

Electronic Supplementary Information

for

Nano-NiFe₂O₄ catalyzed microwave assisted one-pot regioselective synthesis of novel 2-alkoxyimidazo[1,2-*a*] pyridines under aerobic conditions

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ESI 1. Detailed method for the preparation of NiFe₂O₄:

The NiFe₂O₄ nanoparticles were prepared by simple co-precipitation method^[1] from easily available starting materials ferric chloride (FeCl₃.6H₂O) and nickel chloride (NiCl₂.6H₂O), distilled water and sodium hydroxide. In a typical synthetic protocol, 0.2 M (20 mL water) ferric chloride solution and 0.1 M (20 mL water) solution of nickel chloride were prepared and mixed under vigorous stirring for 2 h at 80 °C. After that, 0.3 M NaOH was added drop by drop into the solutions till the pH is reached upto 12 and brown colour precipitates were formed. Finally, the precipitates were separated by centrifugation and dried in hot air oven for 4 h at 100 °C. Then calcinated it at 550°C for 6 h. Formation of NiFe₂O₄NPs were confirmed by FT-IR, powder XRD, HR-TEM, FESEM and EDAX studies.

ESI 2. Characterization of nano-NiFe₂O₄:

ESI 2.1. Energy-dispersive X-ray spectroscopy (EDAX) of nano-NiFe₂O₄

EDAX of nano-NiFe₂O₄ proved that the material contains only Ni, Fe, O elements and no other impurities are observed in the material.

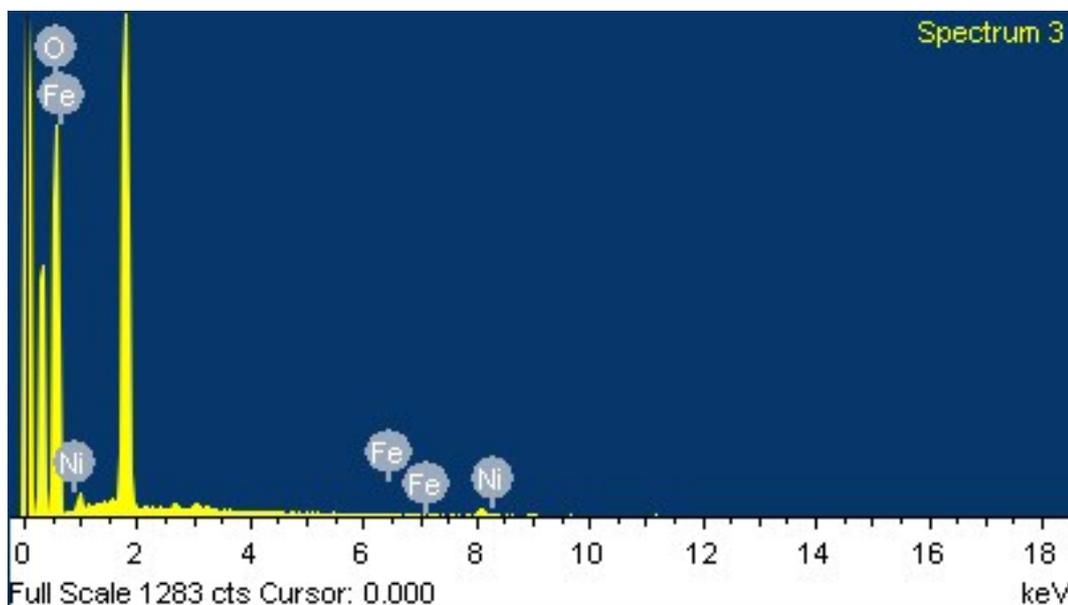


Fig. 1S EDAX of nano-NiFe₂O₄

ESI 2.2.FESEM image of nano-NiFe₂O₄:

The field emission scanning electron microscope image of NiFe₂O₄ also confirms the formation of spherical particles.

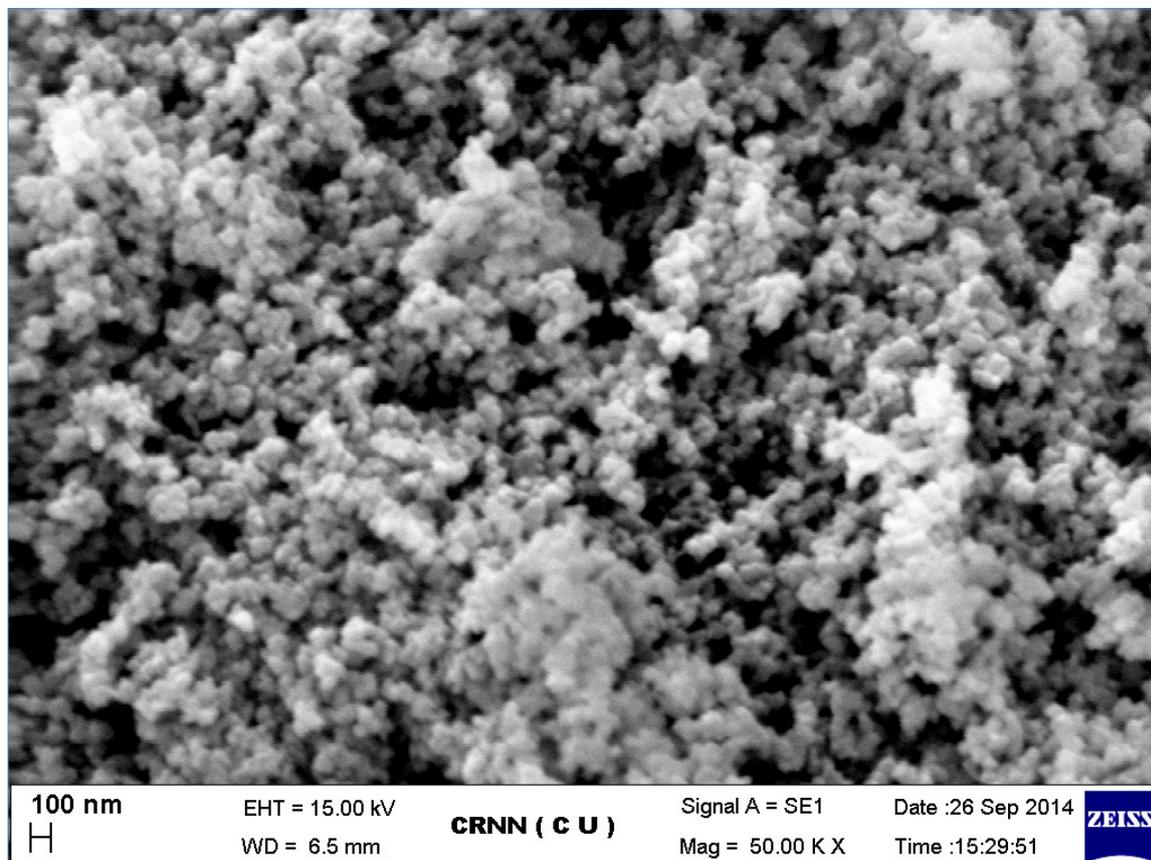


Fig. 2S FESEM image of nano-NiFe₂O₄ :

ESI 2.3. FTIR Spectra of nano-NiFe₂O₄:

An intrinsic stretching vibrations at 600 and 535 cm⁻¹ were assign for tetrahedral site of ferrite (Fe-O) and 817 cm⁻¹ for Fe-OH group were observed in FT-IR of nano-NiFe₂O₄ (Fig. 5S).

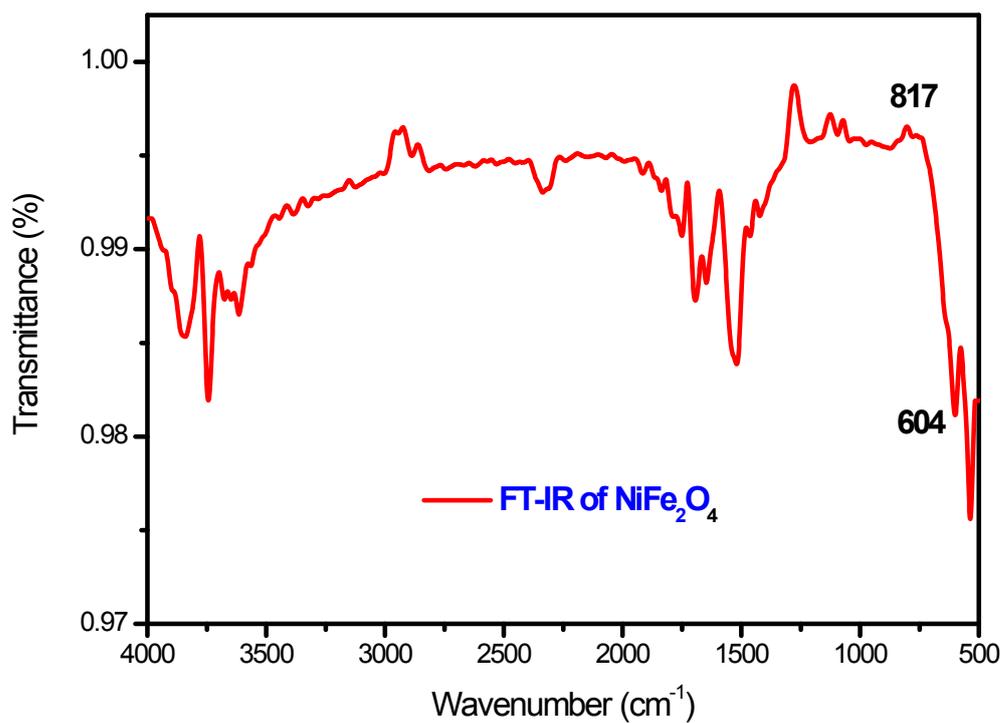


Fig. 3S FTIR Spectra of nano-NiFe₂O₄

ESI 3. Methods for the preparation of CuFe₂O₄:

The preparation of CuFe₂O₄ nanoparticles were carried out following reported procedure^[3]. Solution of Cu(NO₃)₂ (0.001 mol), and FeCl₃·6H₂O (0.002 mol) were prepared and vigorously mixed under stirring for 2 h at 80 °C. Subsequently, 0.3 M NaOH was added drop by drop into the solutions till the pH is reached upto 12 and black precipitate is formed. Then centrifuged and rinsed with distilled water and left in an atmosphere environment to dry. The resulting powder is then calcinated at 850° C in an oven for 2 hours.

ESI 4. Preparation of CoFe₂O₄

The preparation of CoFe₂O₄ nanoparticles were carried out following reported procedure^[4]. First, 2.0 g of anhydrous sodium acetate was dissolved in 30 mL of ethylene glycol, and the mixture was stirred vigorously at room temperature to give a transparent solution. Subsequently, 1.5 mmol of cobalt hydrous chlorides (CoCl₂·6H₂O) and 3.0 mmol of ferric hydrous chloride (FeCl₃·6H₂O) was added slowly to the above solution (the stoichiometric molar ratio of Co²⁺/Fe³⁺ was 1 : 2). This mixture was then vigorously stirred at 70°C for at least 2 h to form a homogeneous solution. Then centrifuged and rinsed with distilled water and left in an atmosphere environment to dry. The resulting powder is then calcinated at 850° C in an oven for 2 hours.

ESI 5. General Information:

All reagents were obtained from commercial sources and used as received, except all solvents which were distilled prior to use. All reactions were carried out with oven-dried glassware under air and all the M.W. reactions were carried out in a sealed M.W. tube under closed air system. A monomode microwave reactor (CEM Discover 908005 Microwave Digestion System with Max. Microwave Power 300W, Volts 90/140 VAC, Freq. 50/60 Hz from CEM Corporation, USA) has been used for all the reactions. Technical grade petroleum ether and ethyl acetate were used for column chromatography. Analytical TLC was performed on Merck 60 F254 silica gel plates (0.25 mm thickness). Column chromatography was performed on silica gel (60-120 mesh size, HIMEDIA, India). ¹H NMR spectra were

determined on a Bruker 500, 400 and 300 MHz spectrometer as solutions in CDCl₃. Chemical shifts are expressed in parts per million (δ) and the signals were reported as s (singlet), d (doublet), t (triplet), m (multiplet) and coupling constants J were given in Hz. ¹³C NMR spectra were recorded at 125, 100 and 75 MHz in CDCl₃ solution. Chemical shifts are expressed in parts per million (δ) and are referenced to CDCl₃ (δ = 77.16) as internal standard. All FT-IR was recorded on a BRUKER ALPHA spectrophotometer.

