

A Convergent Total Synthesis of Ouabagenin

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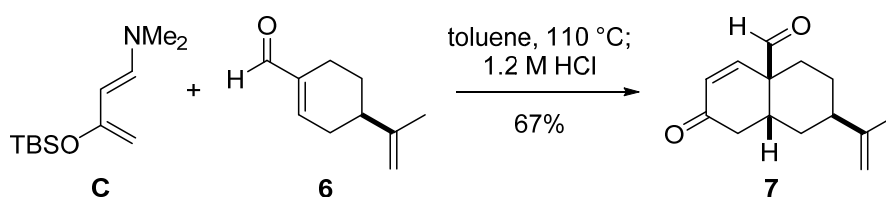
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Supporting Information

58 pages

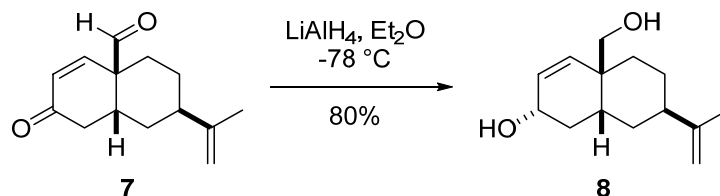
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General Methods: All reactions sensitive to air or moisture were carried out under argon atmosphere under anhydrous conditions, unless otherwise noted. THF, CH₂Cl₂, toluene, DMF and Et₂O were purified by Glass Contour solvent dispensing system (Nikko Hansen & Co., Ltd., Osaka, Japan). All other reagents were used as supplied. Analytical thin-layer chromatography (TLC) was performed using E. Merck Silica gel 60 F254 pre-coated plates. Flash chromatography was performed using 40-50 μ m Silica Gel 60N (Kanto Chemical Co., Inc.) or 32-53 μ m Silica-gel BW-300 (Fuji Silysia Chemical Ltd.). Melting points were measured on Yanaco MP-J3 micro melting point apparatus, and are uncorrected. Optical rotations were measured on JASCO DIP-1000 Digital Polarimeter at room temperature using the sodium D line. Infrared (IR) spectra were recorded as a thin film on a NaCl disk for synthetic intermediates and a KBr disk for ouabagenin (**1**) using JASCO FT/IR-4100 spectrometer. ¹H and ¹³C NMR spectra were recorded on JEOL JNM-ECX-500, JNM-ECA-500, or JNM-ECS-400 spectrometer at room temperature. Chemical shifts were reported in ppm on the δ scale relative to CHCl₃ (δ = 7.26 for ¹H NMR), CDCl₃ (δ = 77.0 for ¹³C NMR), CD₂HOD (δ = 3.31 for ¹H NMR), CD₃OD (δ = 49.0 for ¹³C NMR), C₆D₅H (δ = 7.16 for ¹H NMR), C₆D₆ (δ = 128.06 for ¹³C NMR), (CD₂H)CD₃SO (δ = 2.50 for ¹H NMR) and (CD₃)₂SO (δ = 39.52 for ¹³C NMR), as internal references. Signal patterns are indicated as s, singlet; d, doublet; m, multiplet; br, broaden peak. The numbering of compounds corresponds to that of **1**. High resolution mass spectra were measured on JEOL JMS-T100LP instrument (ESI-TOF) or BRUKER DALTONICS microTOF II (ESI-TOF).

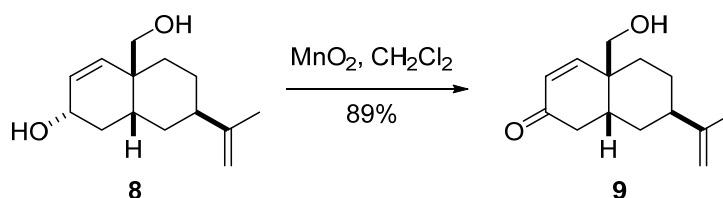


Aldehyde 7. Rawal's diene **C**^{S1} (46.4 g, 204 mmol) was added to a solution of (*R*)-perillaldehyde **6**^{S2} (15.9 g, 106 mmol) in refluxing toluene in five portions every three hours. After the consumption of **6** was confirmed by ¹H-NMR analysis of the reaction mixture, the mixture was cooled to room temperature and concentrated. THF (530 mL) and 1.2 M HCl (270 mL) were successively added to the resultant residue. The mixture was stirred at room temperature for 13 h, and then was concentrated to remove THF. After brine (200 mL) was added, the resultant mixture was extracted with EtOAc (600 mL x3). The combined organic layers were washed with brine (500 mL). The aqueous layers were re-extracted with EtOAc (500 mL). Then, the combined organic layers were

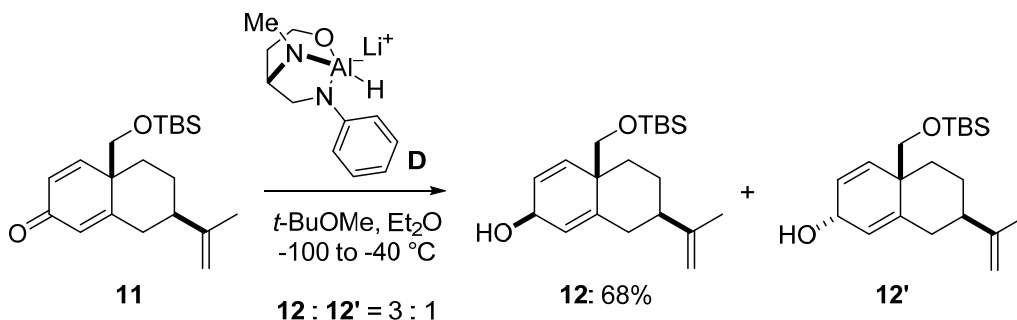
dried over Na₂SO₄, filtrated, and concentrated. The residue was purified by flash column chromatography on silica gel (800 g, hexane to hexane/EtOAc 5/1) to afford **7** (15.5 g, 71.0 mmol) in 67%: colorless oil; $[\alpha]_D^{25}$ -190 (*c* 1.8, CHCl₃). IR, ¹H and ¹³C NMR data of **7** were identical to those of the antipode reported previously.^{S3}



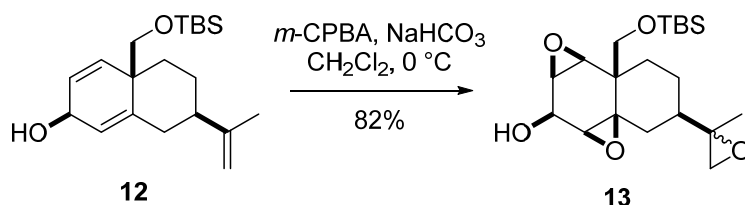
Alcohol 8. LiAlH₄ (6.7 g, 180 mmol) was added to a solution of **7** (15.5 g, 71.0 mmol) in Et₂O (710 mL) at -78 °C. The reaction mixture was stirred at -78 °C for 7 h, and then H₂O (7 mL), aqueous 15 wt% NaOH (7 mL) and H₂O (21 mL) were successively added at -78 °C. The mixture was warmed to room temperature, and was filtered through a pad of Celite with EtOAc (300 mL). The filtrate was concentrated, and the residue was purified by flash column chromatography on silica gel (430 g, CH₂Cl₂ to CH₂Cl₂/EtOAc 2/1) to afford diol **8** (12.6 g, 56.7 mmol) in 80% yield: white crystal; m.p. 97-98 °C; $[\alpha]_D^{25}$ -32 (*c* 2.6, CHCl₃); IR (neat) ν 3345, 2931, 2862, 1644, 1456, 1052, 1032 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.21-1.32 (1H, m, H8_a), 1.27-1.35 (1H, m, OH) 1.45-1.51 (1H, m, H6_a), 1.52-1.62 (3H, m, H8_b and 9), 1.71 (3H, s, CCH₃), 1.73 (1H, ddd, *J* = 13.8, 13.8, 5.2 Hz, H6_b), 1.79-1.84 (2H, m, H4), 2.02-2.08 (1H, m, H5), 2.08 (1H, m, H7), 3.49 (1H, d, *J* = 10.9 Hz, H19_a), 3.71 (1H, dd, *J* = 10.9, 4.6 Hz, H19_b), 4.36 (1H, m, H3), 4.67-4.71 (2H, m, CCH₃=CH₂), 5.51 (1H, dd, *J* = 10.3, 2.3 Hz, H2), 5.76 (1H, dd, *J* = 10.3, 1.7 Hz, H1); ¹³C NMR (125 MHz, CDCl₃) δ 20.9, 27.3, 30.3, 31.5, 33.4, 35.6, 39.3, 40.4, 66.7, 68.7, 108.5, 132.6, 136.0, 150.1; HRMS (ESI) calcd for C₁₄H₂₂O₂Na [M+Na]⁺ 245.1512, found 245.1512.



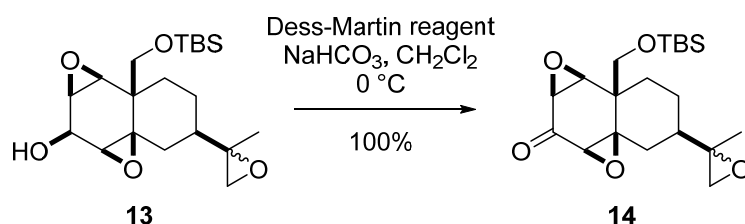
Enone 9. MnO₂ (90% purity, 50.4 g, 522 mmol) was added to a solution of diol **8** (7.74 g, 34.8 mmol) in CH₂Cl₂ (120 mL) at room temperature. The reaction mixture was stirred at room temperature for 7 h, and then was filtered through a pad of Celite with CH₂Cl₂ (300 mL), MeOH (500 mL) and a 3 : 1 mixture of CHCl₃ and MeOH (500 mL). The filtrate was concentrated, and the residue was purified by flash column chromatography on silica gel (100 g, hexane to hexane/Et₂O 1/2) to afford enone **9** (6.86 g, 31.1 mmol) in 89% yield:



Allylic alcohol 12. LiAlH_4 (1.0 M in Et_2O , 140 mL, 140 mmol) was added to a solution of (*S*)-4-anilino-3-methylamino-1-butanol (27.2 g, 140 mmol) in *t*-BuOMe (100 mL) at 0 °C over 40 min. The mixture was warmed to room temperature, and stirred for 3 h. After the mixture was cooled to -100 °C, a solution of dienone **11** (7.76 g, 23.2 mmol) in *t*-BuOMe (17 mL) was added over 10 min. The reaction mixture was stirred at -100 °C for 4 h and -40 °C for 20 h, and then MeOH (5 mL) and H_2O (30 mL) were successively added. After being warmed at room temperature, the resultant solution was filtered through a pad of Celite with Et_2O (1 L). The filtrate was washed with brine (200 mL), dried over Na_2SO_4 , and filtrated. Hexane (1 L) and Et_3N (10 mL) were added to the solution, and the combined solution was filtered through a pad of silica gel (50 g, hexane/ Et_2O / Et_3N 100/100/1) to remove (*S*)-4-anilino-3-methylamino-1-butanol. The filtrate was concentrated to afford a 3 : 1 diastereomeric mixture of allylic alcohol **12** and **12'**. The residue was purified by flash column chromatography on silica gel (250 g, hexane/ Et_3N 100/1 to hexane/ EtOAc / Et_3N 150/10/1) to afford **12** (5.27 g, 15.8 mmol) in 68% yield and impure **12'** (2.0 g): colorless oil; $[\alpha]_{\text{D}}^{26}$ -22 (*c* 1.2, CHCl_3); IR (neat) ν 3348, 2930, 2857, 1644, 1470, 1442, 1255, 1092, 839 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 0.035 (3H, s, CH_3 of TBS), 0.041 (3H, s, CH_3 of TBS), 0.88 (9H, s, *t*-Bu of TBS), 1.29 (1H, ddd, $J = 13.7, 13.7, 4.6$ Hz, H9_a), 1.42-1.52 (1H, m, H8_a), 1.47 (1H, d, $J = 10.9$ Hz, OH), 1.64-1.71 (1H, m, H8_b), 1.74 (3H, s, $\text{CCH}_3=\text{CH}_2$), 1.81 (1H, ddd, $J = 13.7, 3.5, 3.5$ Hz, H9_b), 1.97 (1H, dddd, $J = 12.6, 12.6, 3.5, 3.5$ Hz, H7), 2.12 (1H, d, $J = 13.8, 12.6, 1.2$ Hz, H6_a), 2.22 (1H, ddd, $J = 13.8, 3.5, 2.3$ Hz, H6_b), 3.48 (1H, d, $J = 9.8$ Hz, H19_a), 3.72 (1H, d, $J = 9.8$ Hz, H19_b), 4.39 (1H, ddd, $J = 10.9, 5.2, 4.6$ Hz, H3), 4.71-4.74 (2H, m, $\text{CCH}_3=\text{CH}_2$), 5.68 (1H, d, $J = 9.8$ Hz, H1), 5.82-5.85 (1H, m, H4), 5.97 (1H, ddd, $J = 9.8, 4.6, 1.7$ Hz, H2); ^{13}C NMR (125 MHz, CDCl_3) δ -5.5, -5.4, 18.6, 20.8, 26.0, 27.1, 34.1, 37.3, 42.4, 46.6, 62.6, 65.5, 108.8, 124.1, 127.0, 137.1, 142.7, 149.3; HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{34}\text{O}_2\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 357.2220, found 357.2222.

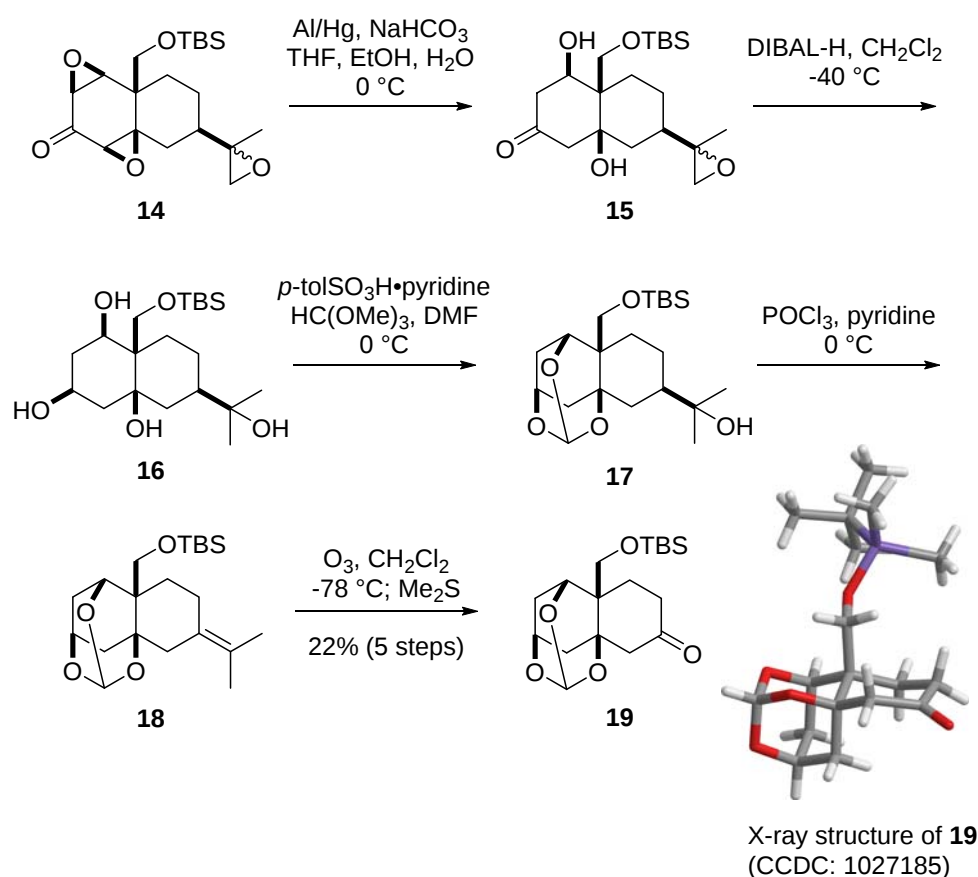


Tris-epoxide 13. *m*-CPBA (77% purity, 12.4 g, 55.2 mmol) was added to a mixture of allylic alcohol **12** (5.27 g, 15.8 mmol) and NaHCO₃ (5.3 g, 63 mmol) in CH₂Cl₂ (160 mL) at 0 °C. The reaction mixture was stirred at 0 °C for 3 h, and then saturated aqueous Na₂S₂O₃ (20 mL) was added. After saturated aqueous NaHCO₃ (40 mL) was added, the resultant solution was extracted with EtOAc (150 mL x3). The combined organic layers were washed with NaHCO₃ (100 mL x3) and brine (200 mL), dried over Na₂SO₄, filtrated, and concentrated. The residue was purified by flash column chromatography on silica gel (100 g, hexane to EtOAc) to afford a 1 : 1 diastereomeric mixture of tris-epoxide **13** (4.93 g, 12.9 mmol) in 82% yield: white solid; m.p. 116-117 °C; $[\alpha]_D^{27}$ -9.9 (*c* 1.0, CHCl₃); IR (neat) ν 3414, 2935, 2860, 1645, 1467, 1254, 1095, 1034, 841 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.09 (3H, s, CH₃ of TBS), 0.10 (3H, s, CH₃ of TBS), 0.91 (9H, s, *t*-Bu of TBS), 1.01-1.13 (2H, m), 1.26 (3/2H, s, CH₃), 1.27 (3/2H, s, CH₃), 1.38-1.48 (2H, m), 1.65-1.78 (1H, m), 1.78-1.92 (1H, m), 2.22-2.28 (1H, m), 2.42-2.52 (1H, m), 2.54 (1H, dd, *J* = 4.5, 4.5 Hz), 2.62 (1H, d, *J* = 4.5 Hz), 2.98-3.03 (1H, m), 3.18-3.20 (1H, m), 3.35 (1H, m), 3.81-3.84 (1H, m), 3.98-4.00 (1H, m), 4.21-4.27 (1H, m); ¹³C NMR (125 MHz, CDCl₃) δ -5.60, -5.59, 18.21, 18.25, 18.35, 22.3, 22.7, 25.8, 28.58, 28.61, 34.0, 34.4, 37.5, 37.6, 43.3, 52.9, 53.1, 54.2, 58.36, 58.39, 60.82, 60.84, 60.95, 61.13, 61.16, 65.1, 65.4, 65.5, some of ¹³C peaks were not observed due to overlap with other peaks; HRMS (ESI) calcd for C₂₀H₃₄O₅SiNa [M+Na]⁺ 405.2068, found 405.2064.



Ketone 14. Dess-Martin periodinane (11.2 g, 26.4 mmol) was added to a 1 : 1 diastereomeric mixture of tris-epoxide **13** (6.71 g, 17.5 mmol) and NaHCO₃ (4.4 g, 52 mmol) in CH₂Cl₂ (180 mL) at 0 °C. The reaction mixture was stirred at room temperature for 5 h, and then saturated aqueous Na₂S₂O₃ (20 mL) was added. After saturated aqueous NaHCO₃ (20 mL) was added, the resultant solution was extracted with EtOAc (150 mL x3). The combined organic layers were washed with brine (150 mL), dried over Na₂SO₄, filtrated, and concentrated. The residue was purified by flash column chromatography on

silica gel (200 g, hexane to hexane/EtOAc 4/1) to afford a 1 : 1 diastereomeric mixture of ketone **14** (6.67 g, 17.5 mmol) in 100 % yield: colorless oil; $[\alpha]_D^{25}$ -10 (c 2.0, CHCl_3); IR (neat) ν 2932, 2884, 2857, 1705, 1468, 1445, 1388, 1254, 1102, 942, 840 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 0.12 (3H, s, CH_3 of TBS), 0.13 (3H, s, CH_3 of TBS), 0.92 (9H, s, t -Bu of TBS), 1.01-1.08 (1H, m), 1.16 (1H, ddd, J = 13.8, 13.8, 4.6 Hz), 1.27 (3H, s, CH_3), 1.38-1.57 (2H, m), 1.72-1.80 (1H, m), 1.88 (1/2H, dd, J = 13.8 Hz, H7), 1.94 (1/2H, dd, J = 13.8 Hz, H7), 2.28-2.34 (1H, m), 2.54-2.57 (1H, m), 2.60-2.64 (1H, m), 3.07-3.10 (1H, m), 3.41-3.43 (1H, m), 3.48 (1H, d, J = 4.0 Hz), 4.01-4.05 (1H, m), 4.06-4.10 (1H, m); ^{13}C NMR (125 MHz, CDCl_3) δ -5.6, 14.1, 18.2, 18.3, 18.6, 21.9, 22.2, 22.6, 25.8, 27.8, 27.9, 31.6, 33.7, 34.0, 39.50, 39.54, 42.9, 43.0, 52.7, 53.1, 55.9, 58.08, 58.11, 61.38, 61.40, 61.44, 64.6, 64.7, 64.6, 64.7, 70.30, 70.34, 200.7, some of ^{13}C peaks were not observed due to overlap with other peaks; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{36}\text{O}_6\text{SiNa}$ $[\text{M}+\text{MeOH}+\text{Na}]^+$ 435.2173, found 435.2168.



Ketone 19. Al/Hg was prepared from small pieces of aluminum foil (2.8 g) by treating with 2 wt% aqueous HgCl_2 (100 mL) for 20 seconds and washing with EtOH and Et_2O . The freshly prepared Al/Hg was added to a solution of a 1 : 1 diastereomeric mixture of ketone **14** (7.14 g, 18.6 mol) in a mixture of THF (214 mL), EtOH (71 mL), H_2O (71 mL) and saturated aqueous NaHCO_3 (14 mL) at 0 °C. The reaction mixture was stirred at 0 °C

for 3 h. After Et₂O (1 L) cooled to -40 °C was added, the resultant solution was filtered through a pad of Celite with Et₂O. The filtrate was washed with brine (400 mL), dried over Na₂SO₄, filtrated, and directly subjected to flash column chromatography on silica gel (50 g, Et₂O) to afford the impure diol **15**, which was used in the next reaction without further purification.

DIBAL-H (1.0 M in hexane, 93 mL, 93 mmol) was added to a solution of the above crude diol **15** in CH₂Cl₂ (370 mL) at -40 °C over 20 min. The reaction mixture was stirred at -40 °C for 10 min, and then saturated aqueous Rochelle's salt (100 mL) was added. The resultant solution was stirred at room temperature for 20 h. The mixture was extracted with EtOAc (350 mL x3), and the combined organic layers were washed with brine (400 mL), dried over Na₂SO₄, filtrated, and concentrated to afford the impure tetraol **16**, which was used in the next reaction without further purification.

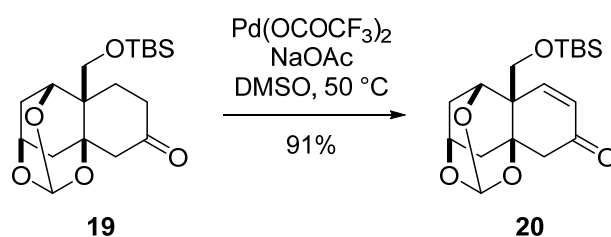
p-tolSO₃H·pyridine (466 mg, 1.85 mmol) was added to a solution of the above crude tetraol **16** and HC(OMe)₃ (100 mL, 920 mmol) in DMF (370 mL) at 0 °C. The reaction mixture was stirred at 0 °C for 2 h, and then saturated aqueous NaHCO₃ (100 mL) was added. The resultant solution was extracted with Et₂O (400 mL x3), and the combined organic layers were washed with H₂O (400 mL x2), brine (400 mL), dried over Na₂SO₄, filtrated, and concentrated. The residue was purified by flash column chromatography on silica gel (200g, hexane/Et₃N 100/1 to hexane/Et₂O/Et₃N 100/100/1) to afford the impure orthoester **17**, which was used in the next reaction without further purification.

POCl₃ (2.1 mL, 23 mmol) was added to a solution of the above crude orthoester **17** in pyridine (74 mL) at 0 °C. The reaction mixture was stirred at 0 °C for 16 h, and then saturated aqueous NaHCO₃ (50 mL) was added. After H₂O (20 mL) was added, the resultant solution was extracted with EtOAc (100 mL x3). The combined organic layers were washed with brine (100 mL), dried over Na₂SO₄, filtrated, and concentrated. The residue was purified by flash column chromatography on silica gel (75 g, hexane to EtOAc) to afford the impure **18**, which was used in the next reaction without further purification:

For characterization of **18**, a small amount of the mixture was purified by flash column chromatography on silica gel (75 g, hexane to EtOAc). **18**: colorless oil; $[\alpha]_D^{26} +1.4$ (*c* 1.8, CHCl₃); IR (neat) ν 2953, 2929, 2856, 1463, 1377, 1250, 1174, 1132, 1100, 1088, 991 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.066 (3H, s, CH₃ of TBS), 0.069 (3H, s, CH₃ of TBS), 0.90 (9H, s, *t*-Bu of TBS), 1.20 (1H, ddd, *J* = 13.2, 13.2, 5.2 Hz, H_{9a}), 1.66 (3H, s, CH₃), 1.69 (3H, s, CH₃), 1.76 (1H, dd, *J* = 13.8, 1.7 Hz, H_{2a}), 1.81 (1H, ddd, *J* = 13.2, 5.7, 2.3 Hz, H_{9b}), 1.94 (1H, ddd, *J* = 13.8, 3.5, 2.3 Hz, H_{4a}), 2.01-2.08 (1H, m, H_{4b}), 2.01-2.08 (1H, m, H_{8a}), 2.11 (1H, d, *J* = 13.7 Hz, H_{6a}), 2.35 (1H, d, *J* = 13.7 Hz, H_{6b}), 2.54-2.62 (2H, m, H_{2b}

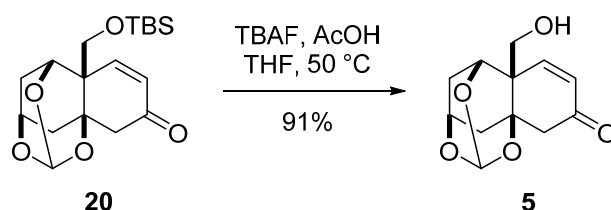
and 8_b), 4.02 (1H, d, $J = 9.8$ Hz, H19_a), 4.15-4.17 (1H, m, H1), 4.21-4.23 (1H, m, H3), 4.42 (1H, d, $J = 9.8$ Hz, H19_b), 5.68 (1H, s, CH); ^{13}C NMR (125 MHz, CDCl_3) δ -5.54, -5.46, 18.3, 20.3, 20.4, 24.3, 25.0, 25.9, 28.8, 34.9, 36.9, 43.0, 59.0, 67.3, 69.8, 74.8, 106.3, 125.6, 126.1; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{36}\text{O}_4\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 403.2275, found 403.2268.

