

The stability study of tubular DNA origami in the presence of protein crystallisation buffer

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1. Materials

All chemicals were obtained from Alfa Aesar (Tianjing, China) and used without further purification. All short oligo-DNA strands were purchased from Invitrogen. M13mp18 viral DNA was purchased from New England Biolabs. Water used in all experiments was Milli-Q deionized (18.2 M Ω .cm).

2. Method

Preparation of tubular DNA origami

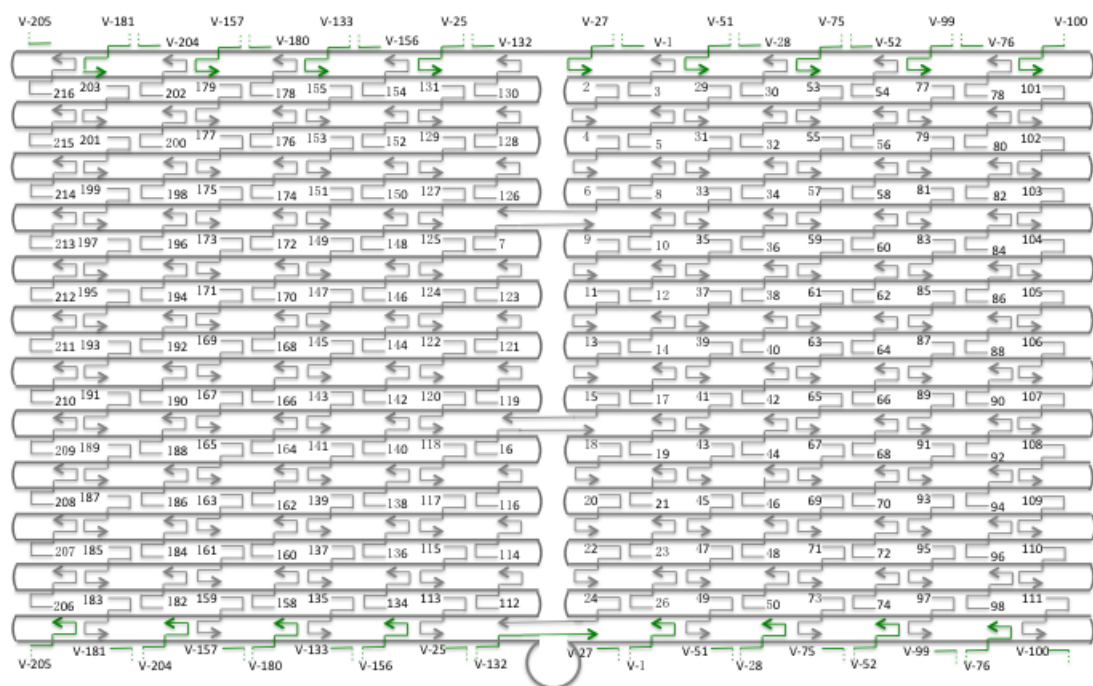
Detailed structures of tubular DNA origami used in this study are shown in Supplementary Fig. S1, and the sequences of the staple strands are listed at the end. DNA origami assembly was done by mixing scaffold and staples to a final concentration of 6.25 nM and 15.625 nM, respectively, in a 1 $\times$ TAE-Mg²⁺ solution buffer (40 mM Tris, pH 8.0, 2 mM EDTA, 12.5 mM Mg(OAc)₂). This mixture was cooled from 90 to 25 °C at a rate of – 1.0 °C /min using a PCR thermal cycler.

The assembled structures were purified from the excess staple strands by centrifugation with Millipore's 100 kD molecule-cutoff Centricon spin-filter in three cycles at a speed of 3,000 g for 10 min at 4 °C in the same 1 $\times$ TAE-Mg²⁺ buffer. The assembled origami structures were then collected at the end of the third cycle of filtration.

AFM measurements

For each measurement, 5 μ L of the sample was deposited onto a freshly cleaved mica surface and left to adsorb for 3 min. 30 μ L of 1 $\times$ TAE-Mg²⁺ buffer was added to the liquid cell and the sample was scanned under ScanAsyst model using a E scanner of AFM (Bruker Multimode 8). The probe used here was ScanAsyst Fluid+ (Olympus). All the images were only flattened by the AFM analysis program without other treatment.