ESI 6. General method for the synthesis of 2-alkoxy-3-arylimidazo[1,2-*a*]pyridines: Representative experimental procedure for the 2-ethoxy-3-phenylimidazo[1,2-*a*]pyridine:

(a) Conventional heating Condition:

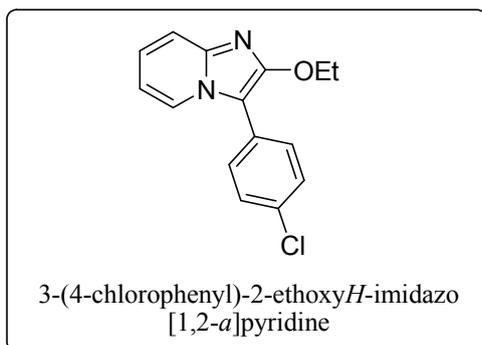
A mixture of β -nitrostyrene (1 mmol), 2-aminopyridine (1.3 mmol), ethanol (2 ml) and NiFe₂O₄ (10 mol %) refluxed for 4 hours under open atmosphere. After completion of the reaction (monitored by TLC) the reaction mixture was cooled to room temperature and extracted with ethyl acetate, washed with water with Brine solution. The organic phase was dried over anhydrous Na₂SO₄. The crude product was obtained by evaporation of solvent in vacuum which was purified by column chromatography over silica gel (60–120 mesh) using mixture of petroleum ether and ethyl acetate (9:1) as an eluting solvent to afford the pure product.

(b) MW irradiation Conditions:

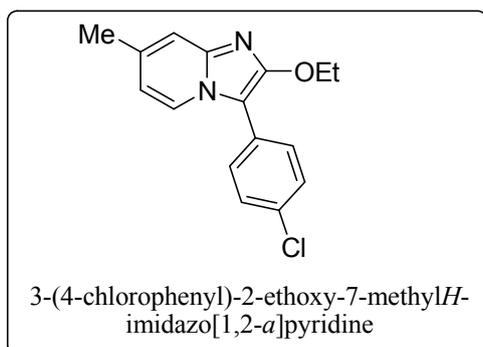
A mixture of β -nitrostyrene (1 mmol), 2-aminopyridine (1.3 mmol) and ethanol (2 ml) was taken in a sealed MW tube with a magnetic bar in presence of NiFe₂O₄ (10 mol %) and was irradiated at under MW condition (70 W) for 5 minutes. After that, the mixture was cooled to room temperature and the catalyst separated by a strong external magnet and finally, product was extracted with ethyl acetate, washed with water with Brine solution. The organic phase was dried over anhydrous Na₂SO₄. The crude product was obtained by evaporation of solvent in vacuum which was purified by column chromatography over silica gel (60–120 mesh) using mixture of petroleum ether and ethyl acetate (9:1) as an eluting solvent to afford pure 2-ethoxy-3-phenylimidazo[1,2-*a*]pyridine. This protocol was followed for all the products listed in Table 2. The products were confirmed by FT-IR, ¹H NMR and ¹³C NMR spectroscopy and elemental analysis. The spectroscopic data and elemental analysis of all the compounds has been given in S7.

ESI 7. Detailed spectral data of the 2-alkoxy-imidazopyridine derivatives listed in Table 2.

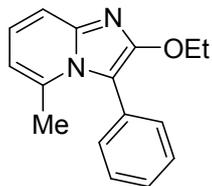
The ^1H and ^{13}C NMR spectra were recorded 300 MHz, 400 MHz or 500 MHz Bruker NMR spectrometer and CDCl_3 was used as solvent.



Gummy mass (entry 1, Table 2, Isolated yield 85%); ^1H NMR (300MHz, CDCl_3): δ 8.31 (d, $J = 3.0$ Hz, 1H), 7.32-7.27 (m, 4H), 7.23 (t, $J = 3.0$ Hz, 1H), 6.88 (t, $J = 3.0$ Hz, 1H), 6.56 (d, $J = 3.0$ Hz), 4.50-4.43(m, 2H), 1.45 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (75MHz, CDCl_3): δ 161.38, 161.02, 148.87, 137.69, 131.69, 131.64, 130.27, 129.21, 128.10, 118.22, 116.48, 63.08, 14.37; Anal calc for $\text{C}_{17}\text{H}_{17}\text{ClN}_2\text{O}$: C, 66.06; H, 4.80; N, 10.27 %; Found: C, 66.01; H, 4.85; N, 10.30 %.

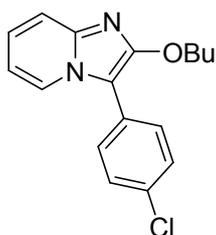


Gummy mass (entry 2, Table 2, Isolated yield 80%); ^1H NMR (300MHz, CDCl_3): δ 8.15 (d, $J = 5.1$ Hz, 1H), 7.18 (d, $J = 6.3$ Hz, 2H), 7.27 (d, $J = 6.3$ Hz, 2H), 6.73 (d, $J = 5.1$ Hz, 1H), 6.44 (s, 1H), 4.43 (q, $J = 6.9$ Hz, 2H), 2.22 (s, 3H), 1.43 (t, $J = 6.9$ Hz, 3H); ^{13}C NMR (75MHz, CDCl_3): δ 161.02, 149.16, 148.59, 139.65, 138.01, 136.40, 130.66, 130.17, 128.41, 119.77, 116.78, 63.16, 21.09, 14.37; Anal calc for $\text{C}_{16}\text{H}_{15}\text{ClN}_2\text{O}$: C, 67.02; H, 5.27; N, 9.75 %; Found: C, 66.91; H, 5.30; N, 9.79 %.



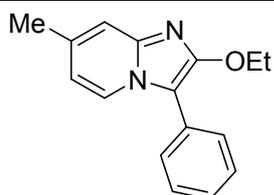
2-ethoxy-5-methyl-3-phenyl*H*-imidazo
[1,2-*a*]pyridine

Gummy mass (entry 3, Table 2, Isolated yield 85%); ¹H NMR (300MHz, CDCl₃): δ 7.32-7.25 (m, 4H), 7.21-7.16 (m, 2H), 6.73 (d, *J* = 6.0 Hz 1H), 6.25 (d, *J* = 8.1 Hz 1H), 4.48-4.41 (m, 2H), 2.45 (s, 3H), 1.41 (t, *J* = 6.9 Hz, 3H); ¹³C NMR (75MHz, CDCl₃): δ 160.86, 160.81, 157.72, 137.90, 131.66, 130.21, 129.23, 128.01, 117.50, 113.09, 62.98, 24.48, 14.36. Anal calc for C₁₆H₁₆N₂O: C, 76.16; H, 6.39; N, 11.10 %; Found: C, 76.12; H, 6.43; N, 11.16 %.



2-butoxy-3-(4-chlorophenyl)*H*-imidazo
[1,2-*a*]pyridine

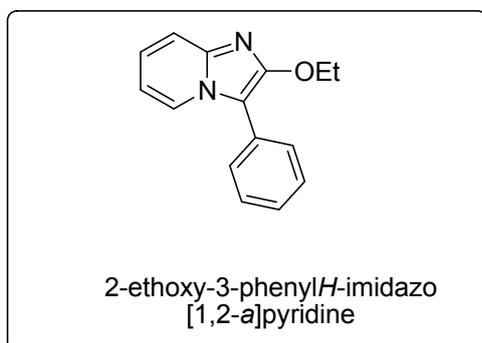
Gummy mass (Table 2, entry 4, yield 84%); ¹H NMR (400MHz, CDCl₃): δ 8.31 (d, *J* = 4.8Hz, 1H), 7.53-7.49 (m, 1H), 7.26-7.18 (m, 4H), 6.92 (t, *J* = 5.2 Hz, 1H), 4.41-4.10 (t, *J* = 6.4 Hz, 2H), 1.82-1.76 (m, 2H), 1.53-1.47 (m, 2H), 1.00-0.97 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100MHz, CDCl₃): δ 161.31, 157.75, 148.82, 138.60, 137.67, 134.40, 132.31, 129.17, 128.10, 118.20, 116.47, 67.44, 30.85, 21.17, 14.45. Anal calc for C₁₇H₁₇ClN₂O: C, 67.88; H, 5.70; N, 9.31 %; Found: C, 67.3; H, 5.75; N, 9.27 %.



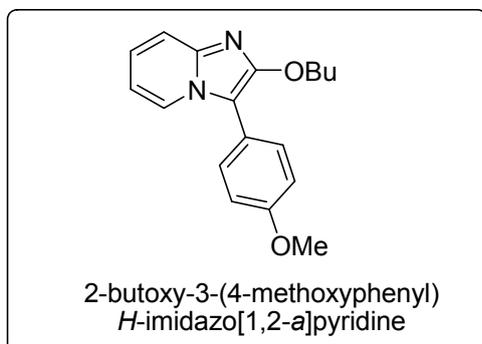
2-ethoxy-7-methyl-3-phenyl*H*-imidazo
[1,2-*a*]pyridine

Gummy mass (Table 2, entry 5, Isolated yield- 85%); ¹H NMR (500MHz, CDCl₃): δ 8.16 (d, *J* = 5.0 Hz, 1H), 7.33-7.26 (m, 3 H), 7.23-7.20 (m, 2H), 6.71-6.70 (d, *J* = 5.0 Hz 1H), 6.412 (s, 1H), 4.47-4.43 (m, 2H), 2.20(s, 1H), 1.45-1.43 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (125MHz, CDCl₃): δ 161.25, 160.70, 149.04, 148.30, 131.78, 130.25, 129.22, 128.94, 127.36, 119.53, 116.92, 63.03, 21.04, 14.39. Anal calc for C₁₆H₁₆N₂O:

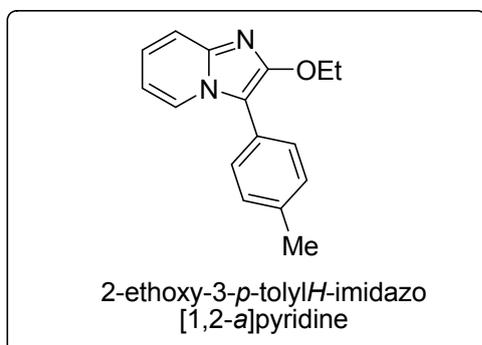
C, 76.16; H, 6.39; N, 11.10 %; Found: C, 76.12; H, 6.44; N, 11.16 %.



Gummy mass (Table 2, entry 6, Isolated yield- 85%); ¹H NMR (500MHz, CDCl₃): δ 8.33 (d, *J* = 4.0 Hz, 1H), 7.47 (t, *J* = 6.0 Hz, 1 H), 7.33-7.21 (m, 5H), 6.89 (d, *J* = 5.0 Hz, 1H), 6.56 (d, *J* = 8.0 Hz, 1H), 4.50-4.46 (m, 2H), 1.27 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (75MHz, CDCl₃): δ 161.37, 160.98, 148.86, 147.92, 131.67, 130.26, 129.19, 128.90, 127.37, 120.00, 116.46, 63.06, 14.35. Anal calc for C₁₅H₁₄N₂O: C, 75.65; H, 6.92; N, 11.76 %; Found: C, 75.60; H, 5.95; N, 11.82 %.