Ozone was bubbled into a stirred solution of the above crude **18** in CH_2Cl_2 (57 mL) at -78 °C for 1 h. After oxygen was bubbled into the reaction mixture for 30 min, Me_2S (3.0 mL, 41 mmol) was added at -78 °C. The resultant solution was warmed to room temperature, stirred for 12 h, and then concentrated. The residue was purified by flash column chromatography on silica gel (75 g, hexane to hexane/EtOAc 2/1) to afford ketone **19** (1.42 g, 4.01 mmol) in 22% yield over 5 steps: crystal; m.p. 118-119 °C; $[\alpha]_{\text{D}}^{26}$ -11 (c 1.2, CHCl_3); IR (neat) ν 2954, 2931, 2897, 2856, 1716, 1470, 1284, 1249, 1146, 1091, 995, 974 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 0.09 (3H, s, CH_3 of TBS), 0.10 (3H, s, CH_3 of TBS), 0.90 (9H, s, t -Bu of TBS), 1.51 (1H, ddd, $J = 13.7, 12.6, 6.3$ Hz, H9_a), 1.82 (1H, dd, $J = 13.8, 1.7, 1.7$ Hz, H2_a), 1.86 (1H, d, $J = 13.8$ Hz, H4_a), 2.11 (1H, ddd, $J = 13.7, 8.1, 1.8$ Hz, H9_b), 2.13-2.19 (1H, m, H4_b), 2.27 (1H, dd, $J = 14.9, 1.8$ Hz, H6_a), 2.42 (1H, ddd, $J = 16.6, 6.3, 1.8$ Hz, H8_a), 2.55 (1H, ddd, $J = 16.6, 12.6, 8.1$ Hz, H8_b), 2.64 (1H, ddd, $J = 13.8, 2.9, 2.9$ Hz, H2_b), 2.68 (1H, d, $J = 14.9$ Hz, H6_b), 4.00 (1H, d, $J = 10.3$ Hz, H19_a), 4.23-4.27 (2H, m, H1 and 3), 4.48 (1H, d, $J = 10.3$ Hz, H19_b), 5.66 (1H, s, CH); ^{13}C NMR (125 MHz, CDCl_3) δ -5.6, 18.2, 23.4, 25.9, 28.5, 35.9, 37.5, 42.5, 50.2, 59.6, 66.4, 69.5, 73.9, 105.8, 207.0; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{30}\text{O}_5\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 377.1755, found 377.1758.

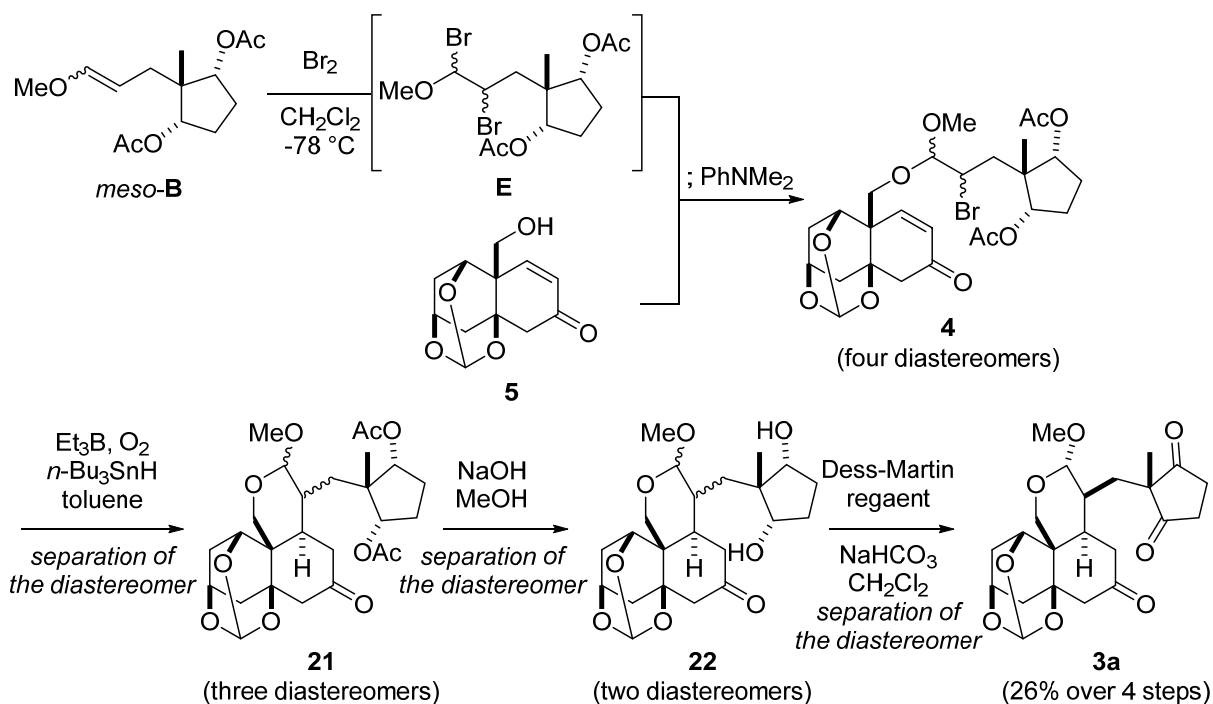


Enone 20. $\text{Pd}(\text{OCOCF}_3)_2$ (550 mg, 1.65 mmol) was added to a solution of ketone **19** (492 mg, 1.39 mmol) and NaOAc (342 mg, 4.17 mmol) in DMSO (28 mL) at room temperature. The reaction mixture was heated to 50 °C, and stirred for 21 h. After $\text{Pd}(\text{OCOCF}_3)_2$ (208 mg, 0.636 mmol) and NaOAc (110 mg, 1.34 mmol) were added, the reaction mixture was further stirred at 50 °C for 21 h. The mixture was cooled to room temperature, and then saturated aqueous NaHCO_3 (30 mL) was added. The resultant solution was extracted with EtOAc (50 mL x3), and the combined organic layers were washed with brine (50 mL), dried over Na_2SO_4 , filtrated, and concentrated. The residue was purified by flash column chromatography on silica gel (50 g, hexane to hexane/ EtOAc 4/1) to afford enone **20** (445 mg, 1.26 mmol) in 91% yield: crystal; m.p. 118 °C; $[\alpha]_{\text{D}}^{24}$ +110 (c 0.83, CHCl_3); IR (neat)

ν 2952, 2929, 2885, 2855, 1679, 1470, 1385, 1283, 1251, 1173, 1132, 1098, 995, 972 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 0.03 (6H, s, CH_3 of TBS x2), 0.85 (9H, s, *t*-Bu of TBS), 1.60 (1H, ddd, $J = 13.8, 1.7, 1.7$ Hz, $\text{H}_{2\text{a}}$), 2.12-2.20 (2H, m, H_4), 2.34 (1H, d, $J = 17.2$ Hz, $\text{H}_{6\text{a}}$), 2.73 (1H, dddd, $J = 13.8, 4.0, 4.0, 2.3$ Hz, $\text{H}_{2\text{b}}$), 2.96 (1H, d, $J = 17.2$ Hz, $\text{H}_{6\text{b}}$), 4.22 (1H, d, $J = 9.7$ Hz, $\text{H}_{19\text{a}}$), 4.24 (1H, d, $J = 9.7$ Hz, $\text{H}_{19\text{b}}$), 4.26-4.27 (1H, m, H_3), 4.40 (1H, d, $J = 4.0$ Hz, H_1), 5.70 (1H, s, CH), 6.11 (1H, d, $J = 9.7$ Hz, H_8), 6.69 (1H, d, $J = 9.7$ Hz, H_9); ^{13}C NMR (125 MHz, CDCl_3) δ -5.7, -5.6, 18.2, 25.8, 30.2, 36.4, 46.4, 47.2, 66.3, 66.6, 69.7, 73.6, 105.8, 130.8, 150.4, 196.4; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{28}\text{O}_5\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 375.1598, found 375.1599.



AB-Ring 5. TBAF (1.0 M in THF, 3.8 mL, 3.8 mmol) was added to a solution of enone **20** (445 mg, 1.26 mmol) and AcOH (290 μL , 5.0 mmol) in THF (13 mL) at room temperature. The reaction mixture was warmed to 50 $^\circ\text{C}$, and stirred for 17 h. After the reaction mixture was cooled to room temperature, pH 7 phosphate buffer solution (13 mL) was added. The resultant solution was extracted with CH_2Cl_2 (13 mL x5), and the combined organic layers were washed with brine (10 mL), dried over Na_2SO_4 filtrated, and concentrated. The residue was purified by flash column chromatography on silica gel (50 g, hexane to EtOAc) to afford AB-ring **5** (273 mg, 1.15 mmol) in 91% yield: crystal; m.p. 187 $^\circ\text{C}$; $[\alpha]_{\text{D}}^{27} +100$ (c 0.49, CHCl_3); IR (neat) ν 3468, 2996, 2957, 2919, 1673, 1427, 1384, 1286, 1224, 1146, 1125, 1070, 1039, 995, 963 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.62 (1H, ddd, $J = 13.8, 1.7, 1.7$ Hz, $\text{H}_{2\text{a}}$), 1.91 (1H, br s, OH), 2.14 (1H, br d, $J = 13.2$ Hz, $\text{H}_{4\text{a}}$), 2.19 (1H, ddd, $J = 13.2, 3.4, 2.3$ Hz, $\text{H}_{4\text{b}}$), 2.37 (1H, d, $J = 17.2$ Hz, $\text{H}_{6\text{a}}$), 2.75 (1H, dddd, $J = 13.8, 4.0, 4.0, 2.3$ Hz, $\text{H}_{2\text{b}}$), 2.96 (1H, d, $J = 17.2$ Hz, $\text{H}_{6\text{b}}$), 4.25 (1H, d, $J = 10.3$ Hz, $\text{H}_{19\text{a}}$), 4.28-4.31 (1H, m, H_3), 4.37 (1H, d, $J = 10.3$ Hz, $\text{H}_{19\text{b}}$), 4.45-4.47 (1H, m, H_1), 5.71 (1H, s, CH), 6.16 (1H, d, $J = 9.8$ Hz, H_8), 6.77 (1H, d, $J = 9.8$ Hz, H_9); ^{13}C NMR (125 MHz, CDCl_3) δ 30.1, 36.2, 46.3, 47.0, 66.22, 66.25, 69.5, 73.6, 105.8, 131.3, 149.7, 196.2; HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{14}\text{O}_5\text{Na}$ $[\text{M}+\text{Na}]^+$ 261.0733, found 261.0731.



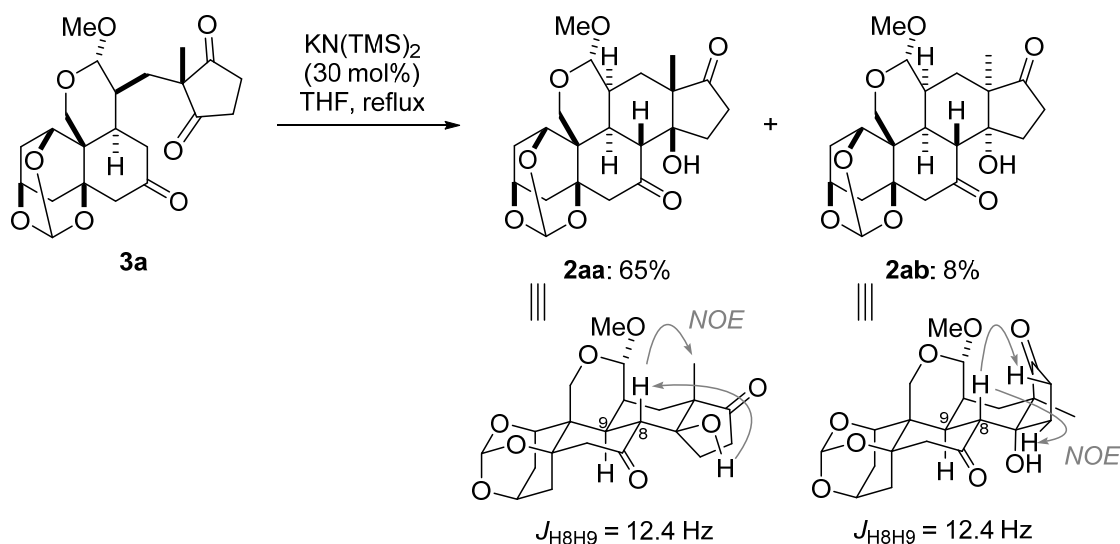
Triketone 3a. A solution of *meso*-**B** (577 mg, 2.13 mmol) in CH_2Cl_2 (2 mL) was added to a solution of Br_2 (88 μL , 1.7 mmol) in CH_2Cl_2 (2 mL) at -78°C over 20 min. After the color of the solution became clear, a solution of AB-ring **5** (204 mg, 0.856 mmol) and PhNMe_2 (430 μL , 3.4 mmol) in CH_2Cl_2 (24 mL) was added to the mixture at -78°C over 30 min. The reaction mixture was warmed to room temperature and stirred for 18 h, and then saturated aqueous NaHCO_3 (30 mL) was added. The resultant mixture was extracted with CH_2Cl_2 (30 mL x3), and the combined organic layers were washed with brine (30 mL), dried over Na_2SO_4 , filtrated, and concentrated. The residue was purified by flash column chromatography on silica gel (70 g, hexane to hexane/ EtOAc 10/1 to 4/1 to 2/1 to 1/1 to 1/2) to afford a mixture of four diastereomers of bromoacetal **4**, which was used in the next reaction without further purification.

A solution of the above crude **4** and $n\text{-Bu}_3\text{SnH}$ (67 μL , 0.25 mmol) in toluene (90 mL) was degassed by freeze-thaw cycles (x3), and then argon gas was charged into the flask. A solution of Et_3B (1.0 M in hexane, 2.5 mL, 2.5 mmol) was added to the mixture. Then a solution of $n\text{-Bu}_3\text{SnH}$ (225 μL , 0.836 mmol) in toluene (saturated with oxygen, 5 mL) was added over 30 min. After the reaction mixture was stirred at room temperature for 30 min, toluene (saturated with oxygen, 5 mL) was added over 30 min. After further 30 min, 2,6-di-*tert*-butyl-4-methylphenol (448 mg, 2.03 mmol) was added as the radical scavenger. The resultant mixture was directly subjected to flash chromatography [a column consecutively packed with silica gel 15 g and 10% (w/w) KF contained silica gel 15 g, hexane to hexane/ EtOAc 10/1 to EtOAc] to afford the impure **21**. The material was further purified by flash column chromatography on silica gel (50 g, hexane to

hexane/EtOAc 10/1 to 4/1 to 2/1 to 1/1 to 1/2) to afford a mixture of three diastereomers of **21**, which was used in the next reaction without further purification.

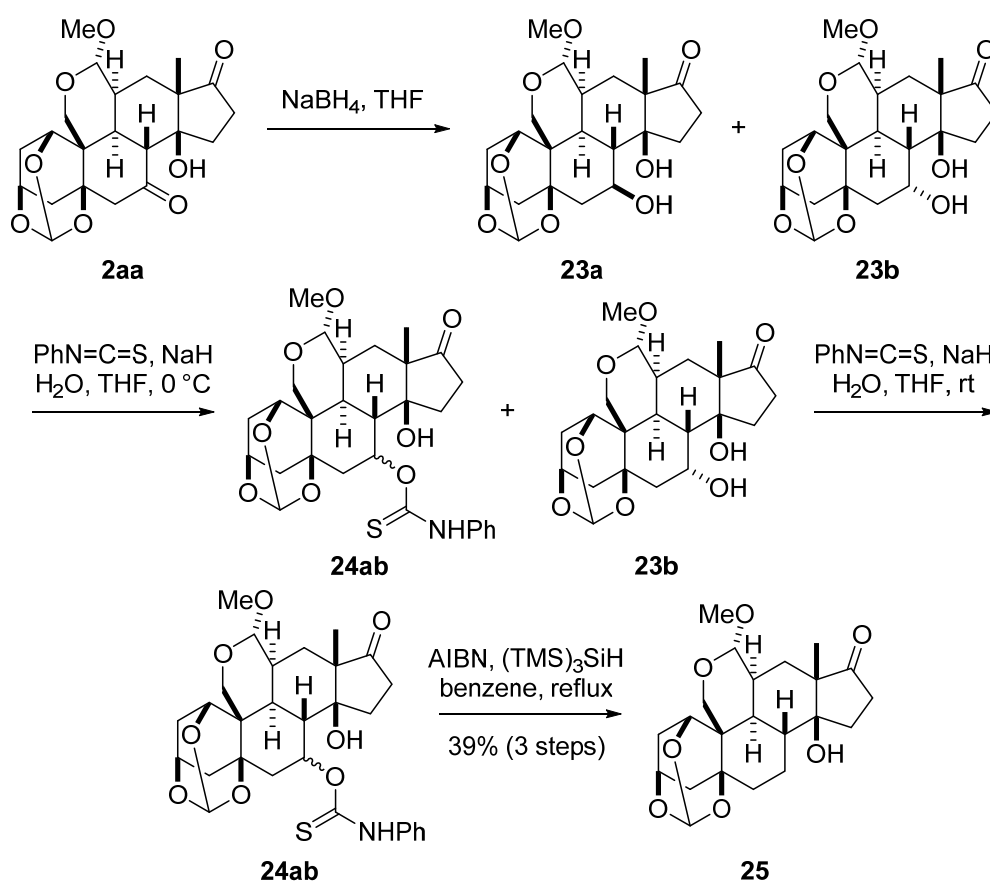
A 0.5 M NaOH aqueous solution (4 mL, 2.0 mmol) was added to a solution of the above crude **21** in MeOH (13 mL) at room temperature. The reaction mixture was stirred at room temperature for 2 h, and then AcOH (6 mL) was added. The pH value of the resultant solution was adjusted to 8-9 by addition of saturated aqueous NaHCO₃. After being diluted with EtOH, the mixture was concentrated. The residue was purified by flash column chromatography on silica gel (15 g, EtOAc) to afford the impure **22**. The material was further purified by flash column chromatography on silica gel (50 g, hexane to hexane/acetone 10/1 to 5/1 to 4/1 to 3/1 to 1/1) to afford a mixture of two diastereomers of **22**, which was used in the next reaction without further purification.

Dess-Martin periodinane (752 mg, 1.77 mmol) was added to a solution of the above crude **22** and NaHCO₃ (297 mg, 3.54 mmol) in CH₂Cl₂ (30 mL) at room temperature. The reaction mixture was stirred at room temperature for 2 h, and then a 1 : 1 mixture of saturated aqueous Na₂S₂O₃ and NaHCO₃ (30 mL) was added. The resultant mixture was extracted with CH₂Cl₂ (30 mL x3), and the combined organic layers were washed with brine (30 mL) and concentrated. The residue was purified by flash column chromatography on silica gel (15 g, hexane to hexane/EtOAc 10/1 to 4/1 to 3/1 to 2/1) to afford triketone **3a** (93 mg, 0.22 mmol) in 26% yield over 4 steps: white crystal: m.p. 196-200 °C; $[\alpha]_D^{24}$ -26 (*c* 1.4, CHCl₃); IR (neat) ν 2966, 2933, 1762, 1721, 1449, 1293, 1253, 1147, 1078, 993, 916 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) 1.09 (3H, s, H18), 1.36 (1H, dd, *J* = 14.6, 2.8 Hz, H12_a), 1.64-1.76 (2H, m, H4_a and 11), 1.85 (1H, dd, *J* = 14.2, 1.8 Hz, H2_a), 1.97 (1H, ddd, *J* = 8.7, 8.7, 3.7 Hz, H9), 2.03 (1H, dd, *J* = 14.2, 11.9 Hz, H12_b), 2.18 (1H, ddd, *J* = 13.7, 3.6, 2.3 Hz, H4_b), 2.26 (1H, d, *J* = 15.1 Hz, H6_a), 2.50-2.82 (7H, m, H2_b, 8, 15 and 16), 2.71 (1H, d, *J* = 15.1 Hz, H6_b), 3.15 (3H, s, OCH₃), 4.10 (1H, d, *J* = 12.8 Hz, H19_a), 4.177 (1H, d, *J* = 12.8 Hz, H19_b), 4.181 (1H, d, *J* = 8.7 Hz, CHOCH₃), 4.24-4.29 (1H, m, H3), 4.40-4.45 (1H, m, H1), 5.61 (1H, s, CH); ¹³C NMR (100 MHz, CDCl₃) δ 25.1, 28.4, 33.0, 33.6, 34.3, 34.7, 35.4, 37.0, 39.0, 41.5, 49.1, 53.5, 55.6, 62.4, 65.9, 70.1, 73.3, 101.5, 105.6, 205.2, 213.7, 216.2; HRMS (ESI) calcd for C₂₂H₂₈O₈Na [M+Na]⁺ 443.1676, found 443.1683.



Steroids 2aa and 2ab. A solution of $\text{KN}(\text{TMS})_2$ (0.67 M, 89 μL , 0.059 mmol) freshly prepared from KH and $\text{HN}(\text{TMS})_2$ in THF^{S5} was added to a solution of triketone **3a** (83 mg, 0.20 mmol) in refluxing THF (4 mL), and the reaction mixture was stirred for 1 h. After the mixture was cooled to room temperature, saturated aqueous NH_4Cl (4 mL) was added. The resultant mixture was extracted with a 2 : 1 mixture of CHCl_3 and EtOH (4 mL x5), and the combined organic layers were washed with brine (2 mL), dried over Na_2SO_4 and concentrated. The residue was purified by flash column chromatography on silica gel (15 g, hexane to hexane/THF 1/1) to afford **2aa** (54 mg, 0.13 mmol) and **2ab** (7.0 mg, 0.017 mmol) in 65% and 8% yields, respectively. **2aa**: crystal; m.p. 268-270 $^\circ\text{C}$ (dec.); $[\alpha]_{\text{D}}^{25}$ -0.48 (*c* 1.1, CHCl_3); IR (neat) ν 3525, 2949, 1736, 1704, 1446, 1379, 1194, 1152, 1067, 993, 976 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 1.14 (3H, s, H18), 1.16 (1H, dd, $J = 15.6, 5.9 \text{ Hz}$, H12_a), 1.66 (1H, d, $J = 12.8 \text{ Hz}$, H4_a), 1.76 (1H, d, $J = 14.2 \text{ Hz}$, H2_a), 1.79 (1H, d, $J = 15.6 \text{ Hz}$, H12_b), 1.81-1.87 (1H, m, H11), 2.05-2.16 (2H, m, H9 and 15_a), 2.20 (1H, ddd, $J = 12.8, 3.7, 2.7 \text{ Hz}$, H4_b), 2.28 (1H, ddd, $J = 14.2, 9.6, 1.4 \text{ Hz}$, H15_b), 2.34 (1H, d, $J = 13.7 \text{ Hz}$, H6_a), 2.42 (1H, ddd, $J = 19.2, 9.6, 1.4 \text{ Hz}$, H16_a), 2.55 (1H, ddd, $J = 19.2, 10.1, 9.6 \text{ Hz}$, H16_b), 2.70 (1H, dddd, $J = 14.2, 3.6, 2.8, 2.7 \text{ Hz}$, H2_b), 2.80 (1H, d, $J = 13.7 \text{ Hz}$, H6_b), 3.19 (1H, d, $J = 12.4 \text{ Hz}$, H8), 3.52 (3H, s, OCH_3), 3.54 (1H, s, OH), 4.13 (1H, d, $J = 12.4 \text{ Hz}$, H19_a), 4.29 (1H, m, H3), 4.75 (1H, m, H1), 4.81 (1H, dd, $J = 12.4, 1.8 \text{ Hz}$, H19), 4.83 (1H, d, $J = 9.1 \text{ Hz}$, CHOMe), 5.69 (1H, s, CH); ^{13}C NMR (100 MHz, CDCl_3) δ 16.7, 27.56, 27.61, 31.1, 32.8, 35.2, 35.3, 39.7, 40.7, 50.8, 51.0, 51.9, 56.8, 63.2, 65.8, 68.7, 73.5, 79.7, 101.9, 105.8, 207.8, 218.4; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{28}\text{O}_8\text{Na}$ $[\text{M}+\text{Na}]^+$ 443.1676, found 443.1681. **2ab**: crystal; m.p. 261-264 $^\circ\text{C}$; $[\alpha]_{\text{D}}^{25}$ -24 (*c* 0.83, CHCl_3); IR (neat) ν 3511, 2965, 2932, 1737, 1702, 1446, 1194, 1149, 1074, 1056, 994, 971 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 1.00 (3H, s, H18), 1.31 (1H, dd, $J = 15.1, 6.4 \text{ Hz}$, H12_a), 1.62-1.79 (3H, m, H2_a, 4_a and 11), 2.13 (1H, ddd, $J = 13.7, 3.6, 2.8 \text{ Hz}$, H4_b), 2.17-2.23 (1H, m, H15_a), 2.20 (1H,

dd, $J = 15.2, 10.5$ Hz, H16_a), 2.28-2.37 (3H, m, 6_a, 9 and 15_b), 2.34 (1H, dd, $J = 15.1, 1.8$ Hz, H12_b), 2.54-2.68 (1H, m, H2_b), 2.59 (1H, ddd, $J = 15.6, 8.2, 8.2$ Hz, H16_b), 2.79 (1H, d, $J = 12.4$ Hz, H8), 2.86 (1H, d, $J = 12.4$ Hz, H6_b), 3.51 (3H, s, OCH₃), 3.73 (1H, s, OH), 4.02 (1H, d, $J = 12.8$ Hz, H19_a), 4.18 (1H, d, $J = 8.7$ Hz, CHCOCH₃), 4.24-4.28 (1H, m, H3), 4.69 (1H, dd, $J = 12.8, 1.4$ Hz, H19_b), 4.69-4.73 (1H, m, H1), 5.66 (1H, s, CH); ¹³C NMR (100 MHz, CDCl₃) δ 20.3, 27.6, 28.2, 30.9, 34.4, 34.5, 34.7, 36.7, 41.1, 51.3, 51.5, 52.7, 56.8, 62.8, 66.1, 69.1, 74.3, 77.1, 101.0, 105.9, 208.6, 218.2; HRMS (ESI) calcd for C₂₂H₂₈O₈Na [M+Na] 443.1676, found 443.1679.

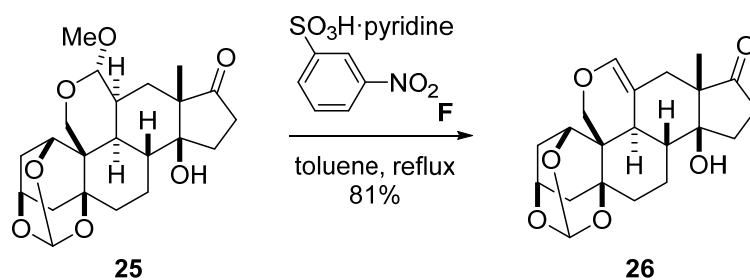


Compound 25. NaBH₄ (12 mg, 0.32 mmol) was added to a solution of **2aa** (88 mg, 0.21 mmol) in THF (20 mL) at room temperature. The reaction mixture was stirred at room temperature for 1 h, and then a 1 : 1 mixture of 1M HCl and CH₂Cl₂ (30 mL) was added. The resultant mixture was extracted with CH₂Cl₂ (15 mL x4), and the combined organic layers were washed with brine (15 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (25 g, hexane to hexane/acetone 1/2) to afford a 2.5 : 1 diastereomeric mixture of **23ab**, which was used in the next reaction without further purification.

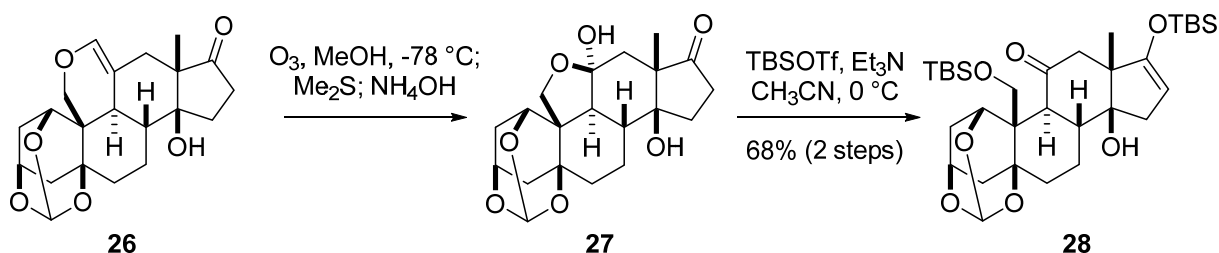
NaH (60% dispersion in mineral oil) was washed with hexane and dried in vacuo. The purified NaH (63 mg, 2.6 mmol) and PhNCS (630 μ L, 5.3 mmol) were successively added

to the above crude **23ab** in THF (13 mL) at 0 °C. The reaction mixture was stirred at 0 °C for 2 h, and then H₂O (1 µL, 0.056 mmol) was added. The mixture was stirred at 0 °C for further 2 h, and then saturated aqueous NH₄Cl (10 mL) was added. The resultant mixture was extracted with CH₂Cl₂ (15 mL x4), and the combined organic layers were washed with brine (15 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (25 g, hexane to hexane/EtOAc 1/2 to hexane/acetone 1/1) to afford a mixture of thiocarbamate **24ab** and alcohol **23b**, which was again subjected to the thiocarbamate formation. NaH (26 mg, 1.1 mmol) and PhNCS (260 µL, 2.2 mmol) were successively added to a solution of the mixture and H₂O (1 µL, 0.06 mmol) in THF (5 mL) at room temperature. The reaction mixture was stirred at room temperature for 1 h, and then saturated aqueous NH₄Cl (6 mL) was added. The resultant mixture was extracted with CH₂Cl₂ (5 mL x4), and the combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (25 g, hexane to hexane/EtOAc 1/1) to afford the impure **24ab**, which were used in the next reaction.

