Supporting Information

Molecular Orientation within Thin Films of Isomorphic Molecular Semiconductors

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Materials and Methods

Solvents and Synthetic Intermediates. Toluene was degassed and dried through a standard solvent system. Chloroform was purchased from Sigma-Aldrich and stored inside a N₂-filled glovebox before use. Chloroform and hexanes used for silica gel column chromatography were analytical grade and used without further purification. Compounds *4-(4,4-Bis(2-ethylhexyl)-4H-silolo[3,2-b:4,5-b']dithiophen-2-yl) -7-bromobenzo[c][1,2,5]thiadiazole* (1)^{S1} and *5'-hexyl-2,2'-bithiophene-5-trimethylstannane* (2)^{S2} were prepared using the reported procedures. Stille coupling reaction was conducted on a Biotage® Microwave Initiator.

¹H, and ¹³C Nuclear Magnetic Resonance (NMR) spectra were recorded on a Bruker Avance 500 MHz spectrometer by using either CDCl₃, CD₂Cl₂ or C₂D₂Cl₄. Chemical shifts (δ) were expressed as parts per million (ppm), and calibrated using residual solvent signals by referring tetramethylsilane (TMS) as 0 ppm. Coupling constants (*J*) were reported in Hertz (Hz).

Field Desorption Time-of-Flight (FD-TOF) Mass Spectra were measured on a VG70 Magnetic Sector instrument at the Mass Spectrometry Facility of UCSB Department of Chemistry and Biochemistry. Samples were prepared by dissolving each molecule into chloroform with a final concentration of ~ 1 mg/mL.

Solution and Thin-film Ultraviolet/Visible (UV/vis) Absorption measurement were performed by using a Beckman Coulter DU 800 series UV-vis spectrophotometer. Solution samples were measured in dry chloroform with a solid concentration of 0.02 mg/mL at room temperature. Thin films were casted from chloroform solutions (10 mg/mL) on pre-cleaned glass substrates at a spin speed of 1500 rpm for 60 s. The thicknesses of all films were calculated between 50 to 65 nm from X-ray reflectivity measurements, which were used to calculate absorption coefficient (α) in the solid state.

Density Functional Theory (DFT) calculations were done using the Gaussian 09 software suite.^{S3} The hybrid long range corrected CAM-B3LYP functional^{S4} with the basis set 6-31G(d,p) were used for both the optimization of the geometry as well as the time-dependent and natural transition orbital analysis.^{S5}

Ultraviolet Photoelectron Spectroscopy (UPS) analysis was done on a Kratos Axis Ultra equipped with a hemispherical electron energy analyzer and was held at a constant pressure of 1.35×10^{-7} Torr. Neat films of **N0**, **N2a**, **N2b**, and **N4** were spun cast on cleaned and UV/ozone-treated Corning 1737 glass patterned with 140 nm of ITO from solutions with concentrations ~2 mg/mL at 2000 RPM for 60 seconds yielding films with an approximate thickness of 10-15 nm. Films were cast in a glovebox with an N₂ atmosphere and were transferred using an air-free sample holder to minimize exposure to Air. UPS spectra were then obtained using a He I (hv = 21.21 eV) and the electron energy analyzer was held at a constant pass energy of 5 eV.

Differential Scanning Calorimetry (DSC) was performed on a TA Instruments DSC (Model Q-20 with a RCS90 cooling system) with 3~4 mg of each sample with a heating/cooling ramp of 10 °C/min under an N₂ atmosphere. The data from the second cycle of each measurement was collected in order to eliminate thermal history of materials.

Grazing-Incidence Wide Angle X-ray Scattering (GIWAXS) measurements were performed at beamline

7.3.3 of the Advanced Light Source with an incidence angle of $\sim 0.13^{\circ}$ under a helium environment to minimize beam damage and reduce scattering from the air. Thin film samples were prepared by spincasting 10 mg/mL chloroform solution of each material on top of silicon (100) substrates.

2 C₆H₁₃ SnMe₃ Pd(PPh₃)₄, MW 89% 1 3 $\mathbf{R} = \mathbf{H}$ 1) LDA 2) Me₃SnCl R = SnMe 92% NO Pd(PPh₃)₄, MW 65~75% 5 X = CH 6 X = N

Synthesis and Characterization of Molecules N0 and N2b

Scheme S1. Synthetic Procedures for Molecules N0 and N2b.

Synthesis of $5-\{4-(7-hexylthiophen-2-yl)thiophen-2-yl-benzo[c][1,2,5]thiadiazole]-3,3'-bis (2$ ethylhexyl)silylene-2,2'-bithiophene (**3**). Inside a nitrogen-filled glovebox, compound**1**(400 mg, 0.71 mmol, 1.0 eq),**2**(293 mg, 0.71 mmol, 1.0 eq), Pd(PPh₃)₄ (49 mg, 0.04 mmol, 0.06 eq), and dry toluene (4 mL) were combined in a 5 mL Biotage® reaction vial and sealed. The solid suspension was subsequently subjected to microwave irradiation with a heating program as 70 °C for 2 min, 100 °C for 2 min, 130 °C for 5 min, and 160 °C for 40 min. The resulting mixture was purified by using silica gel column chromatography (hexane:chloroform = 10:3 as eluent). Compound**3**(506 mg, 89%) was obtained as a deep red oil. ¹H NMR (CDCl₃, 500 MHz) & 8.15–8.12 (m, 1H), 8.02–8.00 (d, J = 4.0 Hz, 1H), 7.83–7.79 (m, 2H), 7.27–7.25 (d, J = 3.5 Hz, 1H), 7.18–7.17 (d, J = 4.0 Hz, 1H), 7.10–7.08 (m, 1H), 6.73–6.71 (d, J = 3.5 Hz, 1H), 2.84–2.80 (t, J = 7.5 Hz, 2H), 1.74–1.67 (m, 2H), 1.53–1.14 (m, 24H), 1.08–0.96 (m, 4H), 0.93–0.78 (m, 15H). ¹³C NMR (CDCl₃, 130.3, 128.4, 126.6, 126.3, 125.6, 125.3, 125.0, 144.3, 143.8, 143.7, 140.3, 139.5, 137.9, 135.0, 130.8, 130.3, 128.4, 126.6, 126.3, 125.6, 125.3, 125.0, 124.1, 124.0, 36.3, 36.1, 31.9, 30.6, 29.3, 29.2, 29.1, 23.4, 23.3, 22.9, 18.2, 18.1, 18.0, 14.5, 14.4, 11.2.