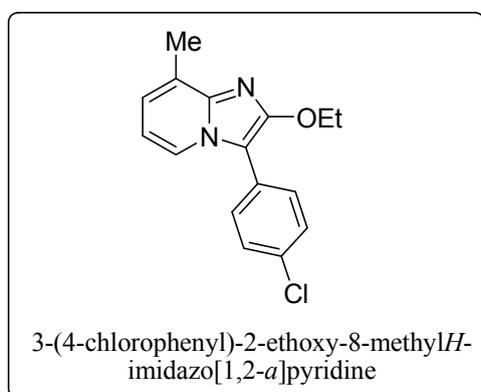


Gummy mass (Table 2, entry 7, Isolated yield- 84%); ¹H NMR (500MHz, CDCl₃): δ 8.34 (d, *J* = 4.0 Hz, 1H), 7.49 (t, *J* = 8.0 Hz, 1H), 7.26 (t, *J* = 4.5 Hz, 1H), 6.89 (d, *J* = 5.5 Hz, 1H), 6.72 (d, *J* = 8.5 Hz, 1H), 6.57 (d, *J* = 8.0 Hz, 1H), 4.38 (t, *J* = 6.5 Hz, 2H), 1.84-1.78(m, 2H), 1.55-1.49 (m, 2H), 1.007-0.978 (t, 3H); ¹³C NMR (125MHz, CDCl₃): δ 165.95, 161.31, 148.56, 144.79, 137.73, 133.67, 130.61, 128.55, 128.49, 118.48, 118.43, 116.57, 67.48, 30.80, 21.16, 19.61. Anal calc for C₁₈H₂₀N₂O₂: C, 72.95; H, 6.80; N, 9.45 %; Found: C, 72.91; H, 6.77; N, 9.40 %.

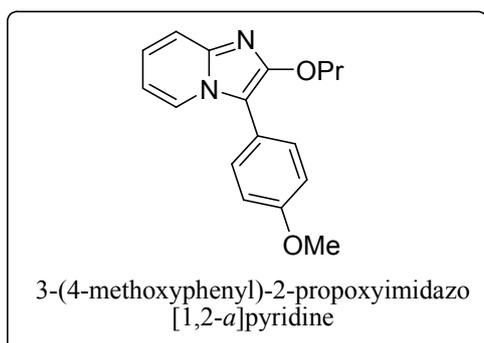


Gummy mass (Table 2, entry 8, Isolated yield- 86%); ¹H NMR (500MHz, CDCl₃): δ 8.17 (d, *J* = 5.0

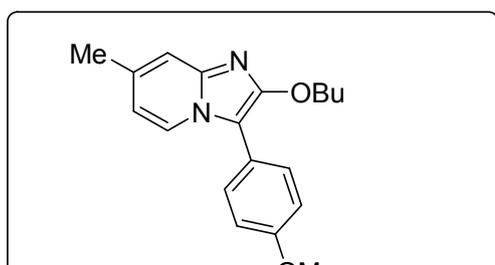
Hz, 1H), 7.28 (t, $J = 8.0$ Hz, 2H), 7.2 (d, $J = 8.0$ Hz, 1H), 7.00 (d, $J = 8.0$ Hz, 1H), 6.88 (d, $J = 5.0$ Hz, 1H), 6.71 (d, $J = 5.0$ Hz, 1H), 4.447-4.404 (m, 2H), 2.25 (s, 3H), 1.42 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 161.28, 149.16, 148.57, 138.46, 138.01, 135.19, 131.61, 130.28, 128.56, 119.33, 116.68, 63.66, 21.13, 14.21. Anal. calc. for $\text{C}_{16}\text{H}_{16}\text{N}_2\text{O}$: C, 76.16; H, 6.39; N, 11.10 %; Found: C, 76.12; H, 6.44; N, 11.06 %.



Gummy mass (Table 2, entry 9, Isolated yield- 81%); ^1H NMR (500 MHz, CDCl_3): δ 7.99 (d, $J = 6.5$ Hz, 1H), 7.91 (d, $J = 8.5$ Hz, 2H), 7.40 (d, $J = 8.5$ Hz, 2H), 6.96 (d, $J = 6.5$ Hz, 1H), 6.70 (t, $J = 6.5$ Hz, 1H), 4.15-4.11 (m, 2H), 2.67 (s, 3H), 1.27 (t, $J = 9.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 161.17, 149.29, 148.55, 139.21, 137.82, 135.90, 130.68, 130.07, 128.63, 119.17, 116.66, 63.93, 21.01, 14.67; Anal. calc. for $\text{C}_{16}\text{H}_{15}\text{ClN}_2\text{O}$: C, 67.02; H, 5.27; N, 9.77 %; Found: C, 66.93; H, 5.23; N, 9.80 %.

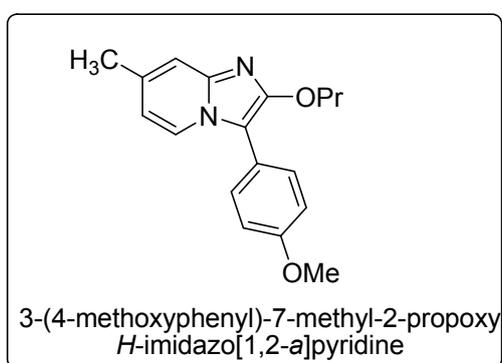


Gummy mass (Table 2, entry 10, Isolated yield- 80%); ^1H NMR (500 MHz, CDCl_3): δ 8.30 (d, $J = 6.5$ Hz, 1H), 7.48 (t, $J = 6.5$ Hz, 1H), 7.25 (d, $J = 8.5$ Hz, 1H), 6.92 (d, $J = 6.5$ Hz, 2H), 6.74 (d, $J = 6.5$ Hz, 2H), 6.55 (d, $J = 6.5$ Hz, 1H), 4.41-4.35 (m, 2H), 3.76 (s, 3H), 1.85-1.67 (m, 2H), 1.00 (t, $J = 9.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 171.28, 148.77, 142.06, 137.89, 136.91, 131.12, 129.11, 128.41, 118.06, 116.53, 113.48, 67.04, 60.51, 21.15, 14.30. Anal. calc. for $\text{C}_{16}\text{H}_{15}\text{ClN}_2\text{O}$: C, 72.32; H, 6.43; N, 9.92 %; Found: C, 72.29; H, 6.46; N, 9.96 %.



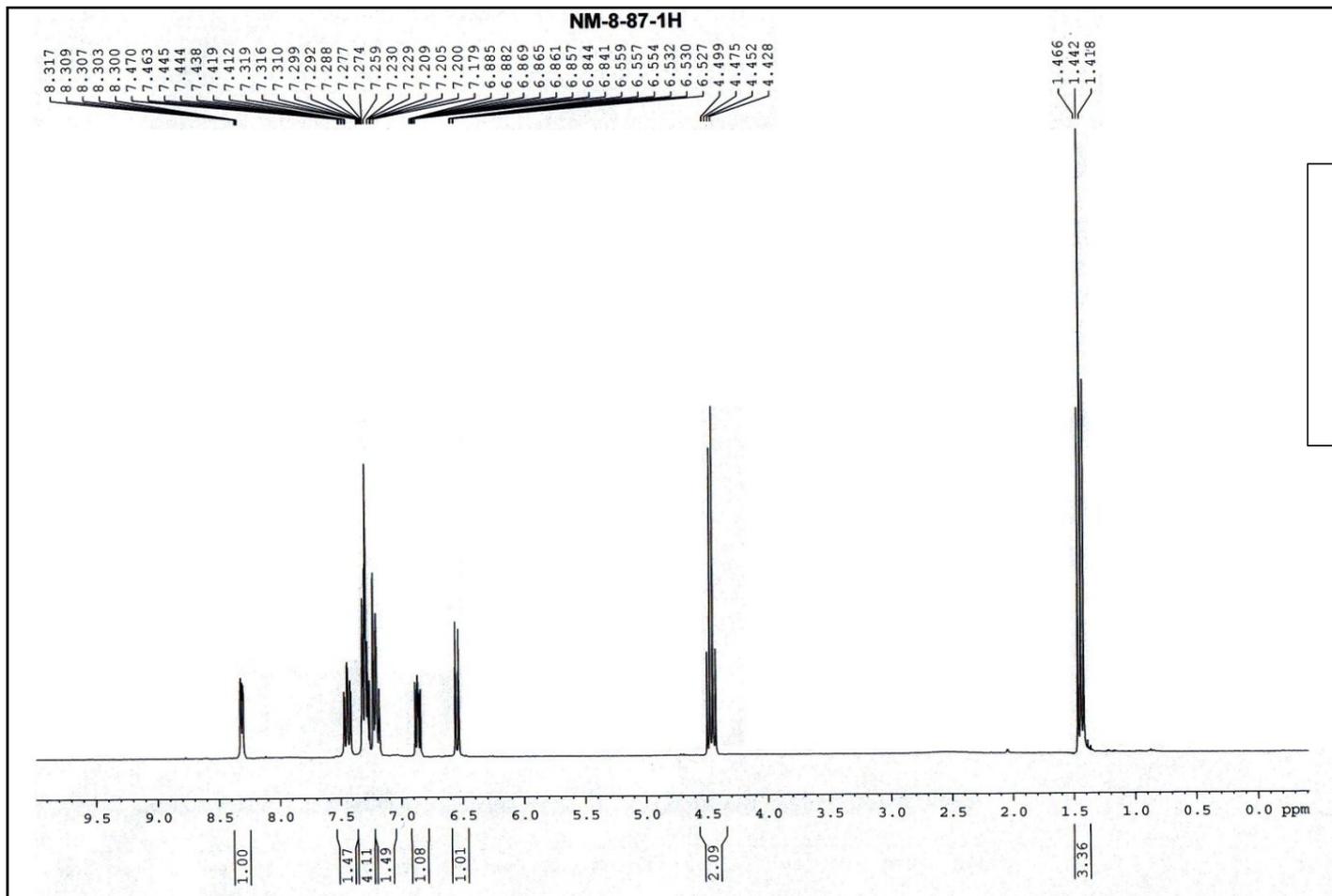
Gummy mass (Table 2, entry 11, Isolated yield- 86%); ^1H NMR (300 MHz, CDCl_3): δ 8.38 (d, $J = 5.1$

Hz, 1H), 7.65 (d, $J = 6.3$ Hz, 2H), 7.58 (d, $J = 6.3$ Hz, 2H), 6.68 (d, $J = 5.1$ Hz, 1H), 6.46 (s, 1H), 4.46 (q, $J = 6.9$ Hz, 2H), 3.85 (s, 3H), 2.31 (s, 3H), 1.76 (m, 2H), 1.5 (m, 1H), 0.91 (t, 3H); ^{13}C NMR (75MHz, CDCl_3): δ 161.15, 148.07, 147.17, 131.14, 129.13, 125.14, 119.43, 118.13, 113.50, 62.95, 55.39, 55.38, 29.84, 21.17, 14.43; Analcalc for $\text{C}_{19}\text{H}_{22}\text{N}_2\text{O}_2$: C, 73.52; H, 7.14; N, 9.03 %; Found: C, 73.44; H, 7.19; N, 9.09 %.



Gummy mass (Table 2, entry 12, Isolated yield- 83%); ^1H NMR (300MHz, CDCl_3): δ 8.18 (d, $J = 4.2$ Hz, 1H), 7.30 (d, $J = 4.8$ Hz, 2H), 7.25 (d, $J = 4.8$ Hz, 2H), 6.69 (d, $J = 5.1$ Hz, 1H), 6.46 (s, 1H), 4.43 (q, $J = 7.2$ Hz, 2H), 3.75 (s, 3H), 2.23 (s, 3H), 1.71 (m, 2H), 0.89 (t, $J = 6.6$ Hz, 3H); ^{13}C NMR (75MHz, CDCl_3): δ 160.98, 148.86, 147.92, 131.64, 130.23, 129.19, 128.92, 128.08, 120.01, 116.43, 60.54, 30.85, 21.18, 19.11, 14.34; Analcalc for $\text{C}_{18}\text{H}_{20}\text{N}_2\text{O}_2$: C, 72.95; H, 6.80; N, 9.45 %; Found: C, 72.91; H, 6.76; N, 9.49.

