

Supplementary Information

**Highly efficient colorimetric detection of cancer cells
utilizing Fe-MIL-101 with intrinsic peroxidase-like catalytic activity over broad
pH range**

Daomei Chen, Bin Li,* Liang Jiang, Deliang Duan, Yizhou Li, Jiaqiang Wang,* Jiao
He and Yanbo Zeng

Yunnan Province Engineering Research Center of Photocatalytic Treatment of
Industrial Wastewater, The Universities' Center for Photocatalytic Treatment of
Pollutants in Yunnan Province, Key Laboratory of Medicinal Chemistry for Natural
Resource, Ministry of Education, School of Chemical Sciences & Technology,
Yunnan University, Kunming 650091, P.R. China.

Corresponding author. Fax: (+)86 871 65031567.

E-

mail: libin36@ynu.edu.cn (B. Li); jqwang@ynu.edu.cn (J. Wang)

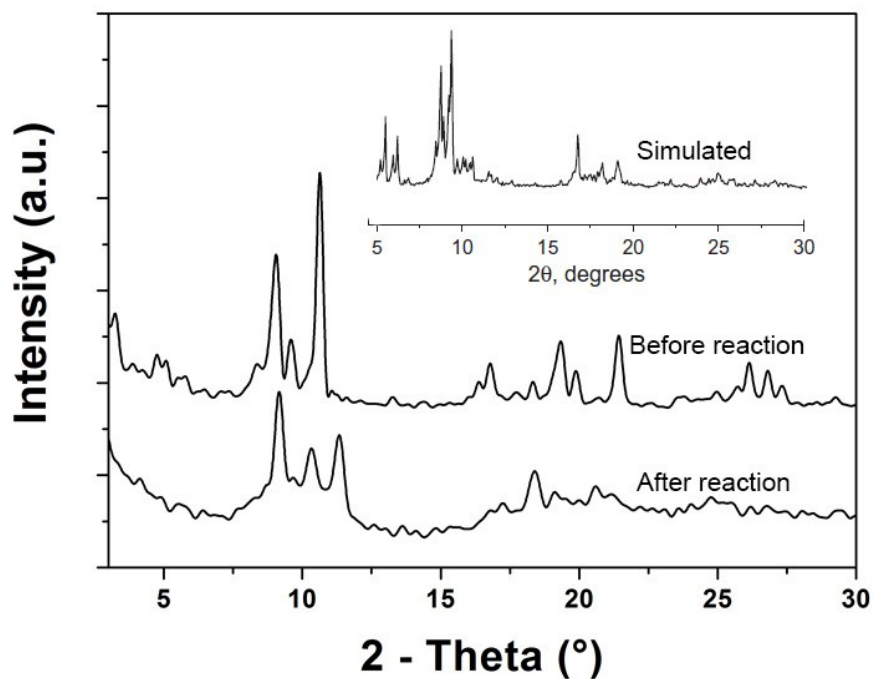


Fig.S1 Powder XRD patterns of Fe-MIL-101 before and after reacted with TMB and H_2O_2 solution at pH 4.0.

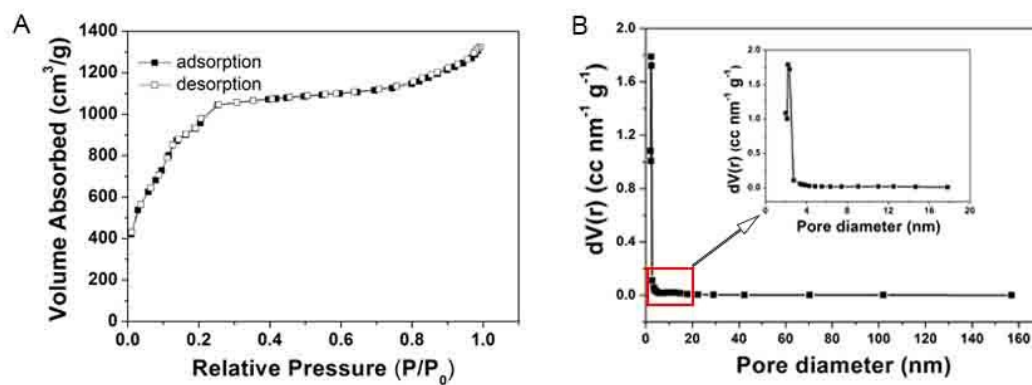


Fig.S2 (A) N_2 adsorption/desorption isotherm of Fe-MIL-101. (B) The BJH pore-size distribution, inset: the pore size distribution of Fe-MIL-101 at the range at 2-20 nm.

