

Supplementary data

Hypersampsones S-W, new polycyclic polyprenylated acylphloroglucinols from *Hypericum sampsonii*

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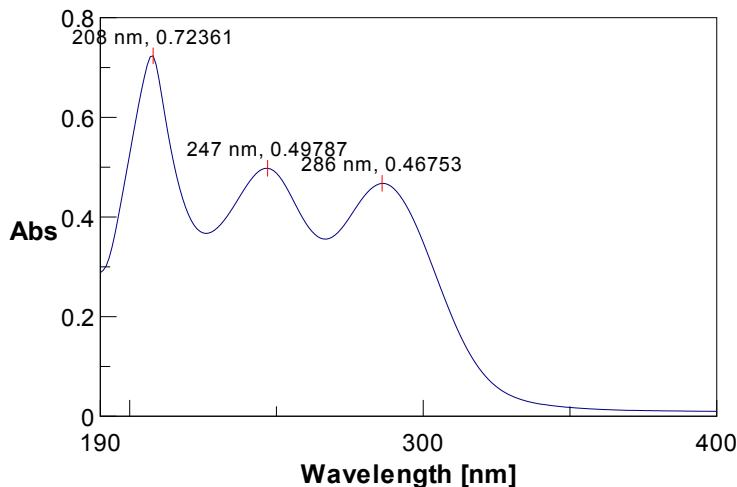
* E-mail address: tyaoxs@jnu.edu.cn; daiyi1004@163.com

List of Supporting Information

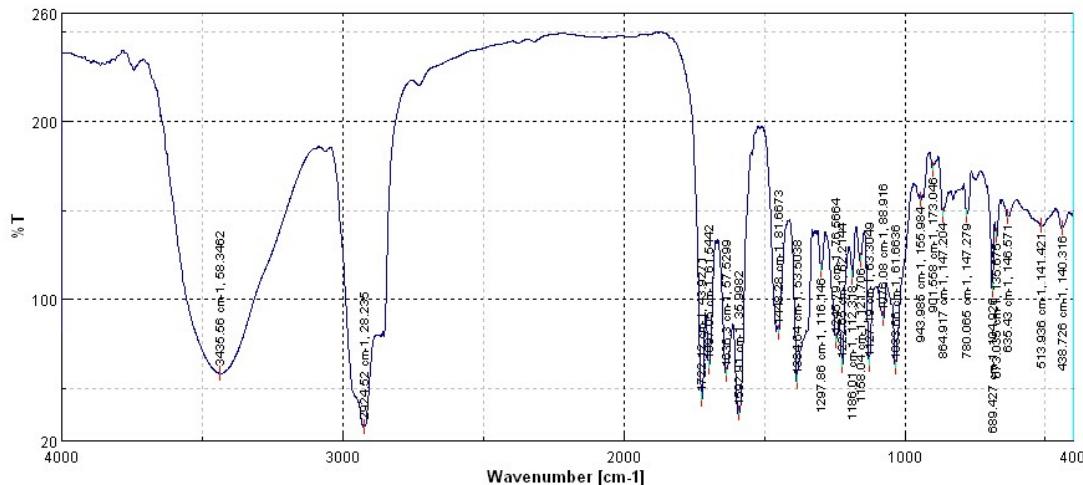
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UV spectrum of hypersampsone S (1) in CH₃OH.



IR (KBr disc) spectrum of hypersampsone S (1).



HR-ESI-MS spectrum of hypersampsone S (1).

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

173 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass)

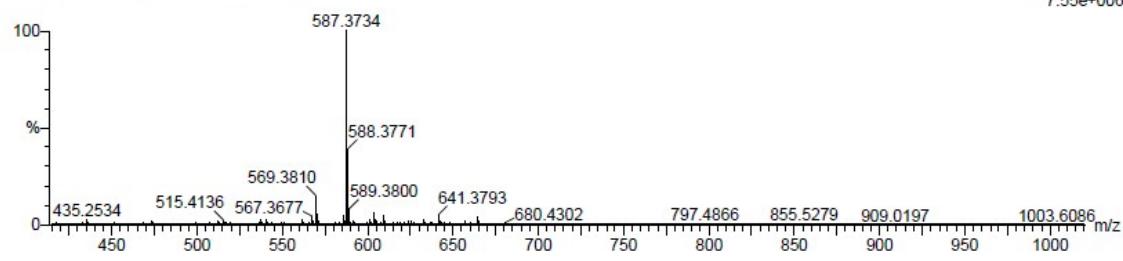
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HSD5F8B-3

20130511-39 266 (2.147) Cm (258:273)

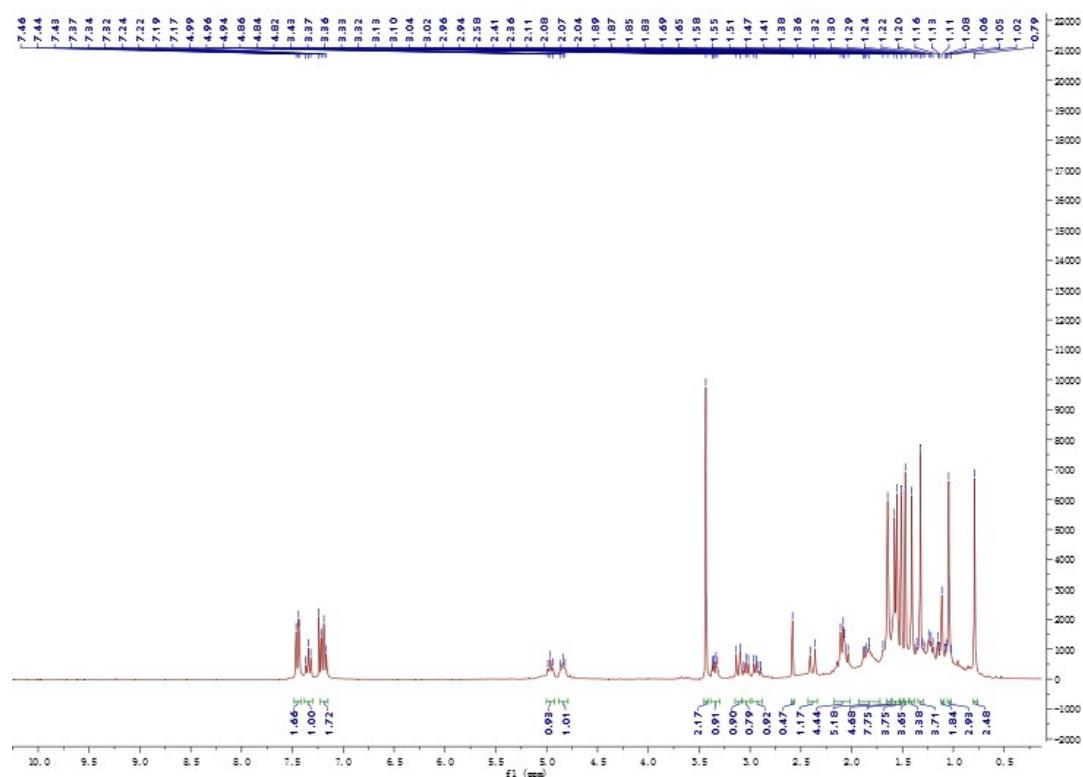
1: TOF MS ES+
7.55e+006



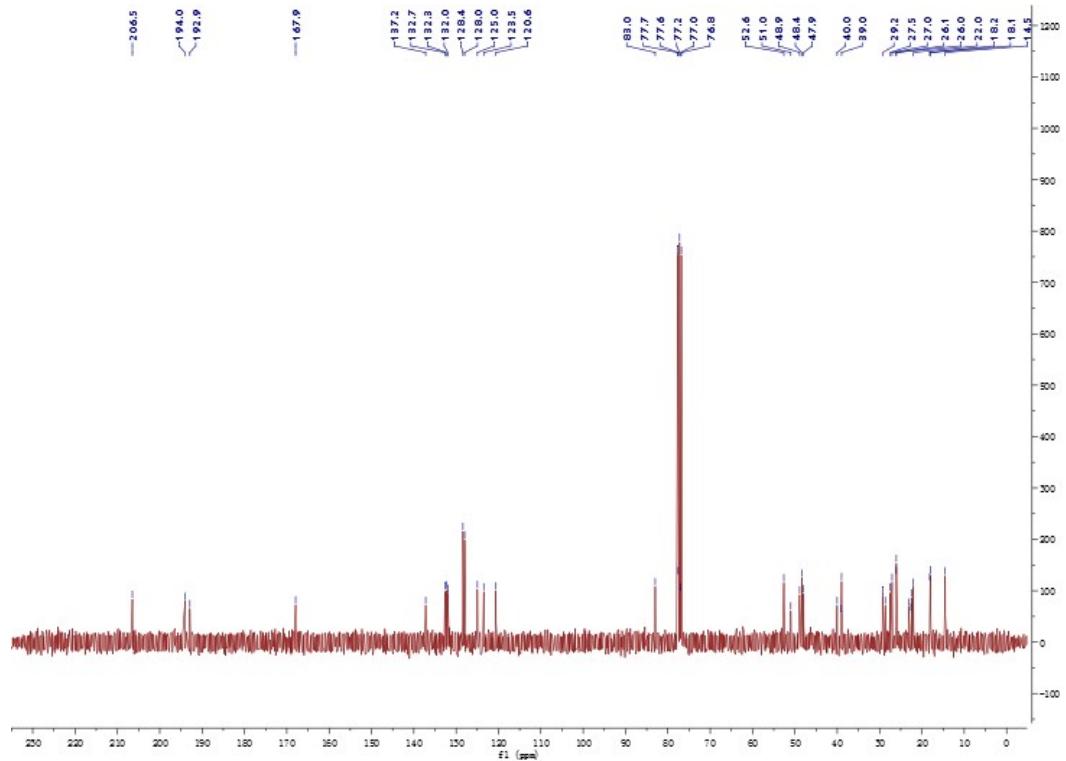
Minimum: -1.5
Maximum: 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf (%)	Formula
587.3734	587.3736	-0.2	-0.3	13.5	1517.1	n/a	n/a	C38 H51 O5

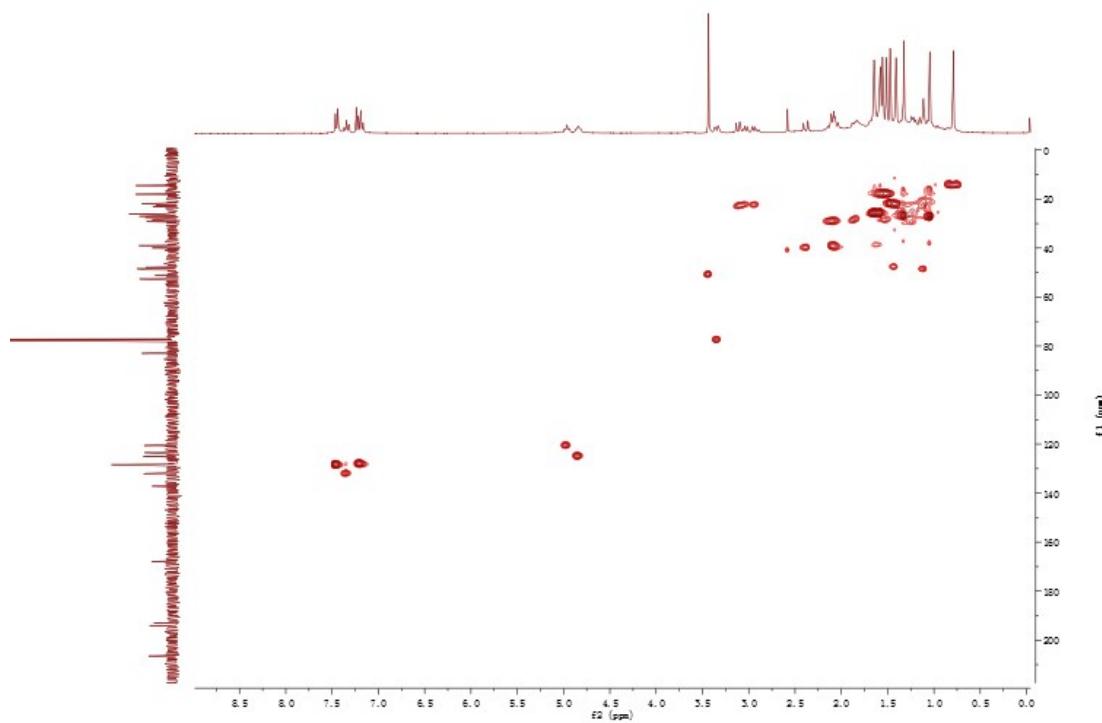
¹H NMR (AV-300, 300 MHz) spectrum of hypersampsone S (1) in CDCl₃



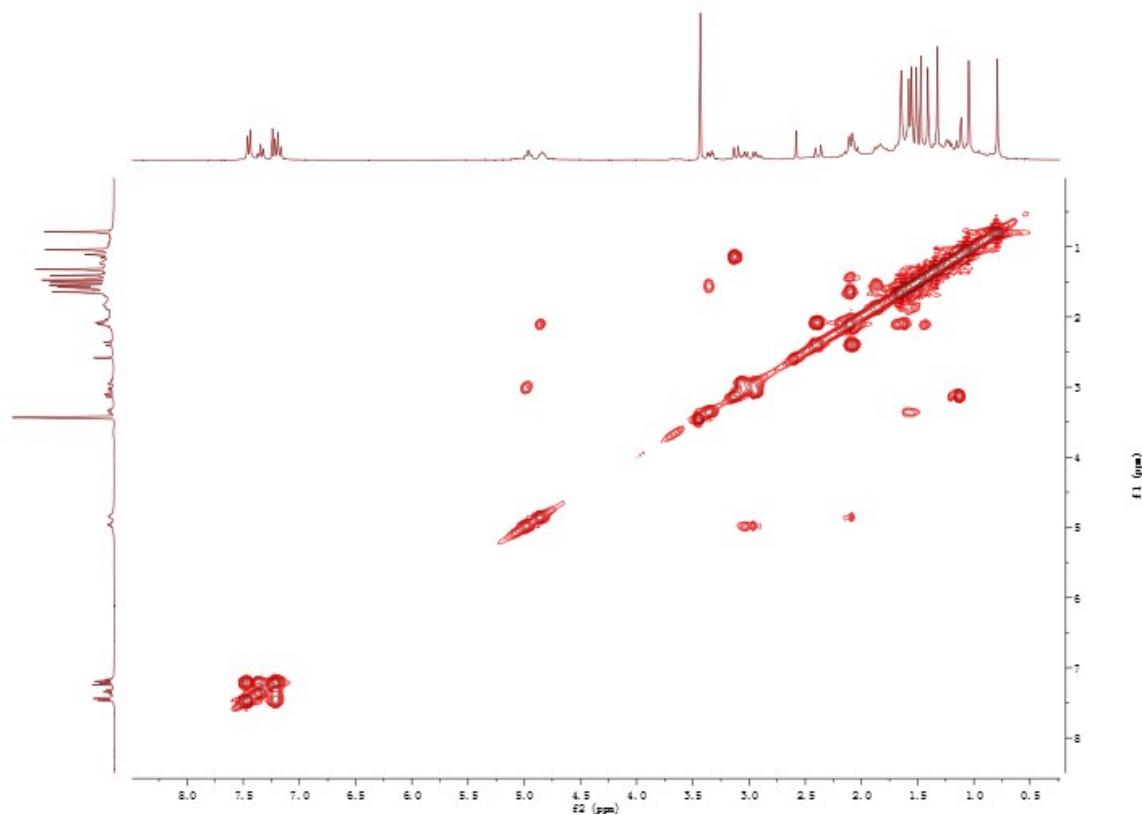
¹³C NMR spectrum (AV-300, 75 MHz) of hypersampsone S (1) in CDCl₃