A solution of the above crude **24ab**, AIBN (11 mg, 0.067 mmol) and (TMS)₃SiH (410 µL, 1.3 mmol) in benzene (13 mL) was degassed by freeze-thaw procedure (x3). The reaction mixture was heated to reflux, and stirred for 1 h. After being cooled to room temperature, the mixture was directly subjected to flash column chromatography on silica gel (20 g, hexane to hexane/EtOAc 1/2) to afford **25** (33 mg, 0.081 mmol) in 39% over 3 steps: white solid: m.p. 260-266 °C; [α]_D²² +14 (*c* 0.60, CHCl₃); IR (neat) ν 3487, 2948, 1731, 1451, 1149, 1069, 1053 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.15 (3H, s, H18), 1.14-1.20 (1H, m, H12_a), 1.38-1.54 (2H, m), 1.67-1.90 (5H, m), 1.80 (1H, dd, *J* = 15.5, 1.7 Hz, H12_b), 2.02-2.08 (2H, m, H4_a and 7_a), 2.14 (1H, ddd, *J* = 13.7, 10.3, 10.3 Hz), 2.26 (1H, d, *J* = 13.8 Hz, H4_b), 2.35 (1H, ddd, *J* = 12.6, 12.6, 4.6 Hz, H8), 2.42-2.48 (2H, m), 2.61-2.68 (1H, m, H2_a), 3.49 (3H, s, OCH₃), 4.04 (1H, d, *J* = 12.0 Hz, H19_a), 4.29-4.33 (1H, m, H3), 4.64 (1H, dd, *J* = 12.0, 1.8 Hz, H19_b), 4.67-4.69 (1H, m, H1), 4.76 (1H, d, *J* = 9.8 Hz, OCHOCH₃), 5.67 (1H, s, CH); ¹³C NMR (125 MHz, CDCl₃) δ 16.9, 19.3, 26.4, 28.0, 31.8, 32.0, 33.0, 34.4, 35.3, 37.4, 39.0, 40.9, 52.4, 56.6, 63.4, 66.4, 68.8, 73.6, 82.1, 102.0, 105.8, 219.7; HRMS (ESI) calcd for C₂₂H₃₀O₇Na [M+Na]⁺ 429.1884, found 429.1892.



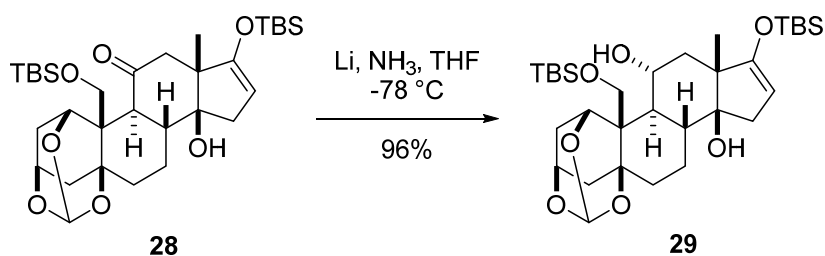
Enol ether 26. Reagent **F** (46 mg, 0.16 mmol) was added to a solution of **25** (33 mg, 0.081 mmol) in toluene (8 mL) at room temperature. The reaction mixture was heated to reflux, and stirred for 30 min. After the mixture was cooled to room temperature, saturated aqueous NaHCO_3 (8 mL) was added. The resultant mixture was extracted by CH_2Cl_2 (8 mL x4), and the combined organic layers were washed with brine (4 mL), dried over Na_2SO_4 , filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (10 g, hexane to hexane/EtOAc 1/2) to afford enol ether **26** (25 mg, 0.067 mmol) in 81% yield: white crystal: m.p. $>300^\circ\text{C}$; $[\alpha]_{\text{D}}^{23} -79$ (c 1.00, CHCl_3); IR (neat) ν 3486, 3058, 2950, 1731, 1670, 1448, 1374, 1308, 1269, 1222, 1149 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 1.02 (3H, s, H18), 1.40-1.56 (2H, m), 1.64-1.76 (3H, m), 1.69 (1H, d, $J = 13.8$ Hz, H12_a), 1.88 (1H, d, $J = 13.8$ Hz, H12_b), 1.88-2.02 (3H, m), 2.13 (1H, ddd, $J = 13.7, 3.6, 2.7$ Hz, H4_a), 2.15-2.26 (1H, m), 2.25 (1H, d, $J = 13.7$ Hz, H4_b), 2.47-2.52 (2H, m), 2.66 (1H, dddd, $J = 14.2, 3.2, 3.2, 3.2$ Hz, H2_a), 4.14 (1H, d, $J = 10.5$ Hz, H19_a), 4.21 (1H, m, H1), 4.33 (1H, m, H3), 4.75 (1H, dd, $J = 10.5, 1.8$ Hz, H19_b), 5.70 (1H, s, CH), 6.16 (1H, s, C=CH); ^{13}C NMR (100 MHz, CDCl_3) δ 13.5, 19.0, 26.8, 28.4, 32.1, 33.6, 35.0, 36.4, 37.2, 39.0, 47.5, 55.3, 63.9, 66.4, 68.5, 73.2, 81.8, 106.0, 109.5, 138.1, 219.9; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{26}\text{O}_6\text{Na}$ $[\text{M}+\text{Na}]^+$ 397.1622, found 397.1624.



TBS-enol ether 28. Ozone was bubbled into a solution of enol ether **26** (25 mg, 0.067 mmol) in MeOH (30 mL) at -78°C for 10 sec. The reaction mixture was stirred at -78°C for 5 min, and then oxygen was bubbled into the reaction mixture for 10 min. After Me_2S (3 mL) was added, the resultant solution was warmed to room temperature, and stirred for 30 min. Then, 28% aqueous NH_3 (3 mL) was added. The mixture was stirred at room temperature for 1 h, and then concentrated. The residue was purified by flash column

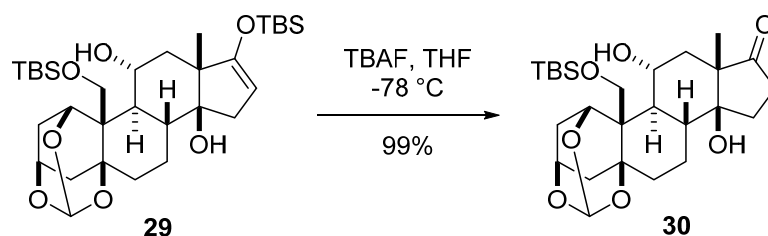
chromatography on silica gel (10 g, hexane to hexane/EtOAc 1/2) to afford the impure **27**, which was used in the next reaction without further purification.

TBSOTf (75 μ L, 0.33 mmol) was added to a solution of the above crude **27** and Et₃N (91 μ L, 0.65 mmol) in CH₃CN (6.5 mL) at -35 °C. The reaction mixture was stirred at -35 °C for 10 min and at 0 °C for 20 min, and then saturated aqueous NaHCO₃ (6 mL) was added. The resultant solution was extracted with EtOAc (6 mL x3), and the combined organic layers were washed with brine (6 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by column chromatography on silica gel (10 g, hexane to hexane/EtOAc 3/1) to afford TBS-enol ether **28** (27 mg, 0.044 mmol) in 68% yield over 2 steps: white solid; m.p. 175-180 °C; [α]_D²⁵ -53 (*c* 0.63, CHCl₃); IR (neat) ν 3491, 2954, 2933, 2890, 2858, 1712, 1658, 1467, 1324, 1254, 1165, 1137, 1087, 997, 841 cm⁻¹; ¹H NMR (500 MHz, C₆D₆) δ 0.12 (3H, s, CH₃ of TBS), 0.135 (3H, s, CH₃ of TBS), 0.139 (3H, s, CH₃ of TBS), 0.20 (3H, s, CH₃ of TBS), 0.95 (9H, s, *t*-Bu of TBS), 0.99 (9H, s, *t*-Bu of TBS), 1.12-1.22 (1H, m), 1.16 (3H, s, H18), 1.25-1.31 (1H, m), 1.68-1.92 (6H, m), 2.03 (1H, dd, *J* = 17.2, 2.9 Hz, H15_a), 2.44 (1H, ddd, *J* = 13.2, 13.2, 4.6 Hz), 2.50 (1H, dd, *J* = 17.2, 2.3 Hz, H15_b), 2.57 (1H, d, *J* = 13.2 Hz, H12_a), 2.59-2.65 (1H, m, H2_a), 2.91 (1H, d, *J* = 13.2 Hz, H12_b), 3.91 (1H, m, H3), 4.22 (1H, d, *J* = 10.3 Hz, H19_a), 4.38 (1H, dd, *J* = 2.3, 2.3 Hz, H16), 4.78 (1H, d, *J* = 10.3 Hz, H19_b), 4.82-4.86 (1H, m, H1), 5.87 (1H, s, CH); ¹³C NMR (125 MHz, C₆D₆) δ -5.5, -5.4, -5.0, -4.5, 18.3, 18.7, 19.9, 20.8, 25.8, 26.1, 28.9, 32.2, 35.3, 38.7, 41.3, 47.8, 48.0, 48.5, 53.0, 61.5, 67.1, 69.0, 74.4, 79.7, 96.0, 106.4, 156.0, 208.1; HRMS (ESI) calcd for C₃₂H₅₄O₇Si₂Na [M+Na]⁺ 629.3300, found 629.3298.

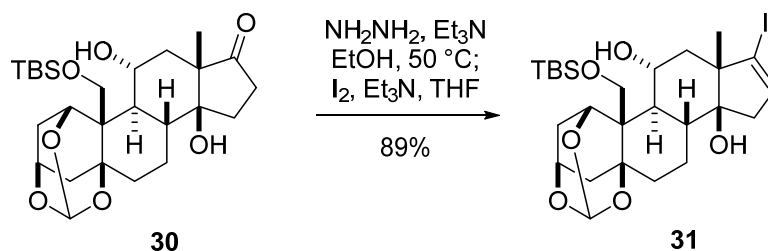


Diol 29. A Schlenk tube equipped with a dry-ice condenser was charged with liquid NH₃ (ca. 10 mL), and then Li metal (51 mg, 7.3 mmol) was added. The resultant blue solution was stirred -78 °C for 1 h, and a solution of TBS-enol ether **28** (27 mg, 0.044 mmol) in THF (3 mL) was added. The reaction mixture was stirred at -78 °C for 30 min, and then saturated aqueous NH₄Cl (7 mL) was added. The resultant mixture was extracted with EtOAc (10 mL x3), and the combined layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (5 g, hexane to hexane/EtOAc 3/1) to afford diol **29** (26 mg, 0.043 mmol) in 96% yield: amorphous; [α]_D²⁷ -26 (*c* 0.87, CHCl₃); IR (neat) ν 3495, 2951,

2933, 2890, 2857, 1637, 1467, 1358, 1303, 1255, 1157, 1004, 839 cm^{-1} ; ^1H NMR (500 MHz, C_6D_6) δ 0.128 (3H, s, CH_3 of TBS), 0.133 (6H, s, CH_3 of TBS x2), 0.16 (3H, s, CH_3 of TBS), 0.951 (9H, s, *t*-Bu of TBS), 0.955 (9H, s, *t*-Bu of TBS), 1.14 (1H, dddd, $J = 13.2, 13.2, 13.2, 4.6$ Hz, H7_a), 1.22 (3H, s, H18), 1.25-1.30 (1H, m), 1.38-1.55 (4H, m), 1.77 (1H, dd, $J = 13.7, 1.2$ Hz, H2_a), 1.82-1.99 (4H, m), 1.95 (1H, dd, $J = 15.5, 2.9$ Hz, H15_a), 2.23 (1H, dd, $J = 13.2, 3.4$ Hz, H12_a), 2.54-2.62 (1H, m, H2_b), 2.82 (1H, d, $J = 4.6$ Hz, OH), 4.02-4.10 (2H, m, H3 and 11), 4.16 (1H, d, $J = 9.7$ Hz, H19_a), 4.37 (1H, dd, $J = 2.9, 1.8$ Hz, H16), 4.73 (1H, d, $J = 9.7$ Hz, H19_b), 5.15 (1H, d, $J = 4.6$ Hz, H1), 5.90 (1H, s, CH); ^{13}C NMR (125 MHz, C_6D_6) δ -5.7, -5.4, -5.0, -4.5, 15.8, 18.3, 18.5, 20.1, 25.8, 26.1, 28.9, 32.2, 35.1, 36.8, 39.8, 45.6, 47.0, 47.1, 51.1, 61.4, 67.0, 69.4, 70.7, 74.3, 82.0, 93.6, 106.1, 161.6; HRMS (ESI) calcd for $\text{C}_{32}\text{H}_{56}\text{O}_7\text{Si}_2\text{Na}$ $[\text{M}+\text{Na}]^+$ 631.3457, found 631.3457.

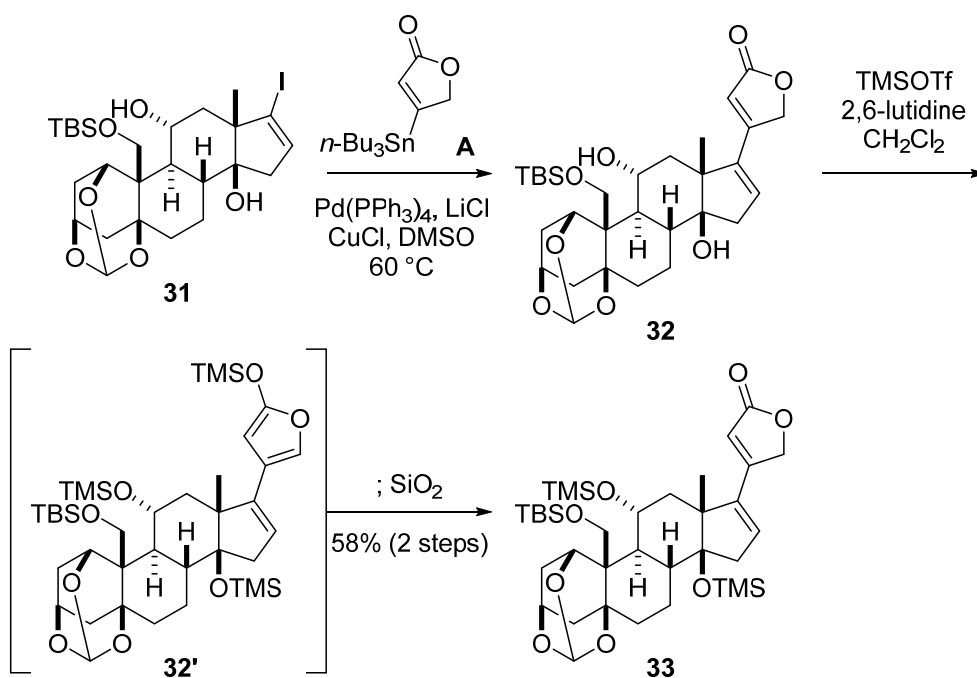


Ketone 30. TBAF (1.0 M in THF, 84 μL , 0.084 mmol) was added to a solution of diol **29** (34 mg, 0.056 mmol) in THF (6 mL) at -78 $^\circ\text{C}$. The reaction mixture was stirred at -78 $^\circ\text{C}$ for 10 min, and then saturated aqueous NH_4Cl (5 mL) was added. The resultant solution was extracted with CH_2Cl_2 (5 mL x3), and the combined organic layers were washed with brine (4 mL), dried over Na_2SO_4 , filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (5 g, hexane to hexane/EtOAc 1/2) to afford ketone **30** (27 mg, 0.055 mmol) in 99% yield: crystal: m.p. 184-188 $^\circ\text{C}$; $[\alpha]_{\text{D}}^{24}$ -3.9 (*c* 1.4, CHCl_3); IR (neat) ν 3482, 2951, 2932, 2883, 2856, 1732, 1469, 1379, 1256, 1154, 1076, 1005, 976, 836 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 0.13 (3H, s, CH_3 of TBS), 0.15 (3H, s, CH_3 of TBS), 0.92 (9H, s, *t*-Bu of TBS), 1.12 (3H, s, H18), 1.34 (1H, dd, $J = 13.2, 12.6$ Hz, H12_a), 1.35-1.52 (3H, m), 1.54 (1H, dd, $J = 13.2, 3.5$ Hz, H12_b), 1.72 (1H, dd, $J = 11.5, 10.9$ Hz, H9), 1.87-1.97 (3H, m), 2.00-2.06 (2H, m, H2_a and 4_a), 2.19 (1H, ddd, $J = 13.8, 10.3, 9.8$ Hz), 2.26 (1H, d, $J = 13.7$ Hz, H4_b), 2.37-2.46 (1H, m), 2.48 (1H, ddd, $J = 19.5, 10.3, 2.3$ Hz), 2.58-2.65 (1H, m, H2_b), 2.69 (1H, brs, OH), 4.04-4.12 (1H, m, H11), 4.10 (1H, d, $J = 10.3$ Hz, H19_a), 4.30-4.34 (1H, m, H3), 4.53 (1H, d, $J = 10.3$ Hz, H19_b), 4.83 (1H, d, $J = 4.0$ Hz, H1), 5.64 (1H, s, CH); ^{13}C NMR (125 MHz, CDCl_3) δ -5.9, -5.5, 14.1, 18.3, 19.0, 25.9, 27.3, 28.5, 31.6, 32.8, 34.8, 40.3, 40.5, 45.1, 46.3, 53.9, 60.7, 66.6, 69.1, 70.1, 73.9, 81.7, 105.4, 218.9; HRMS (ESI) calcd for $\text{C}_{26}\text{H}_{42}\text{O}_7\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 517.2592, found 517.2593.



Vinyl iodide 31. $\text{NH}_2\text{NH}_2 \cdot \text{H}_2\text{O}$ (52 μL , 1.1 mmol) and Et_3N (150 μL , 1.1 mmol) were added to a solution of ketone **30** (27 mg, 0.054 mmol) in EtOH (5 mL) at room temperature. The reaction mixture was heated to 50 $^\circ\text{C}$, and stirred for 7 h. After being cooled to room temperature, the mixture was concentrated.

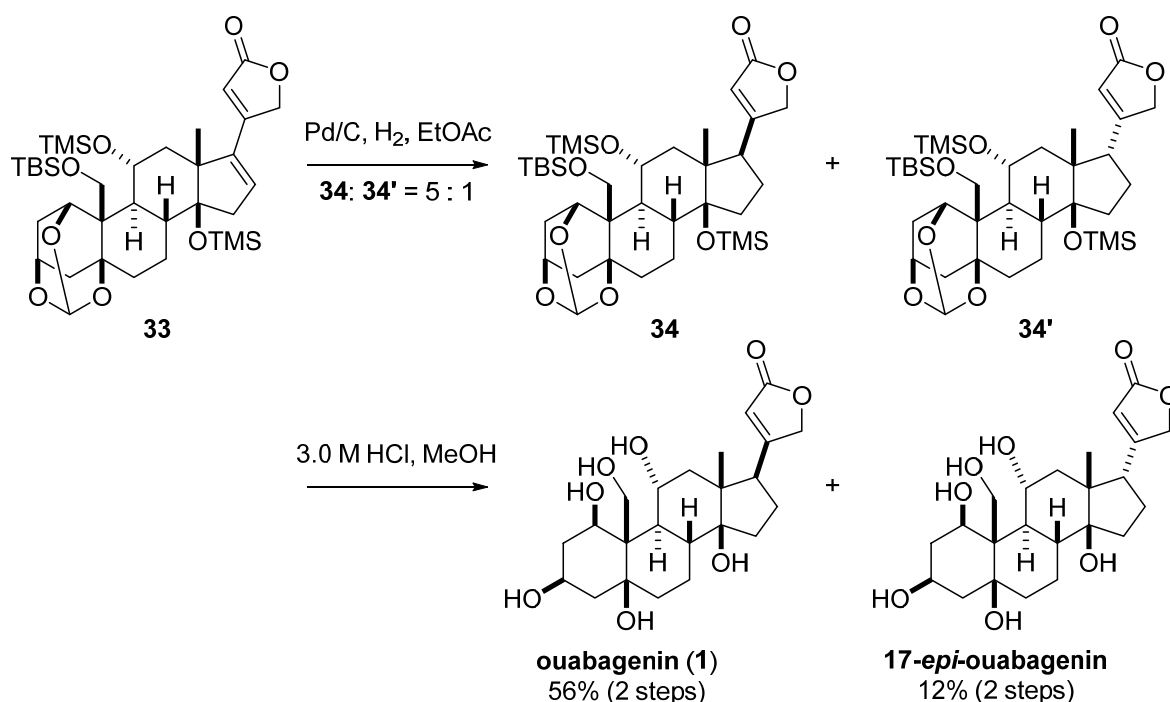
A solution of I_2 (41 mg, 0.16 mmol) in THF (1 mL) was added to a solution of the residue and Et_3N (150 μL , 1.1 mmol) in THF (5 mL). The mixture was stirred at room temperature for 30 min, and then a 1 : 1 mixture of saturated aqueous $\text{Na}_2\text{S}_2\text{O}_3$ and NaHCO_3 (4 mL) was added. The resultant mixture was extracted with CH_2Cl_2 (5 mL x4), and the combined organic layers were washed with brine (5 mL), dried over Na_2SO_4 , filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (5 g, hexane to hexane/EtOAc 2/1) to afford vinyl iodine **31** (29 mg, 0.048 mmol) in 89% yield: white solid: m.p. 233-237 $^\circ\text{C}$; $[\alpha]_{\text{D}}^{23}$ -13 (c 1.5, CHCl_3); IR (neat) ν 3467, 2953, 2930, 2885, 2856, 1461, 1365, 1254, 1146, 1126, 1014, 978 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 0.14 (3H, s, CH_3 of TBS), 0.16 (3H, s, CH_3 of TBS), 0.93 (9H, s, $t\text{-Bu}$ of TBS), 1.09 (1H, dd, J = 13.2, 12.6 Hz, H12_a), 1.13 (3H, s, H18), 1.25-1.37 (2H, m), 1.47-1.61 (2H, m), 1.73 (1H, br s, OH), 1.90-2.07 (5H, m), 2.24 (1H, d, J = 13.8 Hz), 2.31 (1H, dd, J = 16.6, 2.9 Hz, H15_a), 2.58 (1H, dd, J = 16.6, 1.7 Hz, H15_b), 2.59-2.65 (1H, m, H2), 3.04 (1H, d, J = 4.0 Hz, OH), 3.86-3.94 (1H, m, H11), 4.11 (1H, d, J = 10.3 Hz, H19_a), 4.29-4.33 (1H, m, H3), 4.53 (1H, d, J = 10.3 Hz, H19_b), 4.83-4.87 (1H, d, J = 4.0 Hz, H1), 5.64 (1H, s, CH), 6.14 (1H, dd, J = 2.9, 1.7 Hz, H16); ^{13}C NMR (125 MHz, CDCl_3) δ -5.9, -5.5, 18.3, 19.3, 20.4, 25.9, 28.5, 31.6, 34.8, 39.8, 42.9, 45.0, 46.1, 46.6, 55.4, 60.7, 66.6, 68.7, 70.1, 74.0, 81.7, 105.4, 109.9, 133.4; HRMS (ESI) calcd for $\text{C}_{26}\text{H}_{41}\text{IO}_6\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 627.1609, found 627.1613.



Butenolide 33. A Schlenk tube was charged with $\text{Pd(PPh}_3)_4$ (28 mg, 0.024 mmol), LiCl (18 mg, 0.42 mmol) and CuCl (35 mg, 0.35 mmol) in a glove box filled with Ar. A solution of vinyl iodide **31** (29 mg, 0.048 mmol) in DMSO (3 mL) and a solution of butenolide **A** (54 mg, 0.14 mmol) in DMSO (1.8 mL) were successively added, and then the mixture was degassed by the freeze-thaw procedure (x3). The reaction mixture was heated to 60 °C, and stirred for 1 h. After the mixture was cooled to room temperature, pH 7 phosphate buffer (5 mL) was added. The resultant mixture was extracted with CH_2Cl_2 (5 mL x5), and the combined organic layers were washed with brine (5 mL), and dried over Na_2SO_4 . The solution was filtered through a pad of 10% (w/w) KF contained silica gel (5 g), and the filtrate was concentrated. The residue was purified by flash column chromatography on silica gel (5 g, hexane to EtOAc) to afford the impure butenolide **32** (23 mg). A part of the crude material (21 mg) was used in the next reaction without further purification.

