12a in CDCl ₃	S119
Figure S111. The ¹ H- ¹ H COSY spectrum of compound 12a in CDCl ₃	S120
Figure S112. The HMBC spectrum of compound 12a in CDCl ₃	S121
Figure S113. The ¹ H-NMR spectrums of compound 2a and 2b in CDCl ₃	S122
Figure S114. The ¹ H-NMR spectrums of compound 3a and 3b in CDCl ₃	S123
Figure S115. The ¹ H-NMR spectrums of compound 5a and 5b in CDCl ₃	S124
Figure S116. The ¹ H-NMR spectrums of compound 6a and 6b in CDCl ₃	S125
Figure S117. The ¹ H-NMR spectrums of compound 7a and 7b in CDCl ₃	S126
Figure S118. The ¹ H-NMR spectrums of compound 12aa and 12ab in CDCl ₃	S127
Figure S119. The HPLC analysis and ESIMS of the reaction liquid of 12 to 12a	S128
Figure S120. The HPLC analysis of compound 16 and synthetic 15	S129
Figure S121. The HPLC analysis of natural 14 and synthetic 14	S129

Anti-influenza A Virus (H1N1) Assay

The antiviral activity against H1N1 was evaluated by the CPE inhibition assay.⁶ Confluent MDCK cell monolayers were firstly incubated with influenza virus (A/Puerto Rico/8/34 (H1N1), PR/8) at 37 °C for 1 h. After removing the virus dilution, cells were maintained in infecting media (RPMI 1640, 4 µg/mL of trypsin) containing different concentrations of test compounds at 37 °C. After 48 h incubation at 37 °C, the cells were fixed with 100 µL of 4% formaldehyde for 20 min at room temperature. After removal of the formaldehyde, the cells were stained with 0.1% crystal violet for 30 min. The plates were washed and dried, and the intensity of crystal violet staining for each well was measured in a microplate reader (Bio-Rad, USA) at 570 nm. The IC₅₀ value was calculated as the compound concentration required inhibiting influenza virus yield at 48 h post-infection by 50%. Ribavirin was used as the positive control with an IC₅₀ value of 137.3 ± 0.4 µM.

α-Glucosidase Inhibitory Effect Assay

The inhibitory effects were assayed as described preciously.²² The sample was dissolved in sodium phosphate buffer (PBS, pH 6.8) at three concentrations. A volume of 10 µL of the sample solution, 20 µL of PBS and 20 µL of 2.5 mM *p*-nitrophenyl-α-D-glucopyranoside (PNPG) solution (in phosphate buffer) were mixed in a 96-well microplate and incubated at 37 °C for 5 min. A volume of 10 µL of α-glucosidase diluted to 0.2 U/mL by 0.01 M PBS was then added to each well. After incubating at 37 °C for 15 min, the absorbance at 405 nm was recorded by a Spectra max 190 micro plate reader (Molecular Devices Inc.). The blank was prepared by adding phosphate buffer instead of the α-glucosidase and acarbose was used as a positive control. Blank readings (no enzyme) were subtracted from each well and results were compared to the control. The inhibition (%) was calculated as $[1 - (OD_{\text{sample}}/OD_{\text{control}})] \times 100$ %. The IC₅₀ value was calculated as the compound concentration that is required for 50% inhibition and the IC₅₀ value of the acarbose was 0.95 mM.

Antimicrobial Assays

The antimicrobial activities against *Escherichia coli*, *Enterobacter aerogenes*, *Pseudomonas aeruginosa*, *Bacillus subtilis*, *Staphylococcus aureus*, and *Candida albicans* were evaluated by an agar dilution method.²³ The tested strains were cultivated in LB agar plates for bacteria and in YPD agar plates for *C. albicans* at 37 °C. Compounds **1–23** and positive controls (ciprofloxacin lactate for bacteria and ketoconazole for *C. albicans*) were dissolved in MeOH at different concentrations from 100 to 0.05 µg/mL by the continuous 2-fold dilution methods. A 10 µL quantity of test solution was

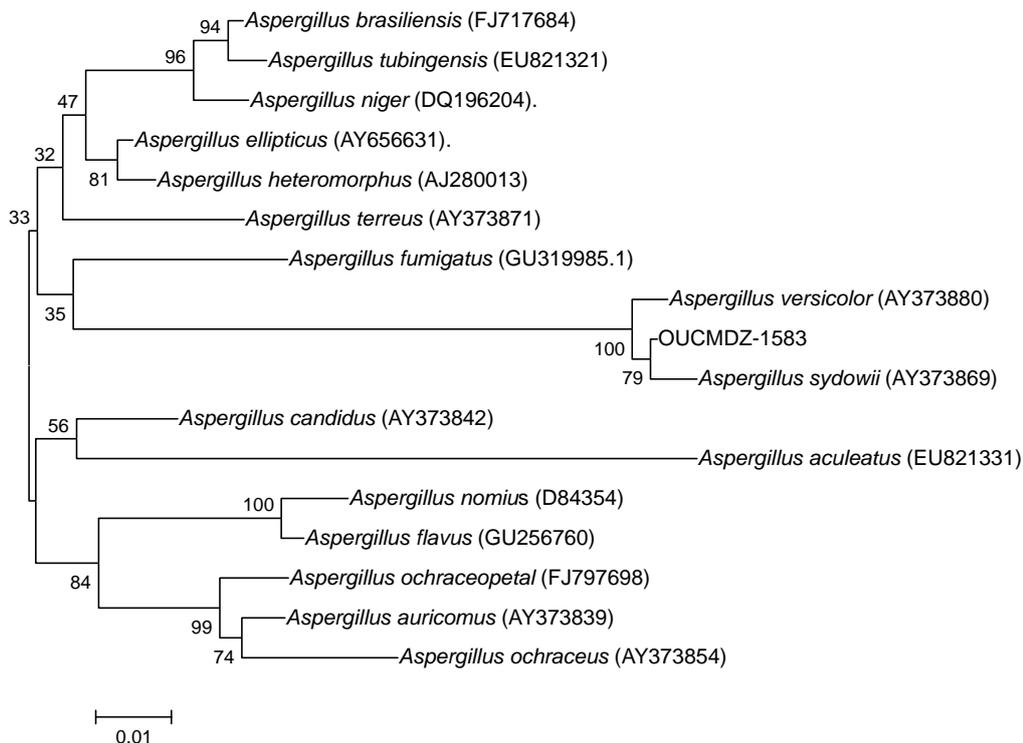
absorbed by a paper disk (5 mm diameter) and placed on the assay plates. After 24 h incubation, zones of inhibition (mm in diameter) were recorded. The minimum inhibitory concentrations (MICs) were defined as the lowest concentration at which no microbial growth could be observed.

Cytotoxic Assays

Cytotoxicity was assayed by the MTT²⁴ and CCK-8 methods.²⁵ In the MTT assay, A549 and MCF-7 cell lines were grown in RPMI-1640 supplemented with 10% FBS under a humidified atmosphere of 5% CO₂ and 95% air at 37 °C, respectively. Cell suspension, 100 μL, at a density of 3×10^4 cell mL⁻¹ was plated in 96-well microtiter plates, allowed to attach overnight, and then exposed to varying concentrations (10^{-5} - 10^{-12} M) of compounds for 72 h. The MTT solution (20 μL, 5 mg/mL in RPMI-1640 medium) was then added to each well and incubated for 4 h. Old medium containing MTT was then gently replaced by DMSO and pipetted to dissolve any formazan crystals formed. Absorbance was then determined on a Spectra Max Plus plate reader at 540 nm. In the CCK-8 assay, K562 cell line was grown in RPMI-1640 supplemented with 10% FBS under a humidified atmosphere of 5% CO₂ and 95% air at 37 °C. Cell suspension, 100 μL, at a density of 5×10^5 cell mL⁻¹ was plated in 96-well microtiter plates and then exposed to varying concentrations (10^{-5} - 10^{-12} M) of compounds after cultivation for 24 h. Three days later, 10 μL of CCK-8 solution was added 4 h before detection. Then the absorbency (A450 value) was measured, and the growth rates of cells were computed.

ITS sequences of *Aspergillus* sp. OUCMDZ-1583

CGTAGGTGAACCTGCGGAAGGATCATTACTGAGTGCGGGCTGCCTCCGGGCGCCCAACCTCCCACCCG
TGAATACCTAACACTGTTGCTTCGGCGGGGAACCCCTCGGGGGCGAGCCGCCGGGGACTACTGAACT
TCATGCCTGAGAGTGATGCAGTCTGAGTCTGAATATAAAATCAGTCAAAACTTTCAACAATGGATCTCT
TGGTTCCGGCATCGATGAAGAACGCAGCGAACTGCGATAAGTAATGTGAATTGCAGAATTCAGTGAAT
CATCGAGTCTTTGAACGCACATTGCGCCCCCTGGCATTCCGGGGGGCATGCCTGTCCGAGCGTCATTGC
TGCCCATCAAGCCCGGCTTGTGTGTTGGGTCGTCGTCACCCCCCGGGGGACGGGCCCCGAAAGGCAGC
GGCGGCACCGTGTCCGGTCTCGAGCGTATGGGGCTTTGTCACCCGCTCGACTAGGGCCGGCCGGGCG
CCAGCCGACGTCTCCAACCATTTTTCTTCAGGTTGACCTCGGATCAGGTAGGGATAACCCGCTGAACTTA
AGCATATCAATAAGCGGAG



Neighbor-joining phylogenetic tree of strain OUCMDZ-1583 based on ITS gene sequences (ca. 565 bp). The values at each node represent the bootstrap values from 1000 replicates, and the scale bar represents 0.0005 substitutions per nucleotide. Phylogenetic analyses were conducted in MEGA4.

Theory and Calculation Details. The calculations were performed by using the density functional theory (DFT) as carried out in the Gaussian 03.^{S1} The preliminary conformational distributions search was performed by HyperChem 7.5 software. All ground-state geometries were optimized at the B3LYP/6-31G(d) level. Solvent effects of methanol solution were evaluated at the same DFT level by using the SCRF/PCM method.^{S2} TDDFT^{S3} at B3LYP/6-31G(d) was employed to calculate the electronic excitation energies and rotational strengths in methanol.

(S1) Gaussian 03, Revision E.01, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, J. A. Montgomery, Jr., T. Vreven, K. N. Kudin, J. C. Burant, J. M. Millam, S. S. Iyengar, J. Tomasi, V. Barone, B. Mennucci, M. Cossi, G. Scalmani, N. Rega, G. A. Petersson, H. Nakatsuji, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, M. Klene, X. Li, J. E. Knox, H. P. Hratchian, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, P. Y. Ayala, K. Morokuma, G. A. Voth, P. Salvador, J. J. Dannenberg, V. G. Zakrzewski, S. Dapprich, A. D. Daniels, M. C. Strain, O. Farkas, D. K. Malick, A. D. Rabuck, K. Raghavachari, J. B. Foresman, J. V. Ortiz, Q. Cui, A. G. Baboul, S. Clifford, J. Cioslowski, B. B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R. L. Martin, D. J. Fox, T. Keith, M. A. Al-Laham, C. Y. Peng, A. Nanayakkara, M. Challacombe, P. M. W. Gill, B. Johnson, W. Chen, M. W. Wong, C. Gonzalez and J. A. Pople, Gaussian, Inc., Wallingford CT, 2004.

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(S3) (a) M. E. Casida, In *Recent Advances in Density Functional Methods*, part I; D. P. Chong, World Scientific: Singapore, 1995; pp 155–192. (b) E. K. U. Gross, J. F. Dobson and M. Petersilka, *Top. Curr. Chem.*, 1996, **181**, 81–172. (c) E. K. U. Gross and W. Kohn, *Adv. Quantum Chem.*, 1990, **21**, 255–291. (d) E. Runge and E. K. U. Gross, *Phys. Rev. Lett.*, 1984, **52**, 997–1000.

Table S1. ^1H (500 MHz) and ^{13}C (125 MHz) NMR Data of Compounds **19–23**

position	19^a		20^b		21^b		22^a		23^a	
	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)	δ_{C}	δ_{H} (J in Hz)
1	128.1, C		168.0, C		167.8, C		170.2, C		169.6, C	
2	148.9, C									
3	195.1, C		81.7, CH	5.05, br d	81.2, CH	5.03, m	77.5, CH	4.73, m	79.8, CH	4.67, m
4	52.5, CH	3.47, d (4.0)	74.5, CH	4.52, br d	74.4, CH	4.52, d (3.0)	42.8, CH ₂	2.90, dd (16.0, 4.0); 2.83, dd (16.0, 12.0)	65.7, CH	4.46, br d (2.5)
4a			127.0, C		131.2, C		131.7, C		136.1, C	
5	55.8, CH	3.74, d (1.6, 4.0)	104.2, CH	6.59, s	104.4, CH	6.57, s	103.0, CH	6.51, s	103.9, CH	6.72, s
6	61.9, CH	4.86, d (1.6)	152.3, C		158.6, C		154.0, C		158.8, C	
7	128.1, CH	6.60, d (16.2)	134.3, C		137.2, C		132.8, C		139.0, C	
8	136.2, CH	6.47, dq (16.2, 6.8)	149.5, C		156.2, C		150.6, C		155.2, C	
8a			101.8, C		102.0, C		102.6, C		102.3, C	
9	19.7, CH	1.87, d (6.8)	19.2, CH ₂	2.57, m; 2.13, m	19.2, CH ₂	2.57, m; 2.13, m	33.0, CH ₂	1.81, ddd (14.7, 10.4, 2.8); 1.56, ddd (14.7, 10.8, 3.6)	38.6, CH ₂	1.94, m; 1.58, m
10	53.3, CH ₂	4.07, d (11.3); 4.27, d (11.3)	78.7, CH	4.09, m	78.7, CH	4.10, m	65.7, CH	3.70, m	66.1, CH	3.70, m
11			39.1, CH ₂	1.67, m; 1.54, m	39.8, CH ₂	1.67, m; 1.52, m	40.4, CH ₂	1.38, m; 1.35, m	40.5, CH ₂	1.40, m; 1.38, m
12			38.1, CH ₂	1.39, m; 1.34, m	38.0, CH ₂	1.40, m; 1.31, m	18.9, CH ₂	1.33, m; 1.30, m	18.9, CH ₂	1.32, m; 1.37, m
13			14.1, CH ₃	0.88, t (7.5)	19.2, CH ₃	0.89, t (7.4)	14.4, CH ₃	0.86, t (7.2)	14.6, CH ₃	0.89, t (7.1)
6-OCH ₃			56.4, CH ₃	3.93, s	56.2, CH ₃	3.87, s	56.5, CH ₃	3.83, s	56.7, CH	3.88, s
7-OCH ₃					60.7, CH ₃	3.93, s			60.5, CH ₃	3.70, s
8-OH				11.11, s		11.3, br s		10.87, br s		11.03, br s

^a recorded in DMSO-*d*₆. ^b recorded in CDCl₃.