ESI 8. Copy of ^1H NMR and ^{13}C NMR spectra of all compounds:

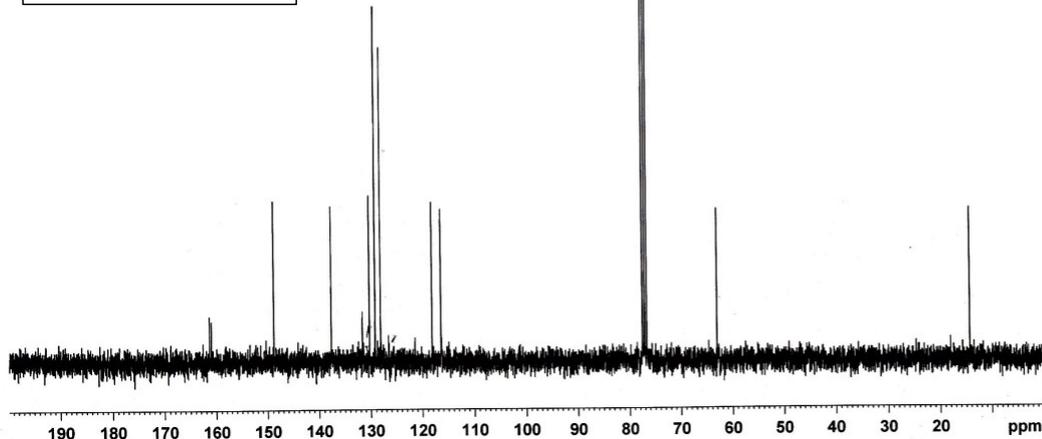
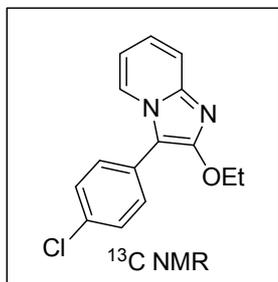


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76.74
63.08

14.37



Current Data Parameters
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PROCNO 1

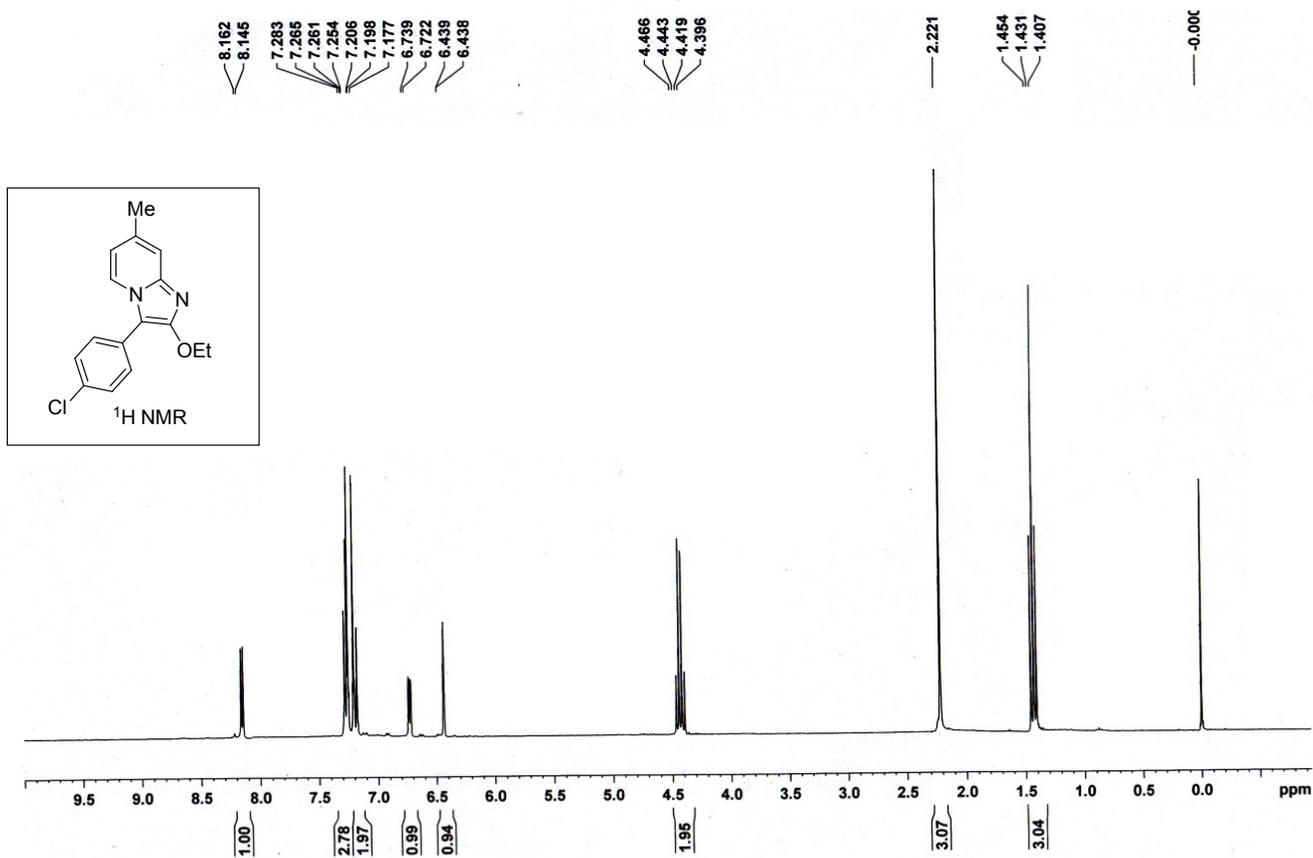
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TE 300.0 K
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D11 0.03000000 sec
TD0 1

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----- CHANNEL f2 -----
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NM-S-81-1H



NM-S-81-13C

Current Data Parameters
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PROCNO 1

F2 - Acquisition Parameters
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AQ 0.7208960 sec
RG 210.78
DW 22.000 usec
DE 6.50 usec
TE 298.7 K
D1 2.00000000 sec
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TD0 1

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SFO1 75.4752949 MHz
NUC1 13C
P1 9.25 usec
PLW1 34.00000000 W

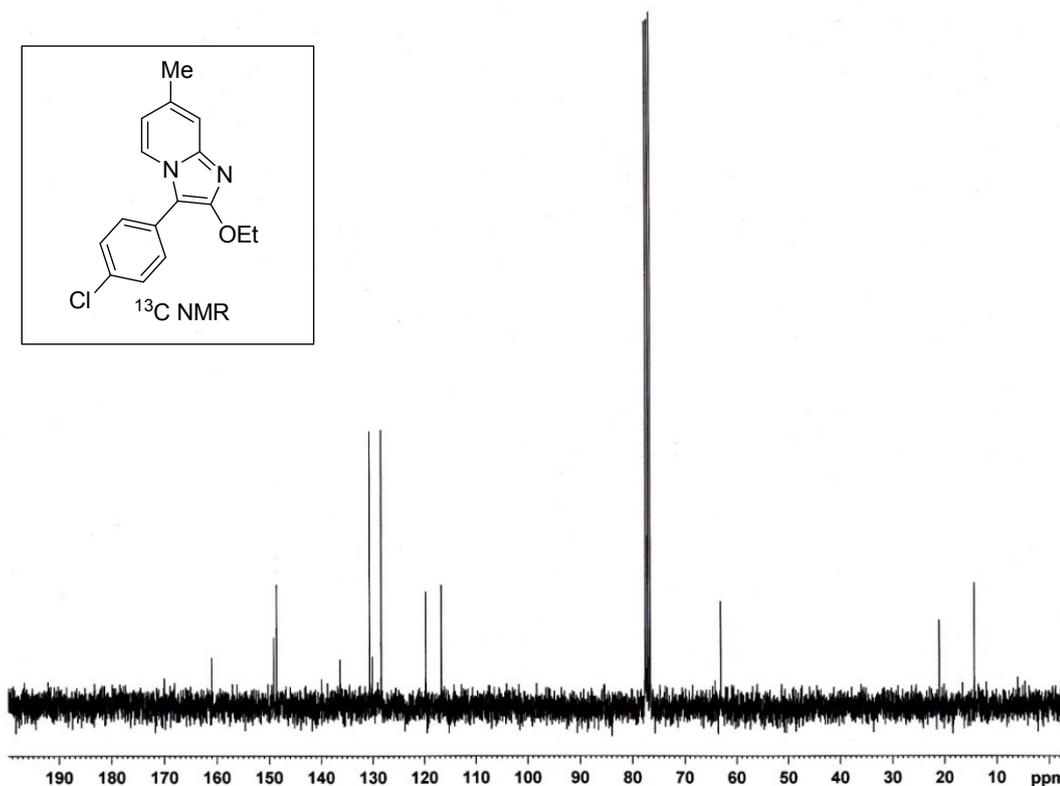
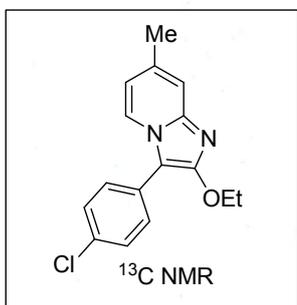
===== CHANNEL f2 =====
SFO2 300.1312005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 6.99840021 W
PLW12 0.17545000 W
PLW13 0.14211001 W

F2 - Processing parameters
SI 32768
SF 75.4677382 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

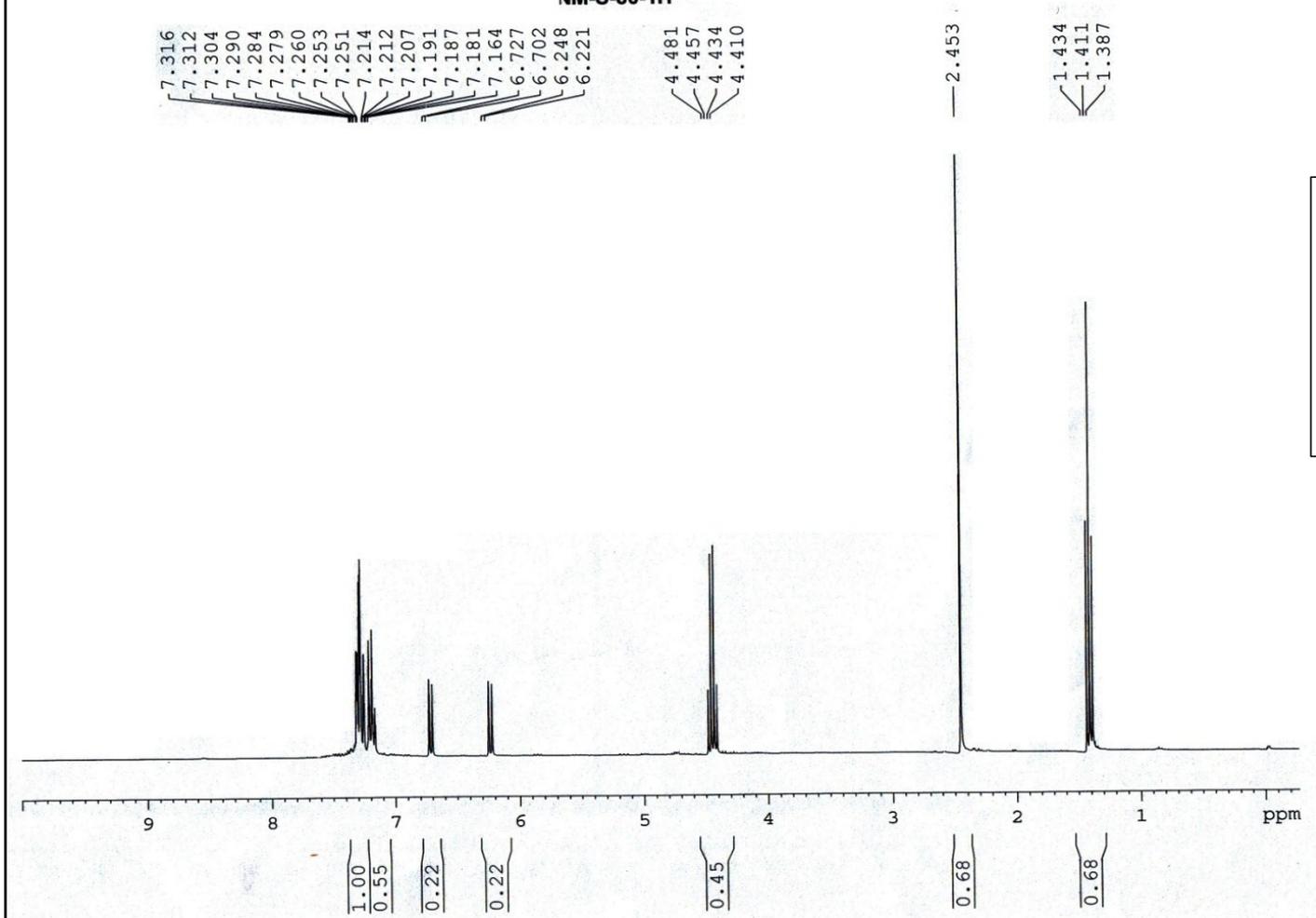
161.02
149.16
148.59
136.40
130.66
130.17
128.41
119.77
116.78

