## **Supporting Information**

- 2 Highly efficient colorimetric detection of ATP utilizing split
- 3 aptamer target binding strategy and superior catalytic

## 4 activity of graphene oxide-platinum/gold nanoparticles

- 5 Siqi Zhang<sup>a</sup>, Kun Wang<sup>a, b</sup>, Jiali Li<sup>a</sup>, Zhenyu Li<sup>a</sup>, Ting Sun<sup>a\*</sup>
- 6 a College of Sciences, Northeastern University, Shenyang, 110819, China
- 7 <sup>b</sup> Department of Chemistry and Environmental Engineering, Changchun University of
- 8 Science and Technology, Changchun, China
- 9 \*Corresponding author
- 10 E-mail: sun1th@163.com
- 11 Tel.: +86-024-83684786;
- 12

1

13 Preparation of Aptamer-1 Modified Magnetic Beads.

The Fe<sub>3</sub>O<sub>4</sub> nanoparticles with the size of 200 nm were synthesized according to solvothermal method<sup>1</sup>. FeCl<sub>3</sub>•6H<sub>2</sub>O (0.54 g), Na acrylate (1.5 g), NaOAc (1.5 g), were dissolved in a mixture of ethylene glycol (20 mL) under magnetic stirring. The obtained mixture solution was transferred into a Teflon stainless-steel autoclave and heated at 200°C for 10 h. When the autoclave was cooled down to room temperature, the black products were washed four times with ethanol and ultrapure water, respectively. Then the magnetic Fe<sub>3</sub>O<sub>4</sub> microspheres were dispersed in a mixture of 1 ethanol (80 mL) and ultrapure water (20 mL) under ultrasonication for 5 min. 2 Ammonia (1 mL) and TEOS (200  $\mu$ L) was added into the mixture solution under 3 mechanical stirring for 12 h. After being separated, the product mixed with APTES 4 (100  $\mu$ L) and ammonia (2 mL) under mechanical stirring for another 12 h. Finally, the 5 Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub>-NH<sub>3</sub> was magnetic separated three times and re-dispersed in 5 mL 6 ethanol/H<sub>2</sub>O solution. The diameter of the Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub>-NH<sub>3</sub> was about 200 nm 7 evaluated by TEM measurements.

The aptamer-1 functionalized  $Fe_3O_4(a)SiO_2$  was prepared as follows. 8 9 Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub>-NH<sub>3</sub> in 100 µL ethanol/H<sub>2</sub>O solution was first mixed with 20 µL Tween 20 (5% in ethanol solution) at room temperature. Then carboxyl modified aptamer-1 10 (final concentration of oligonucleotides 10 µM) in 400 µL 10 mM MES buffer with 11 200 mM EDC and 50 mM NHS was added into the Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub> suspension. The 12 mixture solution was mechanical stirred at 37 °C for 16 hours. To remove excess 13 aptamer, these aptamer-functionalized Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub> were magnetic separated and 14 washed three times using 10 mM Tris-HCl buffer containing 5 mM MgCl<sub>2</sub> and 15 15 16 mM KCl (pH=8.0). The functionalized Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub> nanoparticles were stored in the 17 buffer solution.



2 Scheme S1. (A) Schematic illustration of the preparation of aptamer-1 modified
3 magnetic beads. (B) Schematic illustration of the preparation of aptamer-2 modified
4 graphene oxide/platinum/gold nanoparticle hybrids

5

## 6 2.5 Preparation of PDDA-Functionalized Graphene Oxide.

The graphene oxide was prepared from graphite powder according to the previous work<sup>2</sup>. Then 80 mg of PVP was upon addition of 20 mL of 0.25 mg/mL obtained graphene oxide solution. After magnetic stirring 30 min, the dispersion was centrifuged separation three times and re-dispersed in 5 mL ultrapure water. The PDDA-functionalized graphene oxide was prepared by mixing 0.1 mL of 20 wt % PDDA with 16.8 mL of 0.625 M KCl, followed by adding 4.2 mL of PVP-coated
 graphene oxide. After sonication for 90 min, the PDDA/GO dispersion solution was
 centrifuged separation three times and re-dispersed in 5 mL ultrapure water.

4 2.6 Preparation of Aptamer-2 Modified Graphene Oxide/Platinum/Gold
5 Nanoparticle Hybrids.

The Platinum/gold nanoparticle hybrids were synthesized with citrate reduction according to previous work <sup>3</sup>. A 100 mL sample of aqueous (0.9 mM) HAuCl<sub>4</sub> and  $K_2$ PtCl<sub>6</sub> (0.1 mM) was prepared in a 250 mL round-bottom flask. The solution was continuously heated to a boil while stirring, and 40 mM trisodium citrate solution (10 mL) was added. The boiling and stirring was continued for an additional 30 min after the final color changing from pale yellow to deep red.

