

## Supporting Information

### A Highly Sensitive Hemicyanine-Based Fluorescent Chemodosimeter for Mercury Ions in Aqueous Solution and Living Cells

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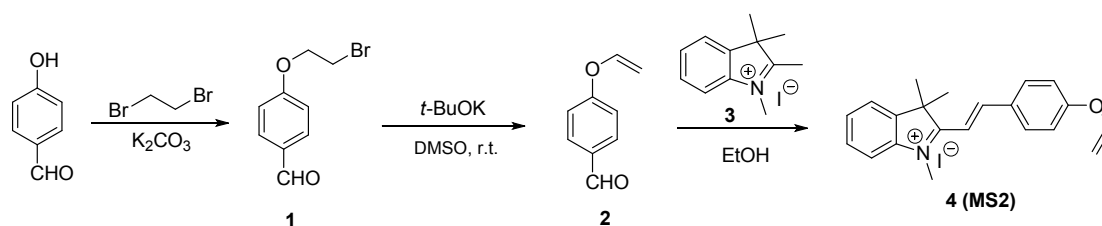
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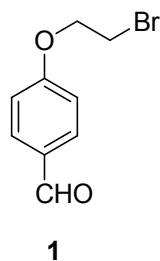
## Materials and methods

All the solvents were of analytic grade. NMR experiments were carried out on a Bruker AV-400 NMR spectrometer with chemical shifts reported in ppm (in CDCl<sub>3</sub>, D<sub>2</sub>O, CD<sub>3</sub>OD, or TMS as an internal standard). Mass spectra were measured on an Agilent 1290 LC-MS spectrometer. All pH measurements were made with a Sartorius basic pH-Meter PB-10. Fluorescence spectra were determined on a PerkinElmer LS55 Fluorescence spectrophotometer. Absorption spectra were collected on a Shimadzu UV 2501(PC)S UV-Visible spectrophotometer. The excitation and emission widths for measurements were all 5 nm. All the cation solutions were prepared from AgNO<sub>3</sub>, CaCl<sub>2</sub>, CdCl<sub>2</sub>, CoCl<sub>2</sub>, CrCl<sub>3</sub>, CuCl<sub>2</sub>, FeCl<sub>2</sub>, FeCl<sub>3</sub>, KCl, LiCl, MgCl<sub>2</sub>, MnCl<sub>2</sub>, NaCl, NiCl<sub>2</sub>, Pb(OAc)<sub>2</sub>, ZnCl<sub>2</sub>, and HgCl<sub>2</sub> in distilled water, with a concentration of 1 mM, respectively. The excitation and emission widths for **MS2** were all 10/5.

## Synthesis and characterization

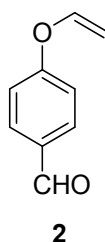


**Scheme S1** Synthesis of **MS2**.

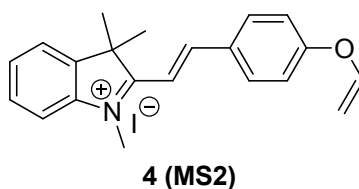


**4-(2-bromoethoxy)benzaldehyde (1)**: 4-hydroxybenzaldehyde (10 g, 0.082 mol) and potassium carbonate (19 g, 0.138 mol) were dissolved in acetone (120 mL) under nitrogen atmosphere, then 1, 2-dibromoethane (41 mL, 0.475 mol) was added and the solution was refluxed for 10 h until all starting material got consumed which was monitored by TLC analysis. The reaction mixture was

washed with water (500 mL), extracted with ether acetate (3 × 150 mL). The extract was dried over sodium sulfate and then concentrated under vacuum. The product was purified by flash chromatography using petroleum ether/ethyl acetate (8:1, v/v) as eluant to give **1** (11.6 g, 62%) as a pale yellow gum; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.82 (s, 1H), 7.78 (d, *J* = 8.2 Hz, 2H), 6.95 (d, *J* = 8.2 Hz, 2H), 4.30 (t, *J* = 5.9 Hz, 2H), 3.61 (t, *J* = 5.8 Hz, 2H).



**4-(vinylloxy)benzaldehyde (2):** Potassium *tert*-butoxide (6.5 g, 57.8 mmol) was dissolved in anhydrous DMSO (75 mL) and then added dropwise into a 200 mL bottom flask containing a DMSO solution (50 mL) of **2** (11.0 g, 48.2 mmol) under nitrogen atmosphere and the mixture was stirred overnight at room temperature. Water (500 mL) was then added to the reaction solution. The mixture was extracted with ether acetate (3 × 150 mL), dried over sodium sulfate and then concentrated under vacuum. The product was purified by flash chromatography using petroleum ether/ethyl acetate (10:1, v/v) as eluant to give **2** (2.6 g, 37%) as a pale yellow solid; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 9.86 (s, 1H), 7.81 (d, *J* = 8.0 Hz, 2H), 7.05 (d, *J* = 8.0 Hz, 2H), 6.63 (dd, *J* = 13.6, 5.6 Hz, 1H), 4.89 (d, *J* = 13.6 Hz, 1H), 4.57 (d, *J* = 5.6 Hz, 1H).