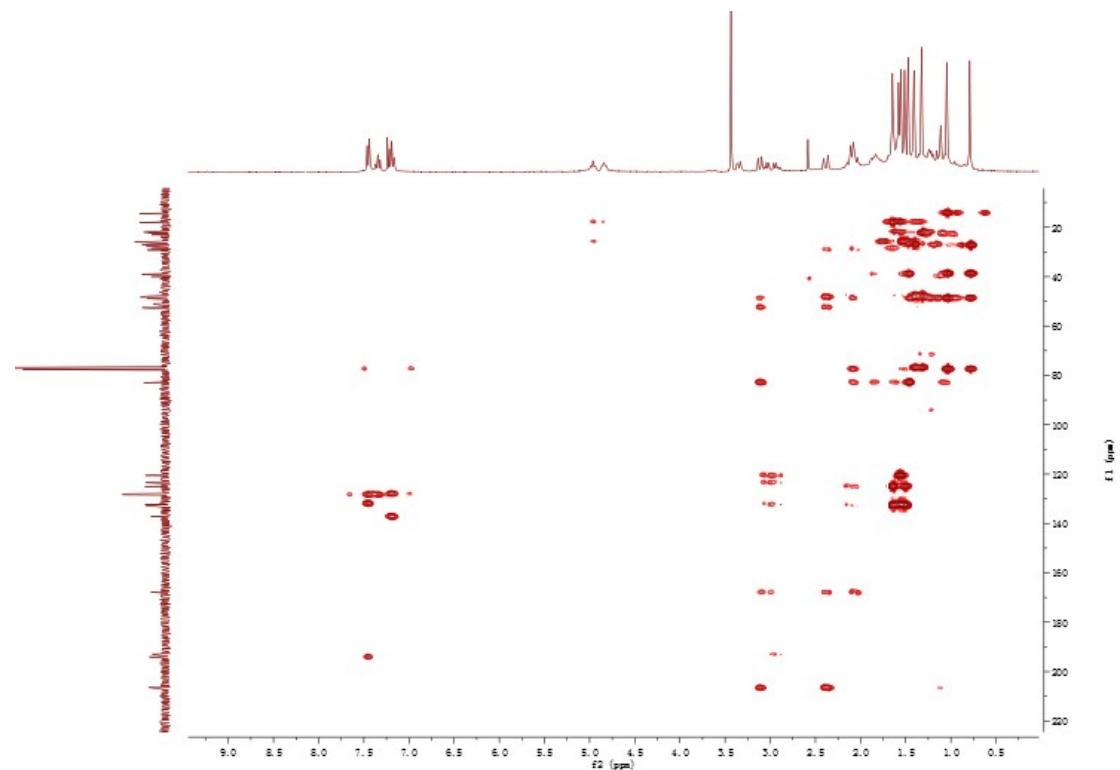
HSQC spectrum (AV-400) of hypersampsone S (1) in CDCl_3



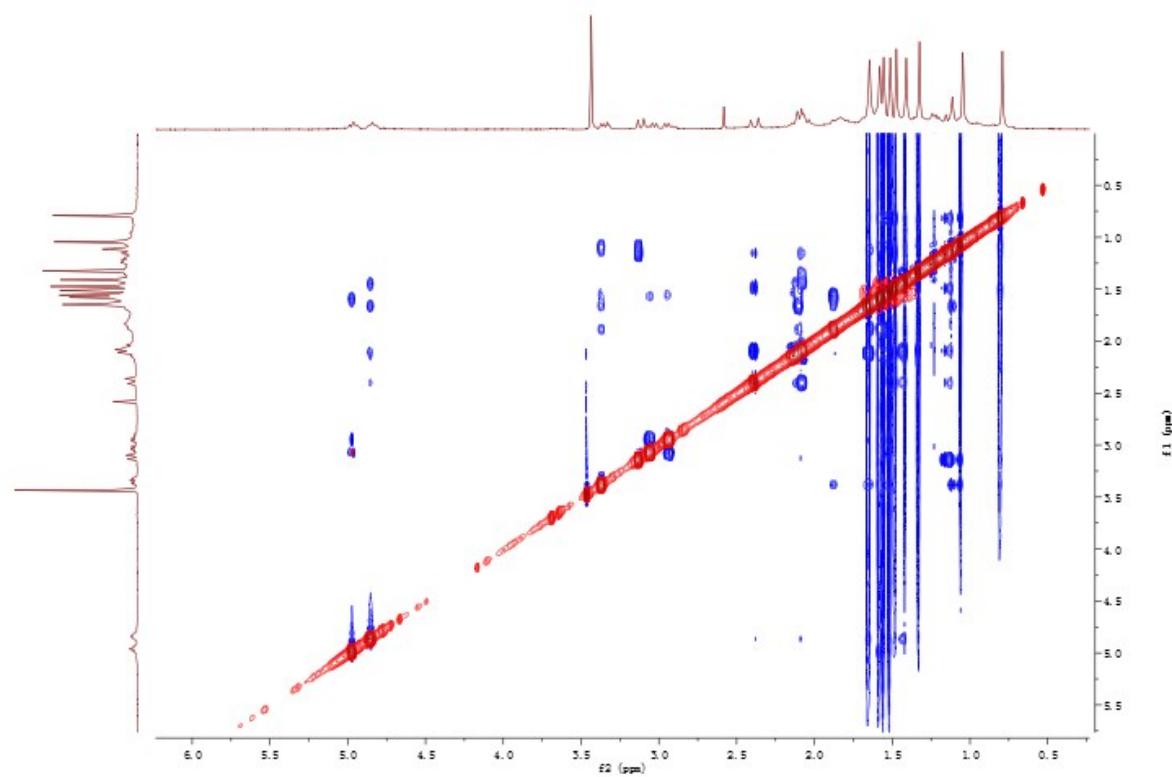
^1H - ^1H COSY spectrum (AV-400) of hypersampsone S (1) in CDCl_3



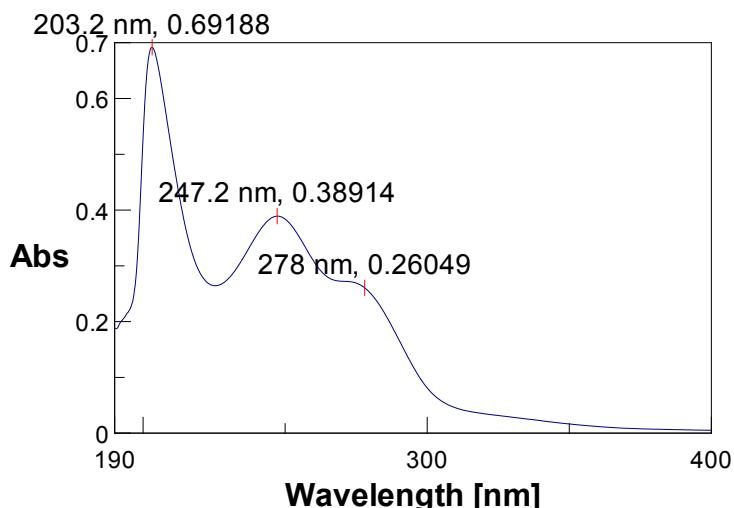
HMBC spectrum (AV-400) of hypersampsone S (1) in CDCl_3



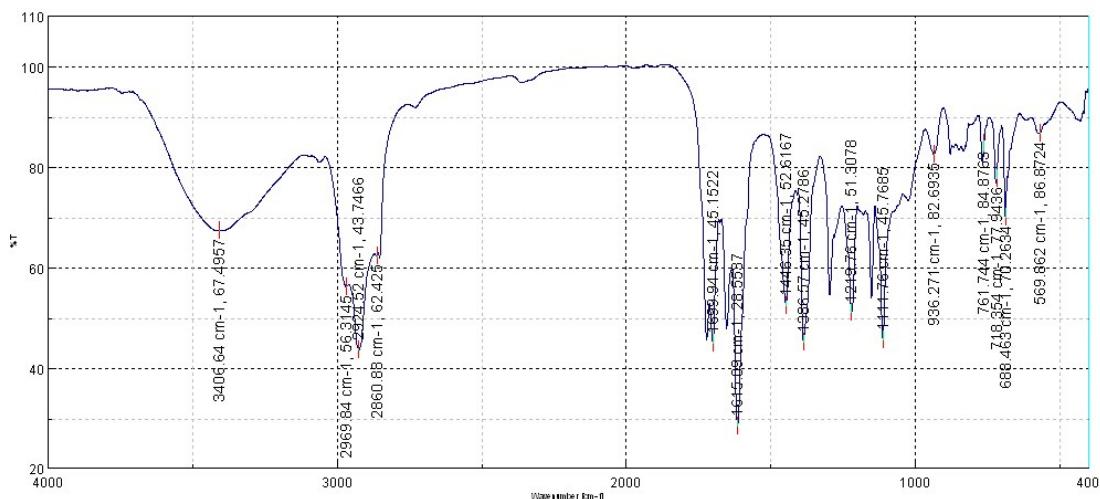
NOESY spectrum (AV-600) of hypersampsone S (1) in CDCl_3



UV spectrum of hypersampsone T (2) in CH₃OH.



IR (KBr disc) spectrum of hypersampsone T (2).



HR-ESI-MS spectrum of hypersampsone T (2).

Single Mass Analysis

Tolerance = 10.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

133 formula(e) evaluated with 2 results within limits (up to 50 closest results for each mass)

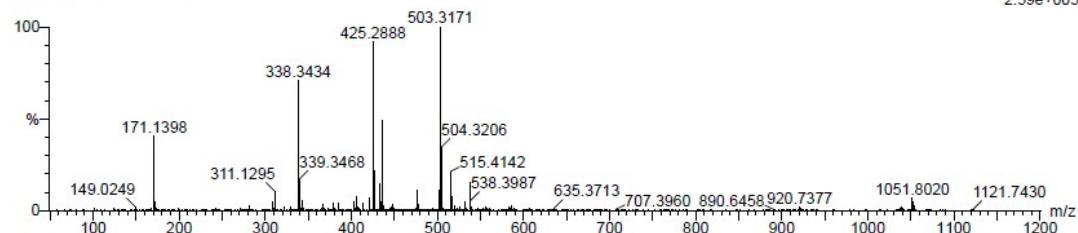
Elements Used:

C: 0-500 H: 0-1000 O: 0-200

HSD1E-4

20131016-02 275 (2.214) Cm (274:277)

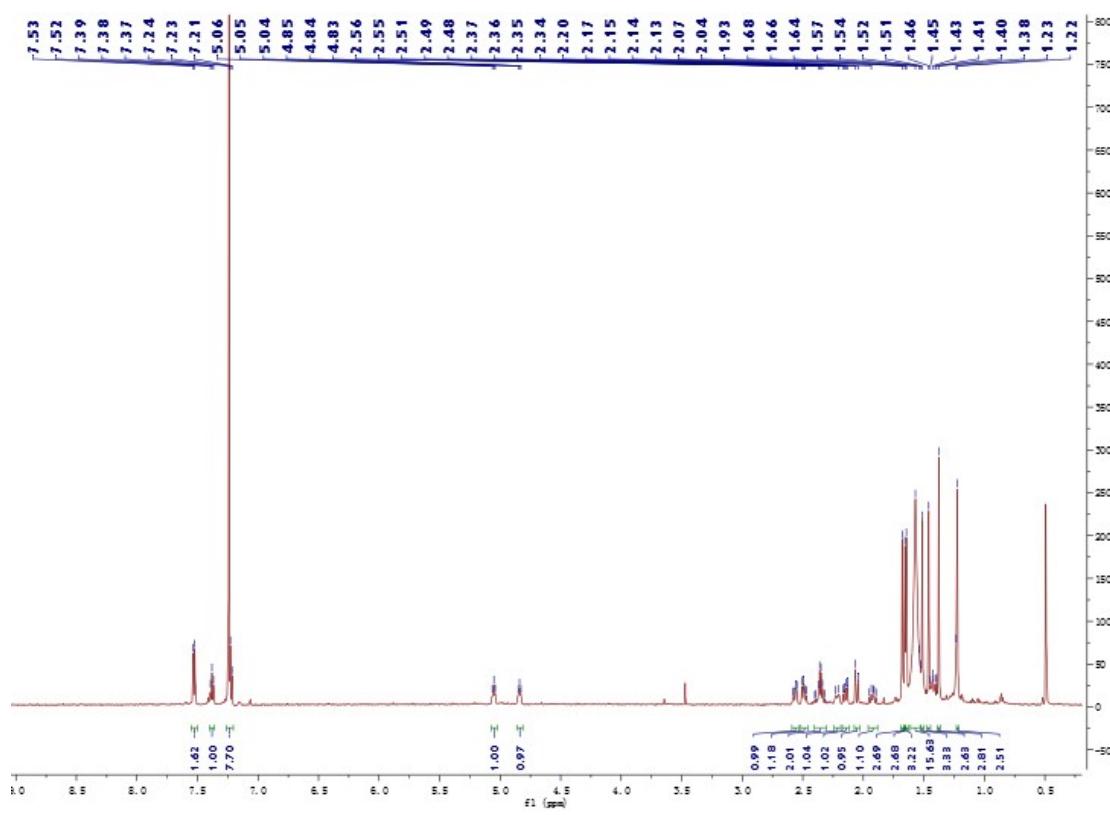
1: TOF MS ES+
2.59e+005



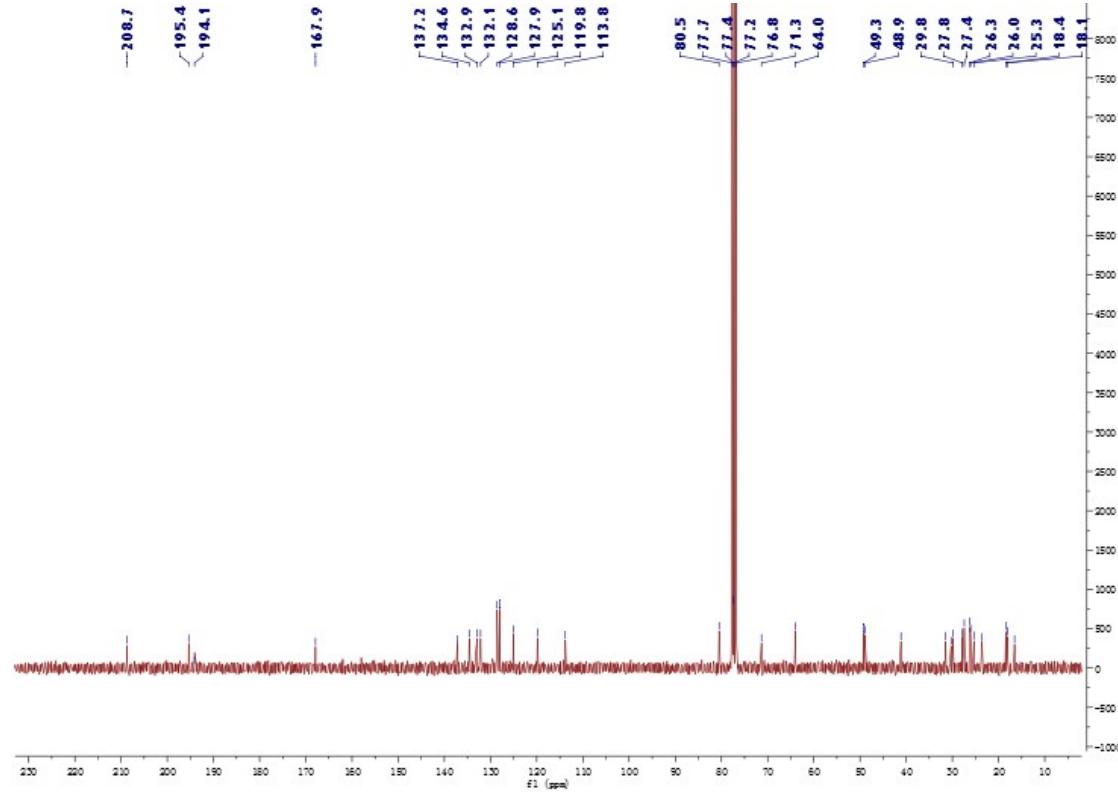
Minimum: -1.5
Maximum: 5.0 10.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf (%)	Formula
503.3171	503.3161	1.0	2.0	12.5	447.6	0.001	99.88	C ₃₃ H ₄₃ O ₄
	503.3220	-4.9	-9.7	3.5	454.3	6.691	0.12	C ₂₆ H ₄₇ O ₉