77.58
77.16
76.74
63.16

21.09
14.37



NM-S-86-1H



Me



NM-S-86-13C

160.86
160.81
157.72

137.90
131.66
130.21
129.23
128.01

117.50
113.09

77.59
77.16
76.74

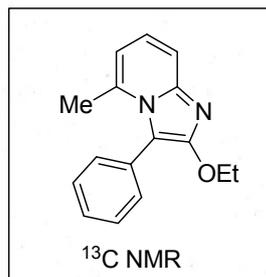
62.98

24.48

14.36

Current Data Parameters
NAME NM-S-86-13C
EXPNO 1
PROCNO 1

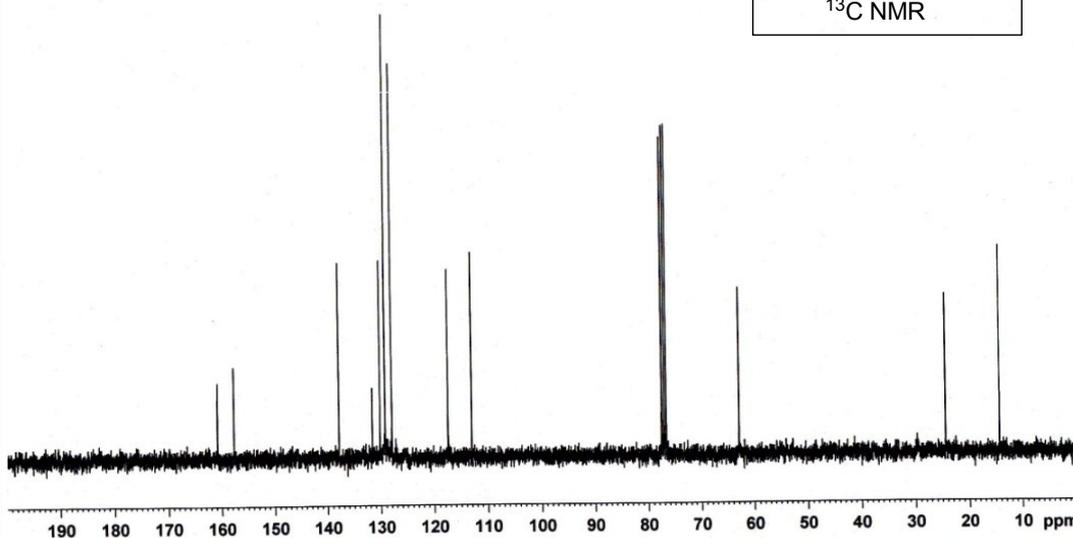
F2 - Acquisition Parameters
Date_ 20150302
Time 15.12
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 63
DS 4
SWH 22727.273 Hz
FIDRES 0.693581 Hz
AQ 0.7208960 sec
RG 210.78
DW 22.000 usec
DE 6.50 usec
TE 298.7 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1



===== CHANNEL f1 =====
SFO1 75.4752949 MHz
NUC1 13C
P1 9.25 usec
PLW1 34.00000000 W

===== CHANNEL f2 =====
SFO2 300.1312005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 6.99840021 W
PLW12 0.17545000 W
PLW13 0.14211001 W

F2 - Processing parameters
SI 32768
SF 75.4677418 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



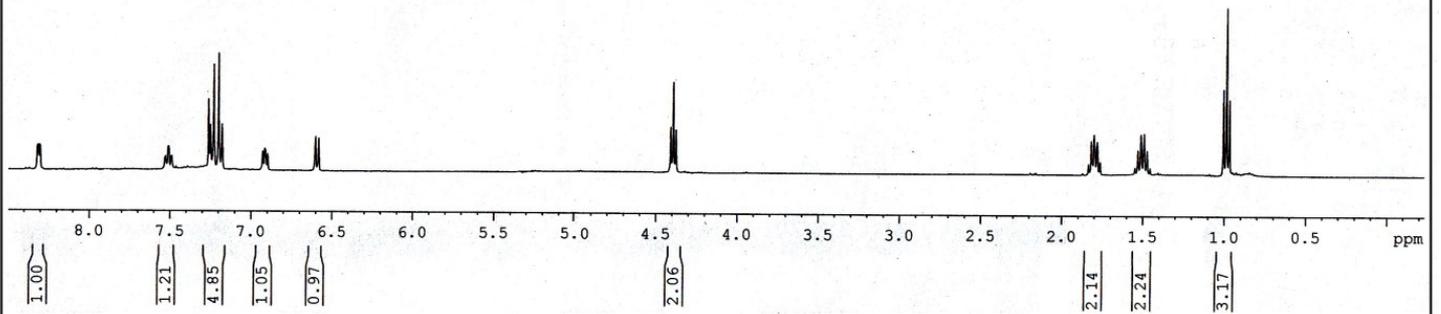
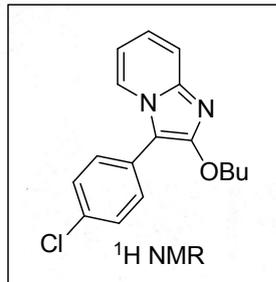
NM-S-94-1H (ND) 400MHz.

8.316
8.314
8.304
7.531
7.527
7.509
7.492
7.489
7.260
7.250
7.228
7.198
7.180
7.176
6.927
6.914
6.896
6.601
6.582

4.405
4.389
4.372

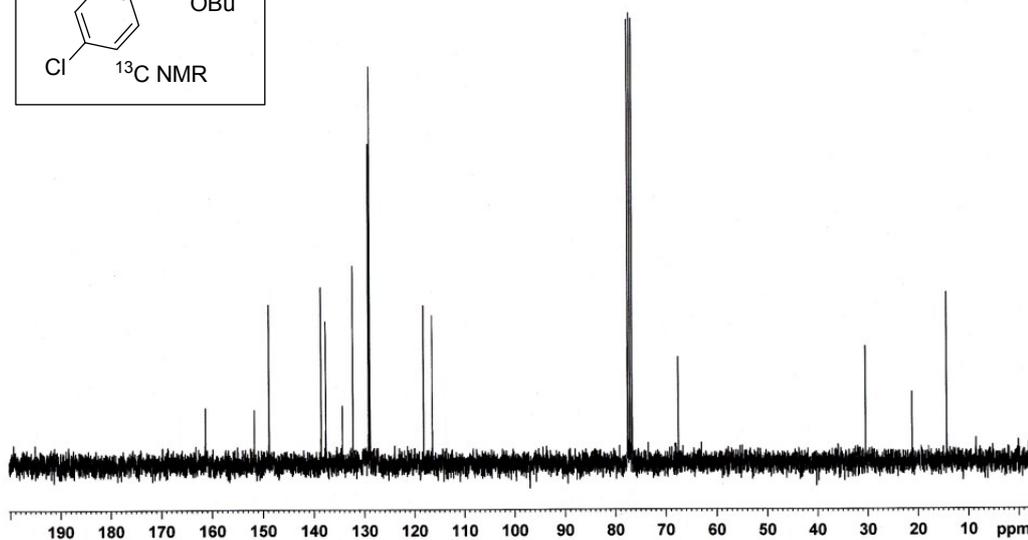
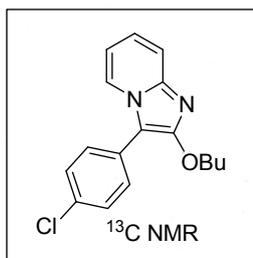
1.818
1.801
1.781
1.764
1.530
1.511
1.492
1.474

1.004
0.986
0.967



NM-S-94-13C

161.31
161.75
148.82
138.60
137.67
134.40
132.31
129.17
128.10
118.20
116.47
77.59
77.16
76.74
67.44
30.85
21.17
14.45



Current Data Parameters
NAME NM-S-94-13C
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150325
Time 11.49
INSTRUM spect
PROBHD 5 mm PABBO BB-
FULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 52
DS 4
SWH 22727.273 Hz
FIDRES 0.693581 Hz
AQ 0.7208960 sec
RG 210.78
DW 22.000 usec
DE 6.50 usec
TE 299.8 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 75.4752949 MHz
NUC1 13C
P1 9.25 usec
PLW1 34.00000000 W

===== CHANNEL f2 =====
SFO2 300.1312005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 6.99840021 W
PLW12 0.17545000 W
PLW13 0.14211001 W

F2 - Processing parameters
SI 32768
SF 75.4677382 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

NM-S-83-1H(SR) 500MHz

8.164
8.154

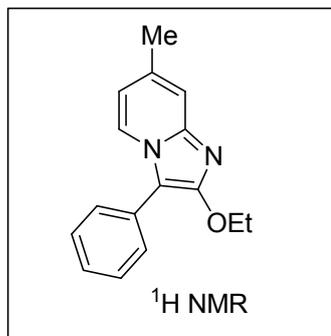
7.333
7.317
7.301
7.287
7.261
7.230
7.215

7.199
6.708
6.698
6.412

4.469
4.455
4.441
4.426

2.202
2.187

1.453
1.439
1.425



9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 ppm

1.00

3.28

2.40

0.96

0.96

2.03

3.39

3.16

NM-S-83-13C-240315-7- (ND)

149.04
148.30
131.78
130.25
129.22
129.13
128.94
128.80
128.08
127.36
119.53
116.92

77.47
77.16
76.84

63.03

21.04
14.39

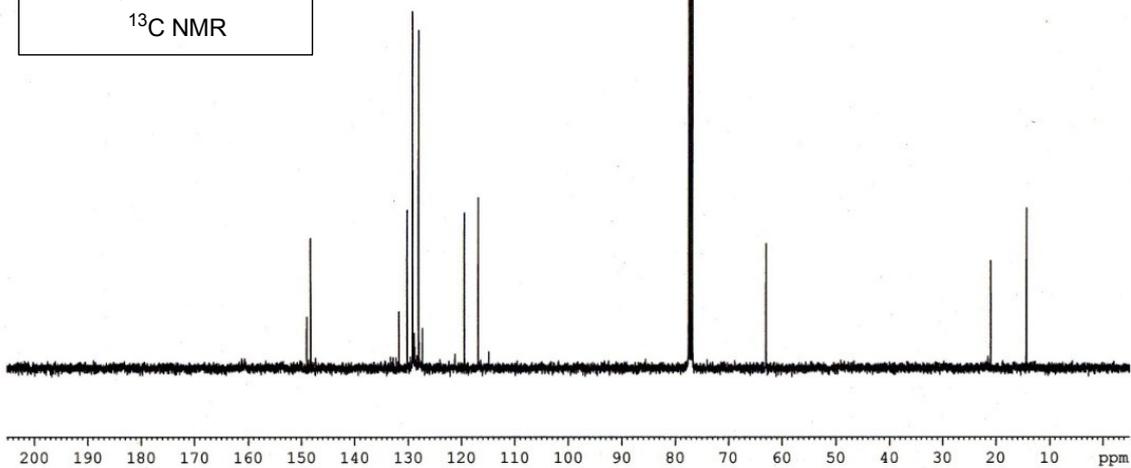
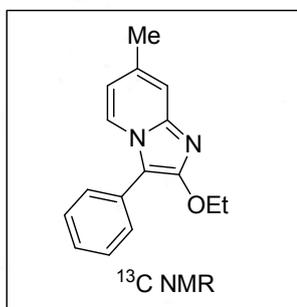
Current Data Parameters
NAME Mar24-2015
EXPNO 7
PROCNO 1

F2 - Acquisition Parameters
Date 20150325
Time 10.53
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 421
DS 4
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 0.6815744 sec
RG 203
DW 20.800 usec
DE 6.50 usec
TE 0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