**MS2 (4):** The mixture of **2** (1.0 g, 6.8 mmol) and 1,2,3,3-tetramethyl-3H-indol-1-ium iodide **3**<sup>1</sup> (1.8 g, 6.1 mmol) of dry ethanol (100 mL) solution was refluxed under nitrogen atmosphere for 12 h. The solvent was then evaporated in vacuo. The crude product was purified by flash chromatography using dichloromethane/methanol (20:1, v/v) as eluant to give **4** (2.3 g, 87%) as a

white solid;

$R_f = 0.48$  (DCM/MeOH 10:1);

M.p. = 70-72 °C. (Decomposed);

$^1\text{H}$  NMR (400 MHz, MeOD)  $\delta$  8.52 (m, 1H), 8.30 (d,  $J = 8.6$  Hz, 2H), 7.92 (m, 2H), 7.75 (m, 2H), 7.33 (d,  $J = 8.6$  Hz, 2H), 7.05 (dd,  $J = 13.6, 6.0$  Hz, 1H), 5.05 (d,  $J = 13.6$  Hz, 1H), 4.77 (d,  $J = 6.0$  Hz, 1H), 4.34 (s, 3H), 2.00 (s, 6H);

$^{13}\text{C}$  NMR (100 MHz, MeOD)  $\delta$  184.0, 162.7, 154.9, 147.9, 144.9, 143.4, 134.3, 131.0, 130.7, 124.1, 118.3, 116.2, 112.5, 98.5, 54.0, 26.8;

HR-MS (TOF-ESI): *Calcd.* for  $([\text{M-I}]^+)$ , 304.1696; Found, 304.1744.

## Photophysical properties of MS2

**Table S1** Photophysical properties of the probe.

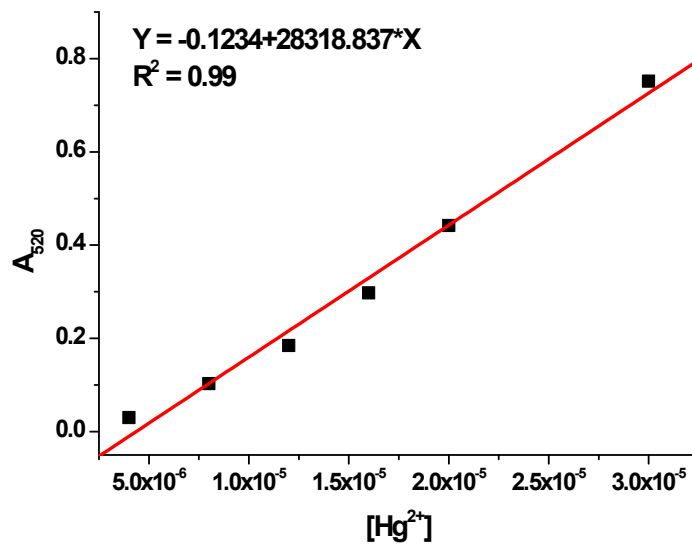
entry	$\lambda_{\text{ab}}$ (nm)	$\lambda_{\text{em}}$ (nm)	$\Phi^{\text{a}}$	$\epsilon / \text{M}^{-1} \text{cm}^{-1}$
<b>MS2</b>	400	551	0.004	31007
<b>MS2+Hg<sup>2+</sup></b>	520	551	0.018 <sup>b</sup>	42067

(a) The quantum yield ( $\Phi$ ) of **MS2** and **MS2-Hg<sup>2+</sup>** system were determined according to the literature.<sup>2</sup> (b)  $\Phi$  was determined in the present of 3.0 equiv. of Hg<sup>2+</sup>.

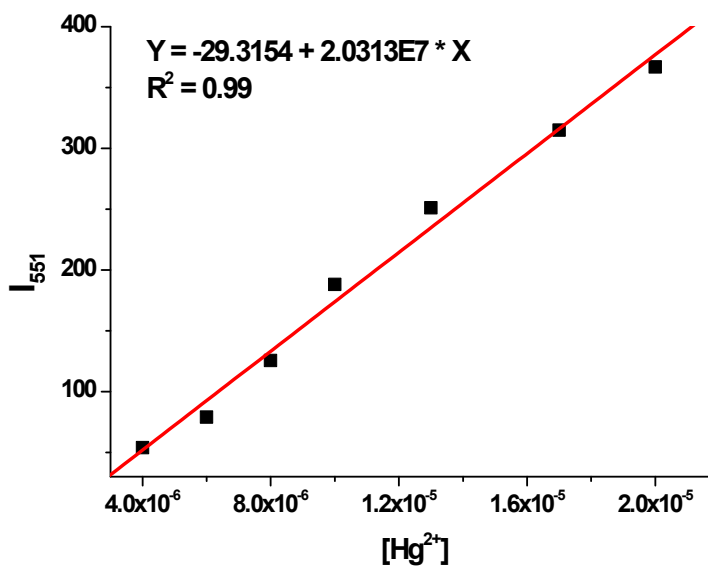
$$\Phi_{\text{Sample}} = \frac{\Phi_{\text{QS}} \cdot A_{\text{QS}} \cdot F_{\text{Sample}} \cdot \lambda_{\text{exQS}} \cdot \eta_{\text{Sample}}^2}{A_{\text{Sample}} \cdot F_{\text{QS}} \cdot \lambda_{\text{exSample}} \cdot \eta_{\text{QS}}^2}$$

Where  $\Phi$  is quantum yield; A is absorbance at the excitation wavelength; F is integrated area under the corrected emission spectra;  $\lambda_{\text{ex}}$  is the excitation wavelength;  $\eta$  is the refractive index of the solution; the Sample and QS refer to the sample and the standard, respectively. We chose Rhodamine 6G in EtOH as standard, which has the quantum yield of 0.95.<sup>3</sup>