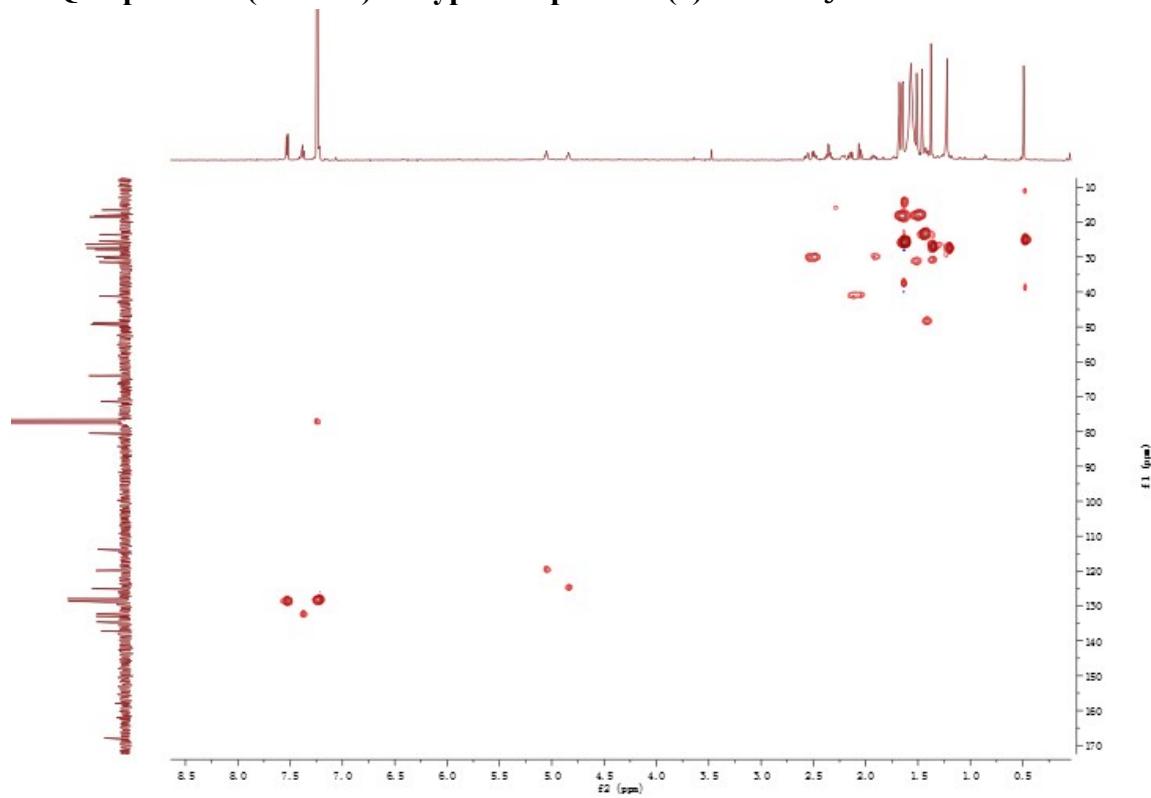
¹H NMR (AV-600, 600 MHz) spectrum of hypersampsone T (2) in CDCl₃



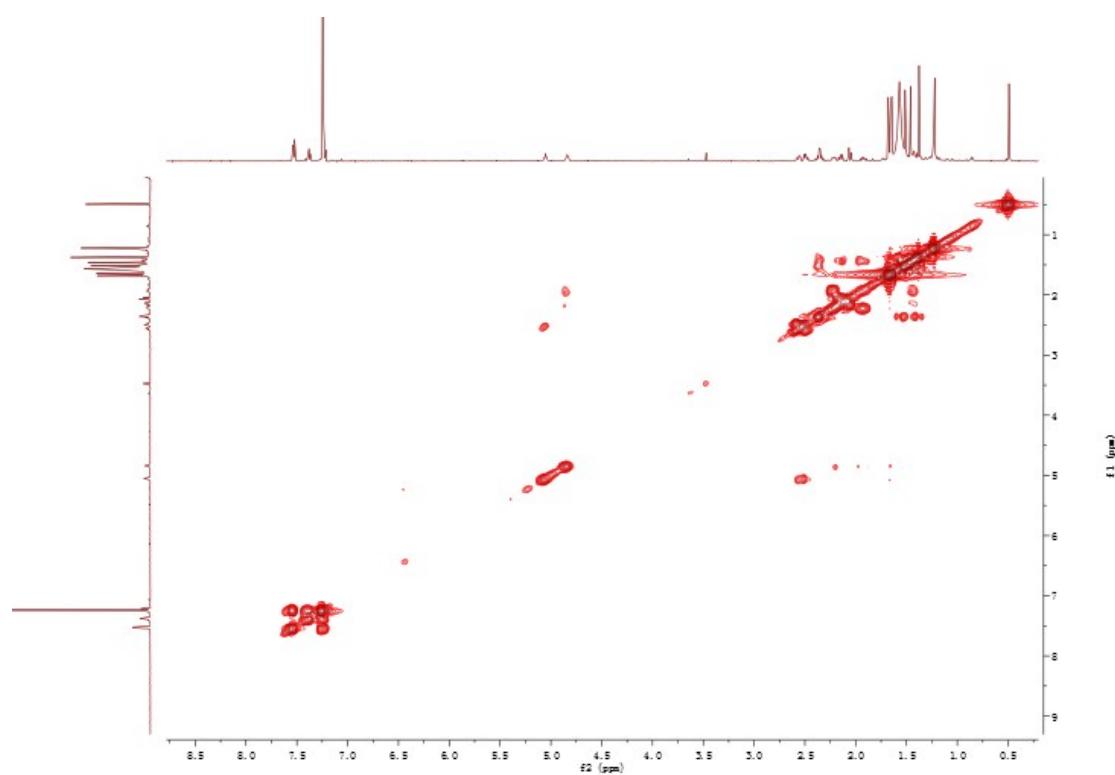
¹³C NMR spectrum (AV-300, 75 MHz) of hypersampsone T (2) in CDCl₃



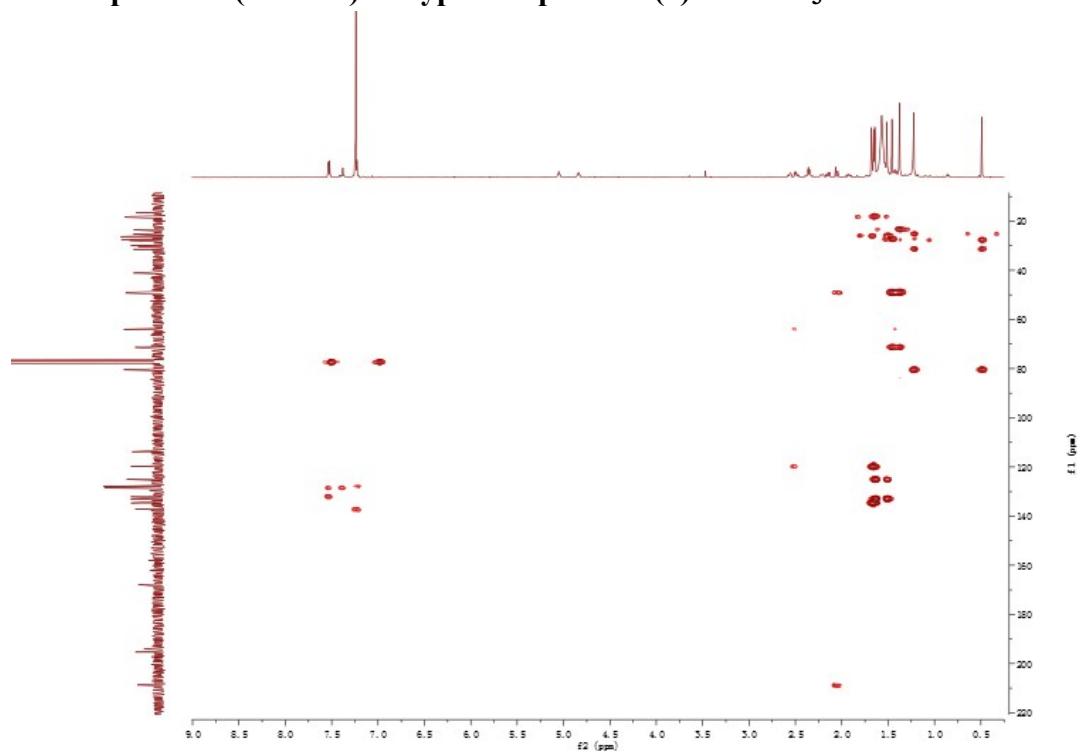
HSQC spectrum (AV-400) of hypersampsone T (2) in CDCl_3



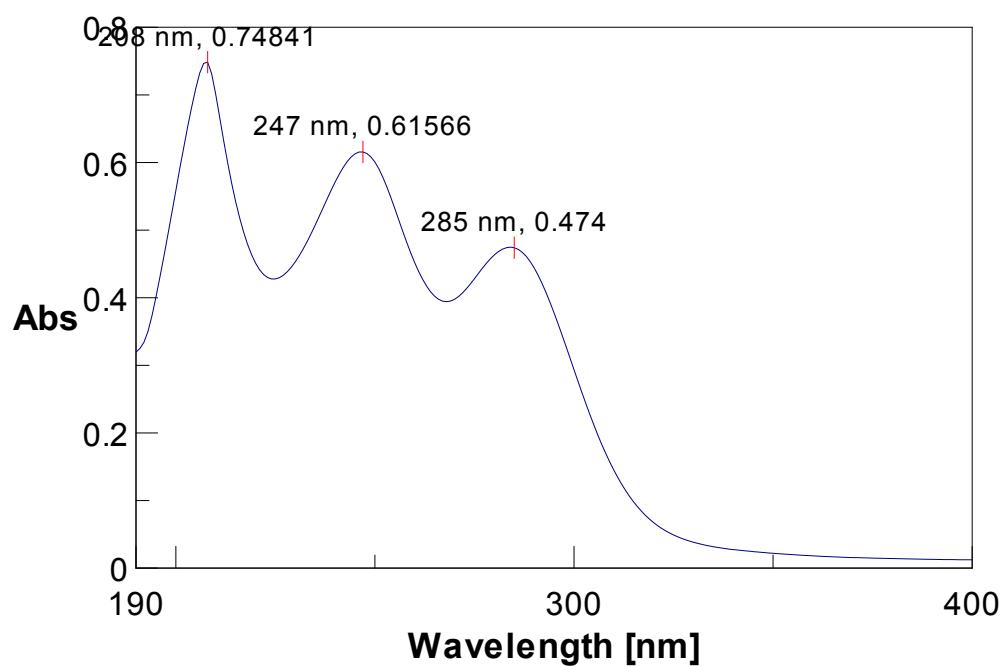
^1H - ^1H COSY spectrum (AV-400) of hypersampsone T (2) in CDCl_3



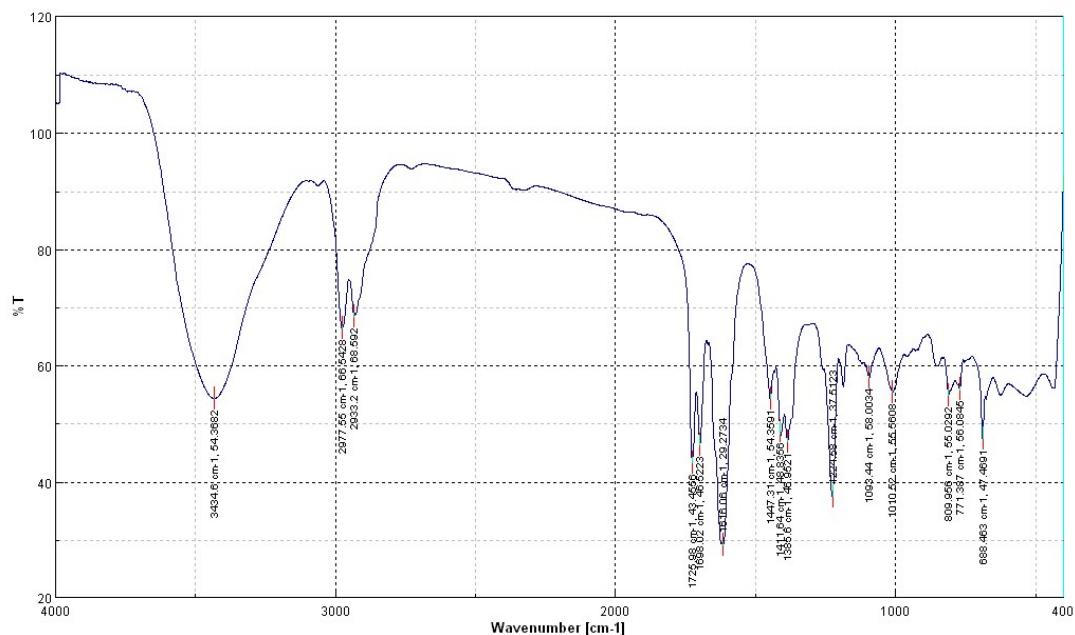
HMBC spectrum (AV-400) of hypersampsone T (2) in CDCl_3



UV spectrum of hypersampsone U (3) in CH_3OH .



IR (KBr disc) spectrum of hypersampsone U (3).



HR-ESI-MS spectrum of hypersampsone U (3).