Figure S1. The ^1H -NMR spectrum of compound **1** in CDCl_3

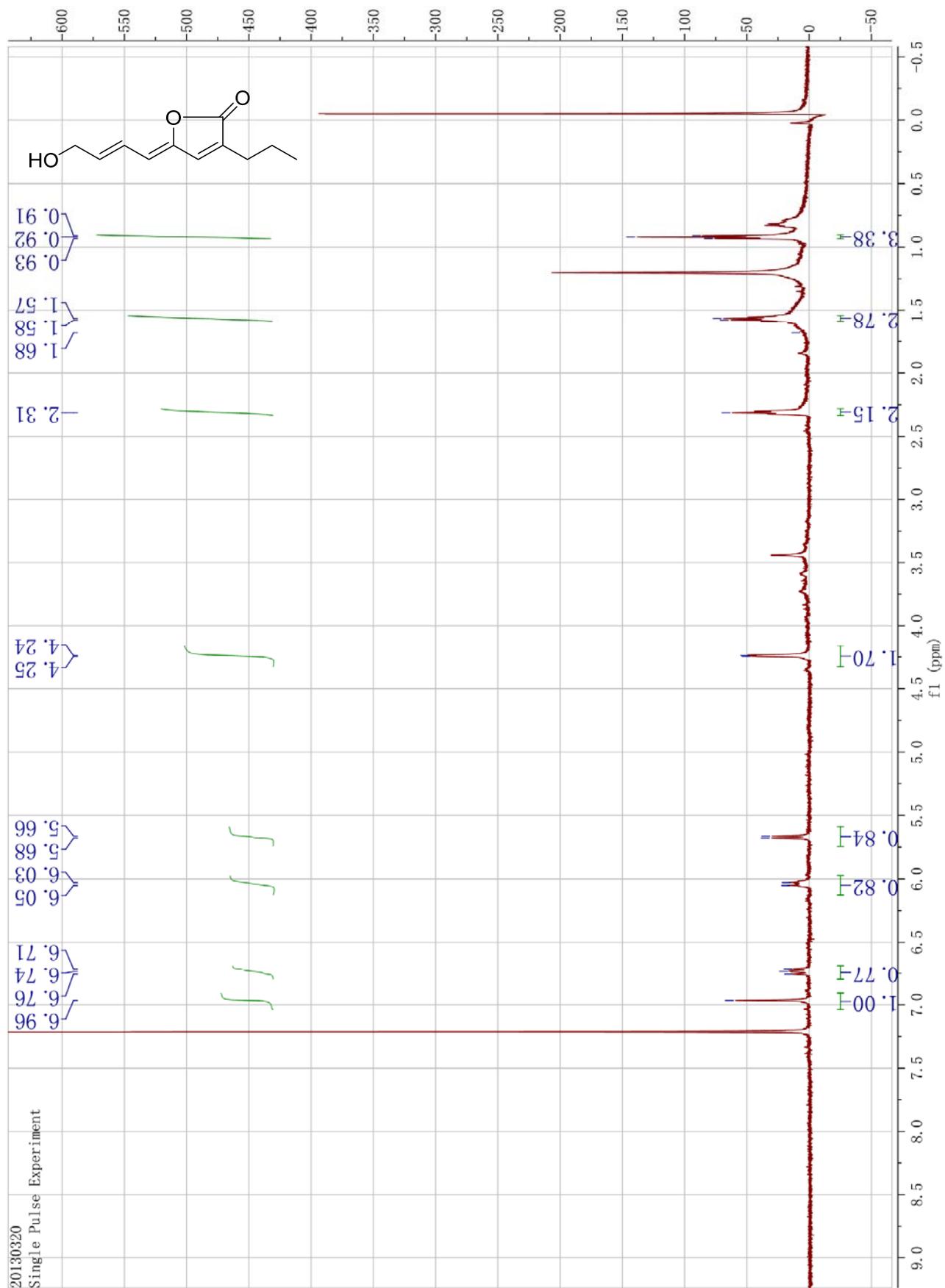


Figure S2. The DEPTQ spectrum of compound **1** in CDCl₃

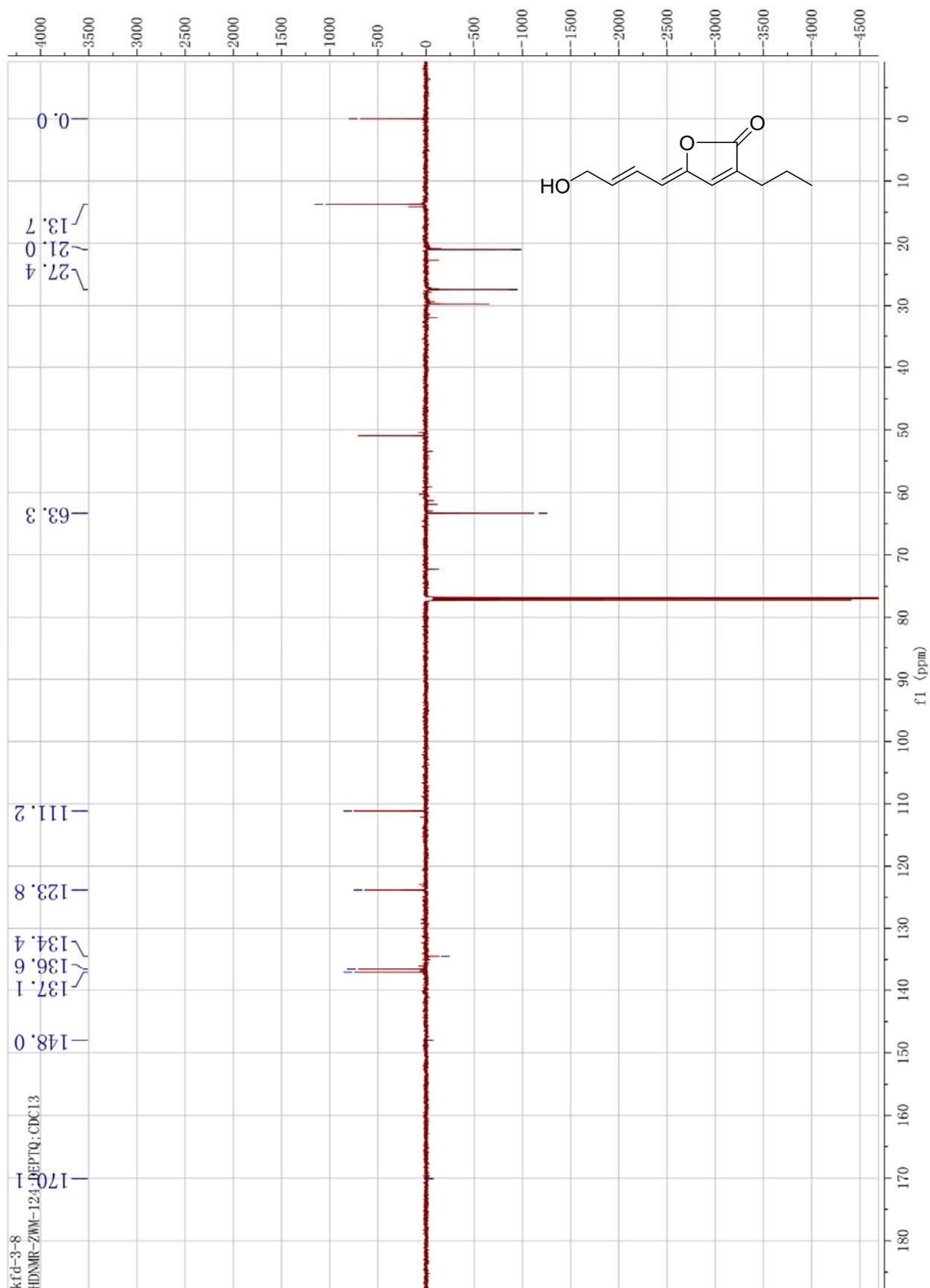


Figure S3. The HSQC spectrum of compound 1 in CDCl_3

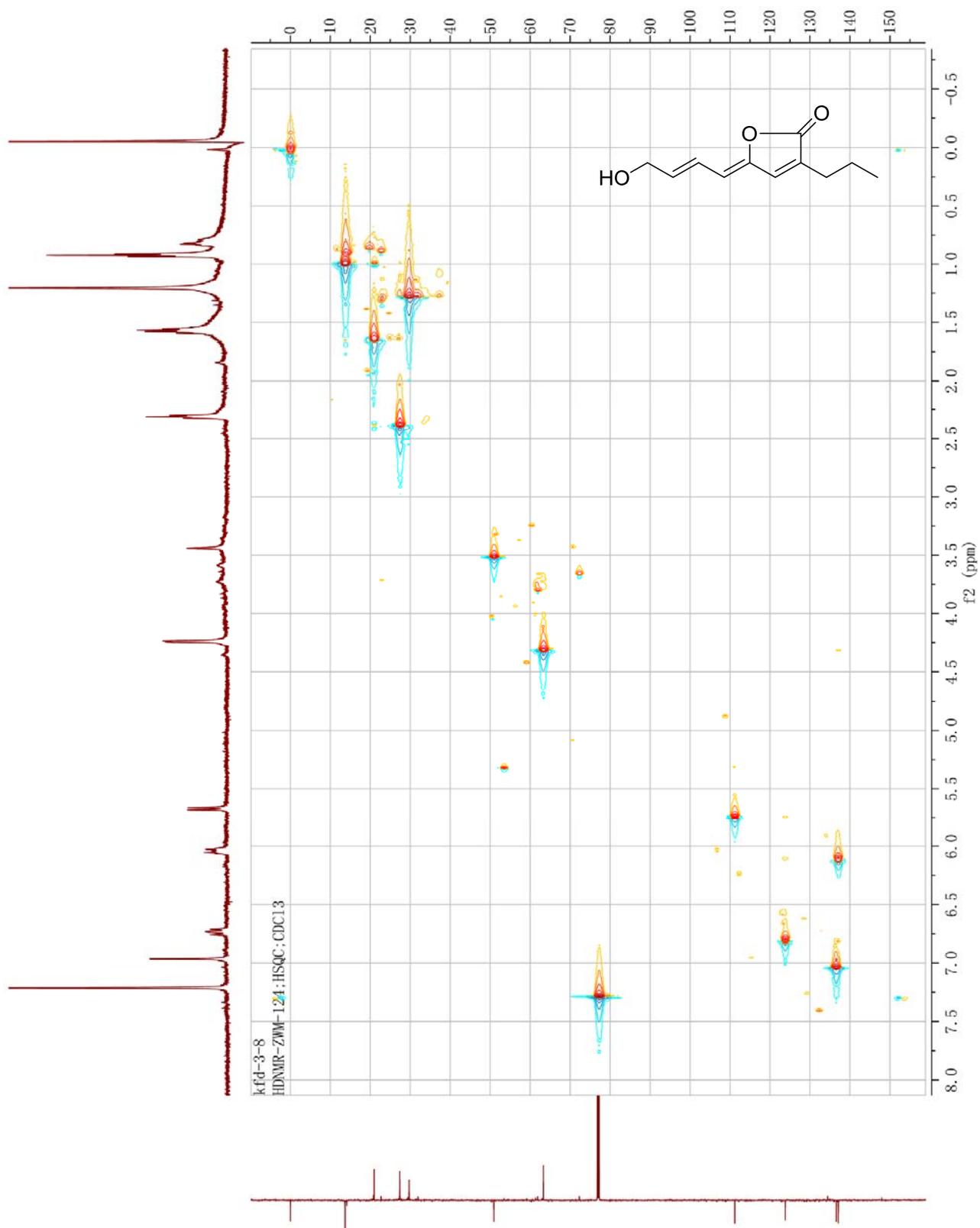


Figure S4. The HMBC spectrum of compound **1** in CDCl₃

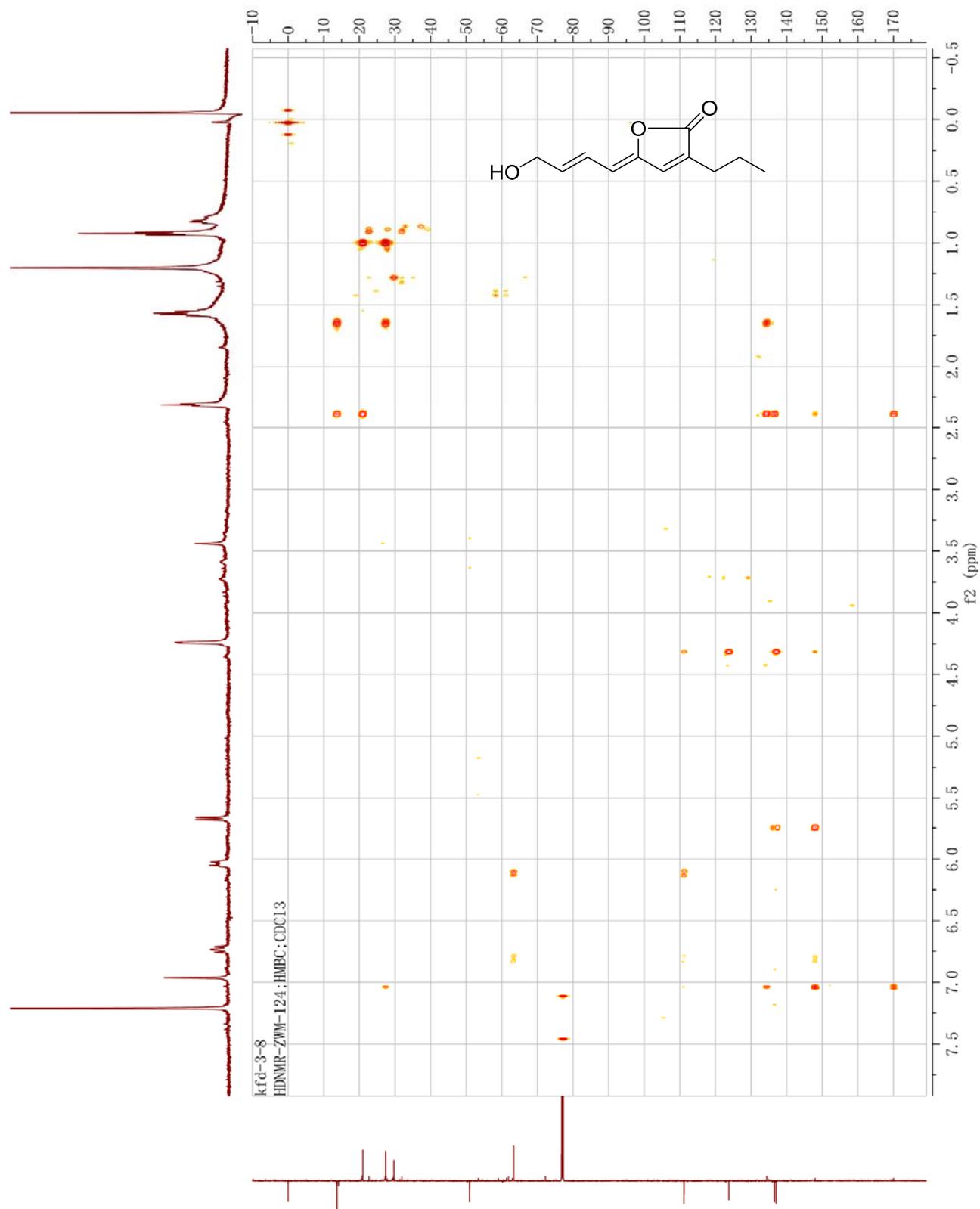


Figure S5. The ^1H - ^1H COSY spectrum of compound **1** in CDCl_3

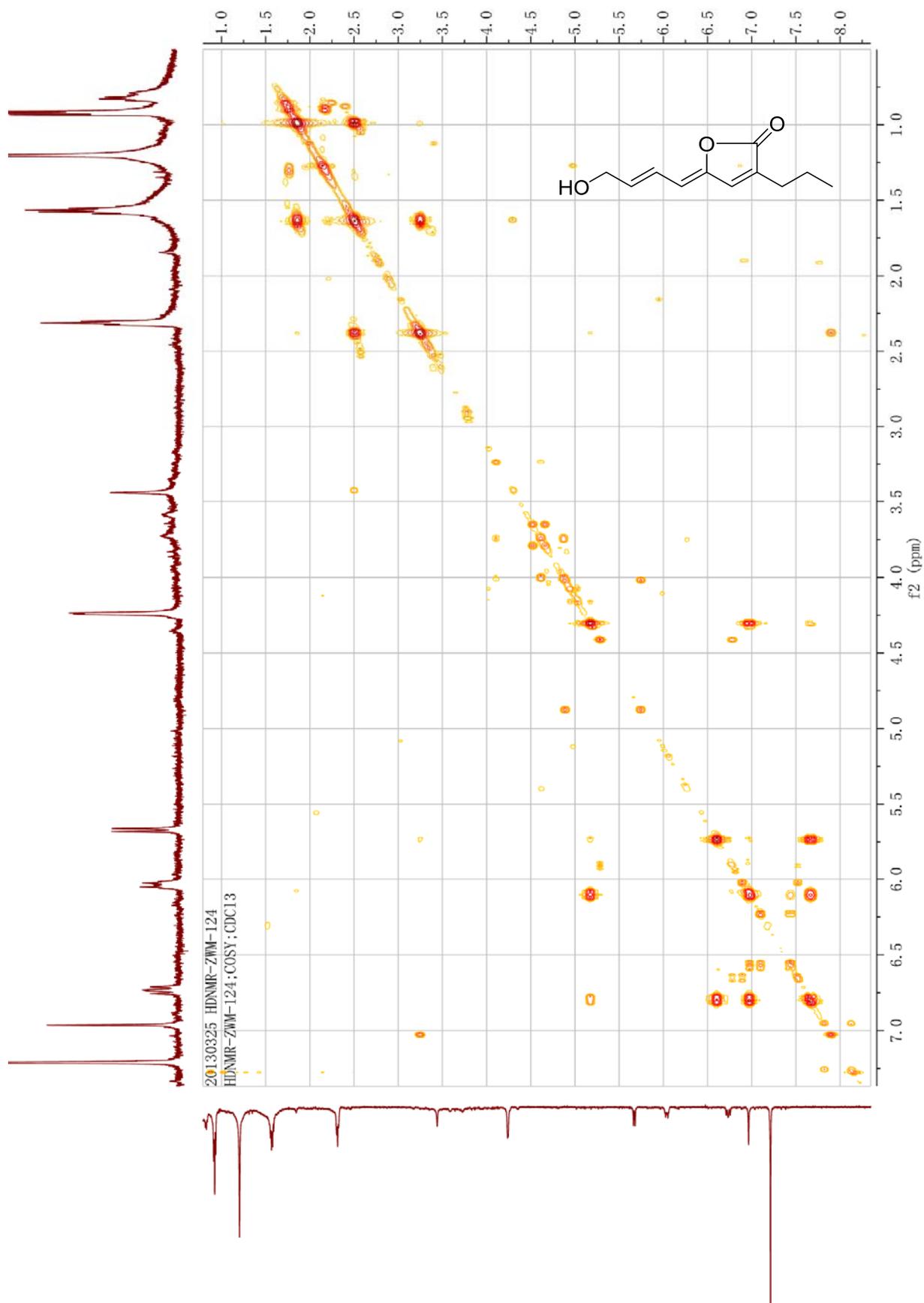


Figure S6. The NOE difference spectrum of compound **1** in CDCl₃

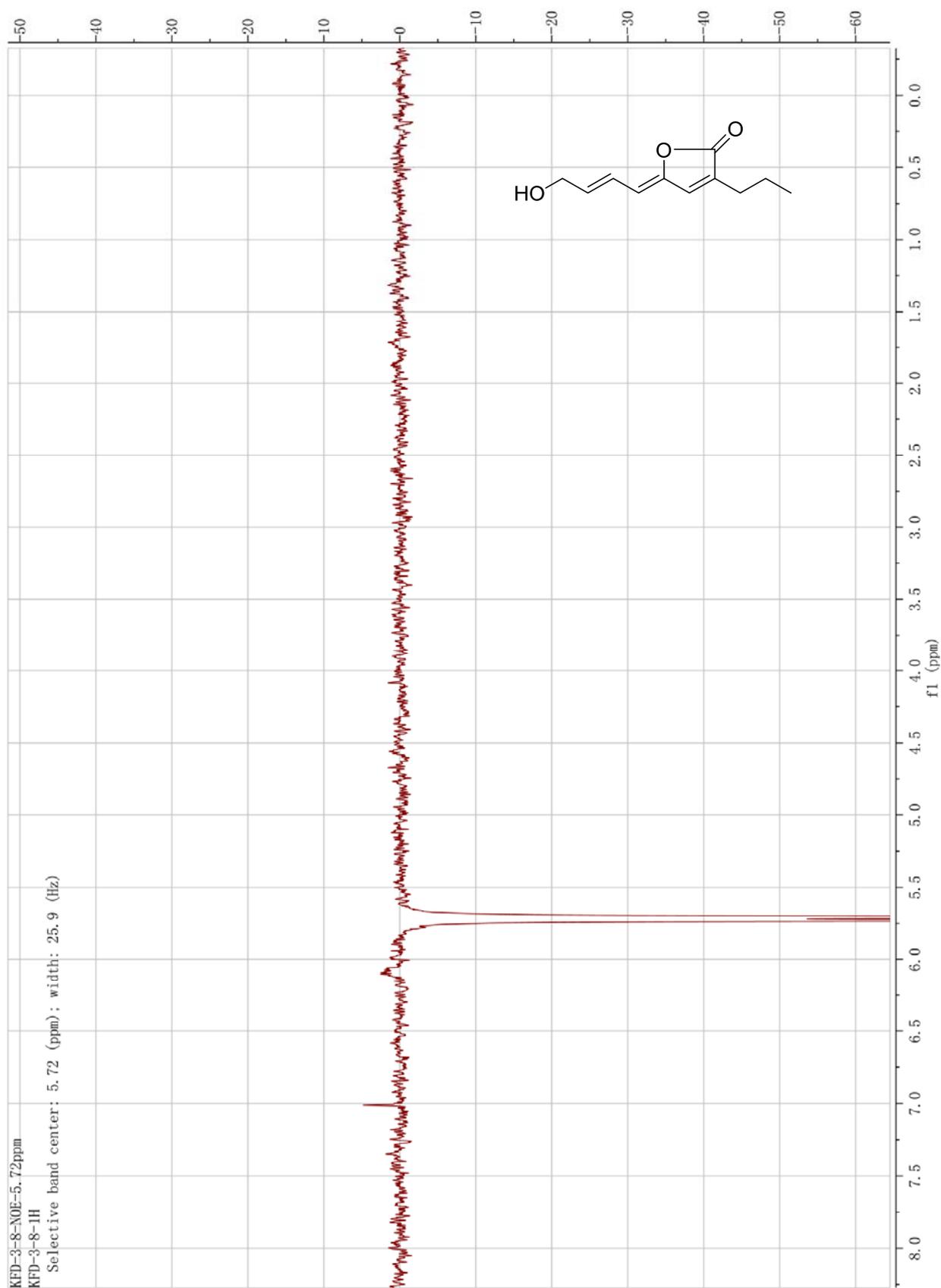


Figure S7. The ^1H -NMR spectrum of compound **2** in CDCl_3

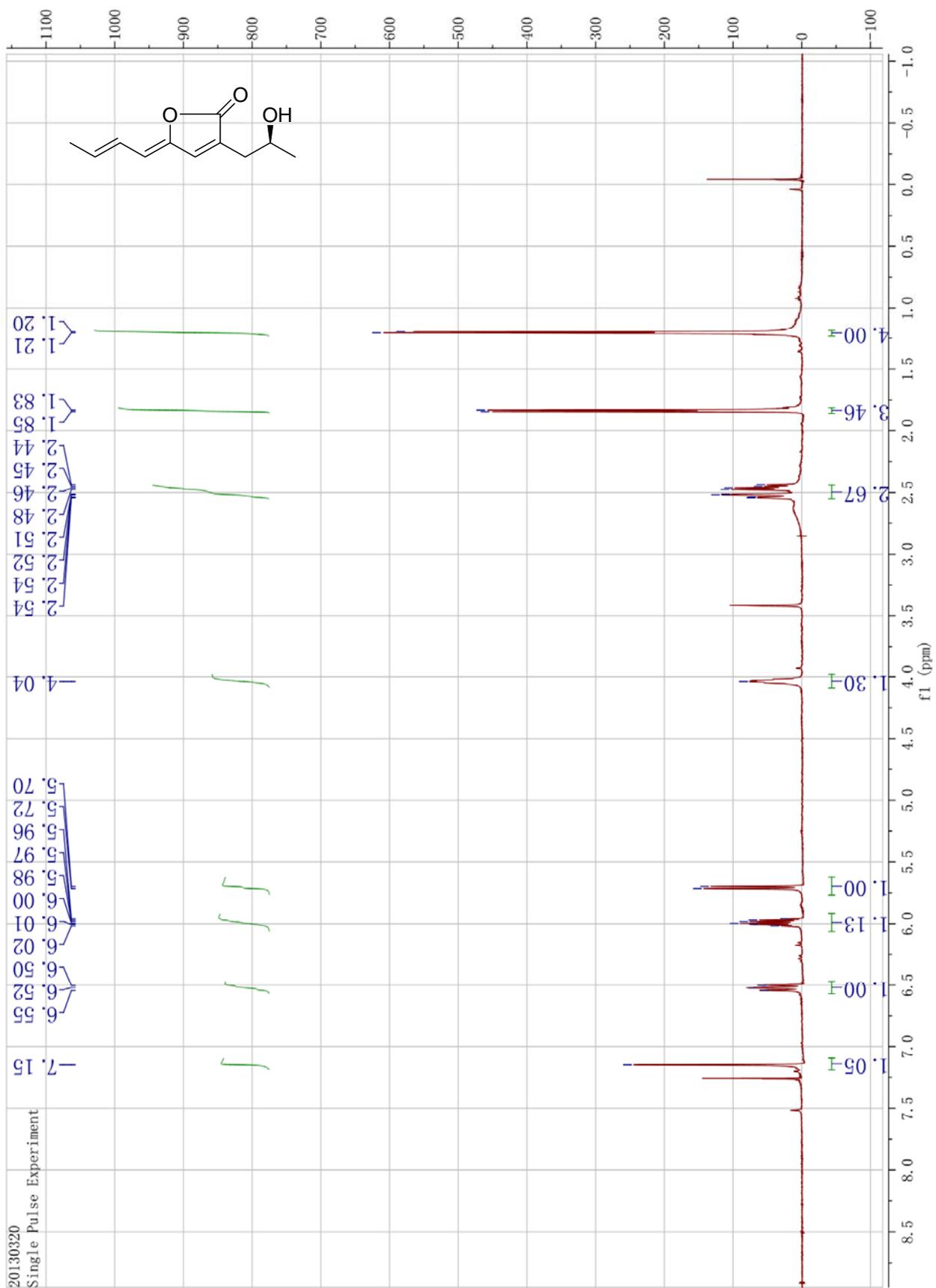


Figure S8. The ^{13}C -NMR spectrum of compound **2** in CDCl_3

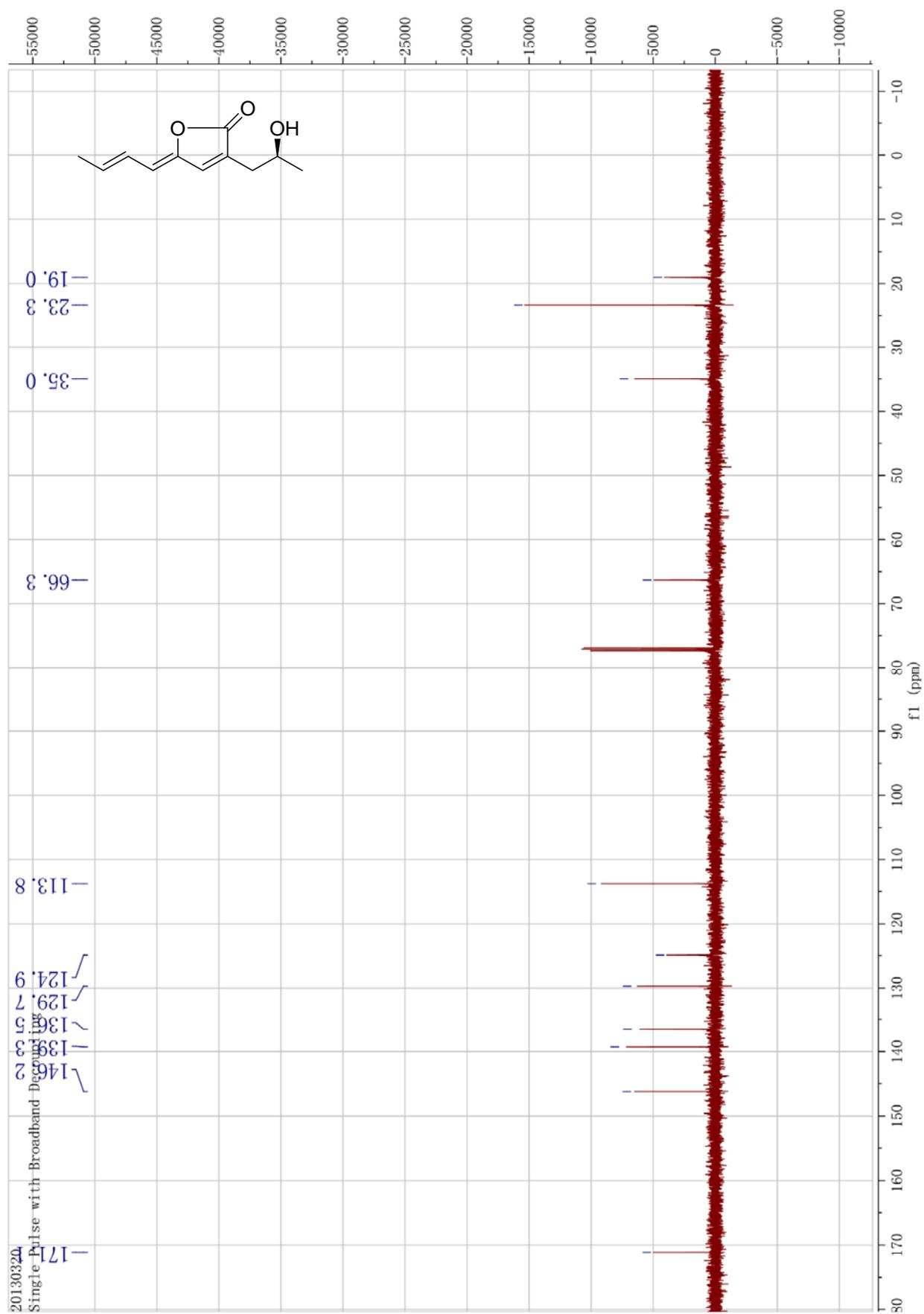


Figure S9. The DEPT spectrum of compound **2** in CDCl₃

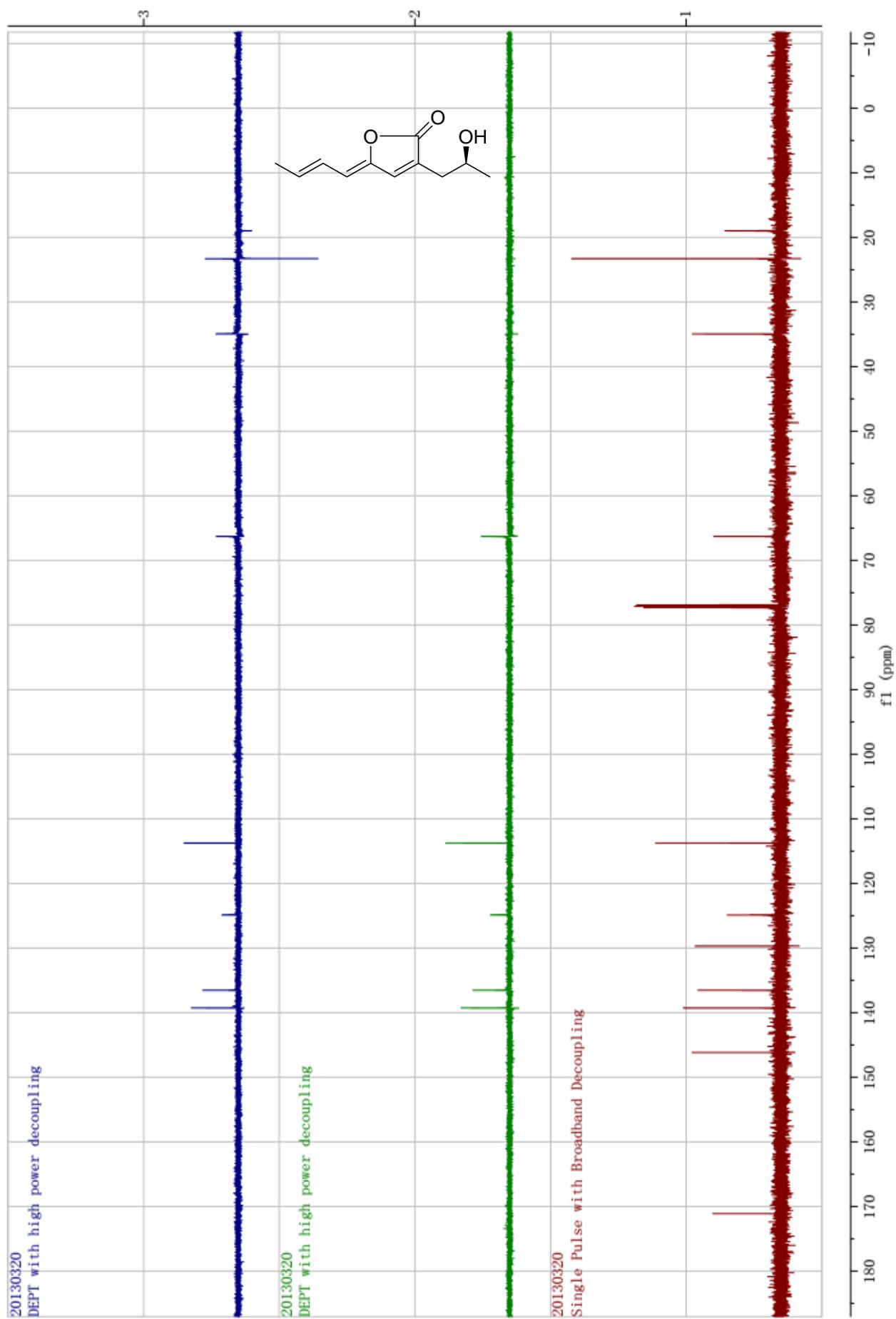


Figure S10. The HMQC spectrum of compound **2** in CDCl₃

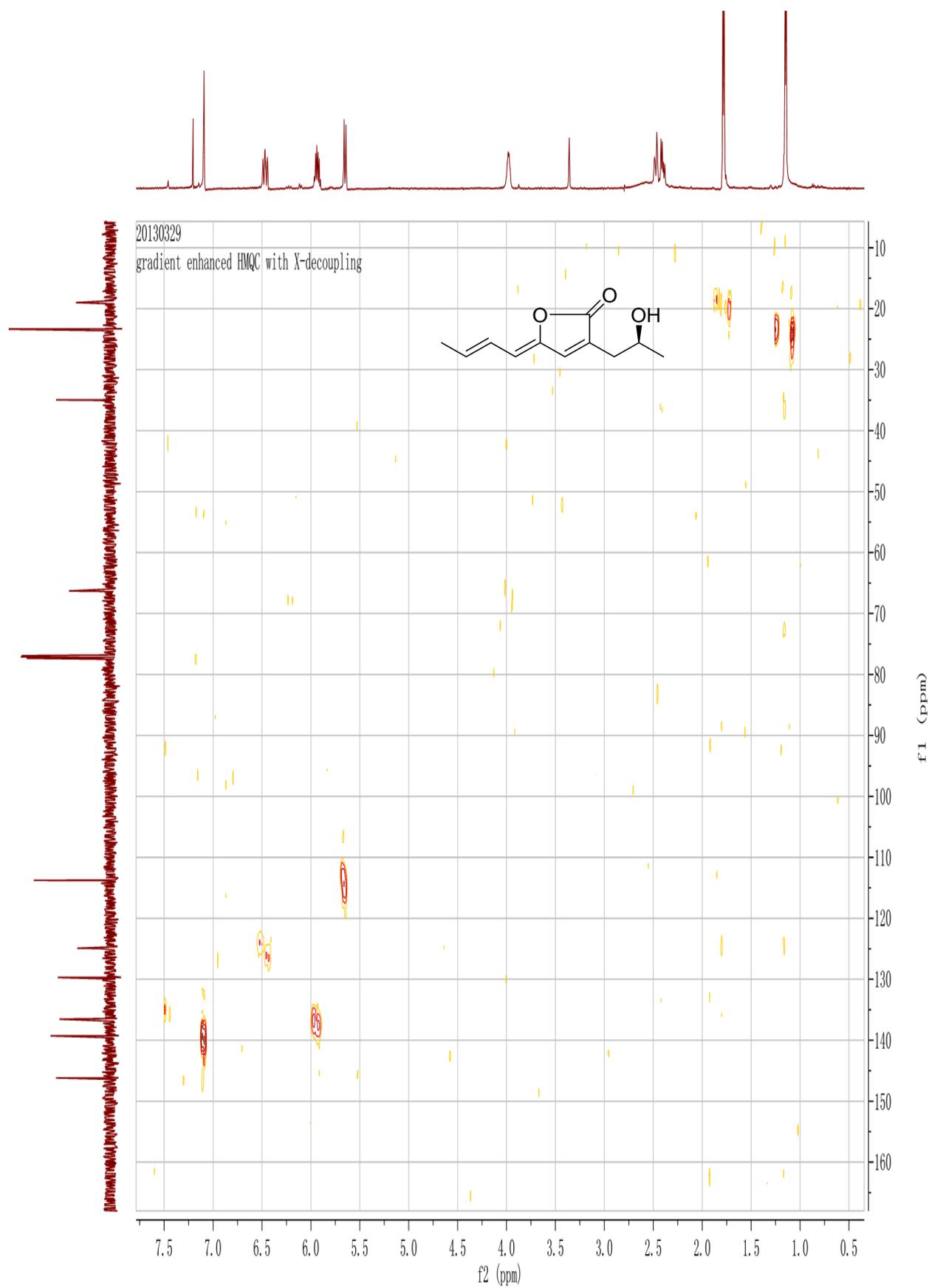


Figure S11. The ^1H - ^1H COSY spectrum of compound **2** in CDCl_3

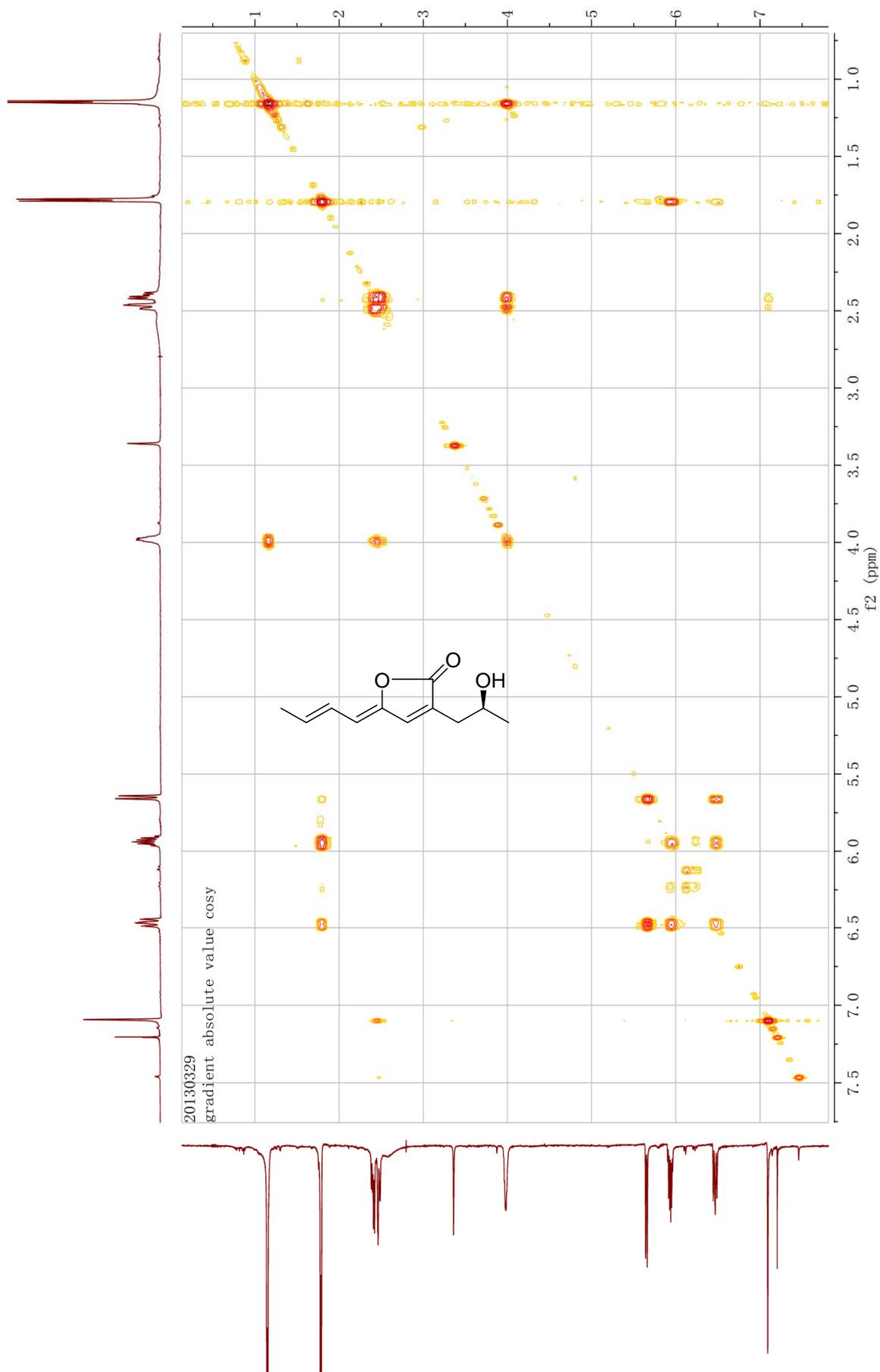


Figure S12. The HMBC spectrum of compound **2** in CDCl₃

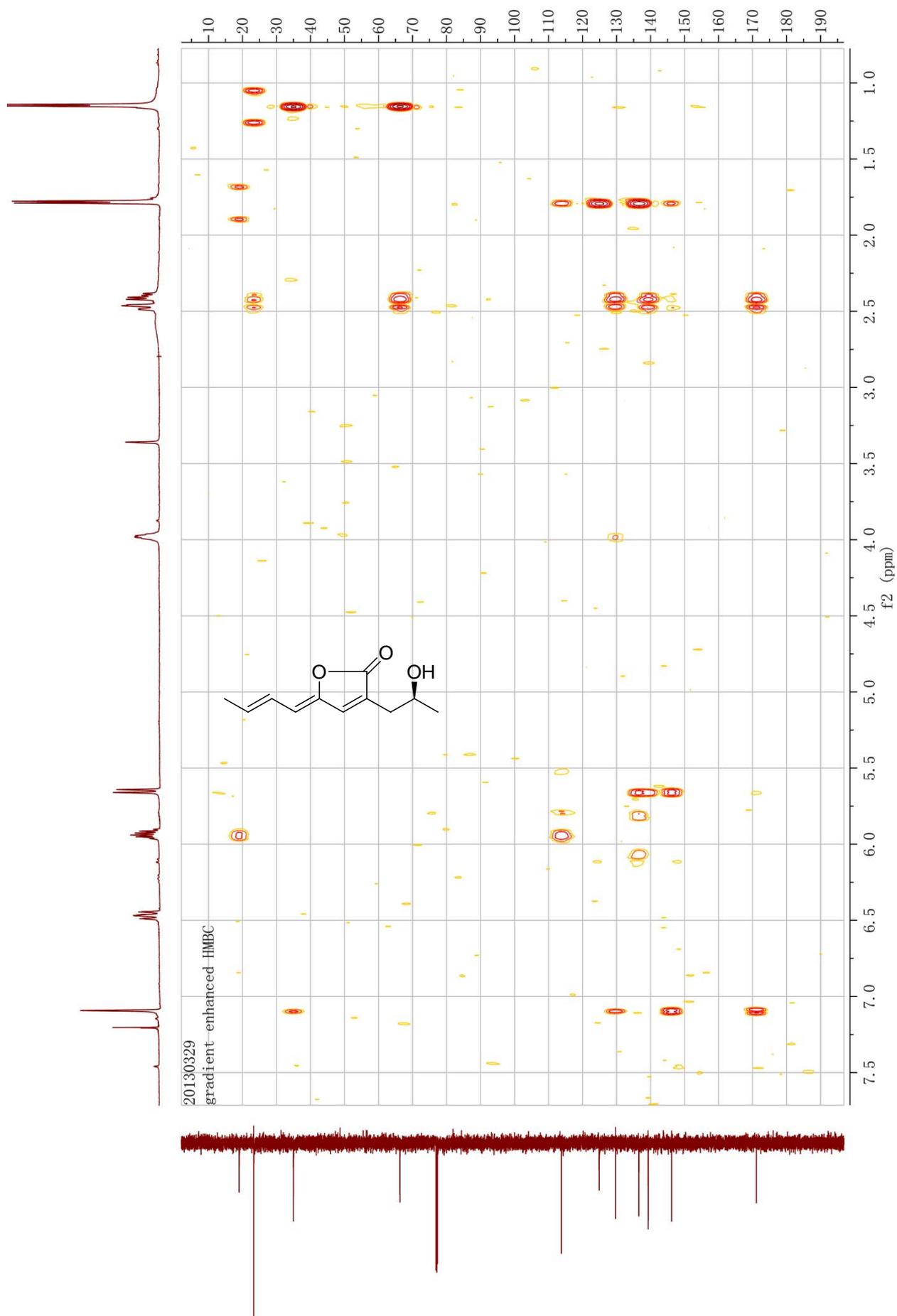


Figure S13. The NOE difference spectrum of compound **2** in CDCl₃

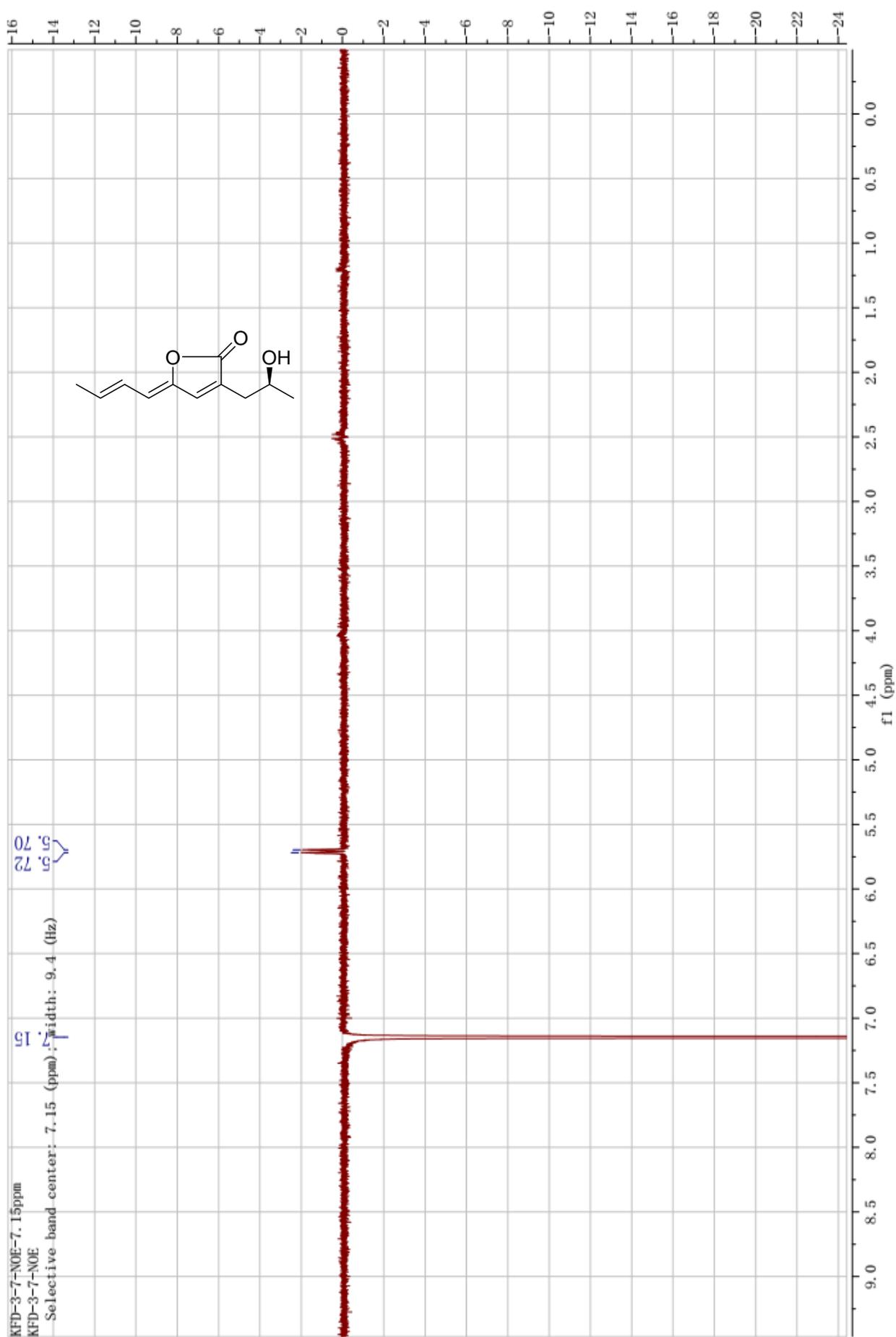


Figure S14. The ^1H -NMR spectrum of compounds **3** and **4** in CDCl_3

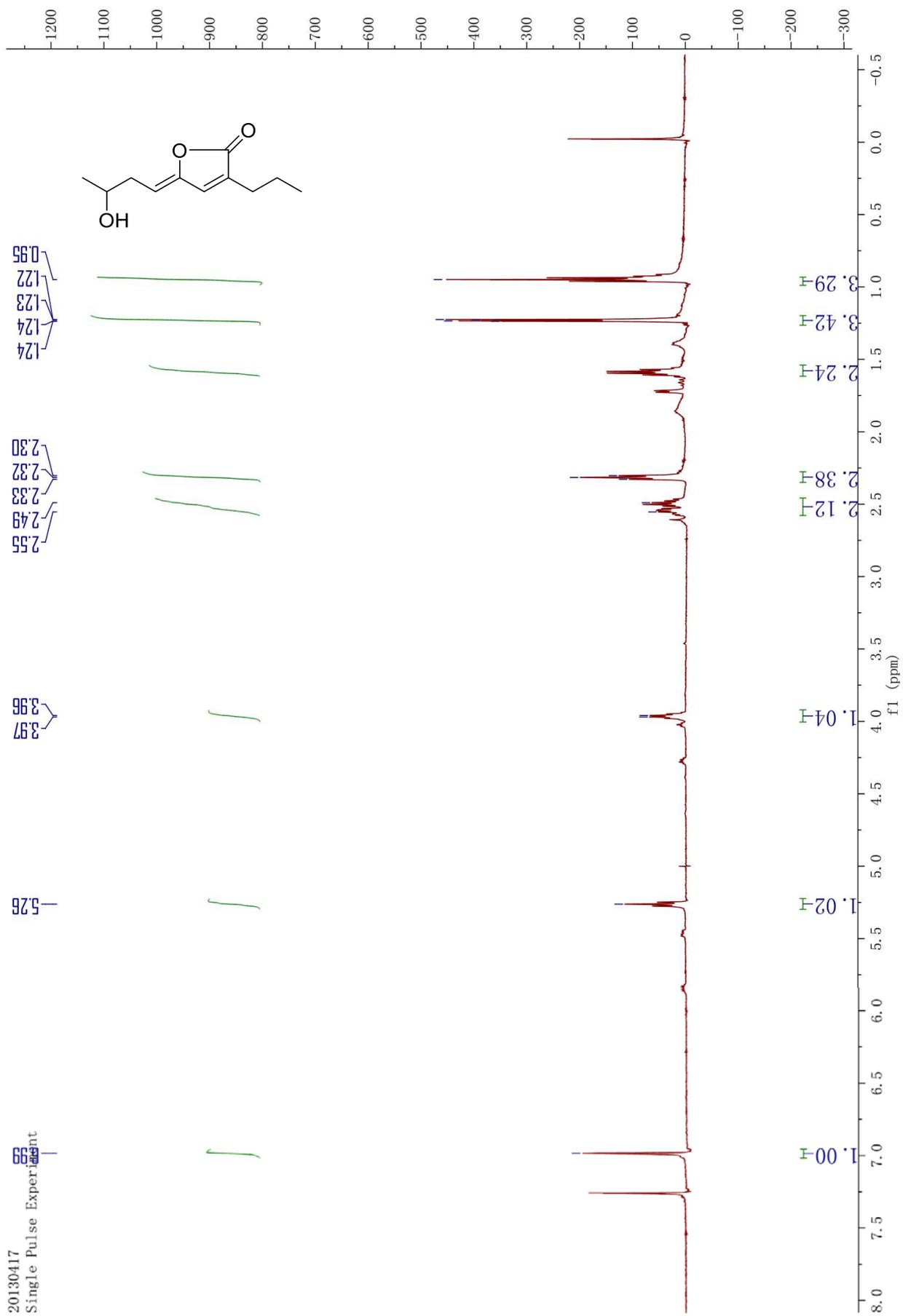
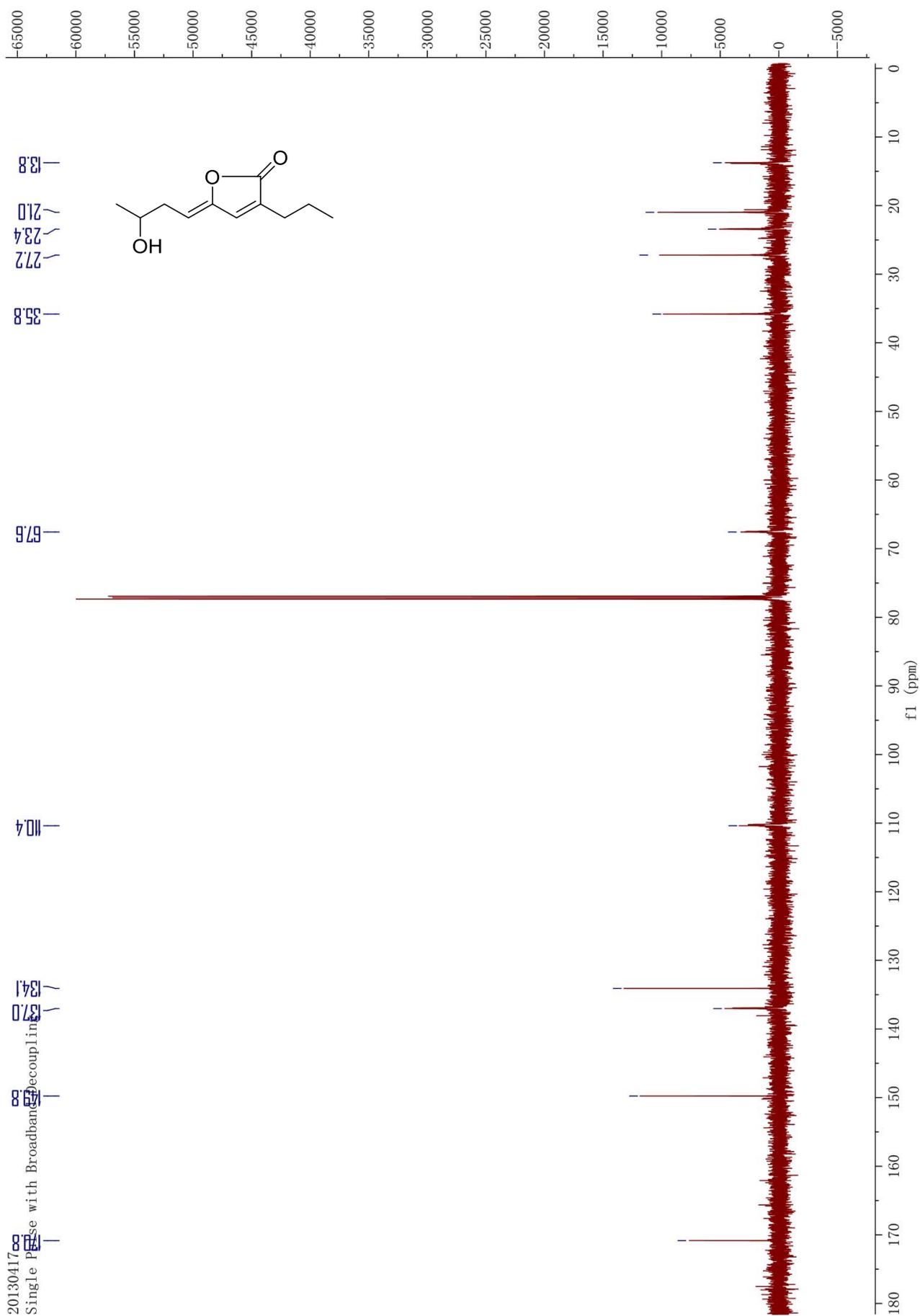


Figure S15. The ^{13}C -NMR spectrum of compounds **3** and **4** in CDCl_3



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Single Pulse with Broadband
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Figure S16. The DEPT spectrum of compounds **3** and **4** in CDCl₃

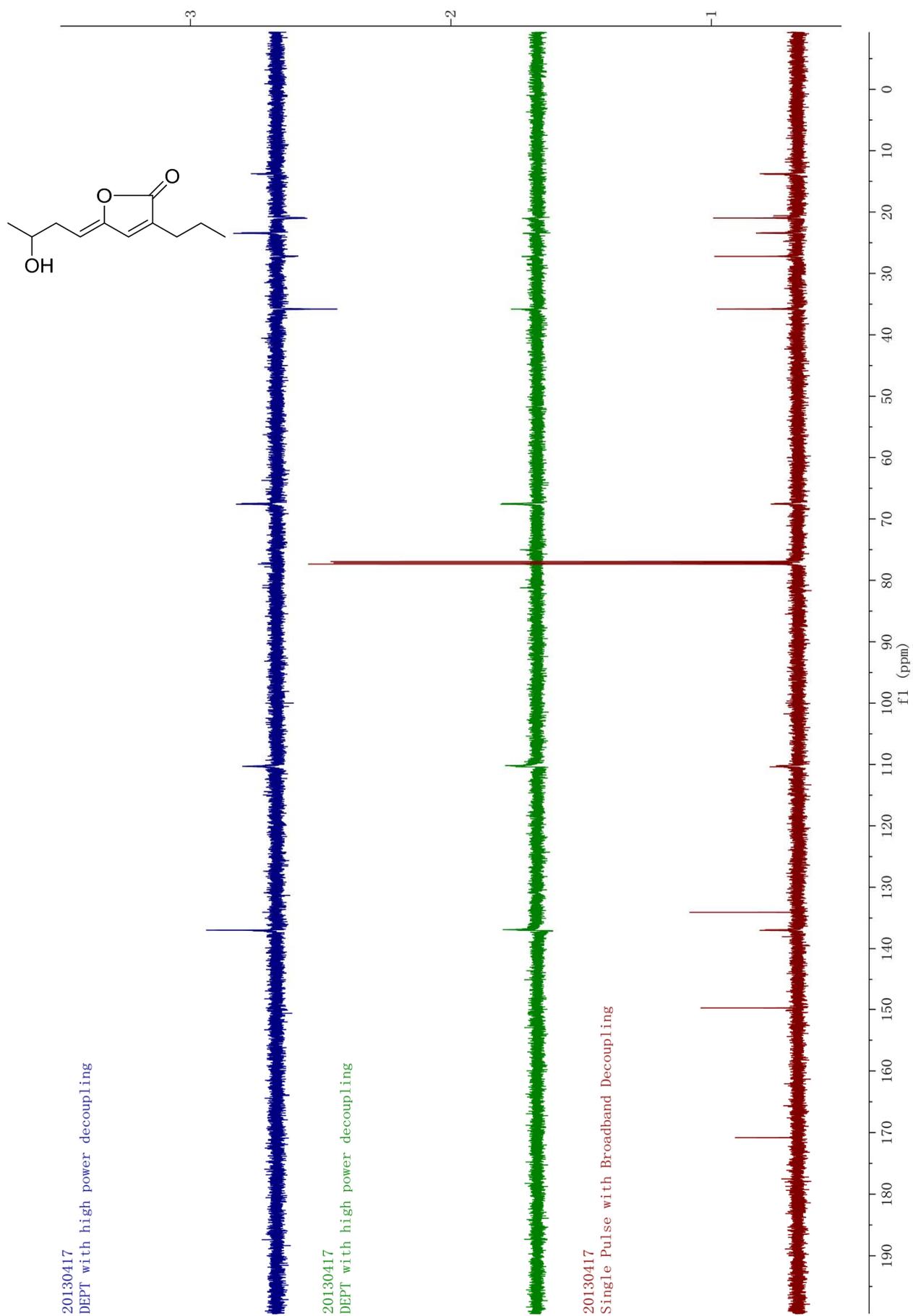


Figure S17. The HMQC spectrum of compounds 3 and 4 in CDCl₃

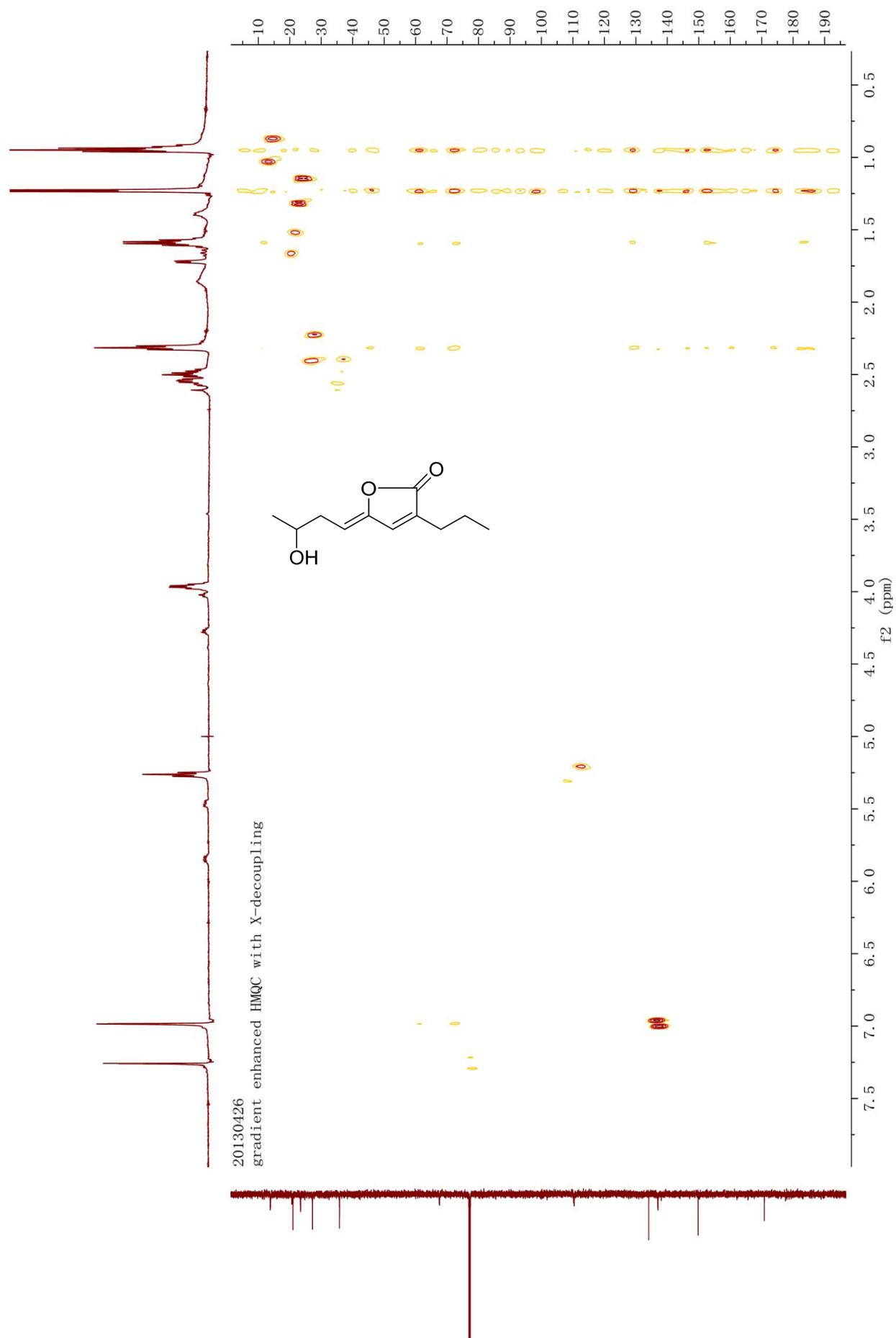


Figure S18. The ^1H - ^1H COSY spectrum of compounds **3** and **4** in CDCl_3

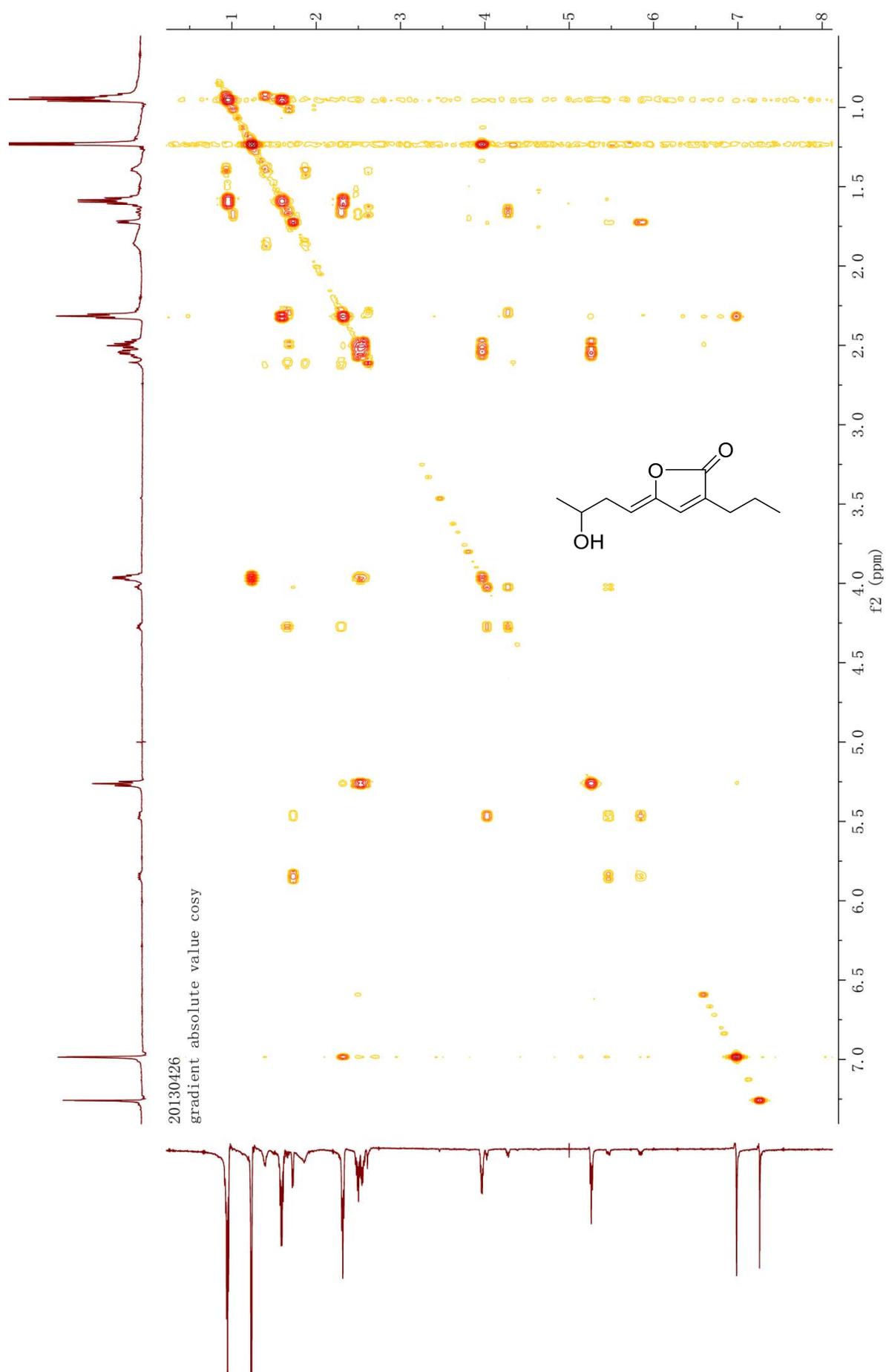


Figure S19. The HMBC spectrum of compounds **3** and **4** in CDCl₃

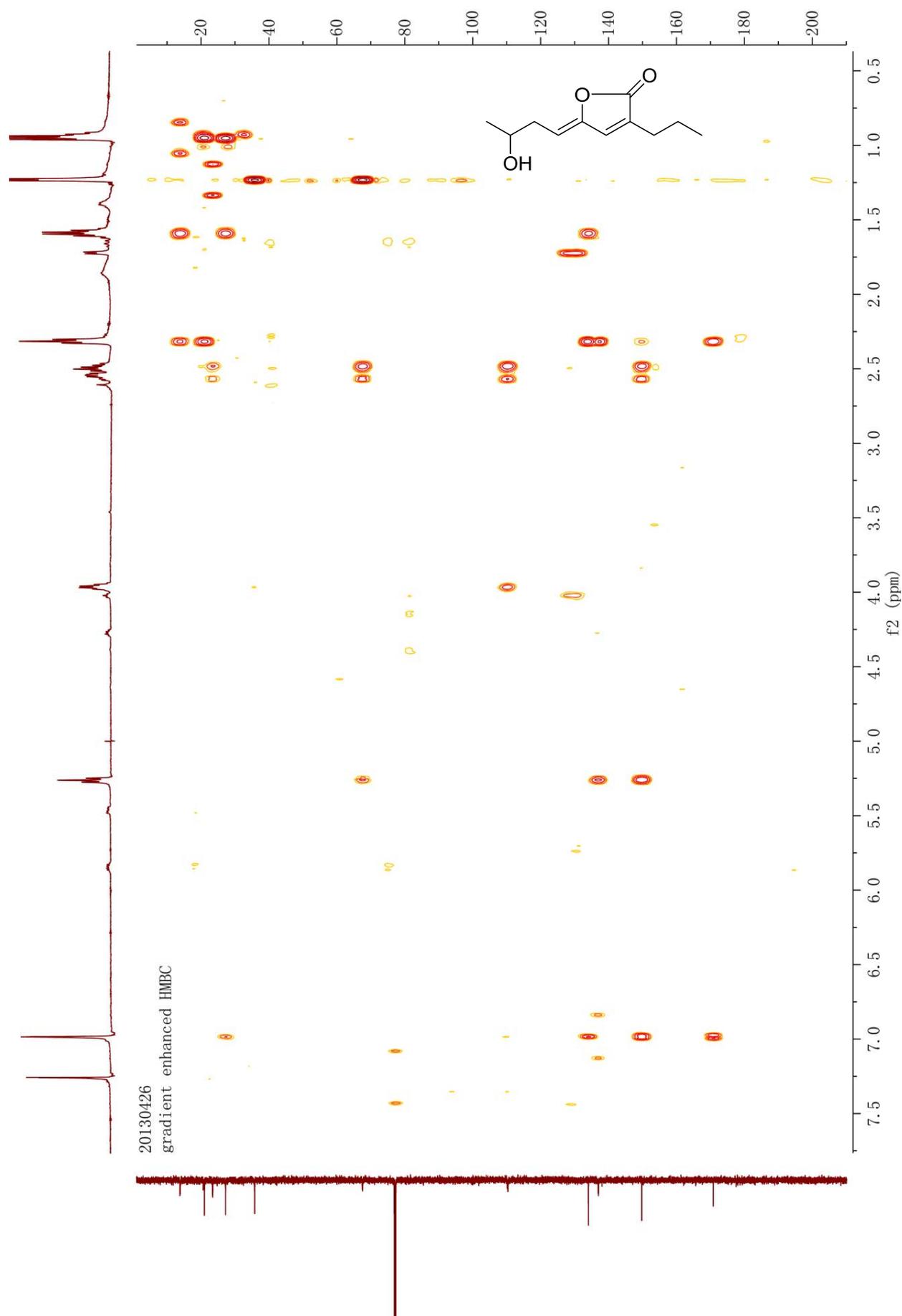


Figure S20. The NOE difference spectrum of compounds **3** and **4** in CDCl₃

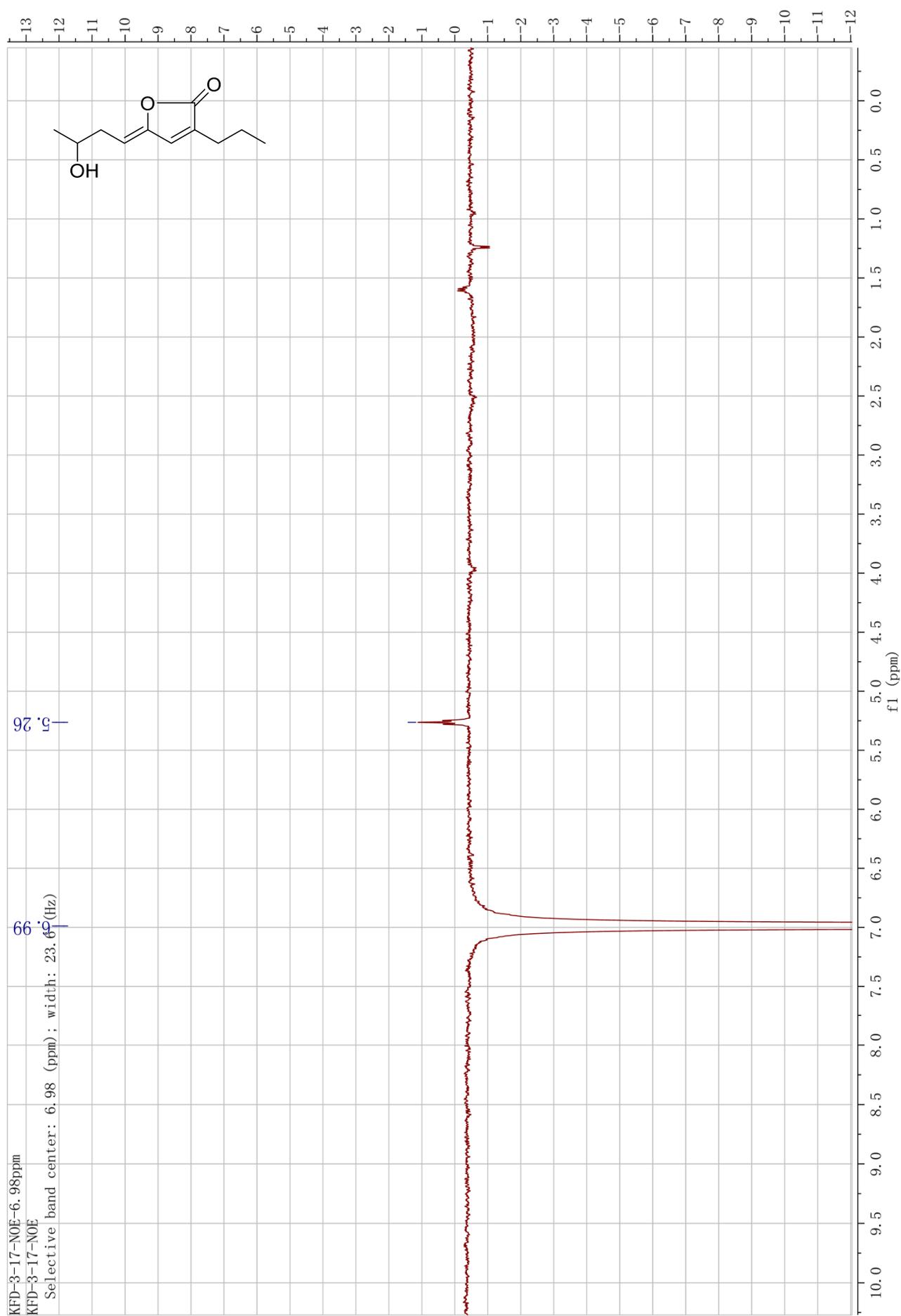


Figure S21. The ^1H -NMR spectrum of compound **5** in CDCl_3

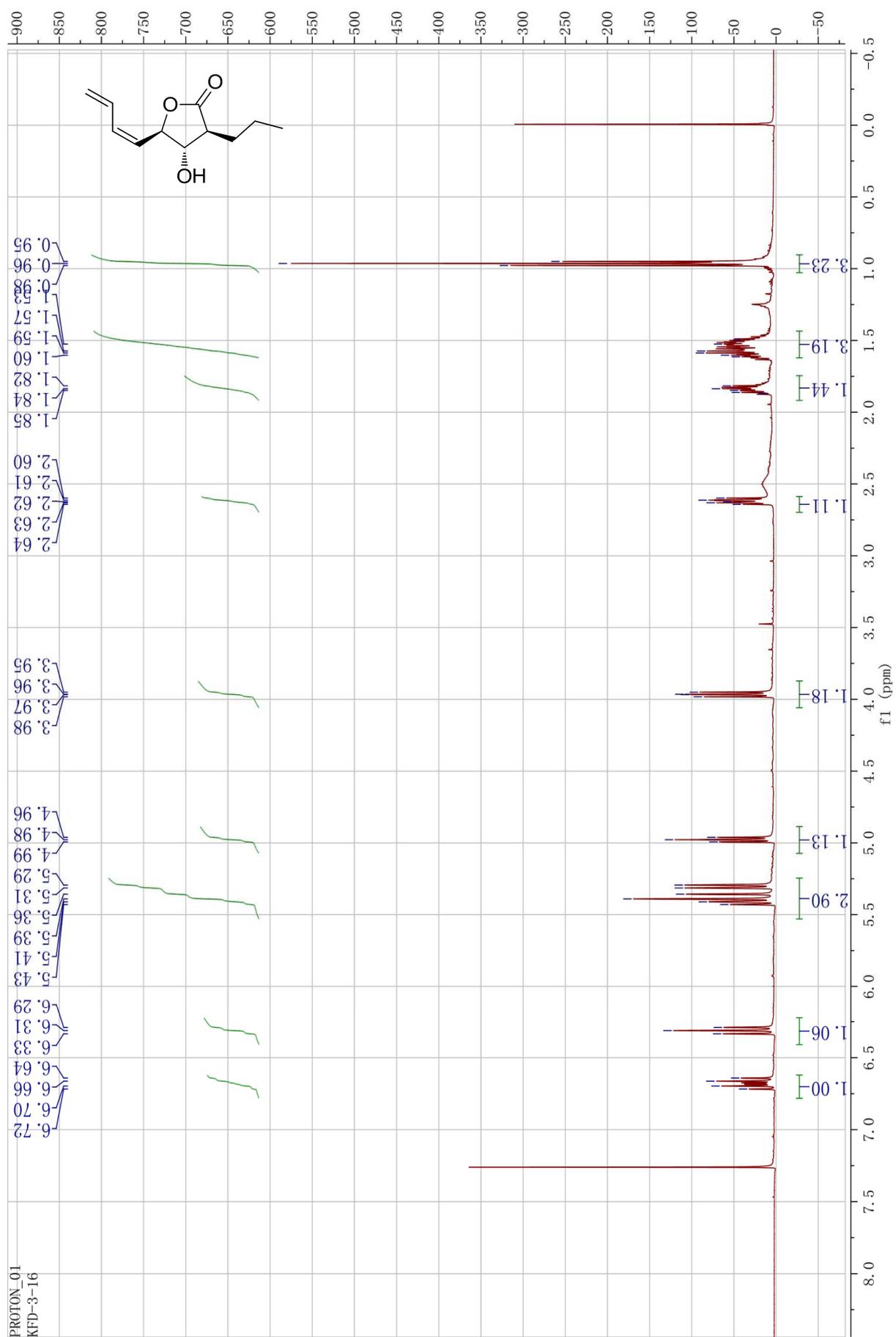


Figure S22. The ^{13}C -NMR spectrum of compound **5** in CDCl_3

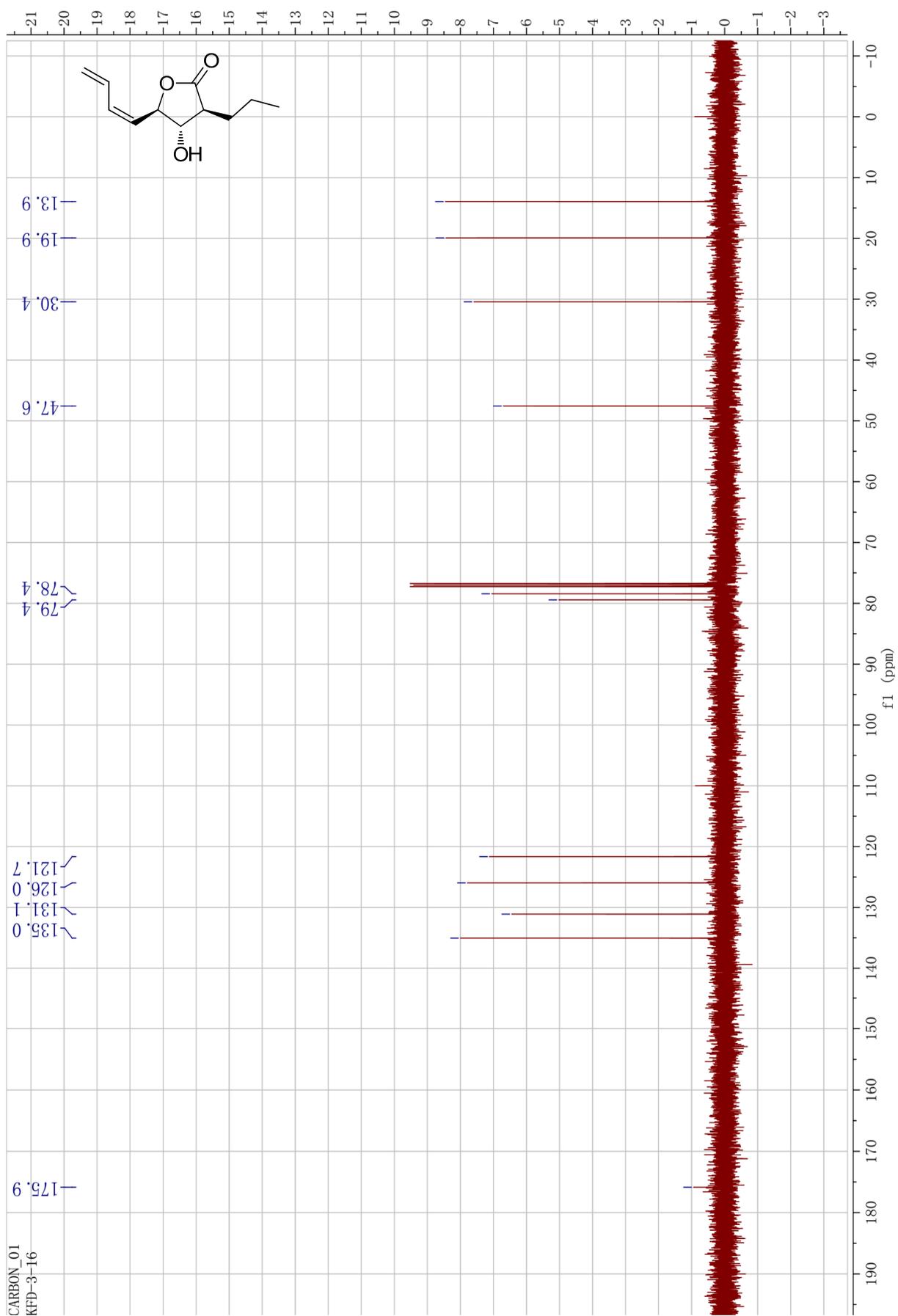


Figure S23. The DEPT spectrum of compound 5 in CDCl₃

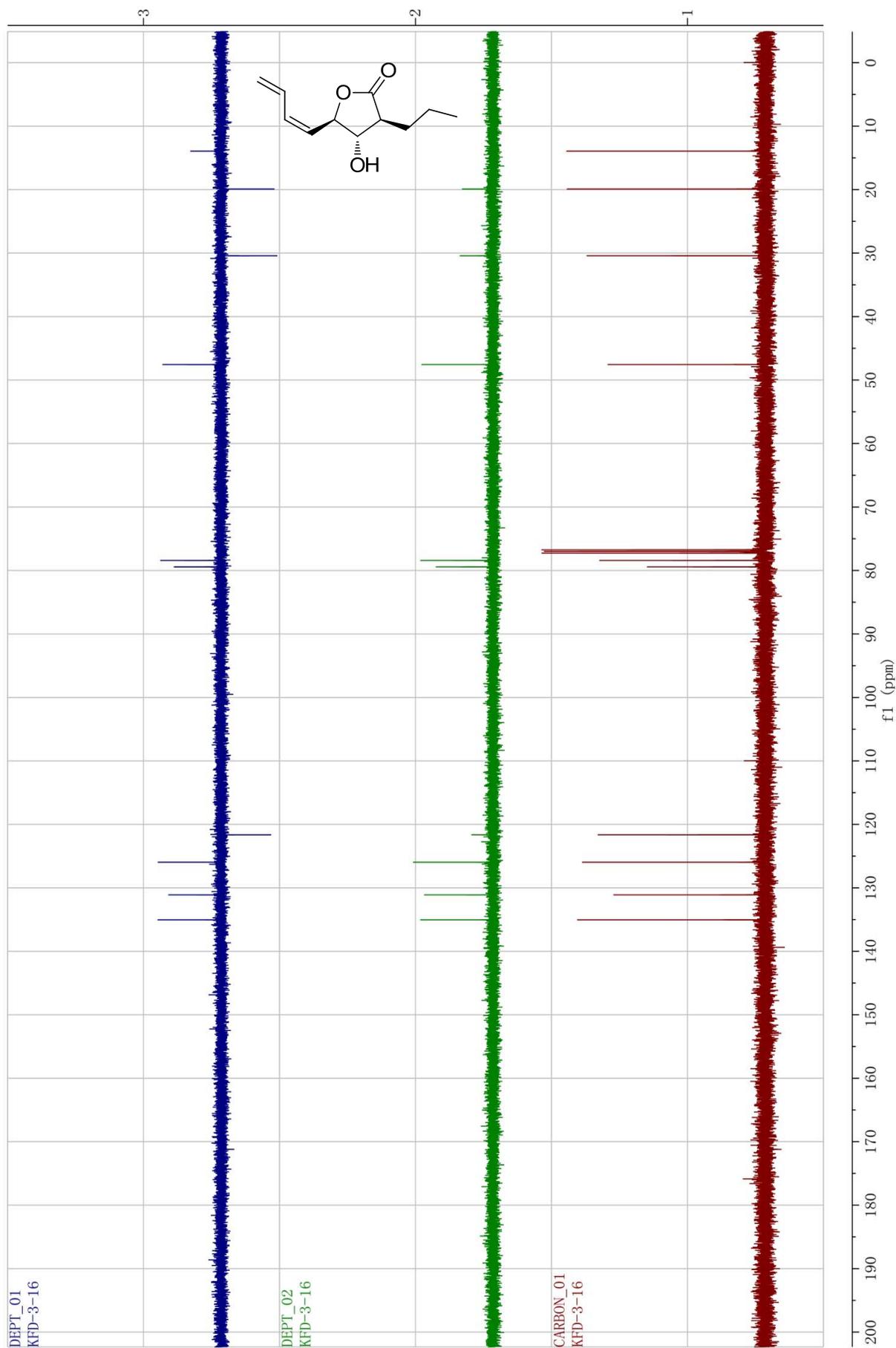


Figure S24. The HMQC spectrum of compound **5** in CDCl₃

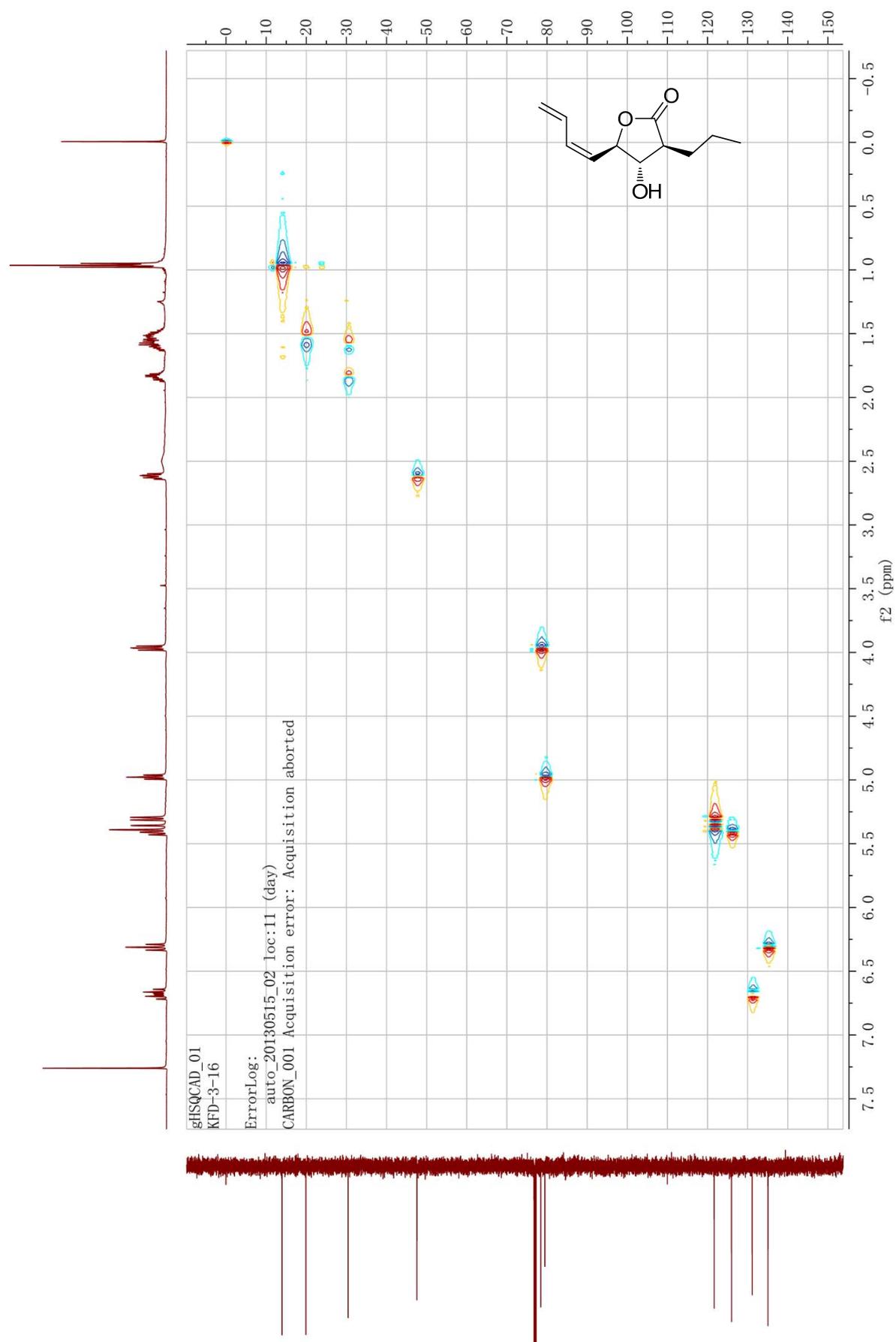


Figure S25. The ^1H - ^1H COSY spectrum of compound **5** in CDCl_3

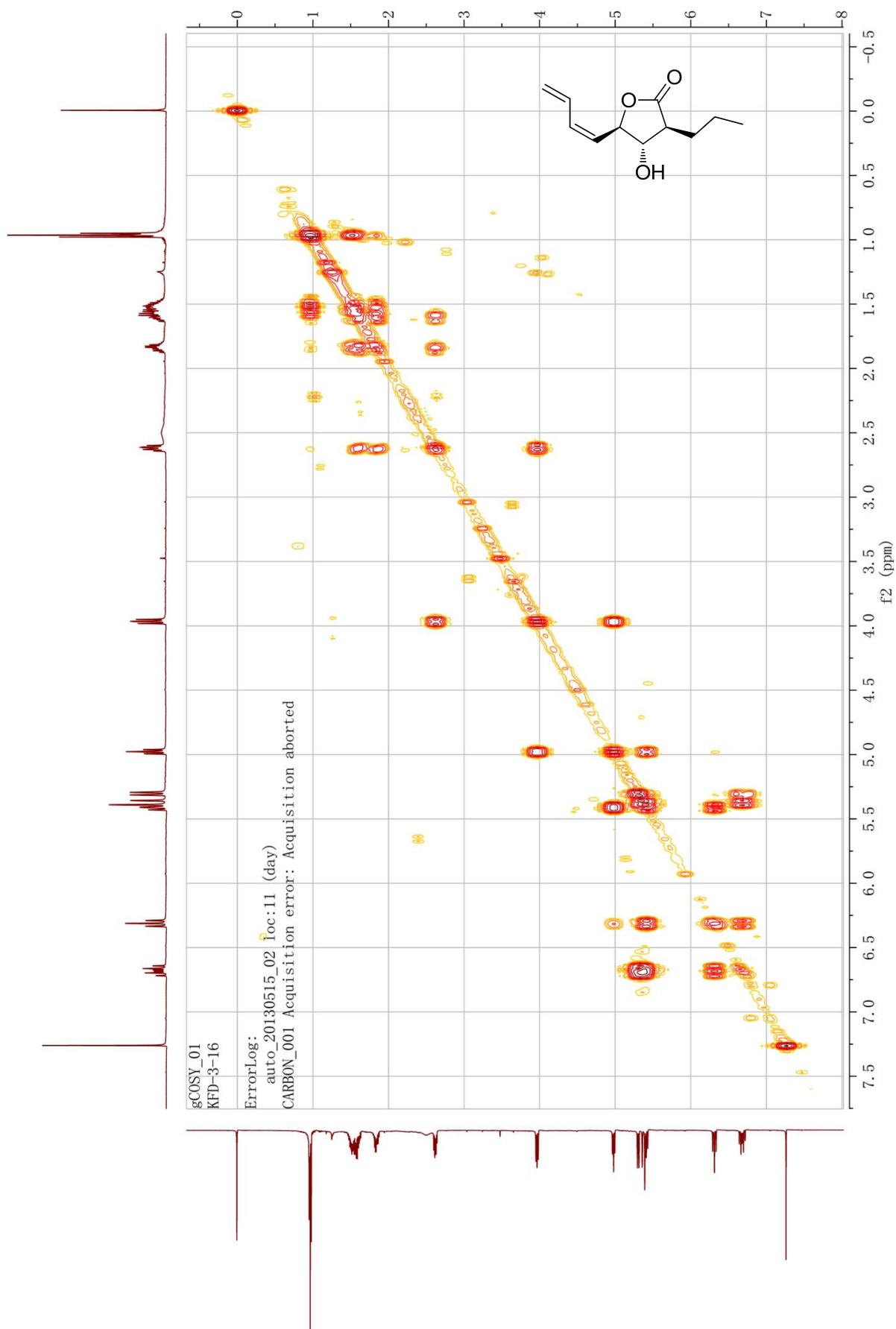


Figure S26. The HMBC spectrum of compound **5** in CDCl₃

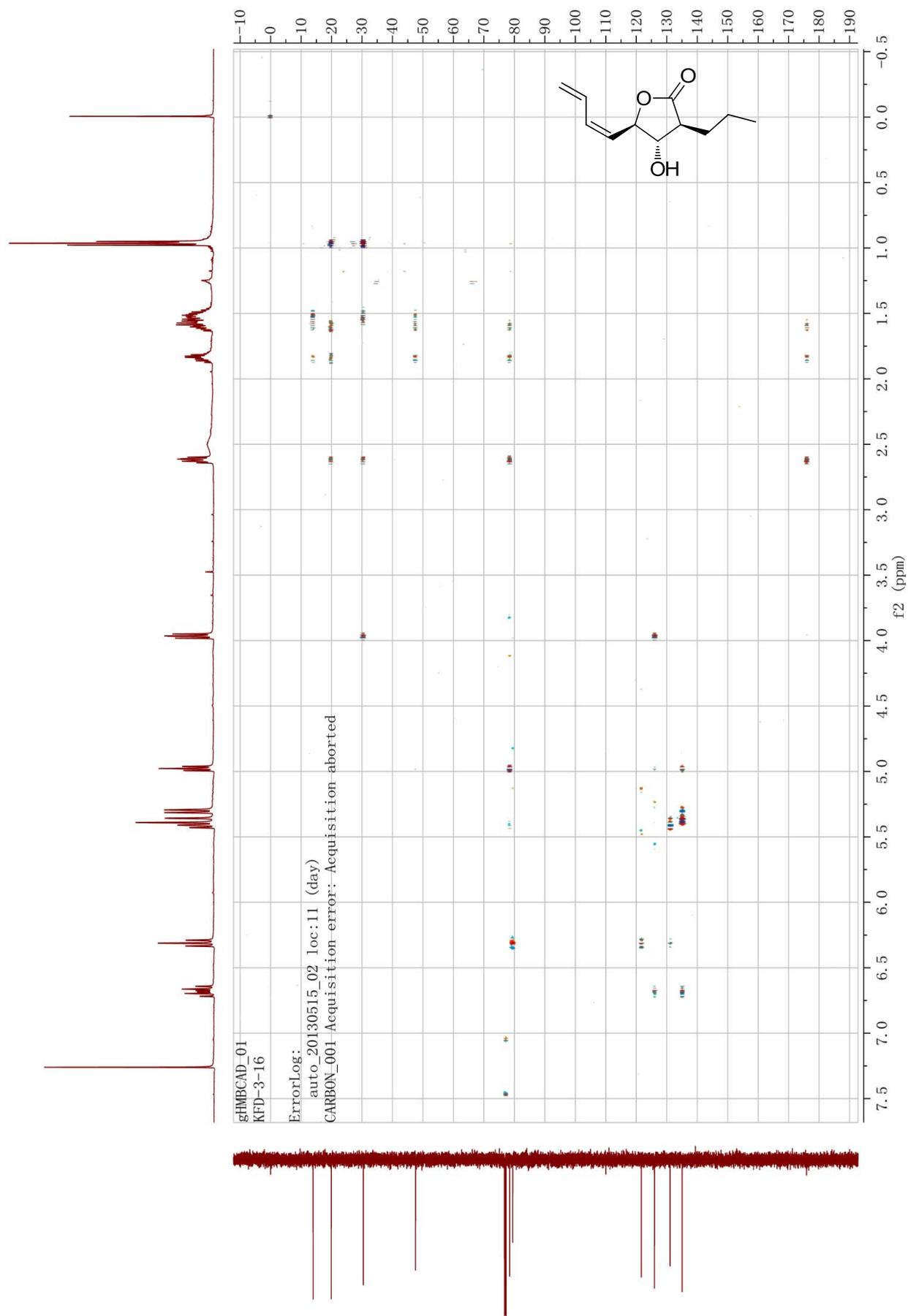


Figure S27. The NOE difference spectrum of compound **5** in CDCl₃ (I)