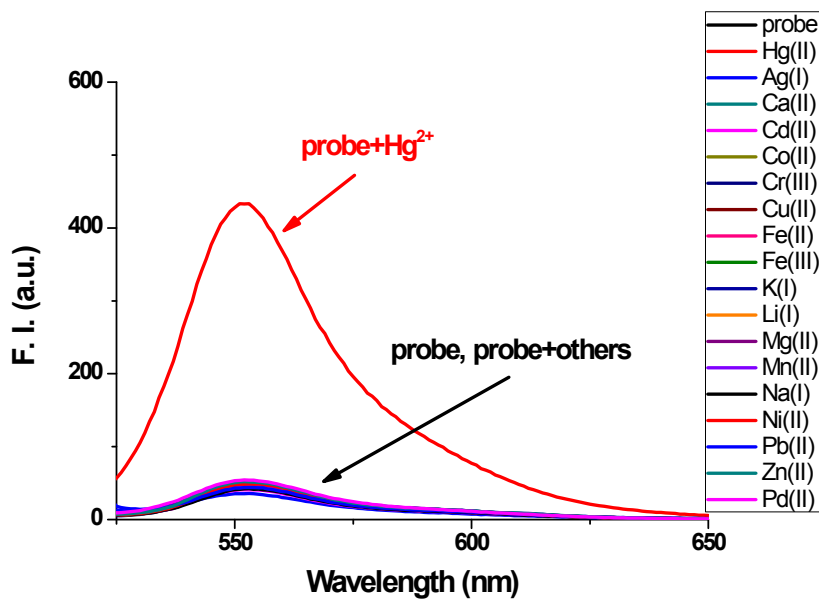
## Additional spectroscopic data



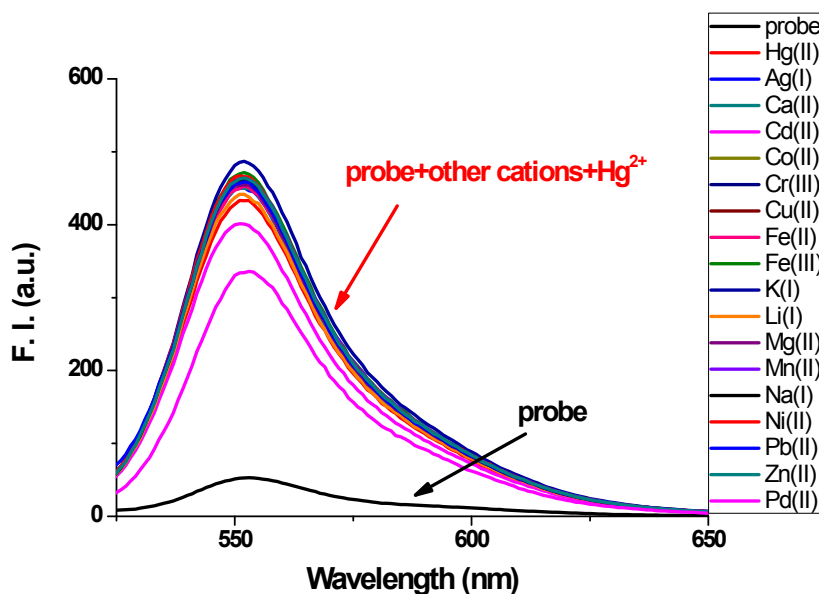
**Fig. S1** UV-vis absorption of MS2 (20.0  $\mu\text{M}$ ) at 520 nm as a function of  $\text{Hg}^{2+}$  concentration (0-30.0  $\mu\text{M}$ ) in PBS buffer solution (10 mM, pH 7.4, containing 1%  $\text{CH}_3\text{CN}$ ).



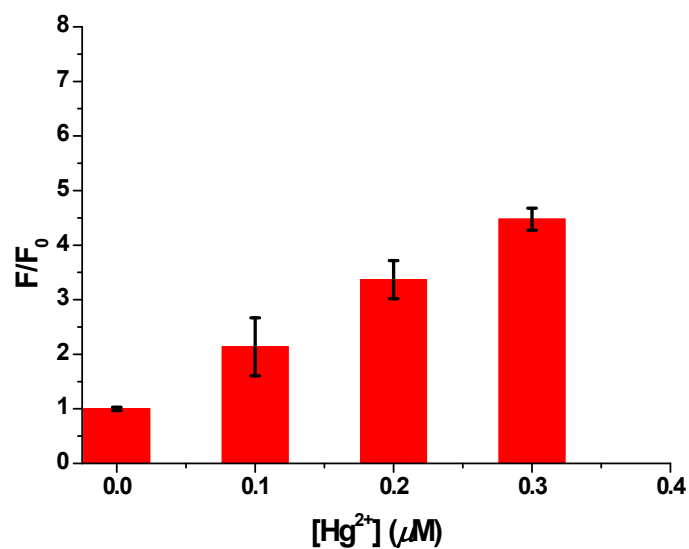
**Fig. S2** Fluorescent intensity of MS2 at 551 nm as a function of  $\text{Hg}^{2+}$  concentration (0-20.0  $\mu\text{M}$ ) under the same condition as the  $\text{Hg}^{2+}$  titration.



**Fig. S3** Fluorescence responses of MS2 (10.0  $\mu\text{M}$ ) with 3.0 equiv. of metal ions in PBS buffer solution (10 mM, pH 7.4, containing 1%  $\text{CH}_3\text{CN}$ ). Metal ions include  $\text{Hg}^{2+}$ ,  $\text{Ag}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{K}^+$ ,  $\text{Li}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Zn}^{2+}$ , and  $\text{Pd}^{2+}$ , ( $\lambda_{\text{ex}} = 510 \text{ nm}$ ).



**Fig. S4** Fluorescence responses of MS2 (10.0  $\mu\text{M}$ ) in the presence of 3.0 equiv. of metal ions ( $\text{Ag}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{K}^+$ ,  $\text{Li}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Zn}^{2+}$ , and  $\text{Pd}^{2+}$ ) in PBS buffer solution (10 mM, pH 7.4, containing 1%  $\text{CH}_3\text{CN}$ ), followed by 3.0 equiv. of  $\text{Hg}^{2+}$  ( $\lambda_{\text{ex}} = 510 \text{ nm}$ ).