a



b

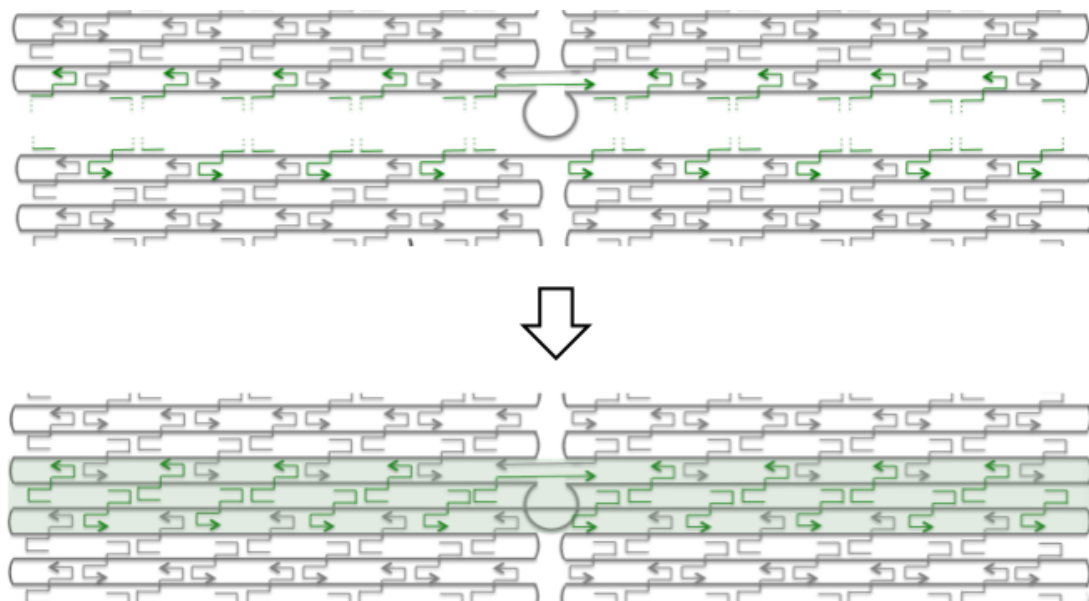


Fig. S1 (a) Design details of the tubular DNA origami with sticky ends between the top and bottom edges. The strand numbers are labeled at the 5' terminal. The green lines were the sticky ends. (b) Design principle of the edge connections during the formation of tubular DNA origami.

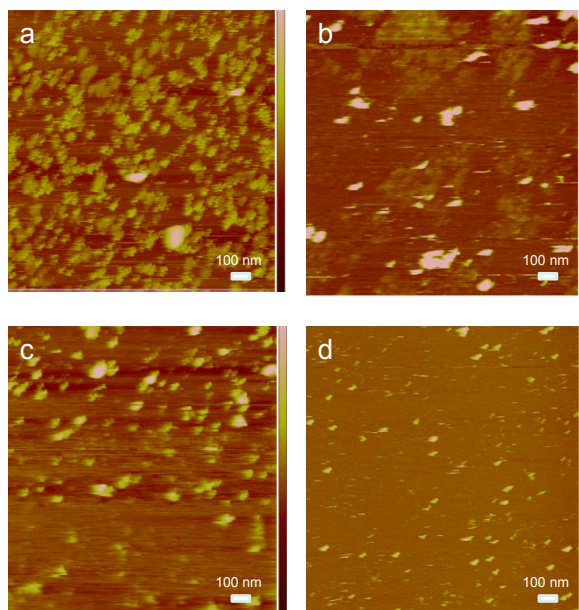


Fig. S2 AFM images of DNA origami assembled in protein crystallization cocktails of (a) lysozyme, (b) thaumatin, (c) human serum albumin, (d) catalase. All the samples are characterised after buffer exchange for at least 24 hrs.

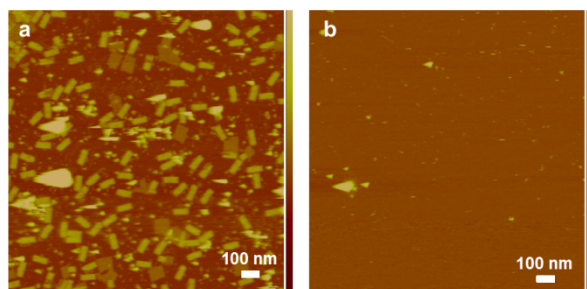


Fig. S3 AFM images of DNA origami after (a) gradient buffer exchange and (b) dialysis buffer exchange of catalase crystallization cocktails All the samples are characterised after buffer exchange for at least 24 hrs.