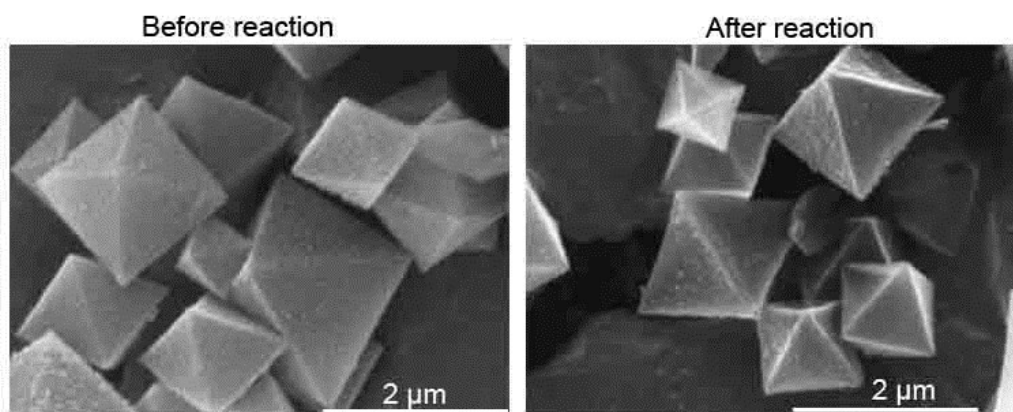


Fig.S3 SEM of Fe-MIL-101 before and after reacted with TMB and H₂O₂ solution at pH 4.0.

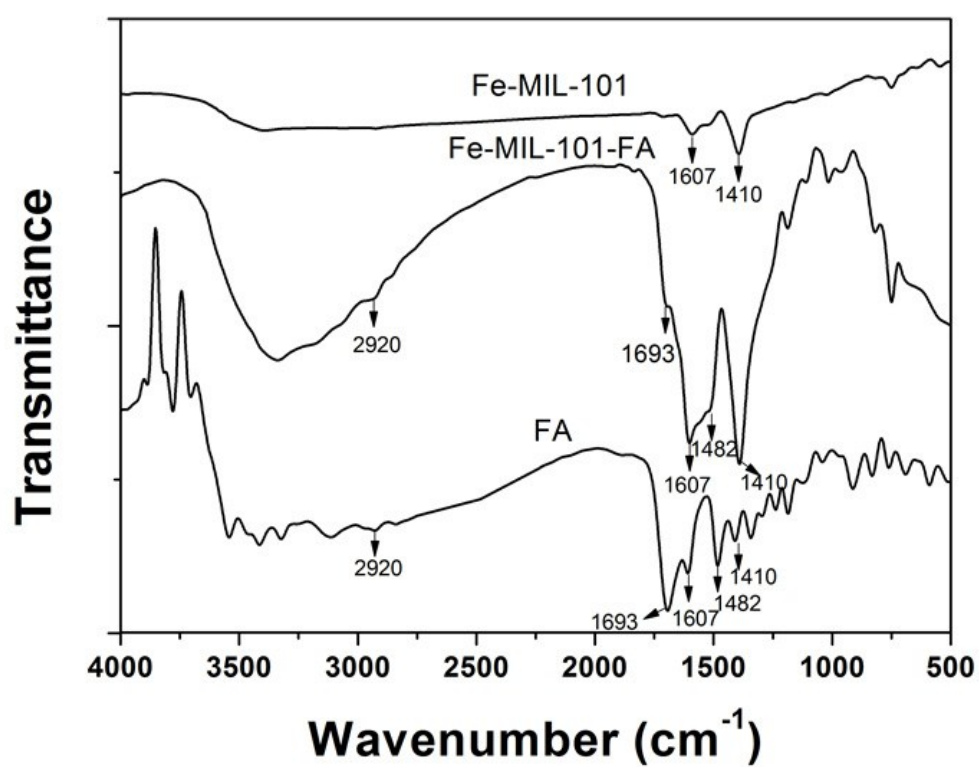


Fig.S4 FT-IR spectra of Fe-MIL-101, Fe-MIL-101-FA and FA.

Table S1 The size and zeta potential of Fe-MIL-101 and Fe-MIL-101-FA in PBS and in DMEM cell culture medium +10%FBS.