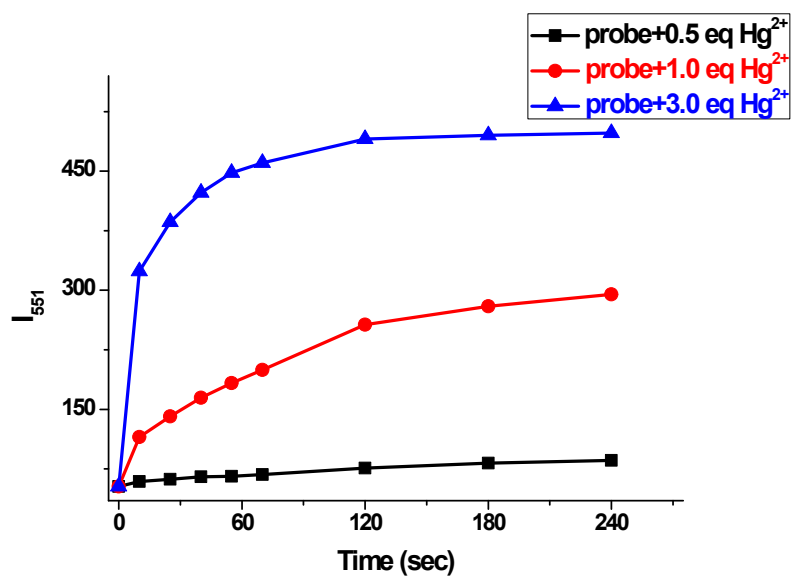


**Fig. S5** Detection limit of **MS2** determination. **MS2** (0.5 μM) in PBS (10 mM, pH 7.4) was treated with various concentrations of Hg<sup>2+</sup> and the fluorescence intensity at emission maxima was recorded and the detection limit was determined to be the concentration of Hg<sup>2+</sup> that induced 3-fold fluorescence increase. Experiments were done in triplate.

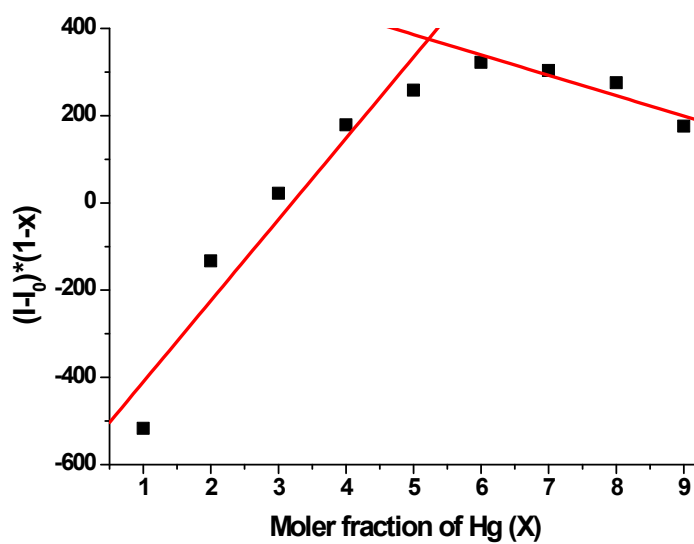


**Fig. S6** The photos of the corresponding solutions in Fig. 6 (from left to right including **MS2** only, and **MS2** with Hg<sup>2+</sup>, Ag<sup>+</sup>, Ca<sup>2+</sup>, Cd<sup>2+</sup>, Co<sup>2+</sup>, Cr<sup>3+</sup>, Cu<sup>2+</sup>, Fe<sup>2+</sup>, Fe<sup>3+</sup>, K<sup>+</sup>, Li<sup>+</sup>, Mg<sup>2+</sup>, Mn<sup>2+</sup>, Na<sup>+</sup>, Ni<sup>2+</sup>, Pb<sup>2+</sup>, Zn<sup>2+</sup>, and Pd<sup>2+</sup>, respectively).