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

140 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass)

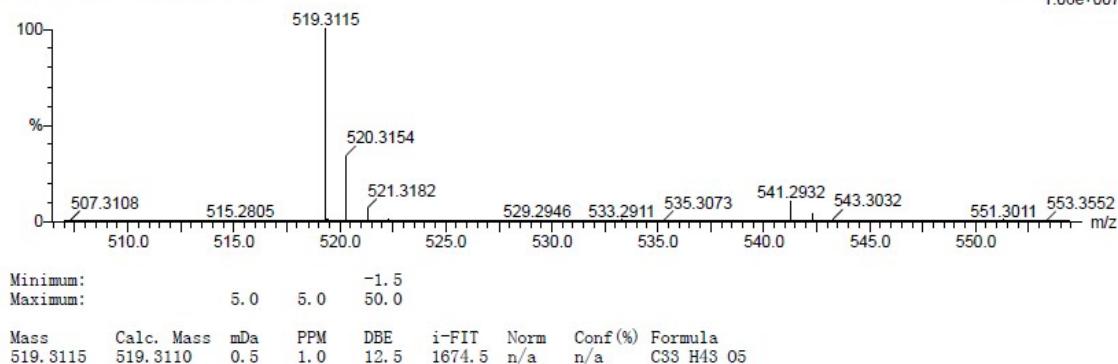
Elements Used:

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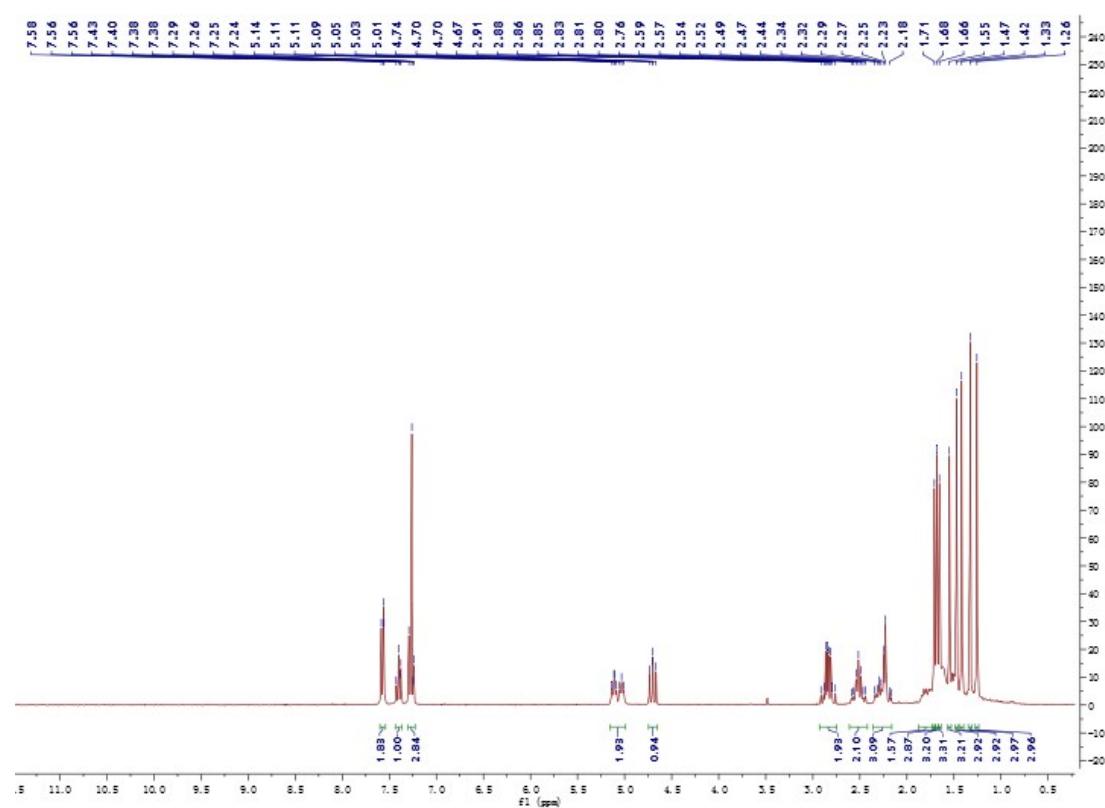
HSD5F6B3-7

20130422-6 245 (1.978) Cm (228:253)

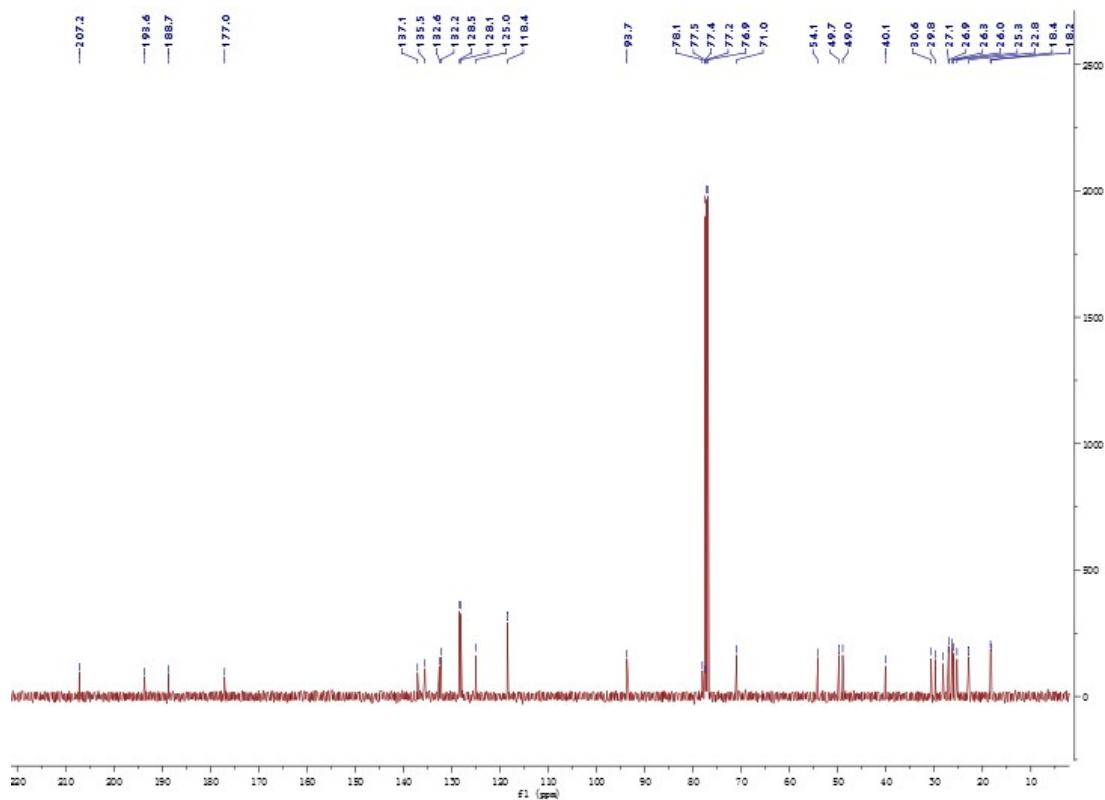
1: TOF MS ES+
1.06e+007



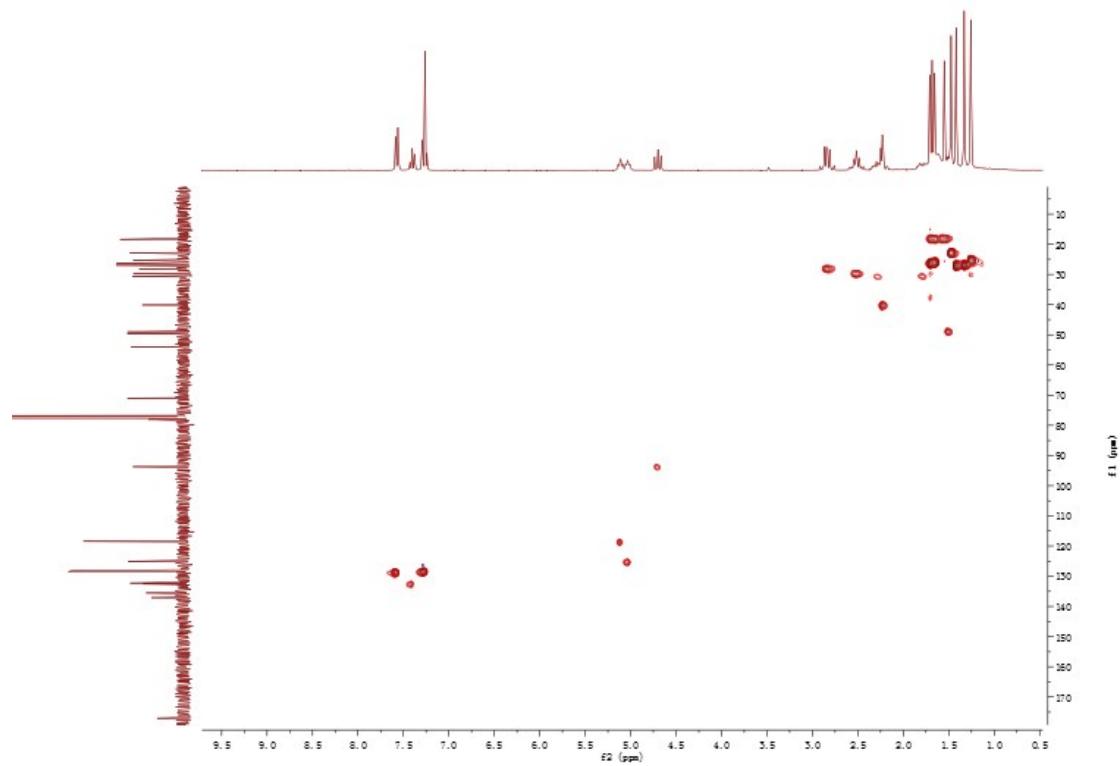
¹H NMR (AV-300, 300 MHz) spectrum of hypersampsone U (3) in CDCl₃



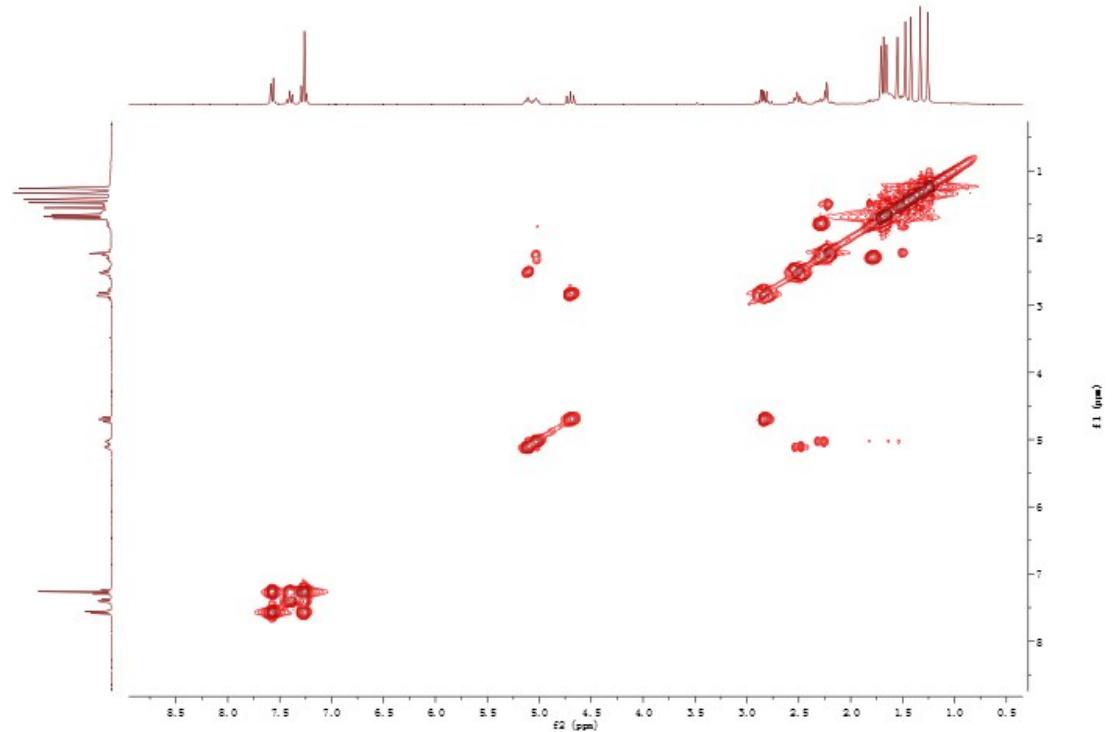
¹³C NMR spectrum (AV-400, 100 MHz) of hypersampsone U (3) in CDCl₃



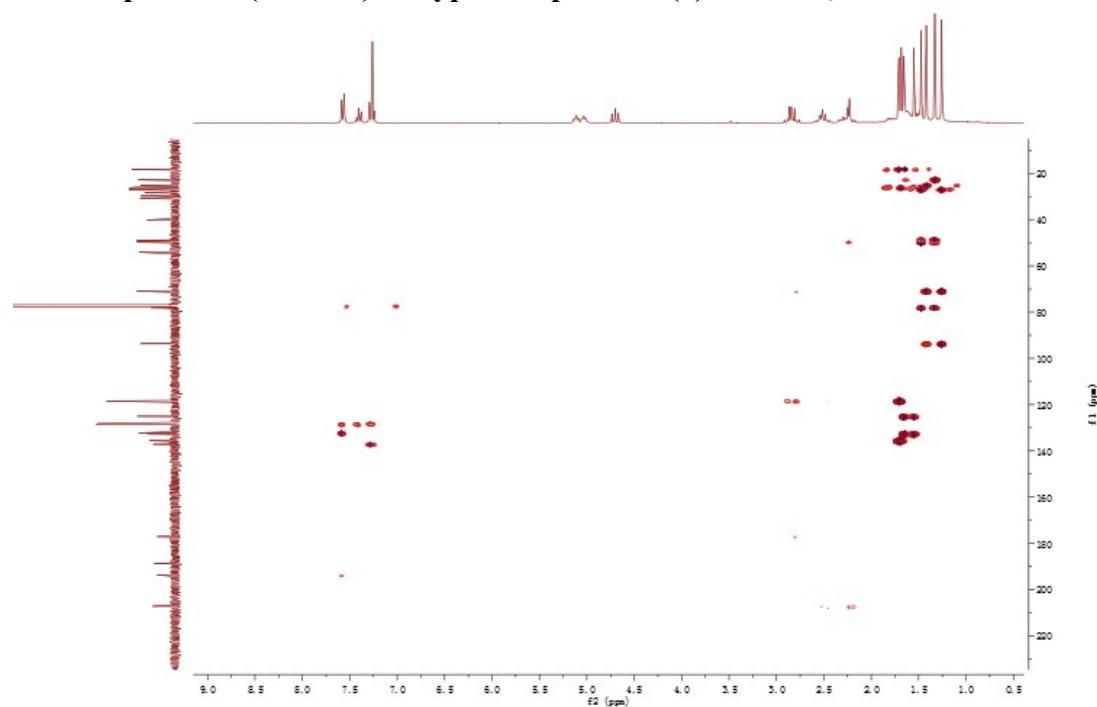
HSQC spectrum (AV-400) of hypersampsone U (3) in CDCl_3



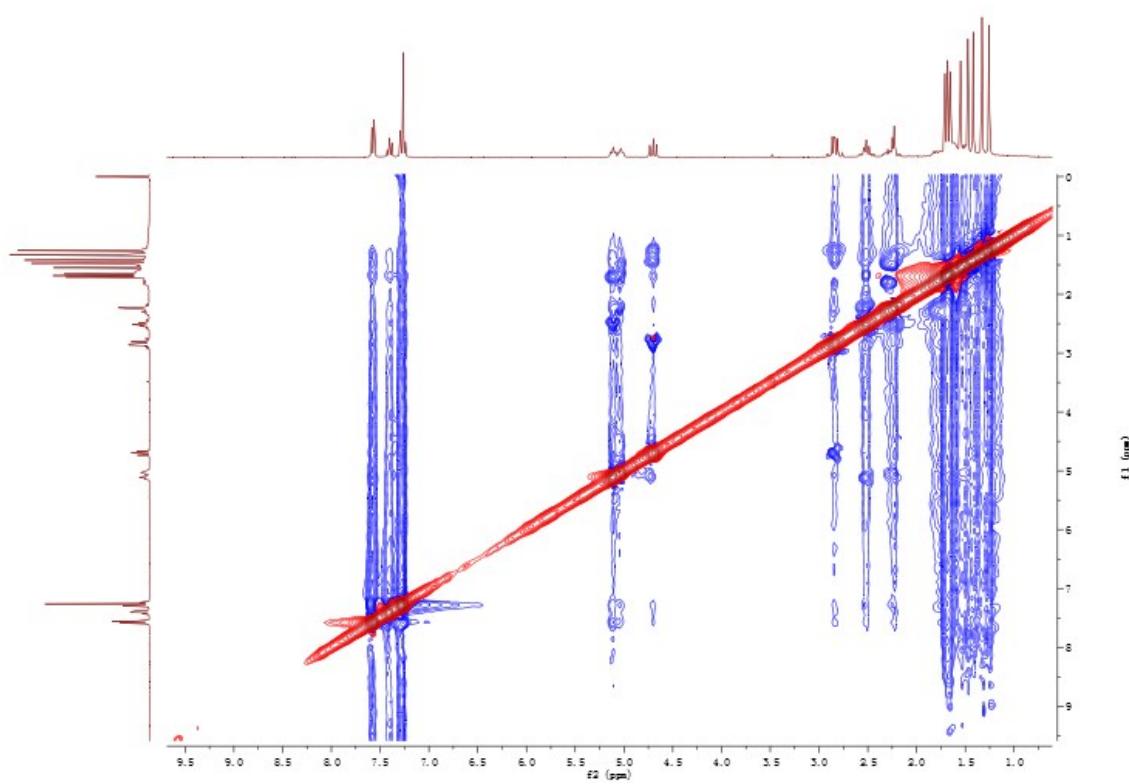
^1H - ^1H COSY spectrum (AV-400) of hypersampsone U (3) in CDCl_3