TMSOTf (36 μL , 0.20 mmol) was added to a solution of the above crude butenolide **32** and 2,6-lutidine (47 μL , 0.40 mmol) in CH_2Cl_2 at -78 °C. The reaction mixture was warmed to room temperature, stirred for 20 min, and then saturated aqueous NaHCO_3 (3 mL) was added. The resultant solution was extracted with CH_2Cl_2 (3 mL x3), and the combined organic layers were washed with brine (3 mL), dried over Na_2SO_4 , filtered, and concentrated to give siloxyfuran **32'**. Silica gel (10 g, Merck 40-63 μm Silica-gel 60) was added to a solution of the **32'** in CH_2Cl_2 (20 mL). After CH_2Cl_2 was removed in vacuo, the resultant solid was stood at room temperature for 10 h. Then, Et_2O was added. The suspension was passed through a pad of cotton, and the filtrate was concentrated. The residue was purified by flash column chromatography on silica gel (3 g, hexane to

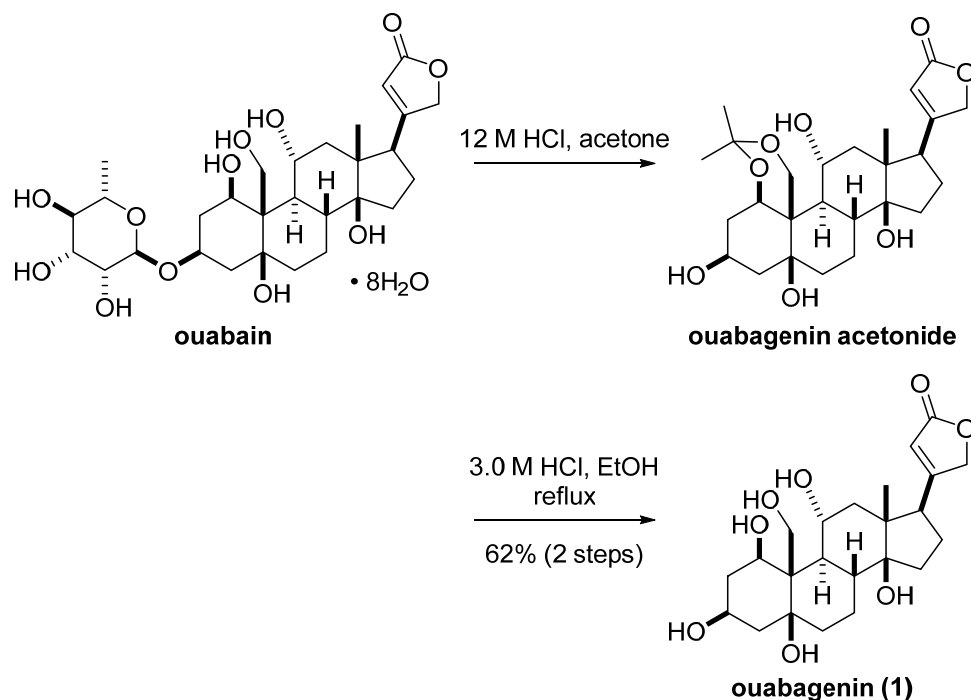
hexane/Et₂O 1/1) to afford the impure **33**. The material was further purified by flash column chromatography on silica gel (1.5g, CH₂Cl₂ to CH₂Cl₂/EtO₂ 4/1) to afford **33** (18 mg, 0.026 mmol). The yield of **33** was calculated to be 58% over 2 steps: white solid: m.p. 186-189 °C; [α]_D²⁶ +23 (*c* 0.90, CHCl₃); IR (neat) ν 2953, 2930, 2856, 1784, 1752, 1623, 1252, 1156, 1081, 985, 839 cm⁻¹; ¹H NMR (500 MHz, C₆D₆) δ -0.06 (9H, s, CH₃ of TMS), 0.08 (9H, s, CH₃ of TMS), 0.18 (3H, s, CH₃ of TBS), 0.21 (3H, s, CH₃ of TBS), 0.95-1.09 (2H, m, H7_a and 12_a), 1.02 (9H, s, *t*-Bu of TBS), 1.10 (3H, s, H18), 1.37-1.42 (1H, m), 1.48 (1H, dd, *J* = 10.0, 9.5 Hz, H9), 1.85 (1H, d, *J* = 14.0 Hz, H2_a), 1.86-1.92 (1H, m), 1.98-2.10 (5H, m), 2.15 (1H, d, *J* = 18.0 Hz, H15_a), 2.51-2.59 (2H, m, H2_b and 8), 4.13 (1H, m, H3), 4.27 (1H, ddd, *J* = 11.0, 10.0, 4.0 Hz, H11), 4.33 (2H, s, H21), 4.45 (1H, d, *J* = 11.5 Hz, H19_a), 4.80 (1H, d, *J* = 11.5 Hz, H19_b), 4.89 (1H, d, *J* = 4.5 Hz, H1), 5.23 (1H, s, H16), 5.91 (1H, s, H22), 5.92 (1H, s, CH); ¹³C NMR (125 MHz, C₆D₆) δ -5.5, -4.9, 1.1, 2.7, 18.2, 18.7, 21.9, 26.4, 29.4, 33.0, 35.4, 38.7, 39.7, 45.1, 47.4, 48.6, 53.5, 63.9, 67.1, 68.6, 70.9, 72.0, 74.8, 89.0, 106.1, 113.0, 131.4, 144.4, 157.4, 173.1; HRMS (ESI) calcd for C₃₆H₆₀O₈Si₃Na [M+Na]⁺ 727.3488, found 727.3482.



Ouabagenin (1).^{S6} A mixture of **33** (18 mg, 0.026 mmol) and Pd/C (5 wt%, 18 mg, 8.5 μ mol) in EtOAc (3 mL) was stirred under H₂ atmosphere at room temperature for 1 h, and then was filtered through a pad of Celite with Et₂O. The filtrate was concentrated, and the residue was purified by flash column chromatography on silica gel (8 g, hexane to hexane/EtO₂ 2/1 to EtOAc) to afford a 5:1 diastereomeric mixture of **34** and **34'**. The

mixture was dissolved in MeOH (1.00 mL), and 9/10 volume of the solution (0.90 mL) was used in the next reaction.

3.0 M HCl (1.0 mL) and MeOH (3.0 mL) were added to the above solution of the mixture of **34** and **34'** in MeOH (0.9 mL). The reaction mixture was stirred at room temperature for 15 h, and then saturated aqueous NaHCO₃ (2 mL) was added. The pH value of the resultant solution was adjusted to 6 by addition of 1.0 M HCl. The solution was extracted with a 2 : 1 mixture of CHCl₃ and EtOH (4 mL x6), and the combined organic layers were dried over Na₂SO₄, and filtered through a pad of silica gel (0.5 g) with MeOH. The filtrate was concentrated, and the residue was purified by flash column chromatography on silica gel (1 g, CH₂Cl₂/MeOH 3/1) to afford a mixture of ouabagenin (**1**) and 17-*epi*-ouabagenin. The mixture was further purified by HPLC (Inertsil ODS-3, 20 mm x 250 mm, MeOH/H₂O 20/80 to 25/75, 15 mL/min) to afford 17-*epi*-ouabagenin (*t_R* = 24 min, 1.2 mg, 2.8 μmol) and ouabagenin (**1**, *t_R* = 34 min, 5.5 mg, 0.013 mmol). The yields of ouabagenin (**1**) and 17-*epi*-ouabagenin were calculated to be 56% and 12% yields, respectively, over 2 steps. **Ouabagenin (1)**: [α]_D²⁶ -11 (*c* 0.28 DMSO/CHCl₃ 2/1); IR (neat) ν 3390, 2944, 1732, 1623, 1437, 1122, 1079, 1026 cm⁻¹; ¹H NMR (400 MHz, CD₃OD) δ 0.95 (3H, s, H18), 1.24-1.39 (1H, m), 1.43-1.64 (4H, m), 1.66-1.81 (3H, m), 1.82-2.24 (8H, m), 2.92 (1H, dd, *J* = 8.7, 5.5 Hz, H17), 3.99-4.11 (1H, br s), 4.18 (1H, d, *J* = 11.9 Hz, H19_a), 4.22-4.32 (1H, br s), 4.37 (1H, d, *J* = 11.9 Hz, H19_b), 4.91 (1H, dd, *J* = 18.8, 1.4 Hz, H21_a), 5.02 (1H, dd, *J* = 18.8, 1.4 Hz, H21_b), 5.91 (1H, dd, *J* = 1.4, 1.4 Hz, H22), one proton peak was not observed due to overlap with HDO peak or broadening of the peak; ¹³C NMR (125 MHz, (CD₃)₂SO/CDCl₃ 2/1) δ 16.9, 22.6, 26.0, 32.3, 47.1, 48.3, 49.0, 49.7, 60.5, 65.3, 66.1, 70.7, 72.9, 74.9, 83.4, 116.3, 173.6, 175.0, five ¹³C peaks were not observed due to overlap with the solvent peak and broadening of the peaks; HRMS (ESI) calcd for C₂₃H₃₄O₈Na [M+Na]⁺ 461.2146, found 461.2140. **17-*epi*-Ouabagenin**: ¹H NMR (400 MHz, CD₃OD) δ 1.10 (3H, s, H18), 1.24-1.38 (3H, m), 1.45-1.80 (5H, m), 1.81-2.21 (8H, m), 3.14 (1H, dd, *J* = 10.1, 9.2 Hz, H17), 3.95-4.27 (2H, m), 4.19 (1H, d, *J* = 11.9 Hz, H19_a), 4.37 (1H, d, *J* = 11.9 Hz, H19_b), 4.83 (1H, dd, *J* = 18.3, 1.0 Hz, H21_a), 4.97 (1H, dd, *J* = 18.3, 1.8 Hz, H21_b), 5.95-5.98 (1H, m, H22), one proton peak was not observed due to overlap with HDO peak or broadening of the peak; HRMS (ESI) calcd for C₂₃H₃₄O₈Na [M+Na]⁺ 461.2146, found 461.2147.

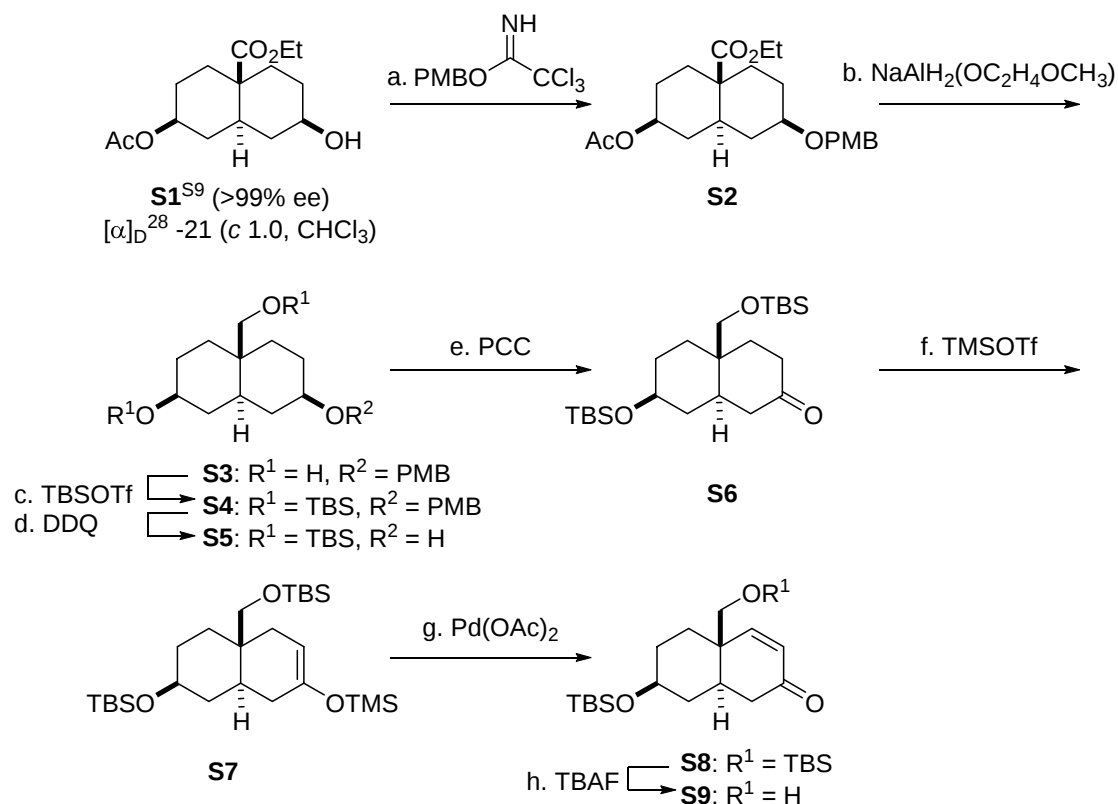


Synthesis of authentic ouabagenin (1).^{S6,7} 12 M HCl (0.3 mL) was added to a solution of ouabain octahydrate (537 mg, 0.737 mmol) in acetone (25 mL) at room temperature. The reaction mixture was stirred for 9 days at room temperature, and then was cooled to 0 °C for 1 h. After the solvent was removed by decantation, the resultant solid was washed with acetone (20 mL). The remaining acetone was azeotropically removed with EtOH (40 mL) to afford ouabagenin acetonide, which was used in the next reaction without further purification.

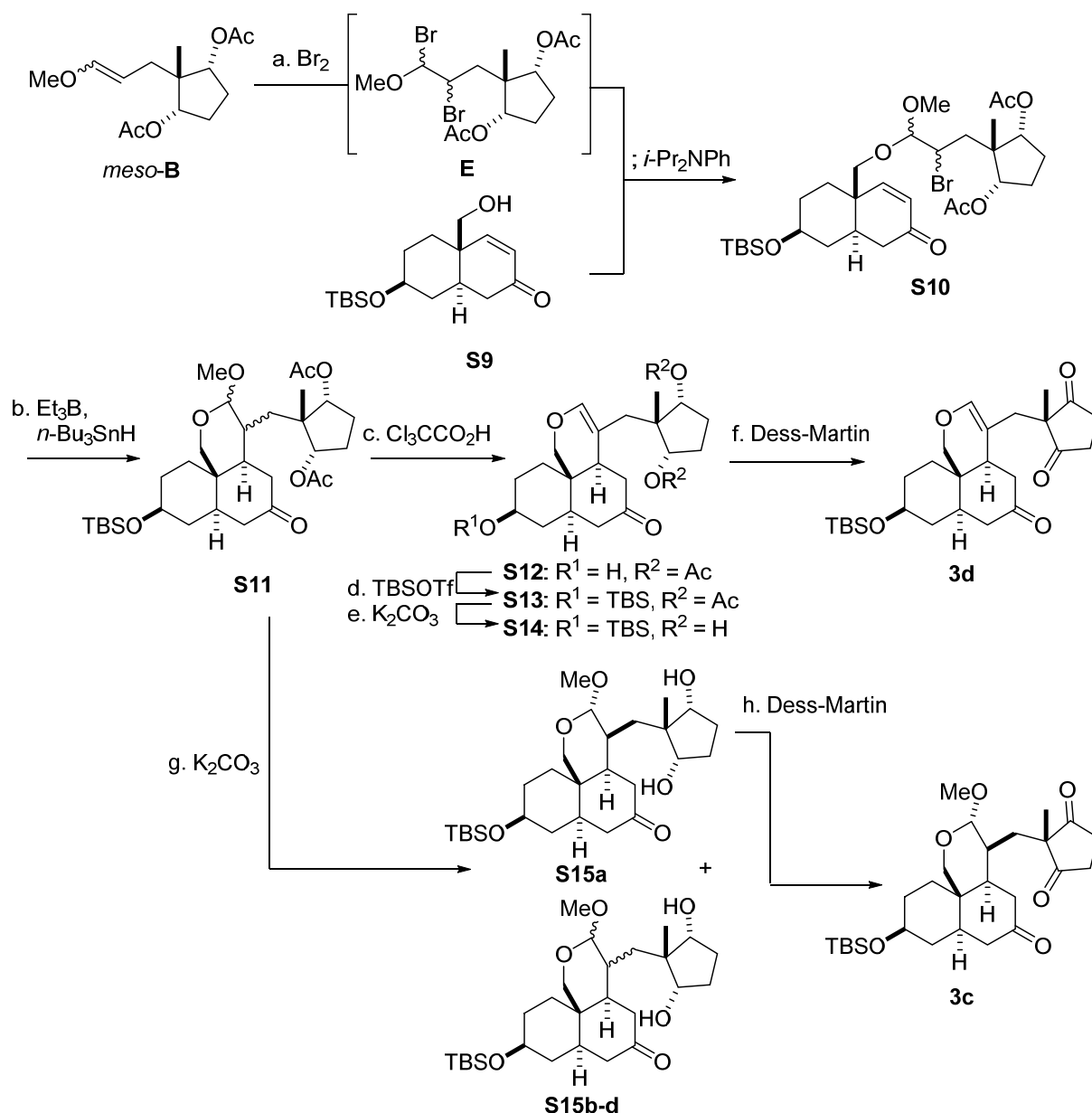
The solid of ouabagenin acetonide was dissolve in EtOH (40 mL). The mixture was heated to reflux, and stirred for 1 h. Then, 0.5 M HCl in MeOH (1 mL) was added. The reaction mixture was stirred at reflux temperature for 1 h, and then 3.0 M aqueous HCl (6 mL) was added. The mixture was stirred at reflux temperature for further 1 h. After the mixture was cooled to room temperature, solid NaHCO₃ was added. The pH value of the resultant solution was adjusted to 6 by addition of 1.0 M HCl and solid NaHCO₃. The solution was extracted with a 2 : 1 mixture of CHCl₃ and EtOH (25 mL x6), and the combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, and filtered through a pad of silica gel (0.5 g) with MeOH. The filtrate was concentrated, and the residue was purified by flash column chromatography on silica gel (15 g, CH₂Cl₂ to CH₂Cl₂/MeOH 10/1 to 3/1) to afford ouabagenin (**1**, 201 mg, 0.458 mmol) in 62% yield over 2 steps^{S8}: [α]_D²⁶ -6.3 (*c* 0.87 DMSO/CHCl₃ 2/1); IR (neat) ν 3398, 2946, 1734, 1625, 1440, 1122, 1025 cm⁻¹; ¹H NMR (400 MHz, CD₃OD) δ 0.95 (3H, s, H18), 1.23-1.40 (1H, m), 1.40-2.50 (15H, m), 2.92 (1H, dd, *J* = 8.7, 5.5 Hz, H17), 3.99-4.15 (1H, br s), 4.18 (1H, d, *J* = 11.9 Hz, H19_a), 4.25-4.40 (1H, br s), 4.37 (1H, d, *J* = 11.9 Hz, H19_b), 4.91 (1H, d, *J*

= 18.3 Hz, H21_a), 5.02 (1H, dd, J = 18.3 Hz, H21_b), 5.92 (1H, s, H22), one proton peak was not observed due to the overlap with HDO peak or broadening of the peak; ^{13}C NMR (125 MHz, $(\text{CD}_3)_2\text{SO}/\text{CDCl}_3$ 2/1) δ 16.9, 22.6, 26.1, 32.3, 34.5, 39.2 (deduced from the HMBC correlation), 47.1, 48.3, 49.0, 49.7, 60.5, 65.2, 66.1, 70.8, 73.0, 75.0, 83.4, 116.3, 173.6, 175.0, three ^{13}C peaks were not observed due to broadening of the peaks; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{34}\text{O}_8\text{Na}$ $[\text{M}+\text{Na}]^+$ 461.2146, found 461.2154.

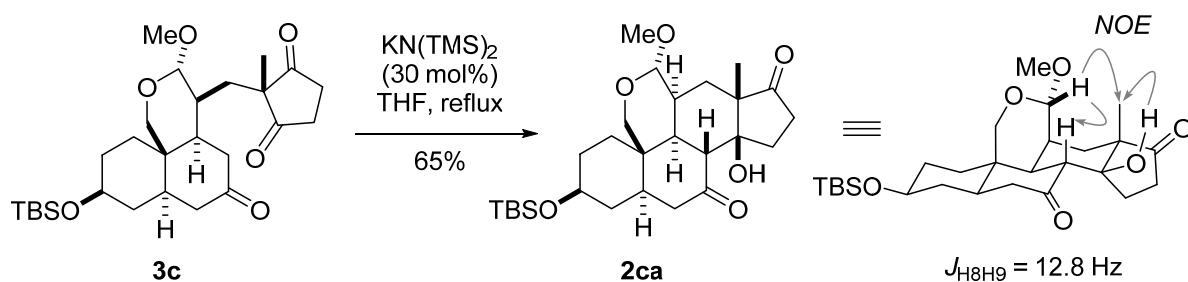
Model Study for the Stereoselective C-Ring Formation.



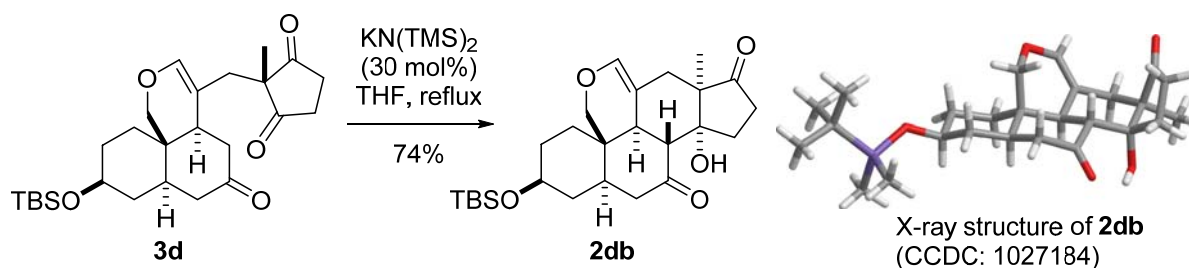
Scheme S1. Synthesis of the model AB-ring from **S1**.^{S9} Reagents and conditions: (a) (4-methoxyphenyl)methyl 2,2,2-trichloroethanecarboximidate, CSA, CH₂Cl₂; (b) NaAlH₂(OC₂H₄OCH₃), toluene, 100 °C; (c) TBSOTf, 2,6-lutidine, CH₂Cl₂, 52% (3 steps); (d) DDQ; CH₂Cl₂, pH 7 phosphate buffer; (e) PCC, CH₂Cl₂, 71% (2 steps); (f) TMSOTf, Et₃N, CH₂Cl₂, 0 °C; (g) Pd(OAc)₂, O₂, DMSO, 72% (2 steps); (h) TBAF, THF, 56%.



Scheme S2. Synthesis of the model compounds. Reagents and conditions: (a) **meso-B** (3-5 equiv), Br₂, CH₂Cl₂, -78 °C; **S9** (1.0 equiv), *i*-Pr₂NPh; (b) Et₃B, *n*-Bu₃SnH, O₂; (c) Cl₃CCO₂H, toluene, reflux; (d) TBSOTf, 2,6-lutidine, CH₂Cl₂, 0 °C; (e) K₂CO₃, MeOH, 42% (5 steps from **S9**); (f) Dess-Martin reagent, NaHCO₃, CH₂Cl₂, 96%; (g) K₂CO₃, MeOH; (h) Dess-Martin reagent, NaHCO₃, CH₂Cl₂, 19% for **3c** (4 steps from **S9**);



Steroid 2ca. A solution of KN(TMS)_2 (0.99 M, 8 μL , 8 μmol) freshly prepared from KH and HN(TMS)_2 in THF was added to a solution of triketone **3c** (13 mg, 0.026 mmol) in refluxing THF (270 μL), and the reaction mixture was stirred for 2 h. After the mixture was cooled to room temperature, saturated aqueous NH_4Cl (0.3 mL) was added. The resultant mixture was extracted with EtOAc (10 mL x3), and the combined organic layers were washed with brine (10 mL), dried over Na_2SO_4 and concentrated. The residue was purified by flash column chromatography on silica gel (1 g, hexane/EtOAc 5/1) to afford **2ca** (8.2 mg, 0.017 mmol) in 65% yield: crystal; m.p. 209-210 $^\circ\text{C}$; $[\alpha]_{\text{D}}^{25}$ 15 (c 0.72, CHCl_3); IR (neat) ν 3525, 2931, 2857, 1739, 1699, 1467, 1381, 1254, 1197, 1102, 1056, 1002 cm^{-1} ; ^1H NMR (400 MHz, C_6D_6) δ 0.09 (6H, s, CH_3 of TBS x2), 0.29 (1H, ddd, $J = 13.7, 13.7, 2.8$ Hz), 0.74 (1H, dd, $J = 15.1, 6.0$ Hz, $\text{CH}_\text{A}\text{H}_\text{B}\text{CCH}_3$), 0.84-0.94 (1H, m), 1.01 (9H, s, $t\text{-Bu}$ of TBS), 1.02-1.09 (1H, m), 1.16-1.28 (2H, m, COCHCH), 1.37 (3H, s, CH_3), 1.50-1.79 (5H, m, $\text{COCH}_2\text{CH}_\text{A}\text{H}_\text{B}$), 1.84 (1H, d, $J = 15.1$ Hz, $\text{CH}_\text{A}\text{H}_\text{B}\text{CCH}_3$), 1.86-1.92 (1H, m, $\text{CH}(\text{OCH}_3)\text{CH}$), 2.09-2.20 (2H, m, $\text{COCH}_\text{A}\text{H}_\text{B}\text{CH}_\text{A}\text{H}_\text{B}$), 2.33 (1H, ddd, $J = 13.3, 3.6, 3.6$ Hz), 2.46 (1H, ddd, $J = 19.2, 9.6, 9.6$ Hz, $\text{COCH}_\text{A}\text{H}_\text{B}\text{CH}_2$), 2.57 (1H, $J = 13.3$ Hz, COCHCH), 3.07 (1H, d, $J = 12.4$ Hz, $\text{CH}_\text{A}\text{H}_\text{B}\text{OCH}$), 3.33 (3H, s, OCH_3), 3.38 (1H, dddd, $J = 5.5, 5.5, 5.5, 5.5$ Hz, CHOTBS), 3.70 (1H, dd, $J = 12.4, 1.4$ Hz, $\text{CH}_\text{A}\text{H}_\text{B}\text{OCH}$), 3.80 (1H, s, OH), 4.44 (1H, d, $J = 9.6$ Hz, CHOCH_3); ^{13}C NMR (125MHz, C_6D_6) δ -4.4, -4.3, 17.2, 18.3, 26.1, 27.9, 30.9, 31.4, 32.3, 33.0, 35.8, 36.4, 38.5, 41.6, 44.8, 48.6, 50.5, 52.0, 56.1, 61.2, 71.1, 80.2, 102.5, 211.4, 217.3; HRMS (ESI) calcd for $\text{C}_{27}\text{H}_{44}\text{O}_6\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 515.2799, found 515.2799.



Steroid 2db. A solution of KN(TMS)_2 (0.97 M, 16 μL , 0.016 mmol) freshly prepared from KH and HN(TMS)_2 in THF was added to a solution of triketone **3d** (24 mg, 0.053

mmol) in refluxing THF (0.53 mL), and the reaction mixture was stirred for 2 h. After the mixture was cooled to room temperature, saturated aqueous NH_4Cl (0.5 mL) was added. The resultant mixture was extracted with EtOAc (10 mL x3), and the combined organic layers were washed with brine (3 mL), dried over Na_2SO_4 and concentrated. The residue was purified by flash column chromatography on silica gel (2 g, hexane/EtOAc 5/1 to 3/1 to 1/1 to EtOAc) to afford **2db** (18 mg, 0.039 mmol) in 74% yield, respectively: white solid; m.p. 200-202 °C; $[\alpha]_{\text{D}}^{25}$ -37 (*c* 0.36, CHCl_3) ; IR (neat) ν 3507, 2929, 2856, 1740, 1701, 1471, 1378, 1294, 1255, 1192, 1125, 1097, 1055 cm^{-1} ; ^1H NMR (500 MHz, C_6D_6) δ 0.06 (3H, s, CH_3 of TBS), 0.07 (3H, s, CH_3 of TBS), 0.40 (1H, ddd, J = 13.8, 13.8, 3.4 Hz), 1.00 (9H, *t*-Bu of TBS), 1.03 (3H, s, CH_3), 1.11-1.32 (5H, m), 1.52-1.64 (3H, m), 1.78 (1H, ddd, J = 19.5, 10.3, 10.3 Hz), 1.88-1.94 (4H, m), 2.00 (1H, ddd, J = 13.8, 4.0, 3.6 Hz), 2.12 (1H, d, J = 11.4 Hz), 2.10-2.20 (1H, m), 2.59 (1H, d, J = 13.2 Hz), 3.34 (1H, m, CHOTBS), 3.40 (1H, d, J = 11.4 Hz, $\text{CH}_\text{A}\text{H}_\text{B}\text{OCH}=\text{C}$), 3.85 (1H, dd, J = 11.4, 1.8 Hz, $\text{CH}_\text{A}\text{H}_\text{B}\text{OCH}=\text{C}$), 4.29 (1H, s, OH), 6.21 (1H, s, $\text{CH}_2\text{OCH}=\text{C}$); ^{13}C NMR (125MHz, C_6D_6) δ -4.47, -4.37, 18.3, 19.3, 26.1, 30.8, 31.1, 31.5, 33.7, 34.1, 34.7, 38.2, 44.2, 45.5, 46.1, 56.1, 59.0, 61.8, 71.2, 78.5, 112.8, 136.1, 211.7, 215.0; HRMS (ESI) calcd for $\text{C}_{26}\text{H}_{40}\text{O}_5\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 483.2537, found 483.2558.

Computer Simulation

Optimized geometries of the structures of **2aa-2ah** and **2ba-2bh**, and their stabilities were evaluated in silico. The initial structures of **2aa-2ah** and **2ba-2bh** were built by the molecular mechanics simulation using a 1000-steps of mixed torsional/low-mode sampling conformational search and PRCG energy minimization with Merck Molecular Force Field (MMFFs, MacroModel).^{S10,11} The geometry optimizations and frequency calculations of the resulting conformations were performed at the PM6 semiempirical method^{S12} using the Gaussian 09 program package.^{S13} The difference in the Gibbs free energies (ΔG at 339.15 K, 1 atm), which are computed for the gas phase, are given in kcal mol⁻¹.

