Supporting Information

A new rhodamine based chemodosimeter for Ni²⁺ with high sensitivity and selectivity

Ivan Zhang,^{*a*} Yi Wang,^{*a*} Chao Wan,^{*b*} Zhen Xing,^{*c*} Wen Li,^{*a*} Minjie Li*^{*a*} and Sean Xiao-An Zhang*^{*a*}

^a State Key Lab of Supramolecular Structure and Materials, College of Chemistry, Jilin University, Changchun 130012, People's Republic of China

^b College of Environment and Resources, Jilin University, Changchun 130012, People's Republic of China

^c College of Life of Science, Jilin University, Changchun 130012, People's Republic of China

Table of Contents

- 1. Reagents and Apparatus
- 2. Synthesis of Rha-py
- 3. Spectroscopic Materials and Methods
- 4. Optimization of experimental conditions
- 5. Determination of Ni²⁺
- 6. Discrimination of Cu²⁺
- 7. IR spectra, Mass spectroscopy, ¹H NMR, ¹³C NMR and X-Ray crystallographic analyses for Rha-amino

1. Reagents and Apparatus

All chemicals were obtained from commercial suppliers and used without further purification. Spectroscopic grade solvents have been used for spectroscopic experiments. The solutions of metal ions were prepared from their nitrate salts, except for FeCl₂, CoCl₂, HgCl₂ and MnCl₂. Water was purified by Milli-Q system. Other chemicals are of analytical reagent grade and used without further purification. Absorption spectra were measured on an Analitik Jena Specord ®210 plus UV/VIS spectrophotometer, path length was 1 cm. All fluorescence measurements were carried out on a Shimadzu-RF5301 fluorescence spectrometer with both excitation and emission bandwidths set at 3.0 nm. All pH values were measured with Sartorius PB-21. ¹H NMR (500 MHz) and ¹³C NMR (126 MHz) spectra were recorded on a Bruker AVANCE500 using TMS as a standard at room temperature. All chemical shifts are reported in the standard notation of parts per million. LC-HRMS analysis was performed on an Agilent 1290-microTOF-Q II mass spectrometer. MALDI-TOF was obtained by Bruker Autoflex speed TOF/ TOF. IR spectra studies were performed on Vertex 80/80V FT-IR spectrometer with LN-MCT Mid DC detector over the range of 4000-800 cm⁻¹ using a KBr plate. Single-crystal X-ray diffraction data for Rha-amino was recorded on a Rigaku RAXIS-PRID diffractometer using the ω-scan mode with graphitemonochromator Mo·K α radiation ($\lambda = 0.71073$ Å). Melting point was determined using a SGW X-4B microscopy melting point apparatus (Shanghai).

2. Synthesis of Rha-py



Synthesis of Rha-nitro: Synthesis of 3', 6'-bis (diethylamino)-4-nitro-3H-spiro [isobenzofuran-1,9'-xanthen]-3-one (**Rha-nitro**): Rha-nitro was synthesized via a similar route as Nagano's with slightly modification.¹ A mixture of 3-nitrophthalic anhydride (1.93 g, 10 mmol), zinc chloride (1.35g, 10 mmol) and N, N-diethyl-3-aminophenol (1.65 g, 10 mmol) was stirred at 110°C for 0.5h. Then added another part of zinc chloride (1.35 g, 10 mmol) and N, N-diethyl-3-aminophenol (1.65 g, 10 mmol), and stirred for 6 h at 160°C. The reaction was quenched by water. The petunia amorphous solid was refluxed in 2M HCl (100 mL) for 2 h. The mixture was neutralized with sodium hydroxide, and the violet precipitate was collected and dried. Extraction with dichloromethane could also be used for collection. The products were purified by chromatography (silica, 10% CH₃OH in CH₂Cl₂) to afford the violet Rha-nitro (2.83 g, 58% yield); Melt point: 158-159°C. LC-HRMS: m/z calculated for [M + H]⁺ 488.2180, found 488.2186. ¹H NMR: $\delta_{\rm H}$ (500 MHz, DMSO) 8.11 (1 H, d, *J*= 7.8), 7.86 (1 H, t, *J*=7.8), 7.57 (1 H, d, *J*= 7.7), 6.74 (2 H, d, *J*= 9.0), 6.68 – 6.56 (4 H, m), 3.44 (8 H, q, *J*= 6.6), 1.13 (12 H, t, *J*= 7.0). ¹³C NMR: $\delta_{\rm C}$ (126 MHz, DMSO) 163.62, 153.85, 152.30, 150.85, 149.19, 146.43, 134.22, 129.62, 127.25, 124.32, 123.79, 109.85, 106.31, 96.59, 44.19, 12.33.