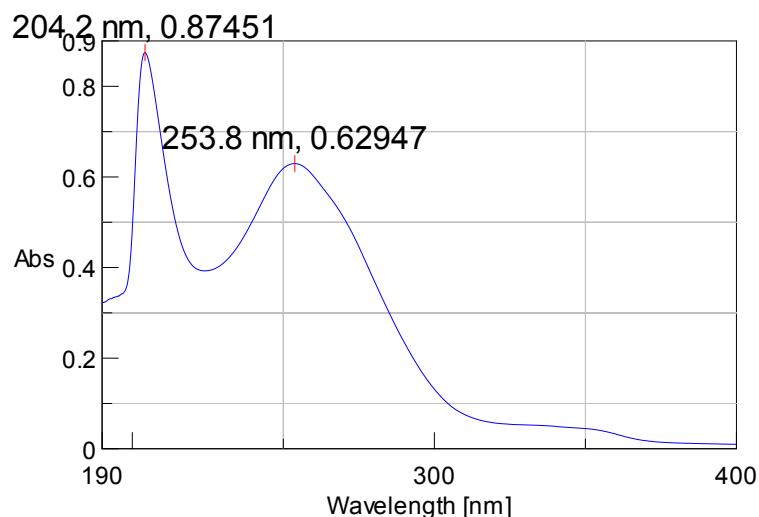
HMBC spectrum (AV-400) of hypersampsone U (3) in CDCl_3



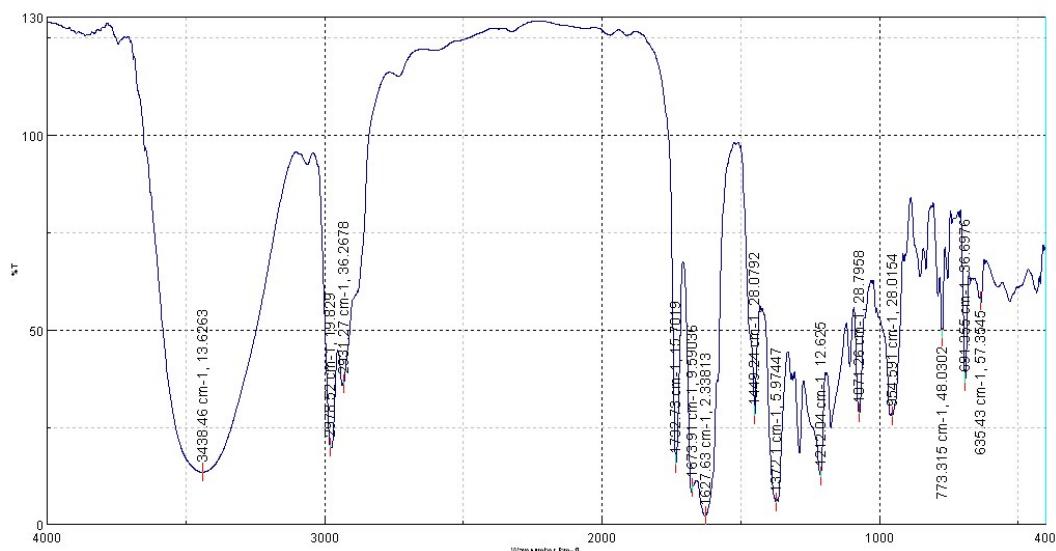
NOESY spectrum (AV-400) of hypersampsone U (3) in CDCl_3



UV spectrum of hypersampsone V (4) in CH₃OH.



IR (KBr disc) spectrum of hypersampsone Q (4).



HR-ESI-MS spectrum of hypersampsone Q (4).

Single Mass Analysis

Tolerance = 5.0 mDa / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

150 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass)

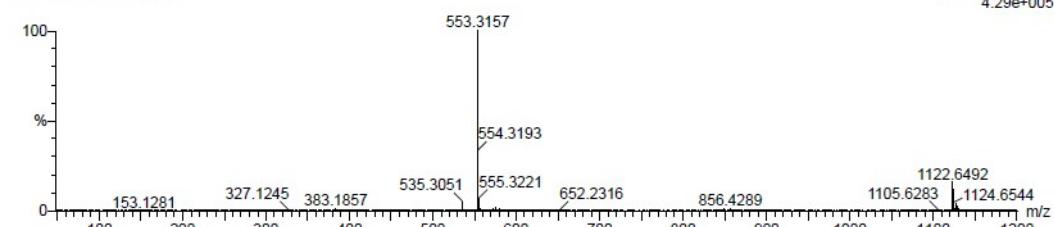
Elements Used:

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HSD7K2-1-4

20141201-28 685 (5.493)

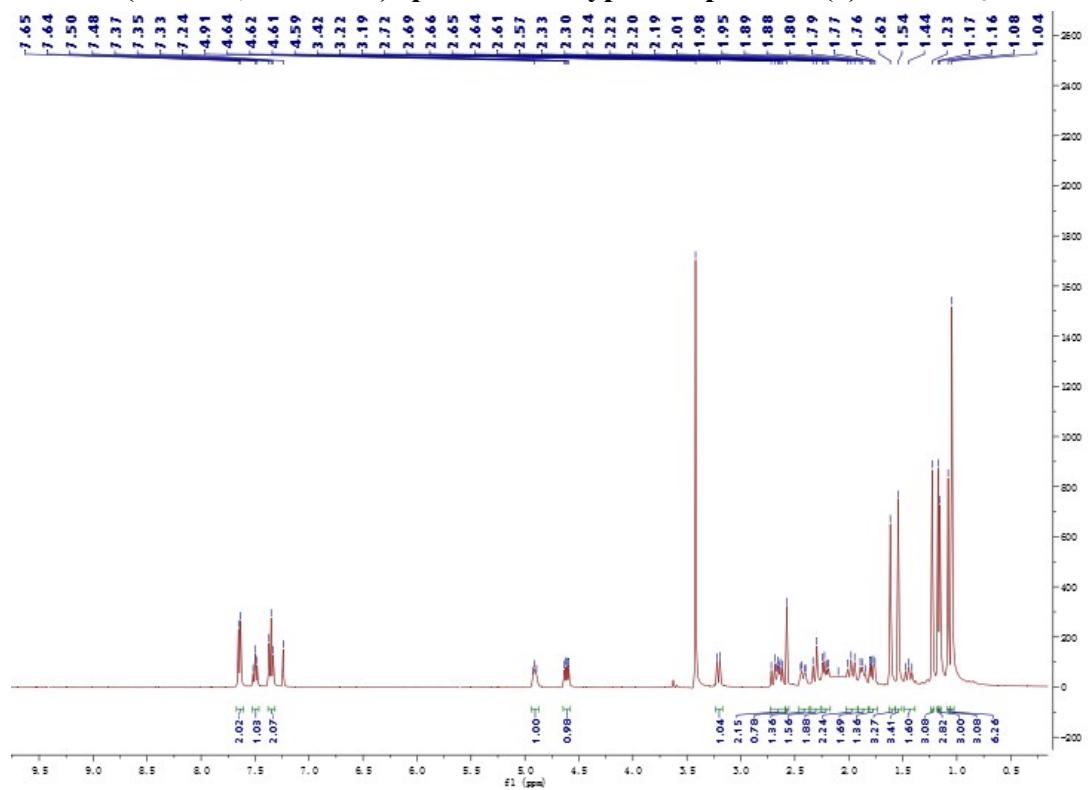
1: TOF MS ES+
4.29e+005



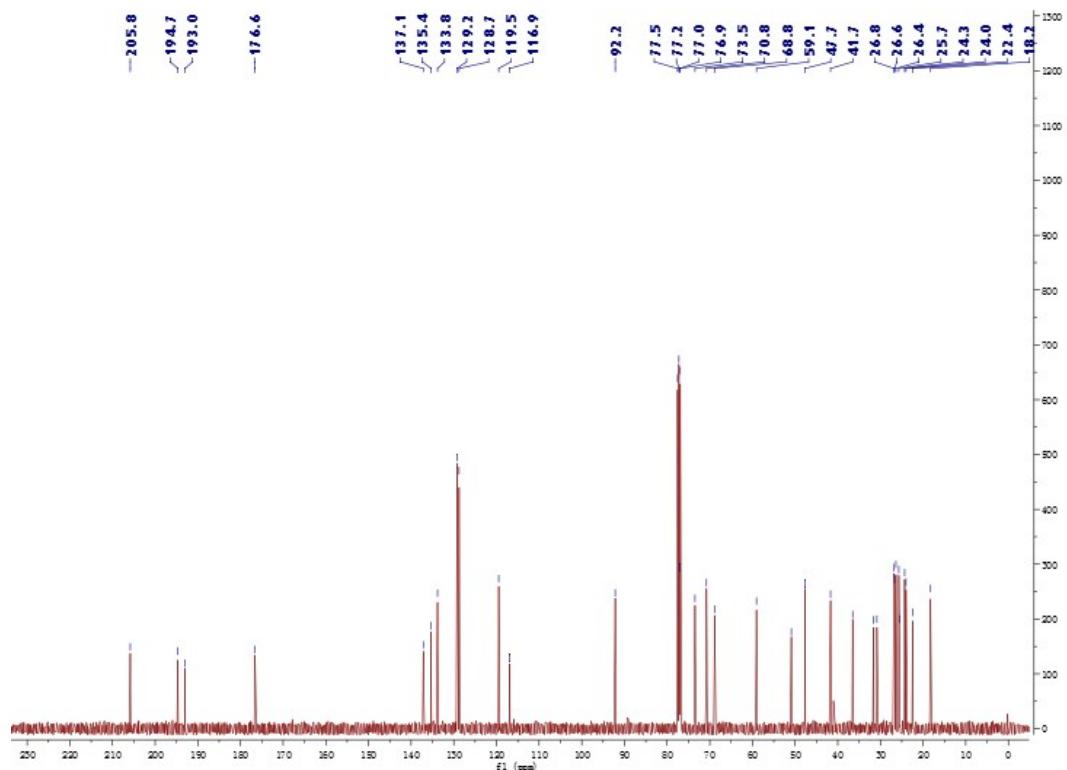
Minimum: -1.5
Maximum: 5.0 5.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf (%)	Formula
553.3157	553.3165	-0.8	-1.4	11.5	276.9	n/a	n/a	C ₃₃ H ₄₅ O ₇

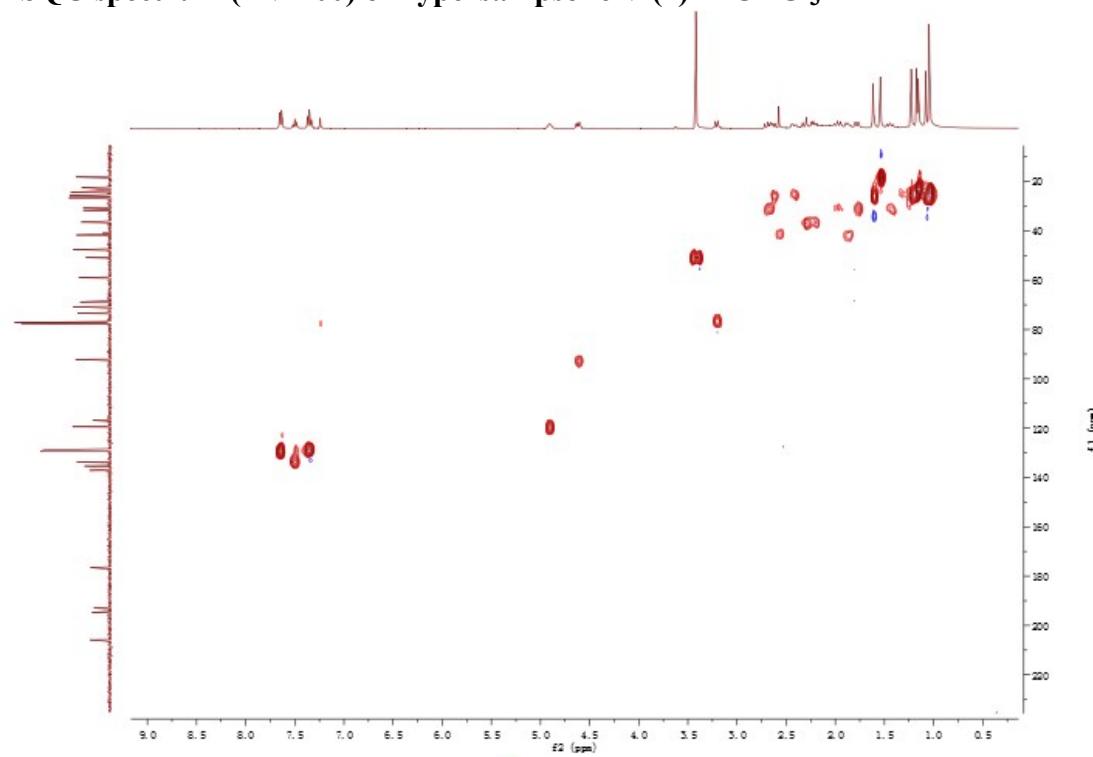
¹H NMR (AV-400, 400 MHz) spectrum of hypersampsone V (4) in CDCl₃



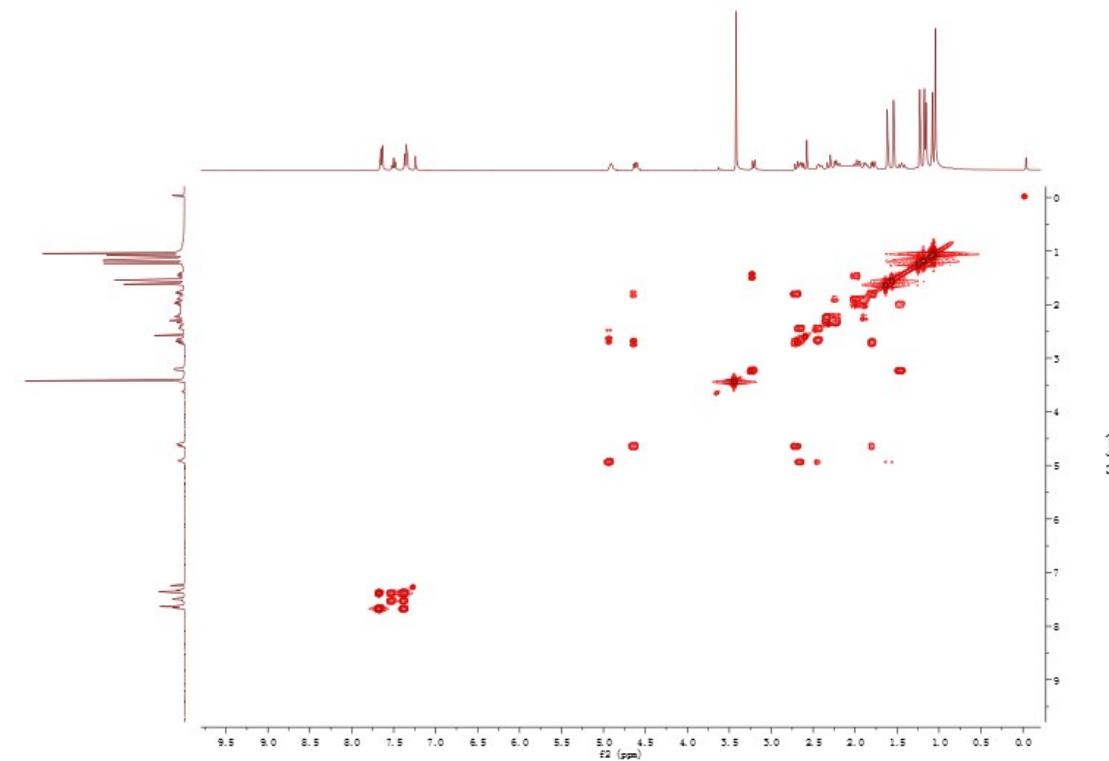
¹³C NMR spectrum (AV-400, 100 MHz) of hypersampsone V (4) in CDCl₃



