

## Supplementary information

**Role of N-acyl-homoserine lactone (AHL) based quorum sensing on biofilm formation on packing media in wastewater treatment process**

Huizhi Hu<sup>1</sup>, Junguo He<sup>1\*</sup>, Jian Liu<sup>1</sup>, Huarong Yu<sup>1</sup>, Jian Tang<sup>1</sup>, Jie Zhang<sup>1</sup>

School of Municipal and Environmental Engineering, Harbin Institute of Technology,  
73 Huanghe Road, Harbin, China.

\*Corresponding author, E-mail: [junguohe@263.net](mailto:junguohe@263.net); Tel: +86-0451-86289099

## Table

**Table S1** Composition of synthetic wastewater

<b>substance</b>	<b>concentration (mg/L)</b>
CH <sub>3</sub> COONa	400
NH <sub>4</sub> Cl	107
MgSO <sub>4</sub> · 7H <sub>2</sub> O	90
CaCl <sub>2</sub>	10.55
K <sub>2</sub> HPO <sub>4</sub>	64
KH <sub>2</sub> PO <sub>4</sub>	28
yeast extract	1
elements solution	0.5 ml

**Table S2** Composition of trace elements solution

<b>substance</b>	<b>concentration (mg/L)</b>
FeCl <sub>3</sub> · 6H <sub>2</sub> O	1.50
CuSO <sub>4</sub> · 5H <sub>2</sub> O	0.03
H <sub>3</sub> BO <sub>3</sub>	0.15
KI	0.18
MnCl <sub>2</sub> · 4H <sub>2</sub> O	0.12
NaMoO <sub>4</sub> · 2H <sub>2</sub> O	0.06
ZnSO <sub>4</sub> · 7H <sub>2</sub> O	0.12
CoCl <sub>2</sub> · 6H <sub>2</sub> O	0.15
EDTA	10.00

**Table S3** Composition of trace elements solution

AHL	Screening time (min)	Retention time (min)	Transitions <sup>1</sup>	Dwell	Cone	Collision
C4-HSL	/	/	172.2 →102.1 172.2 →71.1	0.015	22	12
3OC6-HSL	/	/	214.2 →102.1 214.2 → 71.1	0.015	22	12
C6-HSL	1.81-2.18	2.12	200.2 → 99.1 200.2 →102.1	0.015	20	10
3OC8-HSL	/	/	242.2 →102.1 242.2 → 71.3	0.015	22	12
C7-HSL	/	/	214.2 →113.1 214.2 →102.1	0.015	22	12
C8-HSL	5.25-5.87	5.53	228.2 →127.1 228.2 →102.1	0.015	22	12
3OC10-HSL	/	/	270.3 →102.1 270.3 →169.1	0.015	22	12
C10-HSL	/	/	256.3 →102.1 256.3 →155.1	0.015	22	12
C12-HSL	/	/	283.3 →102.1 284.3 →183.2	0.015	22	12
3-oxo-C12-HSL	13.39-14.46	14.25	298.3 →102.1 298.3 →197.2	0.015	22	12
C14-HSL	18.85-19.16.	19.05	312.4 →102.1 312.4 →211.3	0.015	22	12

<sup>1</sup>Two most prominent transition ions generated from each precursor ion based on the optimized MS/MS parameters were chosen. The first MRM transition was used for quantification while the second MRM transition was used for identity confirmation.

**Table S4** The top 34 most abundant community members (relative abundance >1%) at genus level

Rank	Blank	Phase I			Phase II						Phase III			add-back	
		2W	4W	6W	8W	10W	12W	14W	16W	18W	20W	22W	24W	50 nM	1000 nM
<i>Brevundimonas</i>	0.34	0.53	0.35	0.68	0.67	0.87	1.09	1.55	1.44	1.21	0.98	1.03	1.07	1.14	0.97
<i>Ilumatobacter</i>	0.34	0.23	0.56	0.78	1.78	2.32	2.34	3.55	3.09	2.12	1.89	1.67	1.63	1.51	1.57
<i>Nitrospira</i>	0.45	0.32	0.43	0.45	0.67	0.78	0.85	0.94	0.84	0.90	5.78	6.35	5.71	5.82	4.93 ▼
<i>Longilinea</i>	2.13	2.29	2.66	2.45	1.17	1.21	1.41	2.02	1.92	1.86	1.43	1.43	2.23	3.21 ▲	3.45 ▲▲
<i>Pseudorhodoferax</i>	0.93	0.42	0.58	0.87	1.06	1.46	1.56	1.87	1.68	1.55	1.82	3.42	3.16	0.79 ▼▼	0.73 ▼▼
<i>Pseudorhodoferax</i>	0.56	0.84	0.84	0.89	0.98	0.46	1.66	0.84	0.89	0.98	0.73	0.62	0.74	0.99 ▲	0.92 ▲
<i>Rhodobacter</i>	0.29	0.38	0.32	0.54	1.13	1.87	1.43	1.97	2.07	2.10	0.92	0.48	0.84	1.01 ▲	1.14 ▲
<i>Flavobacterium</i>	0.39	0.43	0.58	0.43	2.40	1.94	1.72	1.67	1.53	1.34	0.94	0.89	0.93	0.73 ▼	0.63 ▼
<i>Cellulophaga</i>	0.69	0.78	0.95	0.84	1.44	1.01	1.10	1.32	1.12	1.08	0.69	0.72	0.54	0.62 ▲	0.67 ▲
<i>Genmatimonas</i>	0.43	0.54	0.53	0.65	0.58	0.68	0.83	0.47	0.69	0.78	0.87	1.23	1.31	1.42	1.23
<i>Dechloromonas</i>	4.52	5.30	5.65	5.58	7.68	8.05	9.46	10.43	8.34	7.96	7.41	6.92	7.00	8.32 ▲	8.12 ▲
<i>Azospira</i>	0.32	0.12	0.32	0.23	0.21	1.09	0.48	0.85	0.67	1.68	2.87	3.30	2.98	3.02	3.11