Synthesis of Rha-amino: Synthesis of 4-amino-3', 6'-bis (diethylamino)-3H-spiro [isobenzofuran -1,9'-xanthen]-3-one (**Rha-amino**): Rha-nitro (2.44g, 5.0 mmol) was reduced with 0.1g 10% Pd/C under hydrogen atmosphere to give Rha-amino.² The residue was filtered, the solution was evaporated and purified by chromatography (silica, 5% CH₃OH in CH₂Cl₂) to give purple powder (1.84g, 80%). Melt point: 229-230°C. LC-HRMS: m/z calculated for $[M + H]^+$ 458.2438, found 458.2427. ¹H NMR: $\delta_{\rm H}$ (500 MHz, CDCl₃) 7.34 (1 H, t, *J*=7.7), 6.69 (3 H, dd, *J*=21.6, 8.4), 6.42 (3 H, d, *J*=7.4), 6.36 (2 H, d, *J*=8.1), 5.38 (2 H, s), 3.36 (8 H, q, *J*=7.0), 1.17 (12 H, t, *J*=7.0). ¹³C NMR: $\delta_{\rm C}$ (126 MHz, DMSO) 170.14, 153.57, 152.41, 148.89, 147.44, 136.20, 128.57, 113.72, 109.73, 108.19, 108.05, 106.14, 96.88, 83.62, 43.77, 12.33.

Synthesis of Rha-py: Synthesis of 3', 6'-bis (diethylamino)-4-((pyridin-2-ylmethyl) amino) -3H-spiro [isobenzofuran -1, 9'-xanthen]-3-one (**Rha-py**). Rha-amino (0.487g, 1.0mmol), Sodium triacetoborohydride (0.424g, 2.0mmol) and 2-pyridinecarboxaldehyde (0.214g, 2.0mmol) was mixed in ClCH₂CH₂Cl with acetic acid catalysis under room temperature for 10h. The reaction was quenched by sodium hydrogen carbonate solution. Extraction with dichloromethane was used for collection. The product was purified by chromatography (silica, 3% CH₃OH in CH₂Cl₂) to give the final product Rha-py (0.42g, 76%); Melt point: 160-161°C. LC-HRMS: m/z calculated for [M + H]⁺ 549.2860, found 549.2851. ¹H NMR: δ_H (300 MHz, DMSO) 8.61 (1 H, d, *J*= 4.4), 7.83 (1 H, td, *J*= 7.7, 1.7), 7.56 (1 H, t, *J*= 5.5), 7.50 - 7.38 (2 H, m), 7.37 - 7.26 (1 H, m), 6.69 (1 H, d, *J*= 8.3), 6.57 (2 H, d, *J*= 8.6), 6.50–6.35 (4 H, m), 6.25 (1 H, d, *J*= 7.3), 4.66 (2 H, d, *J*= 5.5), 3.35 (8 H, dd, *J*= 12.4, 5.3), 1.09 (12 H, t, *J*= 6.9). ¹³C NMR: δ_C (126 MHz, DMSO) 170.72, 158.09, 154.49, 152.90, 149.52, 149.45, 146.98, 137.42, 129.06, 122.89, 122.07, 120.00, 110.39, 110.27, 109.24, 108.59, 106.28, 97.38, 84.30, 47.77, 44.25, 12.82.

3. Spectroscopic Materials and Methods

Experiments involving the measurement of Rha-py absorbance and fluorescence were performed on CH₃CN buffered with HEPES (9/1, v/v, pH=7.1, 20 mM). Stock solution of Rha-py (1×10^{-4} M) was prepared in CH₃CN. The UV-Vis absorption spectra were recorded from 200 nm to 800 nm. The fluorescence emission spectra were recorded at an excitation wavelength of 540 nm with emission wavelength ranging from 550 nm to 800 nm.



4. Optimization of experimental conditions

Figure S1. Plot of absorbance at 550 nm vs. water content in CH_3CN . Black line and square dot represent the Rhodamine B (4 μ M). Blue line and circle dot represent Rha-amino (4 μ M). Contrast to Rhodamine B, the absorbance of Rha-amino increase along with water content, Rhodamine B exist major in its zwitterionic form through the process.



Figure S2. Absorbance of Rha-py (4 μ M) in CH₃CN buffered with HEPES (9/1, v/v, pH=7.1, 20 mM) of different pH in the absence (bottom line, circle, at 550 nm) and presence (top line, square, at 550 nm) of 40 μ M Ni²⁺. The pH values were adjusted by aqueous solution of NaOH (aq, 1M) or HCl (aq, 1M). From the figure, the range between pH=4.5 to pH=7.5 is suitable for the detection of Ni²⁺ by colorimetric method. We chose pH=7.1 as the experiment condition.



Figure S3. Fluorescence spectra of Rha-py (4 μ M) in CH₃CN buffered with HEPES (9/1, v/v, pH=7.1, 20 mM) after the addition of different metal ions (40 μ M). Metal ions: Na⁺, K⁺, Mg²⁺, Ca²⁺, Zn²⁺, Co²⁺, Ni²⁺, Cd²⁺, Mn²⁺, Ag⁺, Hg²⁺, Al³⁺, Cr³⁺, Fe³⁺, Fe²⁺. Excitation was performed at 540 nm.

5. Determination of Ni²⁺



Figure S4. Job's plot of Rha-py and Ni²⁺ with a total concentration of 20 μ M in CH₃CN buffered with HEPES (9/1, v/v, pH=7.1, 20 mM), when the molar fraction of Ni²⁺ was 0.5, the absorbance at 550 nm reached a maximum, indicating a 1:1 metal-ligand ratio.