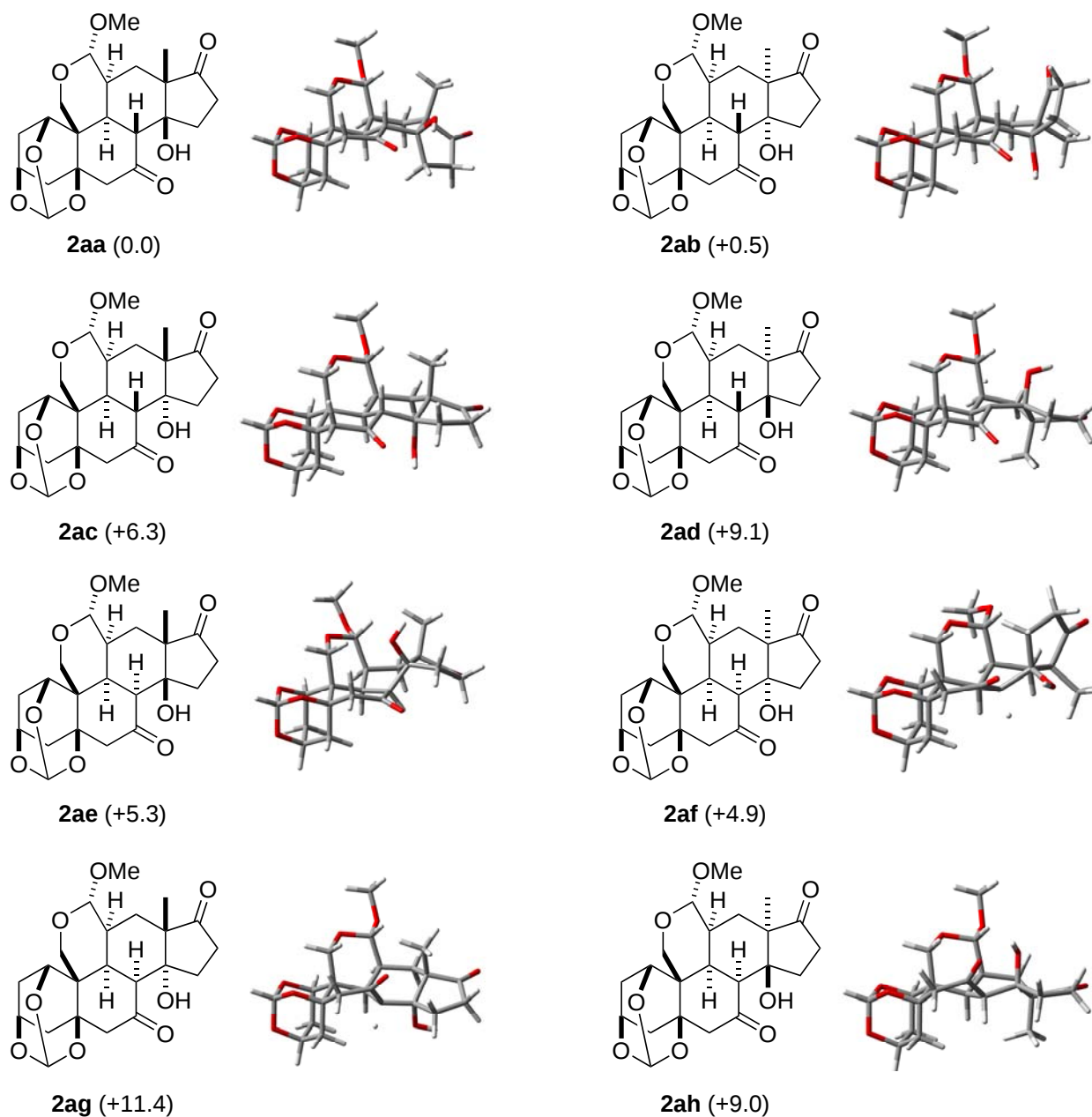


Figure S1. Optimized geometries of the structures of **2aa-2ah**.
Number in parenthesis shows the relative energy (kcal mol⁻¹).

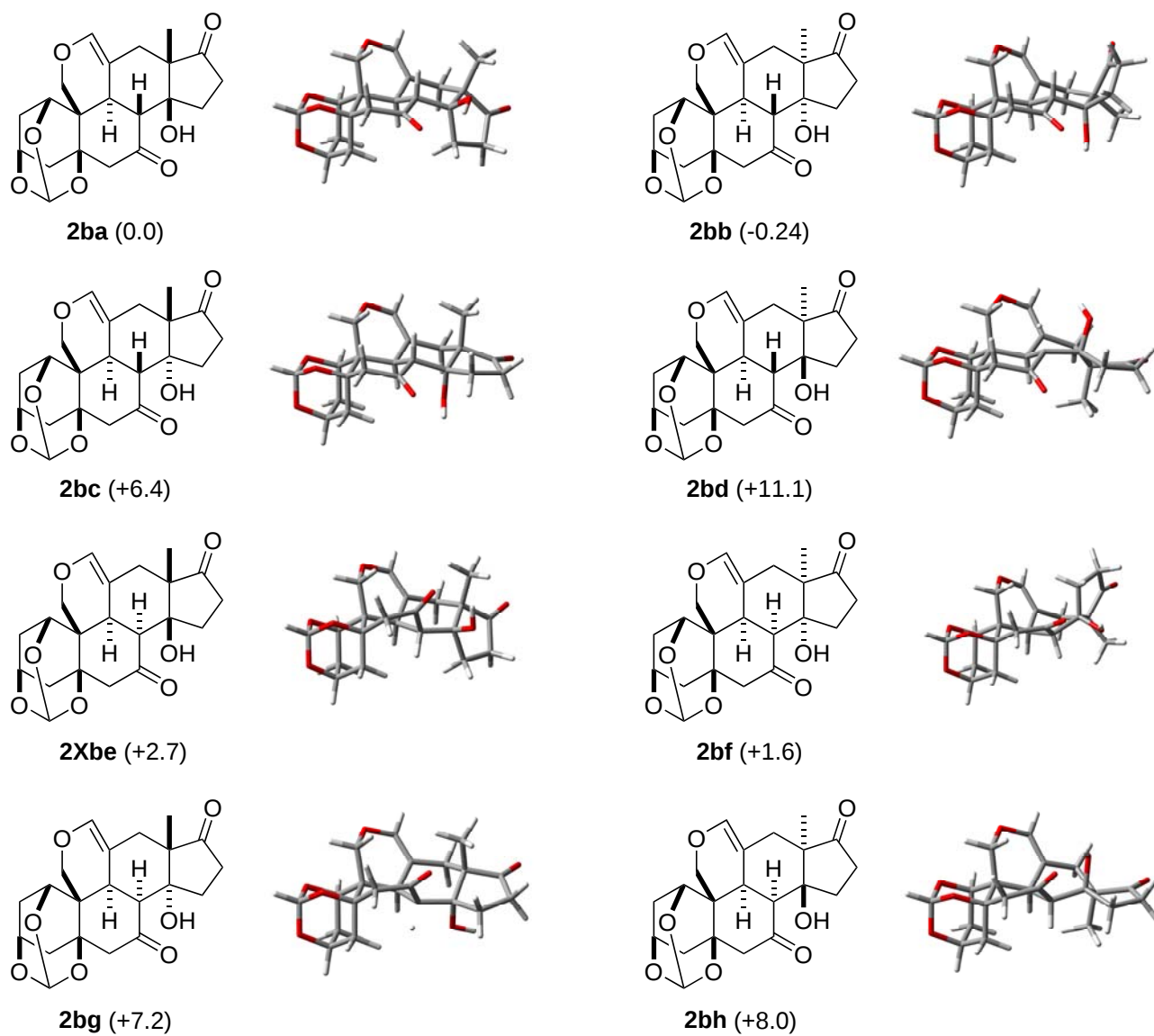


Figure S2. Optimized geometries of the structures of **2ba-2bh**.
Number in parenthesis shows the relative energy (kcal mol⁻¹).

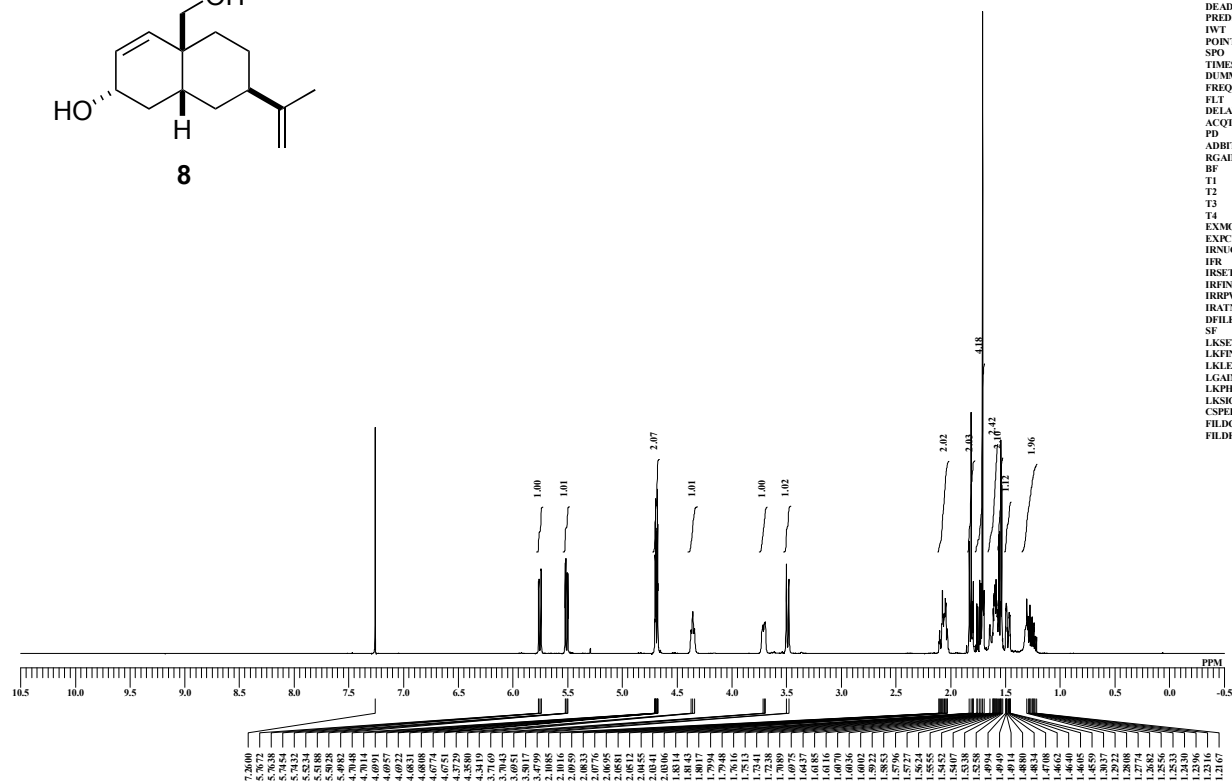
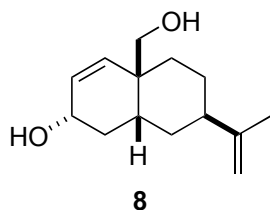
References

- S1. S. A. Kozmin, J. M. Janey and V. H. Rawal, *J. Org. Chem.*, 1999, **64**, 3039.
- S2. M. A. Tius and M. A. Kerr, *Synth. Commun.*, 1988, **18**, 1905.
- S3. K. Tiefenbacher and J. Mulzer, *Angew. Chem., Int. Ed.*, 2008, **47**, 6199.
- S4. K. Mukai, D. Urabe, S. Kasuya, N. Aoki and M. Inoue, *Angew. Chem., Int. Ed.*, 2013, **52**, 5300.
- S5. (a) C. A. Brown, *J. Org. Chem.*, 1974, **39**, 3913; (b) J. Åhman and P. Somfai, *Synth. Commun.*, 1995, **25**, 2301. For a titration method of KN(TMS)₂, see: (c) R. E. Ireland and R. S. Meissner, *J. Org. Chem.*, 1991, **56**, 4566.
- S6. Ouabagenin (**1**) has a propensity to form the corresponding borates with B₂O₃ leached from borosilicate glassware (Pyrex glass). Therefore, the reaction was carried out in quartz glass flask, and soda-lime glassware was used for the purification, and quartz glass tube was used for the NMR experiments. For a related example, see: (a) A. Kawamura, J. Guo, Y. Itagaki, C. Bell, Y. Wang, G. T. Hauptert, Jr., S. Magil, R. T. Gallagher, N. Berova and K. Nakanishi, *Proc. Natl. Acad. Sci. USA*, 1999, **96**, 6654; (b) M. Nagatomo, M. Koshimizu, K. Masuda, T. Tabuchi, D. Urabe and M. Inoue, *J. Am. Chem. Soc.*, 2014, **136**, 5916.
- S7. (a) C. Mannich and G. Siewert, *Chem. Ber.*, 1942, **75**, 737; (b) K. Florey and M. Ehrenstein, *J. Org. Chem.*, 1954, **19**, 1174.
- S8. Vogel and co-workers reported the full assignment of ¹H and ¹³C NMR peaks of ouabagenin (**1**) at 333 K in a 2 : 1 mixture of DMSO-*d*₆ and CDCl₃. D. D. McIntyre, M. W. Germann and H. J. Vogel, *Can. J. Chem.*, 1990, **68**, 1263.
- S9. N. Toyooka, A. Nishino and T. Momose, *Tetrahedron*, 1997, **53**, 6313.
- S10. T. A. Halgren, *J. Comput. Chem.*, 1999, **20**, 730.
- S11 (a) *MacroModel version 10.4*; Schrodinger, LLC, New York, NY; (b) F. Mohamadi, N. G. J. Richards, W. C. Guida, R. Liskamp, M. Lipton, C. Caufield, G. Chang, T. Hendrickson and W. C. Still, *J. Comput. Chem.*, 1990, **11**, 440.
- S12 J. J. P. Stewart, *J. Mol. Model.*, 2007, **13**, 1173.
- S13 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N.

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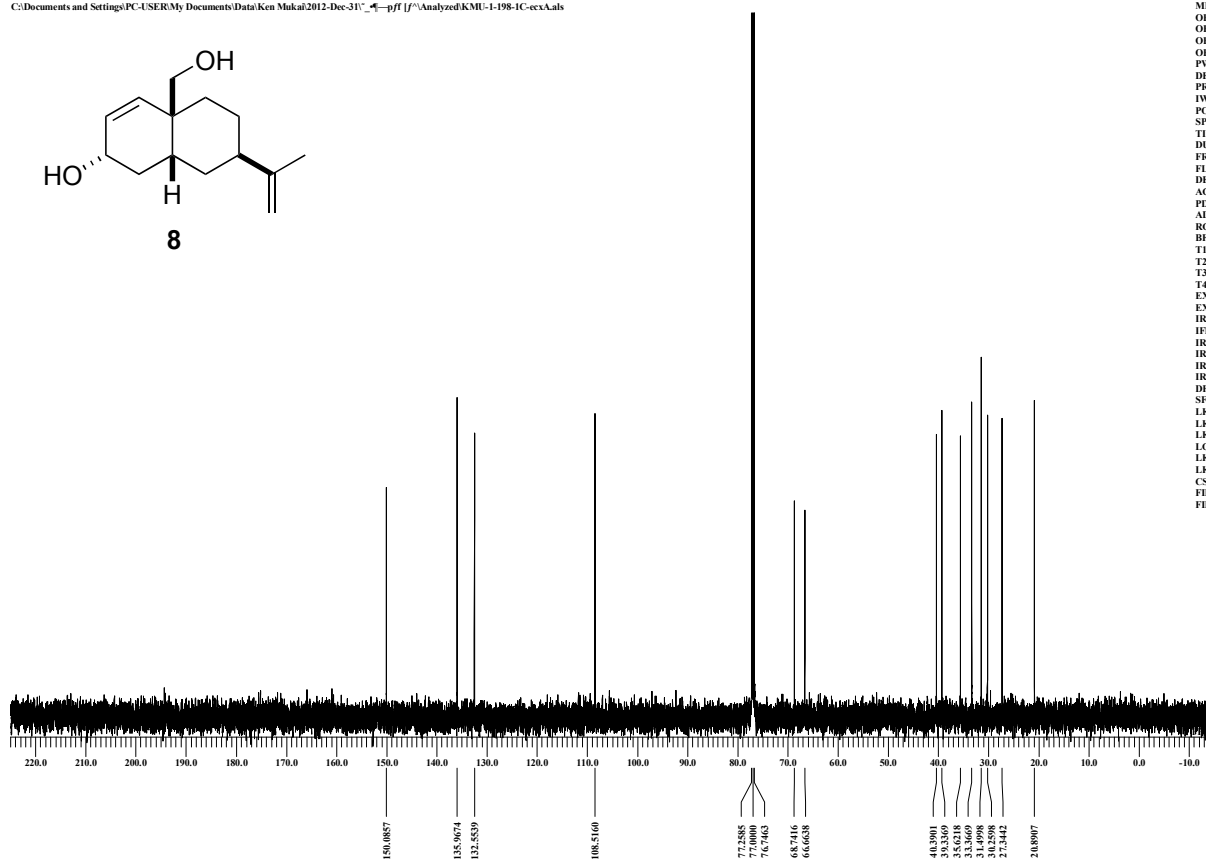
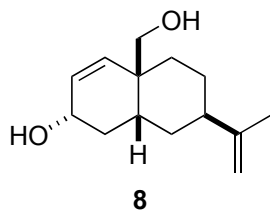
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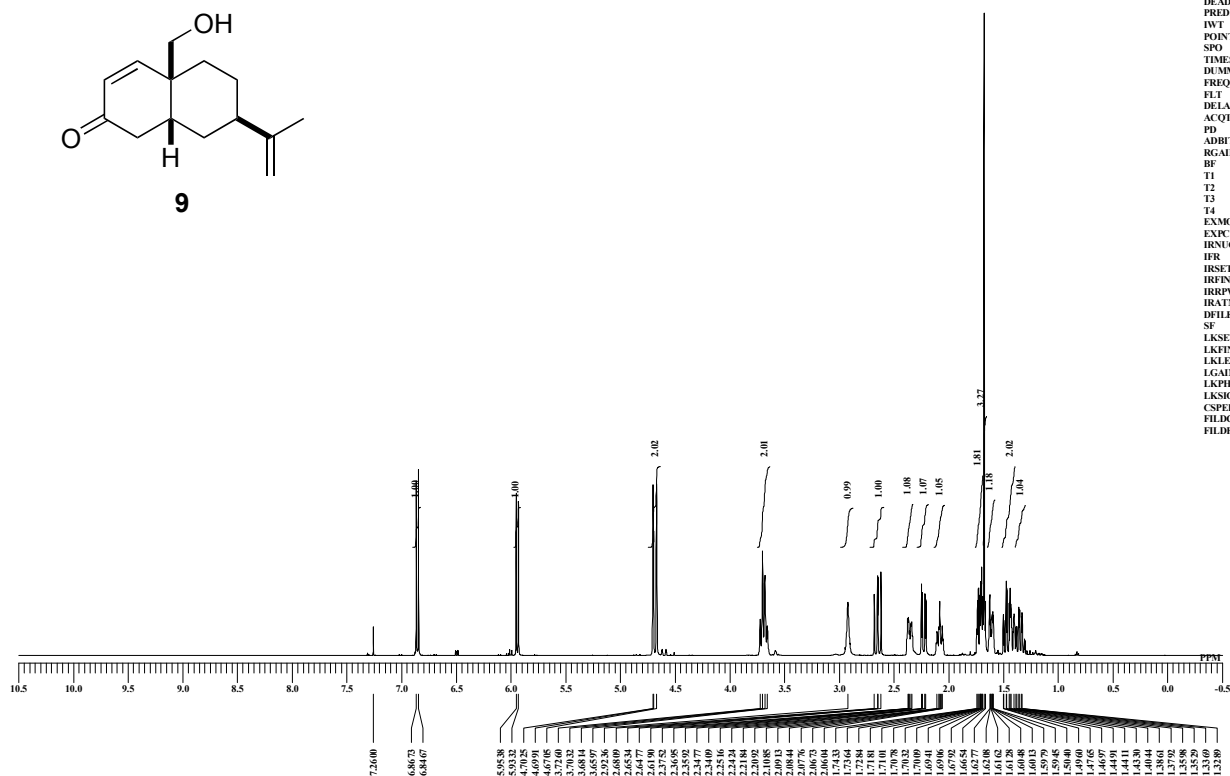
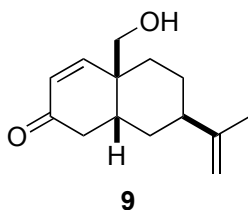
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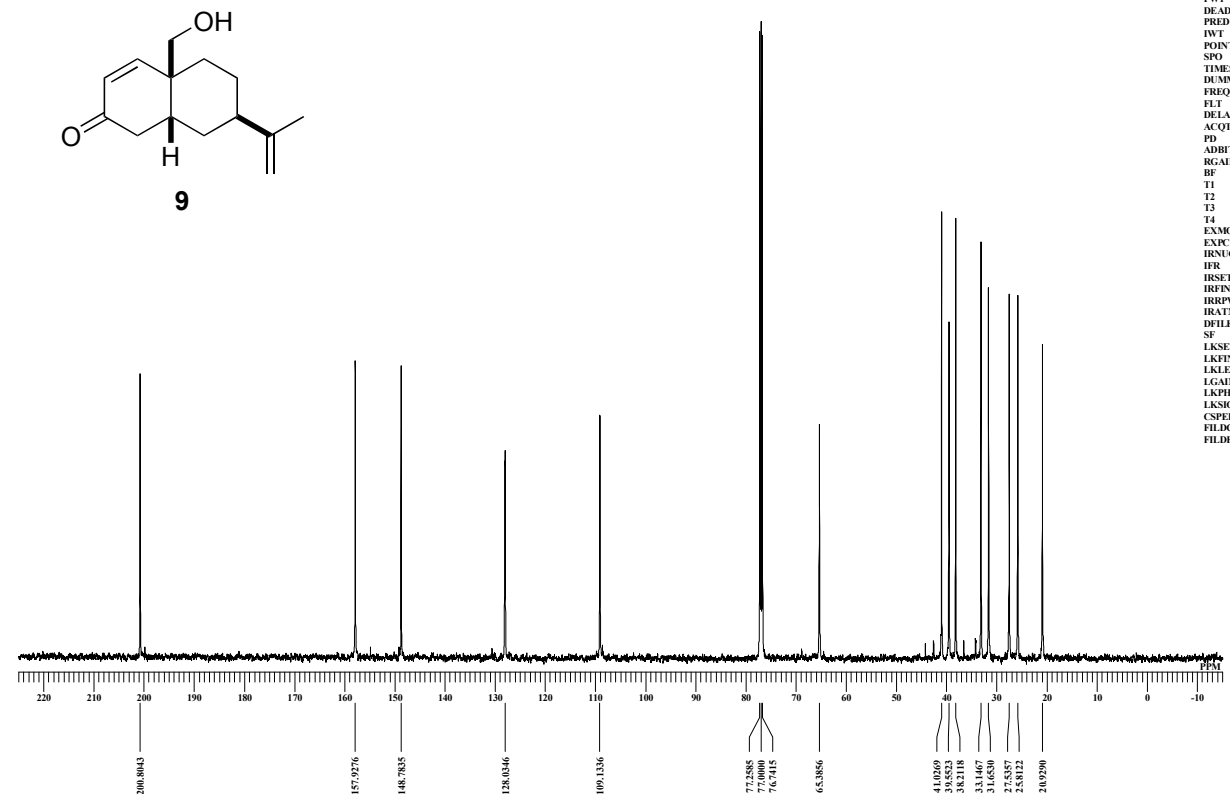
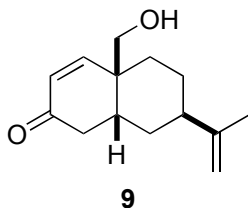
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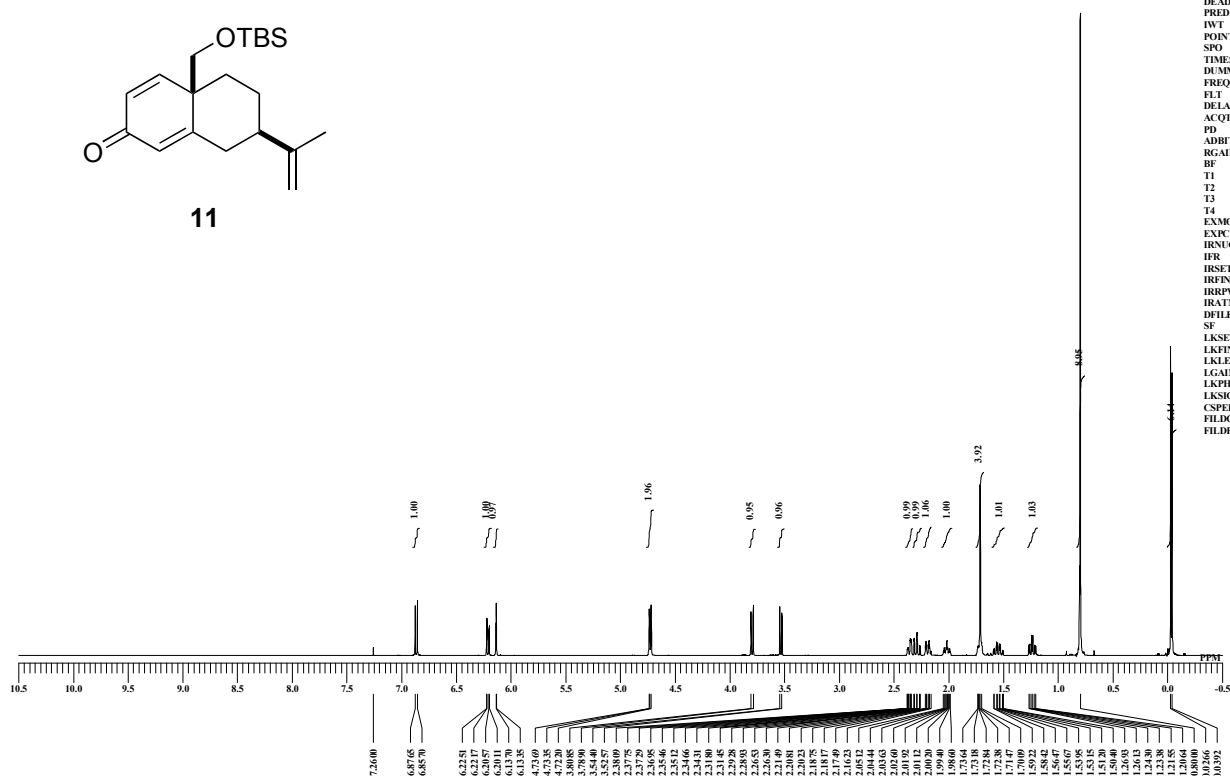
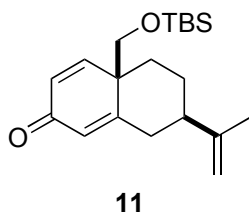
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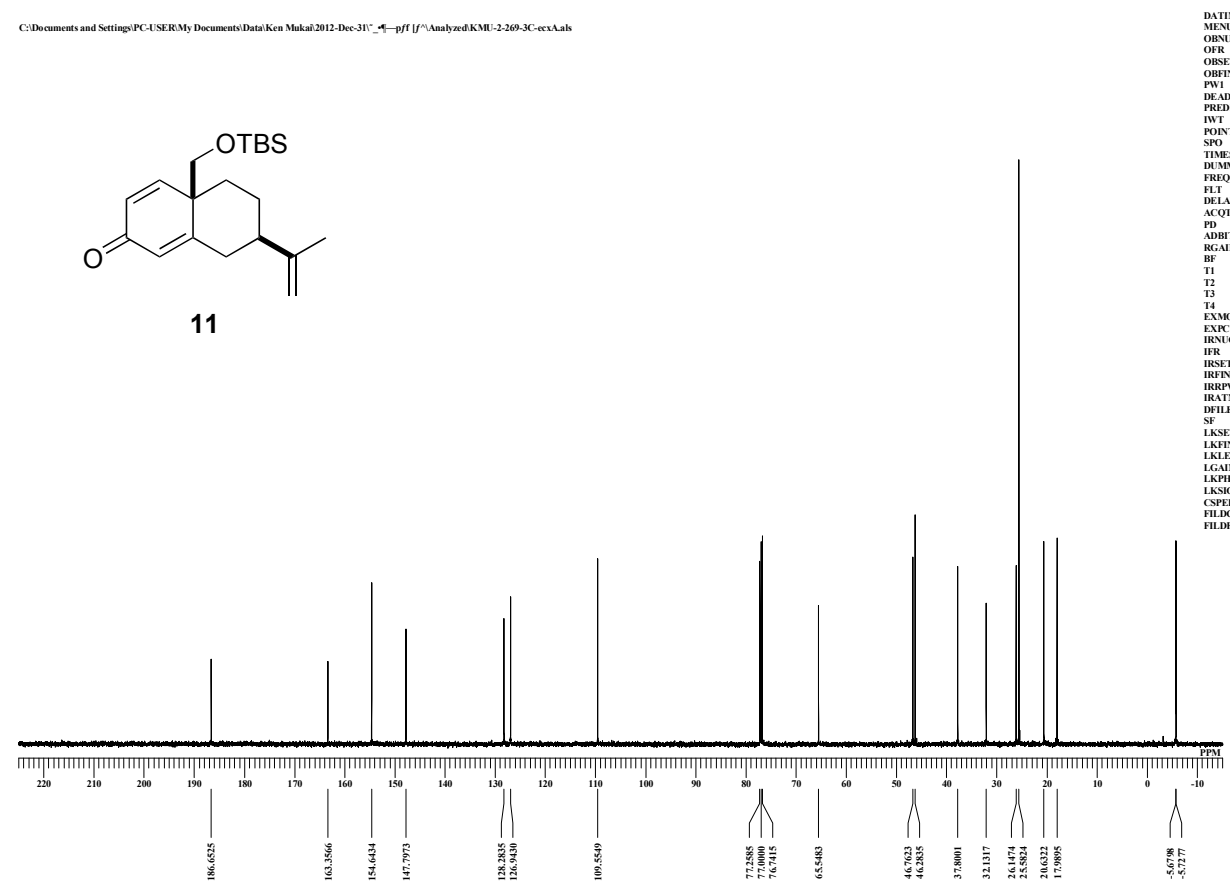
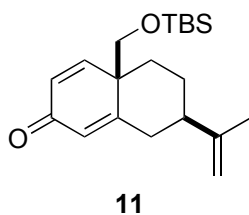
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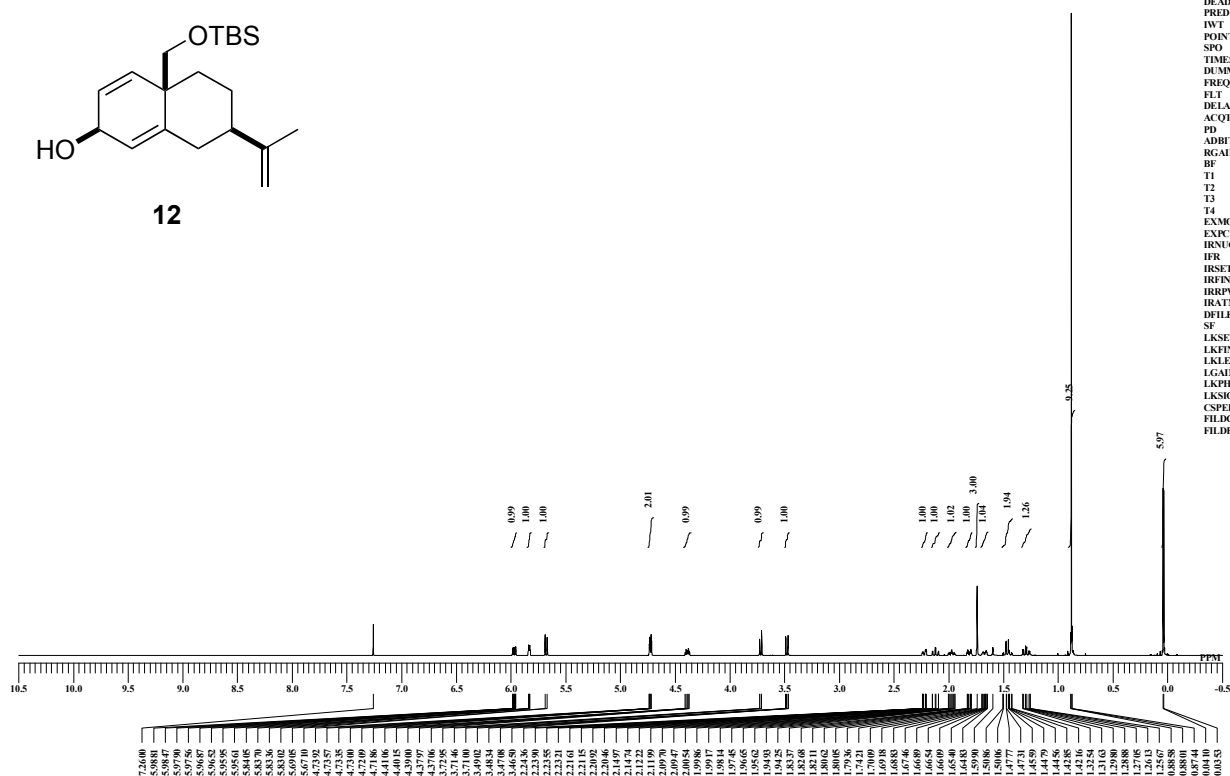
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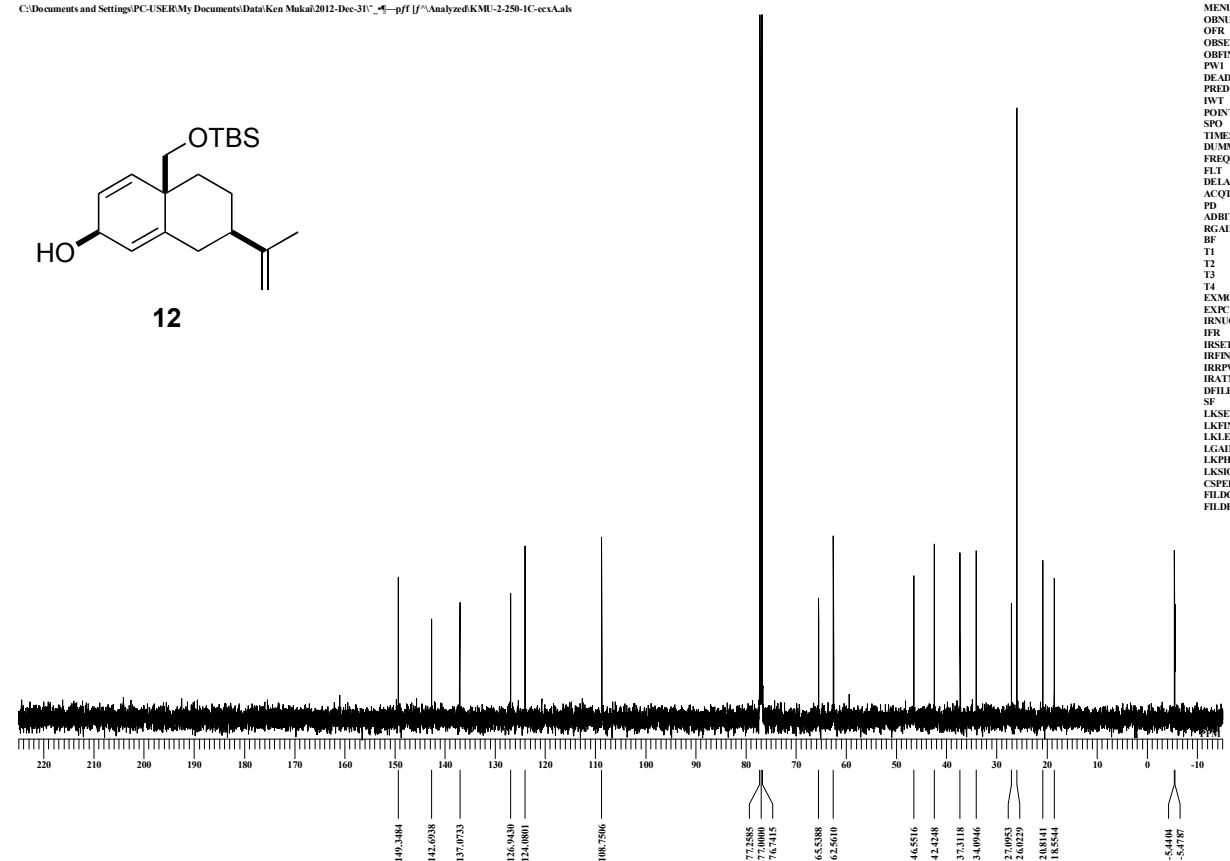
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T4	100.00
EXMOD	single_pulse_ext
EXPCM	
INVCU	IH
IFR	495.13 MHz
IRSET	4.38 kHz
IRFIN	9.64 Hz
IRPWF	92 msec
IRATN	79
DFLE	KML-2-250-1-eexA.es
LFSET	-601.50 KHz
LKSET	-1.8 Hz
LKLEV	0
LGAIN	0
LKPIN	0
LKSG	0
CPSED	0 Hz
FILDC	
T1 DE	