<i>TM7_genera</i>	0.19	0.18	0.21	0.23	0.65	0.87	1.24	2.49	1.56	1.04	1.17	1.22	1.49	1.78	1.95
														▲	▲
<i>Ferruginibacter</i>	1.34	1.26	1.40	1.42	0.89	0.97	1.23	1.43	1.63	1.15	1.61	1.43	1.33	3.37	3.56
														▲▲	▲▲
<i>Thermomonas</i>	0.58	0.43	0.54	0.43	1.34	1.42	1.48	2.52	1.43	1.10	0.54	0.67	0.58	0.47	0.42
														▼	▼
<i>OD1_genera</i>	1.03	1.16	1.23	1.11	0.94	0.84	0.89	0.74	0.59	0.83	0.58	0.38	0.43	0.48	0.67
														▲	▲
<i>Simplicispira</i>	0.38	0.43	0.43	0.73	1.14	0.98	2.44	0.78	1.43	0.84	0.43	0.66	0.75	0.42	0.34
														▼	▼▼
<i>Caenimonas</i>	0.59	0.54	0.67	0.73	1.34	1.05	3.32	1.29	1.60	1.62	2.84	1.08	1.06	1.23	1.34
														▲	▲
<i>Sphingopyxis</i>	0.55	0.45	0.78	0.89	1.54	1.43	1.38	1.82	1.62	1.32	0.93	0.84	0.87	1.11	0.93
														▲	
<i>Hydrogenophaga</i>	3.12	3.34	4.77	4.60	1.25	1.56	2.68	3.17	2.17	1.36	0.92	0.98	0.94	0.99	0.67
															▼
<i>Arcobacter</i>	1.87	1.29	1.23	1.87	0.38	1.48	0.64	1.02	0.48	0.53	0.36	0.48	0.43	0.43	0.58
															▲
<i>Phycisphaera</i>	0.49	0.84	0.47	0.63	0.43	0.74	0.76	0.89	0.94	0.95	1.43	1.23	1.18	1.22	1.43
															▲
<i>Terrimonas</i>	0.47	0.54	0.74	0.87	0.85	1.12	1.67	3.91	4.20	1.23	2.20	2.37	2.34	0.75	0.94
<i>Leptothrix</i>	1.91	1.84	1.88	1.09	0.84	0.73	0.84	0.92	0.83	0.99	4.18	3.96	3.33	1.02	1.04
<i>Bdellovibrio</i>	1.29	1.27	1.19	1.11	0.75	0.78	0.84	0.63	0.62	0.68	0.59	0.63	0.52	0.42	0.57
														▼	
<i>Perlucidibaca</i>	1.20	1.16	1.01	1.88	0.93	0.83	0.94	0.99	0.81	0.98	0.84	0.73	0.69	1.45	1.64
														▲▲	▲▲
<i>Sulfuritalea</i>	2.15	1.78	1.81	2.30	2.20	1.90	1.59	1.03	0.84	2.65	5.27	5.57	6.97	3.29	4.39

<i>Gp4</i>	0.93	0.95	0.74	0.84	1.12	0.72	0.89	1.04	2.30	1.54	0.89	0.56	0.53	0.73	0.85
														▲	▲▲
<i>Haliscomenobacter</i>	12.59	12.65	11.89	11.97	12.67	13.50	10.49	9.30	10.99	14.24	7.49	6.15	4.99	5.02	5.34
<i>Sediminibacterium</i>	1.53	1.52	1.53	1.45	2.20	1.10	1.68	1.12	1.87	1.54	0.95	0.69	0.79	0.83	0.97
															▲
<i>Aeromonas</i>	0.72	0.68	0.92	0.83	1.25	1.56	1.78	2.45	2.23	2.12	1.54	1.45	1.40	1.12	1.45
														▼	
<i>Enterobacter sp.</i>	0.82	0.93	0.69	0.93	1.03	1.10	1.21	1.34	1.26	1.17	0.79	0.76	0.85	0.92	1.32
															▲▲
<i>Nitrosomonas</i>	0.17	0.52	0.66	0.79	1.12	1.35	1.76	1.87	1.42	1.32	1.16	1.14	1.05	2.29	3.45
														▲▲	▲▲
<i>Stenotrophomonas sp.</i>	1.32	1.36	1.97	2.78	2.92	3.58	4.12	5.31	4.25	3.70	1.43	1.40	1.24	3.15	4.12
														▲▲	▲▲

▼ : decrease rate > 10%; ▼▼ : decrease rate > 50%;

▲ : increase rate > 10%; ▲▲ : increase rate > 50%.

## Figure



**Figure S1** - Picture of polypropylene media used in the process