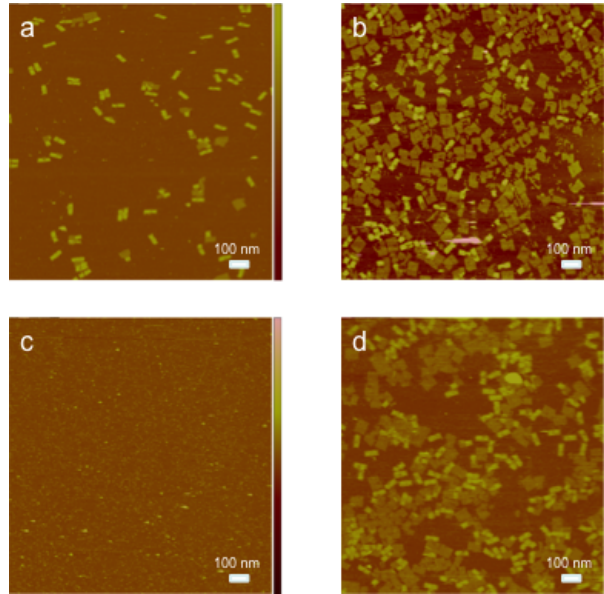


Fig. S4 AFM images of DNA origami in the DNA assembly buffer whose Mg^{2+} was replaced with 200 mM (a) Na^+ , (b) K^+ , (c) Ca^{2+} and (d) NH_4^+ , respectively.

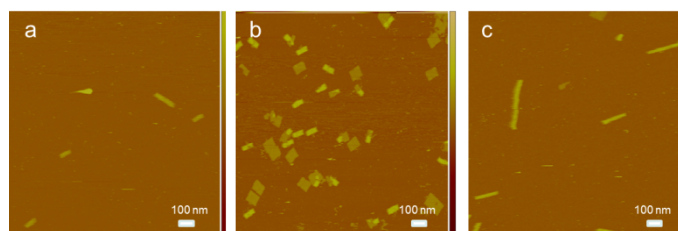


Fig. S5 AFM images of DNA origami in the DNA assembly buffer, whose 40 mM Tris was replaced with 100 mM (a) HEPES, (b) PEPES and (c) MES, respectively.

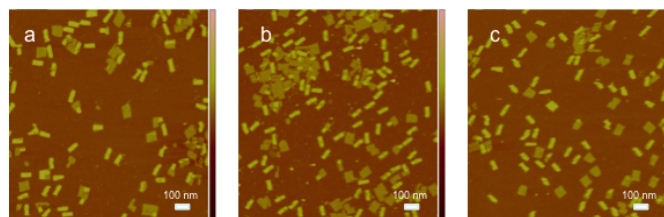


Fig. S6 AFM images of DNA origami with 10 % (v/v) (a) ethanol, (b) MPD, (c) glycerol in the DNA assembly buffer, respectively.

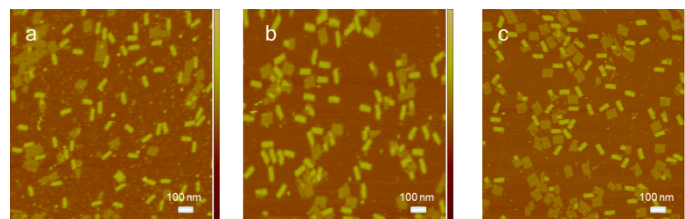


Fig. S7 AFM images of DNA origami with 30 % (w/v) (a) PEG₂₀₀₀, (b) PEG₄₀₀₀, (c) PEG₈₀₀₀ in the DNA assembly buffer, respectively.

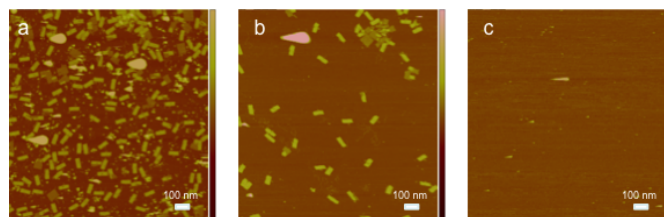


Fig. S8 AFM images of DNA origami with (a) 2 M NaCl, (b) 3 M NaCl, (c) 4 M NaCl in the DNA assembly buffer, respectively.

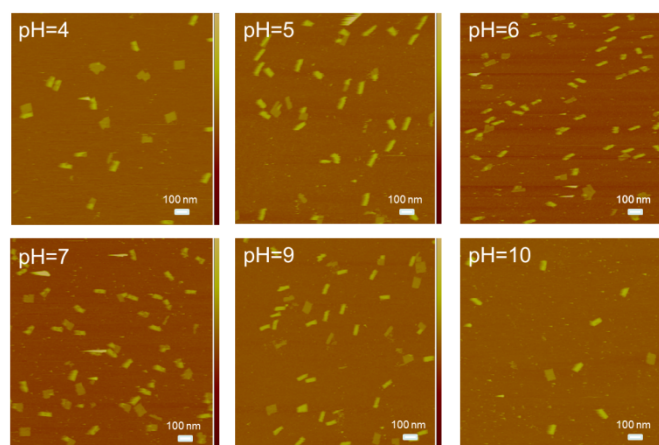


Fig. S9 AFM images of DNA origami assembled at different pH conditions.