Materials	Size (nm)	Zeta potential (mV)	
		PBS	DMEM+10%FBS
Fe-MIL-101	1368 ± 70	□22.4 ± 1.1	□0.071 ± 0.01
Fe-MIL-101-FA	1620 ± 18	□17.6 ± 0.7	0.154 ± 0.03

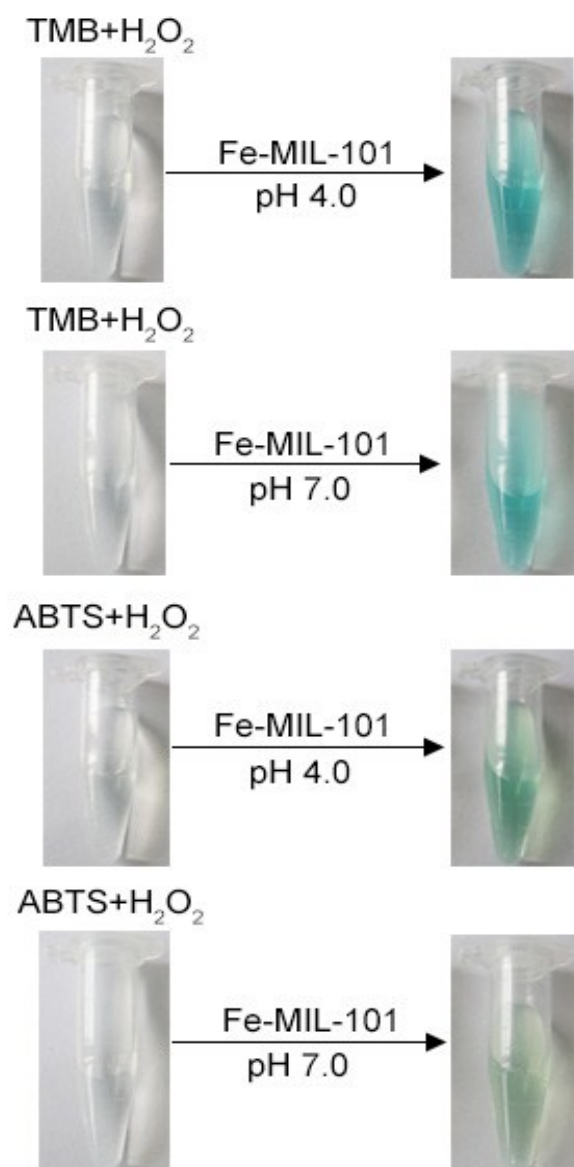


Fig.S5 Typical photographs of oxidation of TMB and ABTS by H₂O₂ catalyzed by Fe-MIL-101 in acetate buffer (pH 4.0) and borate (pH 7.0) solution.

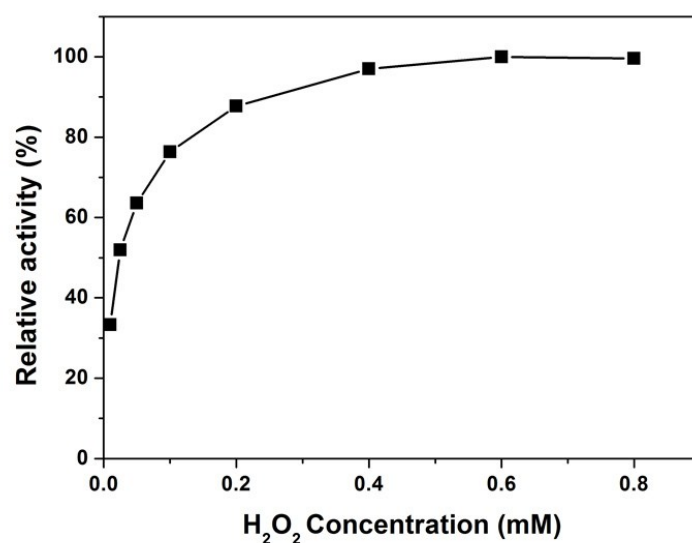


Fig.S6 The effect of H₂O₂ concentration on the peroxidase-like activity of Fe-MIL-101. Experiments were carried out using 20 $\mu\text{g mL}^{-1}$ Fe-MIL-101 in 5 mL buffer with 0.2 mM TMB as a substrate and the reaction time was 5 min.