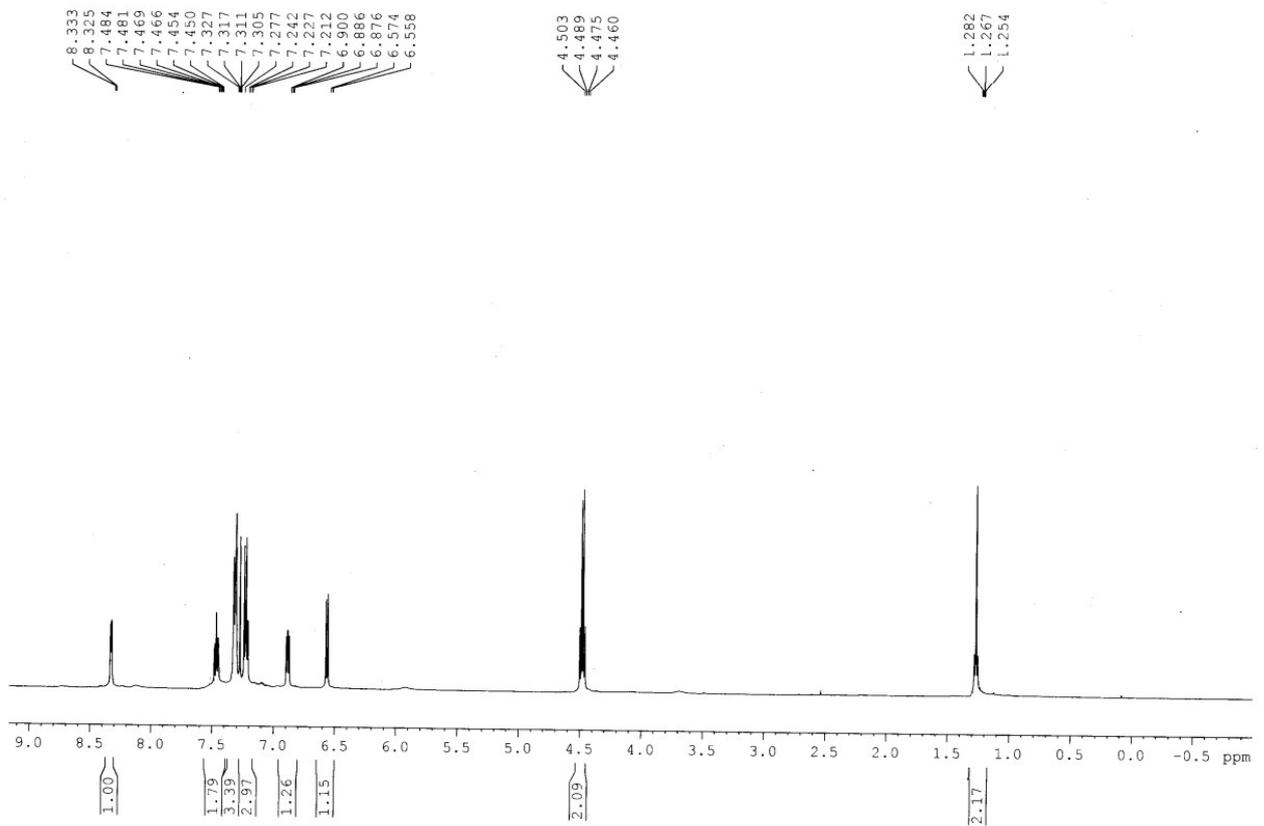
===== CHANNEL f1 =====
SFO1 100.6404326 MHz
NUC1 13C
P1 9.00 usec
PLW1 55.00000000 W

===== CHANNEL f2 =====
SFO2 400.2016008 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 10.00000000 W
PLW12 0.29660001 W
PLW13 0.24025001 W

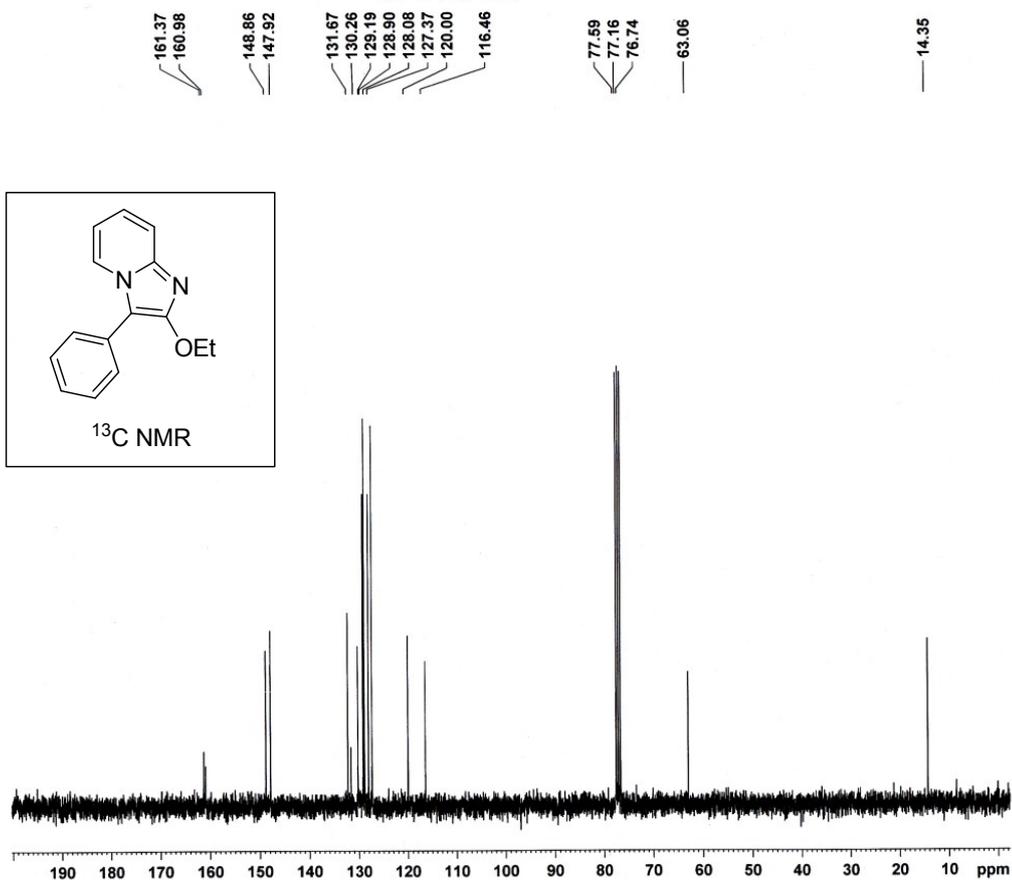
F2 - Processing parameters
SI 32768
SF 100.6303569 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



SJ-S-119-1H(SR) 500MHz



NM-S-88-13C



Current Data Parameters
NAME NM-S-88-13C
EXPNO 1
PROCNO 1

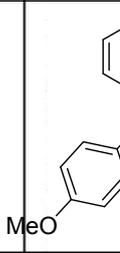
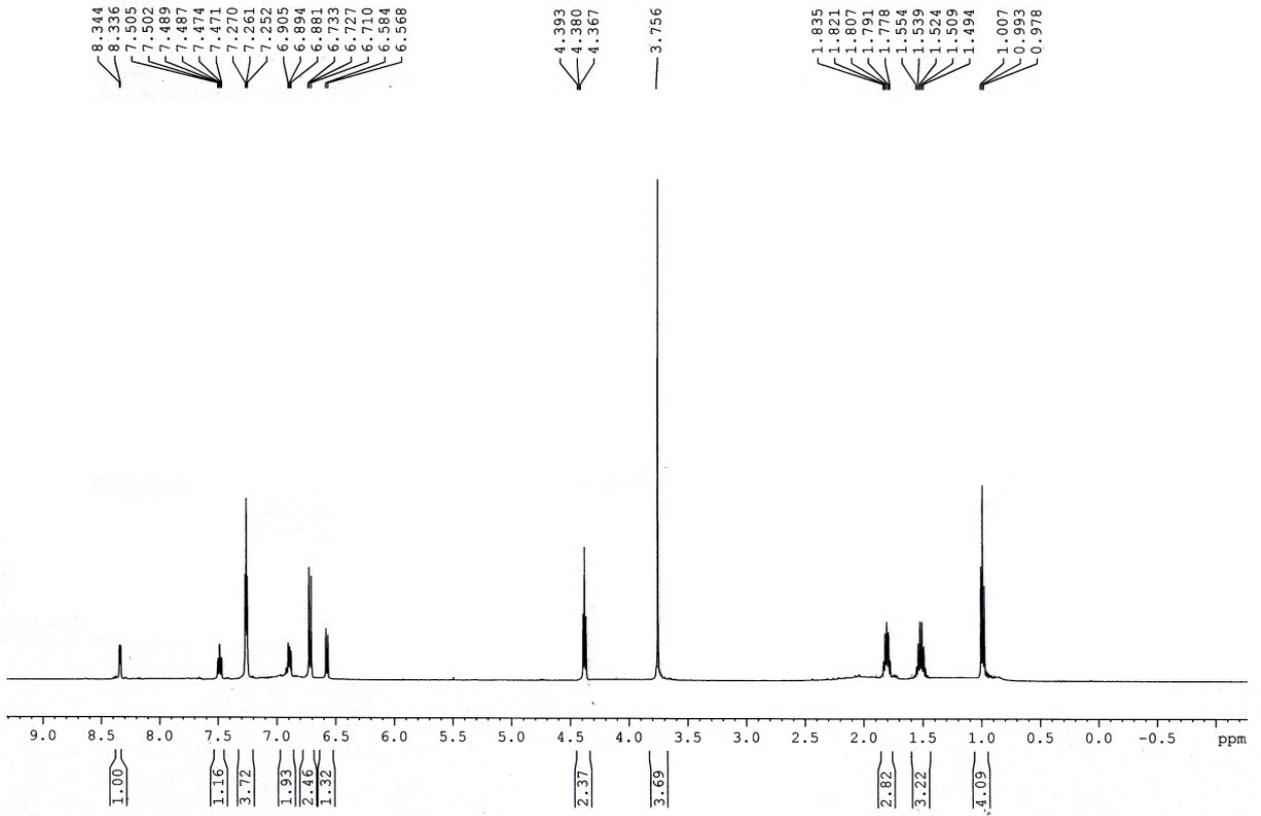
F2 - Acquisition Parameters
Date_ 20150302
Time 15.20
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 84
DS 4
SWH 22727.273 Hz
FIDRES 0.693581 Hz
AQ 0.7208960 sec
RG 210.78
DW 22.000 usec
DE 6.50 usec
TE 298.8 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1

----- CHANNEL f1 -----
SFO1 75.4752949 MHz
NUC1 13C
P1 9.25 usec
PLW1 34.00000000 W

----- CHANNEL f2 -----
SFO2 300.1312005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 6.99840021 W
PLW12 0.17545000 W
PLW13 0.14211001 W

F2 - Processing parameters
SI 32768
SF 75.4677416 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

PM-S-114-1H (SR) 500MHz.



PM-S-113-1H(SR) 500MHz.