Synthesis of 5-{4-(7-hexylthiophen-2-yl)thiophen-2-yl-benzo[c][1,2,5]thiadiazole}-5'-{trimethylstannyl} -3,3'-bis(2-ethylhexyl)silylene-2,2'-bithiophene (4). A solution of compound **3** (460 mg, 0.57 mmol, 1.0 eq) in dry THF (13 mL) was cooled to -35 °C under N₂ and then LDA (0.6 M in THF/hexane, 1.4 mL, 0.86 mmol, 1.5 eq) was added dropwise. The mixture was stirred for 10 min at -35 °C. Trimethyltin chloride (Me₃SnCl, 193 mg, 0.97 mmol, 1.7 eq) in THF (0.6 mL) was added subsequently and the resulting mixture was stirred for 1 hour while warming up to room temperature. The solution was then quenched in water and diluted with ether (200 mL), followed by washing with brine (2 × 100 mL) and water (2 × 100 mL). Compound **4** (506 mg, 92%) was afforded as a purple oil after the solvents were evaporated off. ¹H NMR (CD₂Cl₂, 500 MHz) & 8.21–8.19 (m, 1H), 8.06–8.04 (d, *J* = 4.0 Hz, 1H), 7.88–7.87 (m, 2H), 7.23–7.21 (d, *J* = 4.0 Hz, 1H), 9.20–7.19 (s, 1H), 7.14–7.13 (d, J = 4.0 Hz, 1H), 6.77–6.75 (d, *J* = 3.5 Hz, 1H), 2.86–2.82 (t, *J* = 8.0 Hz, 2H), 1.75–1.69 (m, 2H), 1.52–1.14 (m, 24H), 1.10–0.97 (m, 4H), 0.94–0.78 (m, 15H), 0.48–0.35 (s, 9H).

Synthesis of **N0** *and* **N2b**. Molecules **N0** and **N2b** were prepared according to the same reaction and purification procedures that were previously described in the synthesis of **3** by only varying reactants as follows. The amount and yield obtained for each molecule are shown in parenthesis. **N0** (130 mg, 75%): **4** (150 mg, 0.16 mmol, 2.1 eq), **5** (64 mg, 0.08 mmol, 1.0 eq), and Pd(PPh₃)₄ (7 mg, 0.006 mmol, 0.08 eq); **N2b** (113 mg, 65%): **4** (150 mg, 0.16 mmol, 2.1 eq), **6** (64 mg, 0.08 mmol, 1.0 eq), and Pd(PPh₃)₄ (7 mg, 0.006 mmol, 0.08 eq).

N0: ¹H NMR (C₂D₂Cl₄, 373 K, 500 MHz) δ : 8.29–8.25 (m, 6H), 8.12–8.10 (d, *J* = 4.0 Hz, 2H), 7.94–7.87 (m, 8H), 7.30–7.28 (d, *J* = 3.5 Hz, 2H), 7.22–7.20 (d, *J* = 3.5 Hz, 2H), 6.84–6.82 (d, *J* = 3.5 Hz, 2H), 2.96–2.91 (t, *J* = 8.0 Hz, 4H), 1.87–1.80 (m, 4H), 1.75–1.67 (m, 4H), 1.57–1.20 (m, 74H), 1.05–0.93 (m, 42H). FD-TOF MS: Calculated for C₁₂₄H₁₅₀N₈S₁₄Si₃: 2282.7. Found: 2282.6 (M⁺), 1141.3 (M²⁺), 760.9 (M³⁺).

N2b: ¹H NMR (C₂D₂Cl₄, 373 K, 500 MHz) *δ*: 8.98−8.96 (s, 2H), 8.85−8.82 (m, 2H), 8.29−8.25 (m, 4H), 8.15−8.13 (d, *J* = 4.0 Hz, 2H), 7.98−7.92 (m, 4H), 7.31−7.29 (d, *J* = 3.5 Hz, 2H), 7.22−7.20 (d, *J* = 3.5 Hz, 2H), 6.85−6.83 (d, *J* = 3.5 Hz, 2H), 2.96−2.91 (t, *J* = 8.0 Hz, 4H), 1.88−1.80 (m, 4H), 1.74−1.64 (m,

4H), 1.58–1.19 (m, 74H), 1.05–0.90 (m, 42H). FD-TOF MS: Calculated for C₁₂₂H₁₄₈N₁₀S₁₄Si₃: 2284.7. Found: 2284.6 (M⁺), 1142.3 (M²⁺), 761.6 (M³⁺).



Figure S1. UPS spectra of N0, N2a, N2b, and N4.



Figure S2. DSC traces of N0 and N2b.

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