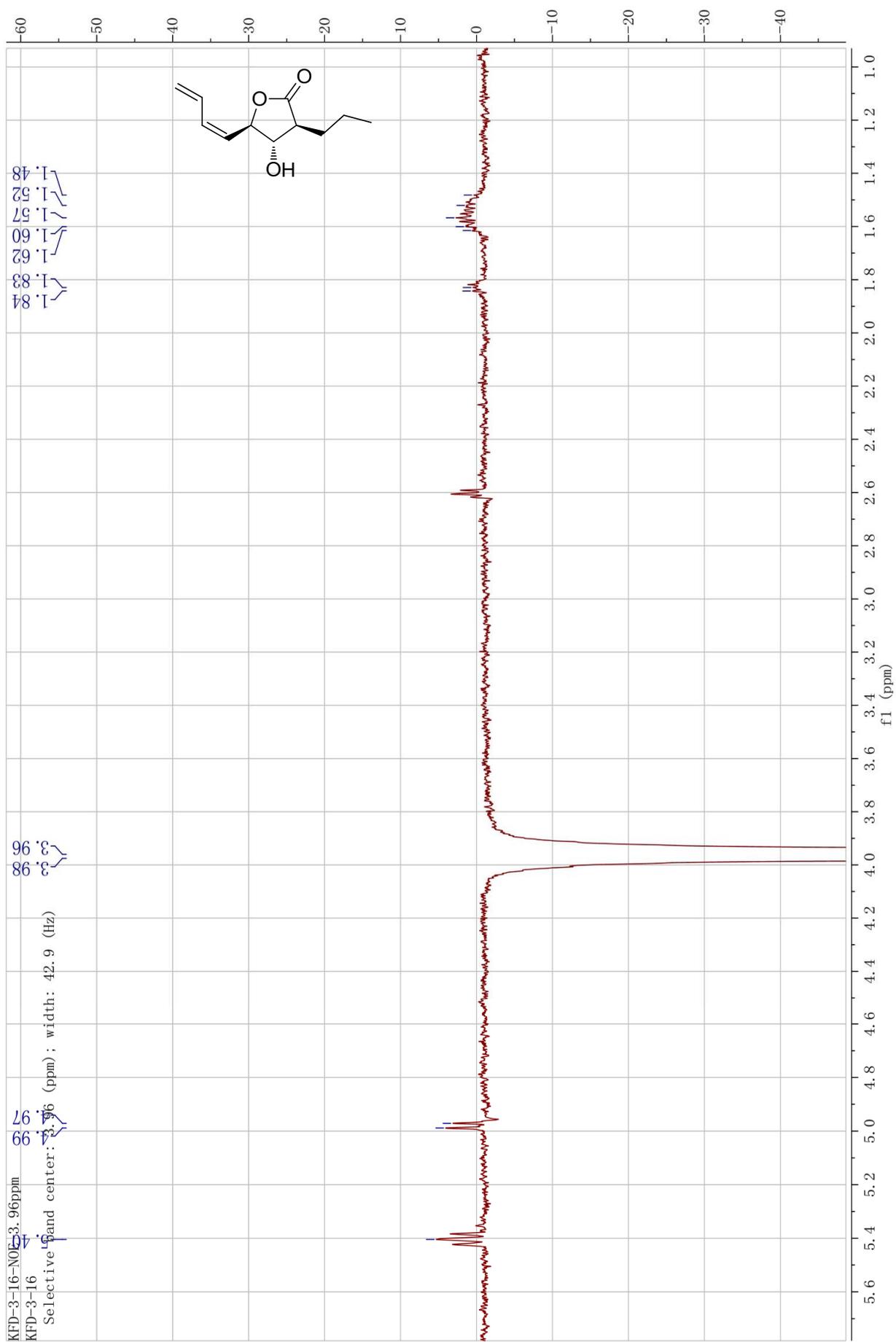


Figure S28. The NOE difference spectrum of compound **5** in CDCl₃ (II)

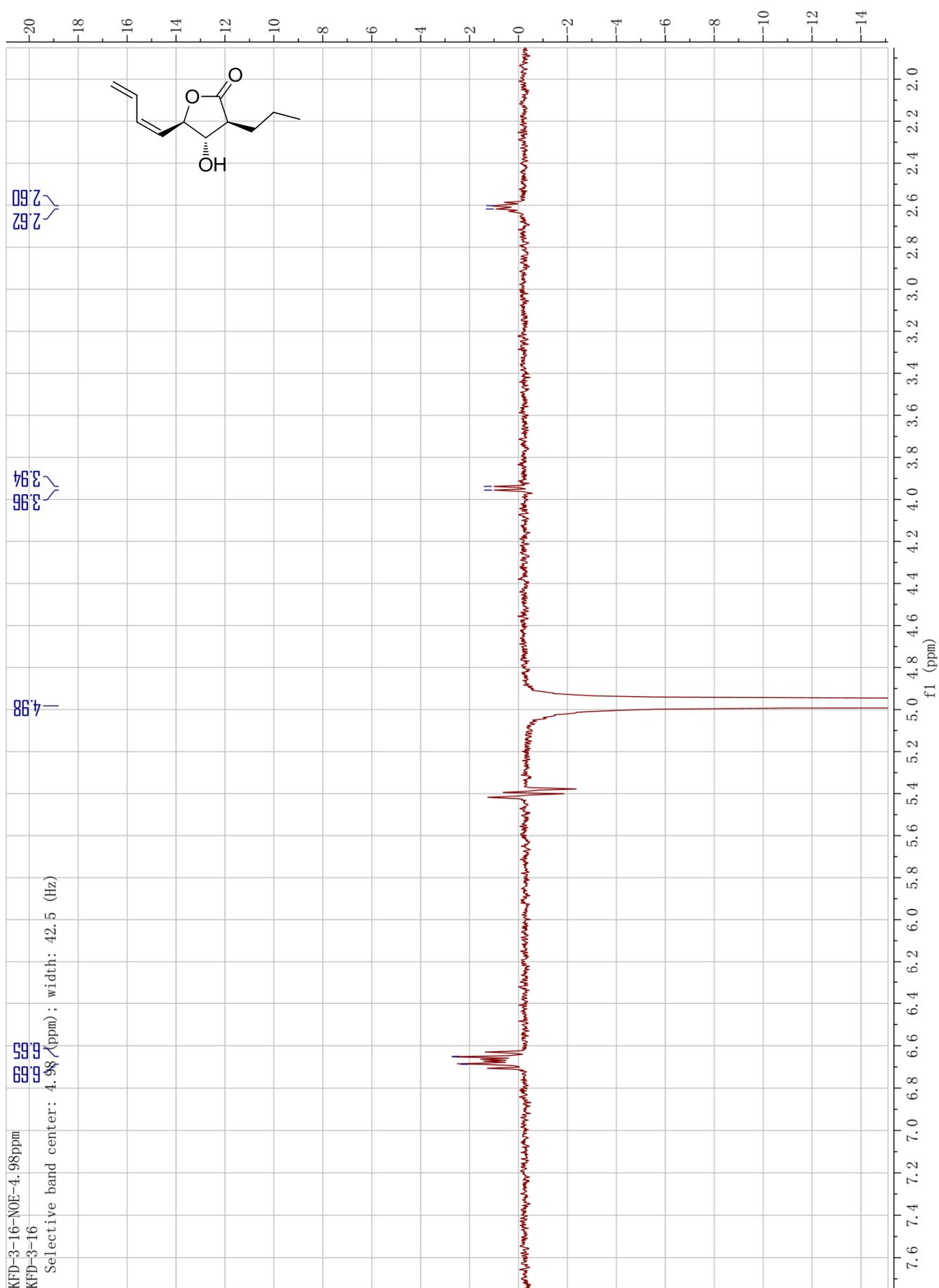


Figure S29. The ^1H -NMR spectrum of compound **6** in $\text{DMSO-}d_6$

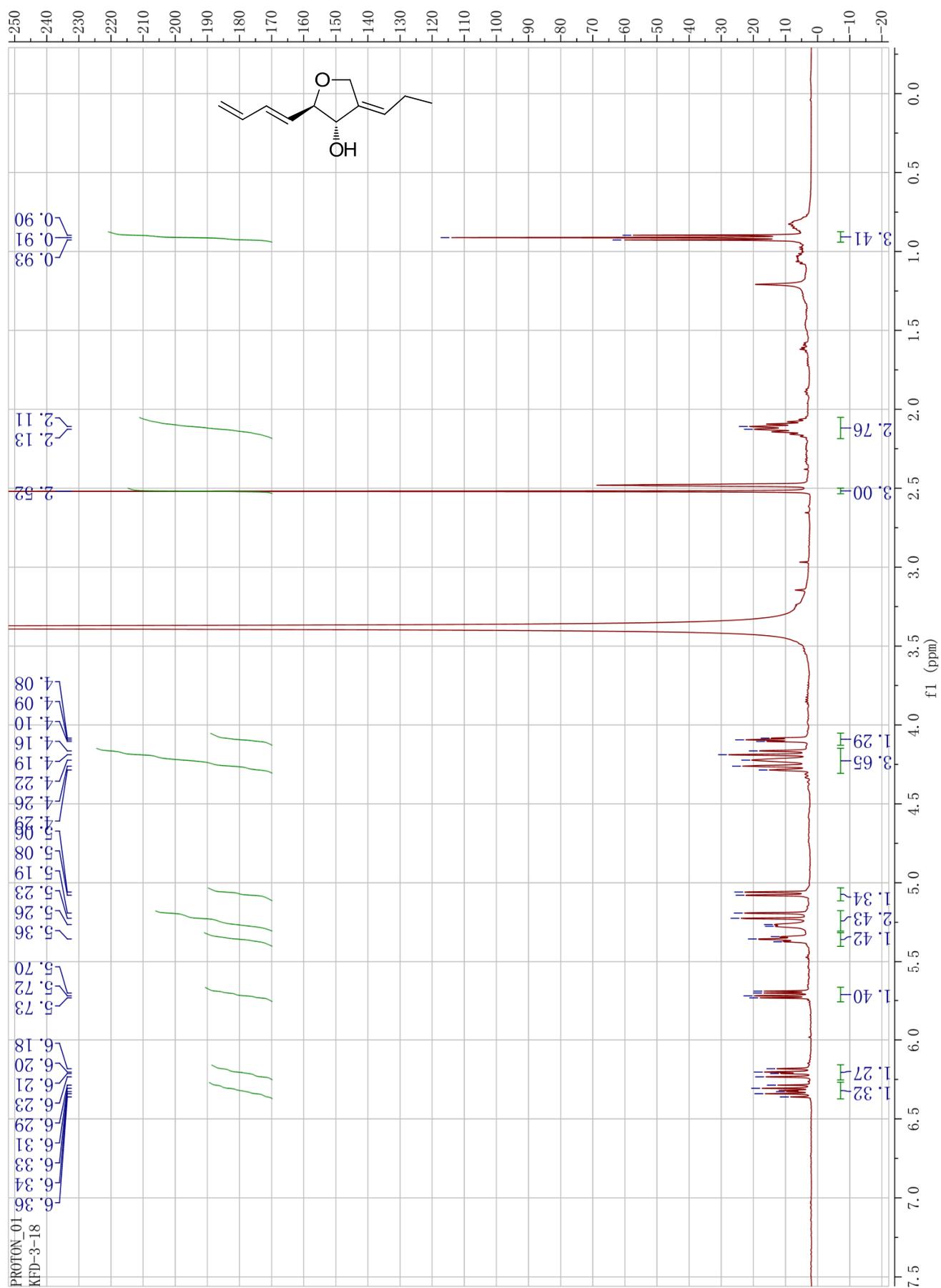


Figure S30. The ^{13}C -NMR spectrum of compound **6** in $\text{DMSO-}d_6$

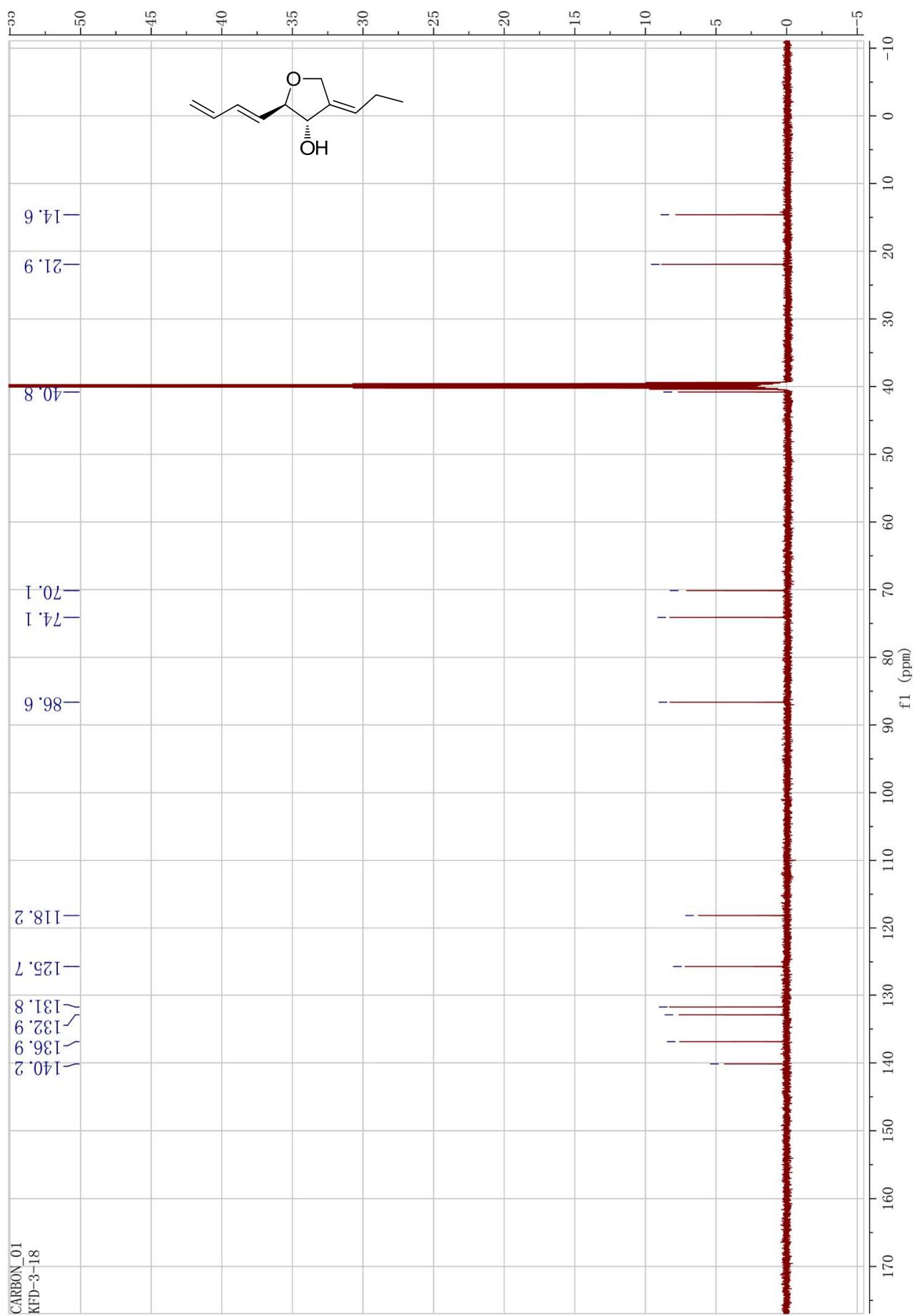


Figure S31. The DEPT spectrum of compound 6 in DMSO-*d*₆

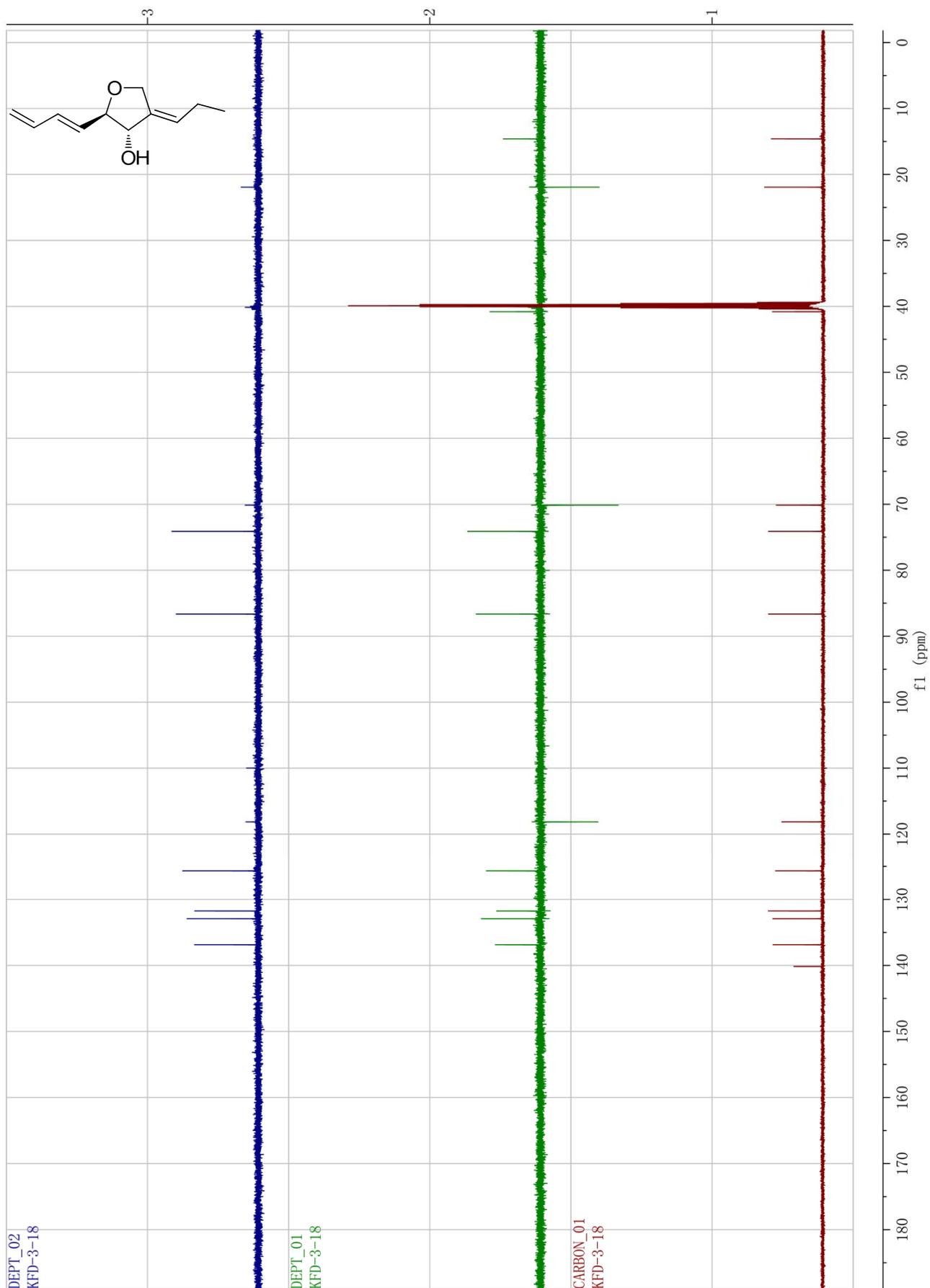


Figure S32. The HMQC spectrum of compound **6** in DMSO-*d*₆

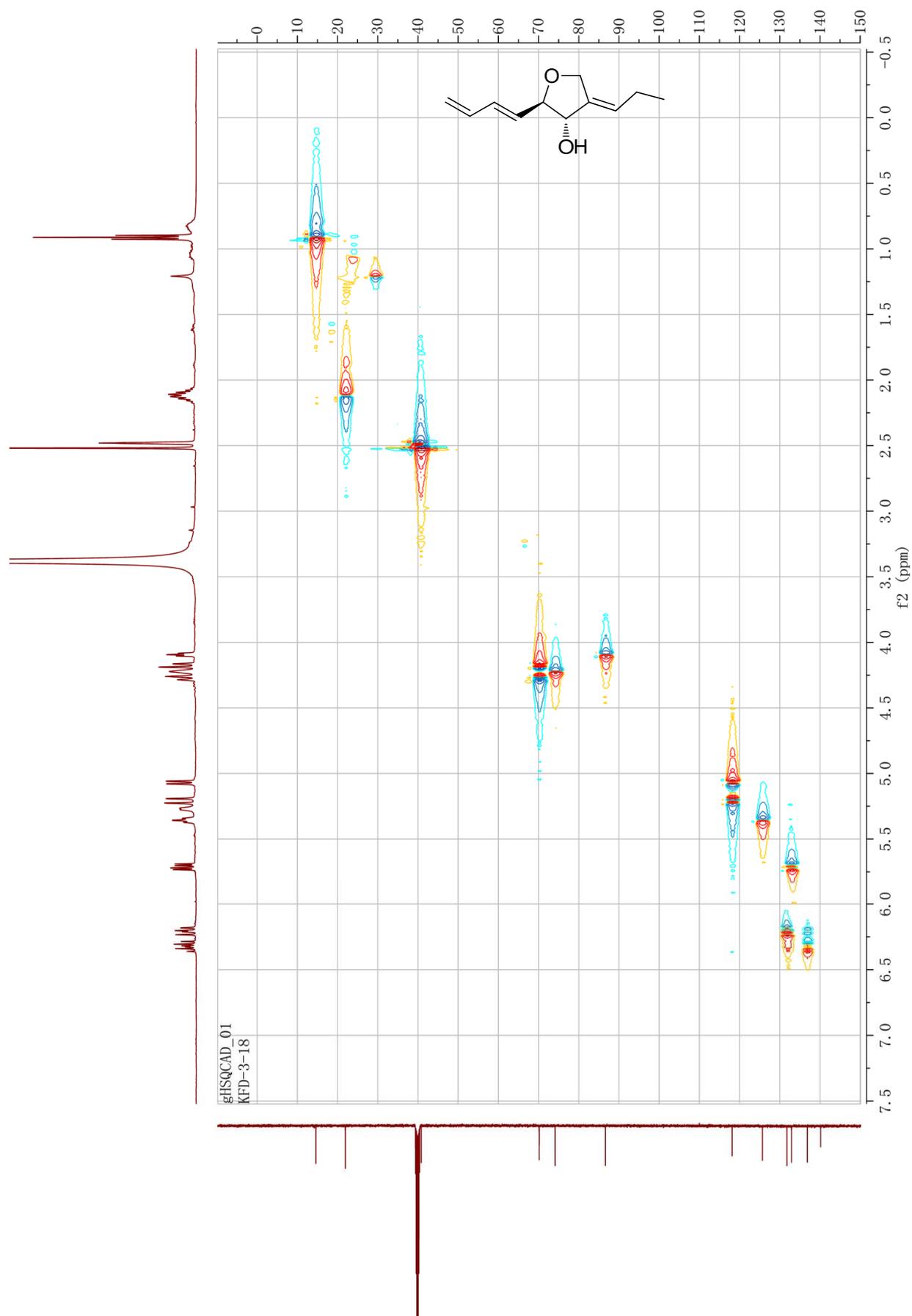


Figure S33. The ^1H - ^1H COSY spectrum of compound **6** in $\text{DMSO-}d_6$

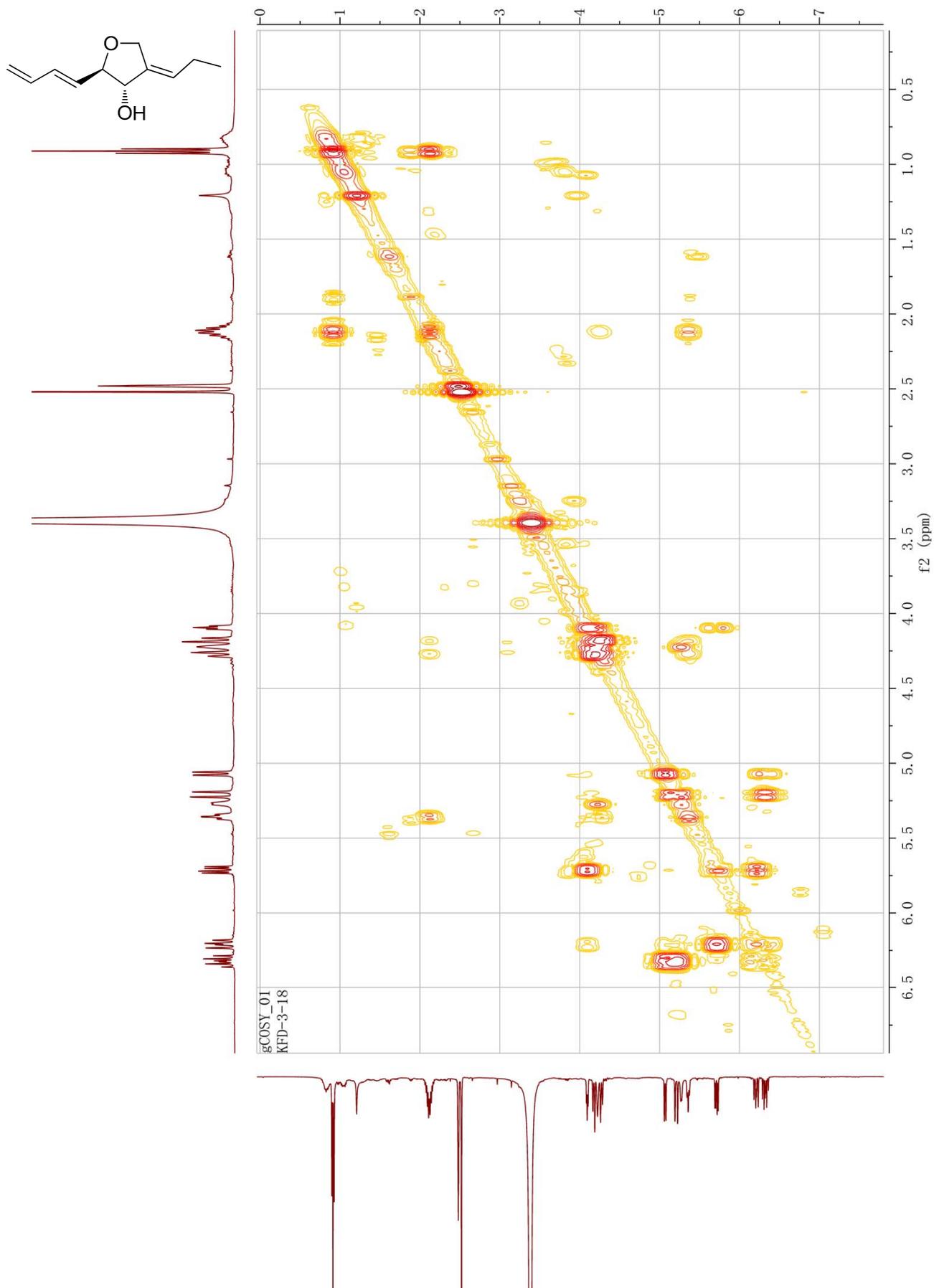


Figure S34. The HMBC spectrum of compound **6** in DMSO-*d*₆

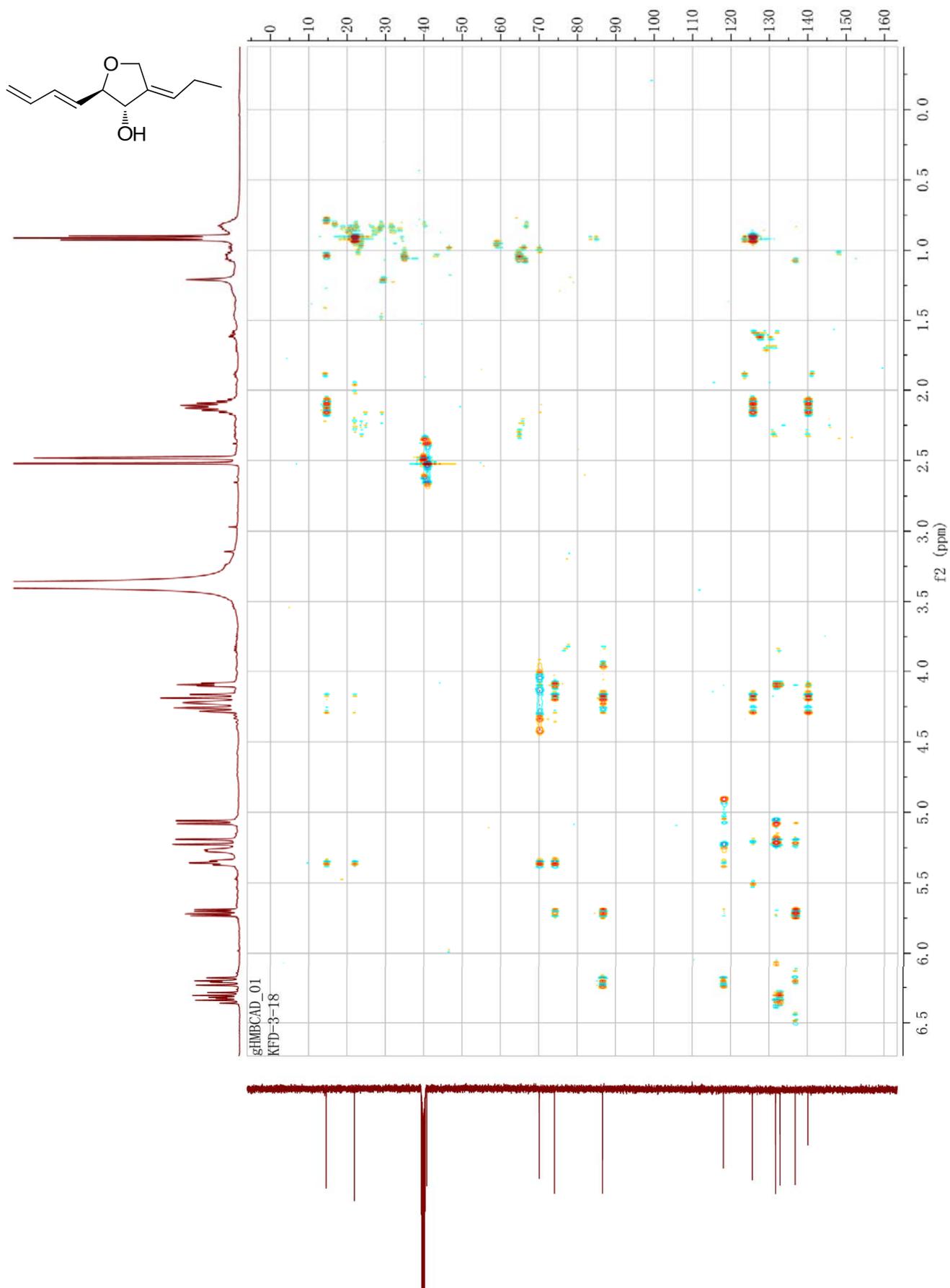


Figure S35. The NOE difference spectrum of compound **6** in DMSO-*d*₆ (I)

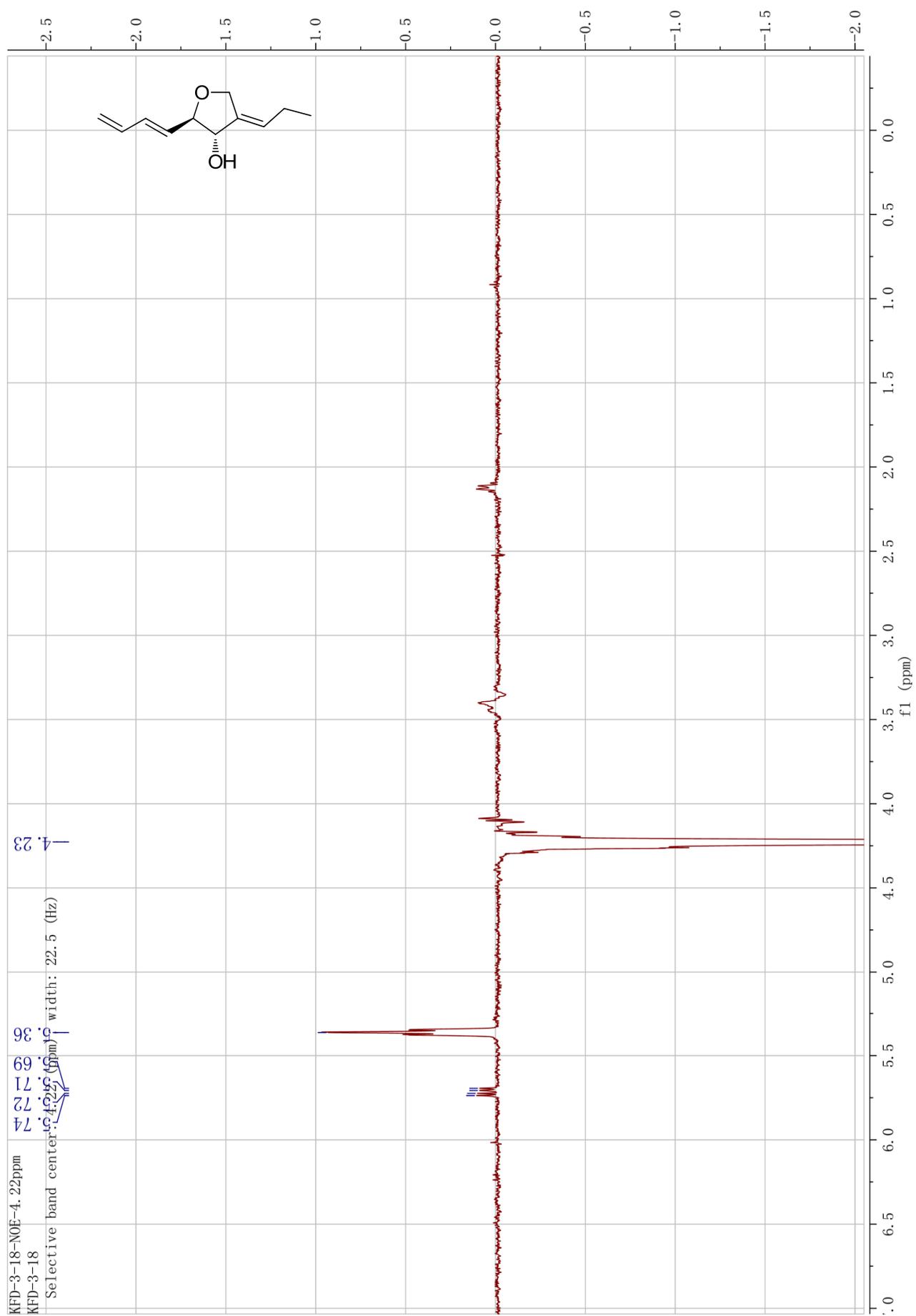


Figure S36. The NOESY difference spectrum of compound **6** in DMSO-*d*₆ (II)