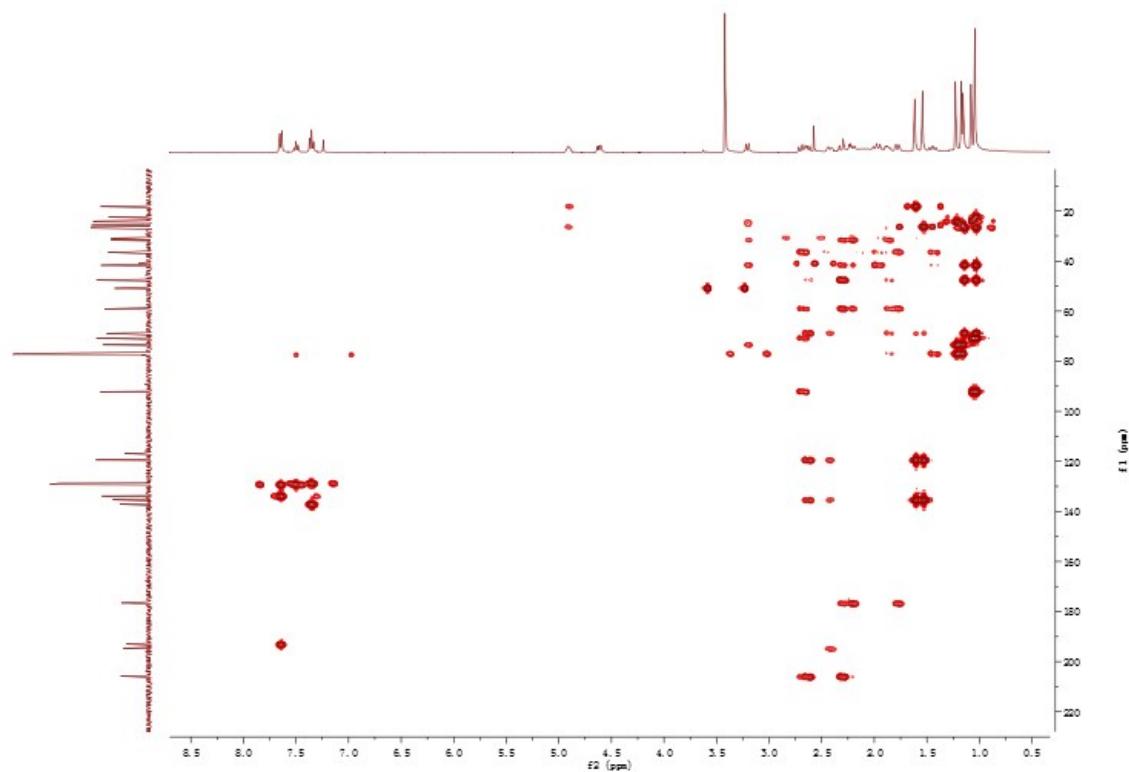
HSQC spectrum (AV-400) of hypersampsone V (4) in CDCl_3



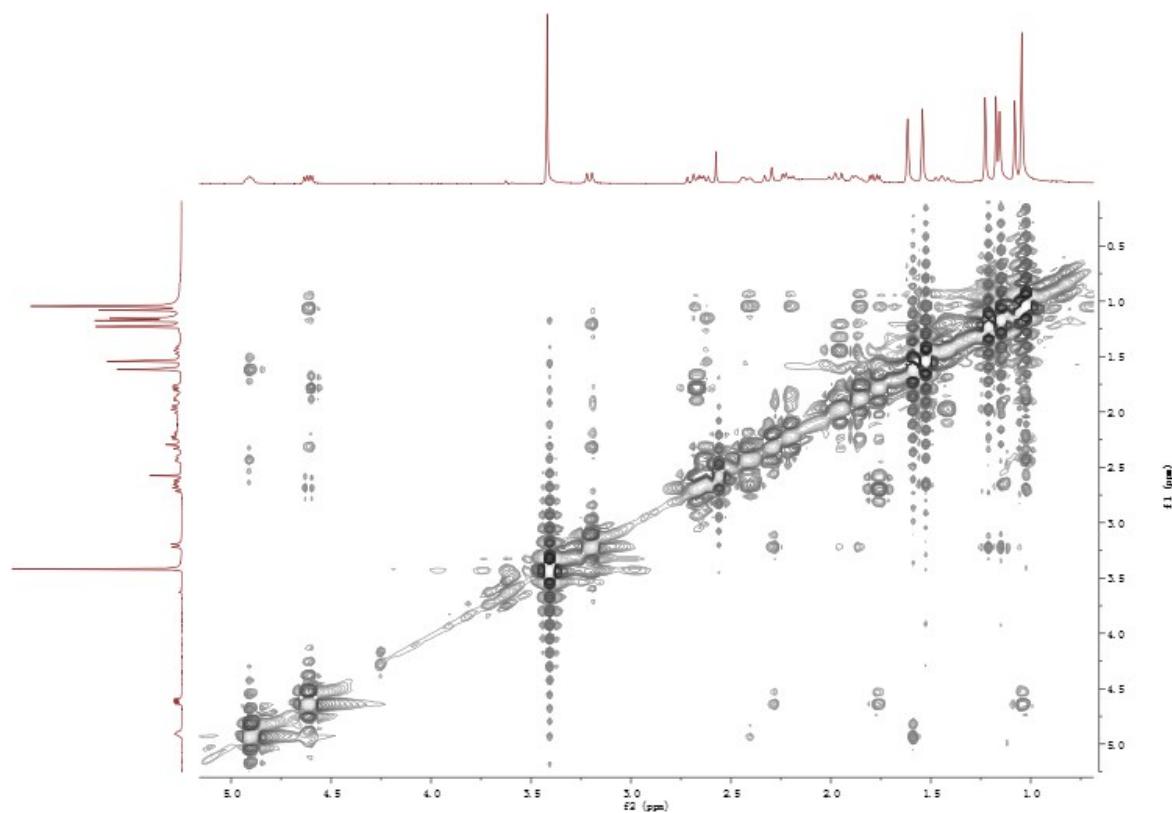
^1H - ^1H COSY spectrum (AV-400) of hypersampsone V (4) in CDCl_3



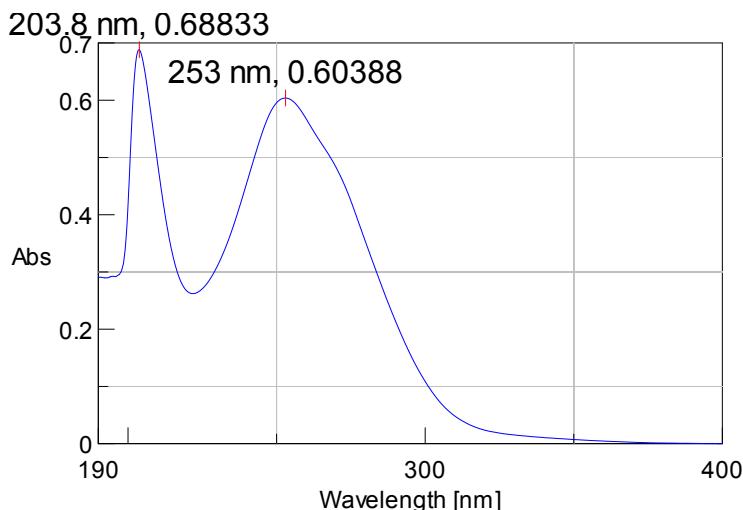
HMBC spectrum (AV-400) of hypersampsone V (4) in CDCl_3



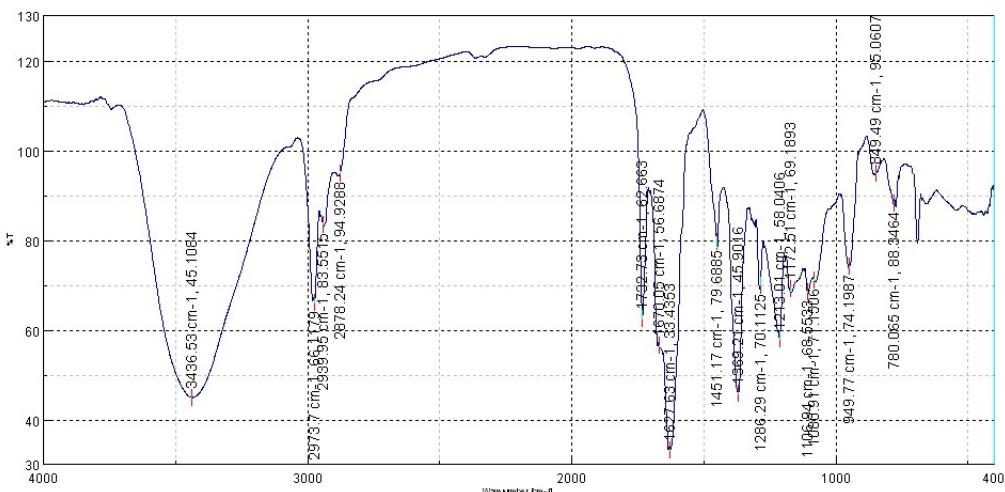
NOESY spectrum (AV-400) of hypersampsone V (4) in CDCl_3



UV spectrum of hypersampsone W (5) in CH₃OH.



IR (KBr disc) spectrum of hypersampsone W (5).



HR-ESI-MS spectrum of hypersampsone W (5).

Single Mass Analysis

Tolerance = 5.0 mDa / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

150 formula(e) evaluated with 1 results within limits (up to 50 best isotopic matches for each mass)

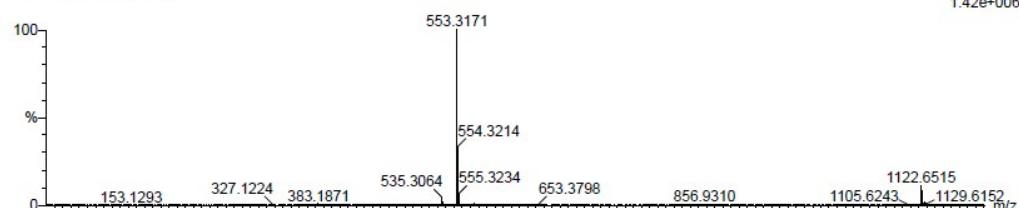
Elements Used:

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HSD7K2-2

20141201-27 710 (5.691)

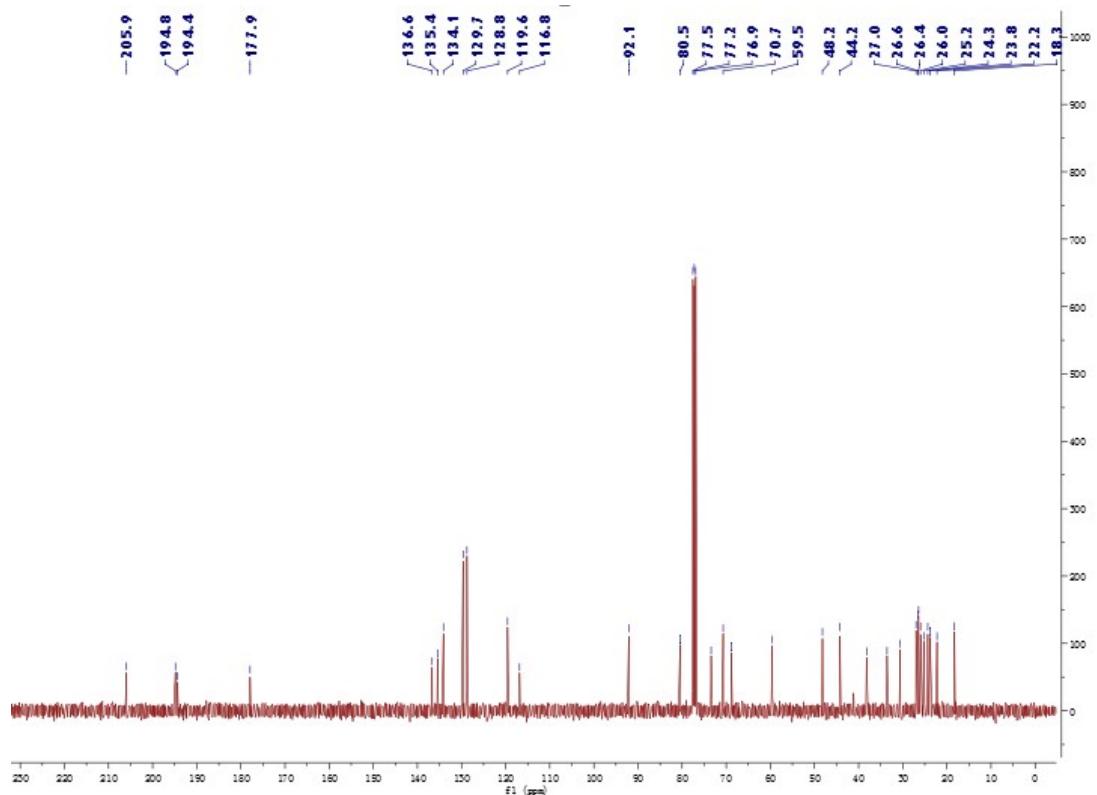
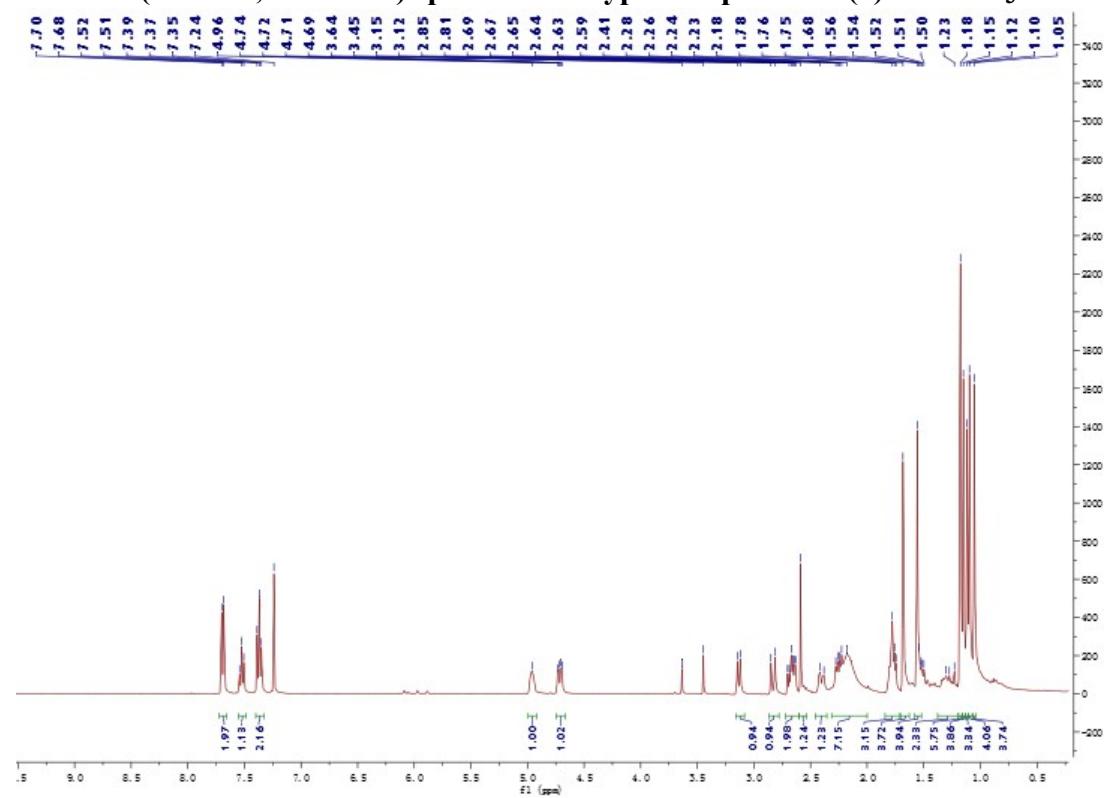
1: TOF MS ES+
1.42e+006



