## **Electronic Supplementary Information**

## Pseudopeptide polymer coating for improving biocompatibility and corrosion resistance of 316L stainless steel

Songtao Liu, Chaoshi Chen, Lijuan Chen, Haikun Zhu, Chong Zhang, Yanmei Wang\*

CAS Key Laboratory of Soft Matter Chemistry, Department of Polymer Science and Engineering,

University of Science and Technology of China, 230026, Hefei, People's Republic of China

\*Corresponding Author. E-mails: wangyanm@ustc.edu.cn

## Synthesis of PMOXA.

A typical procedure for the synthesis of PMOXA with a theoretically number-average molecular weight ( $\overline{M}_{*}$ ) of 2.5 kD, 5 kD, and 10 kD was as follows. MeOTf (Sigma-Aldrich Chemical Co. ,St. Louis, MO) 229.7 µL (2.04 mmol), 114.9 µL (1.02 mmol), or 57.4 µL (0.51 mmol)), MOXA ((Sigma-Aldrich Chemical Co. ,St. Louis, MO) 5.0 mL, 58.8 mmol) and dry acetonitrile (10.0 mL, Sinoreagent, Shanghai, China) were placed into a flame-dried Schlenk flask. The reaction mixture was stirred at 80 °C under nitrogen for 24 h. After, the reaction was quenched by addition of 6.0 mL water and 11.1 g solid Na<sub>2</sub>CO<sub>3</sub> (Sinoreagent, Shanghai, China) and heated to 80 °C for another 20 h. After being cooled to room temperature, the reaction mixture was extracted with CHCl<sub>3</sub> (200 mL × 3, Sinoreagent, Shanghai, China). The combined organic phases were dried over Na<sub>2</sub>SO<sub>4</sub> (Sinoreagent, Shanghai, China), filtered, and then concentrated under vacuum. Finally, the product hydroxy-terminated poly(2-methyl-2-oxazoline) (PMOXA-OH) was isolated by precipitation in ice-cold ether, filtered, and vacuum dried overnight. <sup>1</sup>H NMR spectra of PMOXA were displayed in Fig. S1.



Fig. S1 <sup>1</sup>H NMR spectra of PMOXA with different number-average molecular weight.

The MOXA repeating unit composition of different coating was calculated by below formula:

$$\gamma_{3}\left(\mathbb{P}_{x}\right) = \gamma_{1}\left(\mathbb{P}_{x}\right) \times \left(\mathbb{1} - \eta\left(\mathbb{P}_{x}\right)\right)$$
(S1)

$$\gamma_4(\mathbf{P}_x) = \gamma_2(\mathbf{P}_x) \times (\mathbf{1} - \eta(\mathbf{P}_x))$$
(S2)

Where  $\eta^{(P_x)}$  is the hydrolysis ratio of  $P_x$  (in Table 1);  $\gamma_1^{(P_x)}$  is the H-PMOXA composition in the  $P_x$ /dopamine coating calculated from the value of O/N;  $\gamma_2^{(P_x)}$  is H-PMOXA composition in the  $P_x$ /dopamine coating calculated from the value of C/N;  $\gamma_3^{(P_x)}$  is the MOXA repeating unit composition in the  $P_x$ /dopamine coating calculated from  $\gamma_1^{(P_x)}$ ;  $\gamma_4^{(P_x)}$  is the MOXA repeating unit composition in the  $P_x$ /dopamine coating calculated from  $\gamma_2^{(P_x)}$ . The results were displayed in Table S1.

**Table S1** The content of MOXA repeating unit ( $\gamma_3$  and  $\gamma_4$ ) in different coating.



Fig. S2 The illustration of the modification of slices.



**Fig. S3** The magnified SEM imagtes of the attached platelets on bare 316L SS slice (A,  $\times$  10000 image), PDA coating (B,  $\times$ 15000 image), P<sub>0</sub>/dopamine coating (C,  $\times$ 15000 image), P<sub>1</sub>/dopamine coating (D,  $\times$ 15000 image), P<sub>2</sub>/dopamine coating (E,  $\times$ 15000 image), P<sub>3</sub>/dopamine coating (F,  $\times$ 10000 image), P<sub>4</sub>/dopamine coating (G,  $\times$ 15000), P<sub>5</sub>/dopamine coating (H,  $\times$ 15000 image) modified 316L SS slices.



Fig. S4 The platelet attachment test on the  $P_2$ /dopamine coating modified stent after immersing in PBS for 21 days. As displayed, the result was similar to the  $P_2$ /dopamine coating modified stent after immersing in PBS for 72 h (Fig. 7).



Fig. S5 The bend test of  $P_2$ /dopamine coating modified stent. After bending, the coating on the stent was observed by SEM. There are not cracking, peeling, and flaking in the coating after being bended as shown in 5D.



Fig. S6 The magnified SEM images of HUVECs proliferation on the bare (A), PDA coating modified (B), and  $P_2$ /dopamine coating modified (C) 316L SS slices after 24 h cell culture. The scale bar is 20  $\mu$ m.

To study the cell viability of PMOXA, MTT assay was conducted to estimate the HUVECs viability of P<sub>0</sub>, P<sub>1</sub>, P<sub>2</sub>, and P<sub>3</sub>. This method represents a standard assay of cell viability, which is based on the colorimetric analyses of living cells. Seven concentrations (0.001, 0.01, 0.05, 0.1, 0.2, 0.5, and 1 mg/ml) of the studied polymers were tested. As depicted in Fig. S7, the P<sub>0</sub> and P<sub>1</sub> has good cell viability after 24 h, the survival cells of P<sub>0</sub> and P<sub>1</sub> were increased approximately 10%  $\sim$  20%, implying that the PMOXA possess excellent cell viability. Meanwhile, the survival cells of P<sub>2</sub> and P<sub>3</sub> were lower than that of P<sub>0</sub> and P<sub>1</sub>, but higher than that of the control. Obviously, compared to P<sub>0</sub> and P<sub>1</sub>, cell viability of P<sub>2</sub> and P<sub>3</sub> decreased. This could be ascribed to the hydrolysis ratio and imine groups of H-PMOXA increasing. Therefore, the results above demonstrated that the PMOXA exhibit a good cell viability and the imine groups of H-PMOXA can decrease the cell viability, thus, the P<sub>2</sub> can be used to produce a coating for enhance endothelial cell migration and proliferation, because the imine groups of H-PMOXA will react with PDA and PMOXA segment of P<sub>2</sub> can cover on the surfaces fully.



**Fig. S7** Cell viability assay of HUVECs cultured in cell culture medium with PMOXA and H-PMOXAs with different concentration. The results obtained using MTT test.