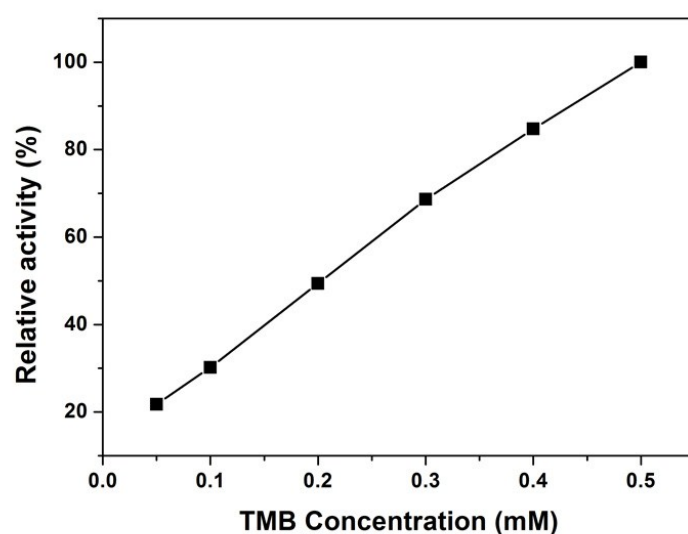


Fig.S7 The effect of TMB concentration on the peroxidase-like activity of Fe-MIL-101. Experiments were carried out using 20 $\mu\text{g mL}^{-1}$ Fe-MIL-101 in 5 mL buffer with 0.4 mM H₂O₂ as a substrate and the reaction time was 5 min.

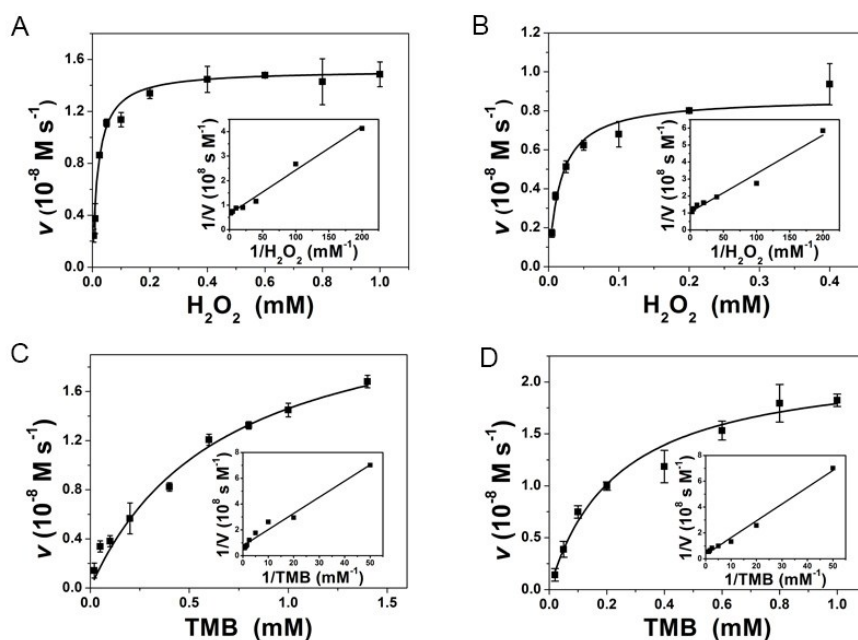


Fig.S8 Steady-state kinetic assays of the Fe-MIL-101-FA. (A, C) The concentration of TMB was 0.2 mM and the H_2O_2 concentration was varied in acetate buffer at pH 4.0 (A) and borate buffer at pH 7.0 (C); inset: double-reciprocal plots of activity of Fe-MIL-101-FA. (B, D) The concentration of H_2O_2 was 0.2 mM and the TMB concentration was varied in acetate buffer at pH 4.0 (B) and borate buffer at pH 7.0 (D), inset: double-reciprocal plots of activity of Fe-MIL-101-FA. Error bars shown represent the standard error derived from three repeated measurements.

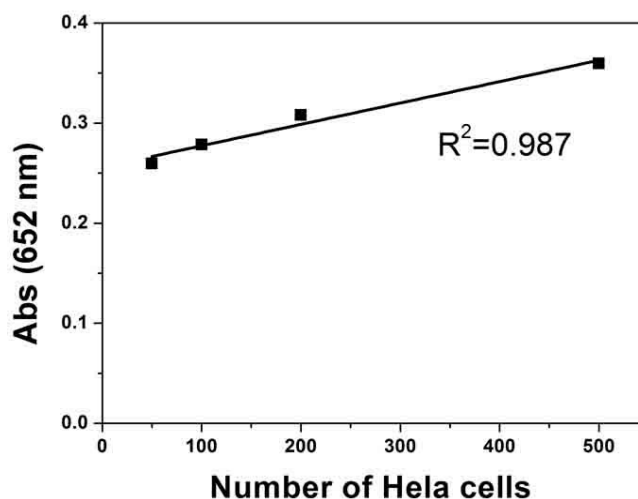


Fig.S9 The relationship between the numbers of HeLa cells and the absorbance intensity at 652 nm generated from the colorimetric assay using FA-conjugated Fe-MIL-101.