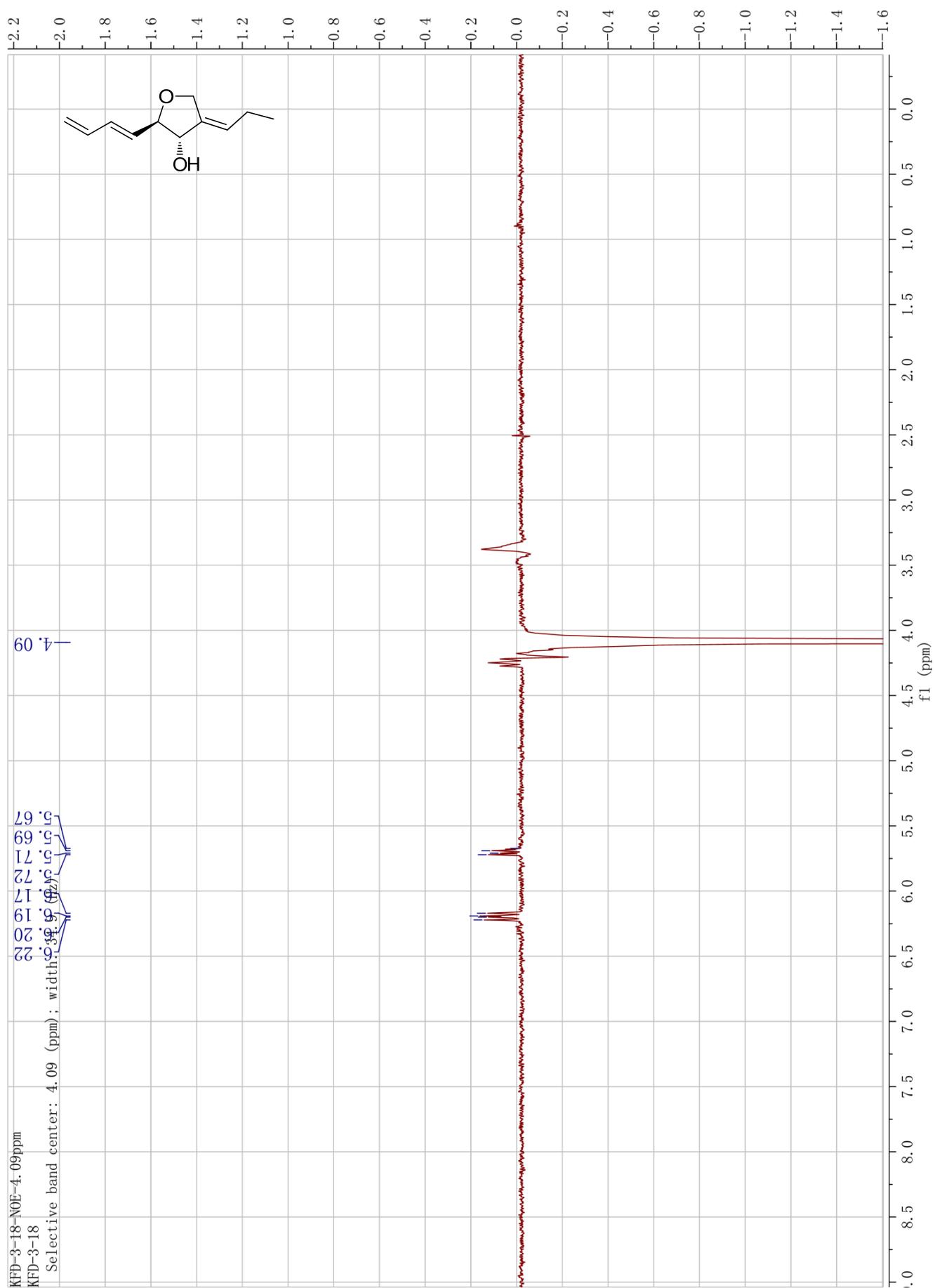


Figure S37. The ^1H -NMR spectrum of compounds **7** and **8** in CDCl_3

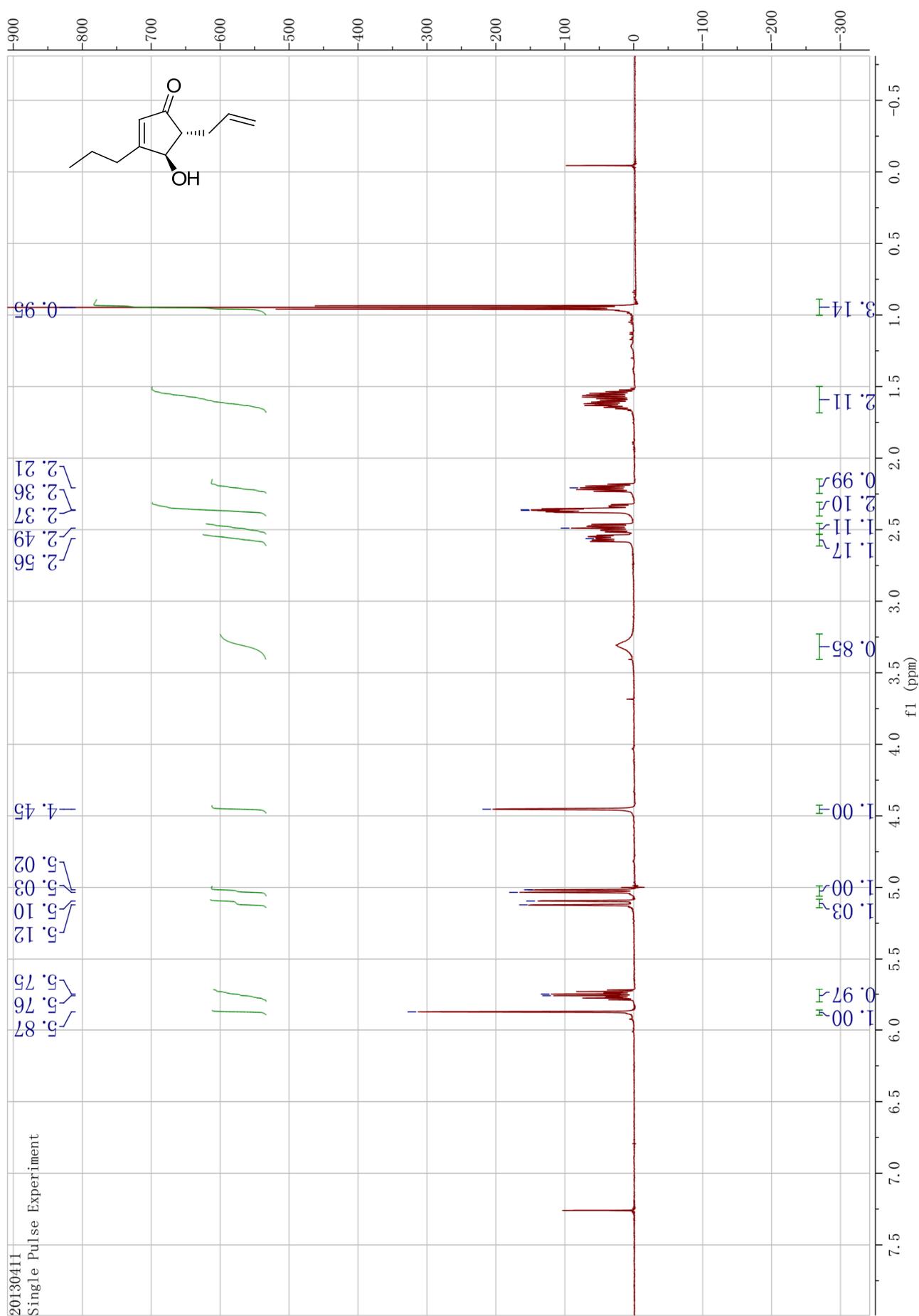


Figure S38. The ^{13}C -NMR spectrum of compounds **7** and **8** in CDCl_3

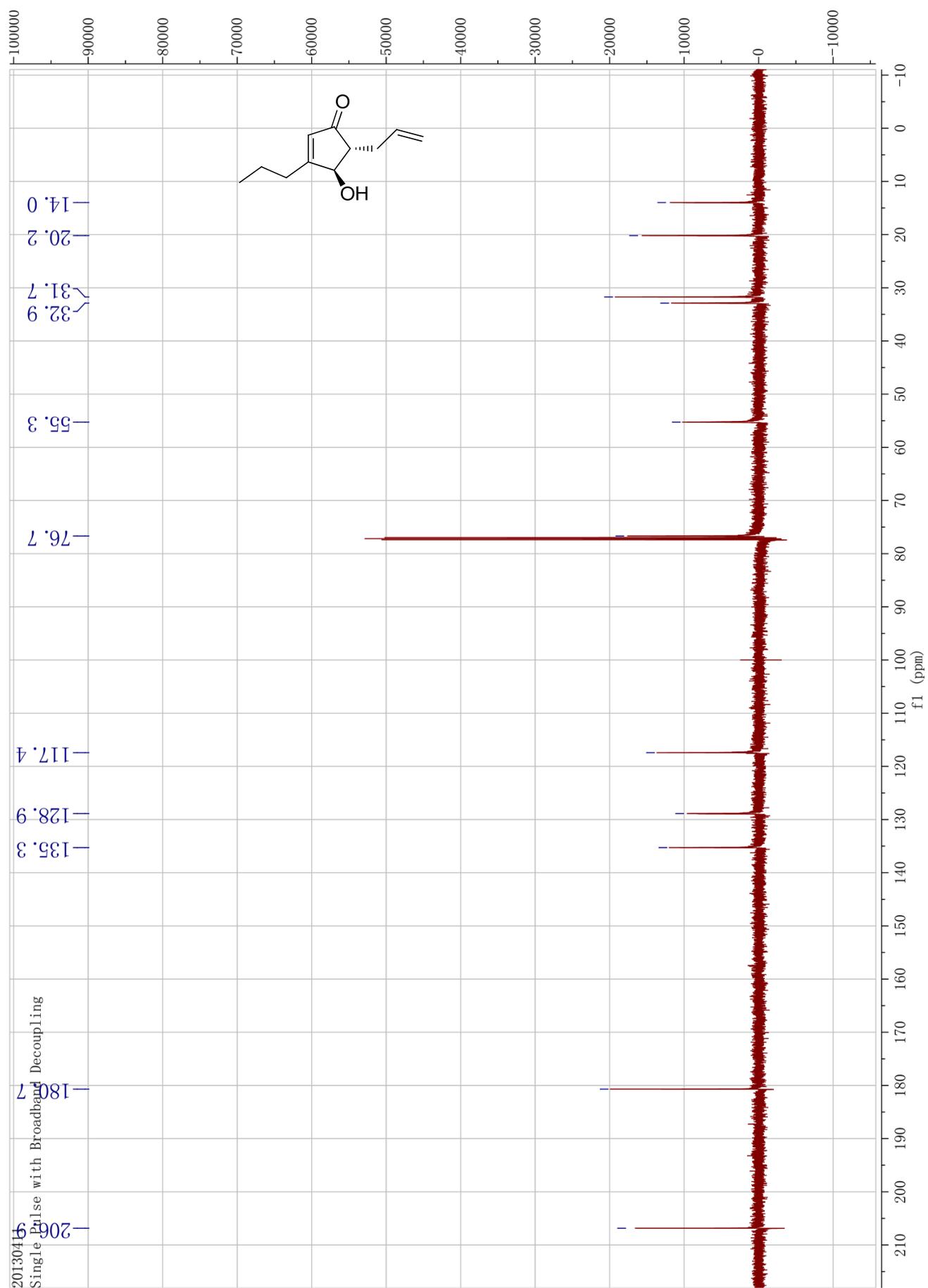


Figure S39. The DEPT spectrum of compounds **7** and **8** in CDCl₃

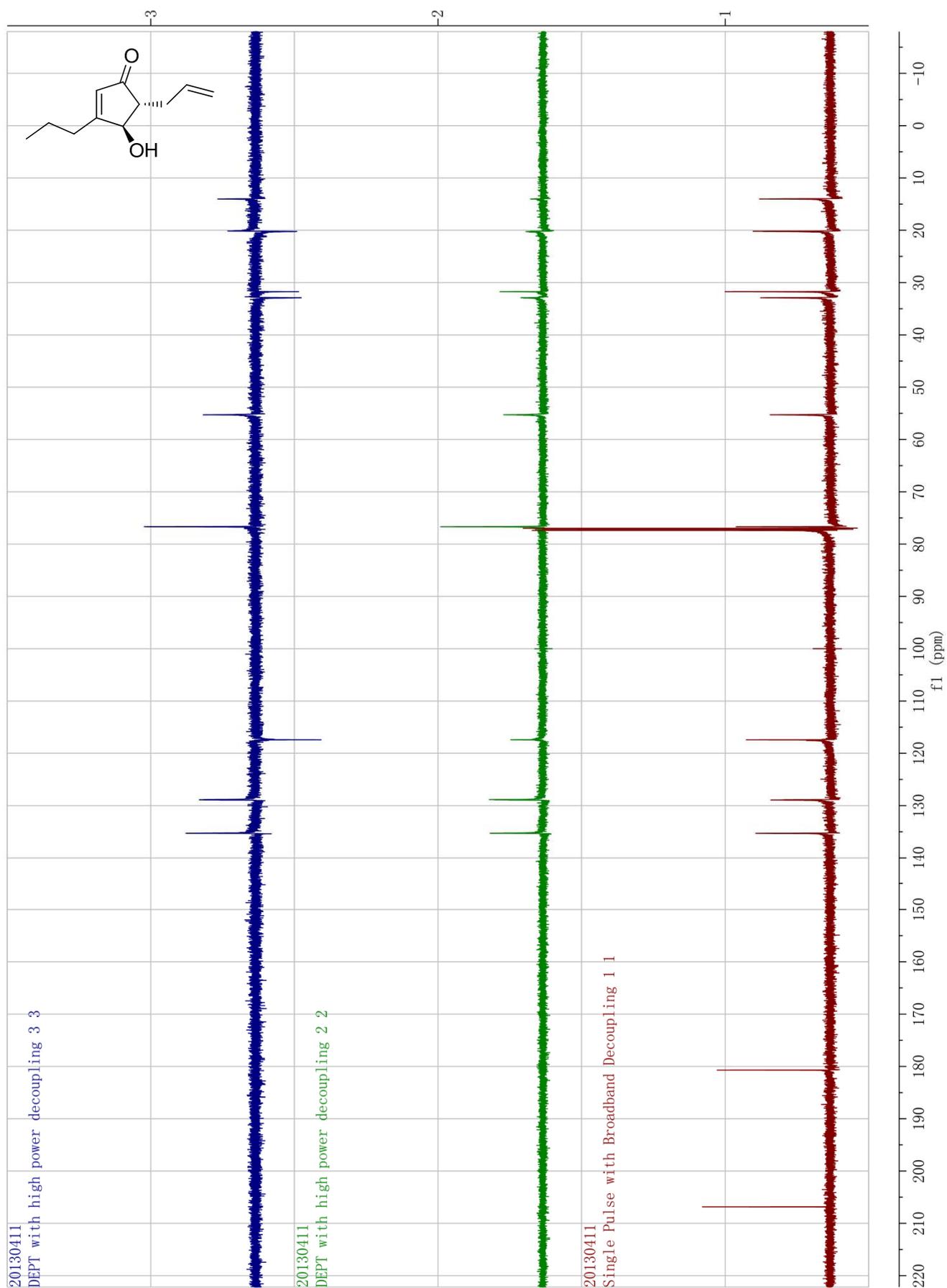


Figure S40. The HMQC spectrum of compounds **7** and **8** in CDCl₃

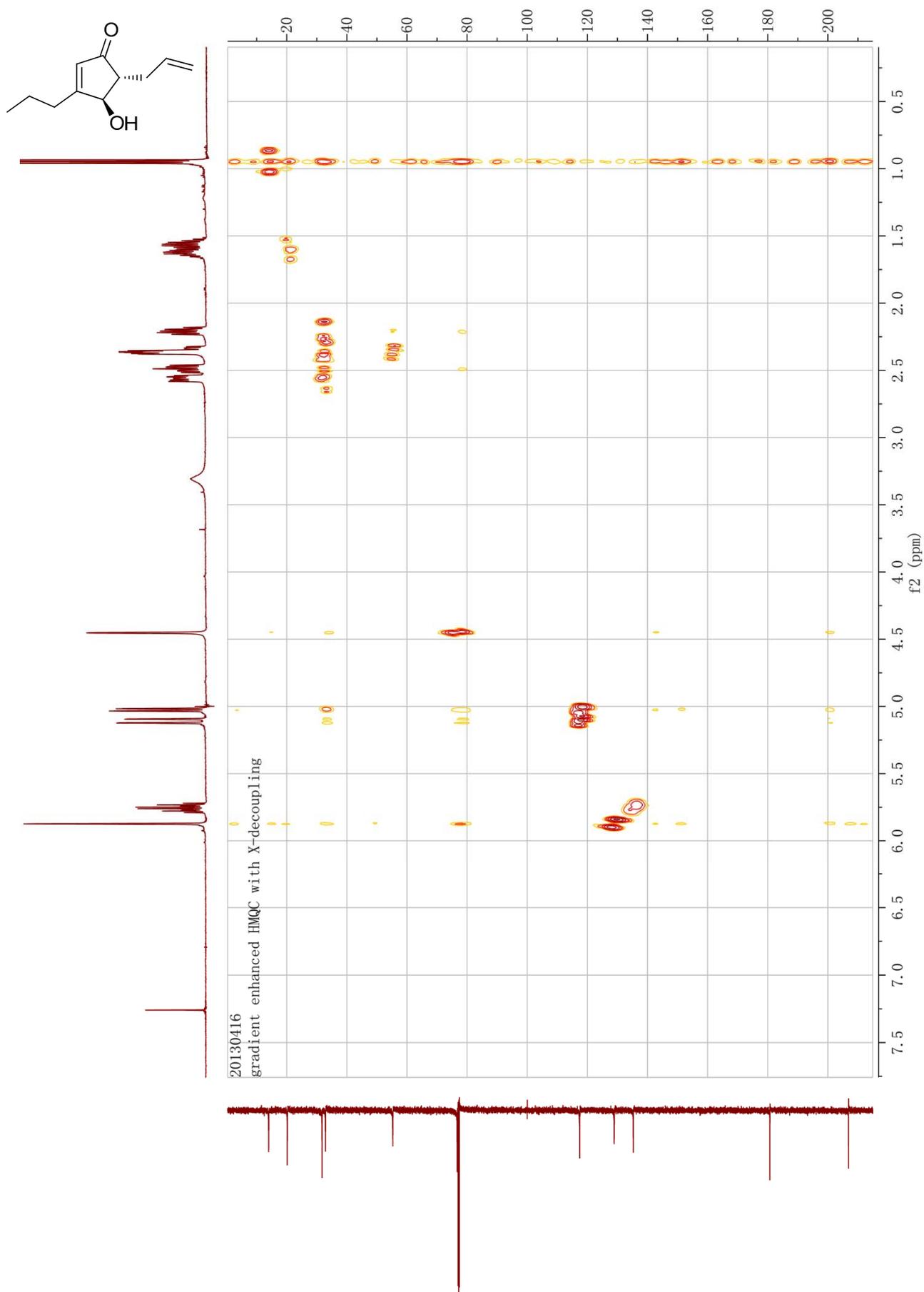


Figure S41. The ^1H - ^1H COSY spectrum of compounds **7** and **8** in CDCl_3

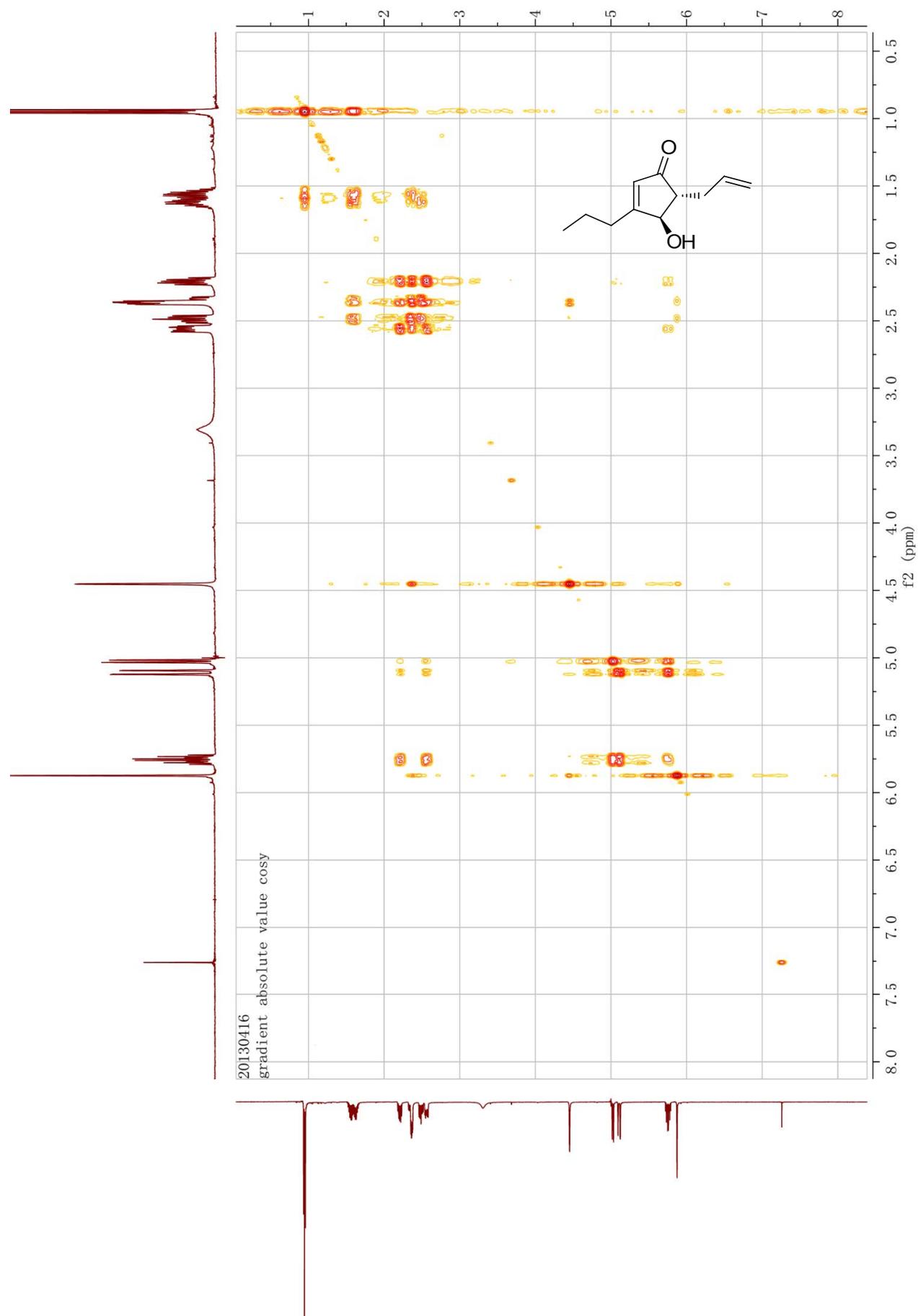


Figure S42. The HMBC spectrum of compounds **7** and **8** in CDCl₃

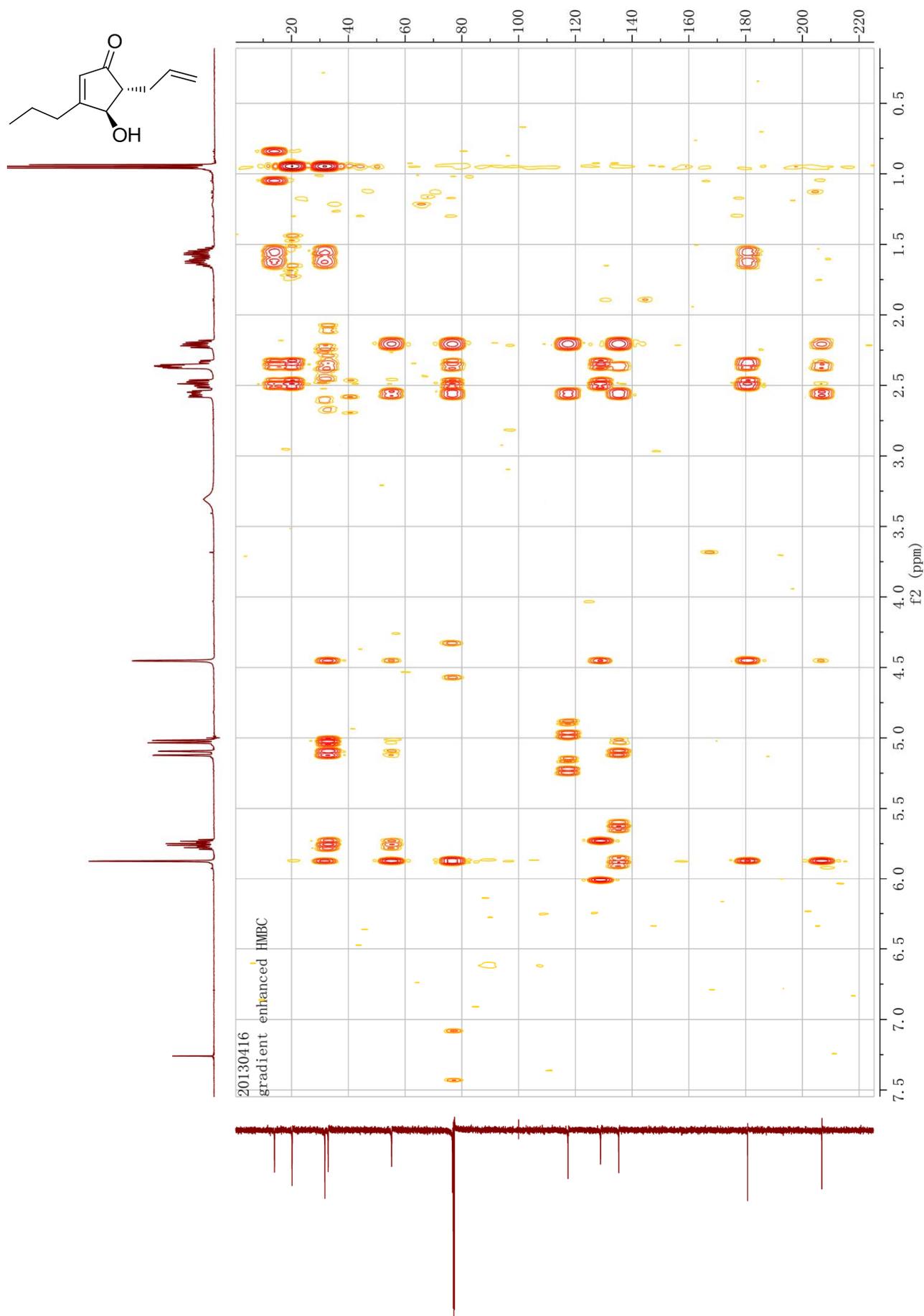


Figure S43. The NOE difference spectrum of compounds **7** and **8** in CDCl₃

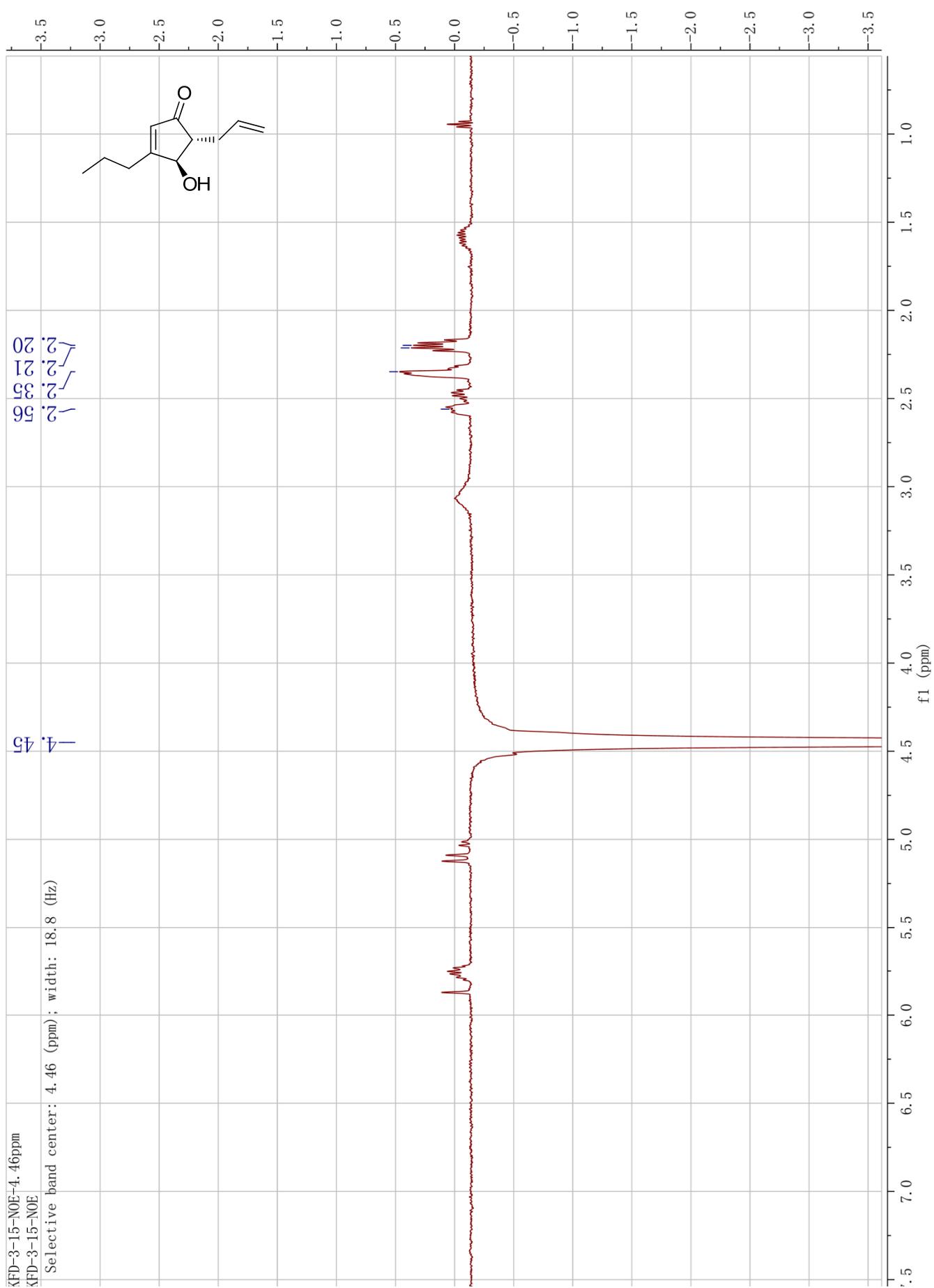


Figure S45. The ^{13}C -NMR spectrum of compound **9** in CDCl_3

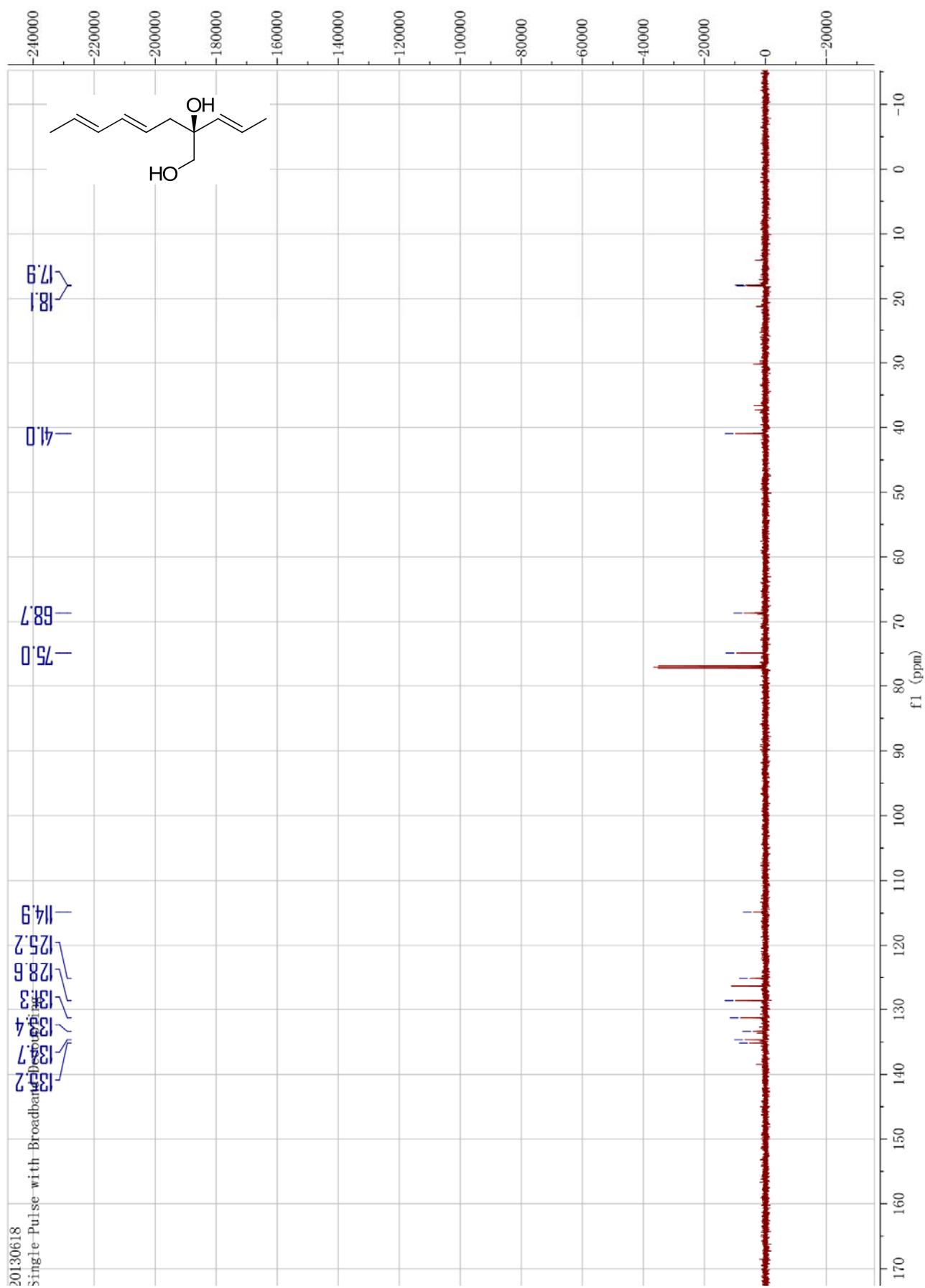


Figure S46. The DEPT spectrum of compound **9** in CDCl₃

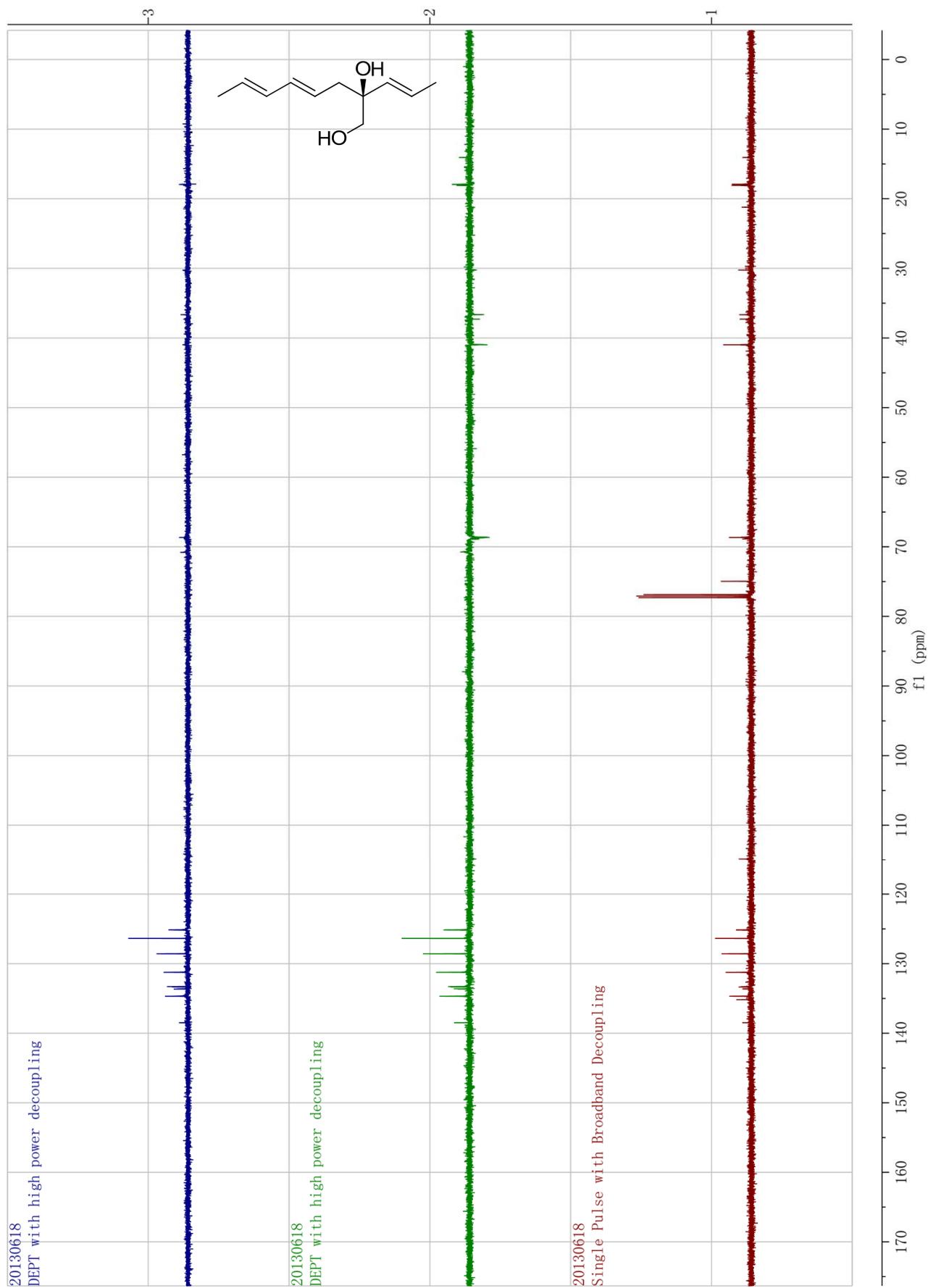


Figure S47. The HMQC spectrum of compound **9** in CDCl₃

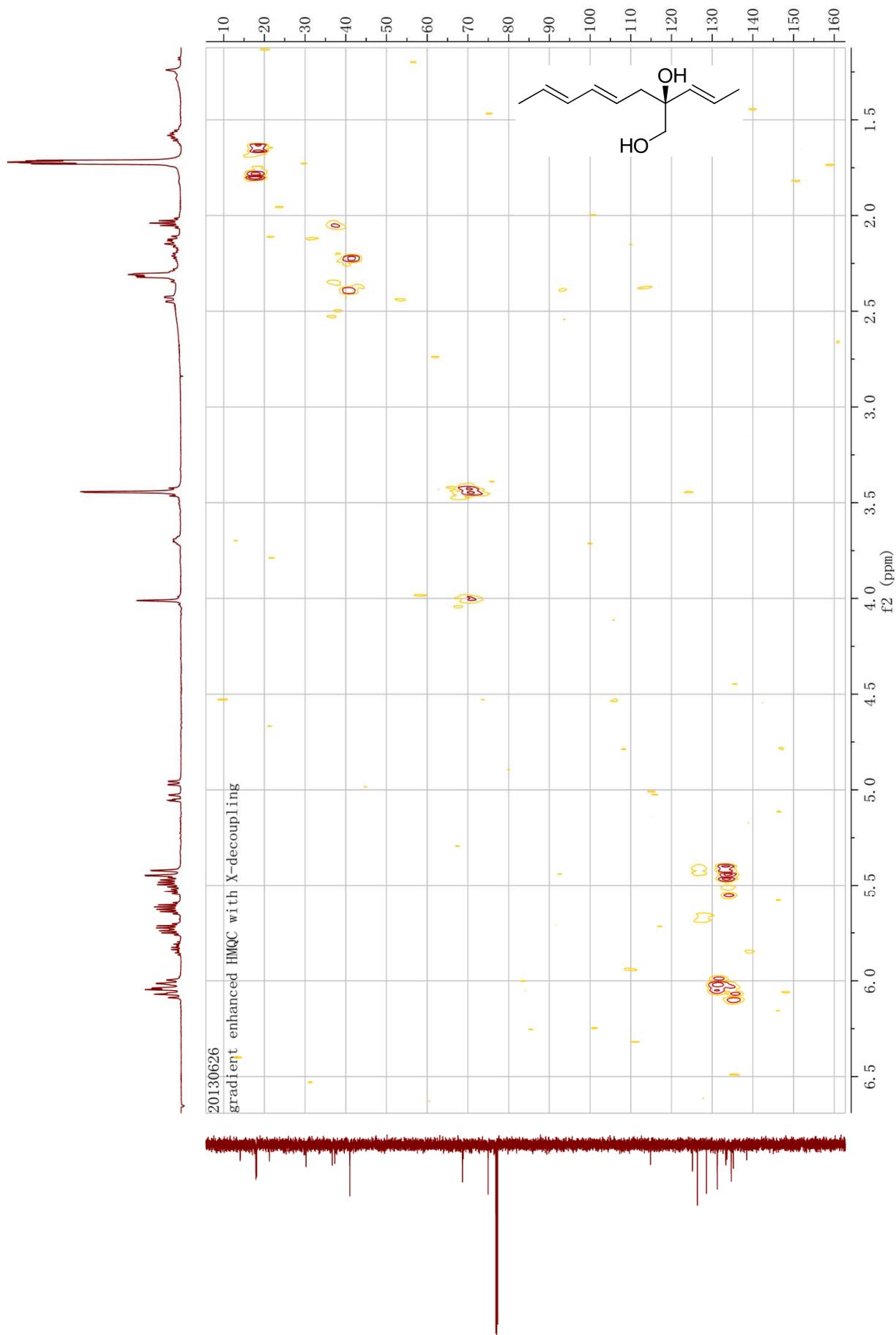


Figure S48. The ^1H - ^1H COSY spectrum of compound **9** in CDCl_3

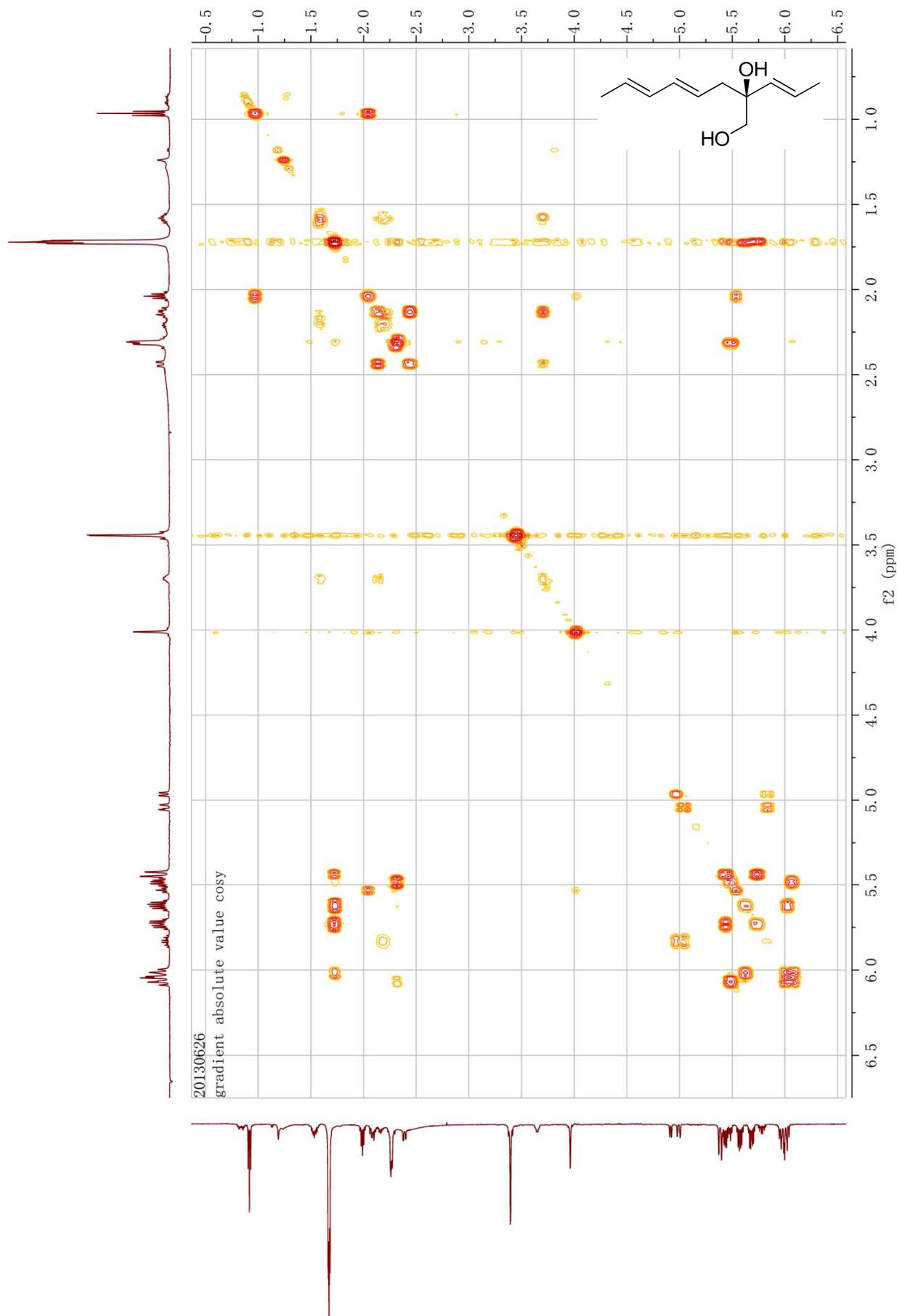


Figure S49. The HMBC spectrum of compound **9** in CDCl₃

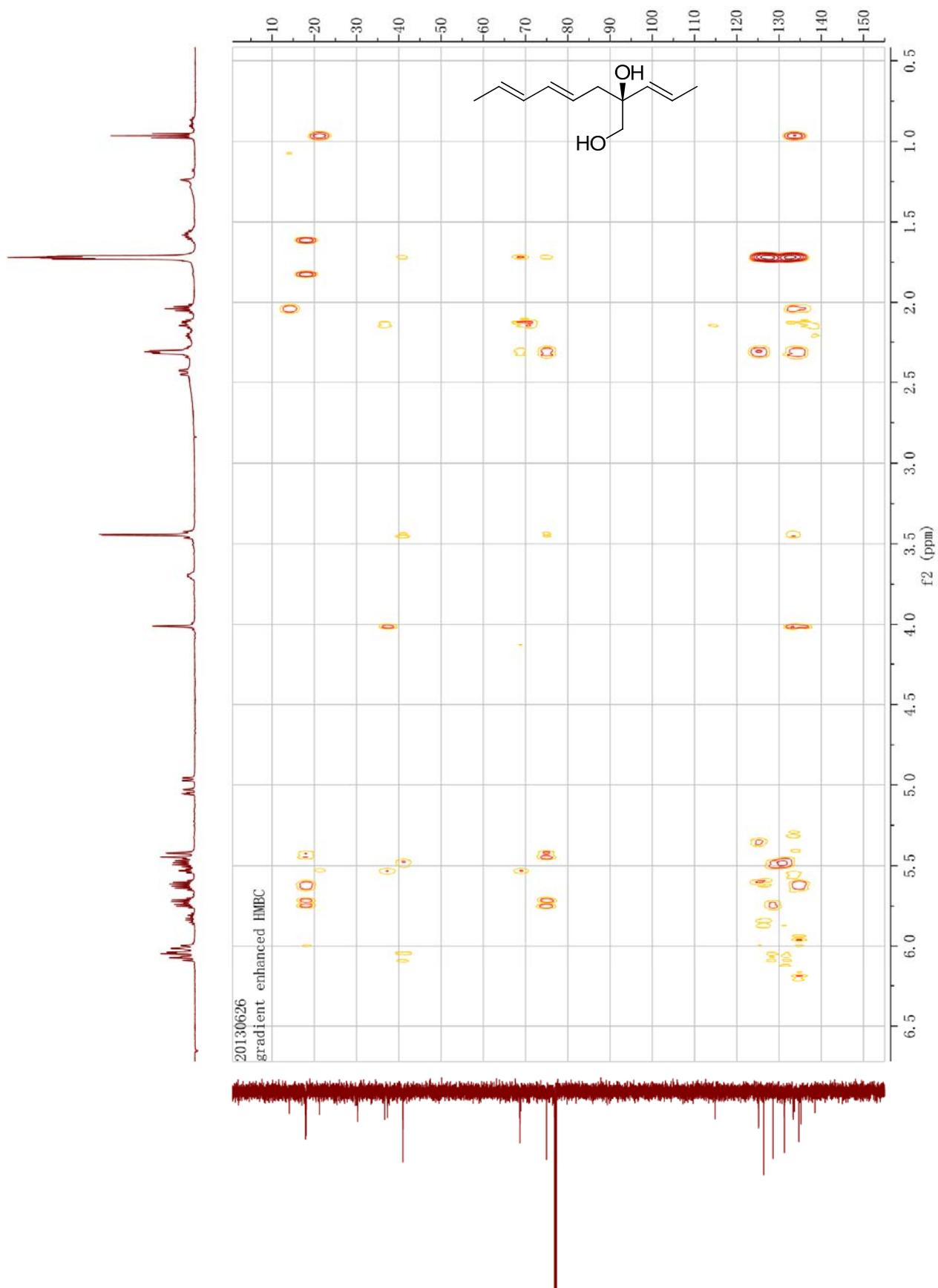


Figure S50. The ^1H -NMR spectrum of compound **10** in $\text{DMSO-}d_6$

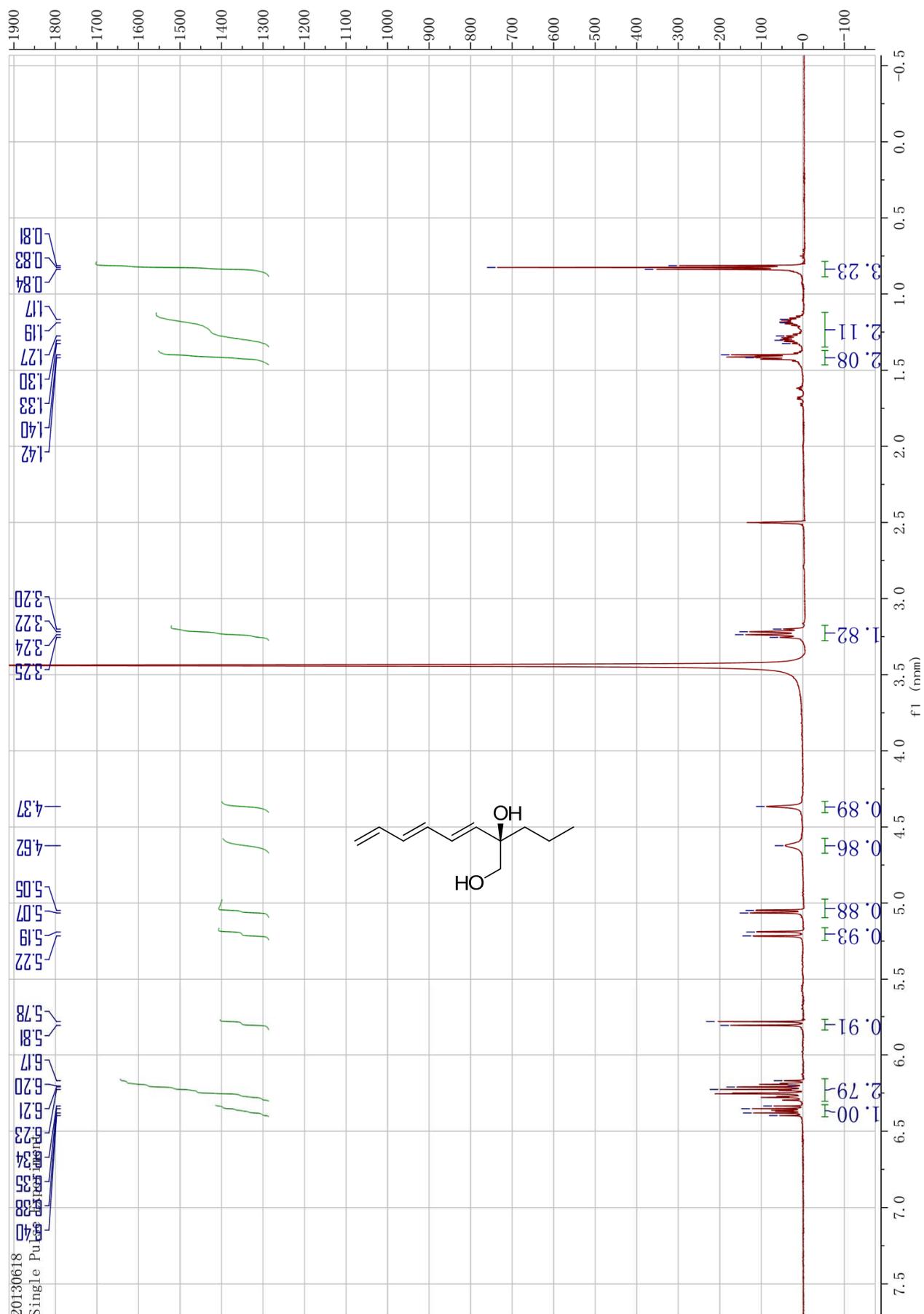


Figure S51. The ^{13}C -NMR spectrum of compound **10** in $\text{DMSO-}d_6$

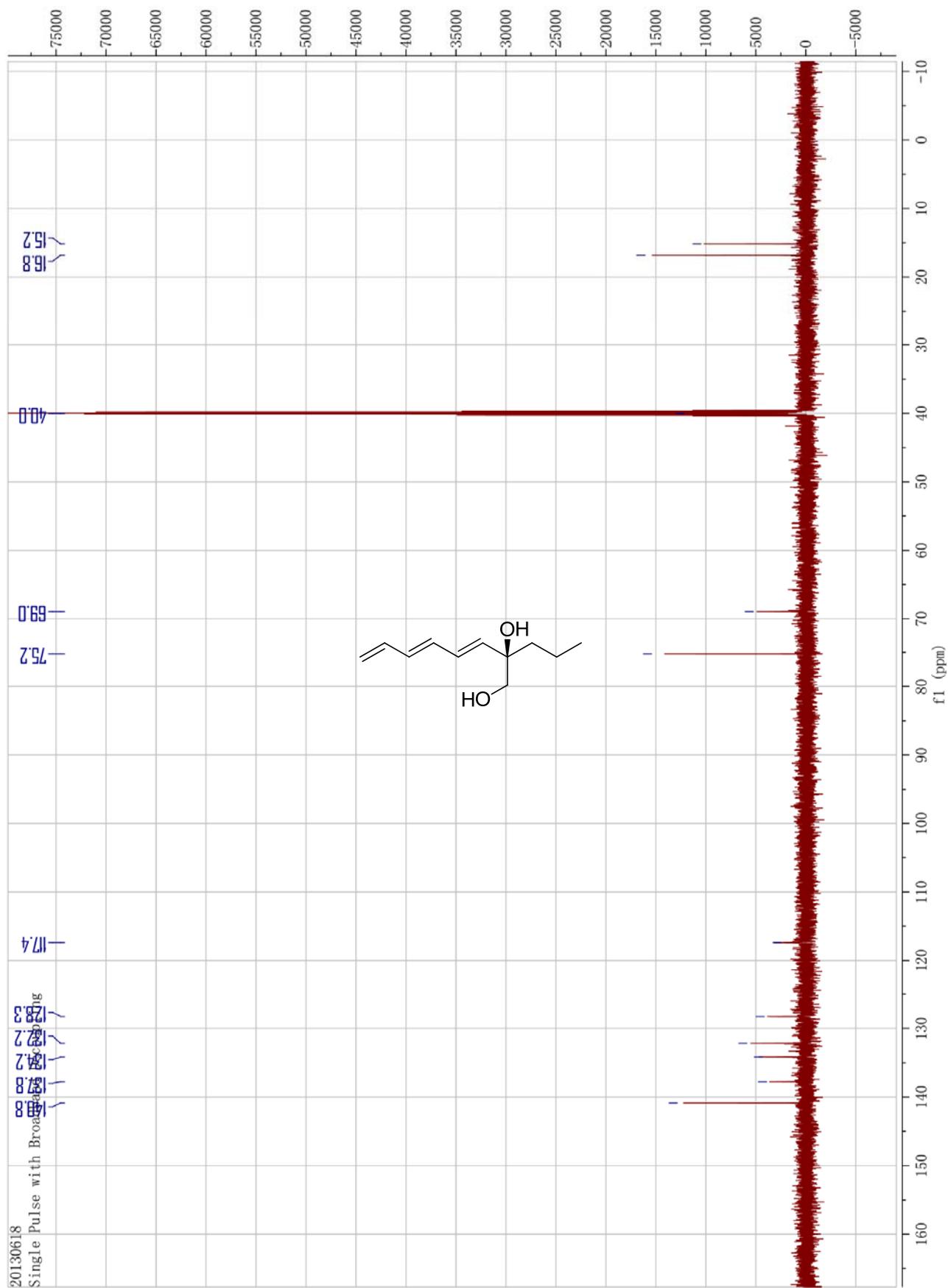


Figure S52. The DEPT spectrum of compound 10 in DMSO-*d*₆