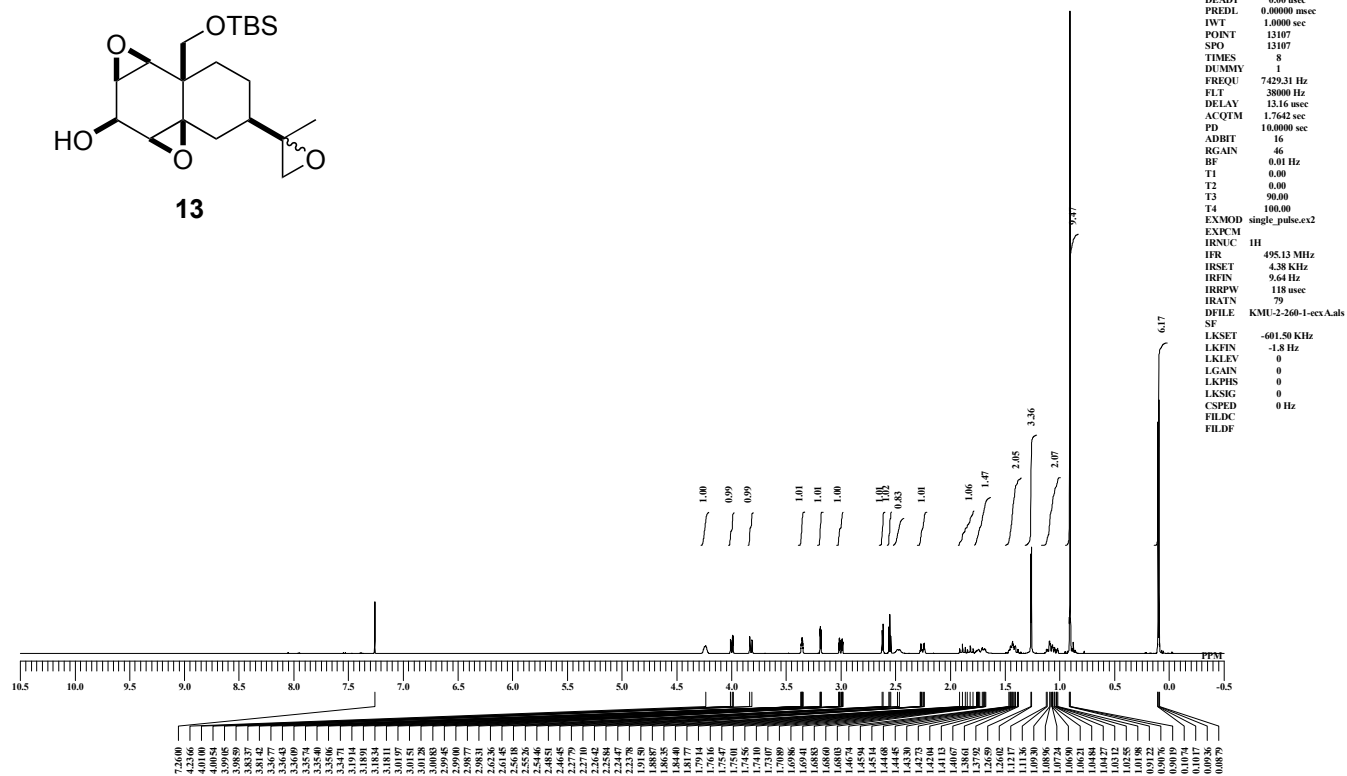
C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\2012-Dec-31\pff\pff\Analyzed\KMU-2-250-1C-ecx\A.als



DATIM	01-09-2017 17:05:56
MENUF	
OBNUC	13C
OFNR	1342.5 MHz
ORSET	3.45 KHz
OBFTN	6.00 Hz
PW1	3.20 Hz
DEADT	0.000 sec
PREDI	0.00000 msec
IWT	1.00000 msec
POINT	26214
SPO	26214
TIMES	128
DUMMY4	0
FREQU	312429.5 Hz
FL1	15790.0 Hz
DELAY	2.640 msec
ACQTM	0.8387 sec
PD	7.000 sec
ADBIT	16
RGAIN	54
BF	1.00 Hz
T1	0.00
T2	0.00
T3	90.00
T4	100.00
EXMOD	single_pulse_dec
EXNC	
RNUC	1H
IFR	495.13 MHz
IRSET	4.98 KHz
IRFTH	9.64 Hz
IRWTR	92 msec
IRATN	7
DFLE	KML-2-250-13-cex-AAS
LKSET	-601.50 KHz
LKFIN	-1.8 Hz
LKE1V	0
LGE1N	0
LKPHS	0
LKSGS	0
CPSPD	0 Hz
FILDC	
FILDF	

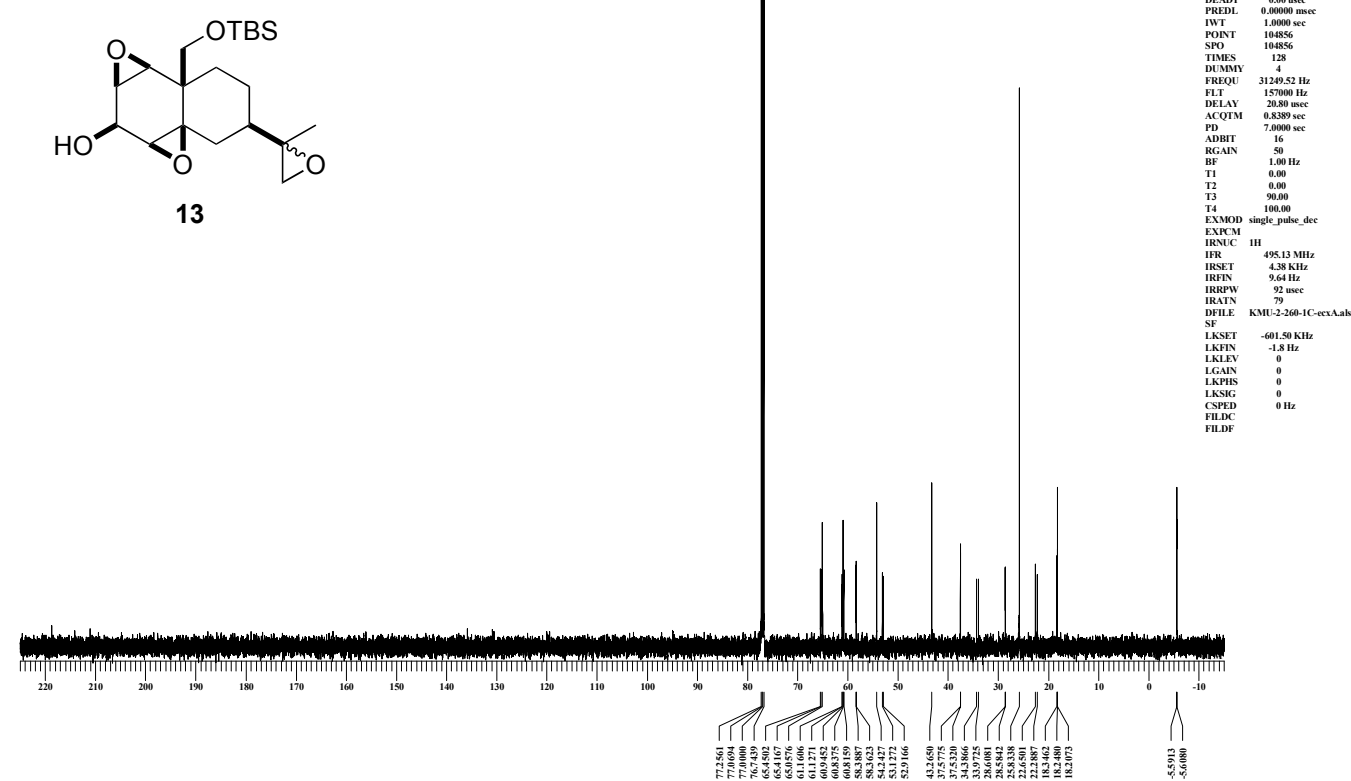
KMU-2-260-1-ecx

C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\2012-Dec-31\pft [1^]Analyzed\KMU-2-260-1-ecx.Aals

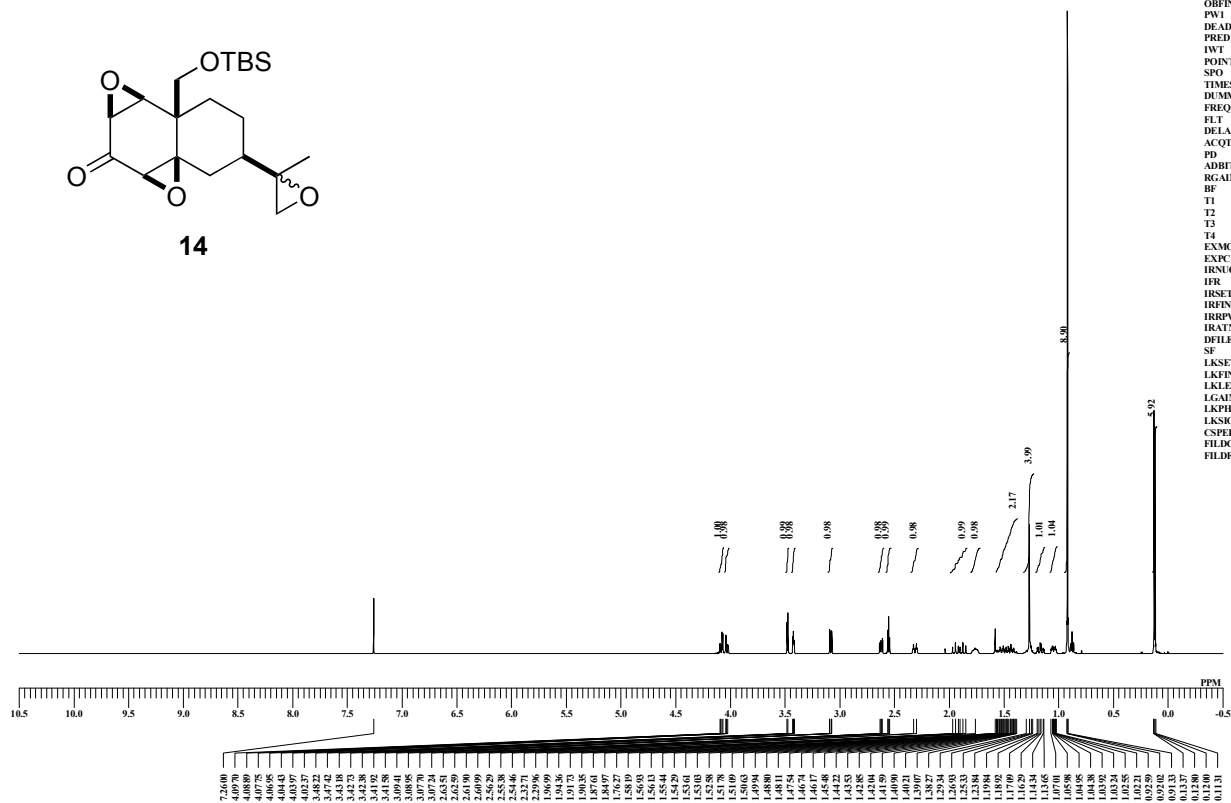


KMU-2-260-1C-ecx

C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\2012-Dec-31\pft [1^]Analyzed\KMU-2-260-1C-ecx.Aals

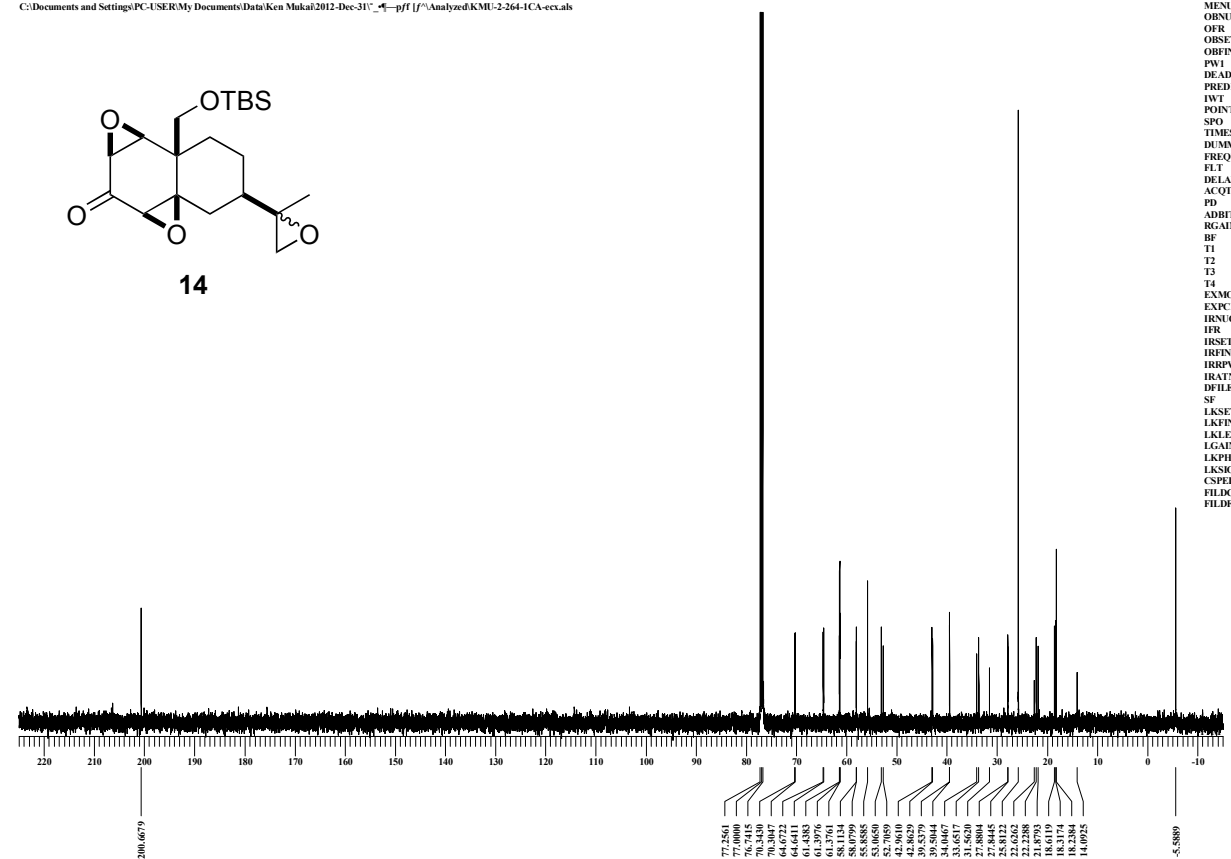


C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\2012-Dec-31* --pff | f^\Analyzed\KMU-2-264-1A-ecx.xls



DATIM	10:09:20 10:15:46:40
MENUP	
ONBUC	IH5
OFR	495.13 MHz
ORSET	4.38 KHz
ORFIN	9.64 Hz
PW1	6.00 usec
DE ADT	0.00 usec
PREDL	0.00 usec
IWT	1.0000 sec
POINT	13107
SPO	13107
TIMES	
DUMMYY	
FREQU	7429.31 Hz
FLT	38000 Hz
DE LAY	13.16 usec
ACQTM	1.7642 sec
PD	10.0000 sec
ADBIT	16
RGAIN	40
BF	0.01 Hz
T1	0.00 usec
T3	0.00 usec
T4	100.00
EXMOD	single_pulse
EXPCM	
IRNVC	IH5
IFR	495.13 MHz
IRSET	4.38 KHz
IRFIN	9.64 Hz
IRBPW	92 usec
IRATN	79
DFILE	KML-2-264-14-ecalls
FLD	
LKSET	-601.50 KHz
LKFIN	-1.8 Hz
LKLEV	0
LKGAIN	0
LKPUS	0
LKSG	0
CPSED	0 Hz
FILED	

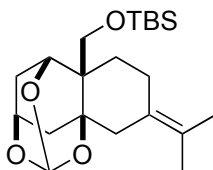
C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\2012-Dec-31* -pff\Analyzed\KMU-2-264-1CA-ecx.als



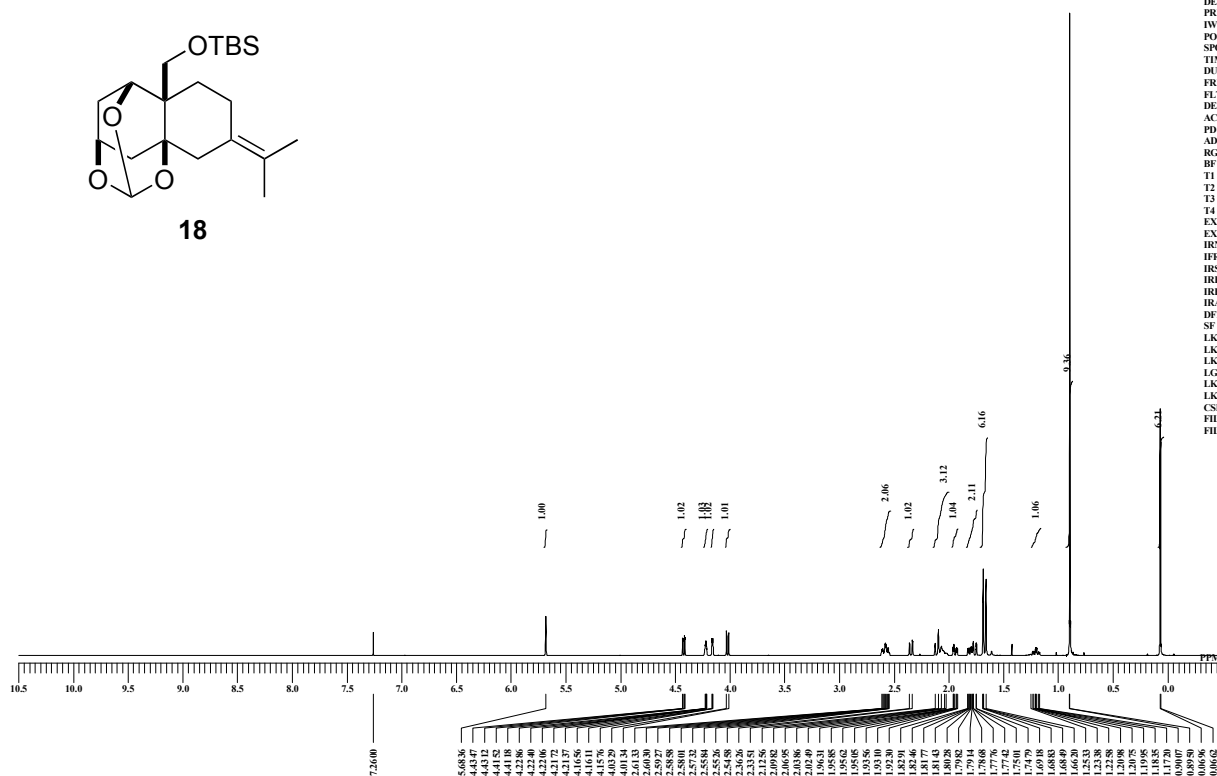
DATIM	10-09-2010 16:46:22
MENUF	
OBNUC	1241
OFR	134.5 MHz
ORSET	3.25 kHz
OFBTF	6.00 Hz
PW1	3.5 Hz
DE-ADT	0.00 msec
PREDL	0.0000 msec
IWT	1.0000 sec
POINT	104856
SPO	104856
TIMES	256
DUMMY4	
FREQU	31429.52 Hz
FLT	15700.0 Hz
DELAY	20.00 msec
ACQTM	0.8380 sec
PD	7.0000 sec
ADBIT	16
RG-MAIN	56
BF	1.00 Hz
T1	0.00
T2	0.00
T3	90.00
T4	100.00
EXMCD	single_pulse_dec
ENCODE	
IRNUC	III
IFR	495.13 Hz
IRSET	4.928 kHz
IRF	6.4 Hz
IRRPW	92 msec
IRATN	79
DFE	KMU-2-264-I-CESAT
LSKET	-601.50 KHz
LKFIN	-1.8 Hz
LK1EV	0
LGINV	0
LKPHS	0
LKSGS	0
CSPED	0 Hz
FILED	