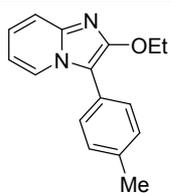
8.171
8.161

7.296
7.280
7.261
7.225
7.209
7.022
7.006
6.892
6.882
6.714
6.704

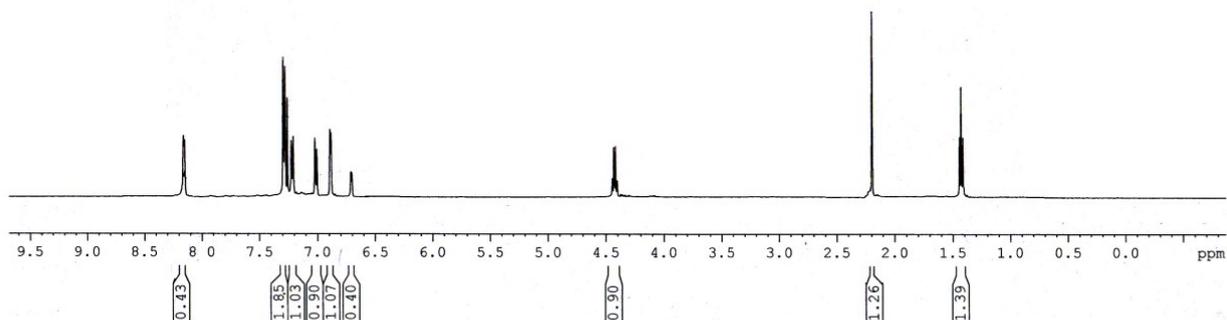
4.447
4.433
4.419
4.404

2.257
2.246

1.441
1.427
1.413
1.395
1.381
1.367



1H NMR



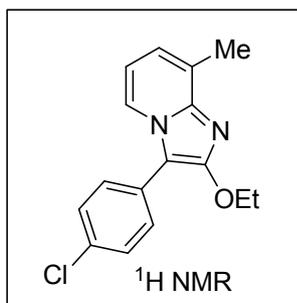
NM-S-82-1H (SR) 500MHz

7.994
7.981
7.917
7.900
7.406
7.389
7.261
6.970
6.957
6.708
6.695
6.681

4.149
4.131
4.113
4.106

2.675
2.653

1.575
1.284
1.274
1.256



8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 ppm

1.01
1.80
1.00

2.86

1.77

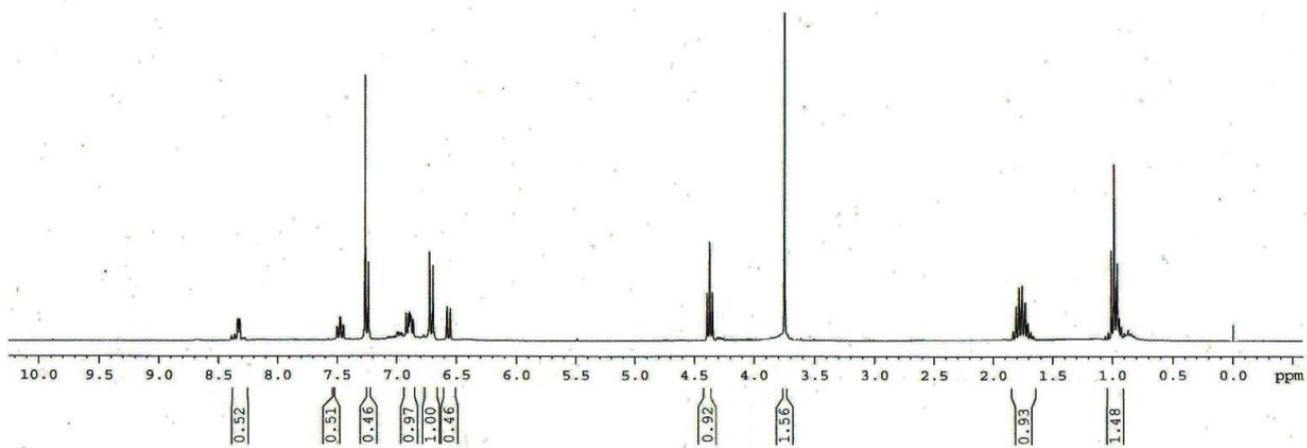
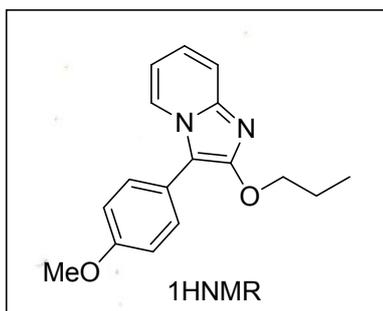
1.31

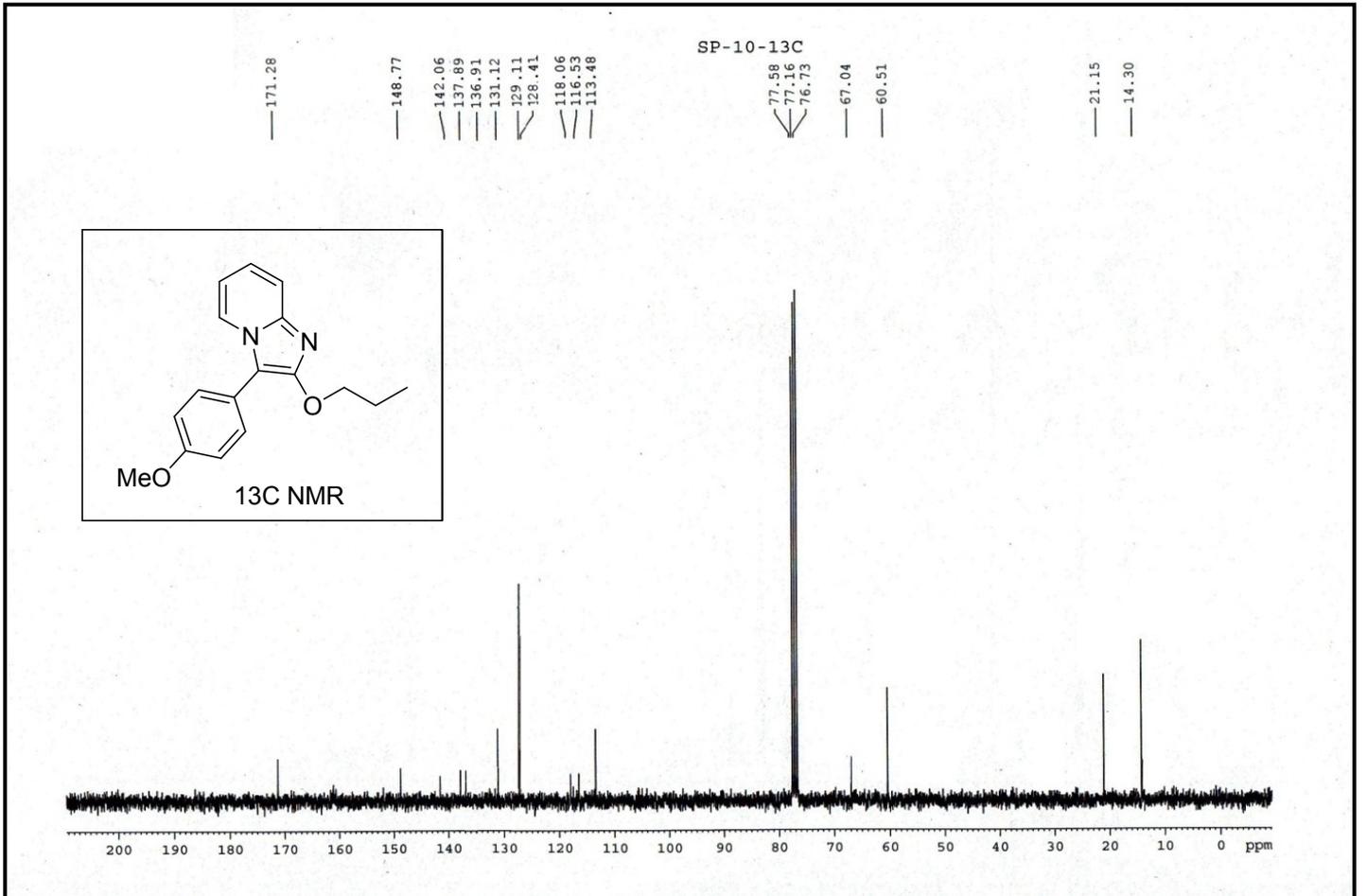
2.26

3.37

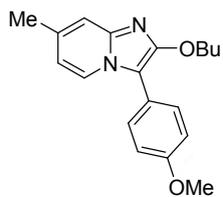
9.30

3.34

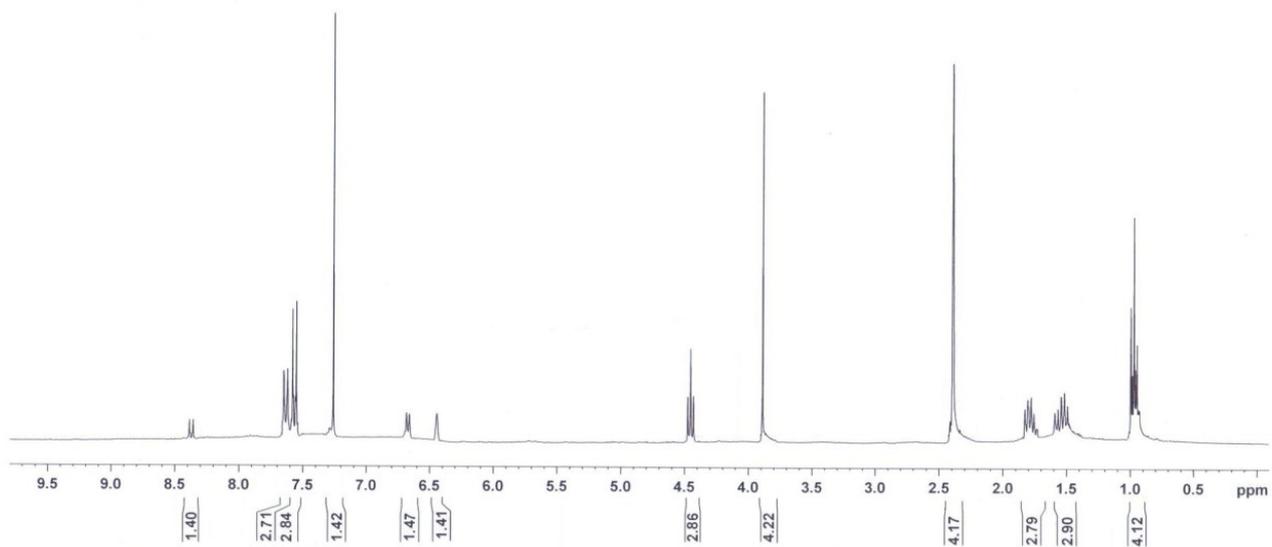




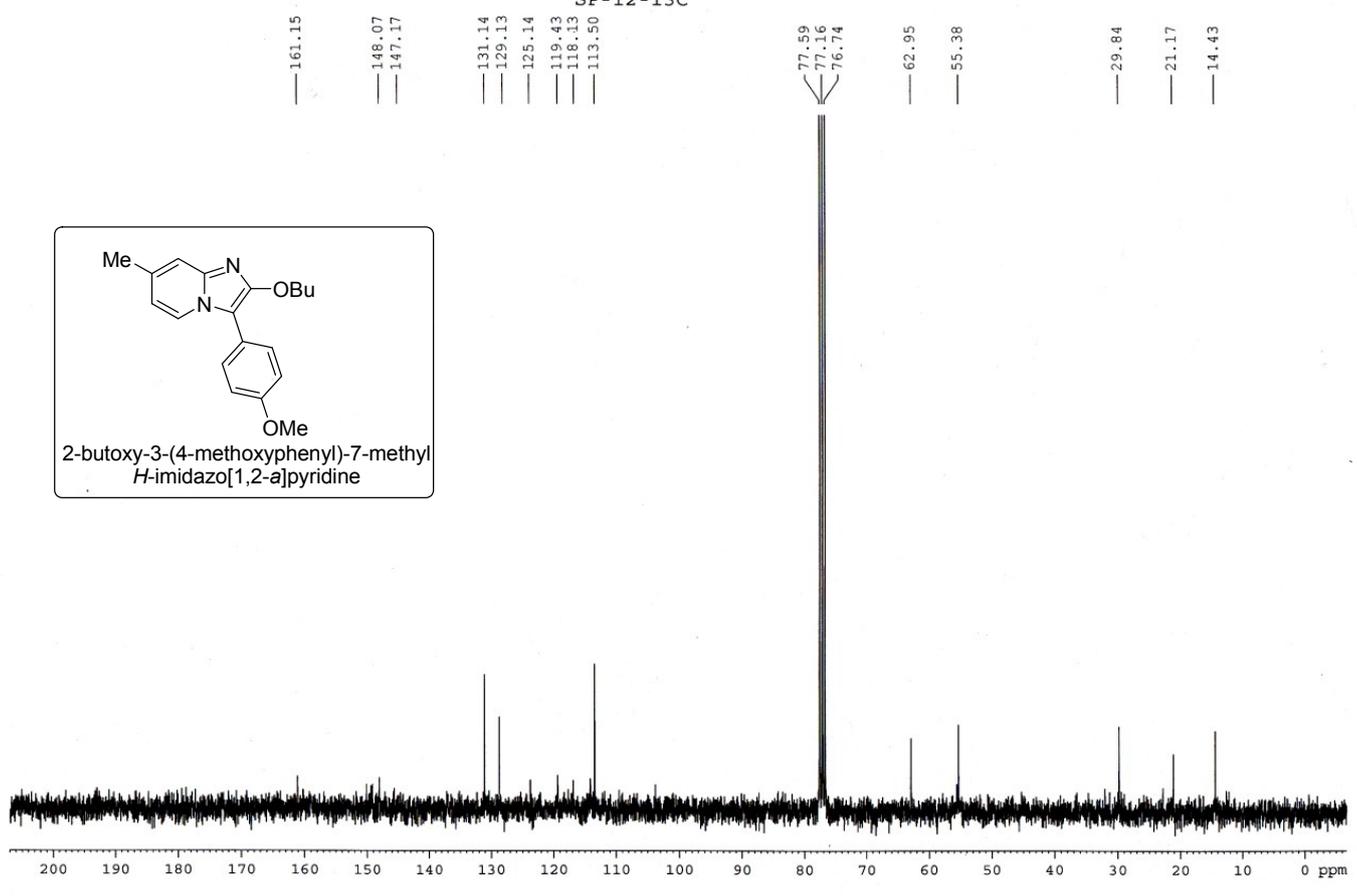
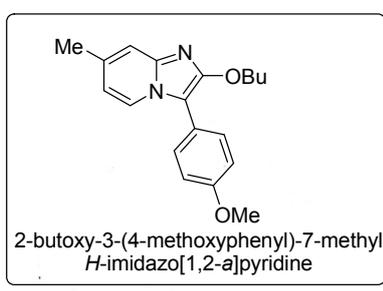
S-136-1H