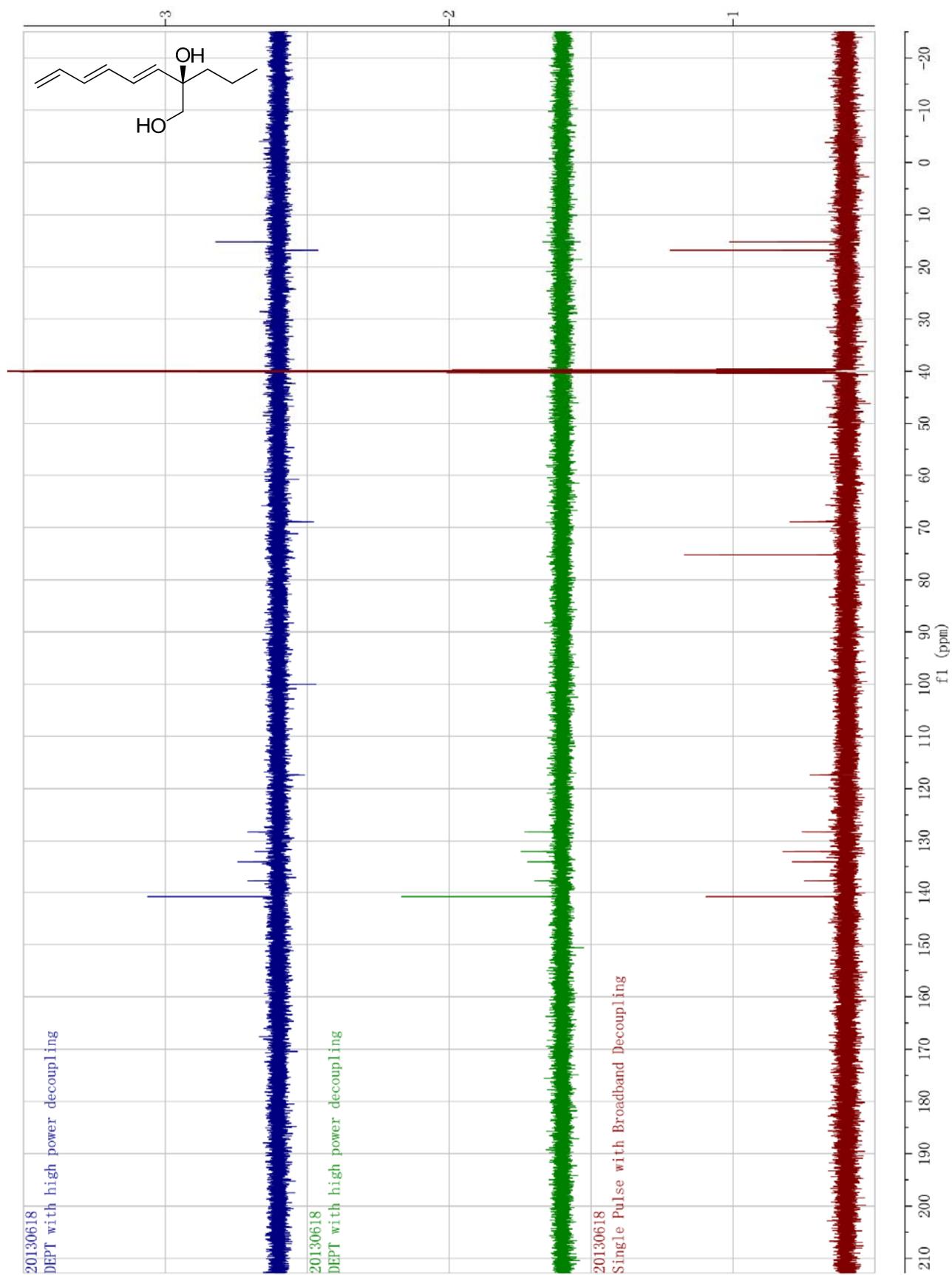


Figure S53. The HMQC spectrum of compound **10** in DMSO-*d*₆

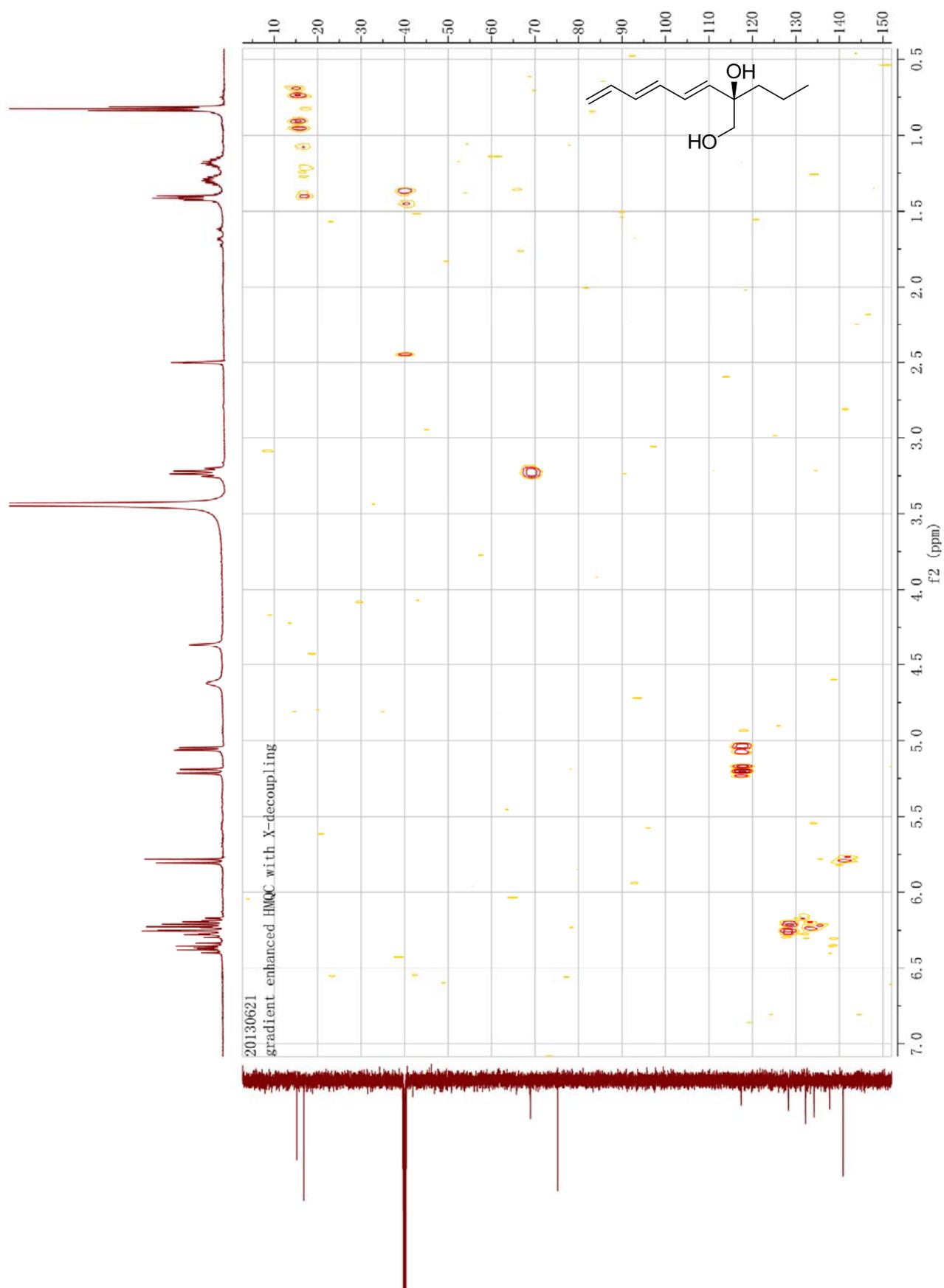


Figure S54. The ^1H - ^1H COSY spectrum of compound **10** in $\text{DMSO-}d_6$

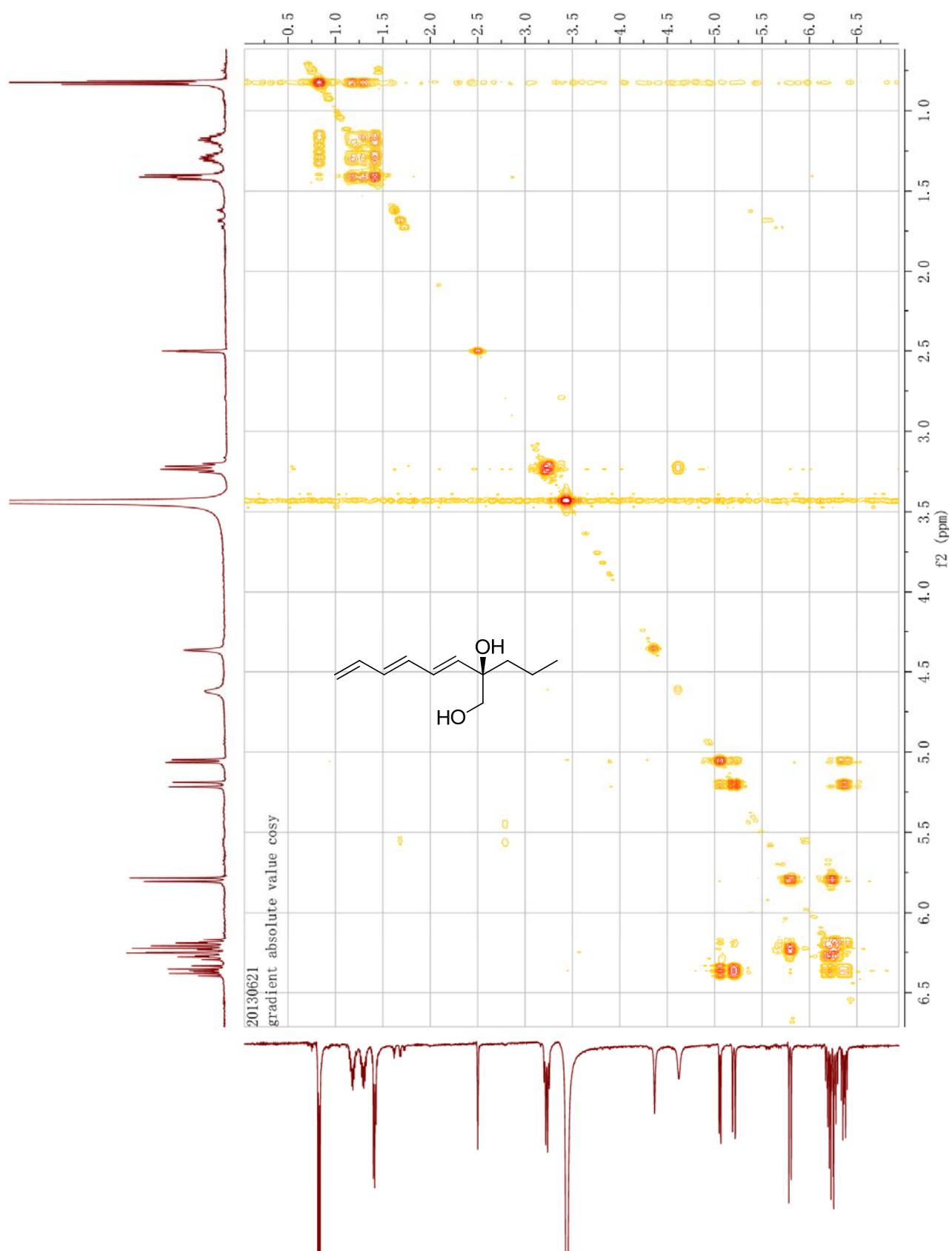


Figure S56. The ^1H -NMR spectrum of compound **11** in $\text{DMSO-}d_6$

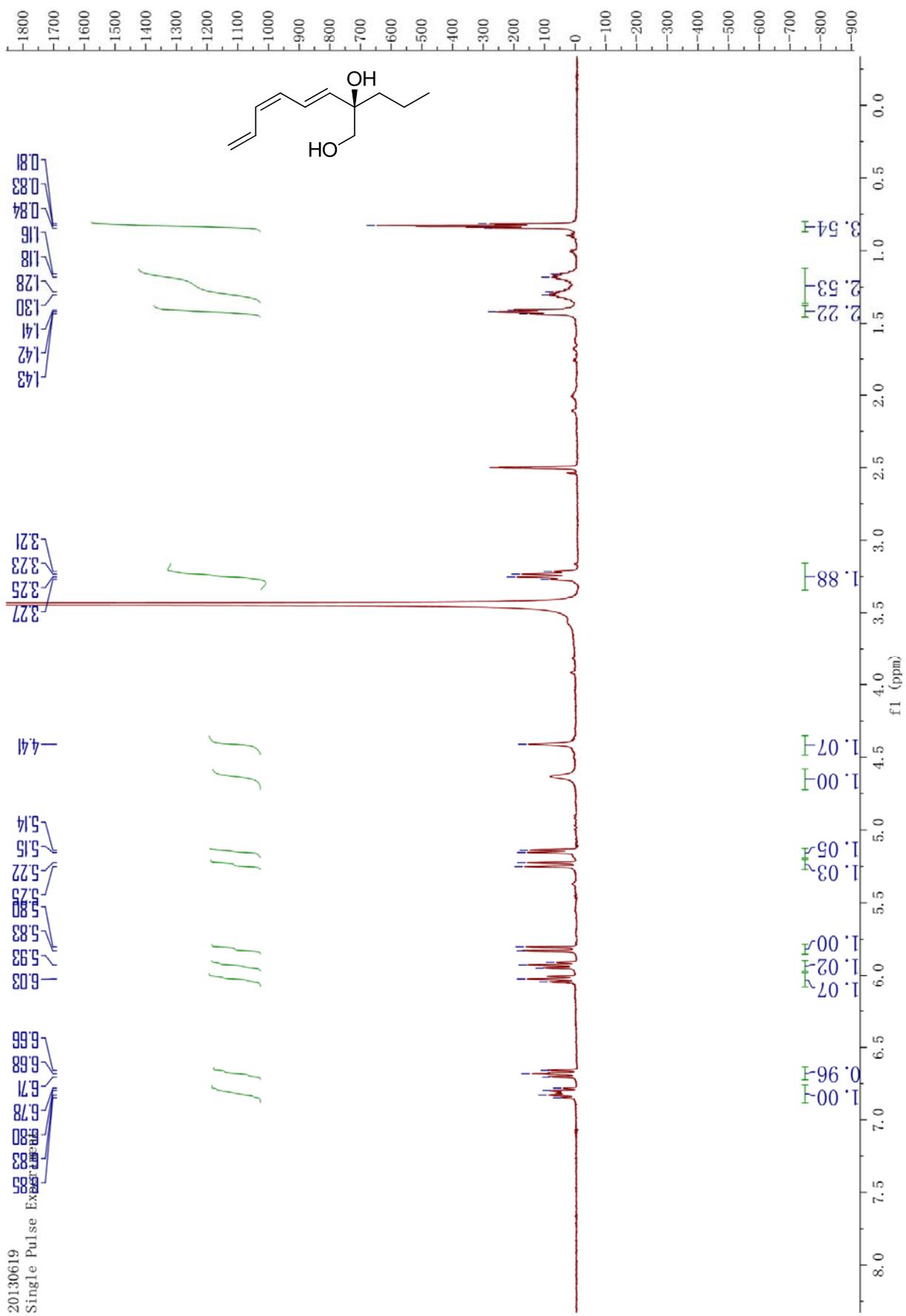


Figure S58. The DEPT spectrum of compound 11 in DMSO-*d*₆

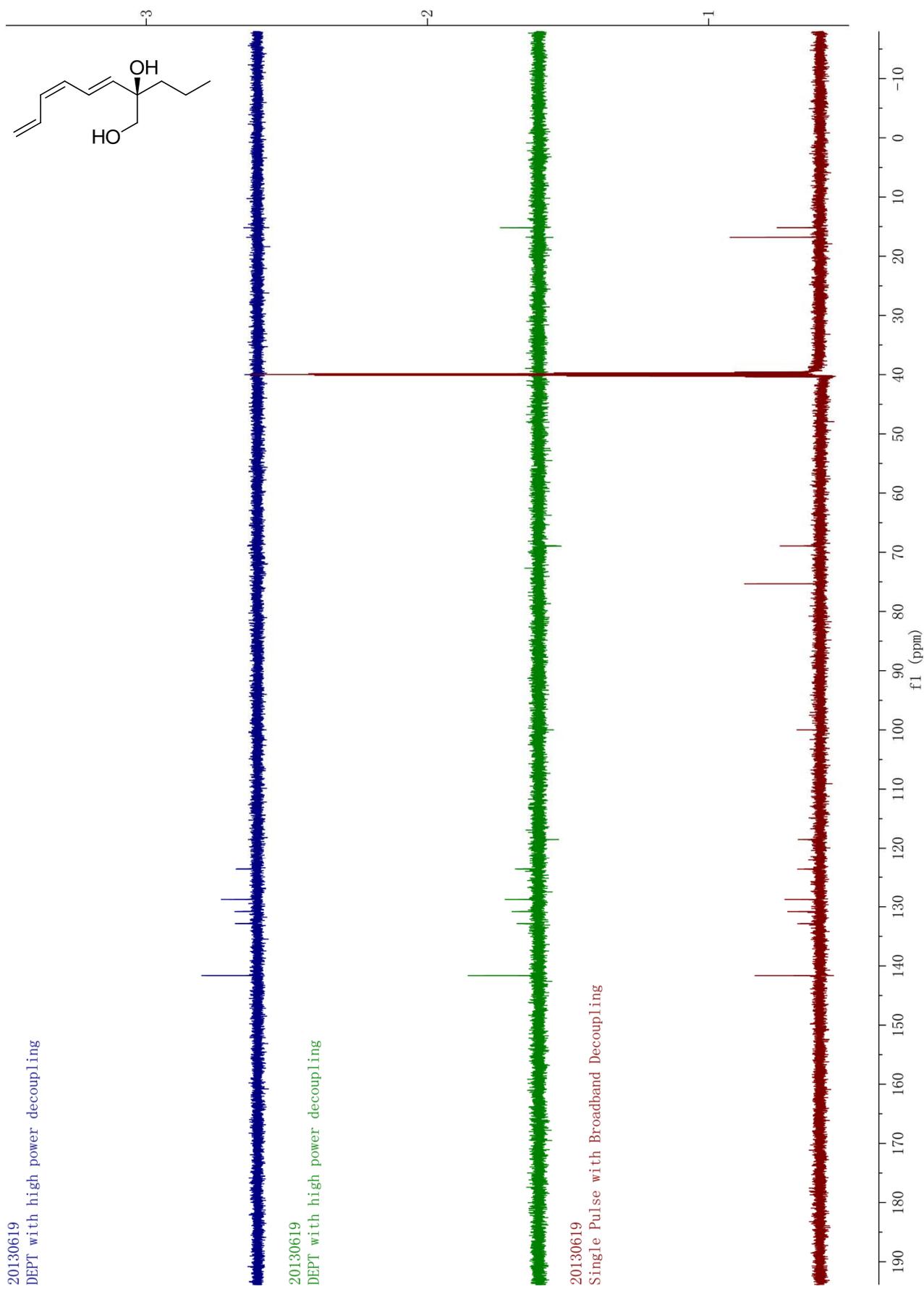


Figure S59. The HMQC spectrum of compound **11** in DMSO- d_6

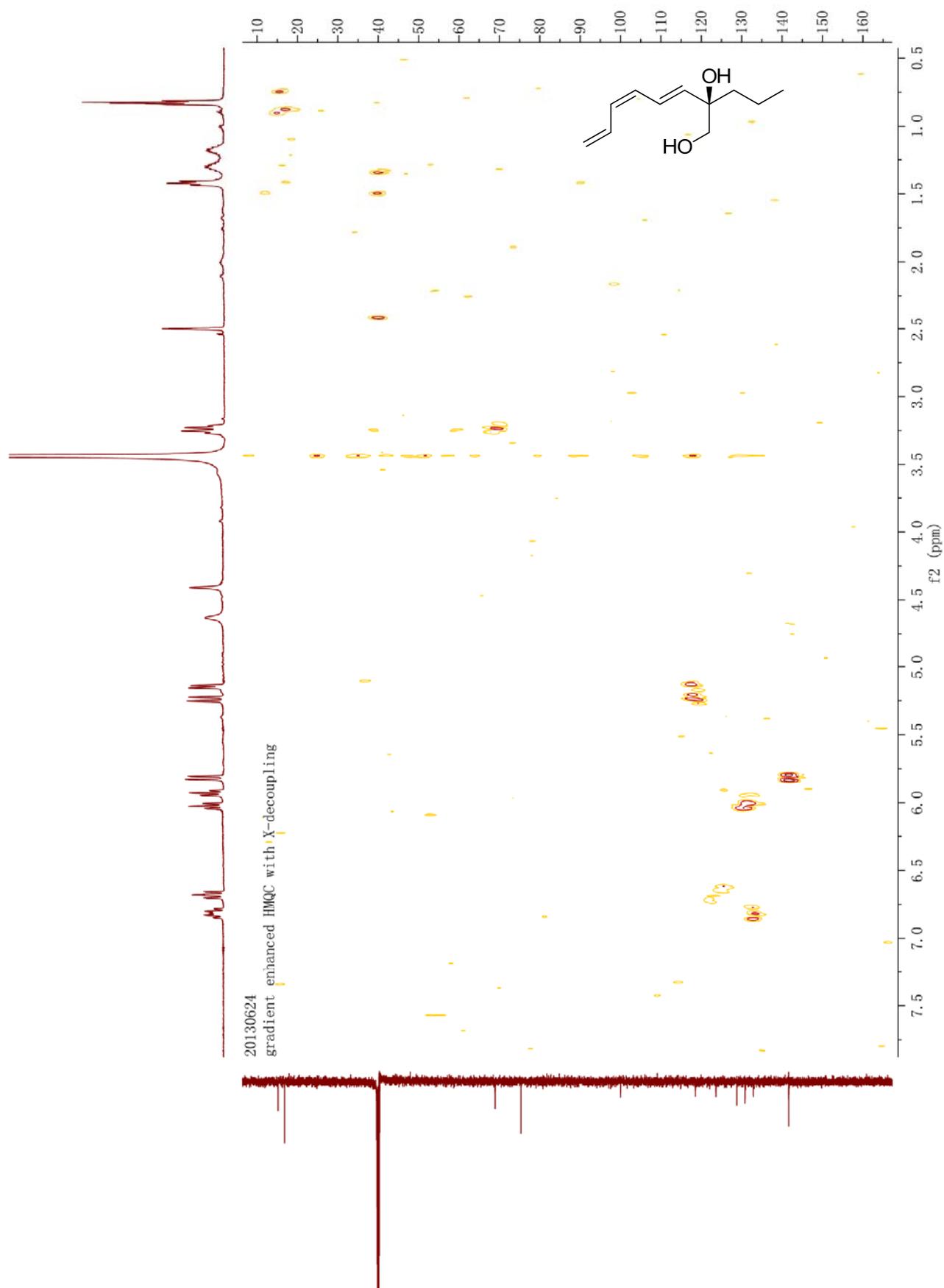


Figure S60. The ^1H - ^1H COSY spectrum of compound **11** in $\text{DMSO-}d_6$

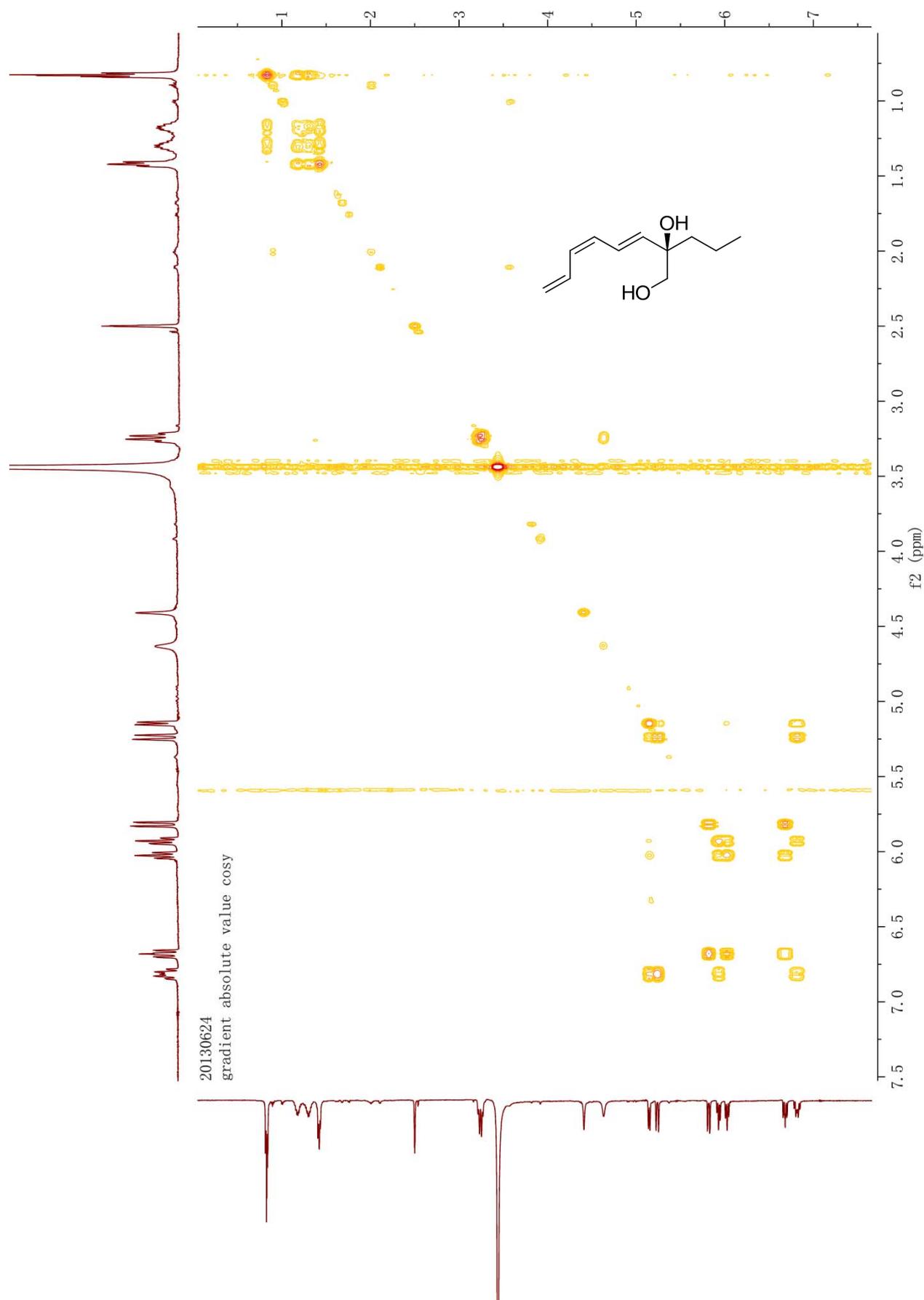


Figure S61. The HMBC spectrum of compound **11** in DMSO-*d*₆

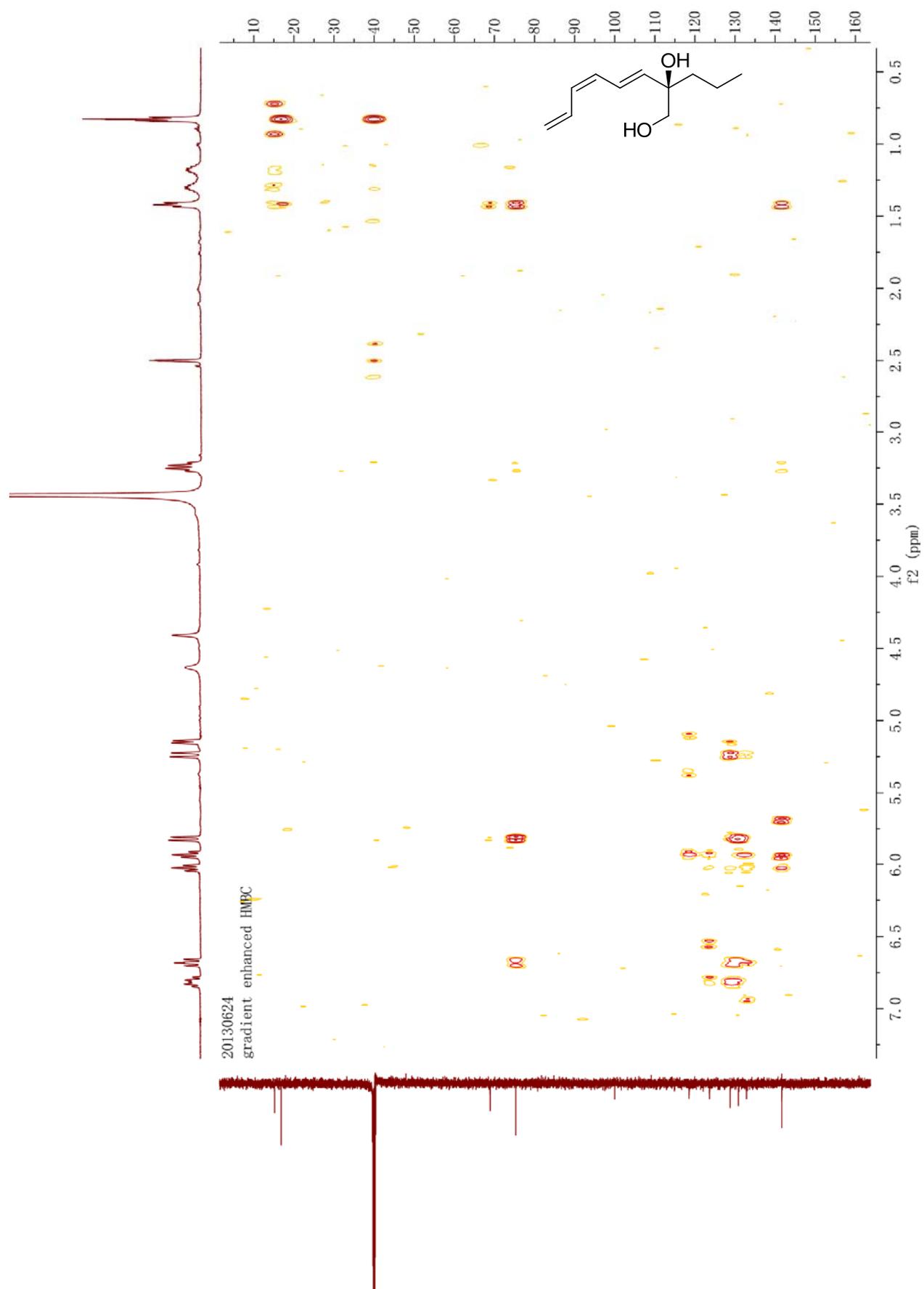


Figure S62. The ^1H -NMR spectrum of compound **12** in $\text{DMSO-}d_6$

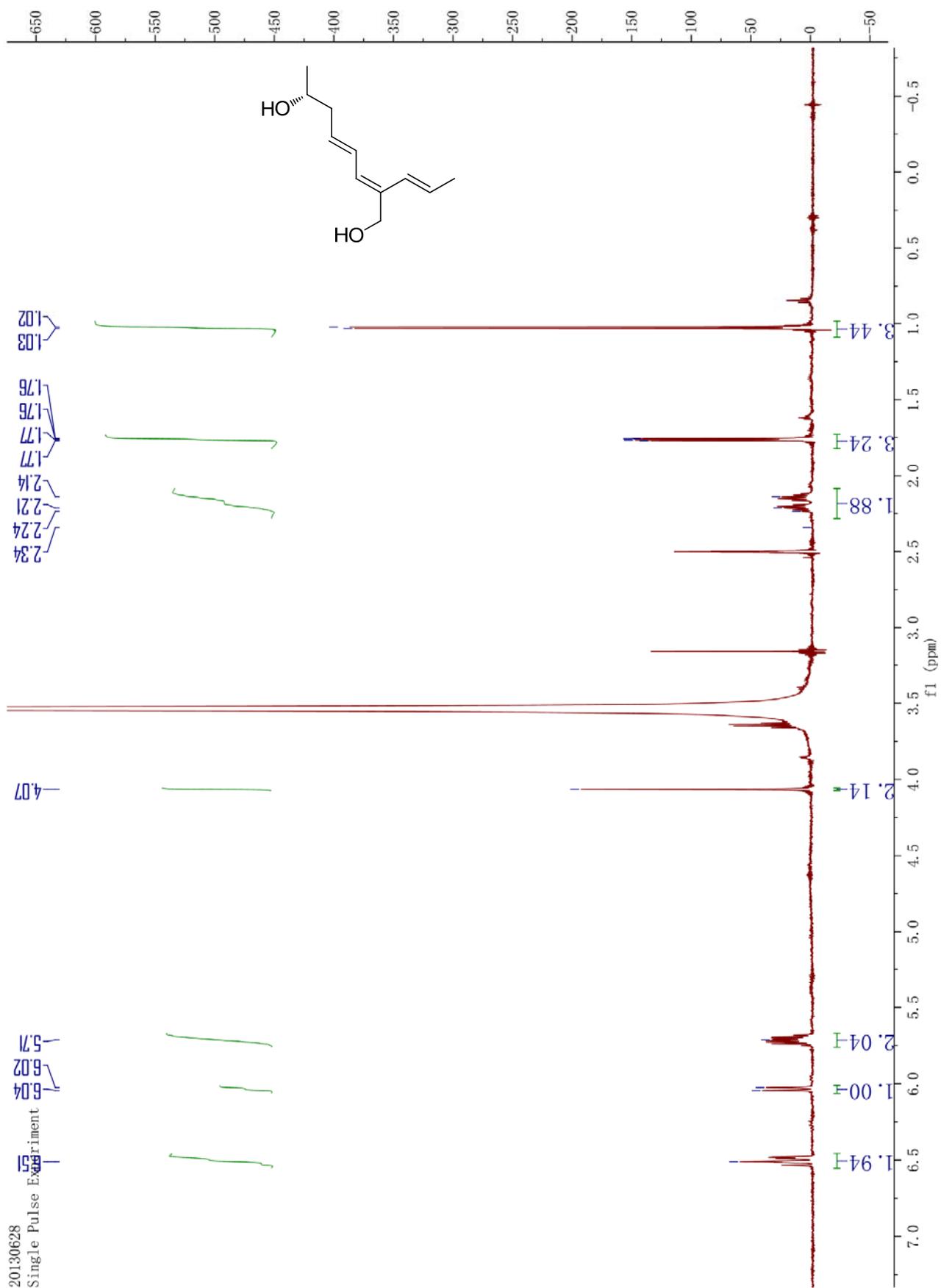


Figure S63. The ^{13}C -NMR spectrum of compound **12** in $\text{DMSO-}d_6$

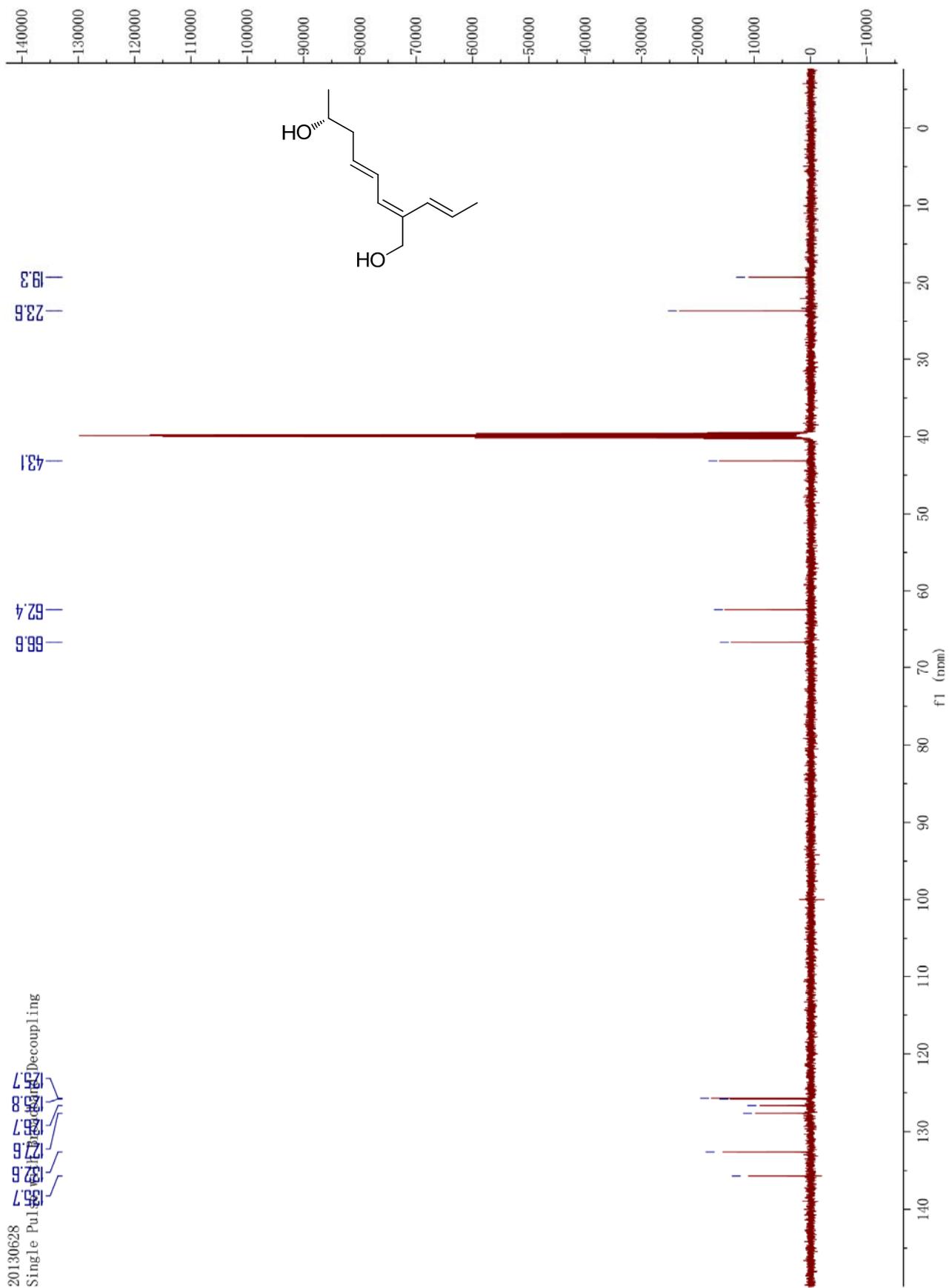


Figure S64. The DEPT spectrum of compound **12** in DMSO-*d*₆

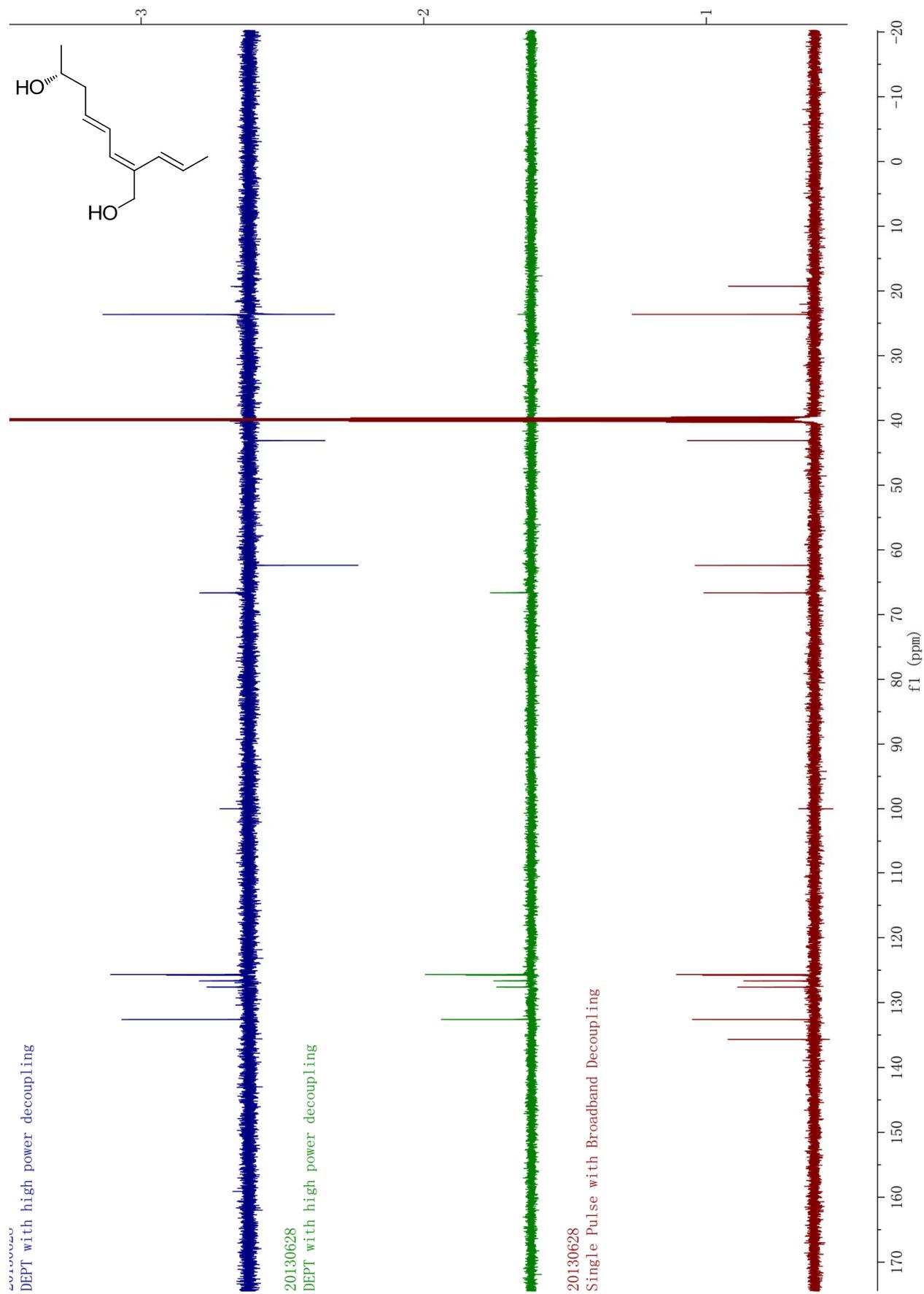


Figure S65. The HMQC spectrum of compound **12** in DMSO-*d*₆

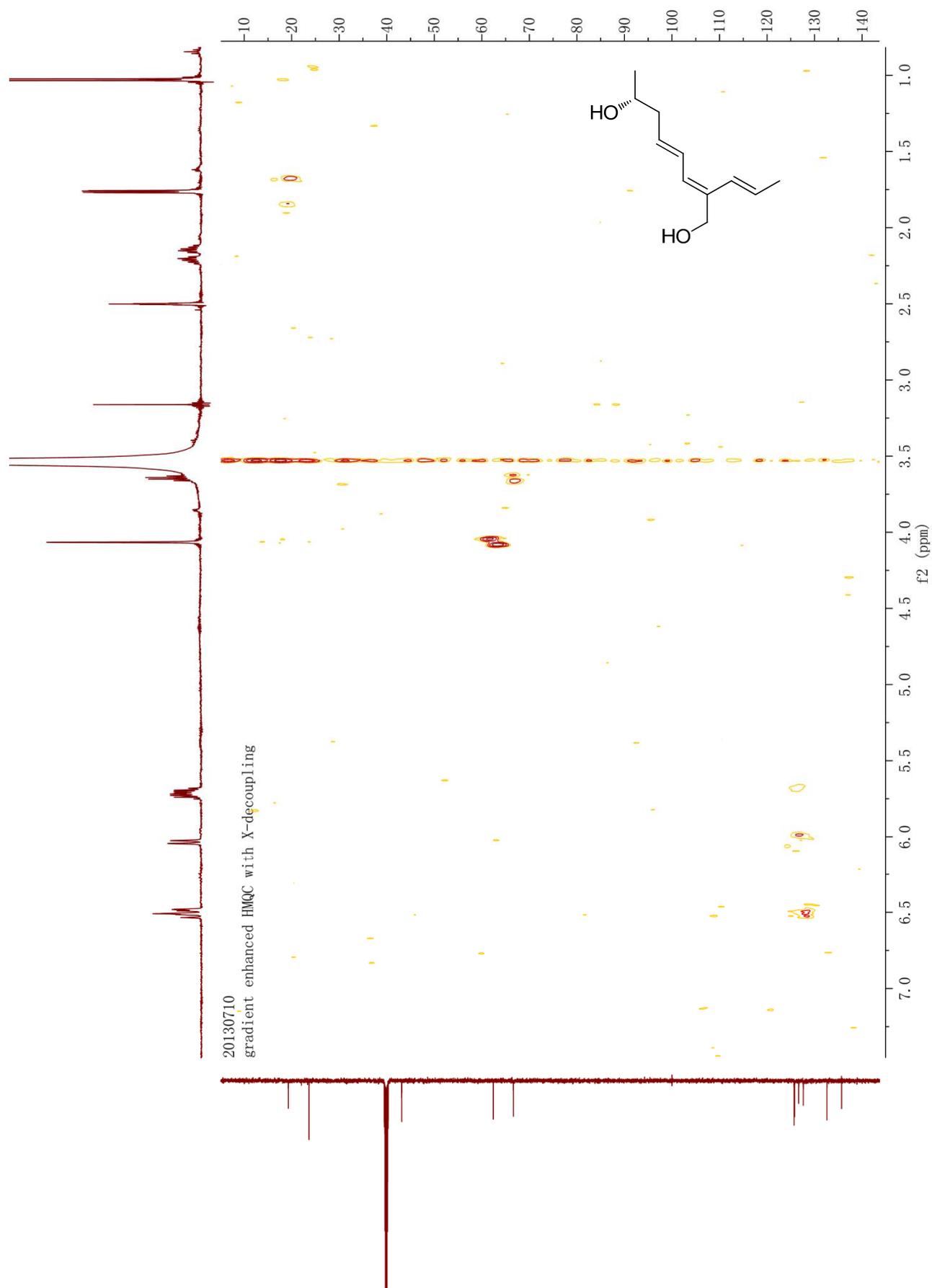


Figure S66. The ^1H - ^1H COSY spectrum of compound 12 in $\text{DMSO-}d_6$

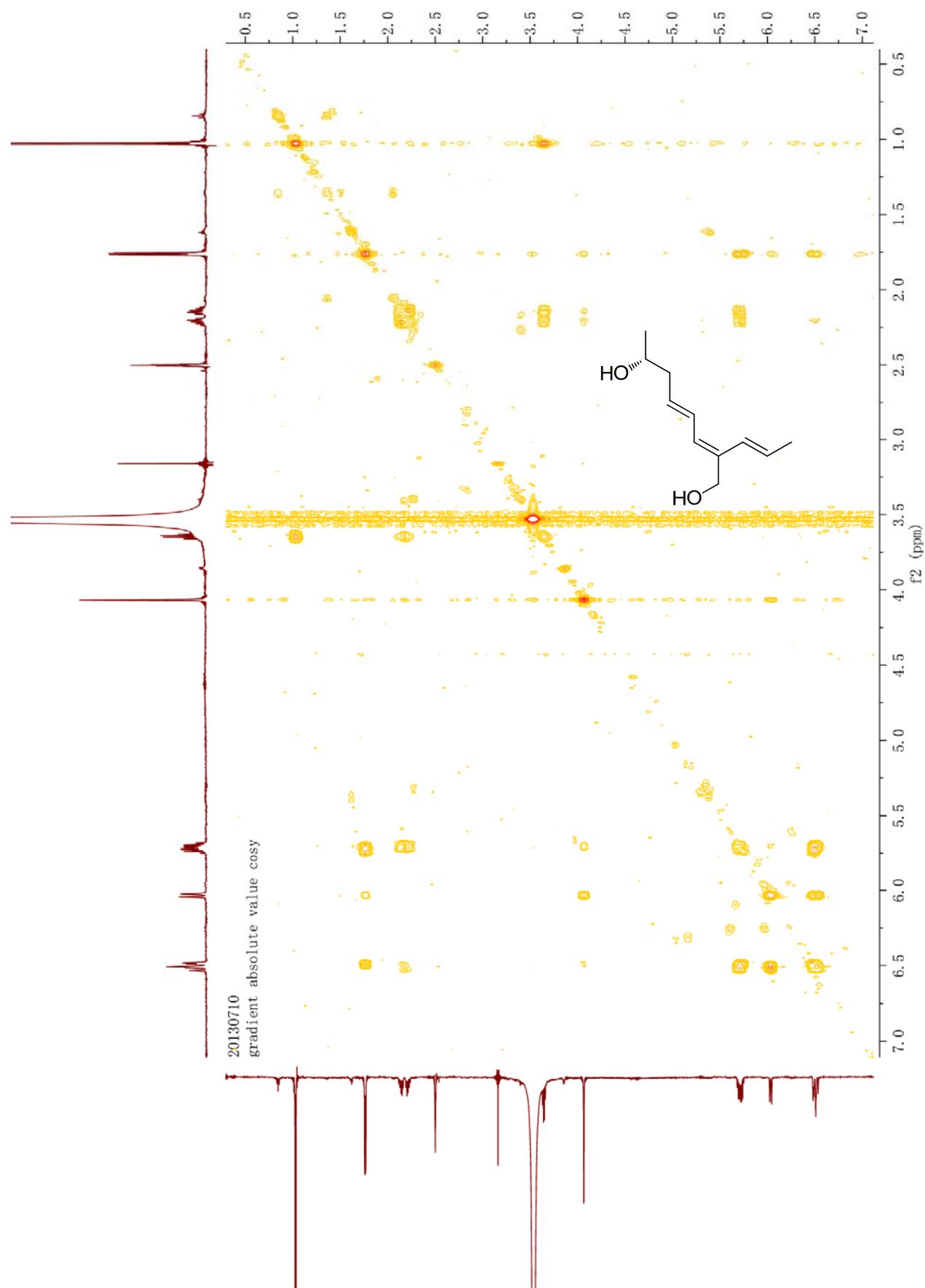


Figure S67. The HMBC spectrum of compound **12** in DMSO-*d*₆

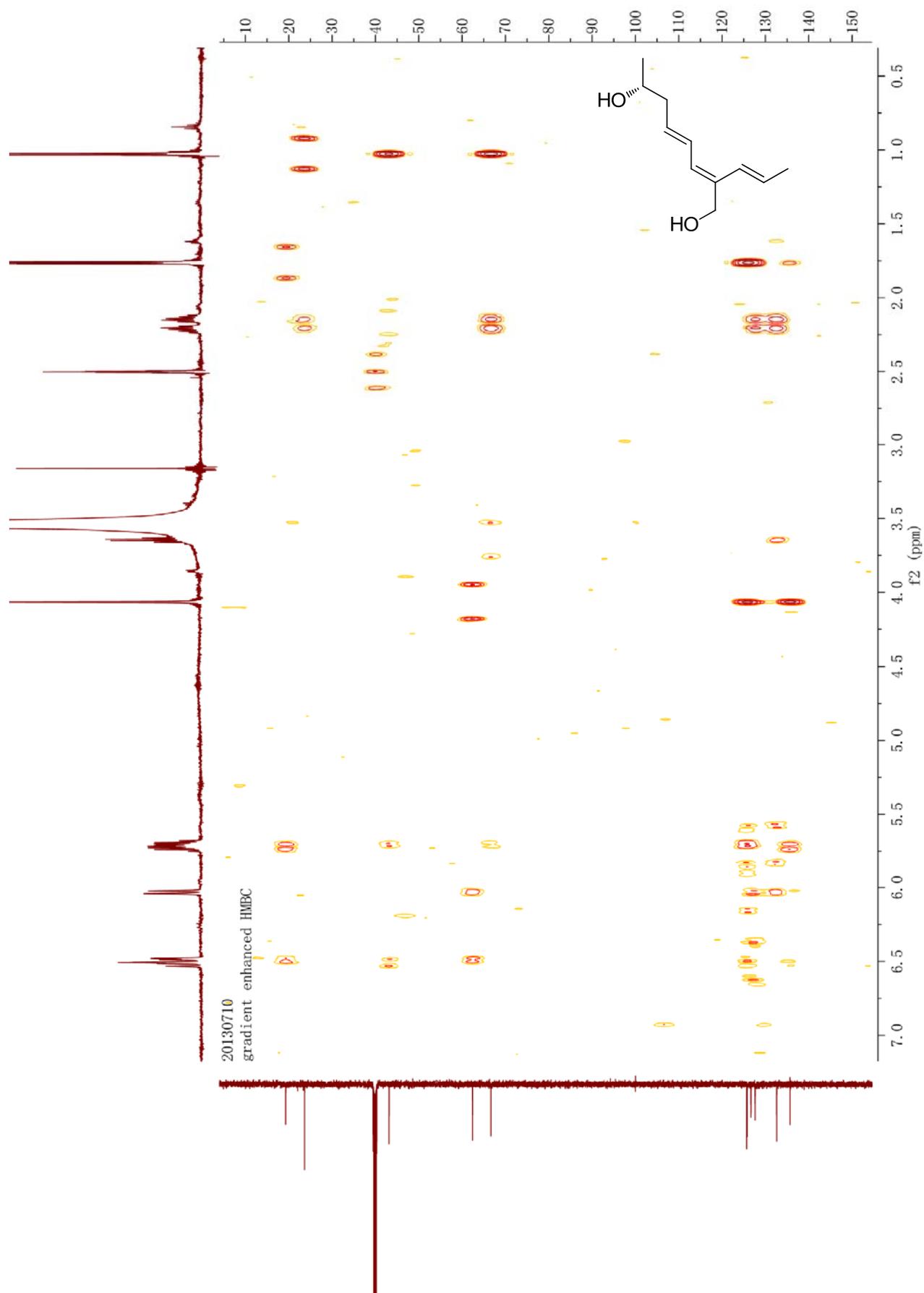


Figure S68. The NOESY spectrum of compound **12** in DMSO-*d*₆

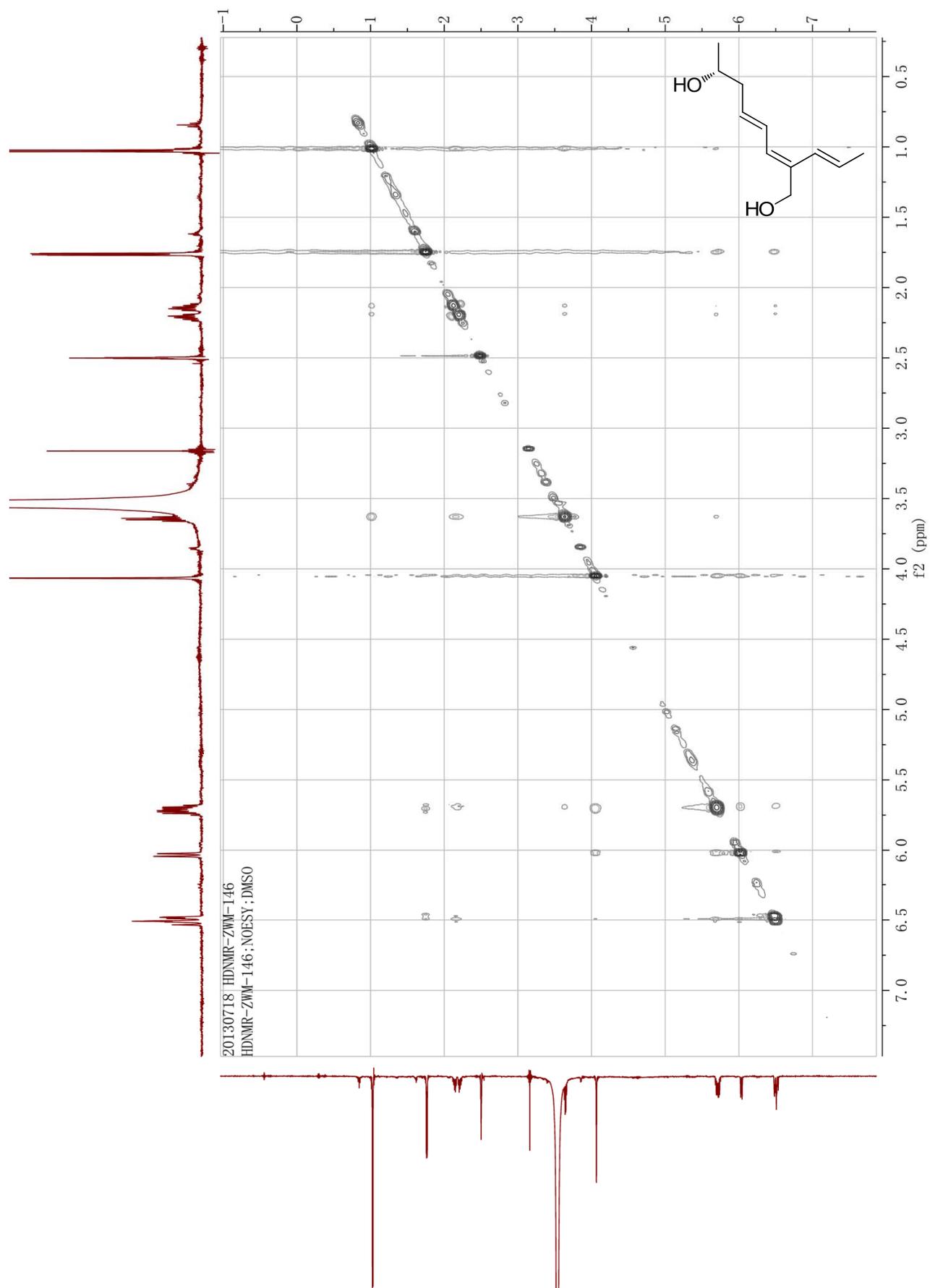


Figure S69. The $^1\text{H-NMR}$ spectrum of compound **13** in $\text{DMSO-}d_6$

