Supplementary Material

Alizarin Red S –Zinc(II) Fluorescent Ensemble For Selective Detection of Hydrogen Sulphide and Assay with H₂S donor

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Synthesis of H₂S Donor



H₂S donor compound benzoic (methyl carbonic) dithioperoxyanhydride was synthesized by following reported procedure¹. Accordingly in a 100 ml round bottom flask 300µl of methoxycarbonylsulfenyl chloride (3.2 mM) was dissolved in 20 ml of distilled diethylether, to this 324 µl of thiobenzoic acid (2.7 mM) was added drop wise at room temperature under stirring at N₂ atmosphere. After 2 hours, the solution was concentrated under high pressure and passed through a short silica column chromatography using Hexane/CHCl₃ as a solvent. After removal of the solvent in rotary evaporator, a colorless oily product was obtained. This product was stored under cold condition. Yield: 350 mg (52 %), ¹H NMR (400 MHz, CDCl₃): δ (ppm) = 3.85 (3H), 7.54 (2H), 7.59 (1H), 7.94 (1H): ESI-MS (-ve) (m/z) = 265.2 [M+Cl]⁺, 337.2 [M+3Cl]⁺

UV-Visible absorption studies

Absorbance study of ARS with Zn: For the absorbance study, stock solution of ARS (0.5 mM) was prepared in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4).For the titration 120 µl of stock solution was diluted to 2000 µL by adding 1878 µL buffer. The titration was done with different concentration of zinc perchlorate (0 – 30 µM) in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4), each spectra was recorded in standard quartz cuvette cell of length 1 cm. The change in absorption was monitored at 530 nm.

Absorbance study of ARS-Zn vs H_2S : For the absorbance study stock solution of ARS (0.5 mM) was prepared and 120µl was diluted to 2000 µl followed by addition of 40µl of zinc perchlorate (1.5 mM) in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4). The titration was done with different concentration of H_2S (0 – 44 µM). The titration was done in standard quartz cuvette cell of length 1 cm.

Fluorescence titration studies

Fluorescence study of ARS with Zn: For the emission study, stock solution of ARS (0.5 mM) was prepared in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4) and for the titration 120 µL of stock solution was diluted to 2000 µL by adding 1878 µL buffer. The titration was done with different concentration of zinc perchlorate (0 - 30 µM) in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4), excitation wavelength was 530 nm with a slit width of 10/10.

Fluorescence study of ARS-Zn(II) with H₂S: For the emission study stock solution of ARS (0.5 mM) was prepared and 120 μ l was diluted to 2000 μ l followed by addition of 40 μ l of zinc perchlorate (1.5 mM) in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4). The titration was done with different concentration of H₂S (0 - 42 μ M). The excitation wavelength was 530 nm with a slit width of 10/10.

Competitive experiments

For the competitive studies with different analytes stock solution of **ARS** (0.5 mM) was prepared and 120µl was diluted to 2000 µl followed by addition of 40µl of zinc perchlorate (1.5 mM) in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4) and 300 µM of different analytes such as nitrate, chloride, bromide, hydroxide, acetate, bicarbonate, glutathione, methionine, nitrite, phosphates, azide, fluoride, hydroxide, thiosulphate, cysteine (150 µM) and H₂S (30 µM) was taken in the titration. Excitation wavelength for the fluorescence titration is 530 nm and slit width was 10/10.

pH studies

105µl of ARS (0.5 mM) was diluted to 1860µl and followed by addition of 35µl of Zinc perchlorate solution prepared in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4). The titration was done with H₂S (27 µM) at various pH between (4-12) using fluorescence spectrophotometer. Excitation wavelength for the fluorescence titration is 530 nm and slit width was 10/10.

Fluorescence study of ARS-Zn(II) with $\rm H_2S$ in presence of Human Serum and BSA

Stock solution of **ARS** (0.5 mM) was prepared in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4). 120µL of this stock solution was diluted to 2 mL followed by addition of zinc perchlorate (30 µM) titrated against H₂S spiked Human serum and bovine serum albumin (6 - 30 µM). The emission intensity at 620 nm was monitored upon excitation at 530 nm. The titration was done with slit width 10/10.

Preparation of Bovine serum albumin (BSA) solution: The BSA solution was prepared by dissolving BSA (1.0 g) in 40 mL MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4) so that the final concentration will be 25 mg/ml. The above solution was taken for the fluorescence studies of probes.

Fluorescence study of ARS-Zn(II) with H₂S donor

For the fluorescence study, 100 μ l of ARS-Zn (60 μ M) was prepared in MeOH:PBS buffer (3/1,(v/v),10 mM, pH = 7.4, 0.04% THF used as a co-solvent) and diluted by adding 134 μ l of buffer and followed by addition of 60 μ l donor (0.001 M, 10 equivalent) and 6 μ l glutathione (0.1 M, 100 equivalent) and was monitor for 30 minutes at 35°C. The excitation wavelength was 530 nm and emission wavelength was 645/30 nm. The titration was done in 96 well- plate using multimode micro plate reader.

Temperature dependent Fluorescence study of ARS-Zn(II) with H₂S donor

For the fluorescence study , 100 μ l of ARS-Zn(II) (60 μ M) was diluted by adding 134 μ l of buffer and followed by addition of 60 μ l donor (0.001 M, 10 equivalent) and 6 μ l glutathione (0.1 M, 100 equivalent) and was monitor for 30 minutes at 30°C, 40 °C and 50°C. The excitation wavelength was 530 nm and emission wavelength was 645. The titration was done in 96 well-plate in multimode micro plate reader.

