

1 **Supplementary Information**

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3 **Fe₃O₄@Zirconium Phosphate Core-shell Nanoparticles for pH-Sensitive and Magnetically**
4 **Guided Drug Delivery Application**

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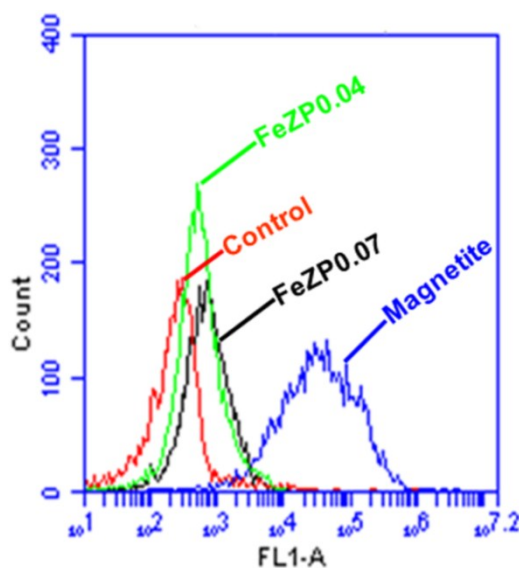
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16 1. Detection of ROS generation

17 The intracellular generation of reactive oxygen species (ROS) from the prepared
18 nanoparticles (viz. magnetite, FeZP0.07, and FeZP0.04) was measured using the oxidation
19 sensitive dye 5-(and -6) chloromethyl-2',7'-dichlorodihydrofluorescein diacetate acetyl ester
20 (CM-H₂DCFDA). For the study, MDA-MB-231 cell lines were treated with the nanoparticles for
21 24 h, after which the cells were trypsinized and suspended in PBS with CM-H₂DCFDA
22 (10 μmol/L) for 1 h and subsequently washed with PBS to remove the unreacted dye. The
23 samples were then acquired on the FL-1 channel of FACSCalibur (BD, Bioscience, USA) and
24 analyzed using Cellquest software. For the analysis, untreated MDA-MB-231 cell lines were
25 taken as the control. The generation of ROS from the nanoparticles was evaluated from the
26 fluorescence measurement of the cell lines and the results are presented in Fig. S1. A dramatic
27 ROS burst was observed in the cells treated with MNPs in contrast to the control, with profound
28 peak shift annotating the intense fluorescence. On the contrary, insignificant peak shift was
29 observed in the cells treated with FeZP0.04 inferring the production of ROS by the nanoparticles
30 to be comparable with that in control cells. Further, FeZP0.07 treatments showed a slight shift of
31 peak that indicates a small production of ROS by the nanoparticles in contrast to control cells.
32 Thus, it can be inferred that MNPs elicits ROS production, which gets significantly reduced with
33 the increase in thickness of zirconium phosphate shell on magnetite core in the core-shell
34 nanoparticles.



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36 **Fig. S1** Quantitative measurement of ROS in MDA-MB-231 cells after 24 h of treatment with
37 the nanoparticles.

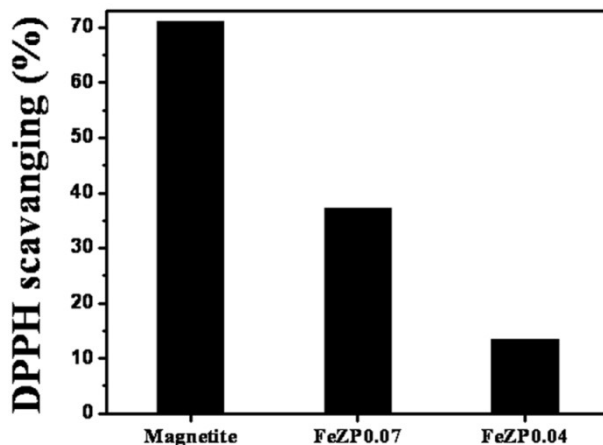
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39 2. DPPH radical scavenging analysis