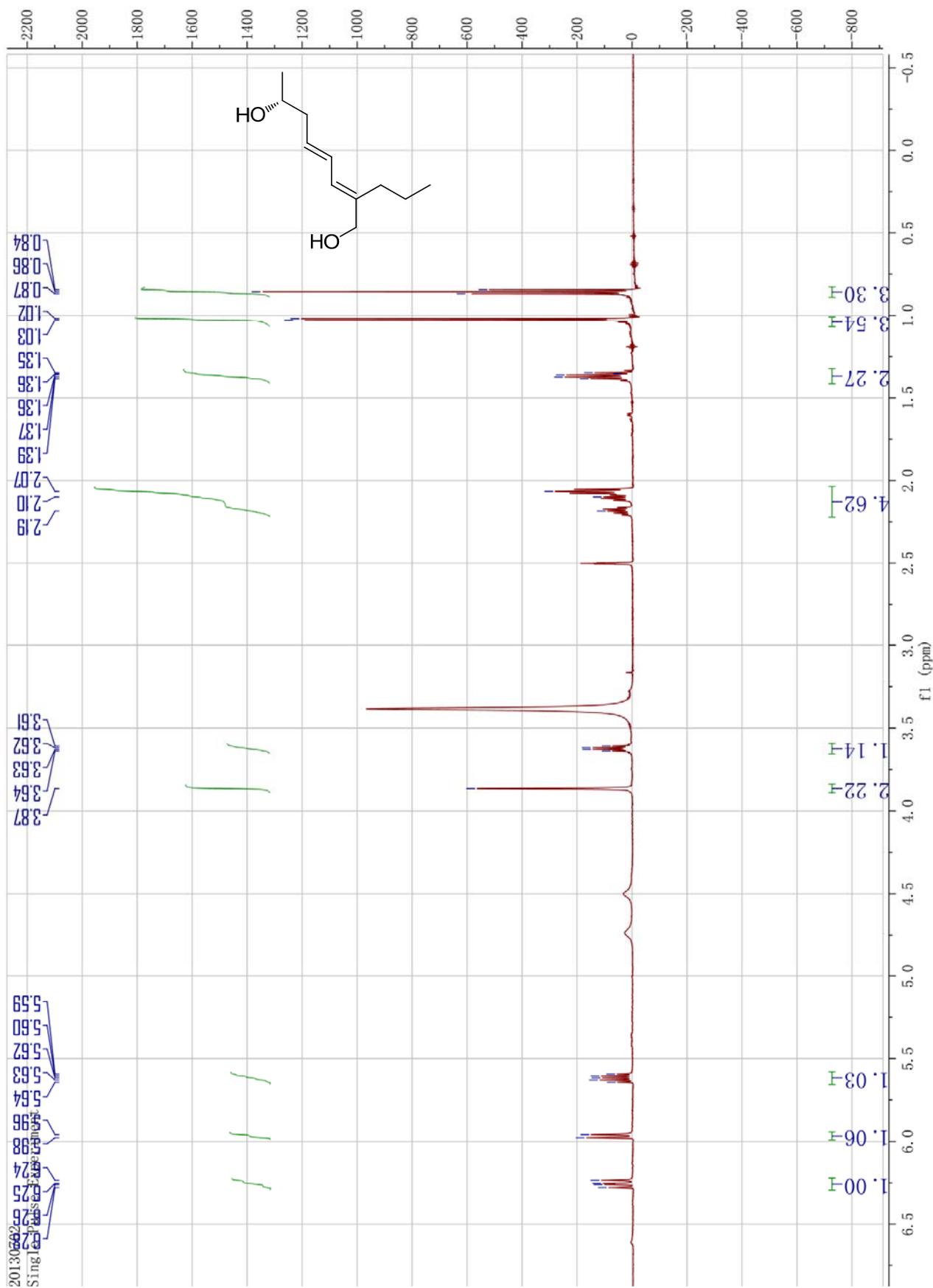


Figure S70. The ^{13}C -NMR spectrum of compound **13** in $\text{DMSO-}d_6$

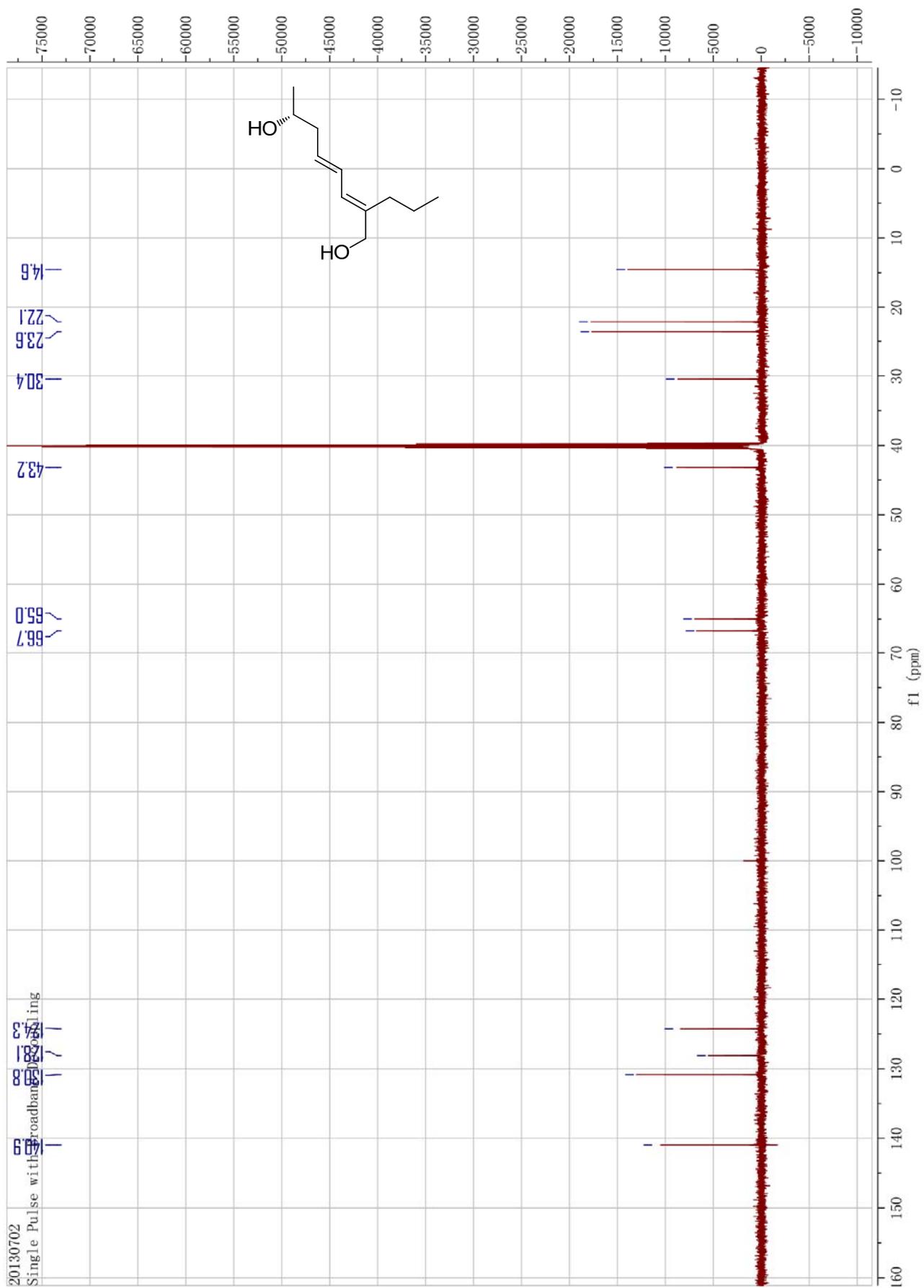


Figure S71. The DEPT spectrum of compound **13** in DMSO-*d*₆

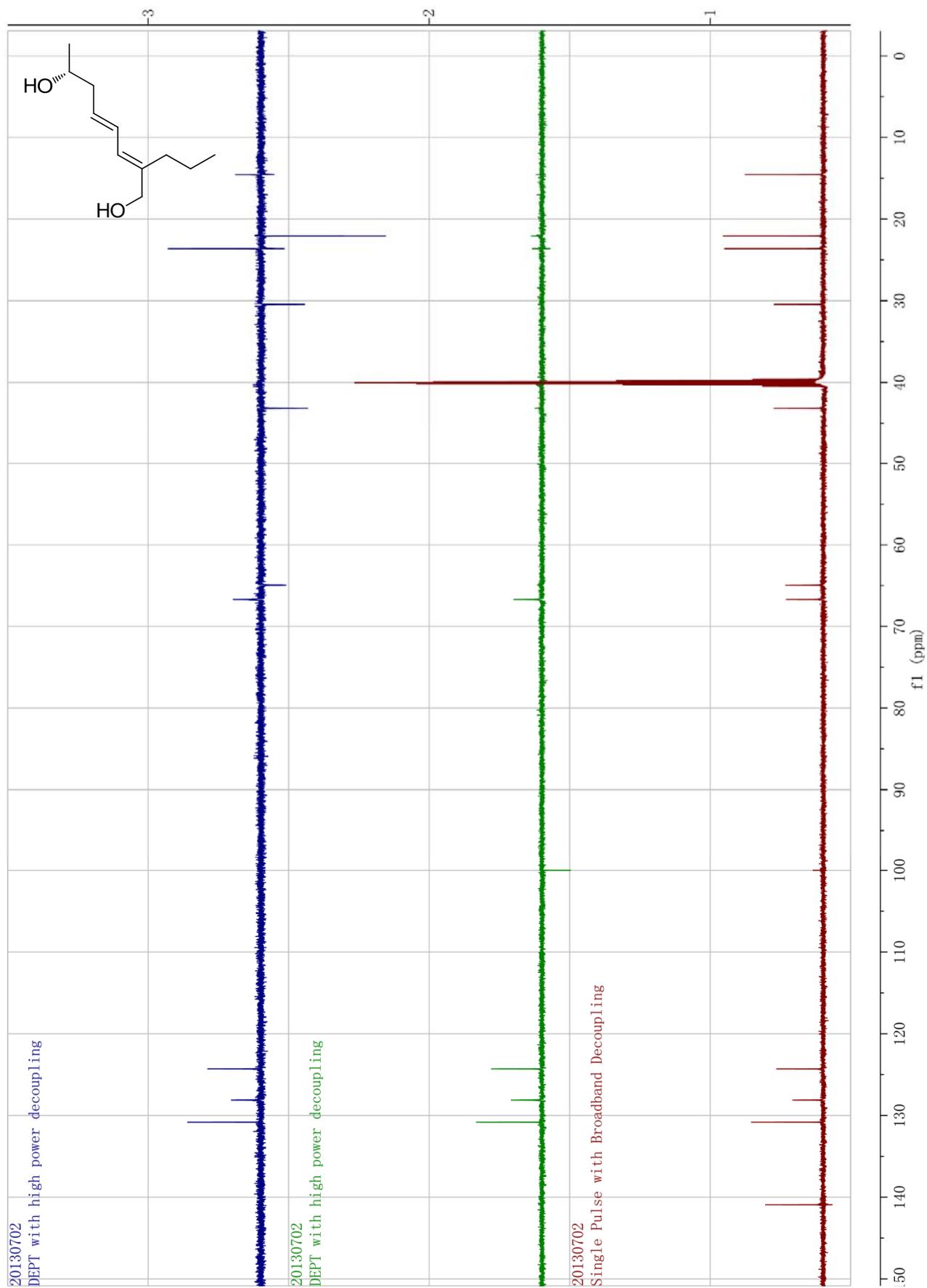


Figure S73. The ^1H - ^1H COSY spectrum of compound **13** in $\text{DMSO-}d_6$

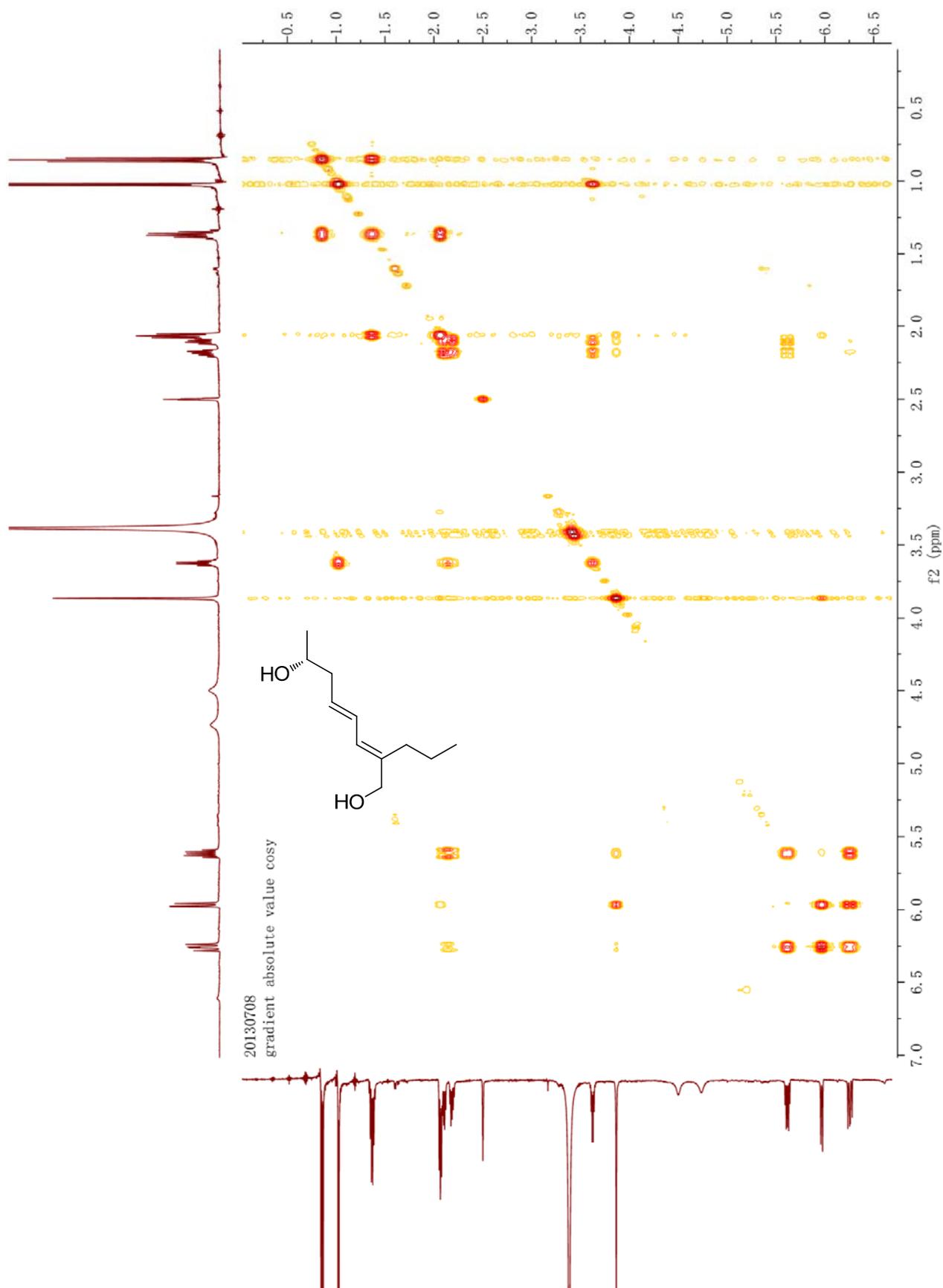


Figure S74. The HMBC spectrum of compound **13** in DMSO-*d*₆

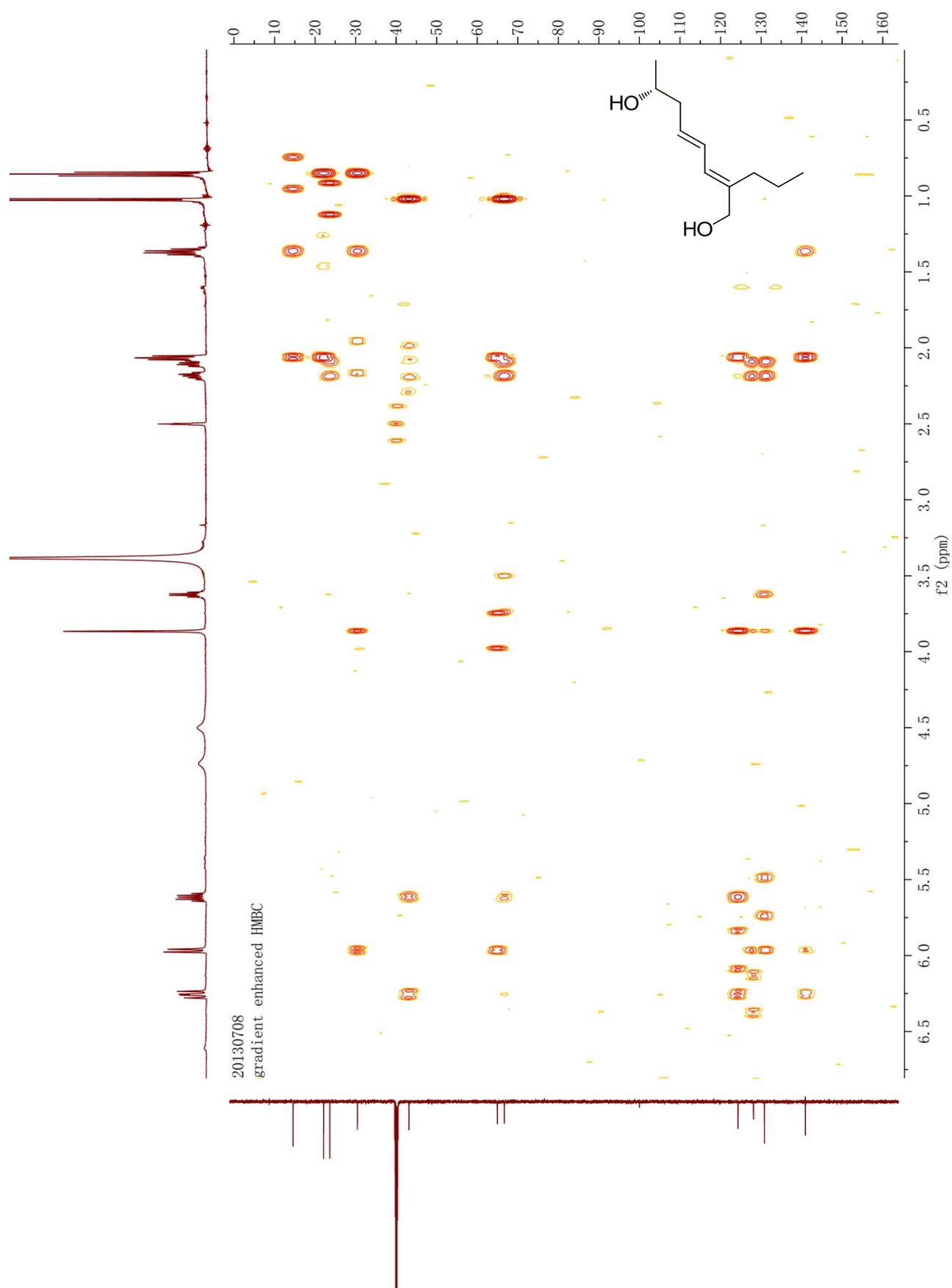


Figure S75. The NOESY spectrum of compound **13** in DMSO- d_6

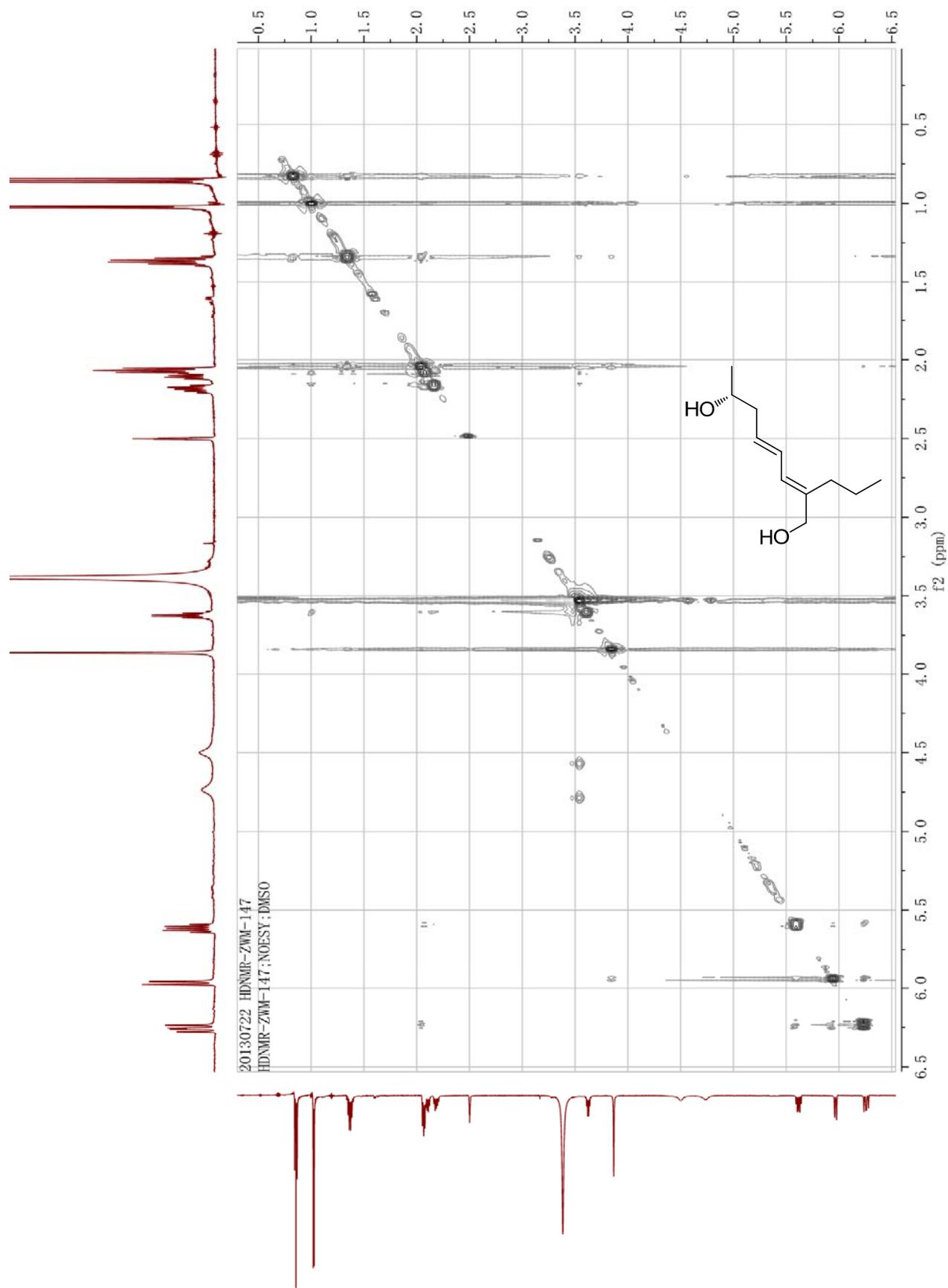


Figure S76. The $^1\text{H-NMR}$ spectrum of compound **14** in $\text{DMSO-}d_6$