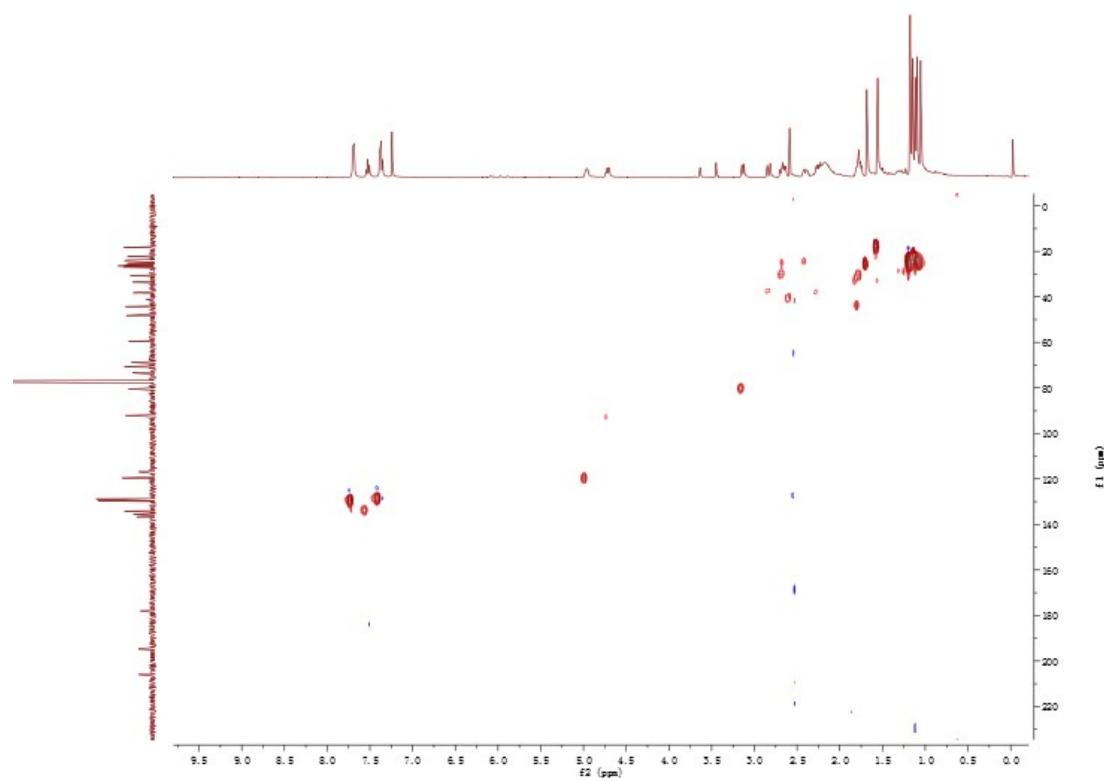
Minimum: 153.1293 Maximum: 5.0 5.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Norm	Conf(%)	Formula
553.3171	553.3165	0.6	1.1	11.5	462.1	n/a	n/a	C ₃₃ H ₄₅ O ₇

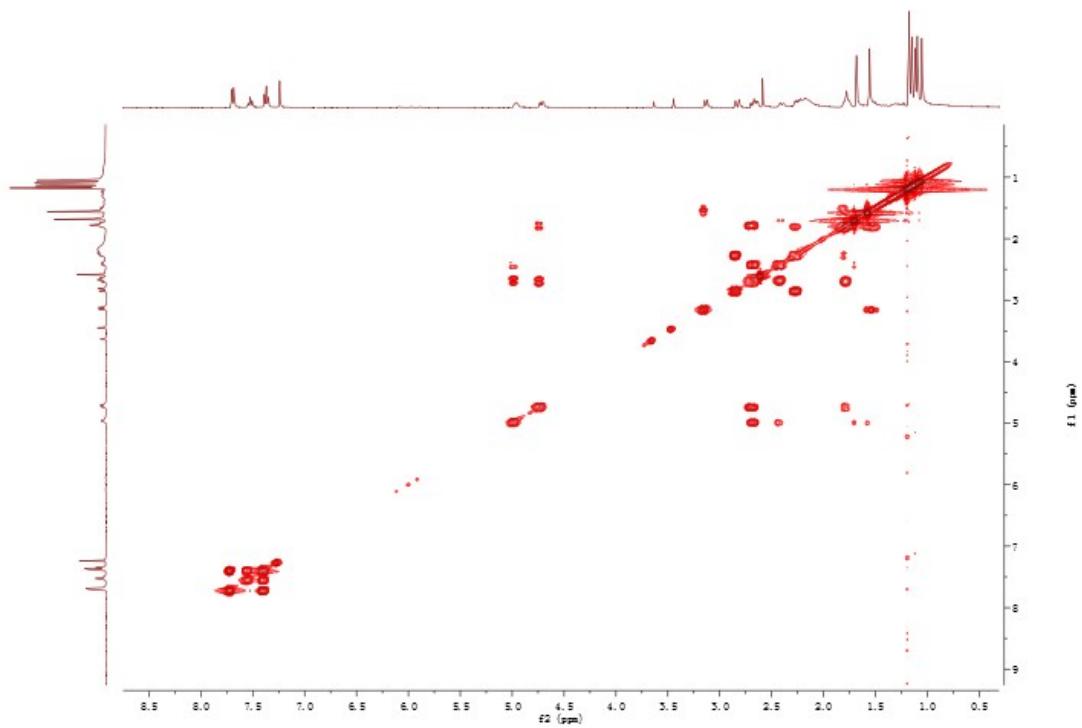
¹H NMR (AV-400, 400 MHz) spectrum of hypersampsone W (4) in CDCl₃



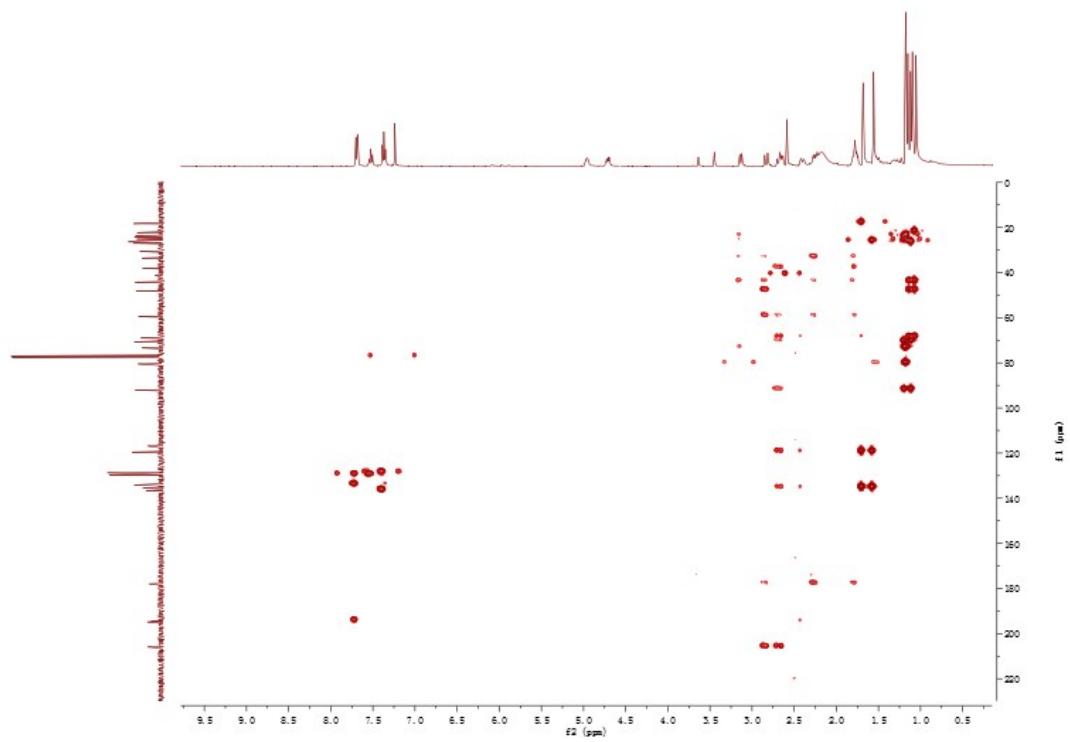
HSQC spectrum (AV-400) of hypersampsone W (5) in CDCl_3



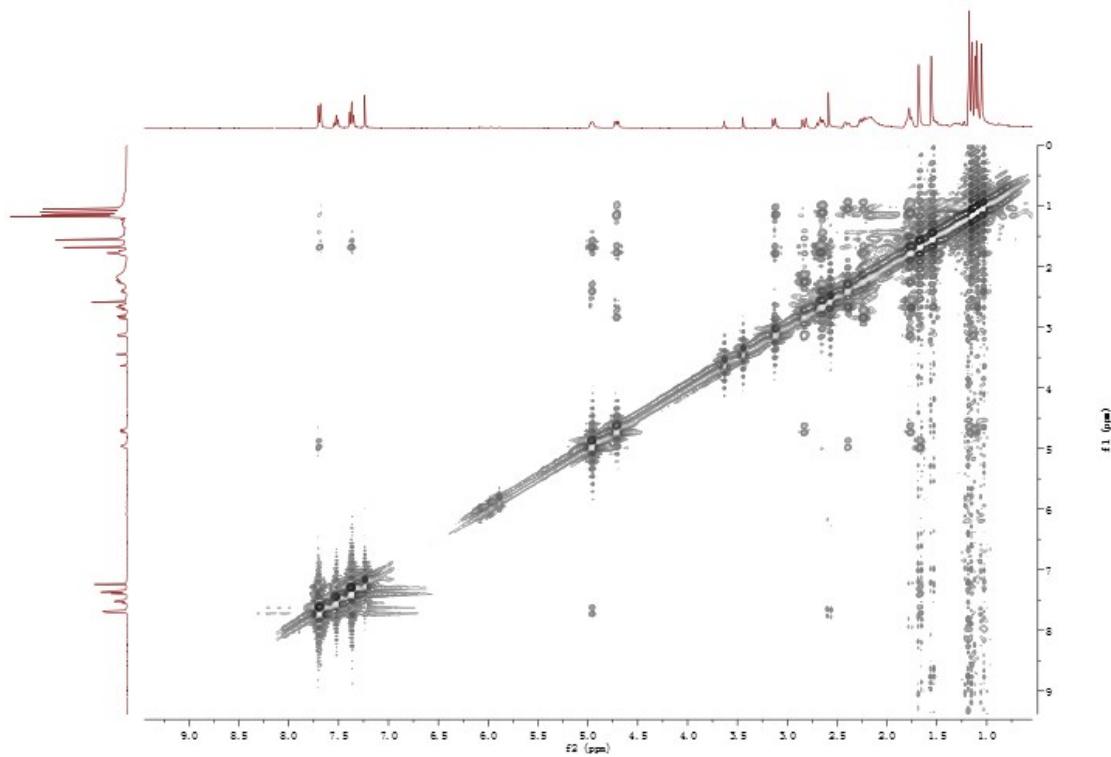
^1H - ^1H COSY spectrum (AV-400) of hypersampsone W (5) in CDCl_3



HMBC spectrum (AV-400) of hypersampsone W (5) in CDCl_3



NOESY spectrum (AV-400) of hypersampsone W (5) in CDCl_3



Computational details of **2** and **3**

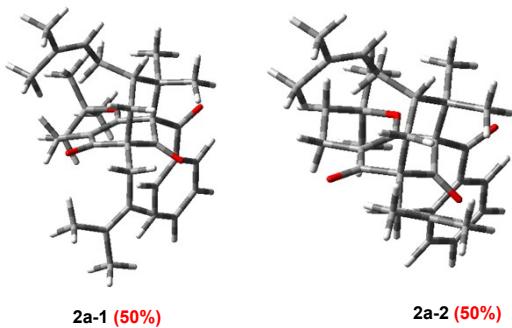
Based on the known relative configuration, two pairs of enantiomers (1R,5S,7S)-**2a**, (1S,5R,7R)-**2b**, (1R,5S,7S,25S)-**3a** and (1S,5R,7R,25R)-**3b** was employed for the conformational random search using the Merck Molecular Force Field method in the Best module of Discovery studio 2.5.5 (Accelrys, San Diego, CA, 2009) software package. The conformations form conformational search results with an energy cut off of 17 kJ/mol (approximately 4 kcal/mol) was selected to further geometry optimization and ECD calculation. As a result, there were 2, 7, 14, 8 low energy conformations with an energy cut off of 4 kcal/mol for **2a**, **2b**, **3a**, **3b**, respectively (Figure 1).

The geometry of the molecules was optimized with Gaussian 09 package¹ at B3LYP/6-31G(d) computational level. The minimum nature of the structure was confirmed by frequency calculations at the same computational level.

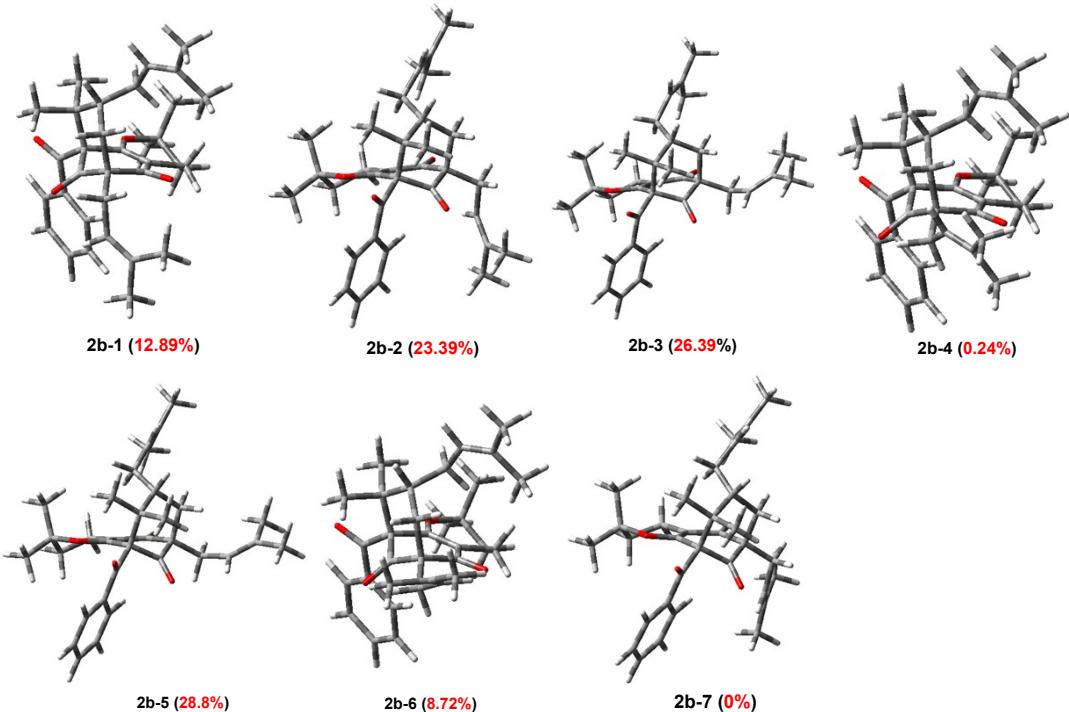
Then quantum chemical theoretical calculations (Diedrich and Grimme, 2003) for ECD were carried out in the solvent medium using time-dependent density functional theory (TDDFT) with B3LYP functional and dgdzvp forcefield basis set with 60 electronic transitions. The energies, oscillator strengths, and rotational strengths of the electronic excitations of all the conformers were calculated using the TD-DFT method at the B3LYP/dgdzvp level.