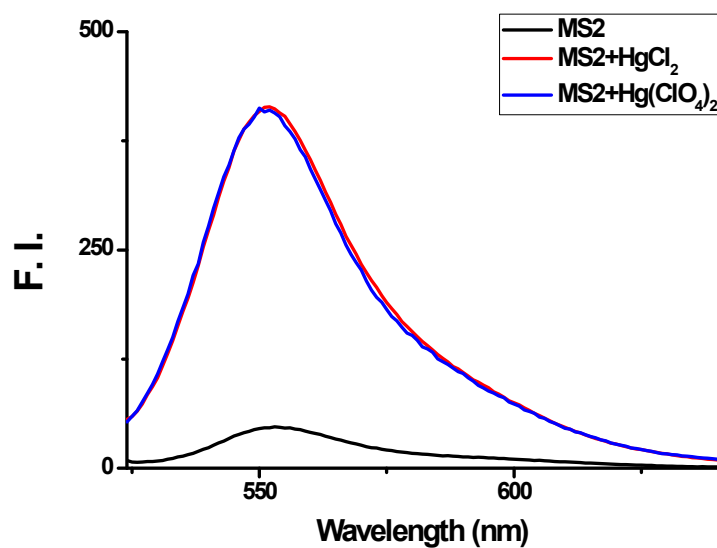




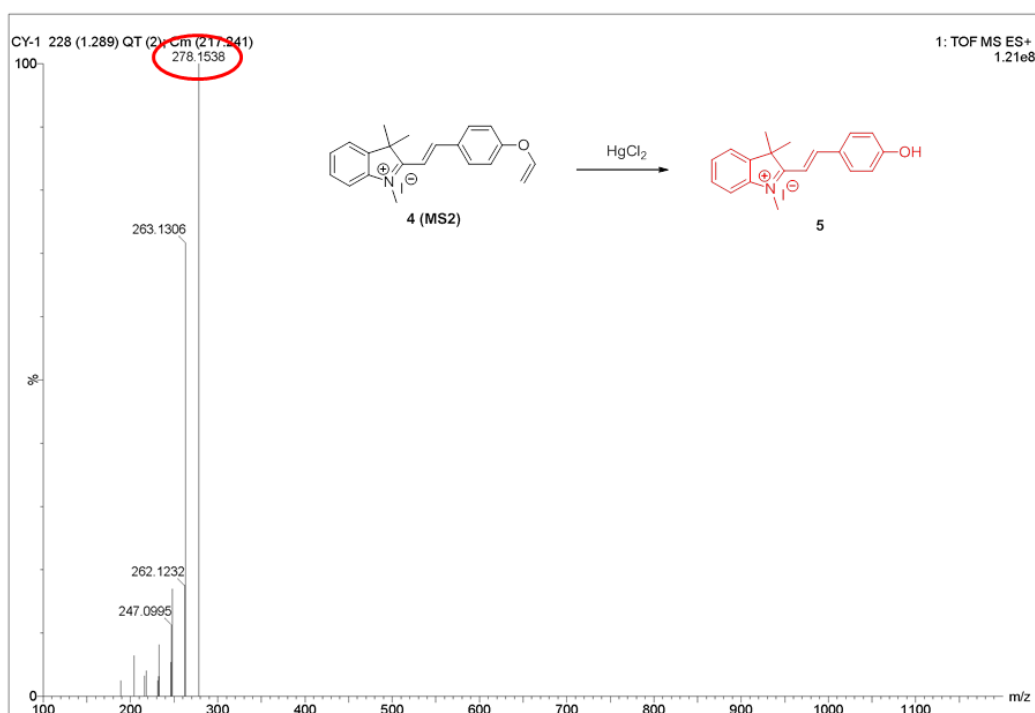
**Fig. S7** Time-dependent fluorescence intensity changes of MS2 (10.0  $\mu\text{M}$ ) upon addition of  $\text{Hg}^{2+}$  (0.5, 1.0, and 3.0 equiv., respectively) in PBS buffer solution (10 mM, pH = 7.4, containing 1%  $\text{CH}_3\text{CN}$ ) ( $\lambda_{\text{ex}} = 510 \text{ nm}$ ).



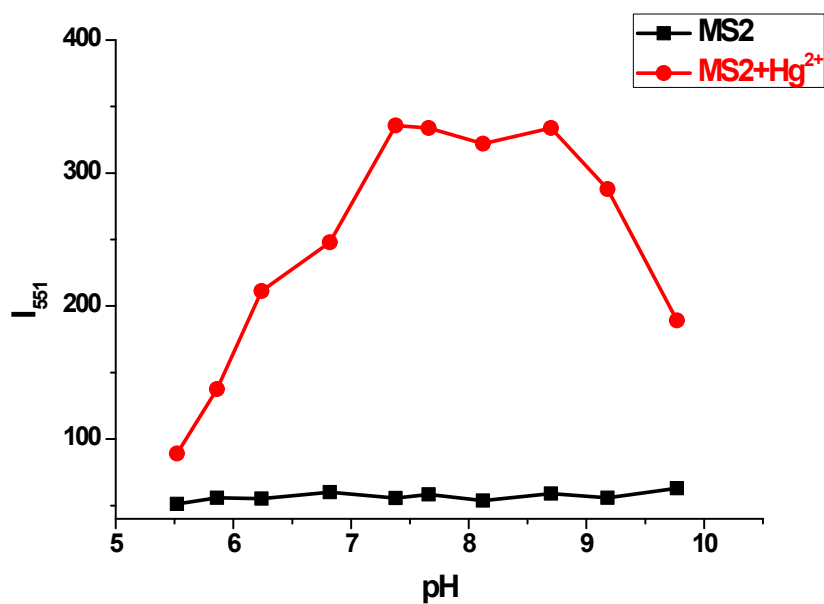
**Fig. S8** The Job's plot of MS2, the total concentration of the sensor and  $\text{Hg}^{2+}$  is 20.0  $\mu\text{M}$  (10 mM PBS buffer solution, pH 7.4, containing 1%  $\text{CH}_3\text{CN}$ ,  $\lambda_{\text{ex}} = 510 \text{ nm}$ ).