KMU-2-285-D.P.-ecx

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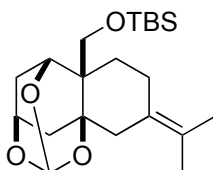
18



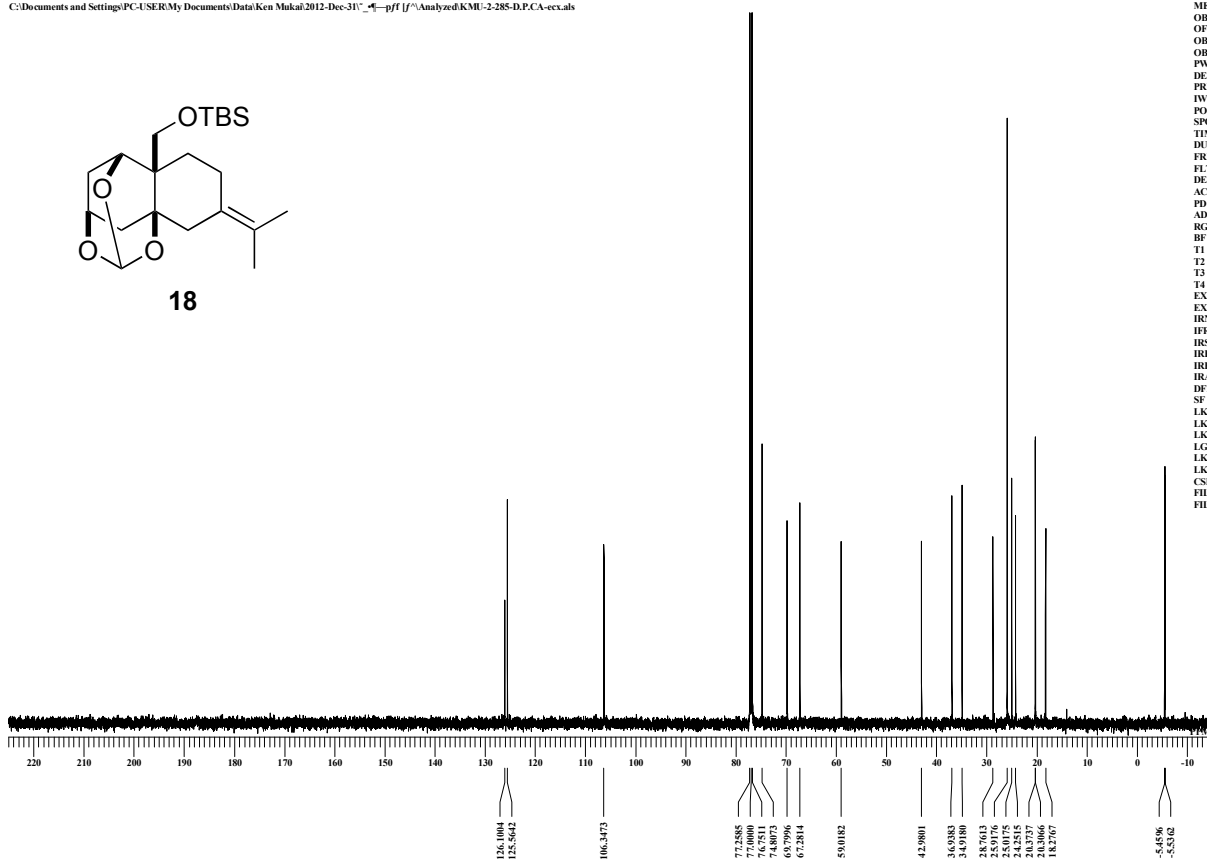
DATIM 06-10-2010 10:23:54
MENUP
ORNUC 1H
OFR 495.13 MHz
OBSET 4.38 KHz
OBFIN 9.64 Hz
PW1 6.00 usec
DEADT 0.00 usec
PREDI 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 8
DUMMY 1
FREQU 7429.31 Hz
FLT 38000 Hz
DELAY 13.16 usec
ACQTM 1.7642 sec
PD 10.0000 sec
ADBIT 16
RGAIN 40
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IR 495.13 MHz
IRSET 4.38 KHz
IRFIN 9.64 Hz
IRRPW 92 usec
IRATN 79
DFILE KMU-2-285-D.P.A-ecx.ab
SF
LKSET -601.50 KHz
LKFIN -1.8 Hz
LKLEV 0
LGAIN 0
LKFIN 0
LKSIG 0
CSPED 0 Hz
FILDC
FILDF

KMU-2-285-D.P.C-ecx

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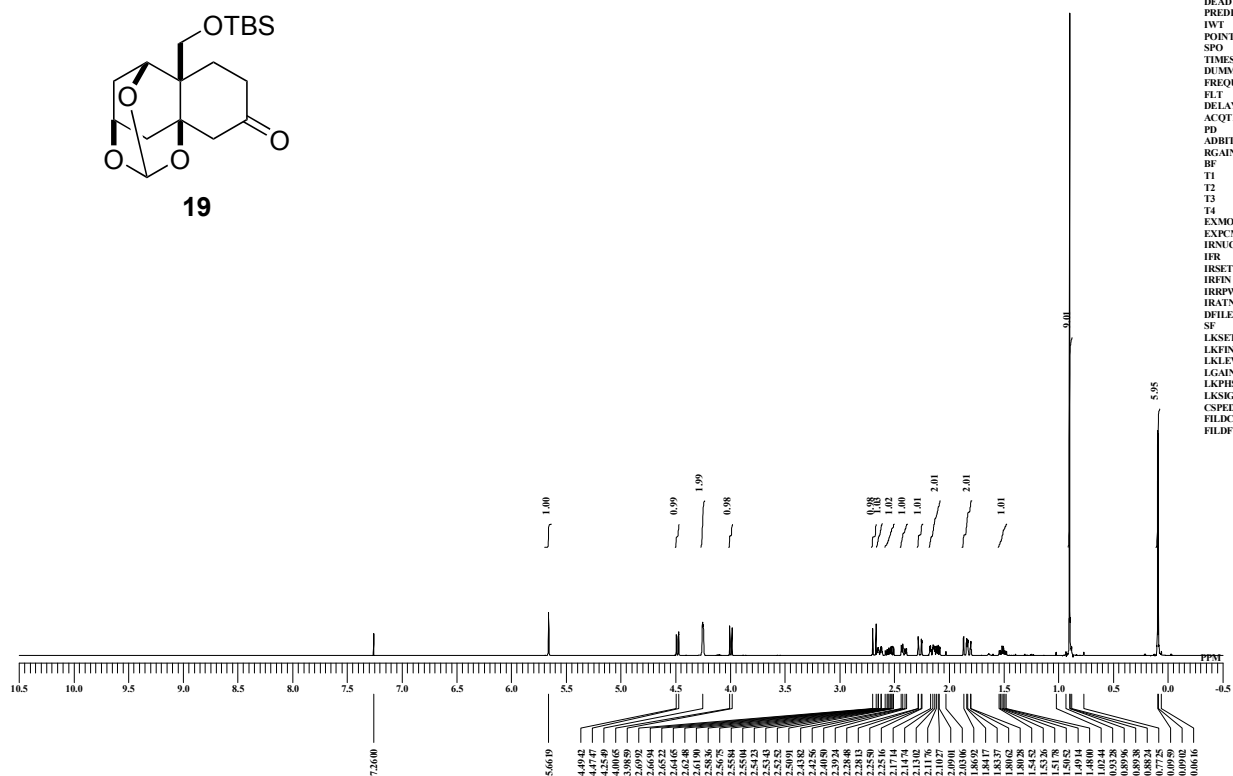


18



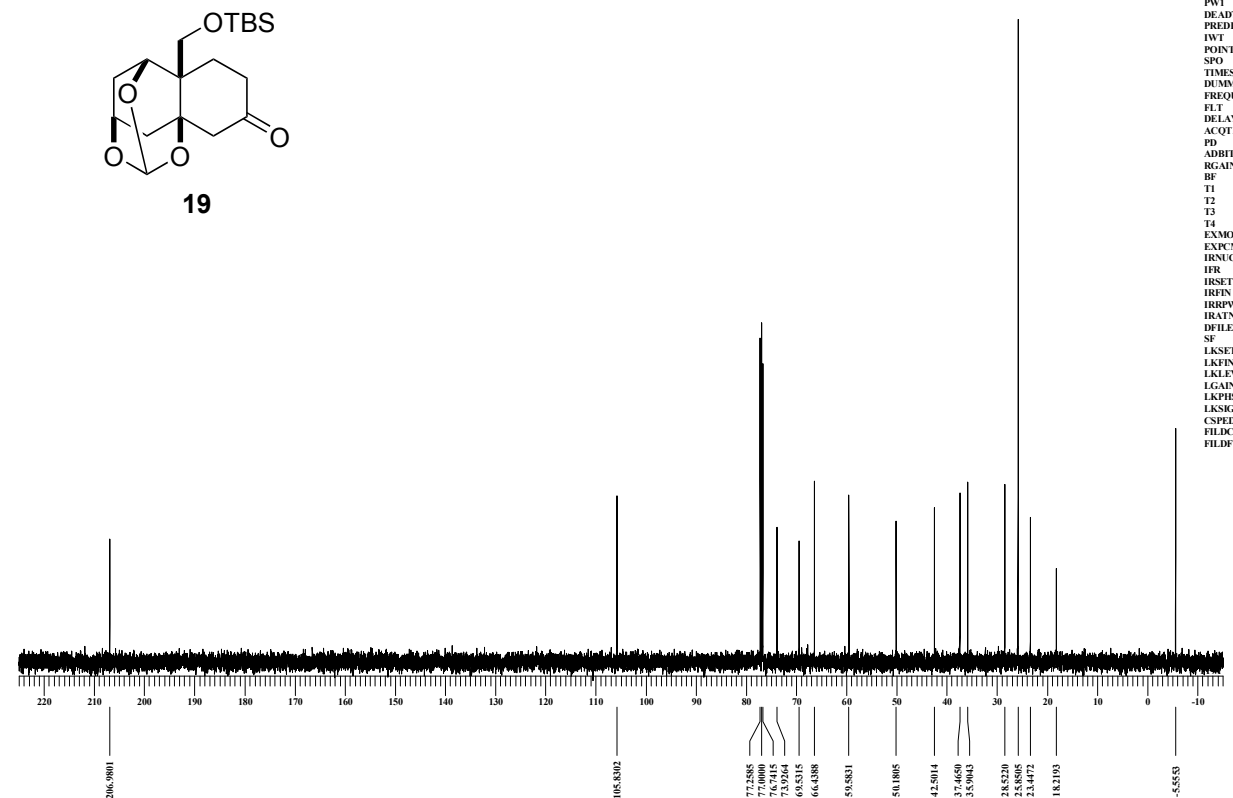
DATIM 06-10-2010 10:59:11
MENUP
ORNUC 13C
OFR 124.51 MHz
OBSET 3.45 KHz
OBFIN 6.00 Hz
PW1 3.25 usec
DEADT 0.00 usec
PREDI 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 26214
TIMES 256
DUMMY 4
FREQU 31249.52 Hz
FLT 157000 Hz
DELAY 20.80 usec
ACQTM 0.8389 sec
PD 7.0000 sec
ADBIT 16
RGAIN 54
BF 1.00 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 1H
IR 495.13 MHz
IRSET 4.38 KHz
IRFIN 9.64 Hz
IRRPW 92 usec
IRATN 79
DFILE KMU-2-285-D.P.C.A-ecx.a
SF
LKSET -601.50 KHz
LKFIN -1.8 Hz
LKLEV 0
LGAIN 0
LKFIN 0
LKSIG 0
CSPED 0 Hz
FILDC
FILDF

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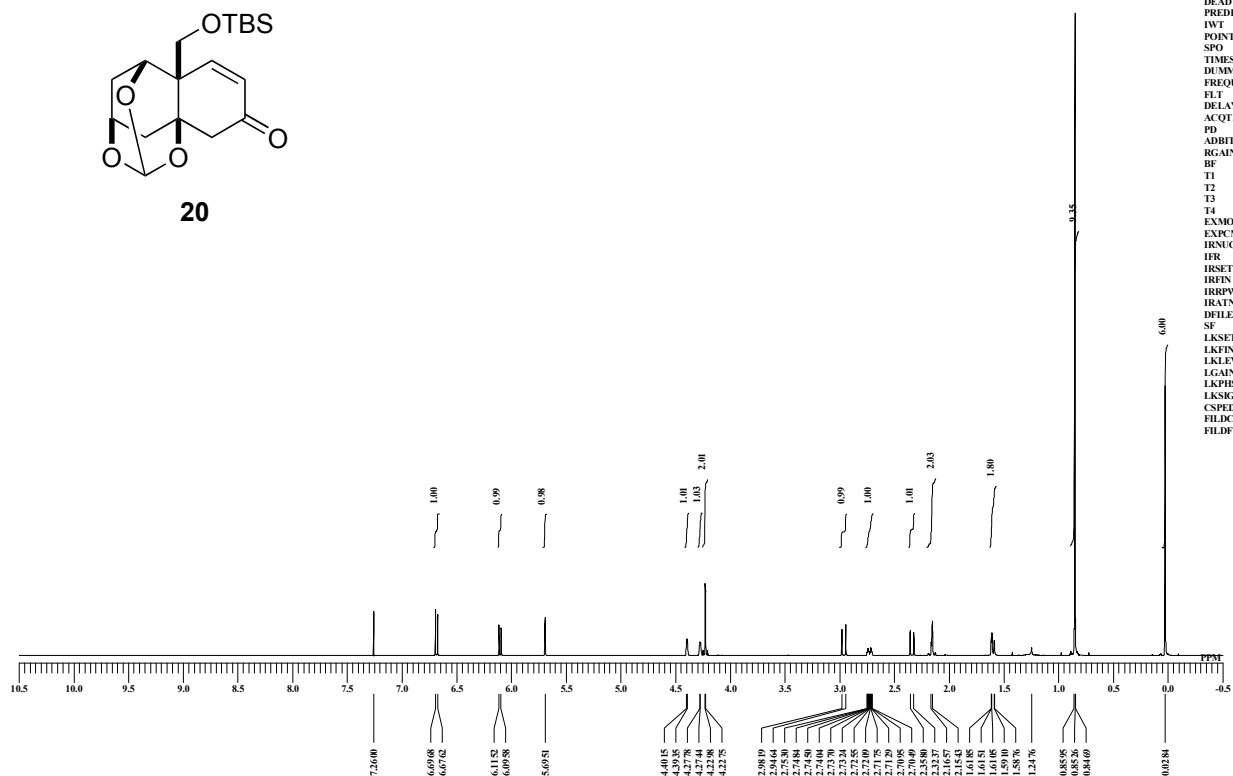
DATIM	07-10-2010 15:57:25
MENUP	
OBNUC	IH5
OFR	495.13 MHz
ORSET	4.38 KHz
ORFIN	9.64 Hz
PW1	6.00 usec
DE ADT	0.00 usec
PDEL1	13.16 usec
IWT	1.0000 sec
POINT	13107
SPO	13107
TIMES	
DUMMY1	
FREQU	7429.31 Hz
FLT	38000 Hz
DE LAY	13.16 usec
ACQTM	1.7642 sec
PD	10.0000 sec
ADBIT	16
RGAIN	40
BF	0.01
T1	0.00 usec
T3	0.00 usec
T0	90.00 usec
T4	100.00 usec
EXMOD	single_pulse
EXPCM	
IRNUC	IH5
IFR	495.13 MHz
IRSET	4.38 KHz
IRFIN	9.64 Hz
IRAPW	92 usec
IRATN	79
DFILE	KML-2-286-1-rct-xxb-As
EXMOD	
IRNUC	-601.50 KHz
LKFEN	-1.8 Hz
LKLEV	0
LGAIN	0
LKPUS	0
LKSG	0
CPSED	0 Hz
FILDE	

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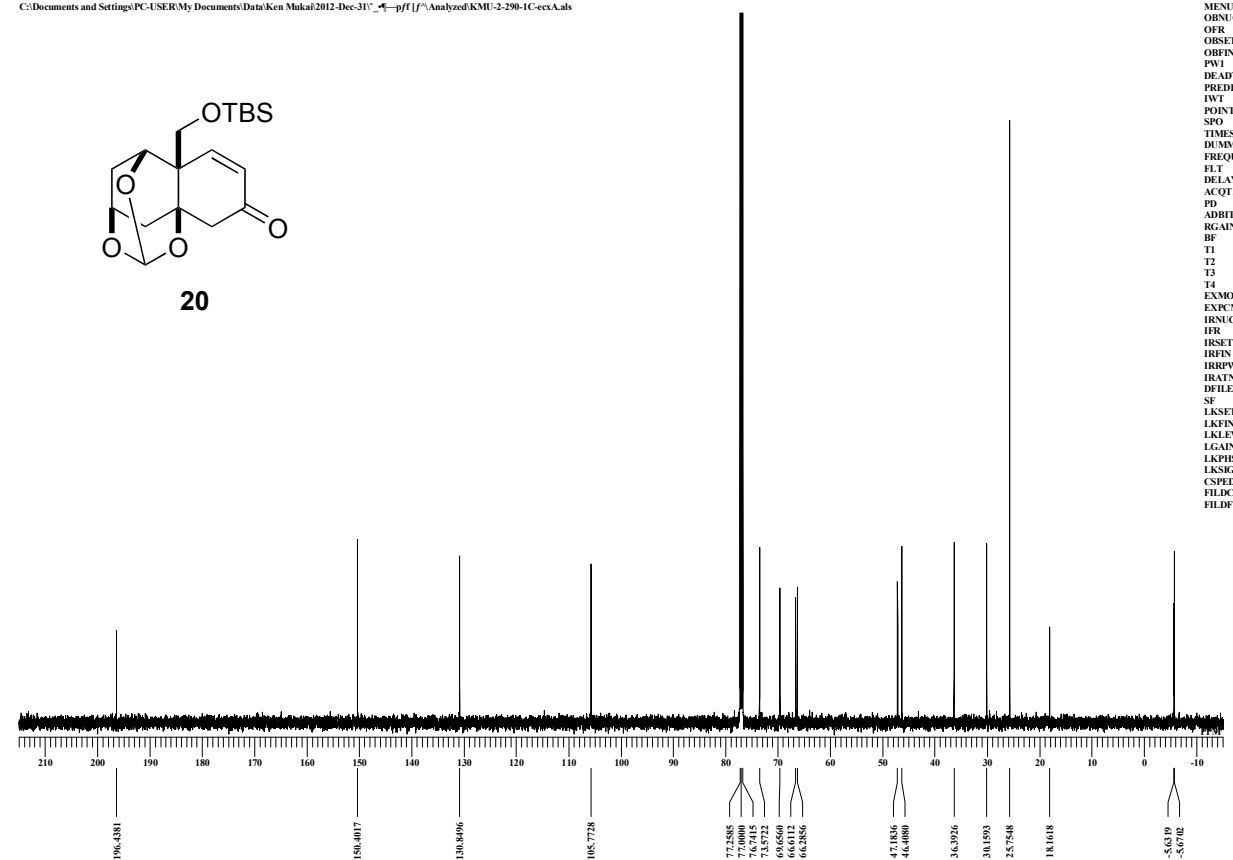


DATIM	07-10-2010 16:13:49
OBNUC	134.51 MHz
OFPR	125.5 kHz
OBSET	3.45 kHz
OBFTN	6.00 Hz
PW1	3.25 usec
DE-BIT	0.000 msec
PREDL	0.0000 msec
IWT	1.0000 sec
POINT	26214
SPO	26214
TIMES	32
DUMMY	4
FREQU	31429.52 Hz
FLT	157800 Hz
DELAY	20.00 msec
ACQTM	0.8389 sec
PDB	2.0000 sec
ADIT	16
RGAIN	50
BF	1.00 Hz
T1	0.00 Hz
T2	0.00 Hz
T3	90.00 Hz
T4	100.00 Hz
EXMCD	single_pulse_dec
INRUC	III
IFR	495.13 MHz
IRSET	4.98 kHz
IRF	0.94 Hz
IRRPW	92 usec
IRATN	79
DFE	KMU-2-286-1-cxx-AAB
LSKET	-601.50 kHz
LKF1N	-1.8 Hz
LKE1V	0
LGE1N	0
LKPHS	0
LKSGS	0
CPSPD	0 Hz
FILDC	
FILDF	

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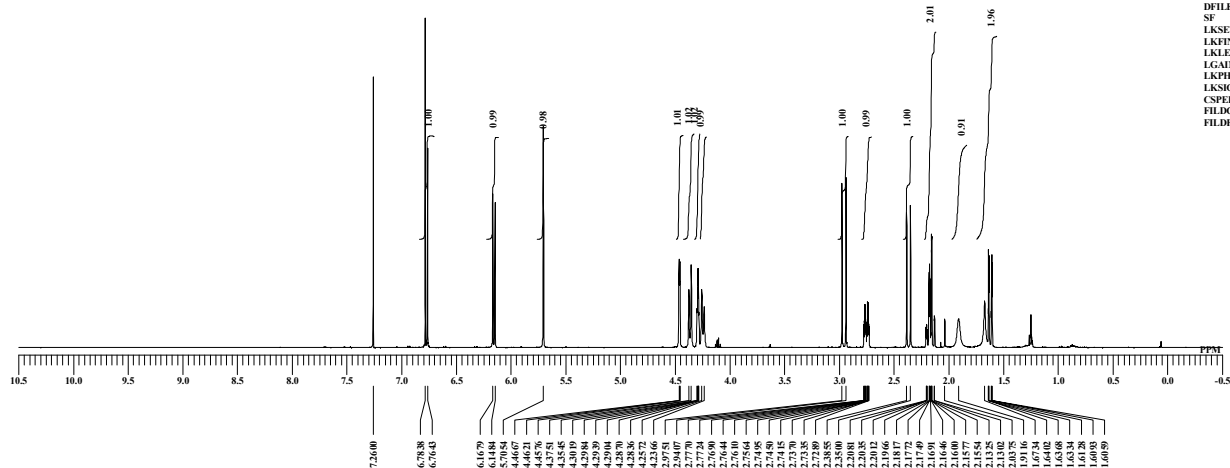


DATIM	10-10-2010 21:32:42
MENUP	
ONBUC	IH5
OFR	495.13 MHz
ORSET	4.38 KHz
ORFIN	9.64 Hz
PW1	6.00 usec
DE ADT	0.00 usec
PREDL	0.0000 MHz
IWT	1.0000 usec
POINT	13107
SPO	13107
TIMES	
DUMMY1	
FREQU	7429.31 Hz
FLT	38000 Hz
DE LAY	13.16 usec
ACQTM	1.7642 sec
PD	10.0000 sec
ADBIT	16
RGAIN	40
BF	0.01 Hz
T1	0.00 usec
T3	0.00 usec
T0	90.00 usec
T4	100.00 usec
EXTMOD	single_pulse_ext
EXPCM	
IRNVC	IH5
IFR	495.13 MHz
IRSET	4.38 KHz
IRFIN	9.64 Hz
IRBPW	92 usec
IRATN	79 usec
DFILE	KML-2-290-1-rct-ExA-ExB
LFSET	-601.50 KHz
LKFIN	-1.8 Hz
LKLEV	0 Hz
LGAIN	0 Hz
LKPUS	0 Hz
LKSG	0 Hz
CPSED	0 Hz
FILCD	

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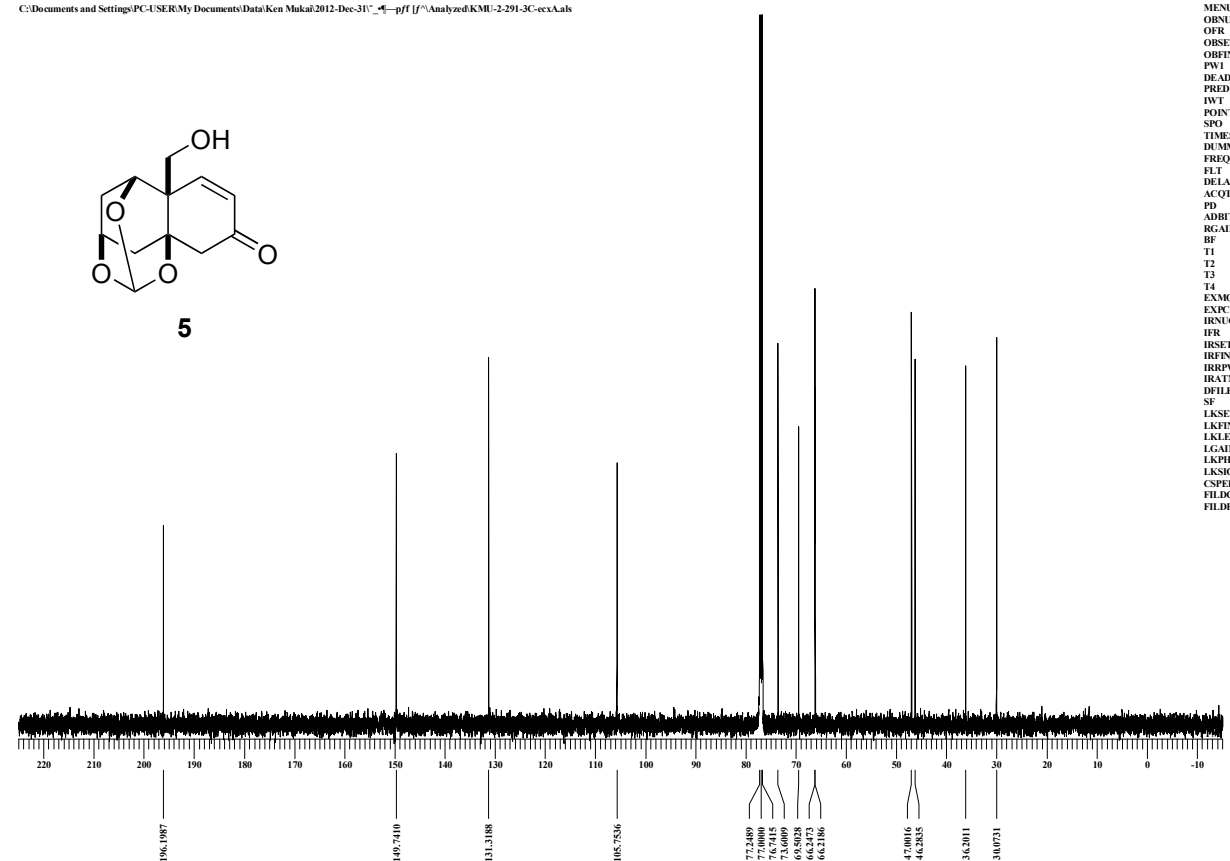
DATUM	10-10-20 12:15:17
MENUF	
OBNUC	1345
OFR	125.3 MHz
OBSET	145.35 kHz
OBFTN	6.25 usec
PW1	3.25 usec
DE-BIT	0.0000 usec
PRELD	0.00000 usec
IWT	1.0000 sec
POINT	26214
SPO	26214
TIMES	26214
DUMMY	4
FREQU	31429.52 Hz
FLT	157800 Hz
DELAY	20.00 usec
ACQTM	0.8389 sec
PBT	7.0000 sec
ADBIT	16
RGAIN	1.00
BF	1.00 Hz
PW1	1.00 usec
T2	0.0000
T3	90.00
T4	100.00
EXMOD	single_pulse_dec
IRNXC	III
IFR	495.13 Hz
IRSET	428.8 kHz
IFR	54.9 Hz
IRATN	92 usec
IRW	99
DFE	KMU-2-290-1C-ecs-A48
LSKET	-601.50 kHz
LKF1N	-1.8 Hz
LKE1V	0
LGC1N	0
LKPHS	0
LKSGS	0
CPSPD	0 Hz
FILDC	
FILDE	