Detailed experimental procedure of cell imaging

Maintenance of C6 cell lines

Glial cell lines (C6) were cultured in RPMI-1640 growth medium supplemented with 10% (v/v) FBS (Fetal Bovine Serum), penicillin, streptomycin and gentamycin in CO₂ incubator at 37 °C, 5% CO₂ and 95% relative humidity in tissue culture flask. Medium was changed on a regular basis and cells were harvested at the log phase of growth for various analysis.

Confocal imaging

Confocal imaging was done according to the method given by Ramsay et al (1988). C6 glial cells (100,000 cells/mL) were incubated in 6 well plates having coverslip in each well for 24 hours and thereafter treated with different concentration of test compound for 20 minutes. Cells washed twice with chilled phosphate buffer saline (PBS), fixed with chilled 4% paraformaldehyde and then washed with chilled PBS and treated with DAPI (10 μ g/mL) for 30 minutes. Then washing of wells was done to remove the excessive dye. Coverslip were placed on slide over floromount. Finally, Slides were observed under Nikon Air Laser Scanning Confocal Microscope System.

C6 glial cells with ARS only

C6 glial cells were incubated with 30 μ M of the **ARS** in buffer for 20 minutes, then confocal microscope images were taken at $\lambda_{ex} = 543$ nm with 40X objective.

C6 glial cells with ARS-Zn(II) complex

ARS-Zn(II) complex was prepared insitu by following method: 6 µl of the stock solution of **ARS** (0.001 M) was added to 188 µl of buffer, to this 6 µl of Zinc perchlorate hexahydrate (0.001 M) was added and the solution allow to stand for 5 minutes. Final concentration of **ARS** was 30 µM and Zn(ClO₄)₂ was 30 µM. **ARS-Zn(II)** complex prepared by above procedure was added to the cell lines and incubated for 20 minutes at 37°C before recording confocal microscope images at $\lambda_{ex} = 543$ nm with 40X objective.

C6 glial cells with ARS-Zn(II) and Na₂S

C6 cell lines were incubated with 30 μ M of **ARS-Zn(II)** and then treated with 44 μ M of Na₂S for 20 minutes at 37°C. The confocal microscope Images were recorded at $\lambda ex = 543$ nm with 40X objective.

Calculation for the detection limit:

The detection limit was calculated using formula:

 $LOD = (3 \times SD)/slope = (3 \times 2.0)/6.52273 \times 10^7 = 92 \text{ nM}$

Where, LOD= Limit of detection and SD = Standard deviation of the blank



SI Figure 1: The UV/visible absorption of ARS $(30\mu M)$ in MeOH:PBS buffer (3/1,(v/v),10mM,pH = 7.4).



SI Figure 2: The emission spectrum of ARS (30μ M) in MeOH:PBS buffer (3/1,(v/v),10mM,pH = 7.4)



SI Figure 3: The emission titration profile of ARS with Zinc



SI Figure 3: Job plot for the binding of ARS with Zinc (II) (2:1 Complex formation). Where, X is the mole fraction.



SI Figure 4: Benesi-Hildebrand plot for the fluorescent titration of ARS with Zinc ion



SI Figure 5: Titration profile for the binding of ARS-Zn(II) with H_2S in MeOH-PBS buffer at RT



SI Figure 6: Fluorescent titration of ARS-Zn(II) with H₂S (0-30 μ M) spiked human serum (a) ARS-Zn (blank), (b) blank+ human serum without spiking H₂S, (c) 6 μ M H₂S spiked, (d) 12 μ M H₂S spiked (e) 18 μ M H₂S spiked (f) 24 μ M H₂S spiked (g) 30 μ M H₂S spiked in Human Serum.



SI Figure 7: Fluorescent titration of **ARS-Zn(II)** with H₂S (0-30 μ M) spiked BSA (1) ARS-Zn (blank), (2) blank+ BSA without spiking H₂S, (a) 6 μ M H₂S spiked BSA, (b) 12 μ M H₂S spiked BSA (c) 18 μ M H₂S spiked BSA(d) 24 μ M H₂S spiked BSA (e) 30 μ M H₂S spiked in BSA.



SI Figure 8 : (a) Blank lead acetate strip paper (b) 0.05 M Na₂S, (c) 0.005 M Na₂S, (d) 0.0005 M Na₂S, (e) 0.0005 M Na₂S. The black color shows the presence of H₂S species in the MeOH-PBS (3:1, v/v, pH 7.4).



SI Figure 9: Effect of methanol percentage in H_2S sensing of ARS-Zn(II) in Me-OH/PBS mixture.



SI Figure 10: Effect of different solvent in probe ARS-Zn(II) with of H₂S (44 μ M). Ratio of different solvent and PBS is (30/70, v/v)



SI Figure 11: Emission spectra **ARS-Zn(II)** with GSH (1mM) and Cysteine (1 mM) and Interference of GSH (1mM) and Cysteine (1 mM) with H_2S in MeOH:PBS buffer (3/1,(v/v),10mM, pH = 7.4)



SI Figure 12: Plot of spiked H_2S in Human serum and BSA versus change in emission intensity at 620 nm.



SI Figure 13: Temperature dependant study of **ARS-Zn(II)** (60 μ M) with H₂S donor (0.001 M, 10 equivalent) and glutathione (0.1 M, 100 equivalent) for 30 minutes at 30°C, 40°C and 50°C.



SI Figure 14: ESI-Mass spectrum of H_2S donor molecule benzoic (methyl carbonic) dithioperoxyanhydride.

¹ T. Roger, F. Raynaud, F. Bouillaud, C. Ransy, S. Simonet, C. Crespo, M.-P. Bourguignon, N. Villeneuve, J.-P. Vilaine, I. Artaud and E. Galardon, *ChemBioChem*, 2013, **14**, 2268-2271.