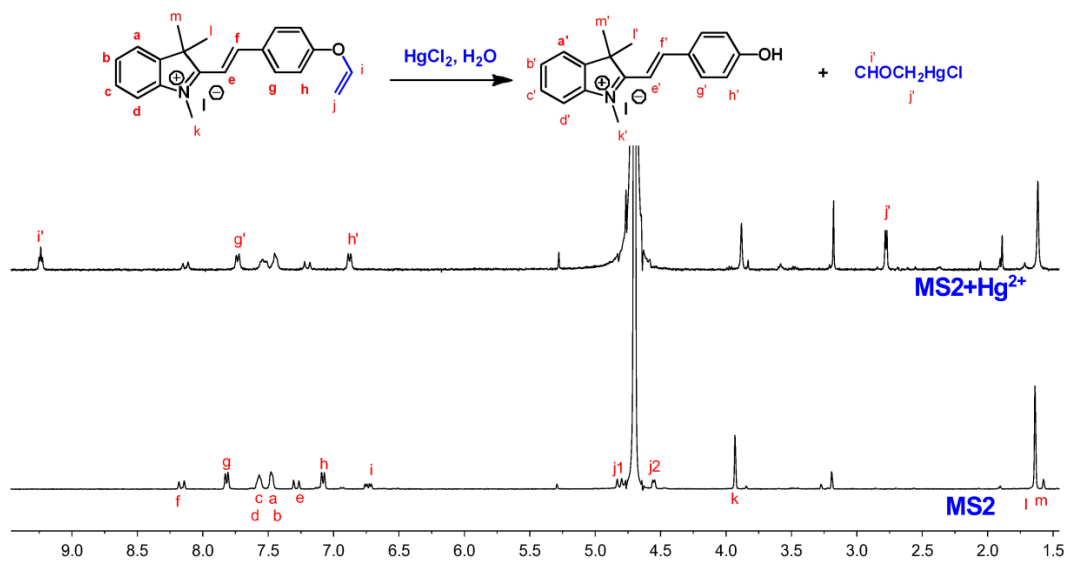
**Fig. S9** The experiments between MS2 (10.0  $\mu$ M) with different source of Hg<sup>2+</sup> (3.0 equiv. HgCl<sub>2</sub> and Hg(ClO<sub>4</sub>)<sub>2</sub>, respectively) in 10 mM PBS buffer solution, pH 7.4, containing 1% CH<sub>3</sub>CN,  $\lambda_{\text{ex}}$  = 510 nm.



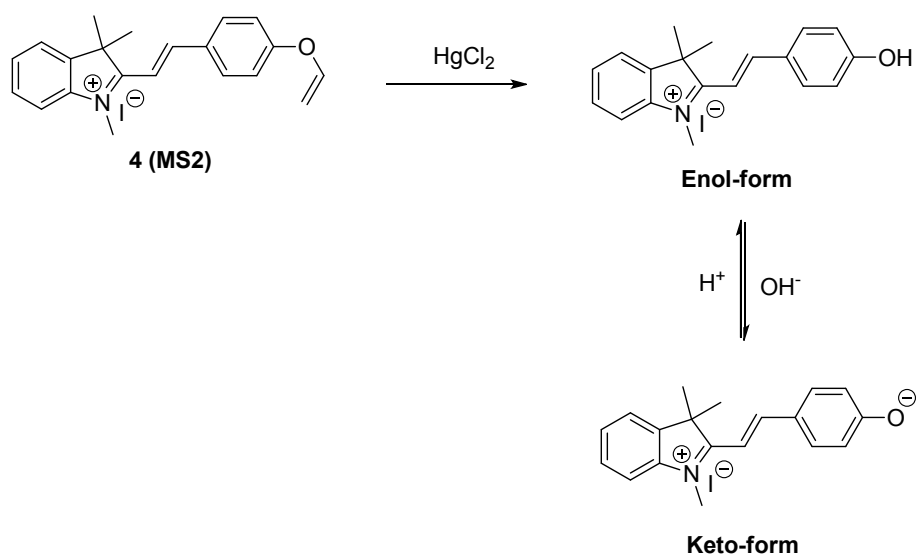
**Fig. S10** HPLC-MS (TOF-ESI) spectrum of MS2-Hg<sup>2+</sup> system shows the free hemicyanine moiety (5). [5-I]<sup>+</sup> = 278.1538 (*calcd.* For 278.1539) (*t* = 1.289 min).



**Fig. S11** Effect of the pH on the fluorescence emission of **MS2** (10.0  $\mu\text{M}$ ) alone and **MS2** (10.0  $\mu\text{M}$ ) reacted with  $\text{Hg}^{2+}$  (2.0 equiv.) ( $\lambda_{\text{ex}} = 510 \text{ nm}$ ).



**Fig. S12** <sup>1</sup>H NMR spectra of **MS2** in the absence and presence of  $\text{Hg}^{2+}$  (1.0 equiv. in  $\text{D}_2\text{O}$ ).



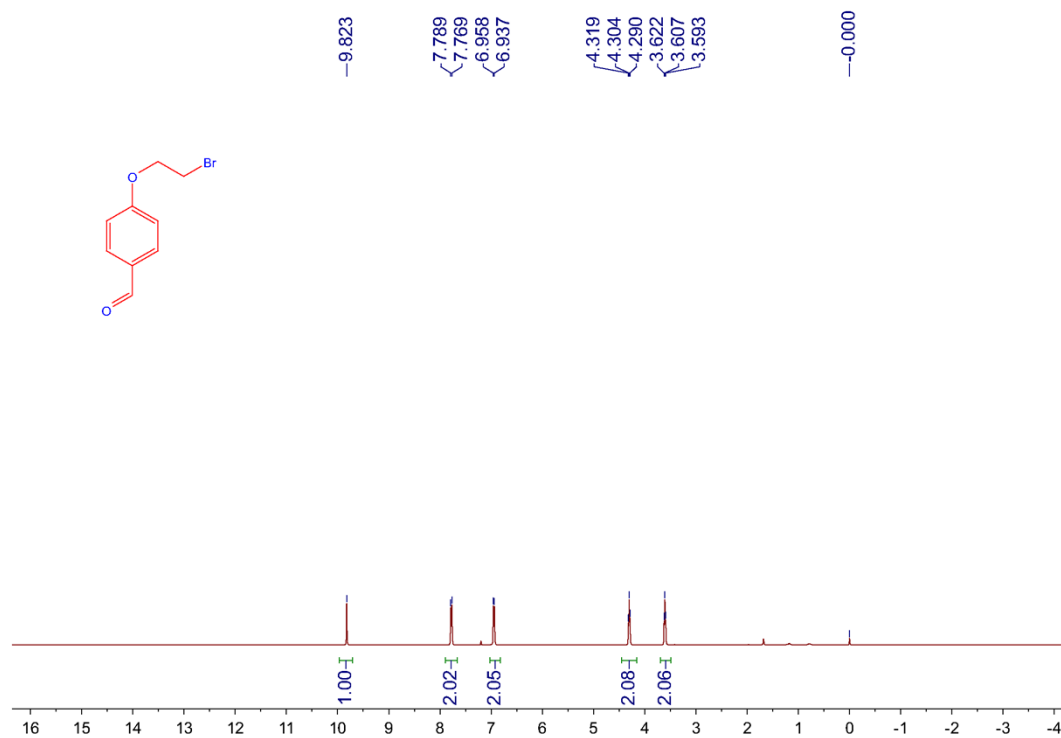
**Scheme S2** Enol-form and Keto-form of free hemicyanine fluorophore.