Sequences

V-1 CGGCCTTGATAGGAACCCATGTACAAACAGTT	29 CTGAAACAGGTAATAAGTTTTAACCCCTCAGA
2 AATGCCCCGTAAACAGTGCCCGTATCTCCCTCA	30 AGTGTACTTGAAAGTATTAAGAGGCCGCCACC
3 TGCCTTGACTGCCTATTTTCGGAACAGGGATAG	31 GCCACCACTCTTTTCATAATCAAACCGTCACC
4 GAGCCGCCCCACCACCGGAACCGCGACGGAAA	32 GTTTGCCACCTCAGAGCCGCCACCGATACAGG
5 AACCAGAGACCCTCAGAACCGCCAGGGGTCAG	33 GACTTGAGAGACAAAAGGGCGACAAGTTACCA
6 TTATTCATAGGGAAGGTAAATATTCATTCACT	34 AGCGCCAACCATTTGGGAATTAGATTATTAGC
7 CATAACCCGAGGCATAGTAAGAGCTTTTAAAG	35 GAAGGAAAATAAGAGCAAGAAACAACAGCCAT
8 ATTGAGGGTAAAGGTGAATTATCAATCACCGG	36 GCCCAATACCGAGGAAACGCAATAGGTTTACC
9 AAAAGTAATATCTTACCGAAGCCCTTCCAGAG	37 ATTATTTAACCCAGCTACAATTTTCAAGAACG
10 GCAATAGCGCAGATAGCCGAACAATTCAACCG	38 TATTTTGCTCCCAATCCAAATAAGTGAGTTAA
11 CCTAATTTACGCTAACGAGCGTCTAATCAATA	39 GGTATTAAGAACAAGAAAAATAATTAAAGCCA
12 TCTTACCAGCCAGTTACAAAATAAATGAAATA	40 TAAGTCCTACCAAGTACCGCACTCTTAGTTGC
13 ATCGGCTGCGAGCATGTAGAAACCTATCATAT	41 ACGCTCAAAATAAGAATAAACACCGTGAATTT
14 CTAATTTATCTTTCCTTATCATTTCATCCTGAA	42 AGGCGTTACAGTAGGGCTTAATTGACAATAGA
15 GCGTTATAGAAAAAGCCTGTTTAGAAGGCCGG	43 ATCAAAATCGTCGCTATTAATTAACGGATTCTG
16 GCTCATTTTCGCATTAAATTTTTGAGCTTAGA	44 CTGTAAATCATAGGTCTGAGAGACGATAAATA
17 AATTACTACAAATCTTACCAGTAATCCCATC	45 CTGATTGAAAGAAATTGCGTAGACCCGAACG
18 TTAAGACGTTGAAAACATAGCGATAACAGTAC	46 ACAGAAATCTTTGAATACCAAGTTCCTTGCTT
19 TAGAATCCCTGAGAAGAGTCAATAGGAATCAT	47 TTATTAATGCCGTCATAGATAATCAGAGGTG
20 CTTTACACAGATGAATATACAGTAAACAATT	48 AGATTAGATTTAAAAGTTTGAGTACACGTAAA
21 TTTAACGTTCTGGGAGAAACAATAATTTTCCCT	49 AGGCGGTCATTAGTCTTTAATGCGCAATATTA
22 CGACAATAAGTATTAGACTTTACAATACCGA	50 GAATGGCTAGTATTAACACCGCCTCAACTAAT
23 GGATTTAGCGTATTAAATCCTTTGTTTCAGG	V-51 CTCAGAGCCATTGCAACAGGAAAAATATTTT
24 ACGAACCAAAACATCGCCATTAAA TGGTGGTT	V-52 GGAAATACACCGCCACCCTCAGAACTGAGACT
V-25 TGAGTTTCCGAGAAAGGAAGGGAACAACTAT	53 CCTCAAGAATACATGGCTTTTGATAGAACCCAC
26 TAGCCCTACCAGCAGAAGATAAAAACATTTGA	54 TAAGCGTCGAAGGATTAGGATTAGTACCGCCA
V-27 CAAGCCCACTGGTAATATCCAGAACGAAGTGA	55 CACCAGAGTTCGGTCATAGCCCCGCCAGCAA
V-28 CCGCCAGCCACCACCCTCATTTTCCTATTATT	56 TCGGCATTCCGCCGCCAGCATTGACGTTCCAG

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138 GGCGATCGCACTCCAGCCAGCTTTGCCATCAA

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