40 The generation of ROS from the nanoparticles (i.e. magnetite, FeZP0.07, and FeZP0.04)
41 was further ascertained from their DPPH (2,2-diphenyl-1-picrylhydrazyl) radical scavenging
42 activity. The purple colored DPPH molecule contains a stable free radical that changes to a
43 yellow colored stable compound upon reaction with a ROS. The decaying of purple coloration of
44 the DPPH solution, that can be monitored spectrophotometrically at 517 nm thus, will give a
45 measure of the ROS produced by the nanoparticles. For the study, 10 mg nanoparticles were
46 added to 2 mL of 20 μ M DPPH solution in ethanol and agitated for 30 min in absence of light.
47 Subsequently, the absorbance of the supernatants was measured at 517 nm and the scavenging
48 percentage of the samples were calculated using the following formula:

$$49 \text{ DPPH scavenging (\%)} = (A_C - A_S / A_C) \times 100$$

50 where, A_C is the absorbance of blank DPPH and A_S is the absorbance of DPPH solution in
51 presence of nanoparticles. The results depicted in Fig. S2 revealed that MNPs exhibit DPPH
52 radical scavenging of 71.1%, whereas the scavenging was observed to be 36.2 and 13.3% in
53 FeZP0.07 and FeZP0.04 respectively. The decrease in DPPH scavenging from MNPs to
54 FeZP0.04 indicated a significant reduction in ROS generation with increase in the thickness of
55 zirconium phosphate shell on magnetite core.



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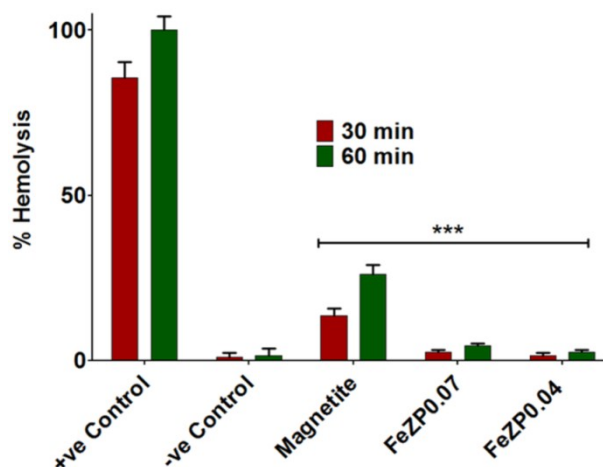
57 Fig. S2 DPPH radical scavenging percentage of the nanoparticles.

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59 3. Hemocompatibility assay

60 Hemocompatibility and hemolysis are major attributes for assessment of therapeutic
61 intervention of nanoparticles in drug delivery application, where nanoparticles with hemolysis of

62 5% or less are considered as non toxic and biocompatible¹ in therapeutic perspective. The
 63 hemocompatibility of the prepared nanoparticles (i.e. magnetite, FeZP0.07, and FeZP0.04) was
 64 ascertained from their hemocompatibility assay. The red blood cells (RBC) were collected from
 65 the blood of BALB/c male mice (6 weeks older) by centrifugation using a ficoll density gradient.
 66 For the assay, the collected RBC pellets were diluted in PBS (5% v/v) and then added to PBS (-
 67 ve control), 1% Triton X-100 (+ve control) and the nanoparticles (suspended in 1X PBS), which
 68 were then incubated at 37 °C for 30 and 60 min. The suspensions were centrifuged and the
 69 absorbance of the supernatants was measured at 570 nm. The results of hemolytic assay (shown
 70 in Fig. S3) revealed 15 and 23% hemolysis by MNPs after 30 and 60 min respectively. On the
 71 contrary, insignificant percent of hemolysis (less than 5%) was observed in FeZP0.07 and
 72 FeZP0.04 treatments even after extended exposure till 60 min, where the hemolysis was lowest
 73 for the FeZP0.04 nanoparticles. Hemolytic assay of the nanoparticles indicates that the FeZP0.04
 74 nanoparticles are hemocompatible and suitable for potential usage in biomedical fields.



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76 **Fig. S3** Hemolytic assay of magnetite, FeZP0.07, and FeZP0.04 nanoparticles, where PBS and
 77 1% Triton X-100 was taken as Negative and Positive controls respectively.

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Each bar indicate the means \pm SD (n = 3) with P < 0.001.

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80 4. References

81 1 M. He, Z. Zhao, L. Yin, C. Tang and C. Yin, *Int. J. Pharm.*, 2009, **373**, 165-173.