### Cell lines and imaging experiments

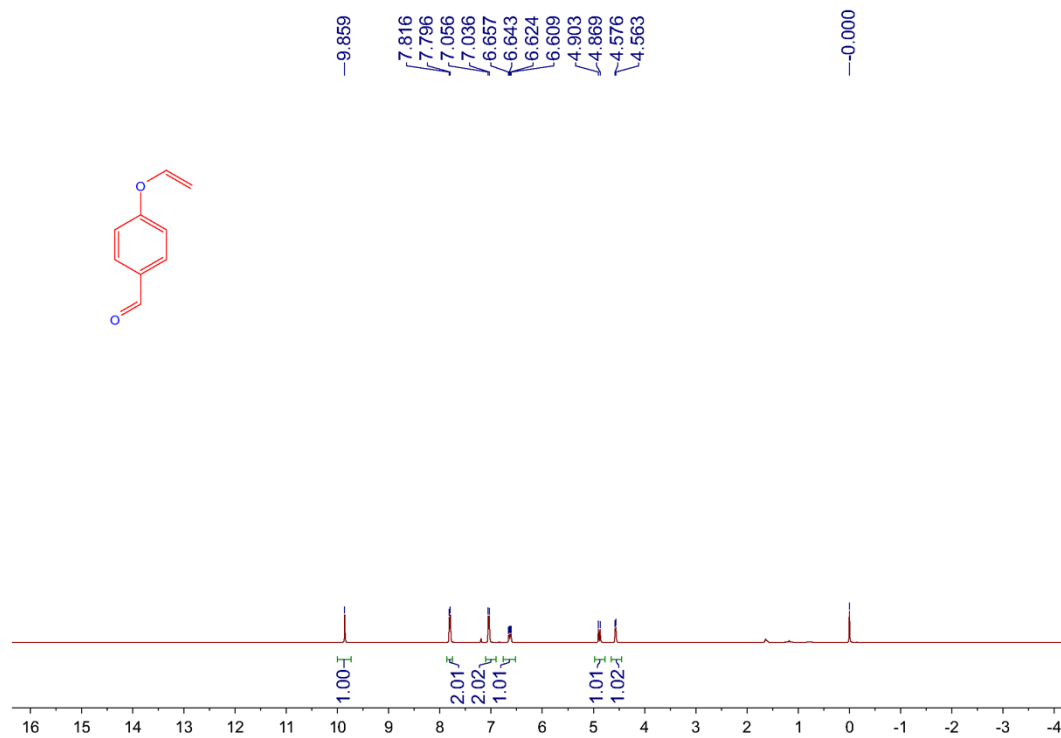
HeLa cells were cultured in DMEM (Invitrogen, Carlsbad, CA), supplemented with 10% fetal bovine serum in a humidified atmosphere of 5% CO<sub>2</sub> at 37 °C. For imaging experiments, exponentially growing cells (at a density of 20000-40000 cells per well, respectively) were seeded in 24-well plate. Cells were cultured at 37 °C in a 5% CO<sub>2</sub> atmosphere for 24 h before they were exposed to reagents. After the staining steps as described in figure captions, the images were collected upon excitation using the corresponding filters for DAPI (blue). Furthermore, MTT assay was employed to evaluate the cytotoxicity of MS2 and it turned out that this probe is virtually nontoxic to HeLa cells even at 20 μM after 24 h incubation.