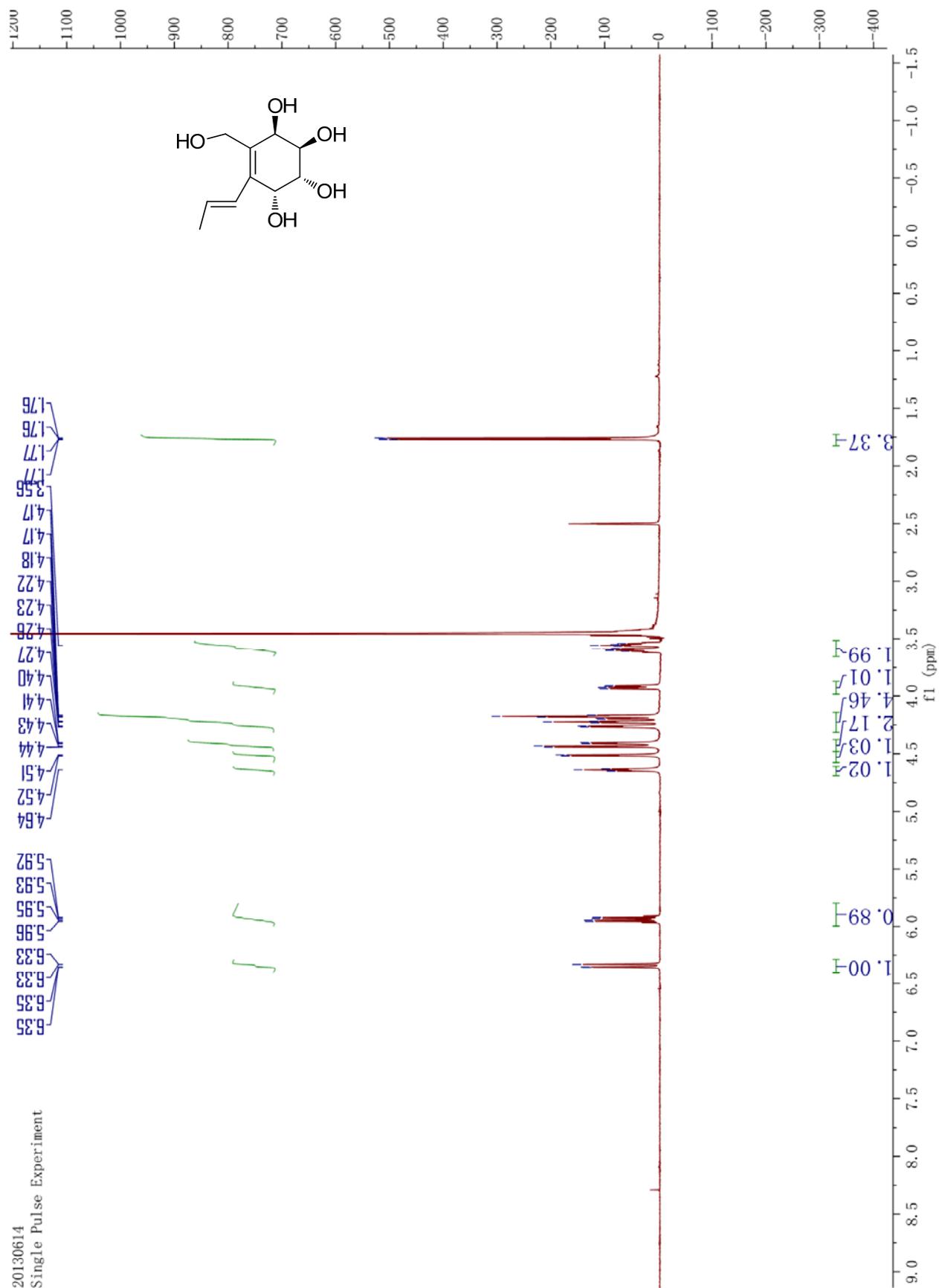


Figure S78. The DEPT spectrum of compound **14** in DMSO-*d*₆

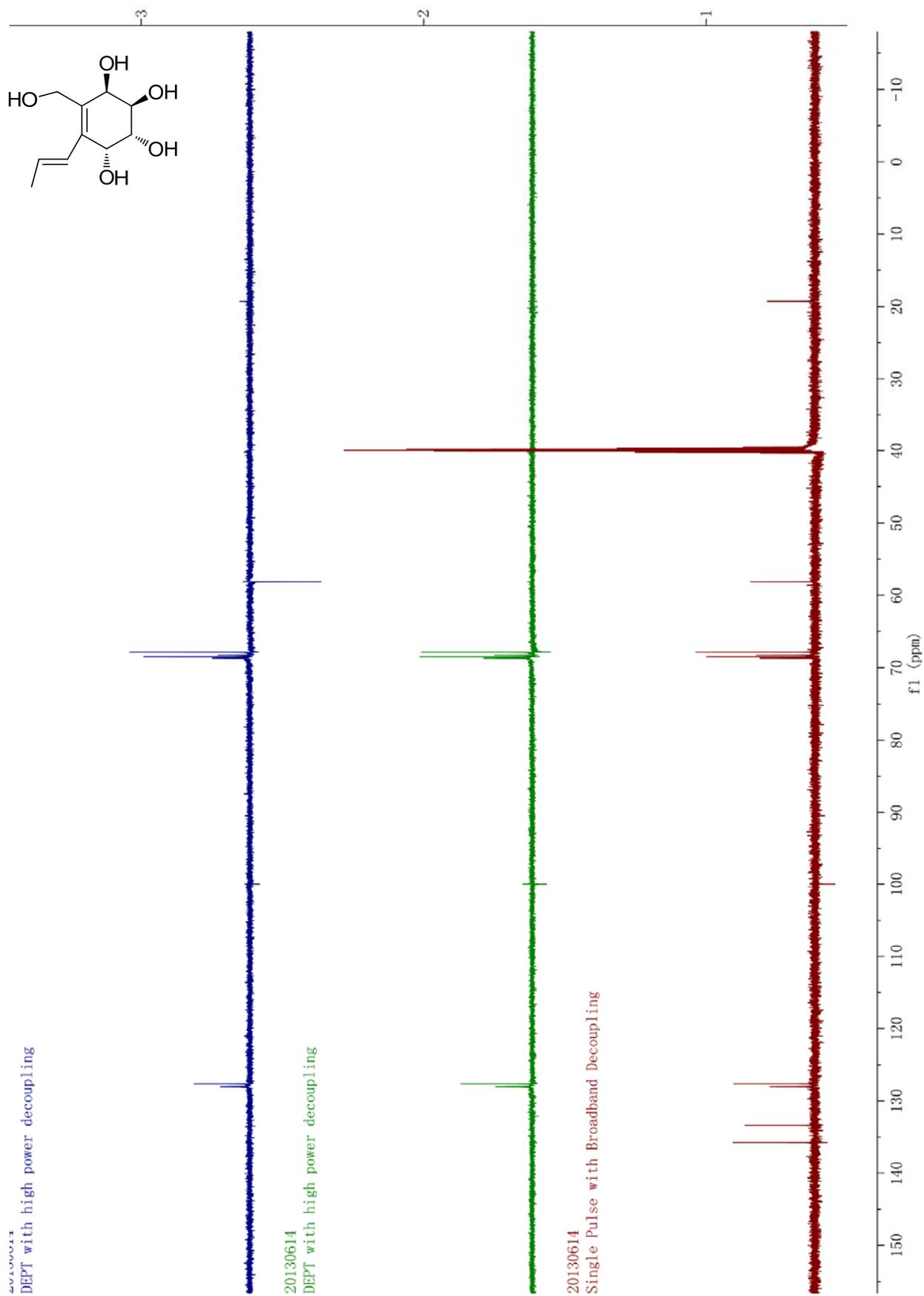


Figure S79. The HMQC spectrum of compound **14** in DMSO-*d*₆

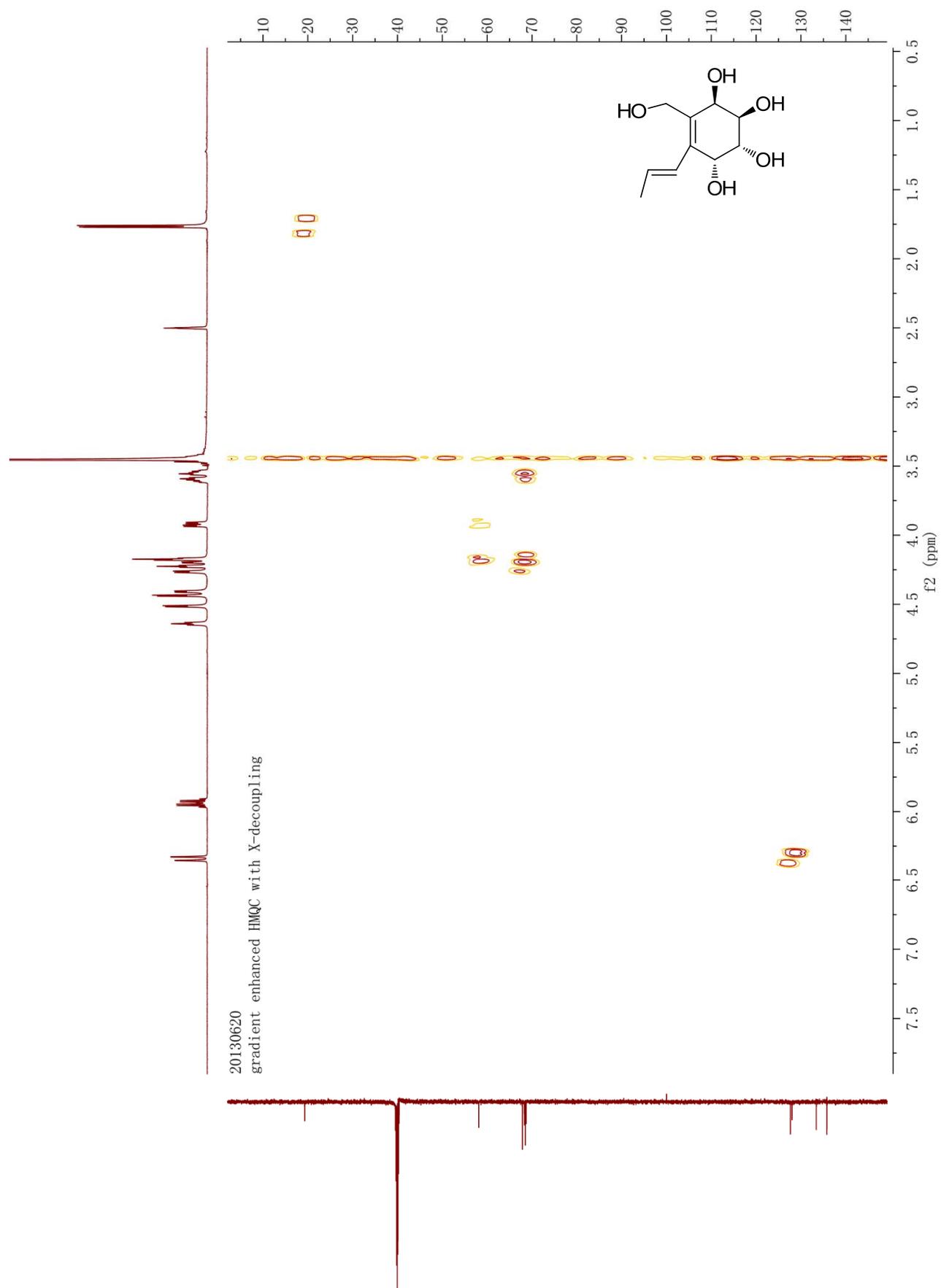


Figure S80. The ^1H - ^1H COSY spectrum of compound 14 in $\text{DMSO-}d_6$

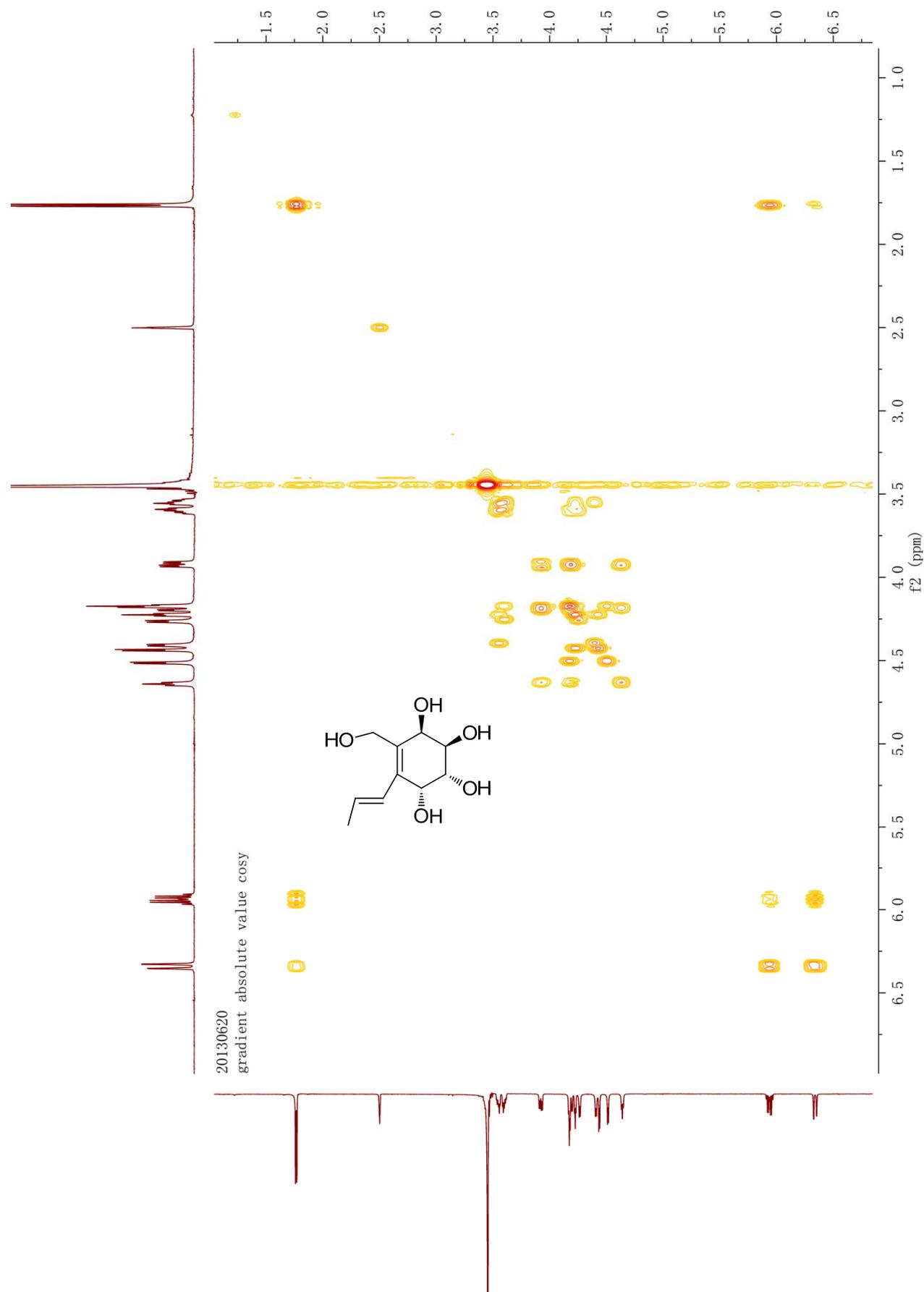


Figure S81. The HMBC spectrum of compound **14** in DMSO-*d*₆

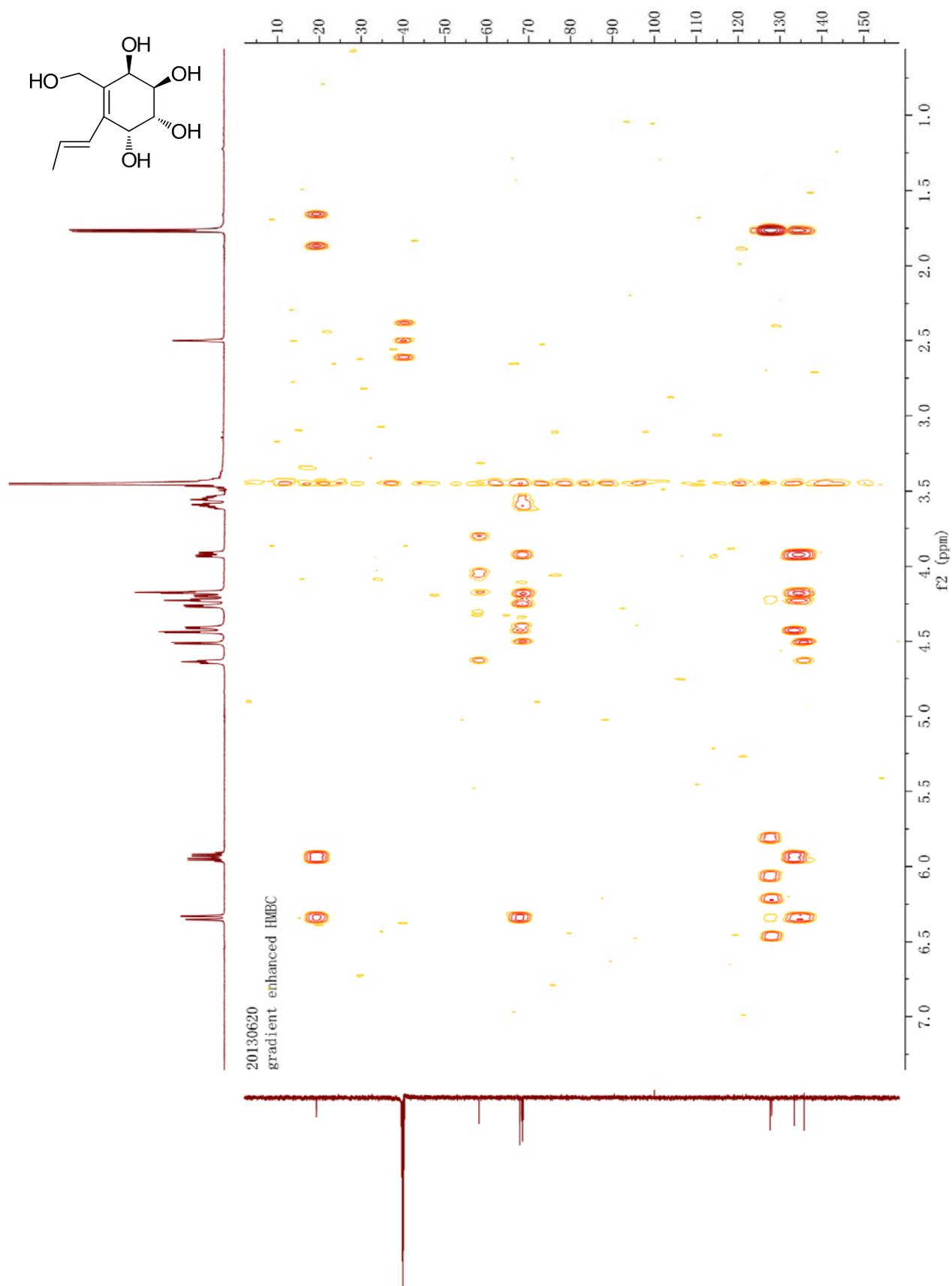


Figure S82. The ^1H -NMR spectrum of compound **15** in $\text{DMSO-}d_6$

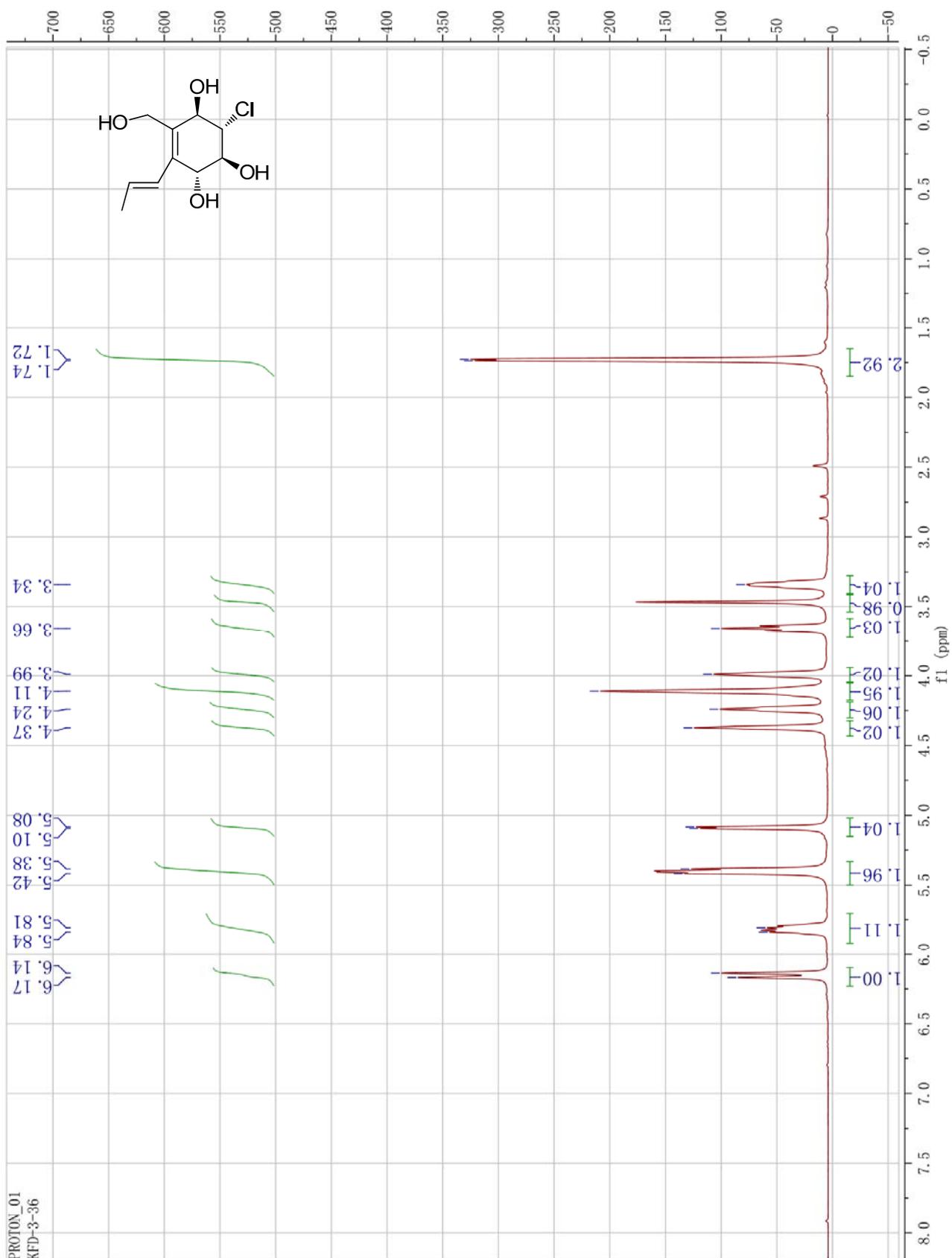


Figure S83. The ^{13}C -NMR spectrum of compound **15** in $\text{DMSO-}d_6$

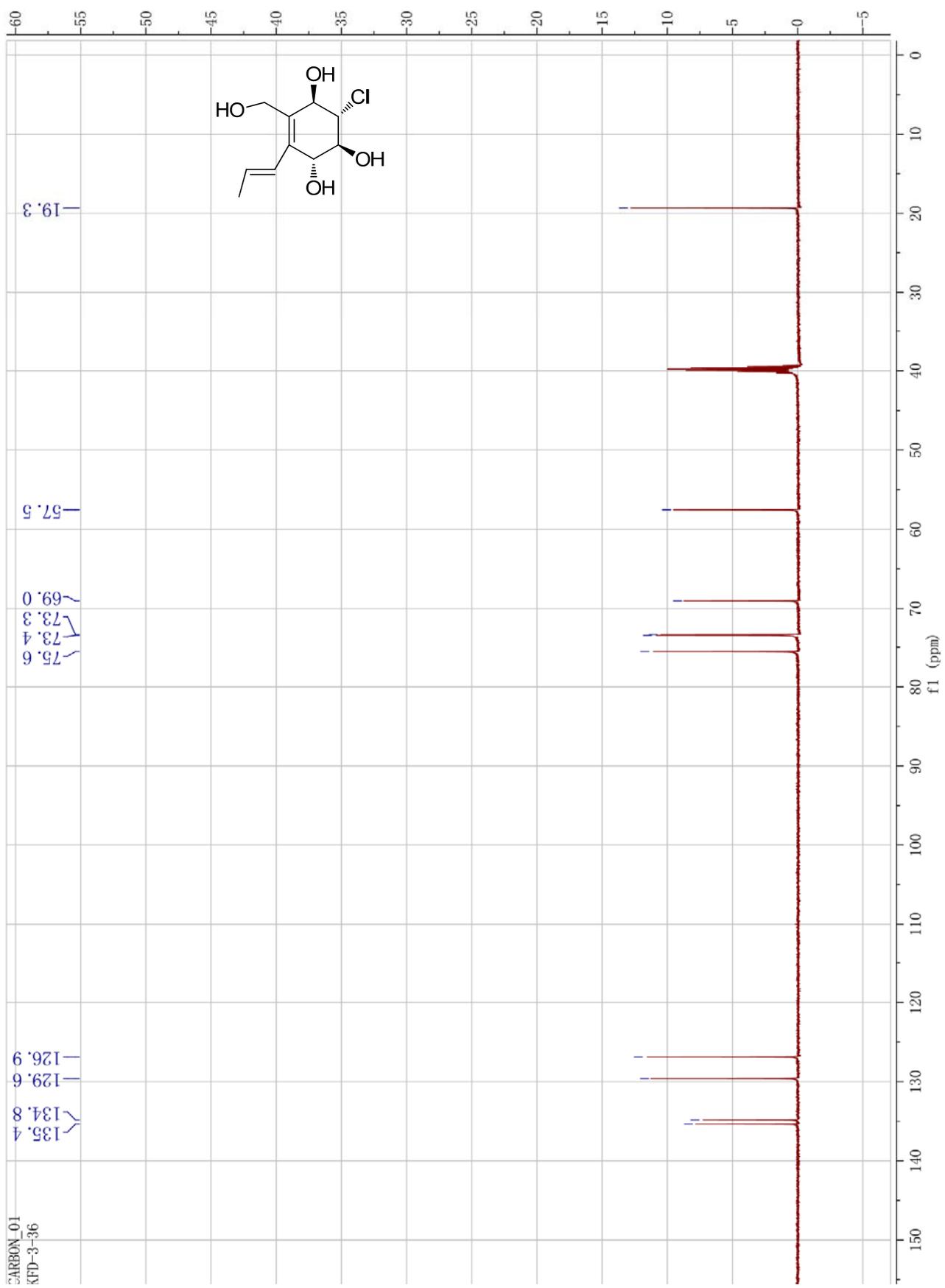


Figure S84. The DEPT spectrum of compound **15** in DMSO-*d*₆

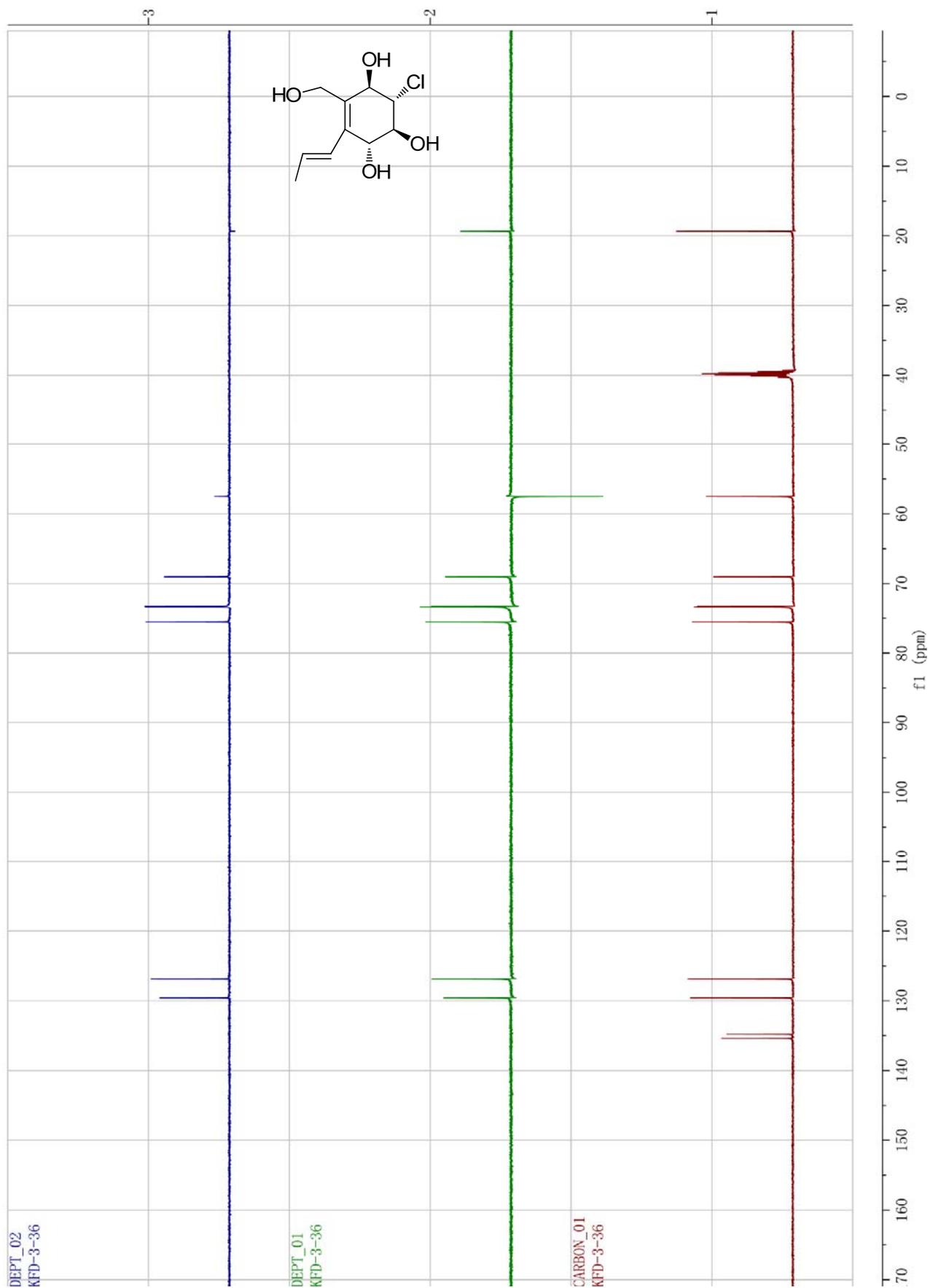


Figure S85. The HMQC spectrum of compound **15** in DMSO-*d*₆

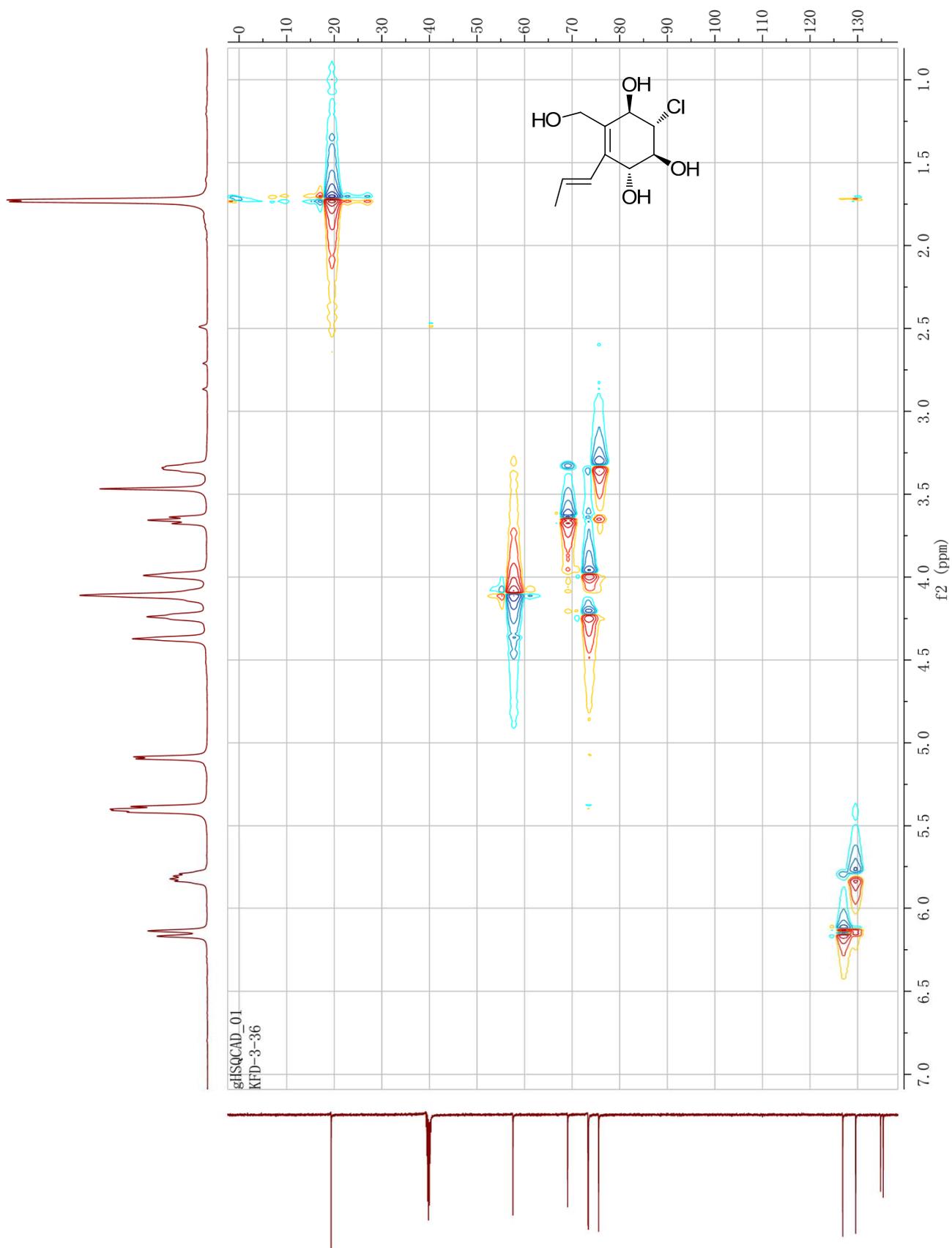


Figure S86. The ^1H - ^1H COSY spectrum of compound **15** in $\text{DMSO-}d_6$

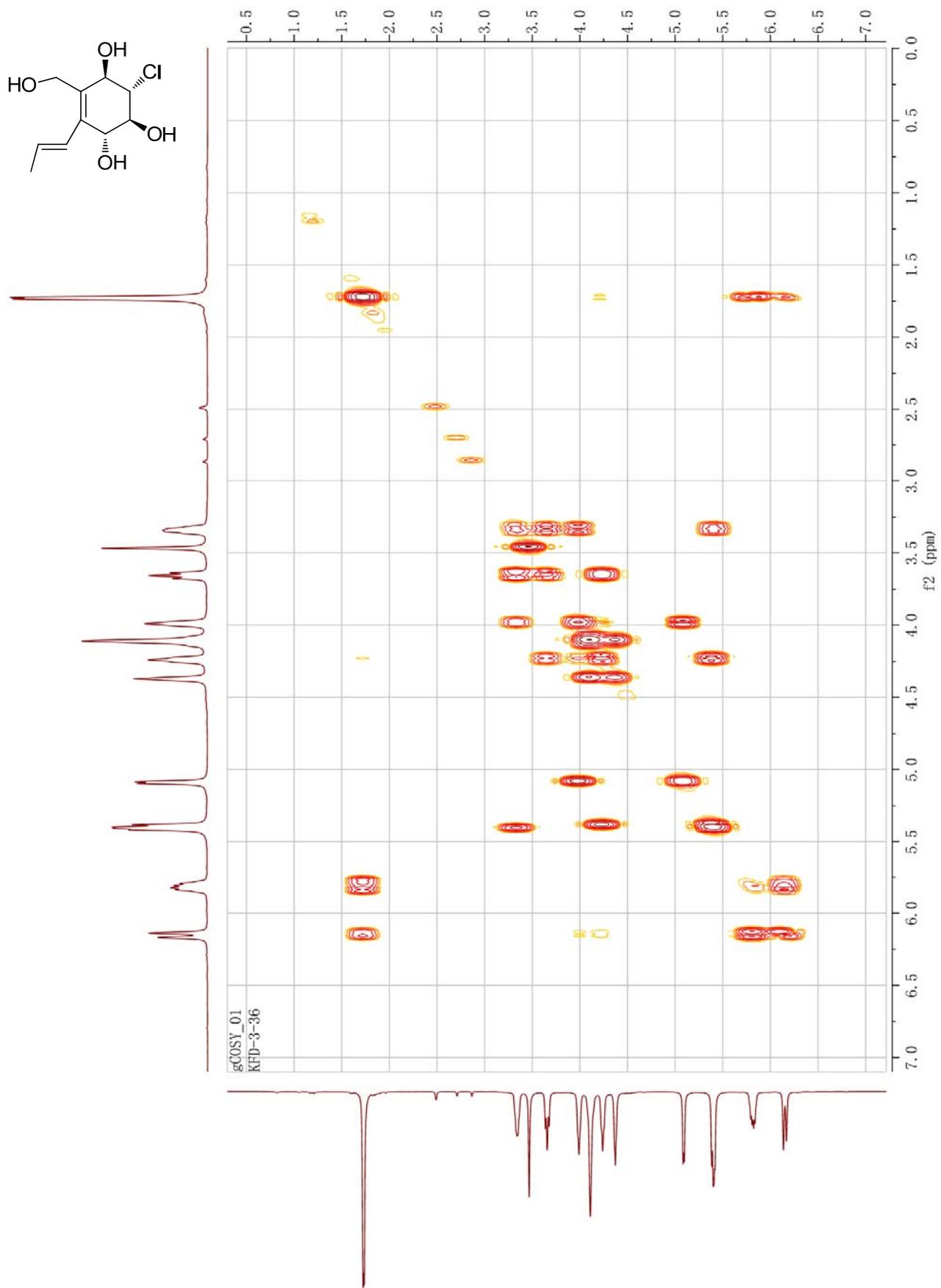


Figure S87. The HMBC spectrum of compound **15** in DMSO-*d*₆

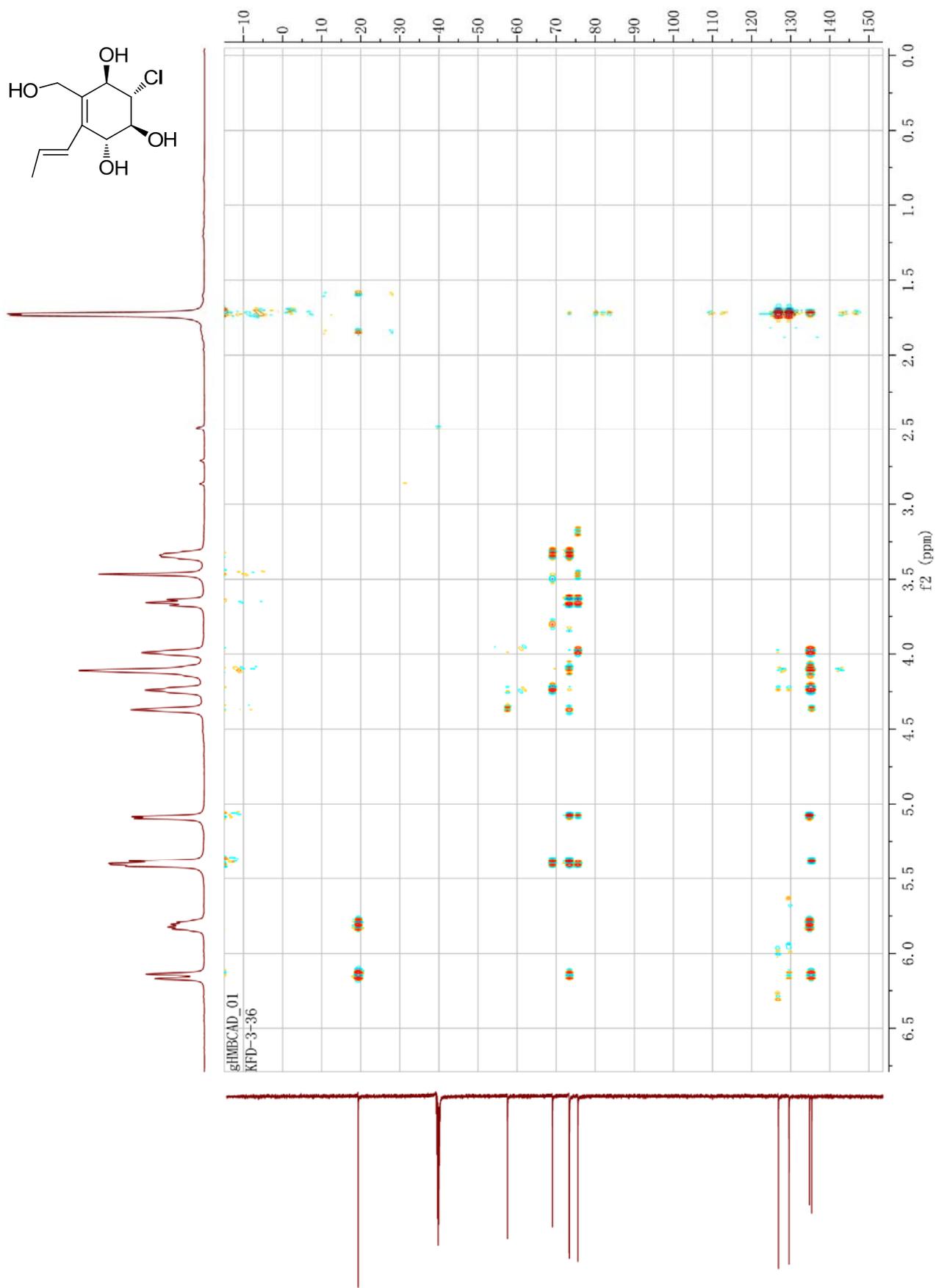


Figure S88. The ^1H -NMR spectrum of compound **16** in $\text{DMSO-}d_6$

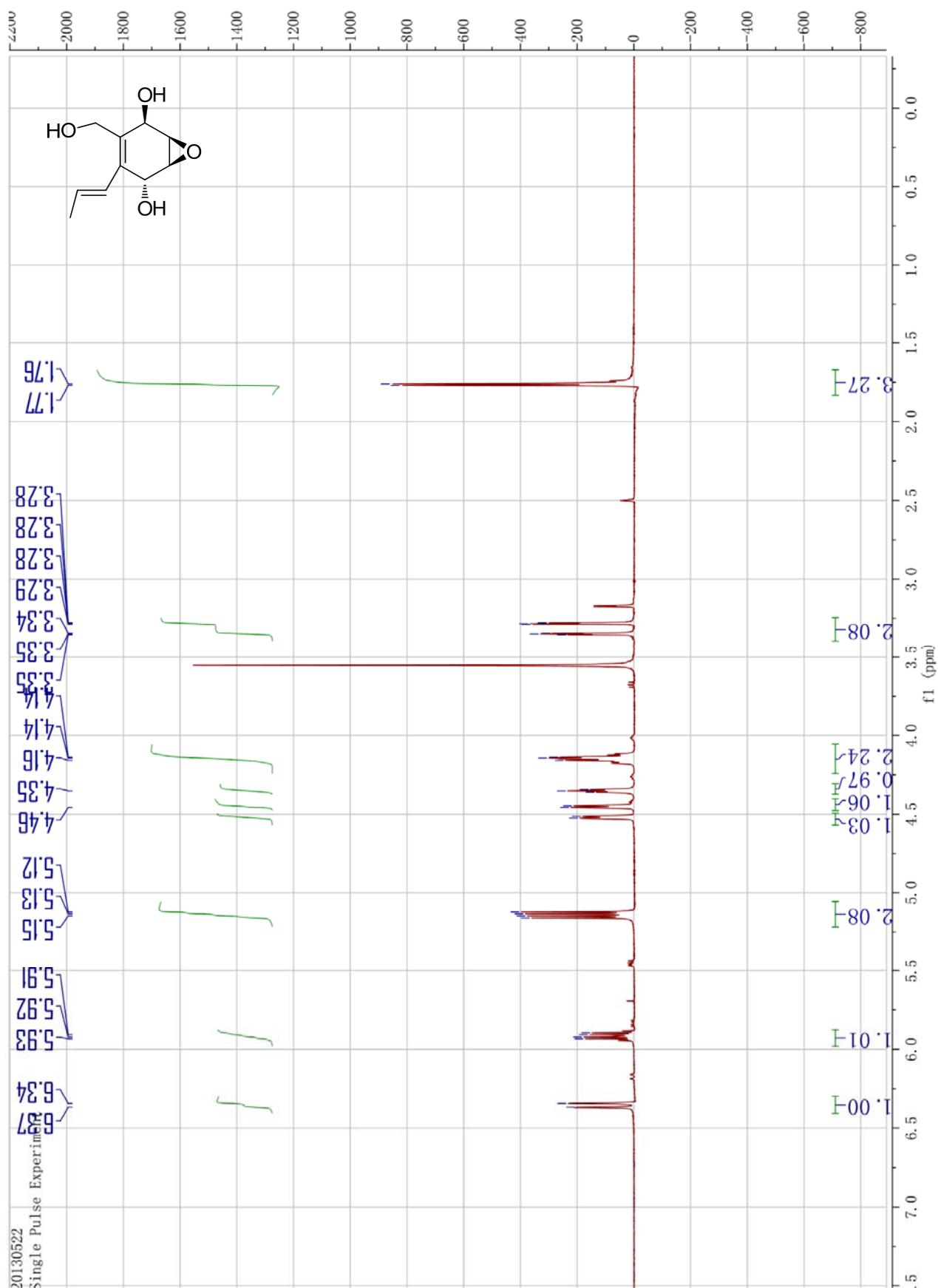


Figure S89. The ^{13}C -NMR spectrum of compound **16** in $\text{DMSO-}d_6$

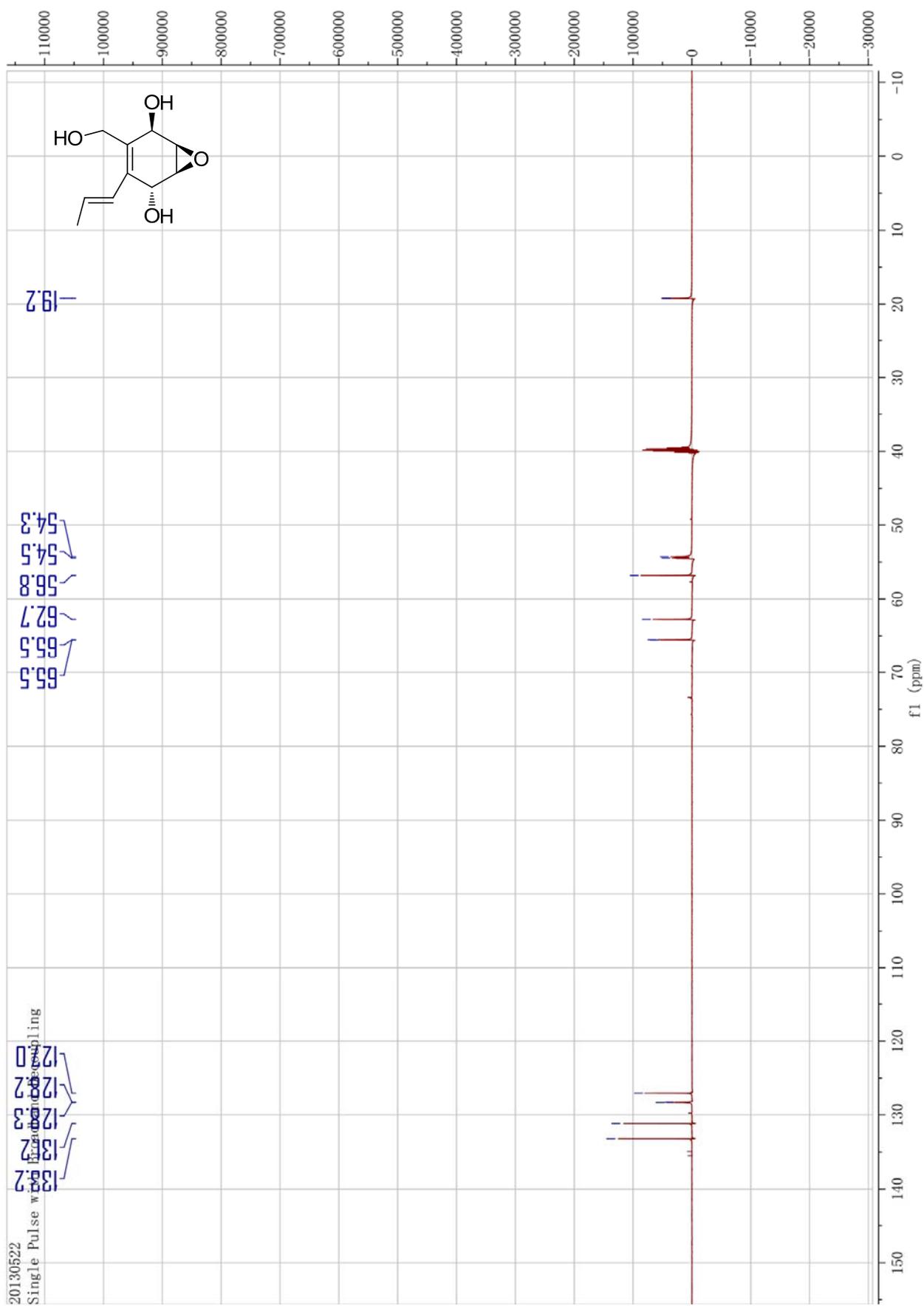


Figure S90. The DEPT spectrum of compound **16** in DMSO-*d*₆

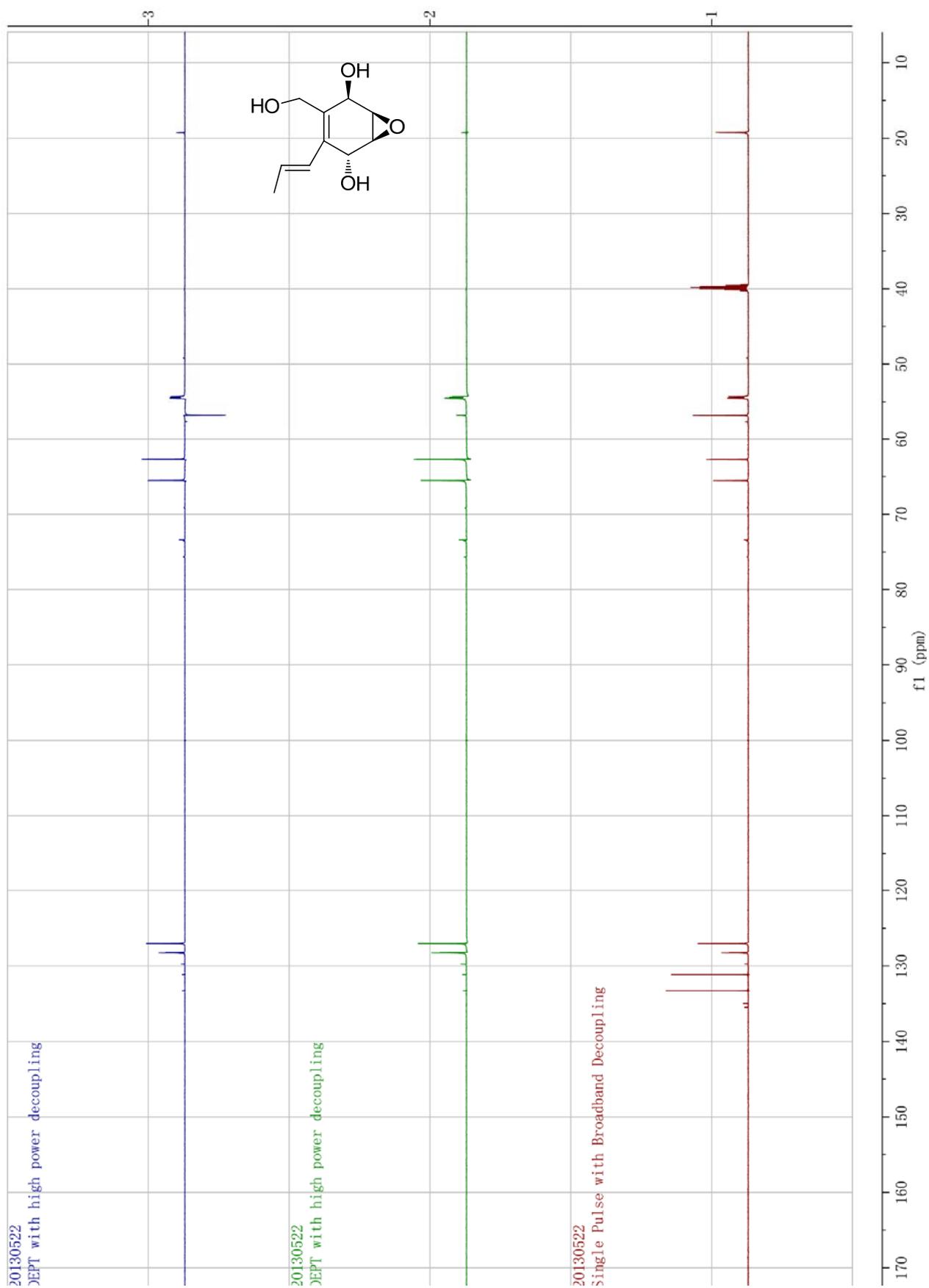


Figure S91. The HMQC spectrum of compound **16** in DMSO-*d*₆

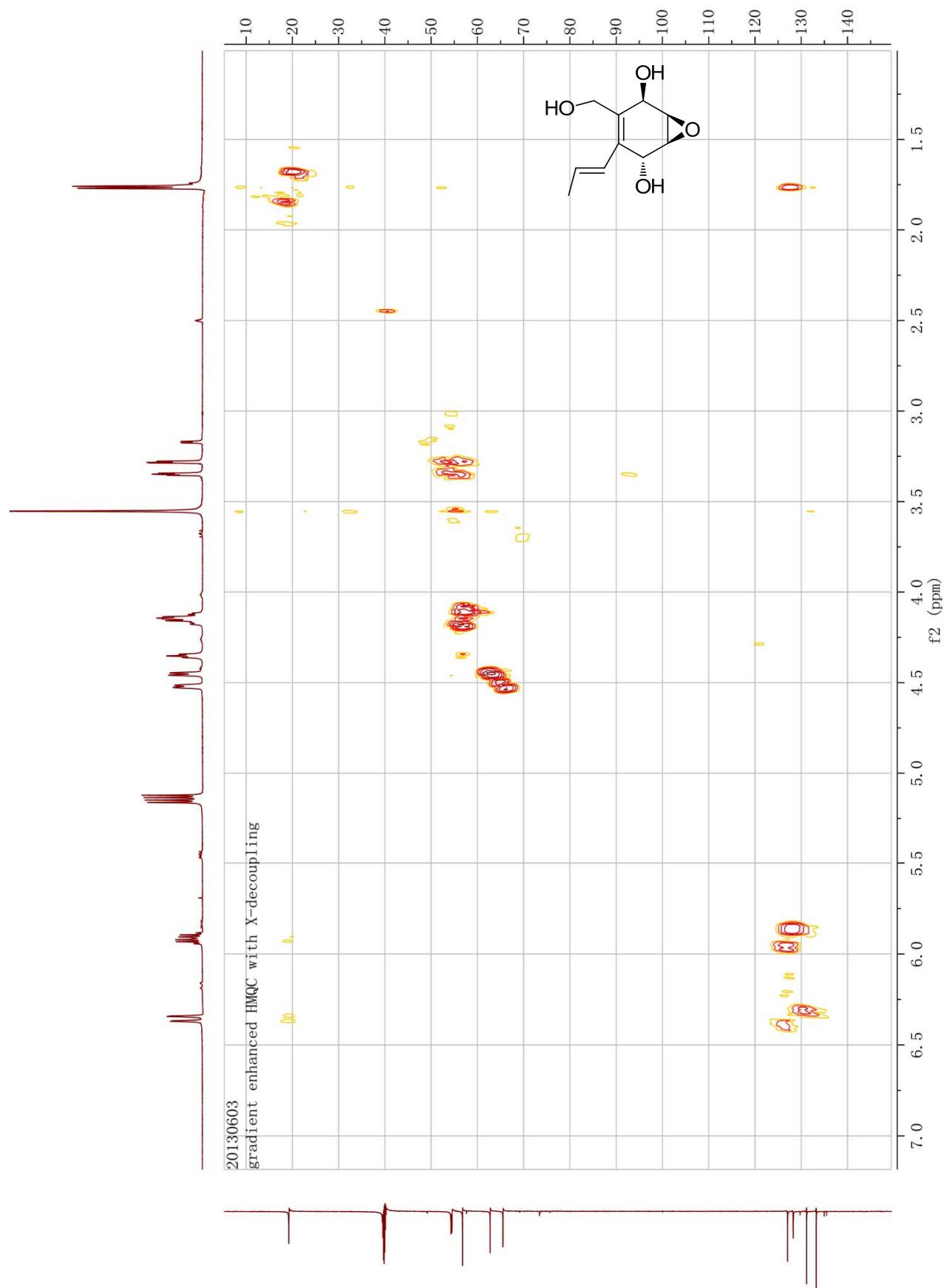


Figure S92. The ^1H - ^1H COSY spectrum of compound **16** in $\text{DMSO-}d_6$

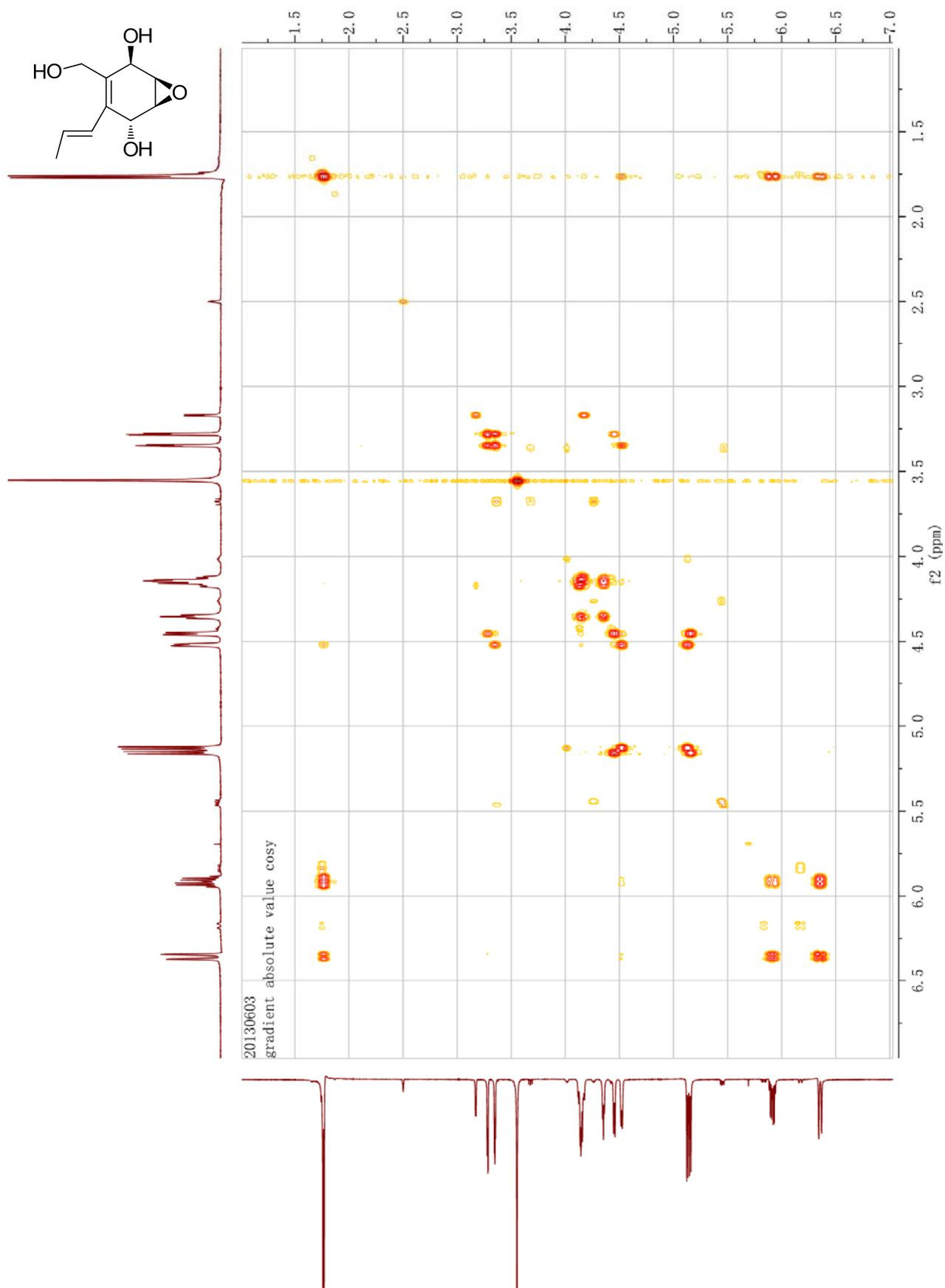


Figure S94. The ^1H -NMR spectrum of compound **17** in $\text{DMSO-}d_6$

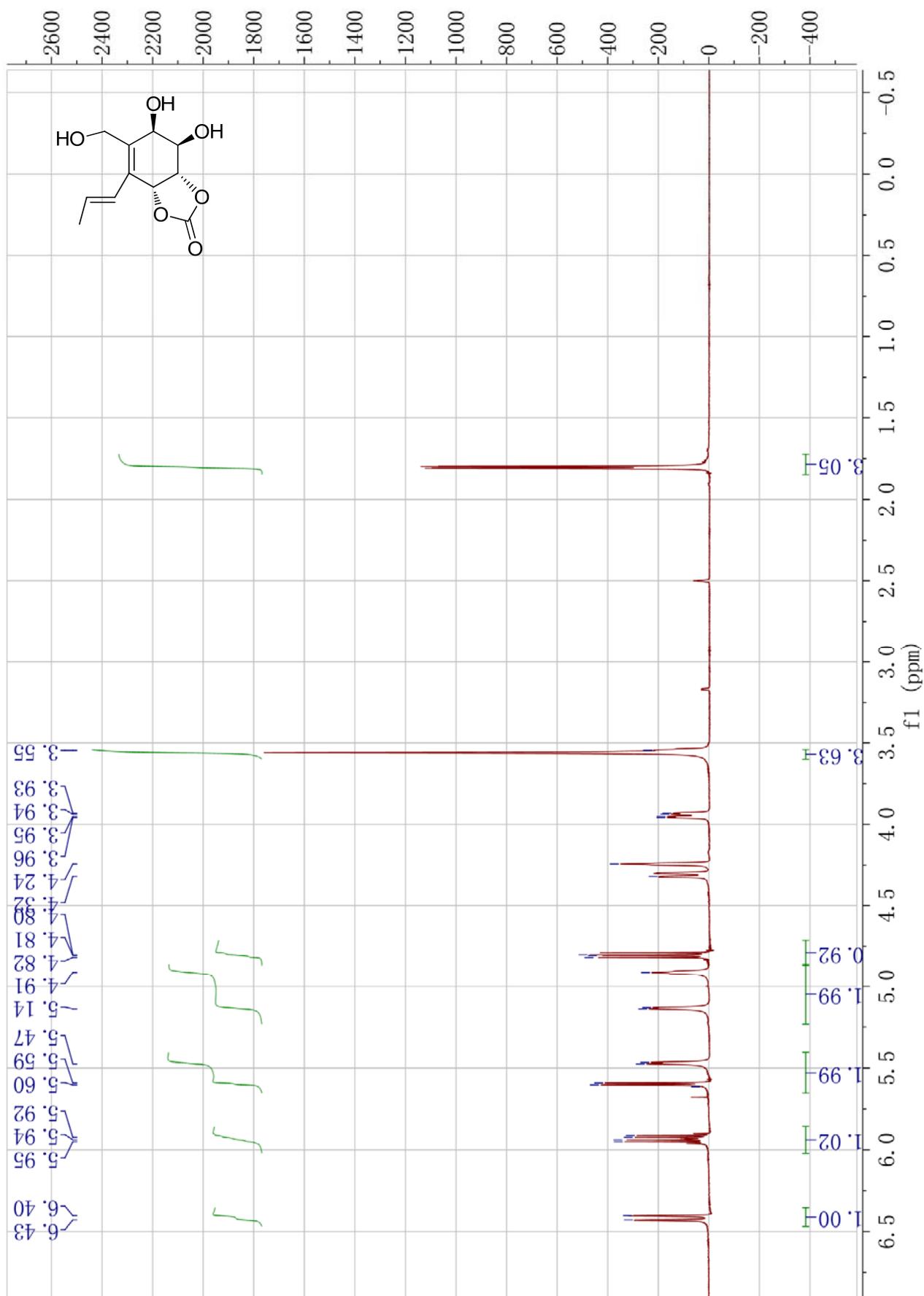


Figure S95. The ^{13}C -NMR spectrum of compound **17** in $\text{DMSO-}d_6$

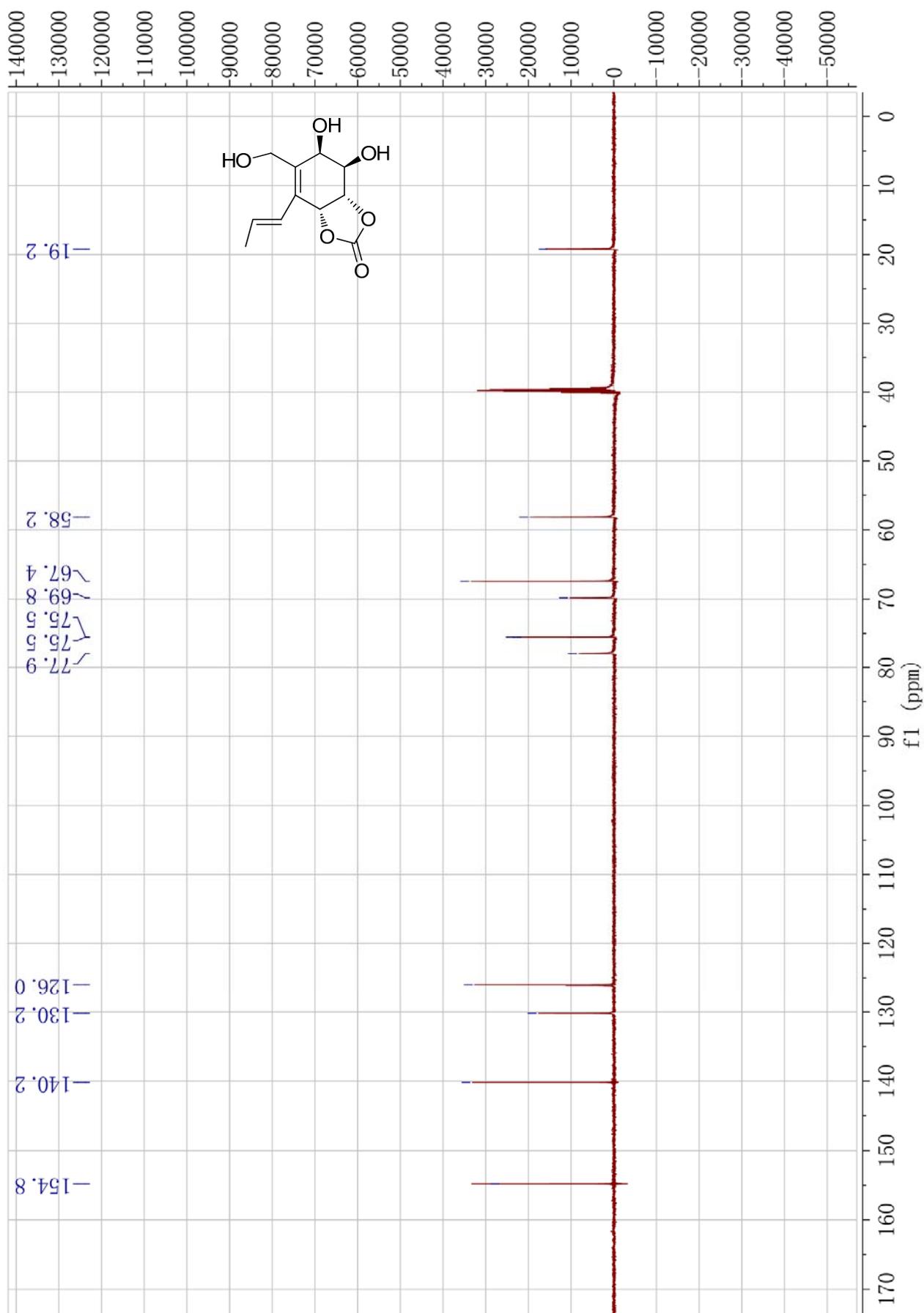


Figure S96. The DEPT spectrum of compound **17** in DMSO-*d*₆

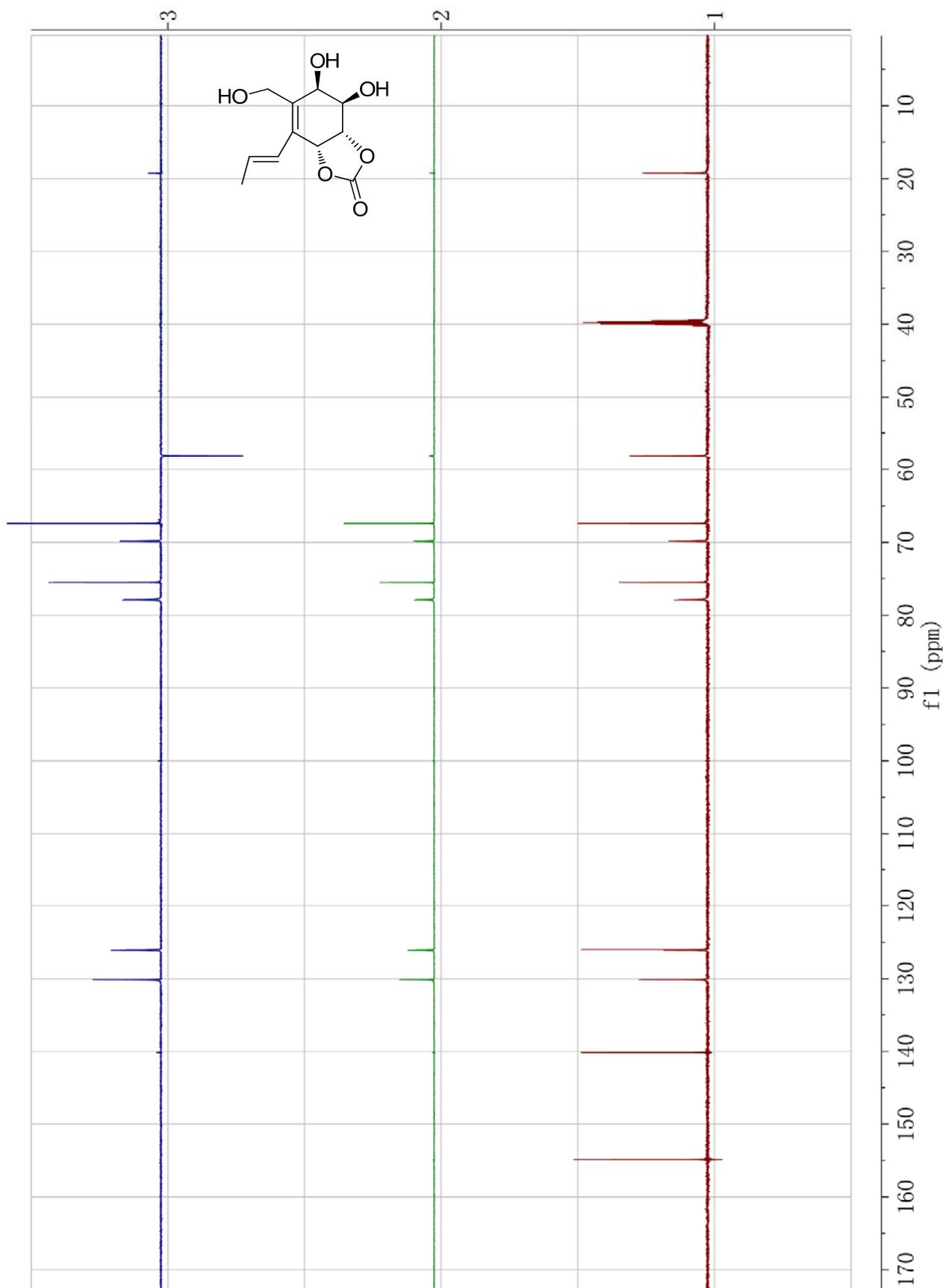


Figure S97. The HMQC spectrum of compound **17** in DMSO-*d*₆

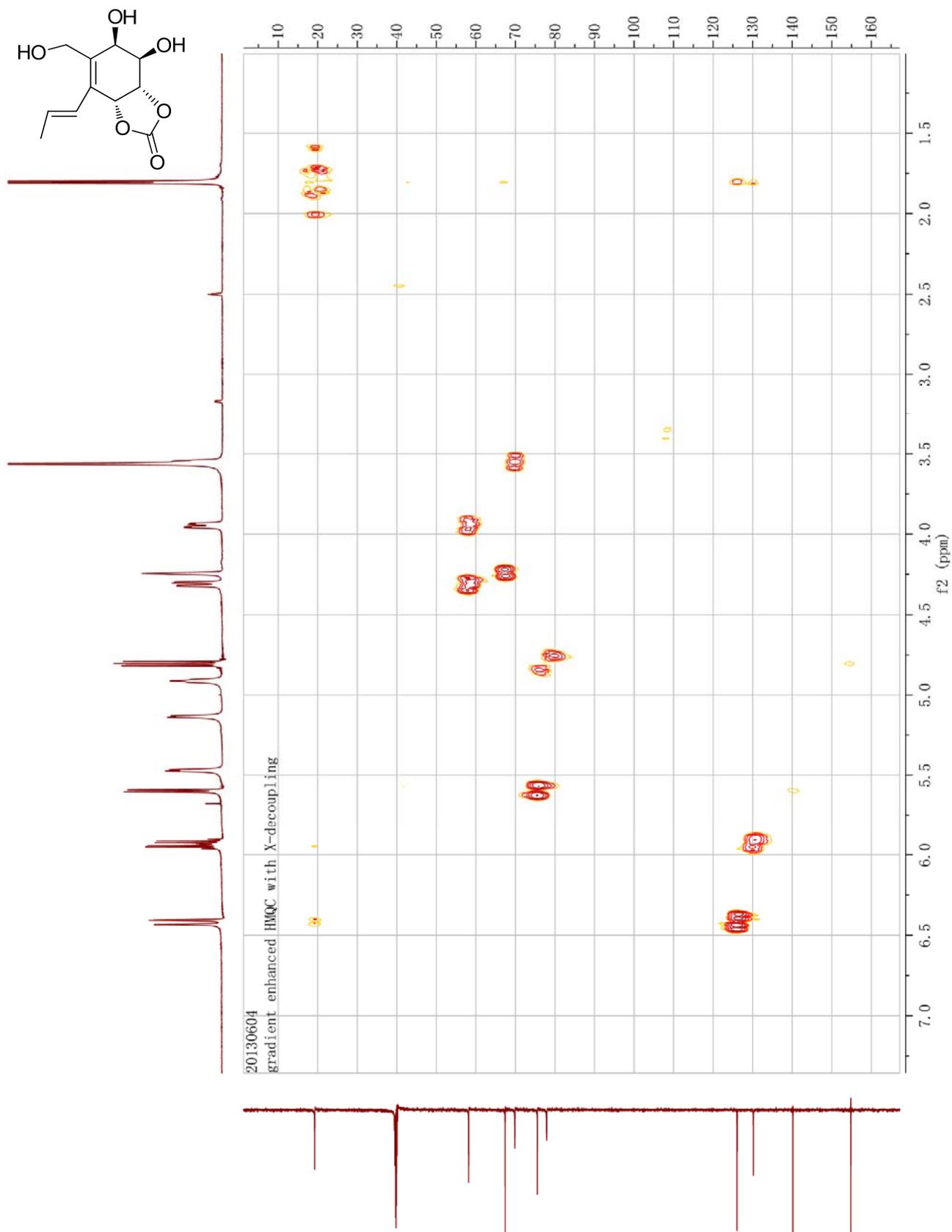


Figure S98. The ^1H - ^1H COSY spectrum of compound 17 in $\text{DMSO-}d_6$

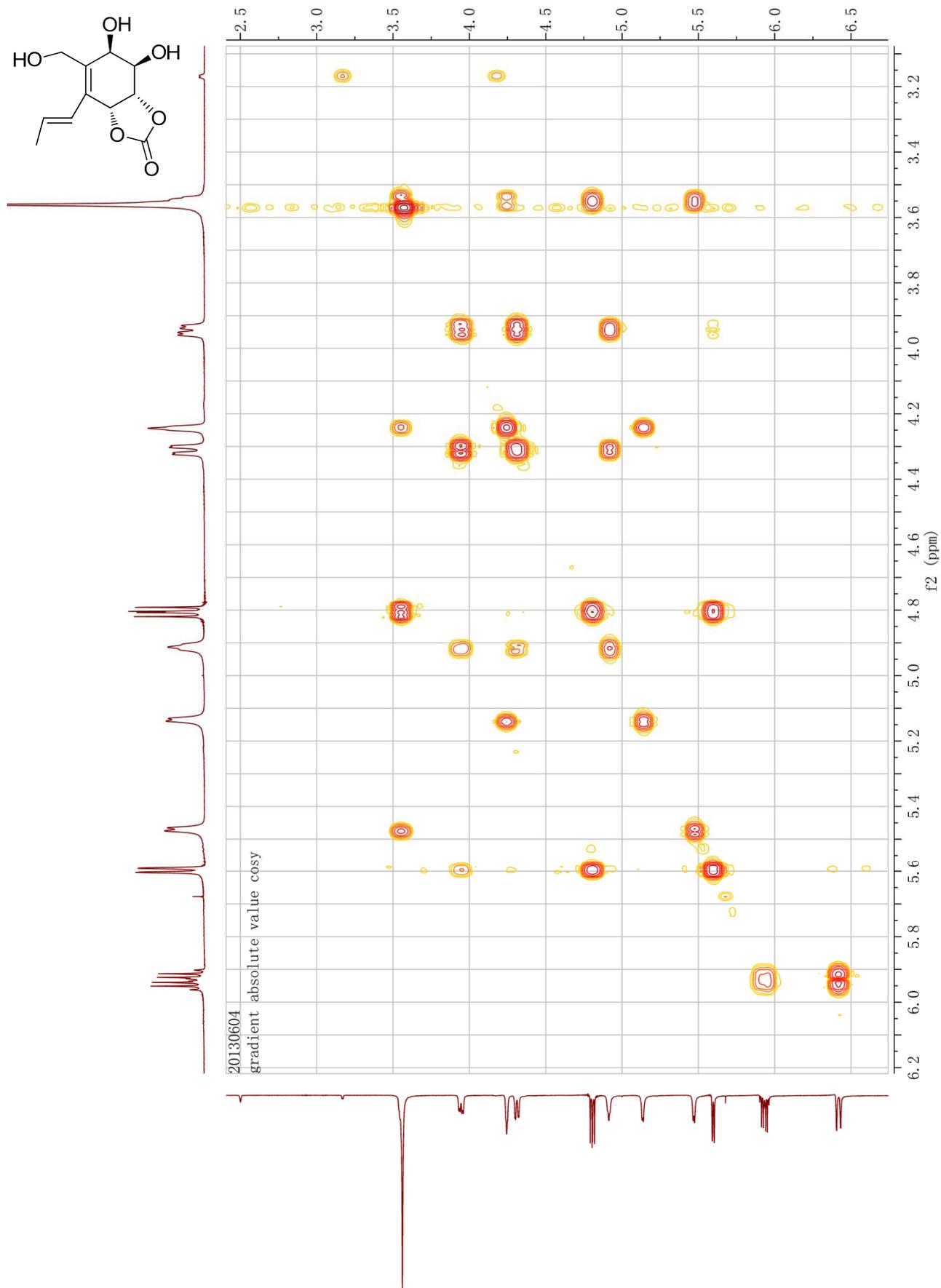


Figure S99. The HMBC spectrum of compound 17 in DMSO-*d*₆

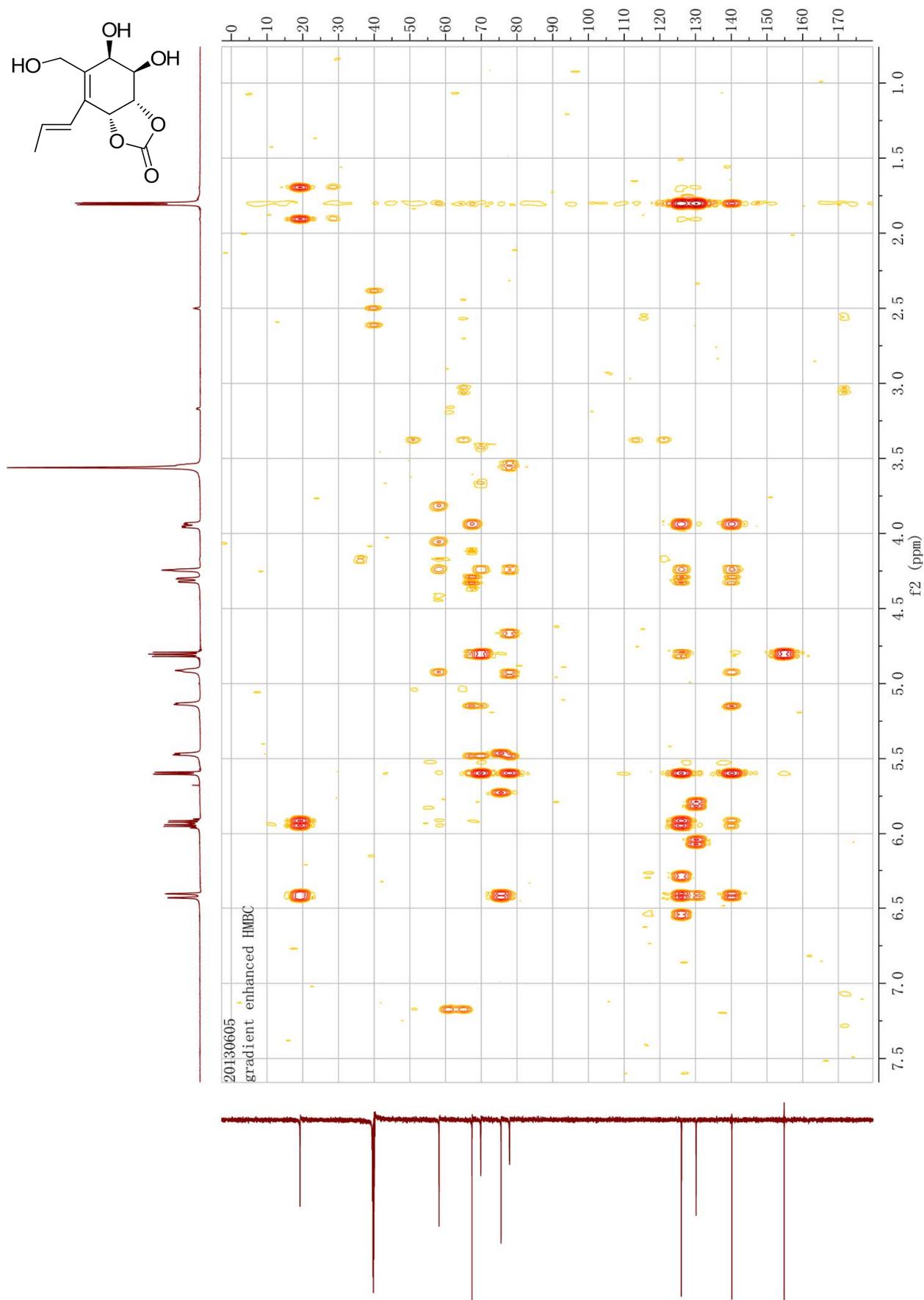


Figure S100. The ^1H -NMR spectrum of compound **18** in CDCl_3

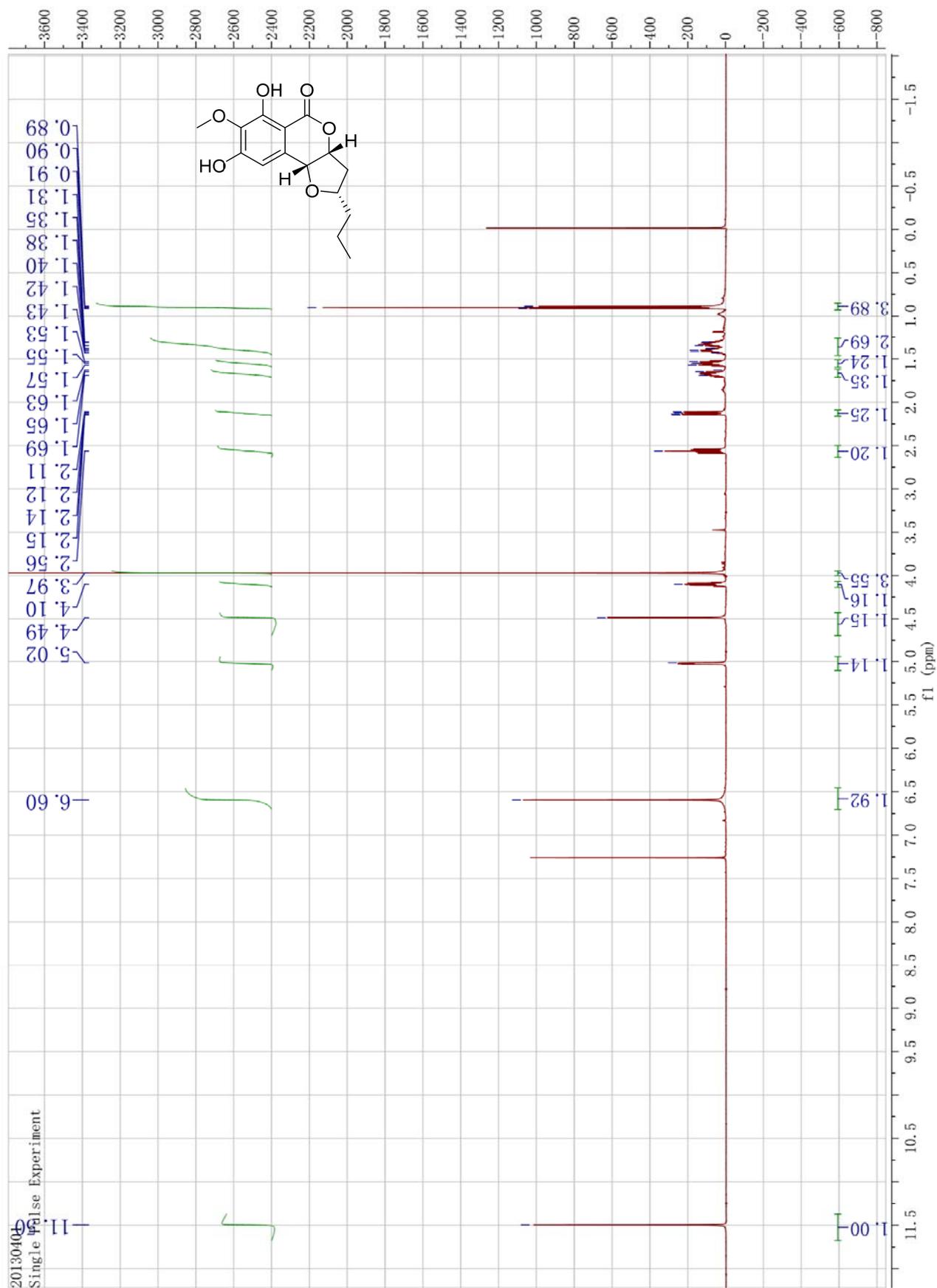


Figure S101. The ^{13}C -NMR spectrum of compound **18** in CDCl_3

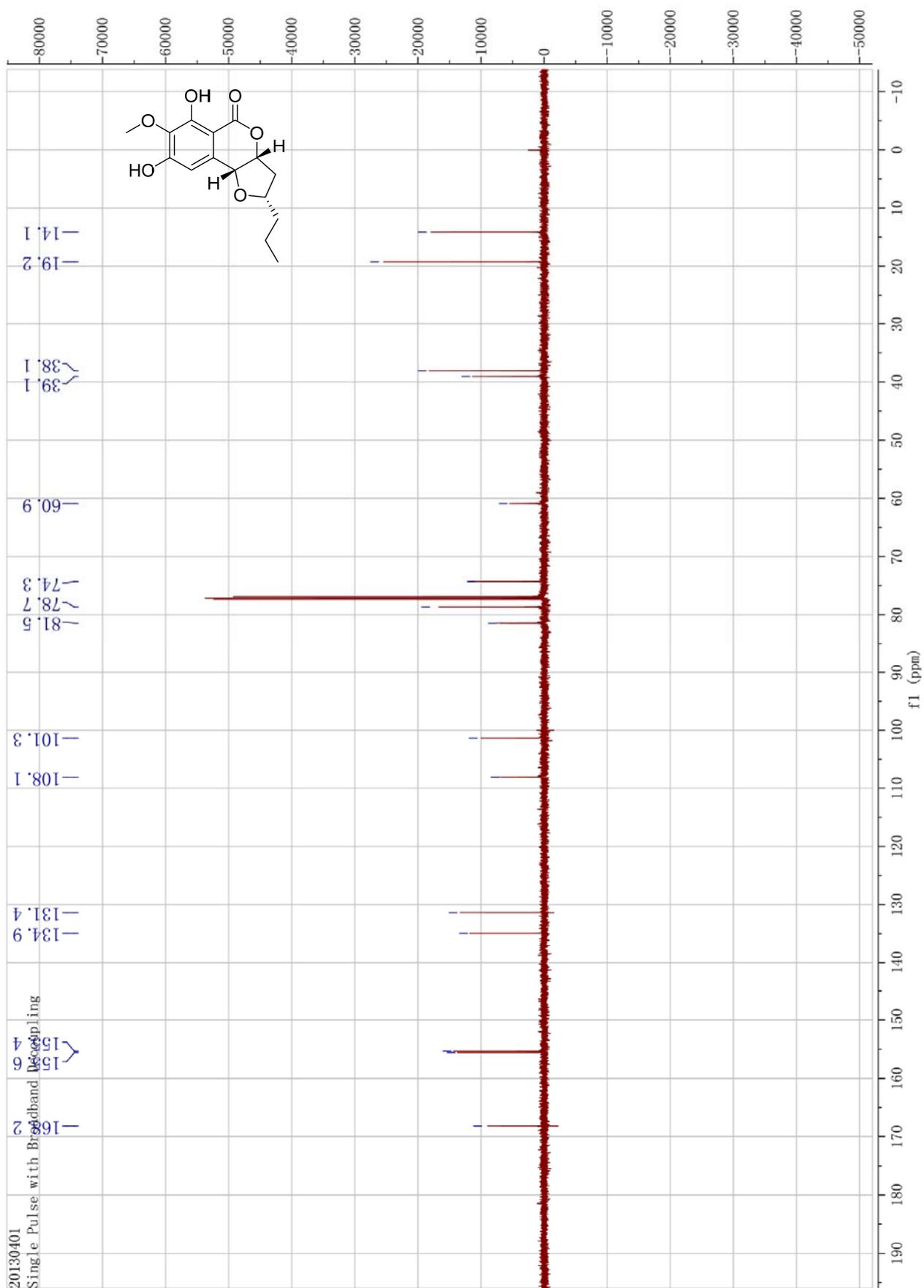


Figure S102. The DEPT spectrum of compound **18** in CDCl_3

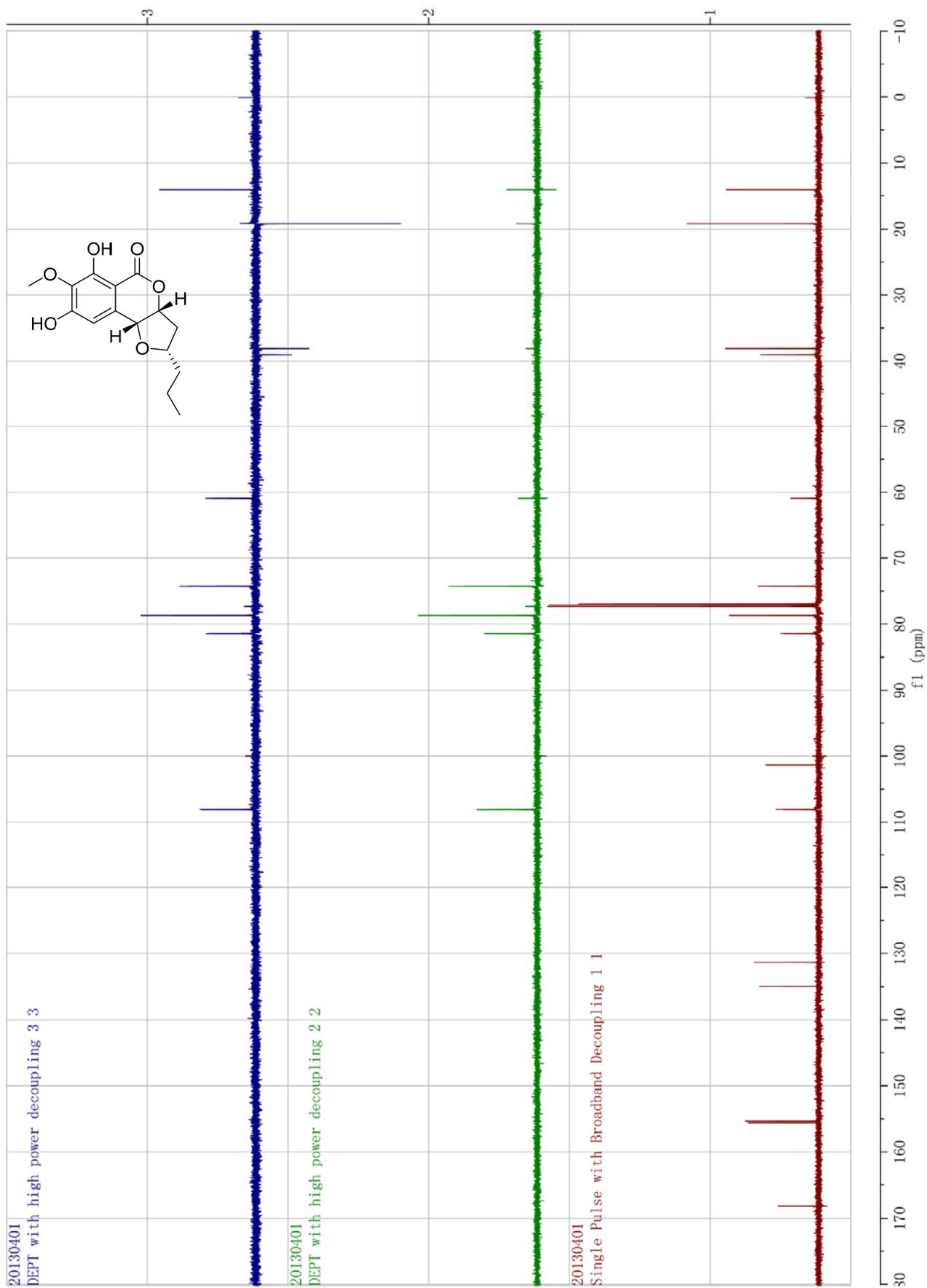


Figure S103. The HMQC spectrum of compound **18** in CDCl₃

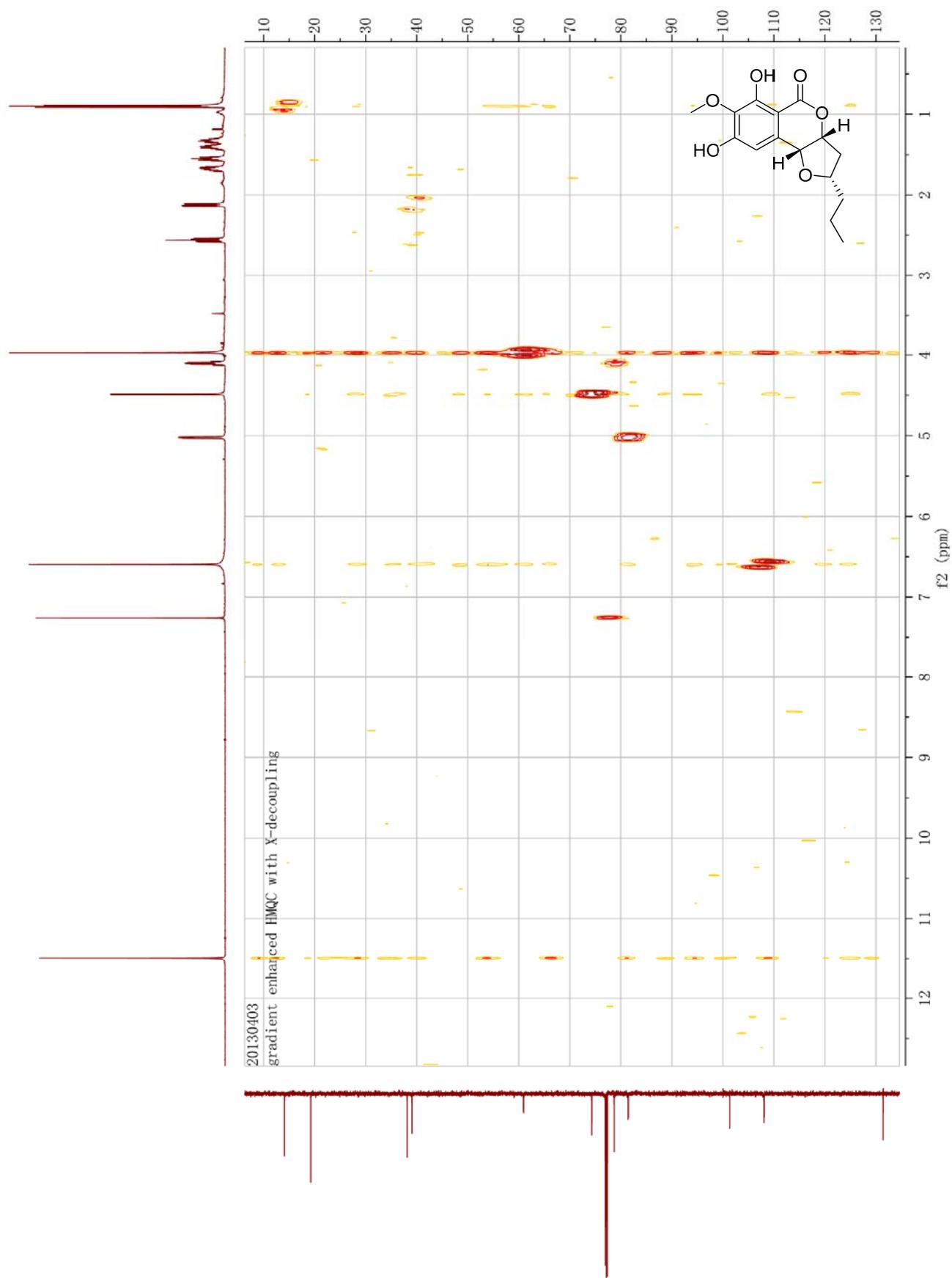


Figure S105. The HMBC spectrum of compound **18** in CDCl₃

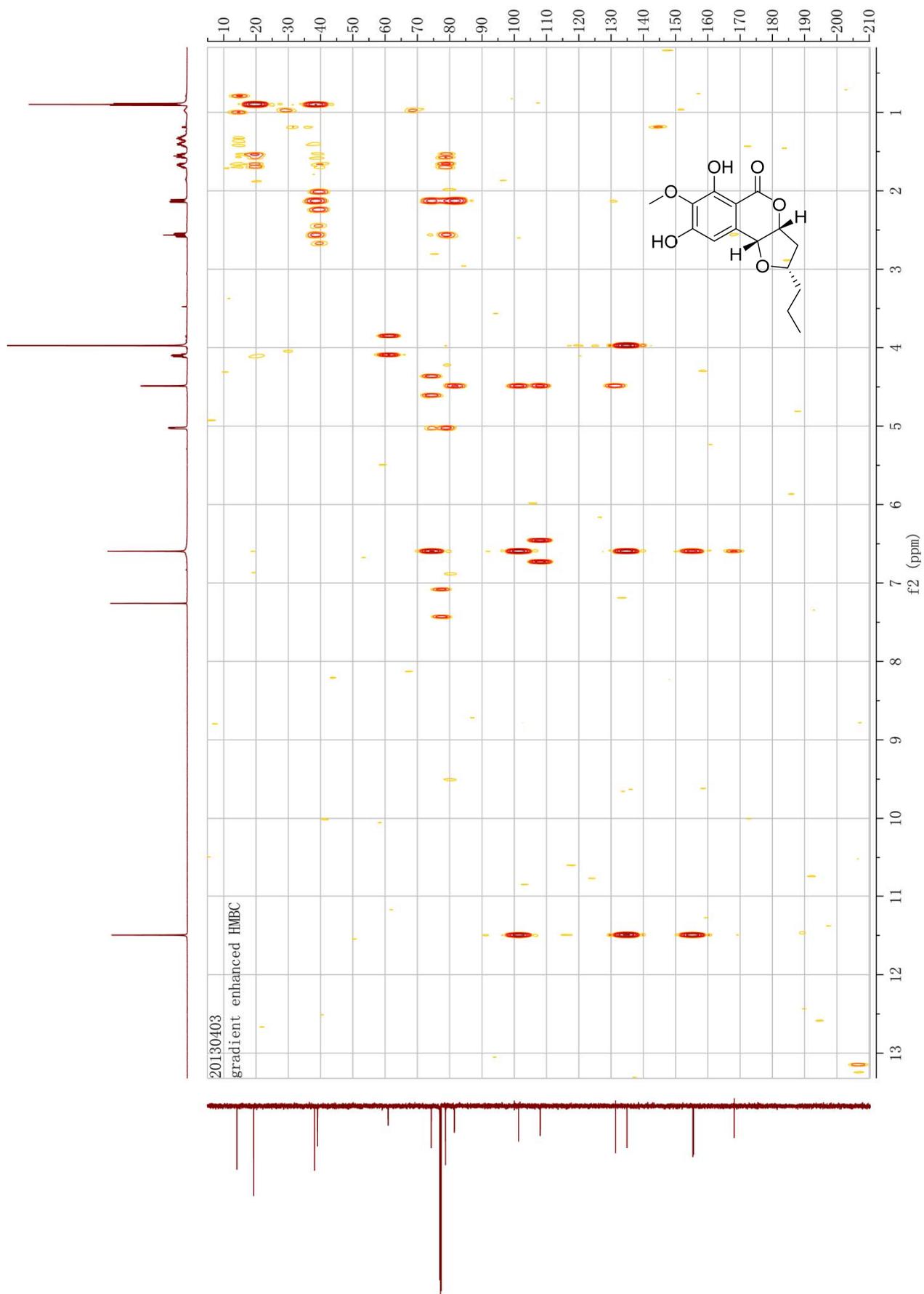


Figure S106. The NOESY spectrum of compound **18** in CDCl₃

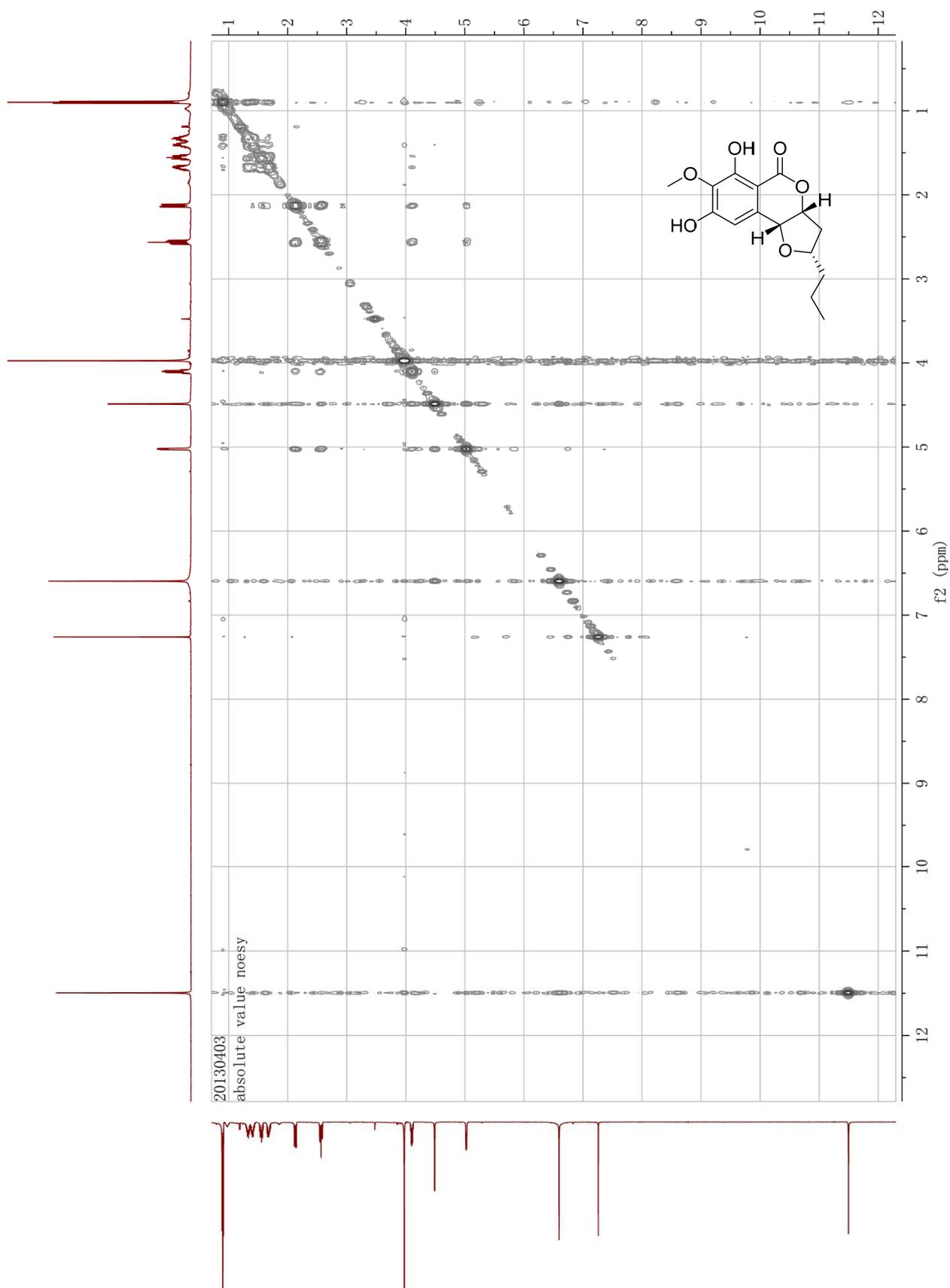
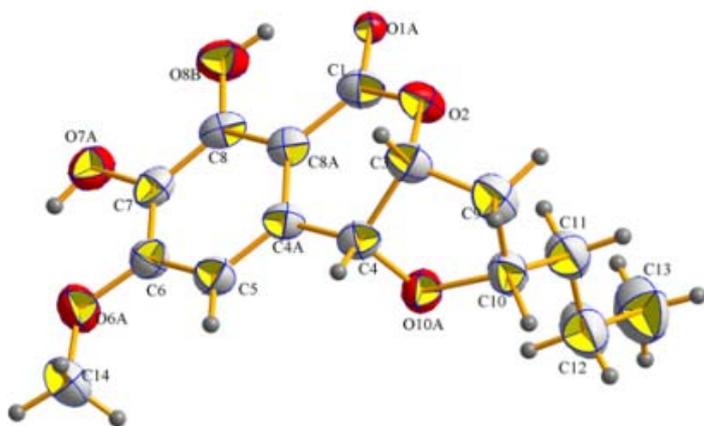