12 1 mL PDDA/GO dispersion solution (1 mg/mL) was added into 10 mL of 13 prepared PtAu<sub>NPs</sub> under magnetic stirring. Then the dispersion solution was sonicated 14 for 10 min before overnight aging. The obtained product was washed three times and 15 dispersed in 2 mL of water.

The aptamer-2 functionalized GO/PtAu<sub>NPs</sub> was prepared by mixing GO/PtAu<sub>NPs</sub> (2 mL) with thiol-aptamer (final concentration of oligonucleotides 10  $\mu$ M) in PBS buffer (0.3 M NaCl, 0.2 PBS, pH=7) under continuous shaking for 16 hours<sup>4</sup>. To remove excess thiol-aptamer, these aptamer-2 functionalized GO / PtAu<sub>NPs</sub> were centrifuging for 45 min at 13000 rpm three times using 10 mM Tris-HCl buffer containing 5 mM MgCl<sub>2</sub> and 15 mM KCl (pH=8.0). The functionalized GO / PtAu<sub>NPs</sub>

| 2  | shortly named as GO/PtAu <sub>NPs</sub> /Aptamer-2. |
|----|---|
| 3  |   |
| 4  |   |
| 5  |   |
| 6  |   |
| 7  |   |
| 8  |   |
| 9  |   |
| 10 |   |
| 11 |   |
| 12 |   |
| 13 |   |
| 14 |   |
| 15 |   |
| 16 |   |
| 17 |   |
| 18 |   |

1 nanoparticles were stored in the buffer solution. The synthesized nanomaterials were



2 Fig. S1 (A) UV-Vis absorption spectra of (a) GO (10 μg/mL), (b) GO@PtAu (10
3 μg/mL), and (c) GO@PtAu@aptamer-2 solutions (10 μg/mL). (B) UV-Vis absorption
4 spectra of (a) Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub> (0.06 mg/mL) and (b) Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub>@aptamer-1 solutions
5 (0.06 mg/mL).

1

As shown in Figure S1 (supporting information), the functionalized GO/PtAu<sub>NPs</sub> and magnetic bead were characterized by measuring the UV absorbance spectra after removing excess DNA. Except for the feature peaks of GO at 270 nm and PtAu at 536 nm, a new and higher peak at 260 nm was detected for the GO/PtAu<sub>NPs</sub>/aptamer-2 (curve c in Figure S1A), indicating the successful anchoring of aptamer-2 on the surface of GO/PtAu<sub>NPs</sub>. A similar peak at 260 nm was observed for aptamer-1-modified magnetic beads (curve b in Figure S1B).

The absorbance value of the functionalized nanoparticles was used to estimate the amount of aptamers after deducting the absorbance value of naked nanoparticles at According to oligonucleotide synthesis reports, the reciprocal of molar extinction coefficients for aptamer-1 and aptamer-2 are 7.0 and 3.9 nmol/OD, respectively. The amount of aptamer-1 on the magnetic beads was 10.94 nmol/mg while the amount of aptamer-2 on the GO/PtAu<sub>NPs</sub> was 57 nmol/mg according to
 simple calculations.



5 Fig. S2. Peroxidase-like activity of (a)GO, (b) naked GO@PtAu and (c) the aptamer-2





2 Fig. S3. Linear relationship between absorbance value of the oxidized TMB product
3 at 652 nm and the concentration of GO/PtAu<sub>NPs</sub>. The error bars represent the standard
4 deviation of three independent measurements.

Figure S3 provides the linear relationship between absorbance value of the
oxidized TMB product at 652 nm and the concentration of GO/PtAu<sub>NPs</sub>. This research
shows that with the increasing concentration of the superior catalyst, the catalytic rate
of TMB is enhanced relatively.

- S. H. Xuan, Y. X. J. Wang, J. C. Yu and K. C. F. Leung, *Chemistry of Materials*, 2009, 21,
   5079-5087.
- 3 2. W. Ren, Y. X. Fang and E. K. Wang, Acs Nano, 2011, 5, 6425-6433.
- 4 3. C.-W. Tseng, H.-Y. Chang, J.-Y. Chang and C.-C. Huang, *Nanoscale*, 2012, 4, 6823-6830.
- 5 4. C. E. Chen, C. Q. Zhao, X. J. Yang, J. S. Ren and X. G. Qu, Advanced Materials, 2010, 22,
- 6 389-393.
- 7