Figure S5. UV-Vis spectra of Rha-py (4 μ M) upon the addition of Ni²⁺. Inset: plot of absorbance at 550 nm vs. number of equivalents of Ni²⁺.

The association constant (K_a) of Rha-py with Ni²⁺ was determined based on the absorbance titration curve using the Benesi-Hildebrand equation as follows³:

$$1/(A_{max} - A_0) = [1/(A - A_0)] \times [1/(Ka \times [Ni^{2+}] + 1)]$$

where A and A_0 represent the absorbance of host in the presence and absence of ions, respectively, A_{max} is the saturated absorbance of host in the presence of excess amount of ions; [Ni²⁺] is the concentration of Ni²⁺ added.



Figure S6. Benesi-Hildebrand plot of Rha-py with Ni²⁺. From the plot, we can get the association constants (K_a) of Rha-py and Ni²⁺ is 4.87×10^4 M⁻¹.



Figure S7. Plot of the absorbance at 550 nm for a mixture of Rha-py (4 μ M) in CH₃CN buffered with HEPES (9/1, v/v, pH=7.1, 20 mM). The detection limit for Ni²⁺ is 7.8 × 10⁻⁸ M.

Table S1. Condition: Rha-py $(4\mu M)$ in CH₃CN buffered with HEPES (9/1, v/v, pH=7.1, 20 mM). The water sample was acquired at South Lake, Changchun and simply filtrated.

Sample	Ni^{2+} added (μM)	Ni^{2+} found (μM)	Recovery (%)	R.S.D. (%)
South Lake	2.00	1.89	94.5	3.7
	4.00	3.84	96.0	3.2

6. Discrimination of Cu²⁺



Figure S8. The blue line represented the change in absorption at 550 nm of Rha-py (4 μ M) in CH₃CN buffered with HEPES (9/1, v/v, pH=7.1, 20 mM) with time in the presence of Ni²⁺ (40 μ M). The red line represented the change in absorption at 540nm of Rha-py (4 μ M) and Cu²⁺ (40 μ M) with time in the same buffer.

7. IR spectra, Mass spectroscopy, ¹H NMR, ¹³C NMR and X-





Figure S9. IR spectra of Rha-py and Rha-py-Ni²⁺ in KBr, red line represented the infrared light absorbance of Rha-py, the blue line represented the infrared light absorbance of Rha-py-Ni²⁺ in KBr disks.



Figure S10. MALDI-TOF spectra of Rha-py and Ni²⁺. The number 605.34 stood for the compound of Ni²⁺ and Rha-py, which confirmed the coordination ratio was 1:1.



Figure S11. LC-MS spectra of (a) Rha-amino, from the spectrum of liquid chromatography, we can find the retention time was 2.7 min. (b) Rha-py, from the spectrum, we can find the retention time was 1.8 min. (c) The LC-MS spectra of Rha-py and Ni²⁺ in CH₃CN buffered with HEPES (9/1, v/v, pH=7.1, 20 mM) (HEPES buffer), from the spectrum, we can find the signal of HEPES and Rha-py, but no Rha-amino. (d) The LC-MS spectra of Rha-py and Cu²⁺ in HEPES buffer, from the spectrum, we can find the signal of HEPES and Rha-amino, but no Rha-py, which illustrated the probe was degraded by Cu²⁺ completely.















Table S2. Summary of crystal data and intensity collection parameters for Rhaamino.

The crystal data for Rha-amino was deposited into Cambridge Crystallographic Data Centre with CCDC numbers of 1036431.

Compound	Rha-amino	
Formula	C28 H31 N3 O3	
Formula mass	457.56	
Space group	P 2 ₁ 2 ₁ 2 ₁	
<i>a</i> / Å	12.122(2)	
b/ Å	13.002(3)	
c/ Å	15.716(3)	
α/ °	90	
β / °	90	
γ/ °	90	
<i>V</i> / Å ³	2476.9(9)	
Ζ	4	
ρ/ g.cm ⁻³	1.227	
$\mu/ \text{ mm}^{-1}$	0.080	
F_{000}	976	
Temp, (K)	293(2)	
No. of collected reflns.	18290	
No. of unique reflns. (Rint)	4351(0.0636)	
Data/restraints/parameters	4351 / 29 / 311	
R_1 , w R_2 [obs I > 2 σ (I)]	0.0831, 0.2154	
R_1 , w R_2 (all data)	0.1277, 0.2457	
Residual peak/ hole e.Å ⁻³	0.532/-0.365	
Goodness-of-fit on F ²	1.063	

References

- H. Kojima, M. Hirotani, N. Nakatsubo, K. Kikuchi, Y. Urano, T. Higuchi, Y. Hirata and T. Nagano, *Anal. Chem.*, 2001, 73, 1967.
- 2. Y. Liu, X. Dong, J. Sun, C. Zhong, B. Li, X. You, B. Liu and Z. Liu, Analyst, 2012, 137, 1837.
- 3. H. A. Benesi and J. H. Hildebrand, J. Am. Chem. Soc., 1949, 71, 2703.