Table S2 Comparison of the kinetic parameters of different nanomaterials that mimic peroxidase at pH 4.0.

Catalysts	Substrates	$K_m(\text{mM})$	$V_{\max} (\text{M s}^{-1})$	Ref.
Fe NPs	TMB	0.38	2.38×10^{-7}	1
Fe NPs	H ₂ O ₂	0.32	4.1×10^{-7}	1
Co NPs	TMB	5.09	9.98×10^{-8}	1
Co NPs	H ₂ O ₂	1.14	1.72×10^{-8}	1
Fe _{0.5} Co _{0.5} NPs	TMB	1.79	4.56×10^{-7}	1
Fe _{0.5} Co _{0.5} NPs	H ₂ O ₂	0.06	1.32×10^{-7}	1
5-Fe-MSN	TMB	0.122	3.31×10^{-7}	2
5-Fe-MSN	H ₂ O ₂	6.67	3.26×10^{-7}	2
FeS	TMB	0.13	/	3
FeS	H ₂ O ₂	7.2	/	3
ZnFe ₂ O ₄ MNPs	TMB	0.85	1.33×10^{-8}	4
ZnFe ₂ O ₄ MNPs	H ₂ O ₂	1.66	7.74×10^{-8}	4
BSA-Au	TMB	0.00253	6.23×10^{-8}	5
BSA-Au	H ₂ O ₂	25.3	7.21×10^{-8}	5
CuInS ₂	H ₂ O ₂	2.01	9.78×10^{-8}	6
RET2-Pt2.9	TMB	0.056	5.82×10^{-7}	7
RET2-Pt2.9	H ₂ O ₂	48.0	5.68×10^{-7}	7
Au@Pt0.17	TMB	0.0095	1.02×10^{-7}	8
Au@Pt0.25	TMB	0.027	1.81×10^{-7}	8
WS ₂ nanosheets	TMB	1.83	4.31×10^{-8}	9
WS ₂ nanosheets	H ₂ O ₂	0.24	4.52×10^{-8}	9
GO-COOH	TMB	0.024	3.45×10^{-8}	10
GO-COOH	H ₂ O ₂	3.9	3.85×10^{-8}	10
GO_MNP-10	TMB	0.144	1.62×10^{-7}	11
PBMNPs	TMB	0.307	1.06×10^{-6}	12
PBMNPs	H ₂ O ₂	323.6	1.17×10^{-6}	12

References

1. Y. Chen, H. Cao, W. Shi, H. Liu, Y. Huang, *Chem. Commun.*, 2013, **49(44)**, 5013-5015.
2. S. Kumari, B.B. Dhar, C. Panda, A. Meena, S. Sen Gupta, *ACS Appl. Mater. Inter.*, 2014, **6(16)**, 13866-13873.
3. Z. Dai, S. Liu, J. Bao, H. Ju, *Chem - A Eur. J.*, 2009, **15(17)**, 4321-4326.
4. L. Su, J. Feng, X. Zhou, C. Ren, H. Li, X. Chen, *Anal. Chem.*, **84(13)**, 2012, 5753-5758.
5. X. Wang, Q. Wu, Z. Shan, Q. Huang, *Biosen. Bioelectron.*, 2011, **26(8)**, 3614-3619.
6. H. Liu, C. Gu, W. Xiong, M. Zhang, *Sensor. Actuat. B-Chem.*, 2015, **209**, 670-676.
7. Y. Fu, X. Zhao, J. Zhang, W. Li, *J. Phys. Chem. C*, 2014, **118(31)**, 18116-18125.
8. W. He, Y. Liu, J. Yuan, J. Yin, X. Wu, X. Hu, K. Zhang, J. Liu, C. Chen, Y. Ji, Y. Guo, *Biomaterials*, 2011, **32(4)**, 1139-1147.
9. T. Lin, L. Zhong, Z. Song, L. Guo, H. Wu, Q. Guo, Y. Chen, F. Fu, G. Chen, *Biosen. Bioelectron.*, 2014, **62**, 302-307.
10. Y. Song, K. Qu, C. Zhao, J. Ren, X. Qu, *Adv. Mater.*, 2010, **22(19)**, 2206-2210.
11. M.I. Kim, M.S. Kim, M. Woo, Y. Ye, K.S. Kang, J. Lee, H.G. Park, *Nanoscale*, 2014, **6(3)**, 1529-1536.
12. X. Zhang, S. Gong, Y. Zhang, T. Yang, C. Wang, N. Gu, *J. Mater. Chem.*, 2010, **20(24)**, 5110-5116.