Figure S107. X-ray data and structure of compound **20**

7-O-Demethylmonocerin (20): Colorless Monoclinic crystals from MeOH-H₂O (1:1, v/v) with molecular formula of C₁₅H₁₈O₆; space group *P2*(1) with $a = 7.7687(7)$ Å, $b = 7.4049(5)$ Å, $c = 12.6454(9)$ Å, $V = 703.08(9)$ Å³, $Z = 2$, $D_{\text{calcd}} = 1.390$ Mg/m³, $\mu = 0.906$ mm⁻¹, and $F(000) = 312$; crystal size: $0.32 \times 0.27 \times 0.07$ mm³. $T = 293(2)$ K. Absolute structure parameter: 0.0(3). These data were obtained on a Bruker APEX DUO area detector diffractometer with graphite monochromatic Cu-K α radiation ($\lambda = 1.54178$ Å) and have been deposited in the Cambridge Crystallographic Data Centre with supplementary publication No 995362. The structure was solved by direct methods (SHELXS-97) and expanded using Fourier techniques (SHELXL-97). The final cycle of full-matrix least-squares refinement was based on 1549 unique reflections ($2\theta < 50^\circ$) and 193 variable parameters and converged with unweighted and weighted agreement factors of $R1 = 0.0409$, $wR2 = 0.1015$, and $R = 0.0499$ for $I > 2\sigma(I)$ data. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



ORTEP drawing of **20**

Figure S108. The $^1\text{H-NMR}$ spectrum of compound **12a** in CDCl_3

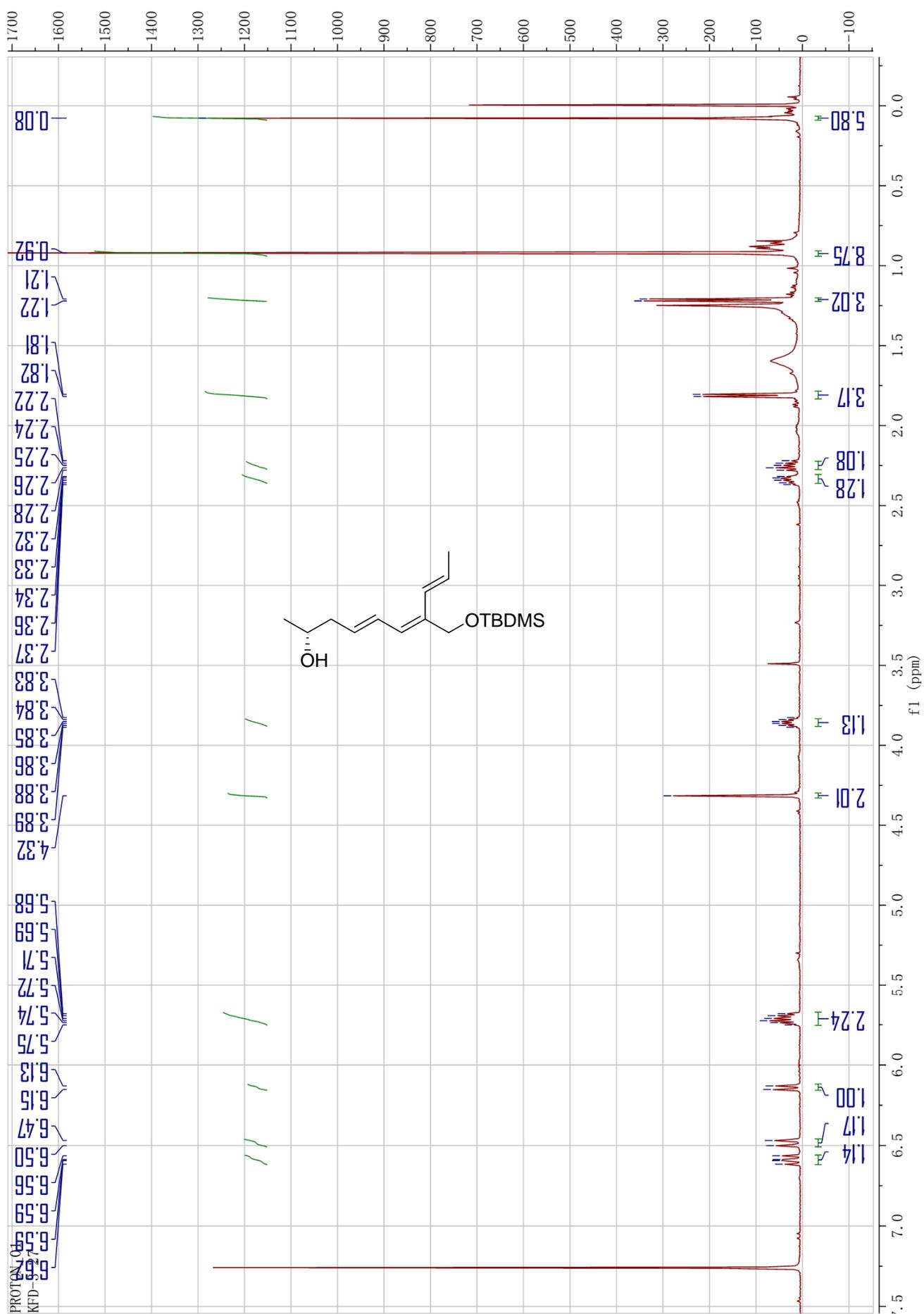


Figure S109. The DEPTQ spectrum of compound **12a** in CDCl₃

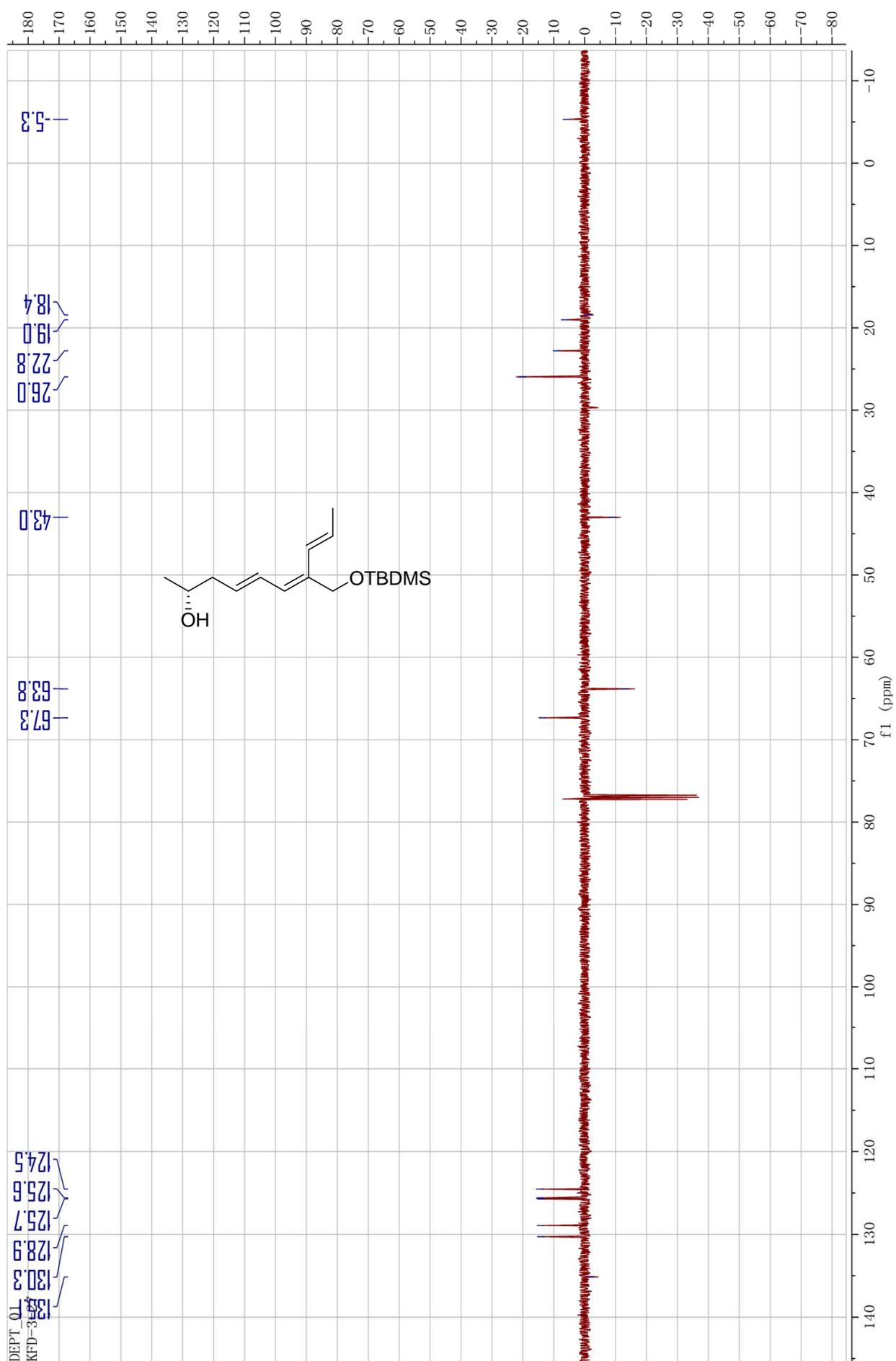


Figure S110. The HMQC spectrum of compound **12a** in CDCl₃

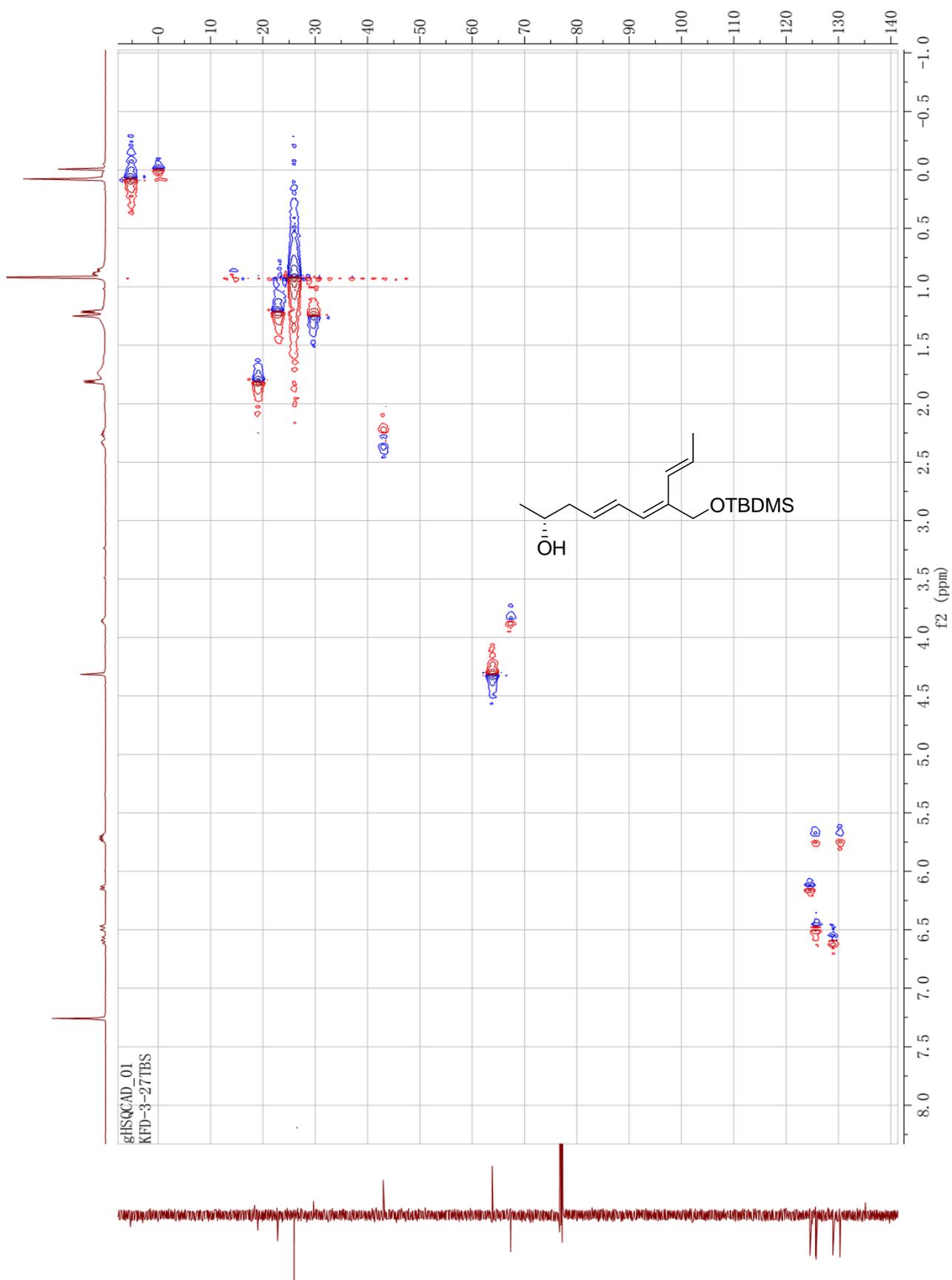


Figure S111. The ^1H - ^1H COSY spectrum of compound **12a** in CDCl_3

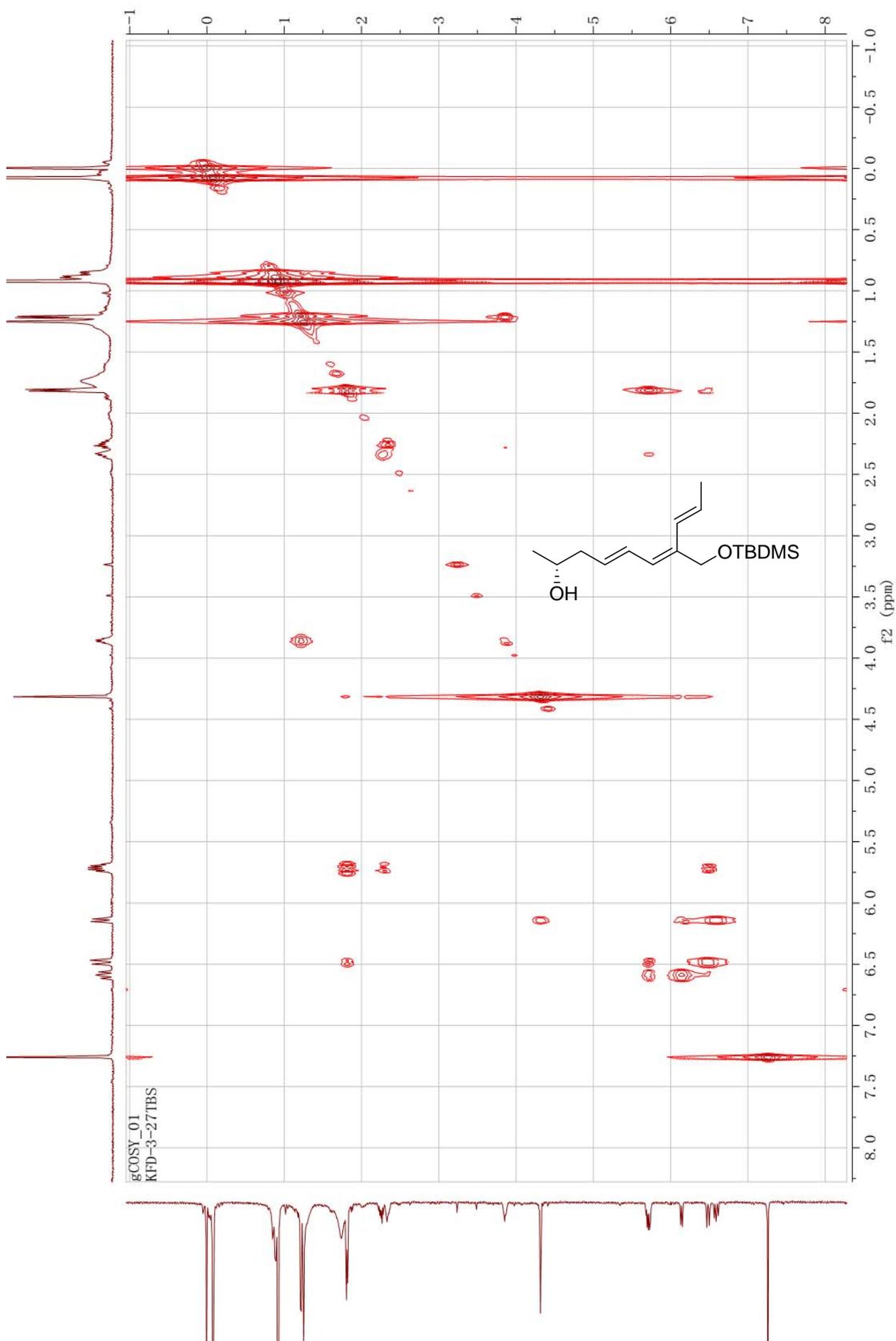


Figure S112. The HMBC spectrum of compound **12a** in CDCl₃

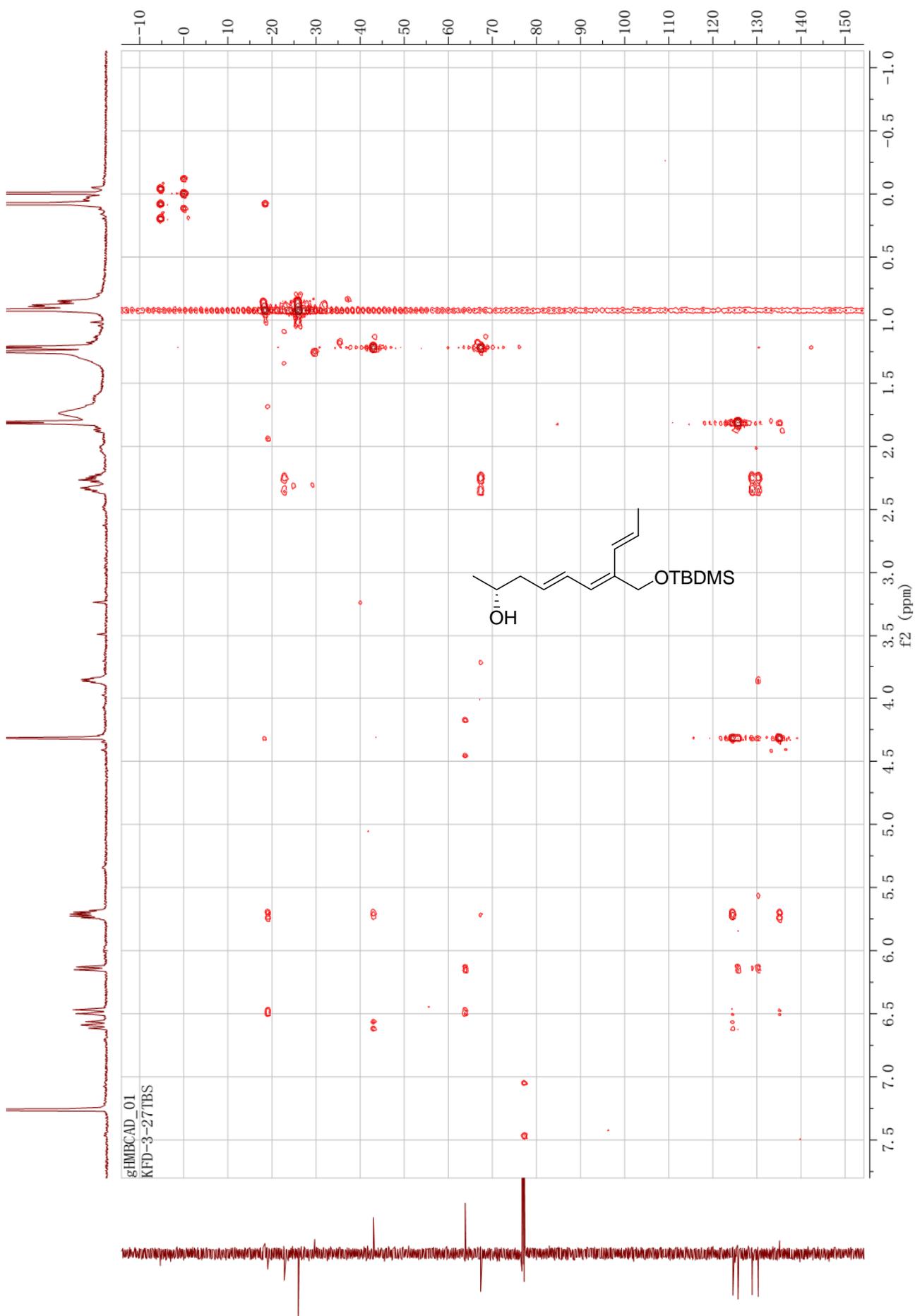


Figure S113. The ^1H -NMR spectrum of compounds **2a** and **2b** in CDCl_3

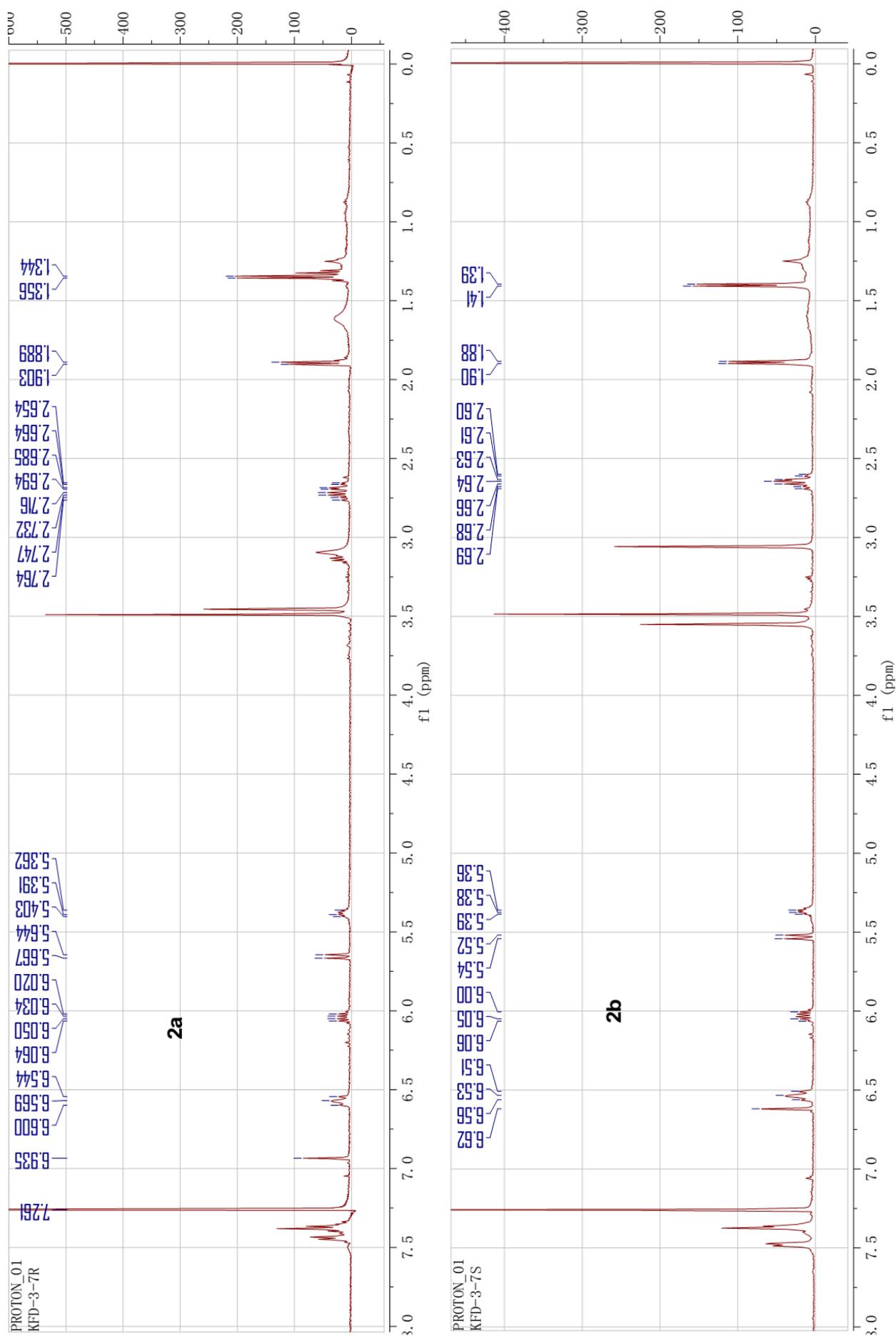


Figure S114. The ^1H -NMR spectra of compounds **3a** and **3b** in CDCl_3

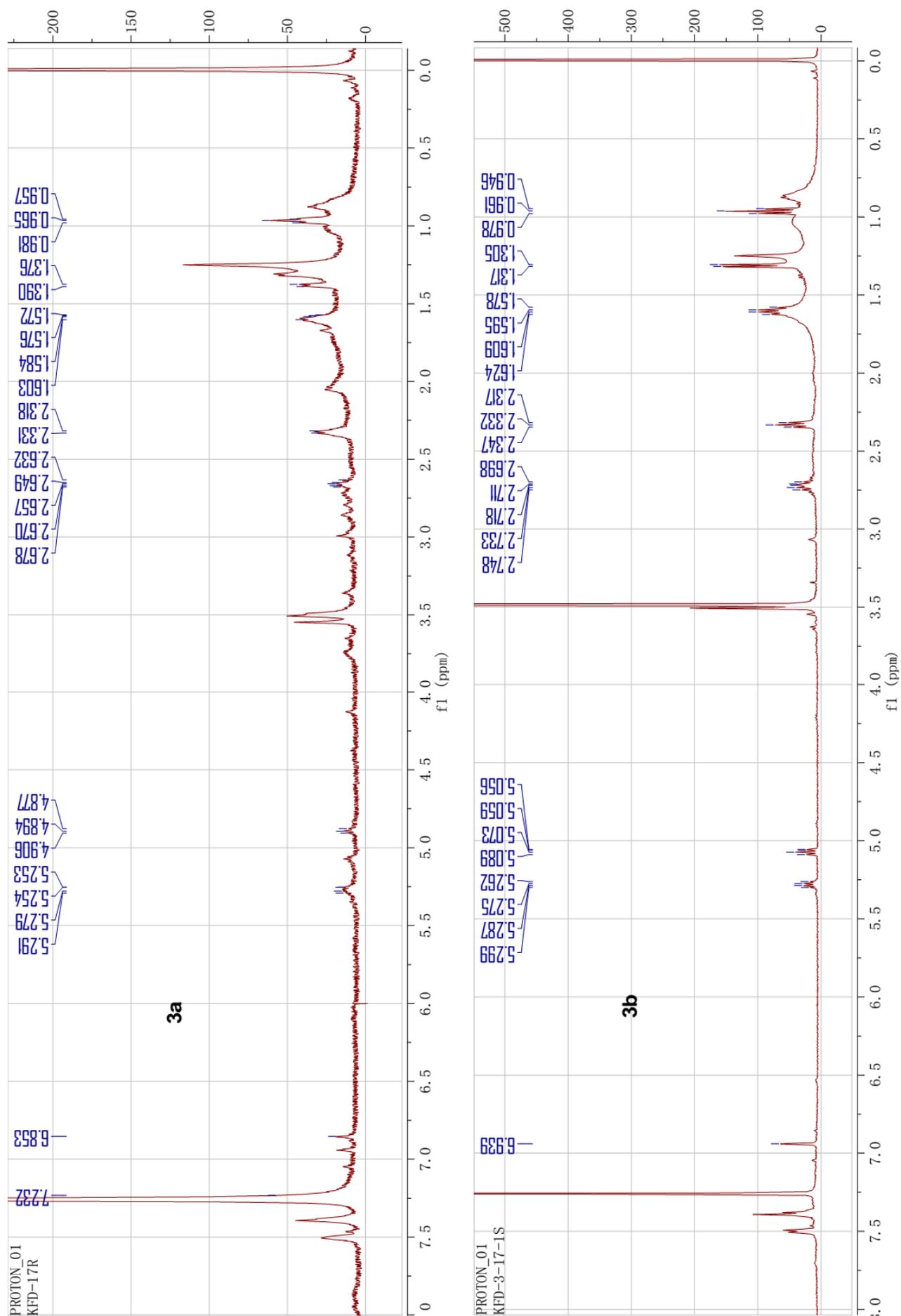


Figure S115. The $^1\text{H-NMR}$ spectra of compounds **5a** and **5b** in CDCl_3

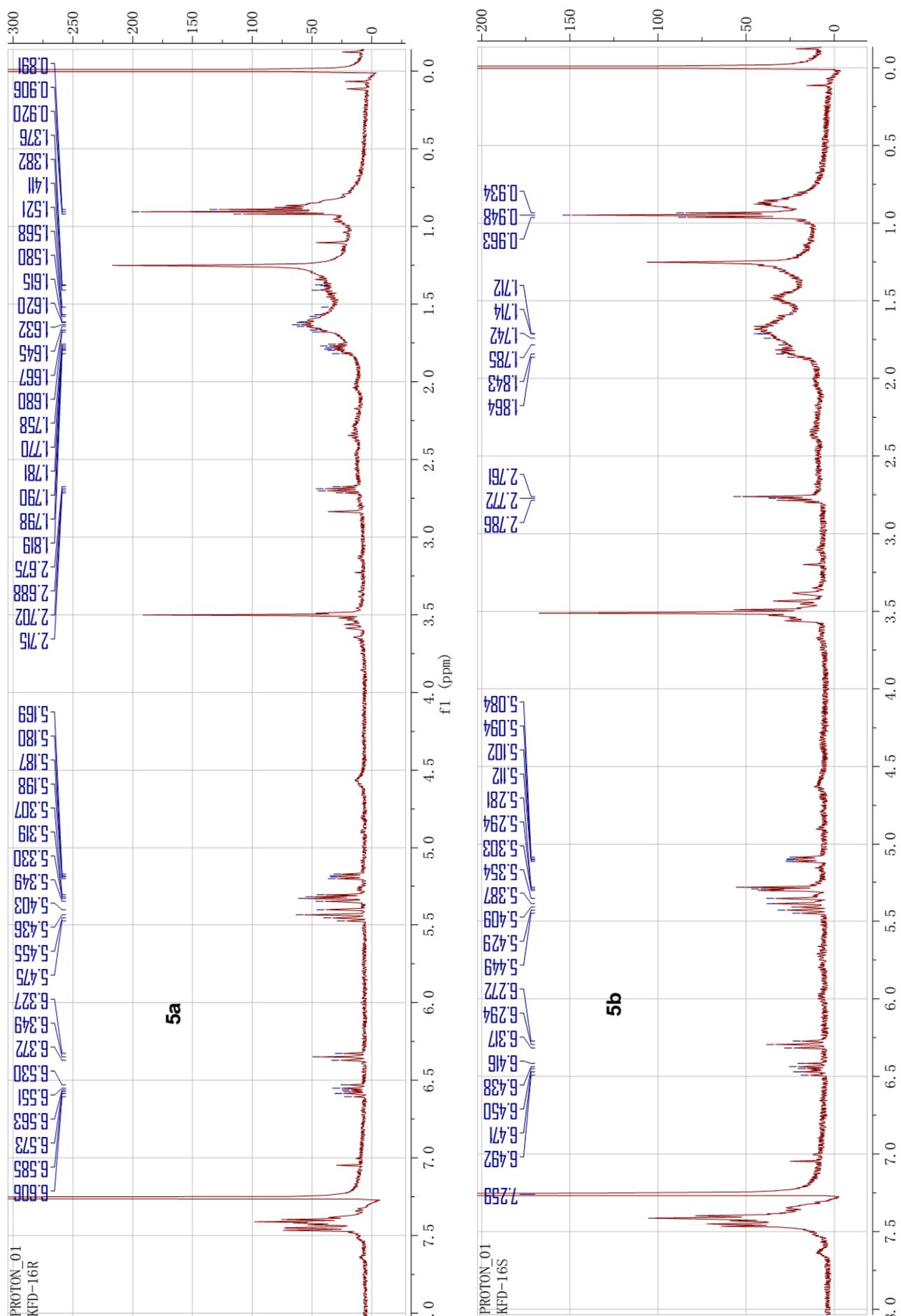


Figure S117. The $^1\text{H-NMR}$ spectra of compounds **7a** and **7b** in CDCl_3

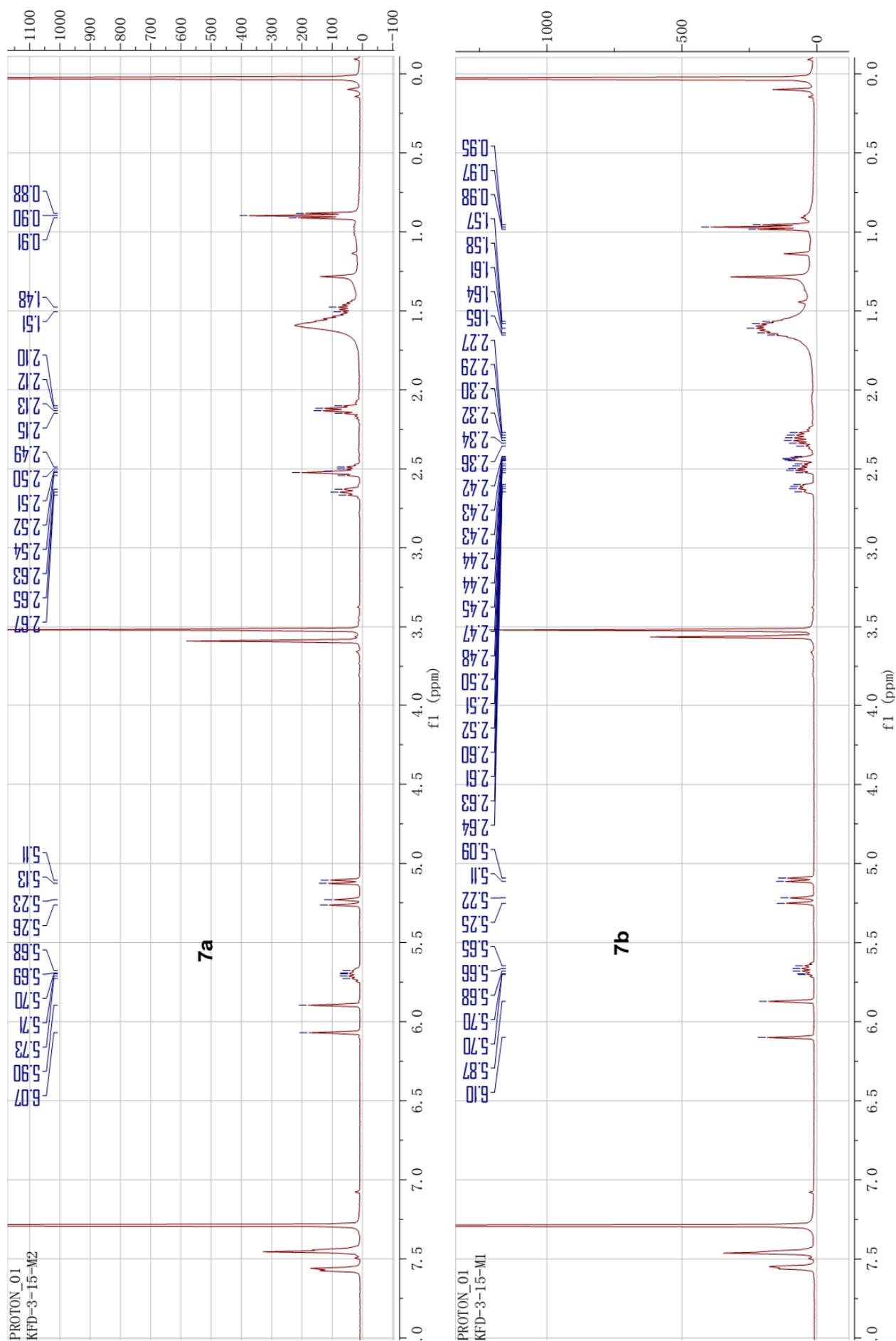


Figure S118. The ^1H -NMR spectra of compounds **12aa** and **12ab** in CDCl_3

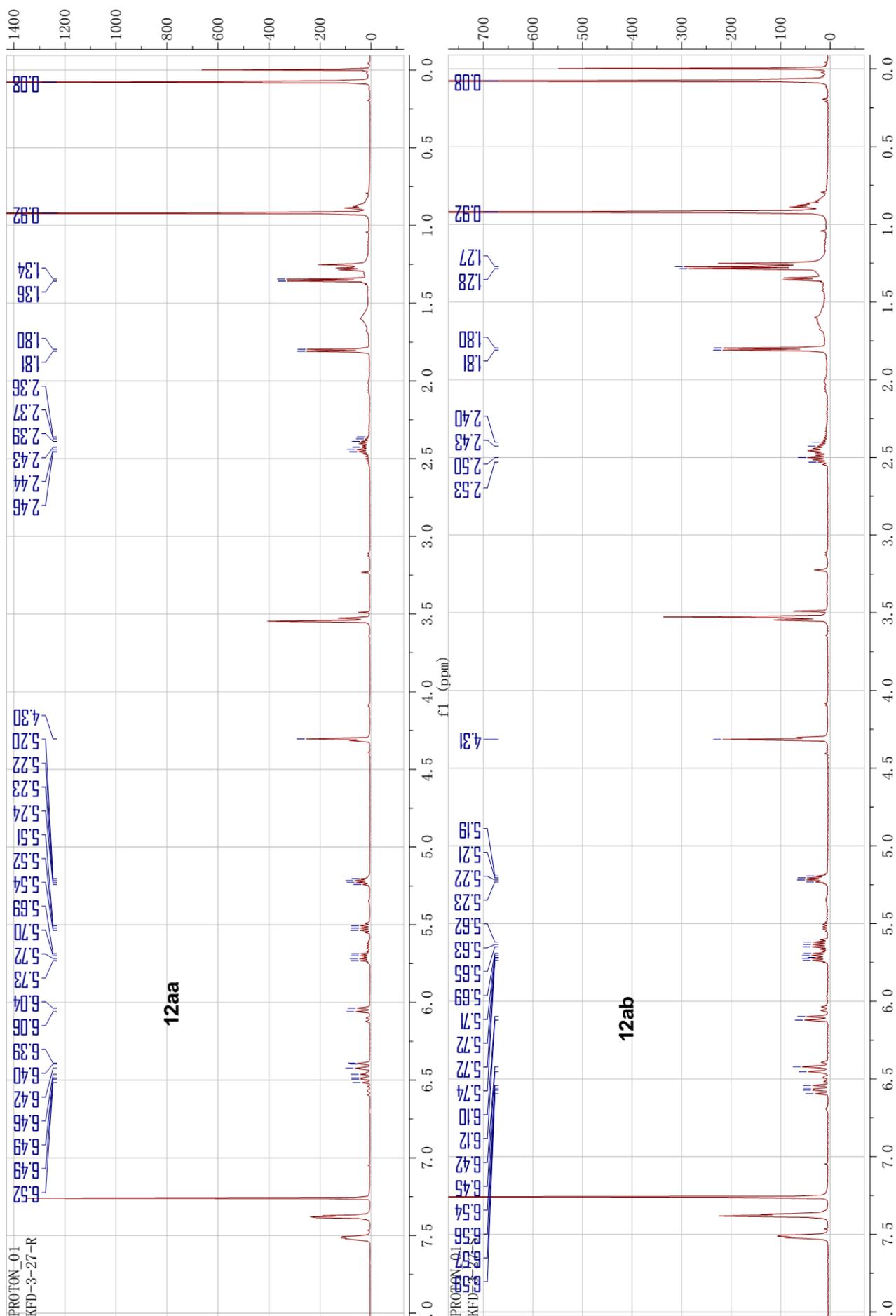


Figure S119. The HPLC analysis and the ESIMS of the reaction liquid of **12** to **12a**.

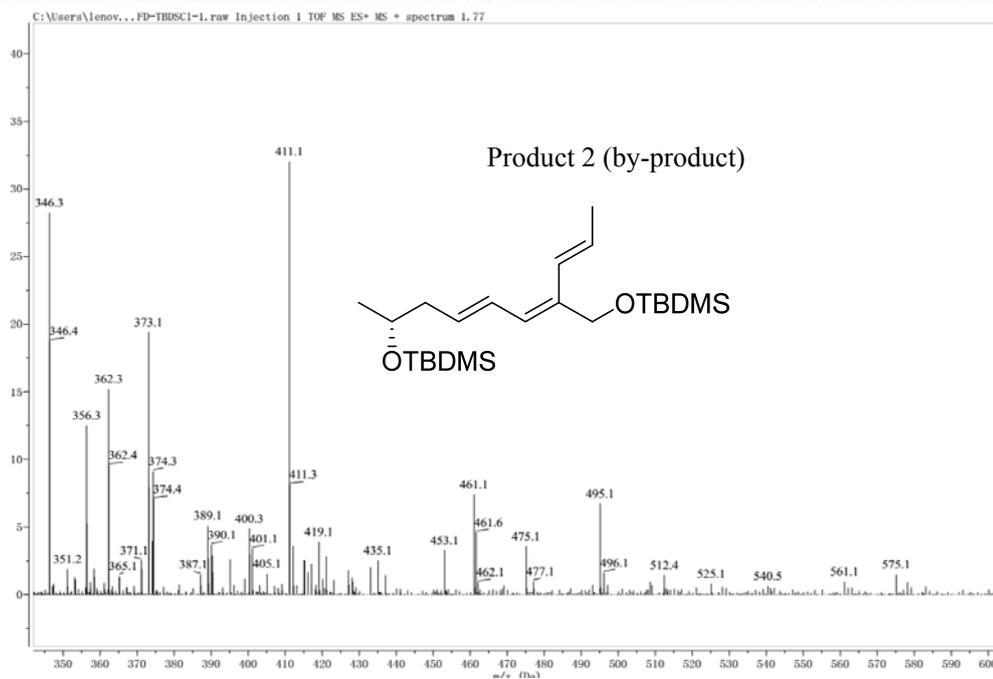
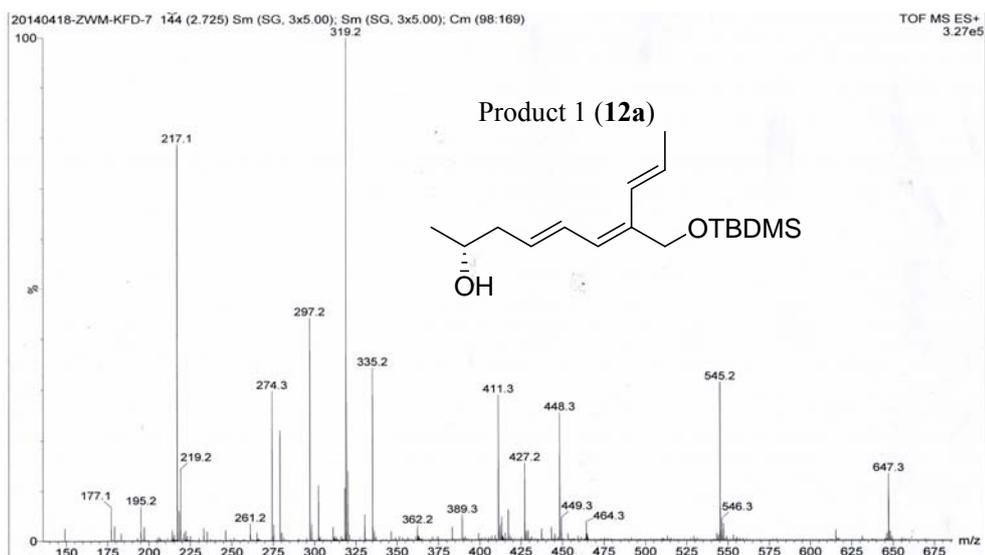
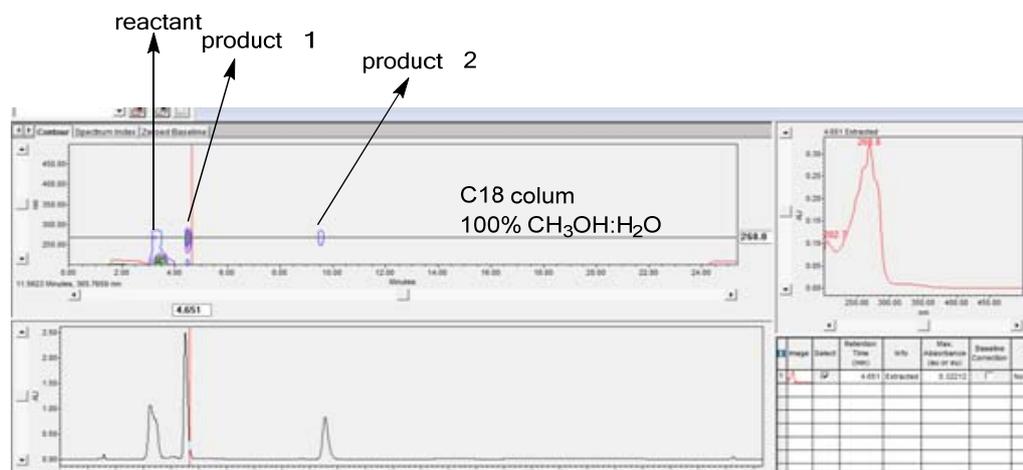


Figure S120. The HPLC analysis of compound **16** and synthetic **15** (ODS, 10% MeOH/H₂O, v/v).

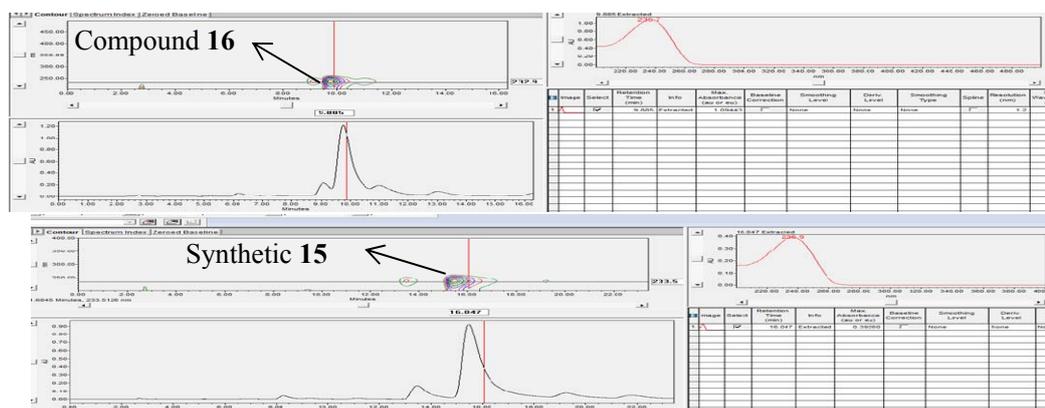


Figure S121. The HPLC analysis of **14** and the synthetic **14** (ODS, 20% MeOH/H₂O, v/v).

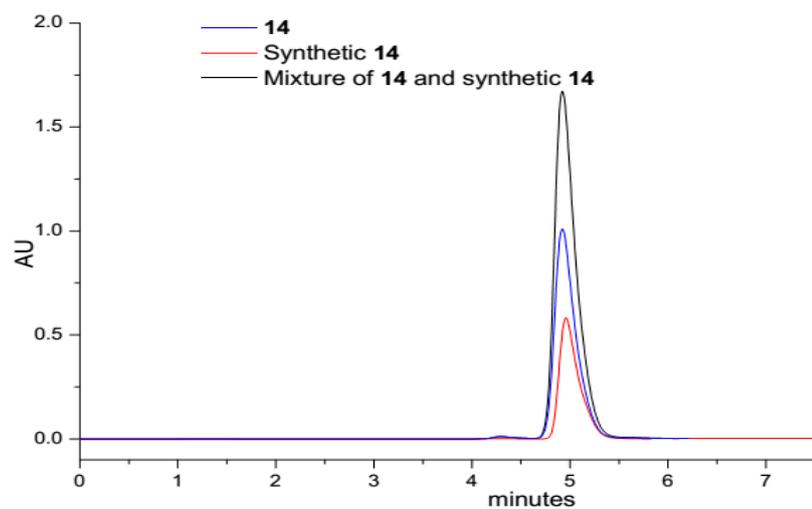


Figure S122. The HPLC analysis of **15** and the synthetic **15** (ODS, 15% MeOH/H₂O, v/v).