Based on the relativity energy, boltzmann weighted average of the different low energy conformations was calculated for **2a**, **2b**, **3a** and **3b**. The ECD for each molecule is calculated based on boltzmann weighted average of the conformations search results. Percentage for each conformations are shown in figure 1. As a result, the overall pattern of calculated ECD spectra of **2a**, **3a** were well matched the experimental data of **2** and **3** (Figure 2).

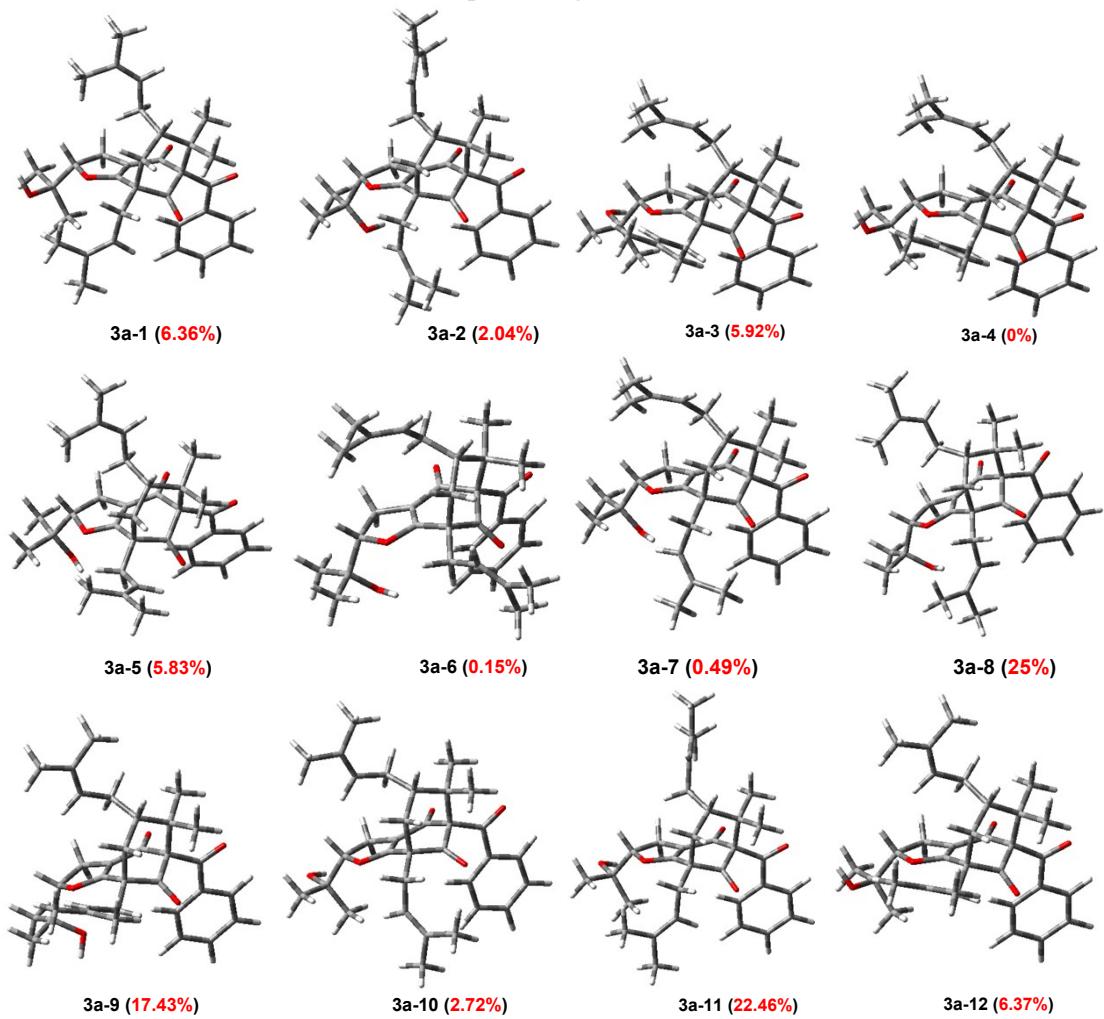
Selected conformations of **2a** and their percentage



Selected conformations of 2b and their percentage



Selected conformations of 3a and their percentage



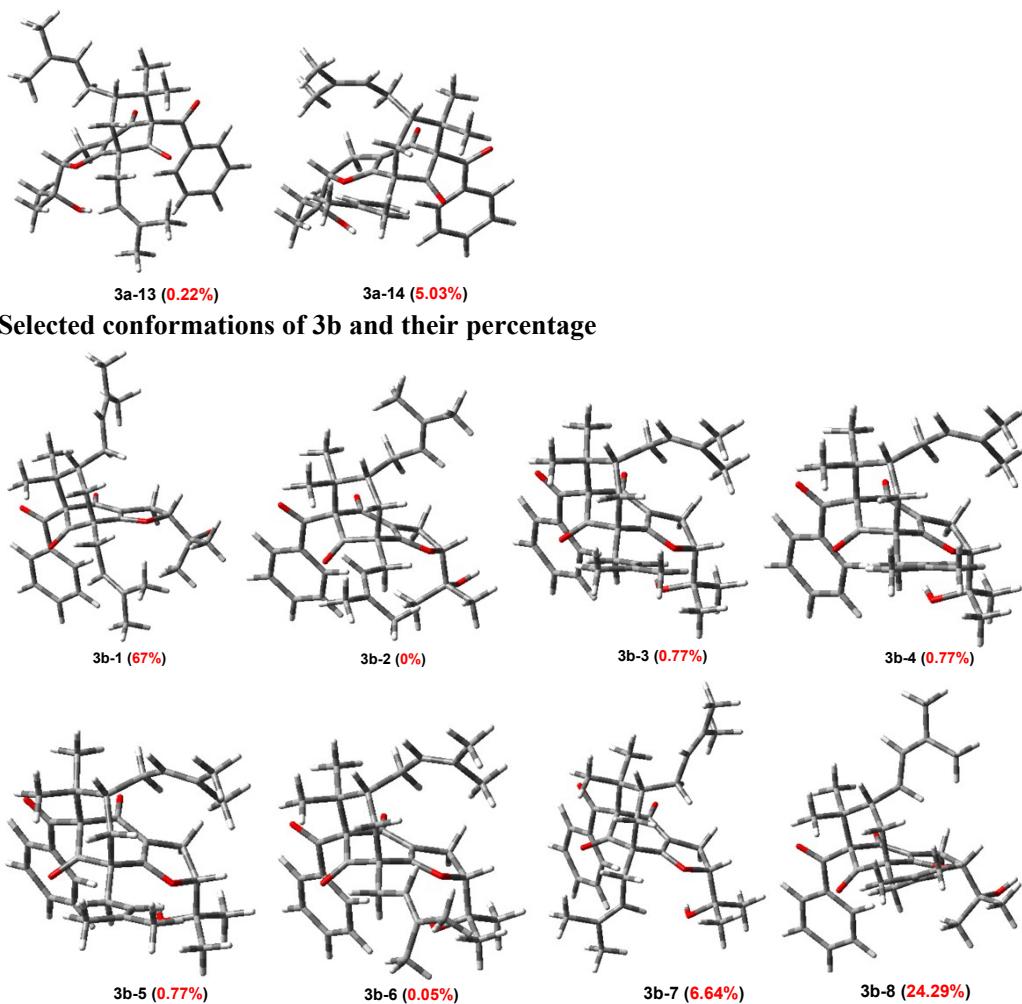


Figure 1. Selected Low energy conformations for 2a, 2b, 3a, 3b and their percentage.

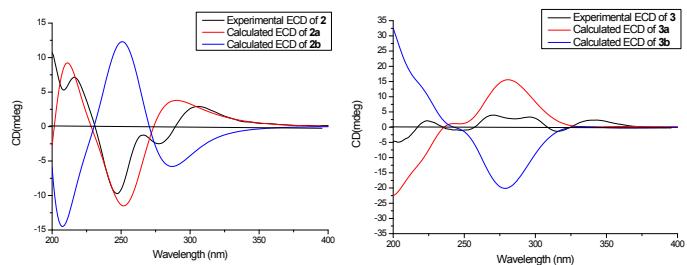


Figure 2. Calculated ECD results of 2 and 3 based on boltzmann weighted average of the selected low energy conformations.

Reference

- (1) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, O.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian, Inc., Wallingford CT, 2009.

Bioassays

RXR α transcriptional activity assay

Cell Culture. The human renal epithelial cells (293T) (ATCC) were cultured in 37 °C in DMEM (Hyclone) containing 10% fetal bovine serum (FBS, Hyclone) for 24 h.

Experimental Methods. The previous dual-luciferase reporter gene assay with some modification was used in the present study ^{1,2}. In brief, approximately 4×10^4 cells / well were seeded in 48-well plates. The two target plasmids, 20 ng pBind RXR α LBD (provided by Dr. Xiao-kun Zhang from the Burnham Institute for Medical Research, Cancer Center, La Jolla, CA, USA.) and 60 ng PG5 LUC (provided by Dr. Xiao-kun Zhang from the Burnham Institute for Medical Research, Cancer Center, La Jolla, CA, USA.), were transfected by Liposome 2000 (Invitrogen) in the cell. After 24 h, the cells were exposed to the test compound for 12 h. Then the cells were rinsed with PBS and lysed by buffered solution (1 × PLB) on the oscillating platform for 15 minutes. According to the introduction of the Dual-Luciferase Reporter Assay System kit(promega), the activities of Firefly luciferase (FL) and Rellina luciferase (RL) were checked.

$$\text{Relative luciferase activity (\%)} = \text{FL} / \text{RL} \times 100\%$$

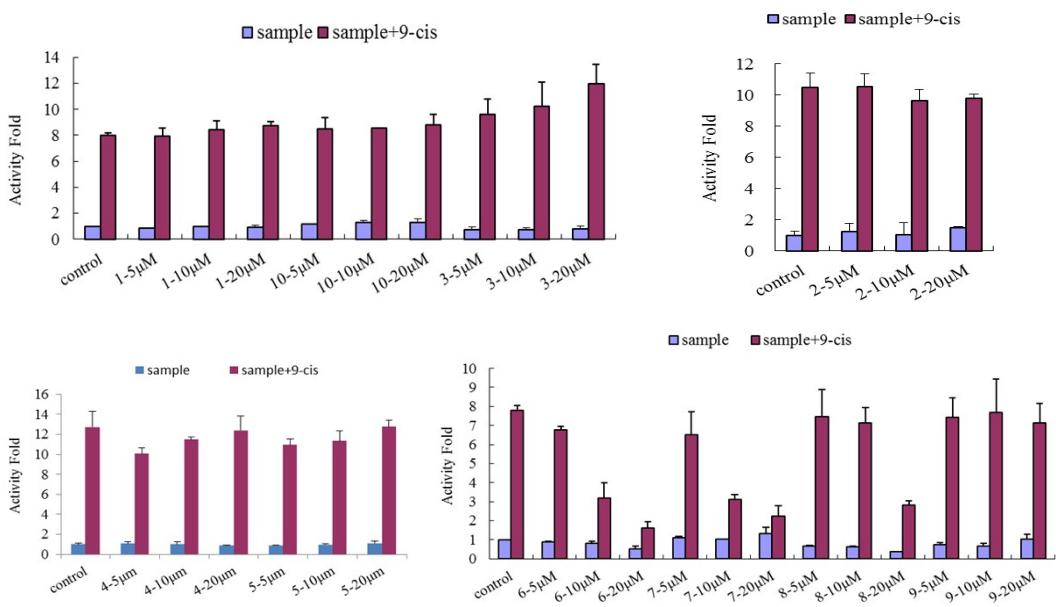


Chart 1. Effects of compounds 1-10 (5, 10, and 20 μM) on the transcriptional activities of RXR α

Cytotoxicity assay

Cell Culture. Human cervical carcinoma HeLa cells were obtained from the {American Type Culture Collection (ATCC, Manassas, VA, USA)} and were cultured in {DMEM (Hyclone)} supplemented with 10% FBS (Fetal Bovine Serum, Hyclone, USA), 100U/mL penicillin (Hyclone), and 100 μ g/mL streptomycin (Hyclone) at 37 °C with 5% CO₂ in a humidified atmosphere. The cells in the exponential phase of growth were used in the experiments.

MTT assay. All test samples were dissolved in dimethyl sulfoxide (DMSO) to make stock solutions and further diluted in culture medium upon assay. HeLa cells were incubated in 96-well cell culture clusters (JET) at a density of 0.5×10^4 cells per well and cultured for 12 h. Thereafter, the cells were treated with 5, 10 and 20 μ M concentrations of **1-10** respectively. After cultured for 48 h, 20 μ L of MTT (Solarbio) solution was added and the cells were incubated for an additional 4 h at 37 °C. Then the supernatant was discarded, and the deposited formazan formed in the cells was dissolved with 100 μ L of DMSO. All optical densities were measured in the MTT assay using a microplate reader (Thermo Multiskan MK3, Thermo Scientific, Helsinki, Finland). The percentage of cell growth rate was calculated as follows:

$$\text{Growth Rate (\%)} = (\text{OD}_{\text{sample}} - \text{OD}_{\text{blank}}) / (\text{OD}_{\text{control}} - \text{OD}_{\text{blank}}) \times 100$$

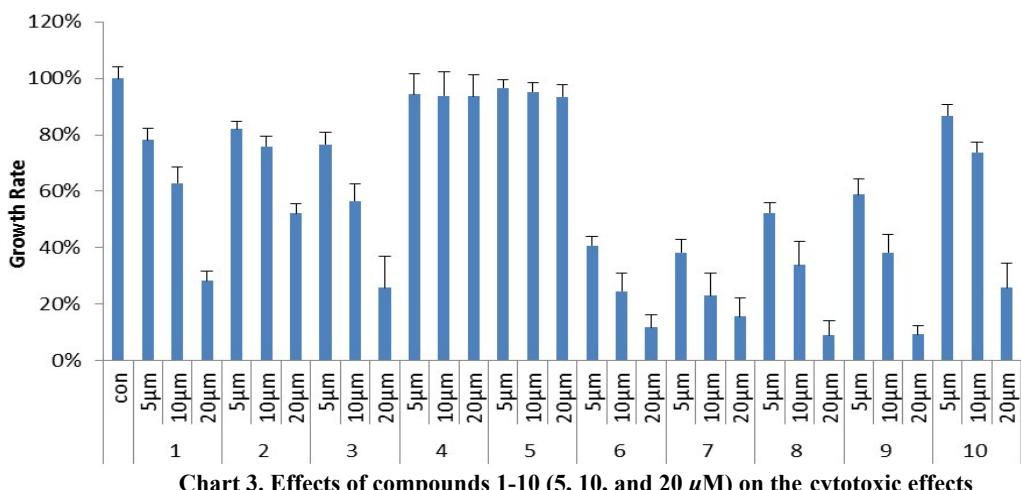


Chart 3. Effects of compounds 1-10 (5, 10, and 20 μ M) on the cytotoxic effects

Reference

- (1) Zhang, X. K.; Lehmann, J.; Hoffmann, B.; Dawson, M. I.; Cameron, J.; Graupner, G.; Hermann, T.; Tran, P.; Pfahl, M. *Nature* **1992**, 358, 587–591.
- (2) Duan, Y.H.; Dai, Y.; Wang, G.H.; Zhang, X.; Chen, H.F.; Chen, J.B.; Yao, X.S.; Zhang, X.K. *J. Nat. Prod.* **2010**, 73, 1283-1287.