## The characterization data of MS2

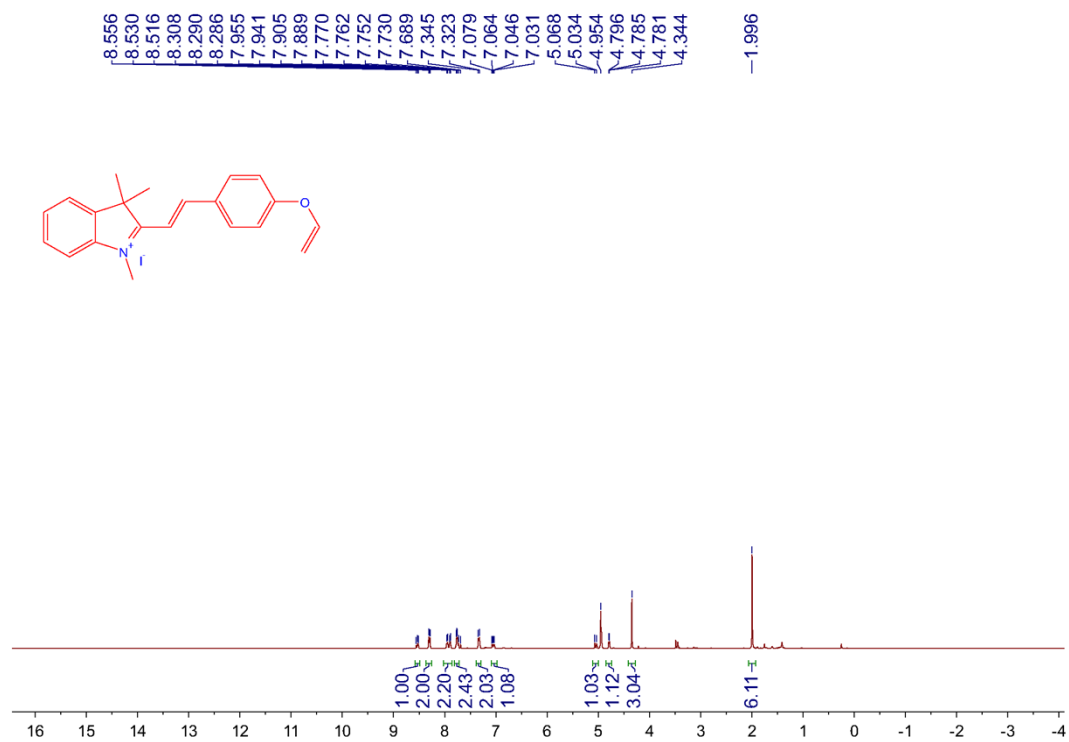
<sup>1</sup>H NMR of 1



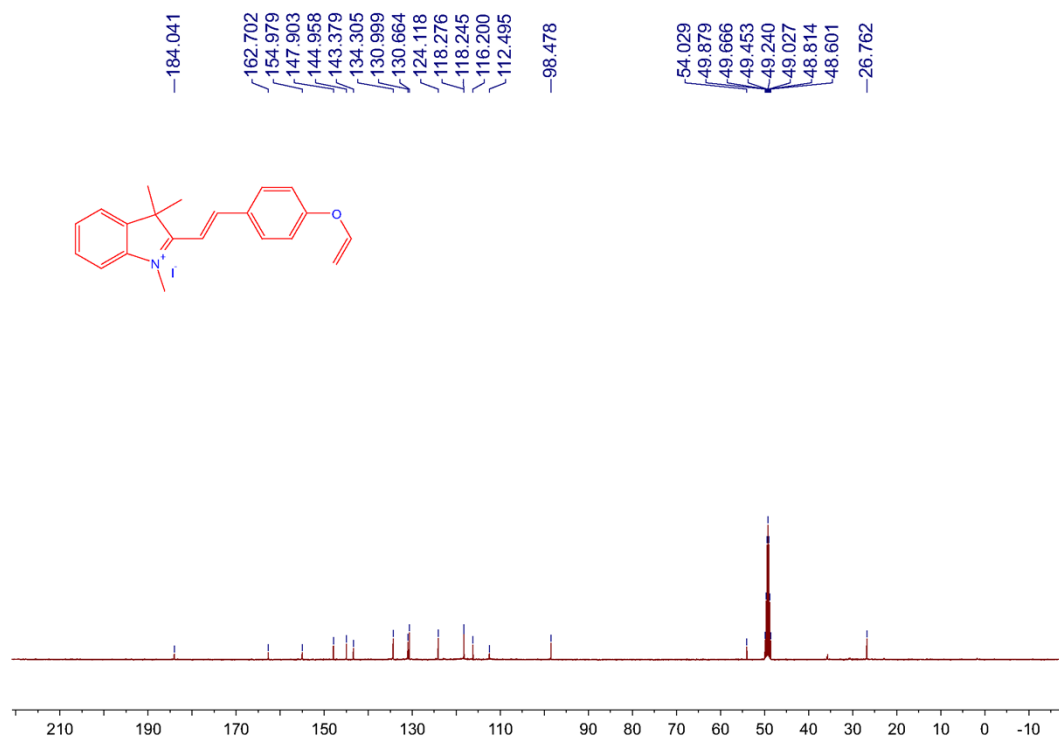
<sup>1</sup>H NMR of 2



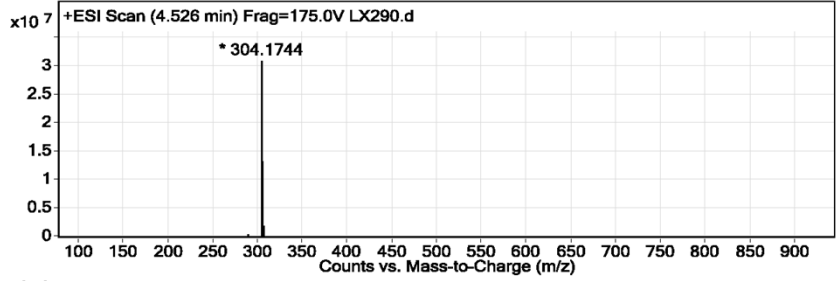
### <sup>1</sup>H NMR of 4 (MS2)



### <sup>13</sup>C NMR of MS2



### Qualitative Analysis Report



Peak List

m/z	z	Abund
304.1744	1	31036770
304.3218		2708289.5
305.1742	1	13436354
306.1782	1	2048707.9

## References

- (1) J. Gu, U. R. Anumala, F. L. Monte, T. Kramer, R. H. Haussen, J. Holzer, V. Goetschy-Meyer, G. Mall, I. Hilger, C. Czech, and B. Schmidt, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 7667-7671.
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- (3) (a) D. F. Eaton, *Pure Appl. Chem.*, 1988, **60**, 1107-1114; (b) D. Magde, R. Wong, and P. G. Seybold, *Photochem. Photobiol.*, 2002, **75**: 327-334.