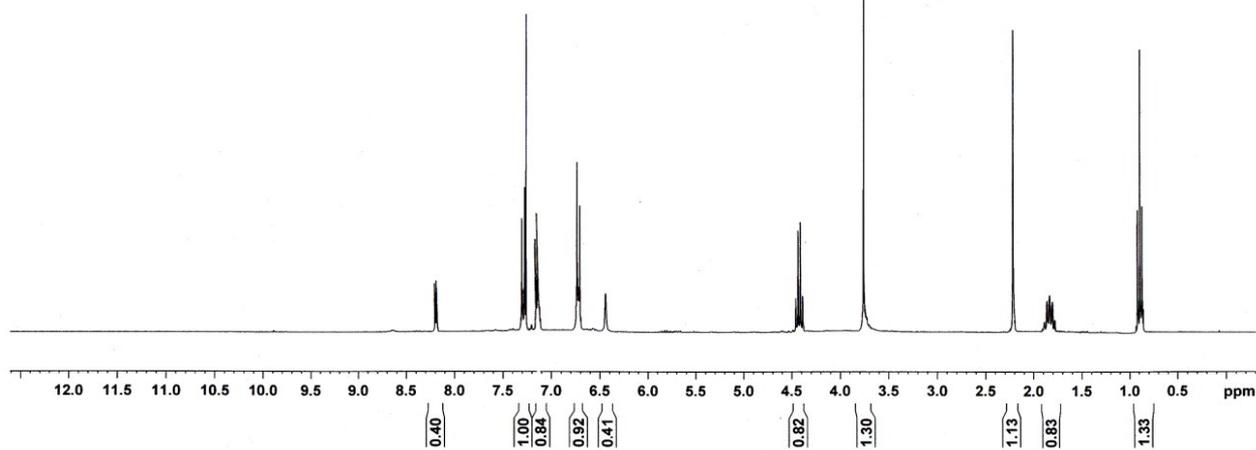
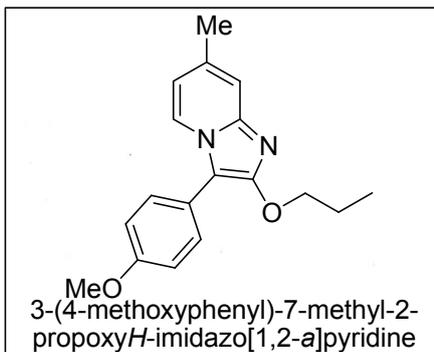
2-butoxy-3-(4-methoxyphenyl)-7-methyl
H-imidazo[1,2-*a*]pyridine



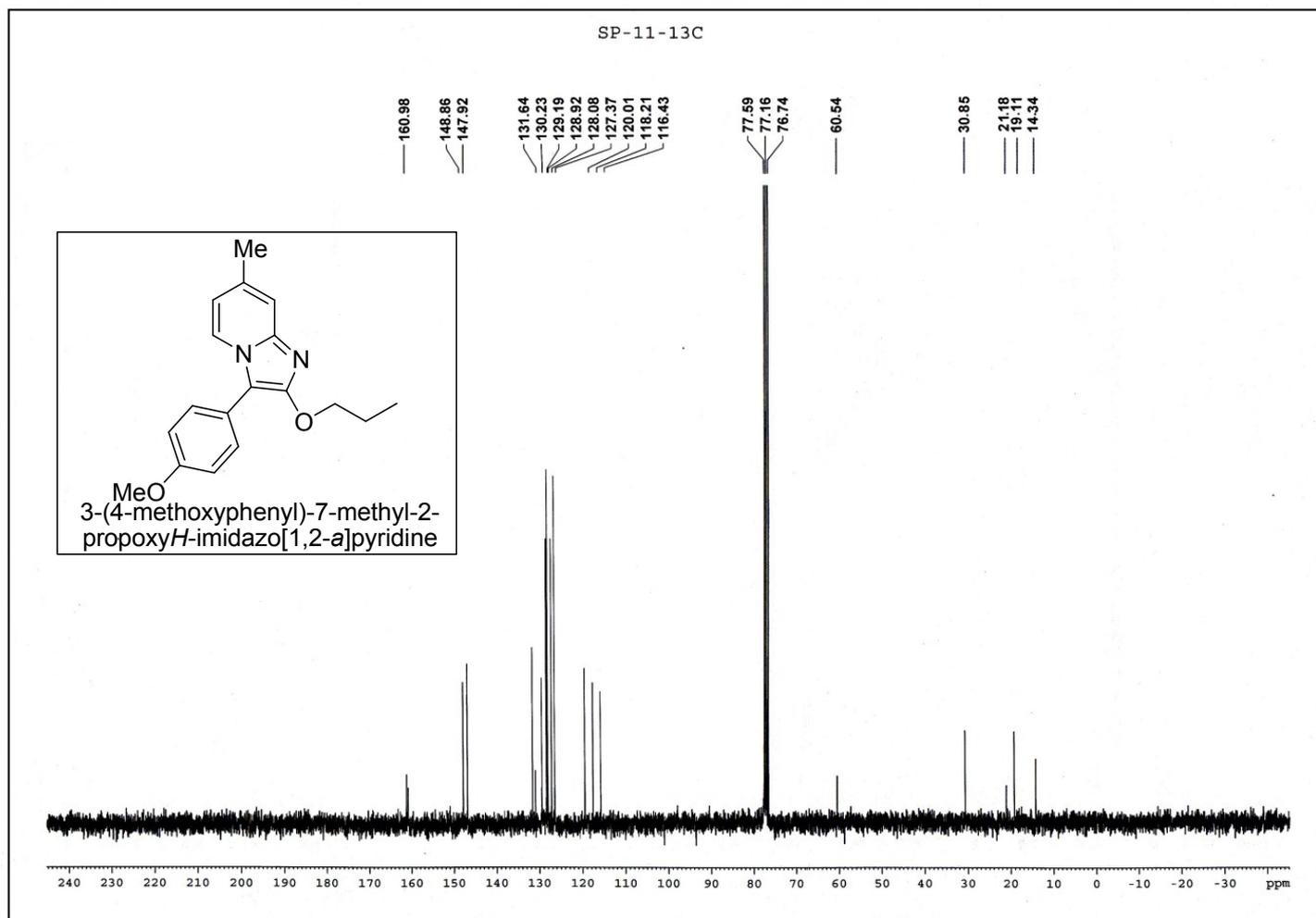
SP-12-13C



SP-12-1H



SP-11-13C



ESI9. Recyclability of Catalyst:

We have examined reusability of the catalyst for the synthesis 2-ethoxy-3-phenylimidazopyridine. After each cycle nano-NiFe₂O₄ was recovered simply by using an external magnet, washed with ethanol and reused for subsequent reactions. The magnetic nano-catalyst was reused for eight times with minimum loss of catalytic activity (Figure 5S).

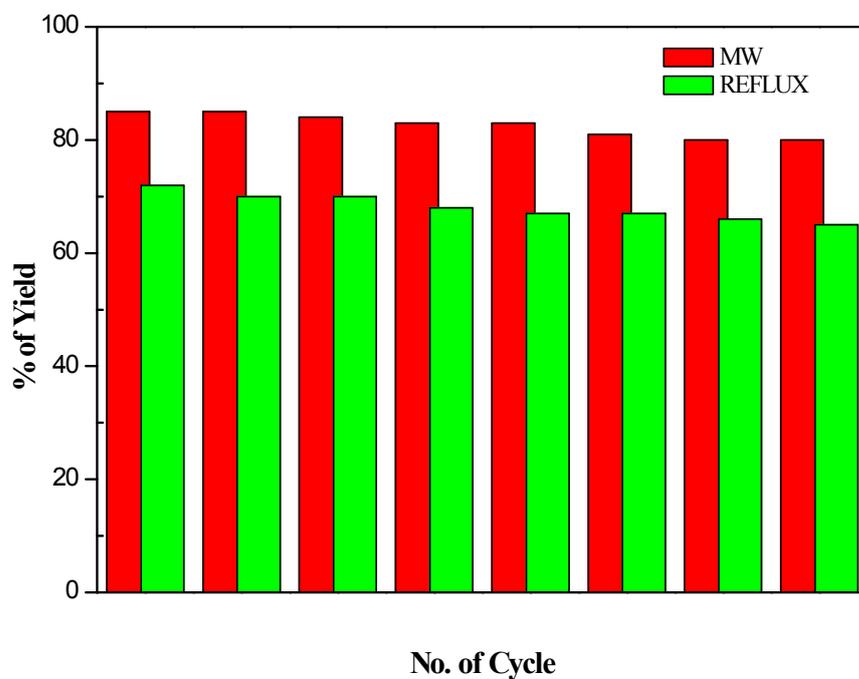


Fig. 4S Recyclability of nano-NiFe₂O₄catalyst

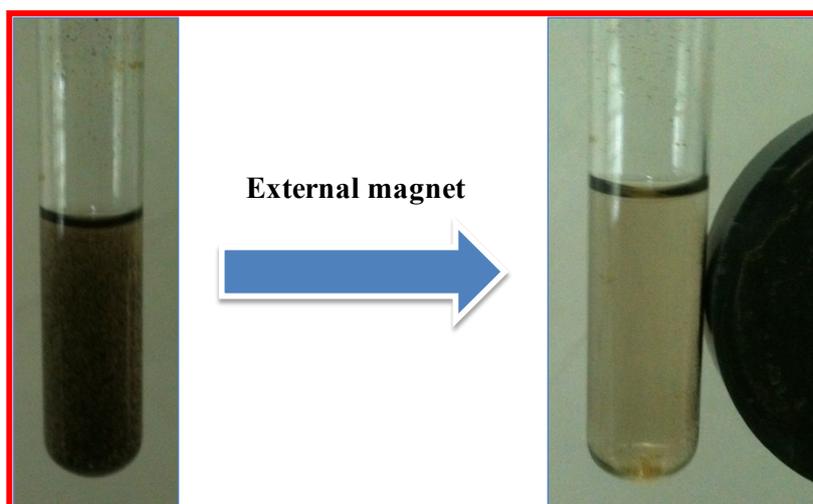


Fig. 5S Separation of magneticof nano-NiFe₂O₄catalyst by using an external magnet.

ESI 10.References:

[1]. For preparation of NiFe₂O₄: P. Sivakumar, R. Ramesh, A. Ramanand, S.Ponnusamy, C. Muthamizhchelvan, *J. Alloys and Compounds*, **2013**, 563, 6–11.

[2] Zi-You Yu, Li-Feng Chen and Shu-Hong Yu, *J. Mater. Chem.A*, **2014**, 2, 10889.

[3] For preparation of CuFe₂O₄: K. Pradhan, S. Paul and A. R. Das, *Catal. Sci. Technol.*, **2014**, 4, 822.

[4] For preparation of CoFe₂O₄: B. Y. Yu, S. Y.Kwak, *Dalton Trans.*, **2011**, 40, 9989