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DATIM	12-10-2010 21:59:21
MENUF	IH
ONBUC	495
OFR	495.13 MHz
ORSET	4.38 kHz
ORFIN	9.64 Hz
PW1	6.00 msec
DE ADT	0.800 msec
PRDL1	0.0000 msec
IWT	1.0000 sec
POINT	13107
SPO	13107
TIMES	
DUMMY1	
FREQU	7429.31 Hz
FLT	38000 Hz
FLT1	13.16 msec
ACQTM	1.7642 sec
PD	10.0000 sec
ADBIT	16
RGAIN	50
BF	0.01
T1	0.00 Hz
T2	0.00
T3	00.00
T4	100.00
EXMOD	single_pulse_ext
EXPCM	
INVC1	IH
IFR	495.13 MHz
IRSET	4.38 kHz
IRFIN	9.64 Hz
IRPWF	92 msec
IRATN	79
DFLE	KML1-2-291.3-xxcAxs
LSKET	-601.50 KHz
LKSET	-1.8 Hz
LKLEV	0
LGAIN	0
LKPIN	0
LKSG	0
CPSED	0 Hz
FILDC	
T1 DE	

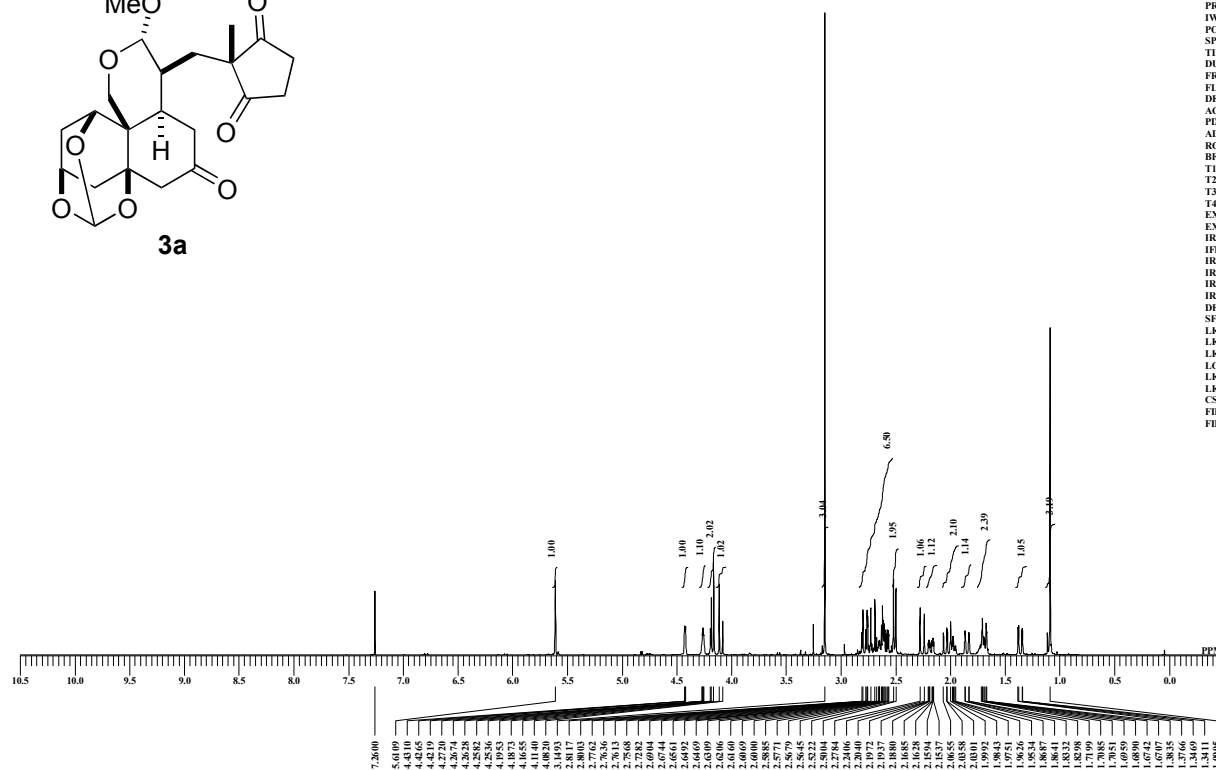
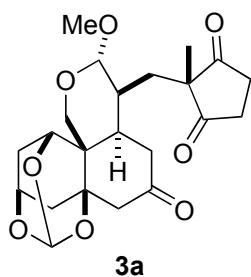
C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\2012-Dec-31\pff\pff\Analyzed\KMU-2-291-3C-ecx\A.als



DATIM	12-10-2010 23:34:52
MENUP	
OBNU/C	12.51
OFR	134.5 KHz
ORSET	0.0000 msec
PW1	3.25 usec
DEADT	0.0000 msec
PREDI	0.0000 msec
IWT	1.0000 sec
POINT	26214
SPO	26214
TIMES	512
DUMMY4	
FREQU	31429.52 KHz
FLI	187800 Hz
DELAY	20.000 usec
ACQTM	0.5300 sec
PD	10.0000 sec
ADBIT	16
RGAIN	50
BF	1.00 Hz
T1	0.00
T2	0.00
T3	90.00
T4	100.00
EXMOD	single_pulse_dec
EXPCM	
RNUC	1H
IRSET	495.13 KHz
IRFIR	4.83 KHz
IRFIR	9.64 Hz
IRWPR	92 msec
IRATN	79
DFLE	KMU-2-291.3-C-rcx-kab
LSKET	-601.50 KHz
LKFIR	-1.8 Hz
LKLEV	0
LKAIN	0
LKPHS	0
LKSGS	0
CSPED	0 Hz
FILDC	
TILDF	

KMU-10-74-1

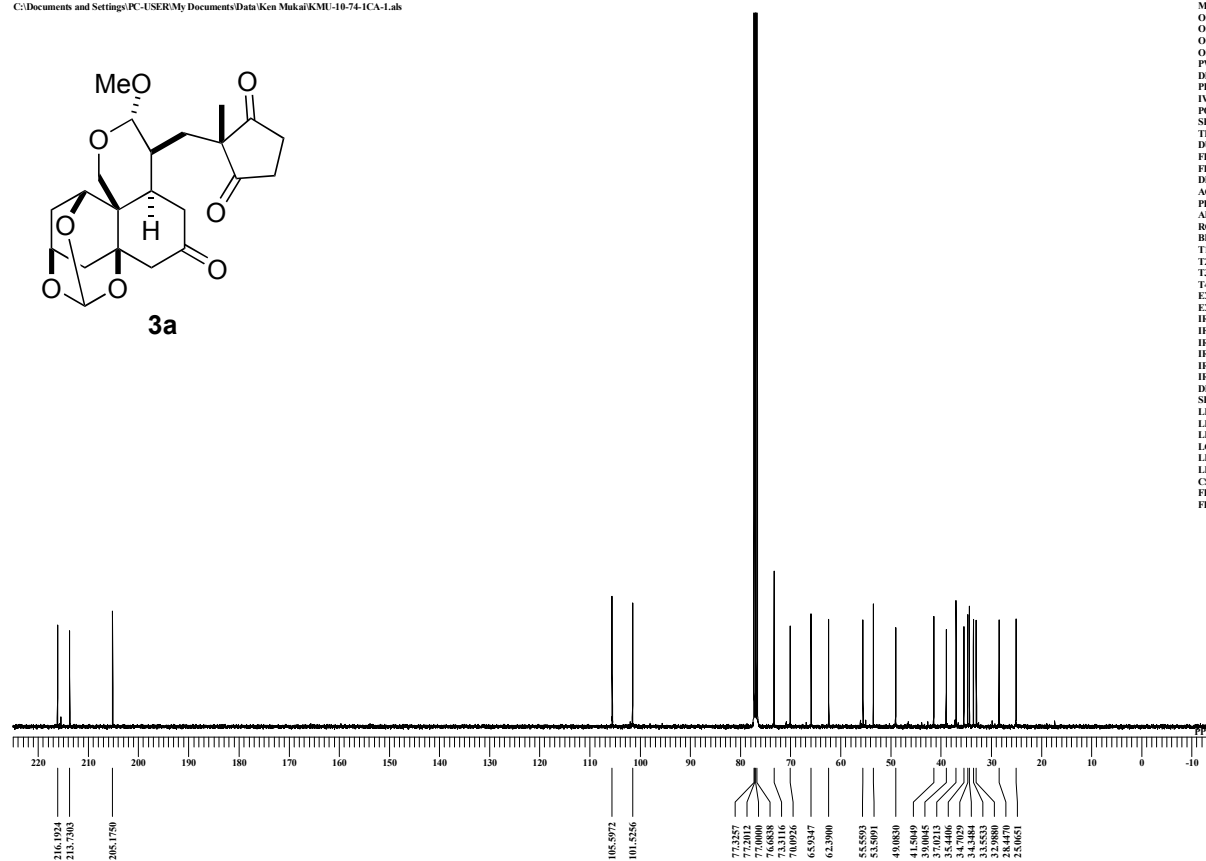
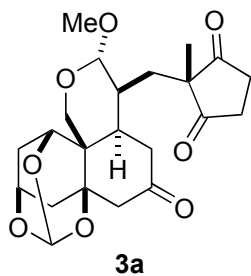
C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\KMU-10-74-1-1Aals



DATIM 19-01-2014 20:30:37
 MENUF
 OBNUC 1H
 OFR 395.88 MHz
 OBSET 6.28 KHz
 OBFIN 0.87 Hz
 PW1 6.44 usec
 DE ADT 0.00 usec
 PREDL 0.00000 msec
 IWT 1.0000 sec
 POINT 13107
 SPO 13107
 TIMES 16
 DUMMY 1
 FREQU 5938.15 Hz
 FLT 30000 Hz
 DELAY 16.68 usec
 ACQTM 2.2073 sec
 PD 10.0000 sec
 ADBIT 16
 RGAIN 30
 BF 0.01 Hz
 T1 0.00
 T2 0.00
 T3 100.00
 T4 100.00
 EXMOD single_pulse.cx2
 EXPCM
 IRNUC 1H
 IFR 395.88 MHz
 IRSET 6.28 KHz
 IRFIN 0.87 Hz
 IRRPW 115 usec
 IRATN 79
 DFILE KMU-10-74-1-1Aals
 SF 13.20 KHz
 LKSET 75.7 Hz
 LKFIN 75.7 Hz
 LKLEV 0
 LGAIN 0
 LKPHS 0
 LKSG 0
 CSPED 0 Hz
 FILDC
 FILDF

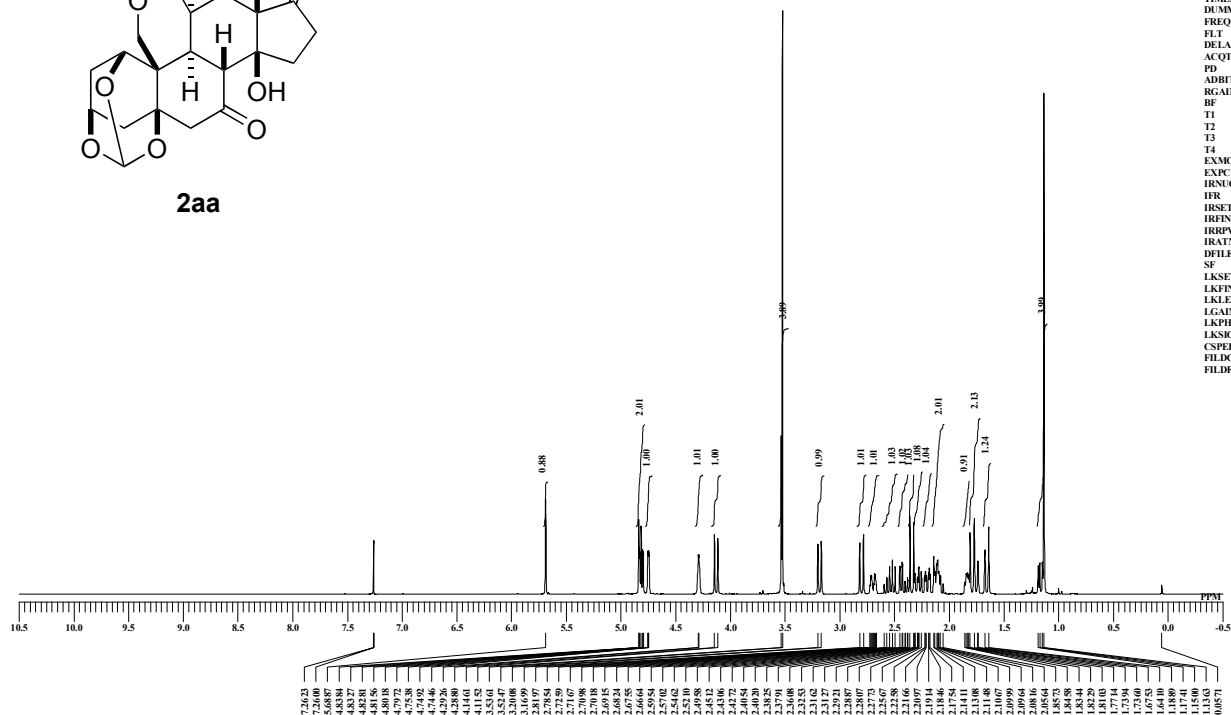
KMU-10-74-1C

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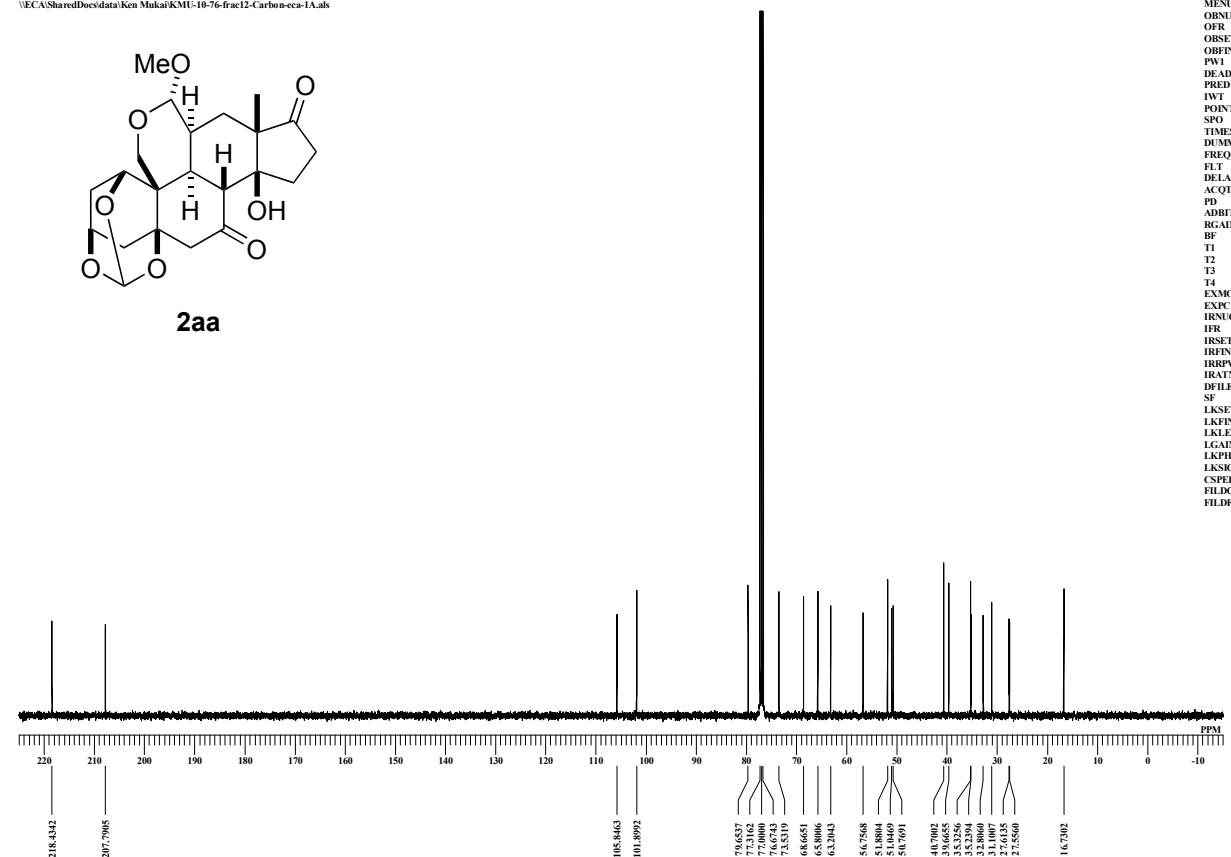
DATIM 20-01-2014 07:28:39
 MENUF
 OBNUC 13C
 OFR 99.55 MHz
 OBSET 5.13 KHz
 OBFIN 0.98 Hz
 PW1 3.03 usec
 DE ADT 0.00 usec
 PREDL 0.00000 msec
 IWT 1.0000 sec
 POINT 26214
 SPO 26214
 TIMES 1024
 DUMMY 4
 FREQU 24999.62 Hz
 FLT 125000 Hz
 DELAY 20.50 usec
 ACQTM 1.0486 sec
 PD 8.0000 sec
 ADBIT 16
 RGAIN 60
 BF 1.00 Hz
 T1 0.00
 T2 0.00
 T3 100.00
 T4 100.00
 EXMOD single_pulse_dec
 EXPCM
 IRNUC 1H
 IFR 395.88 MHz
 IRSET 6.28 KHz
 IRFIN 0.87 Hz
 IRRPW 115 usec
 IRATN 79
 DFILE KMU-10-74-1CA-1Aals
 SF 13.20 KHz
 LKSET 75.7 Hz
 LKFIN 75.7 Hz
 LKLEV 0
 LGAIN 0
 LKPHS 0
 LKSG 0
 CSPED 0 Hz
 FILDC
 FILDF

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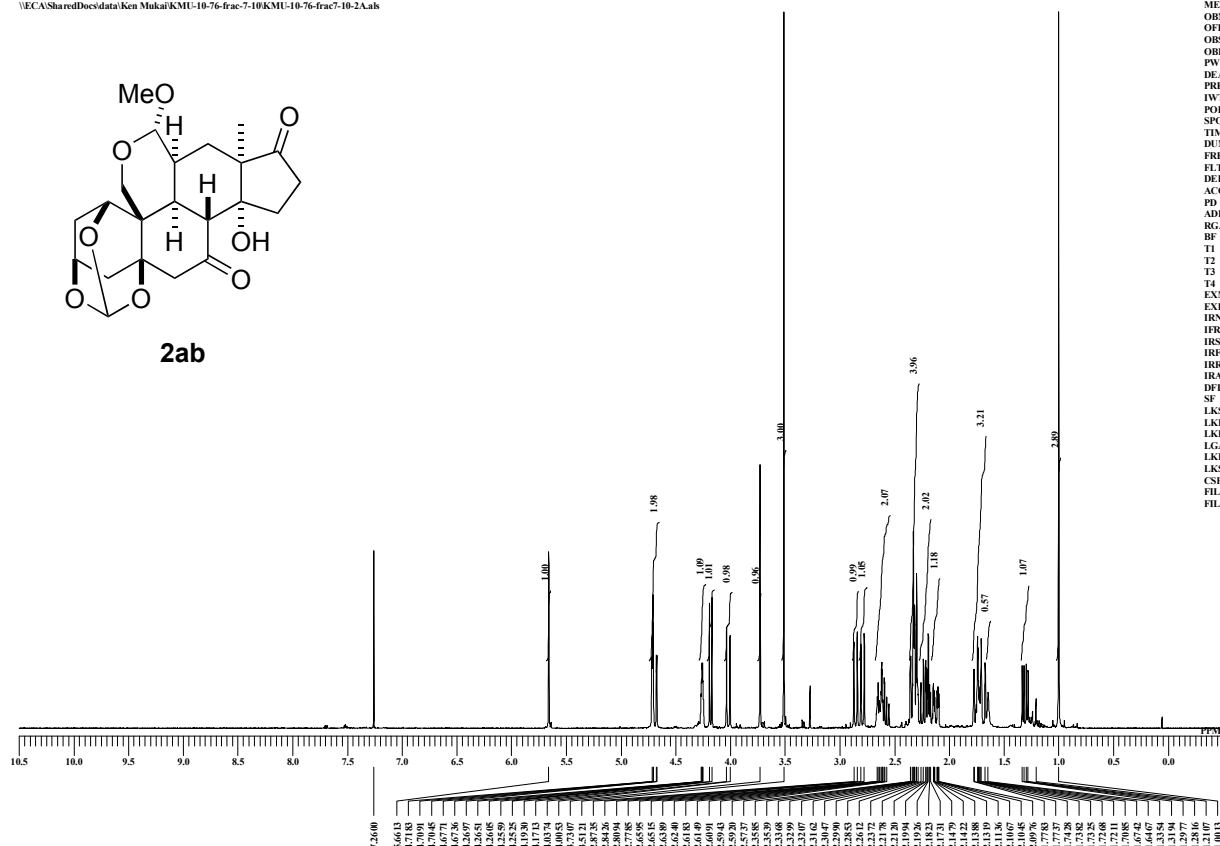
DATIM	06-02-2014 21:22:50
MENUP	
OBH C	
OFR	395.88 MHz
ORSET	6.38 kHz
OFBN	0.84 Hz
PW1	6.44 usec
DEADT	0.00 usec
PRDL	0.00 usec
IWT	1.0000 sec
POINT	13107
SPO	13107
TIMES	
DUMMY1	
FREQU	5938.15 Hz
FLT	3.2003 Hz
DELAY	10.00 usec
ACQTM	2.0075 sec
PD	2.0000 sec
ADBIT	16
RGAIN	22
BF	0.01 Hz
T1	0.00
T2	0.00
T3	100.00
T4	100.00
EXMOD	single_pulse.x2
EXPCM	
IRSET	395.88 MHz
IRFR	6.28 kHz
IRFN	0.87 Hz
IRPWP	6.44 usec
IRATN	7.99
DFILE	KMU-10-76-trace21.1.Aa
LKSET	13.20 kHz
LKFN	75.7 Hz
LKLEW	0.0
LKGIN	0.0
LKPTS	0.0
LKSGS	0.0
CSFED	0 Hz
FILDC	
TILDE	

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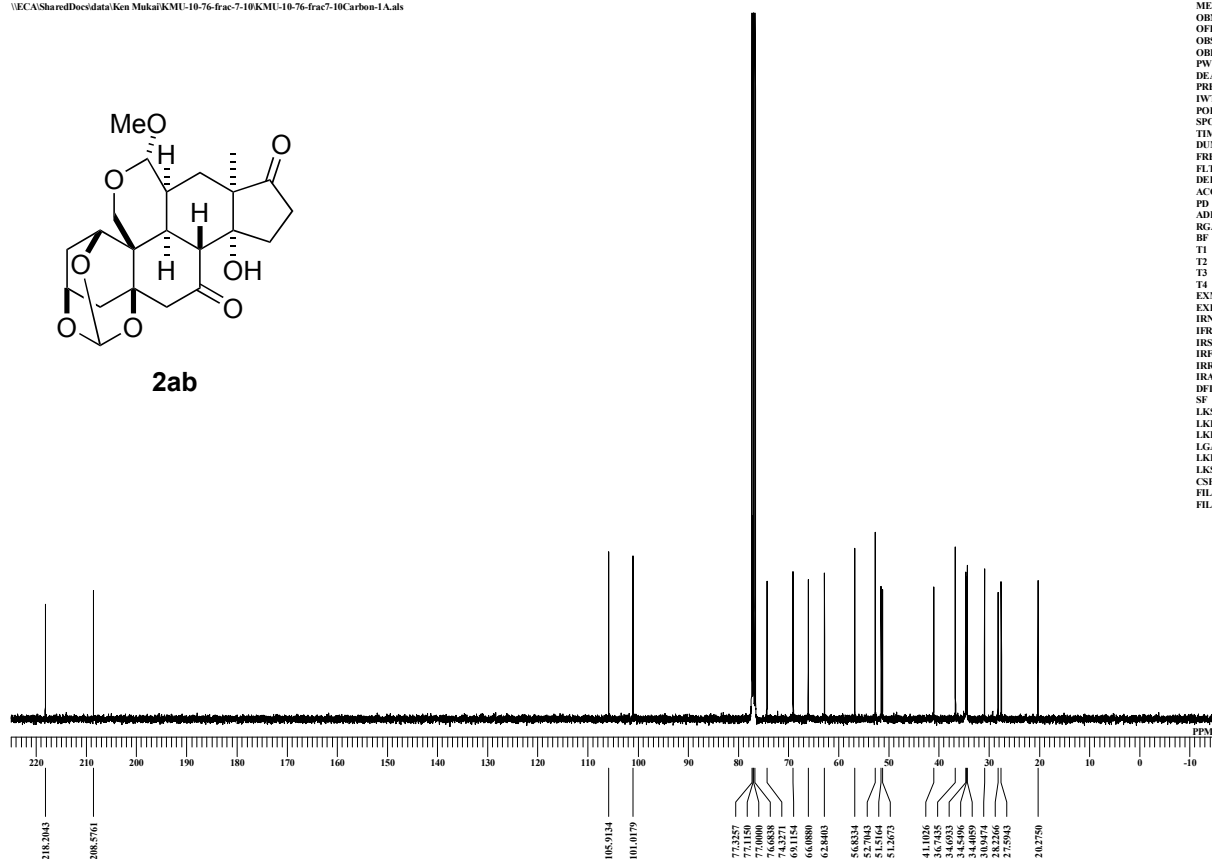
DATIM	06-02-2014 22:33:04
MENUP	
ORNC	13C
OFNU	99.55 MHz
OSNET	5.13 kHz
OFBIN	0.98 Hz
PW1	3.83 usec
DE-BIT	0.00 usec
PRECL	0.00000 msec
IWT	1.0000 sec
POINT	2624
SFO	26234
TIMES	5214
DUMMY	4
FREQ	24999.62 Hz
FLT	12500 Hz
DELAY	20.00 usec
ACQTM	1.0486 sec
PB	7.0000 sec
ADBIT	
RG-AIN	58
BF	1.00 10Hz
T1	0.00
T2	0.00
T3	100.00
T4	100.00
EXMCD	single_pulse_dec
ENMOD	
IRNVC	III
IFR	395.88 MHz
IRSET	6.28 kHz
IFRIN	0.00 Hz
IRHPV	115 usec
IRATN	7.99
DFLE	KMU-10176-fract2-Carbo
LSKET	13.20 kHz
LKFIN	75.7 Hz
LKEIV	0
LKEIN	0
LKPHS	0
LKSGS	0
CFSPD	0 Hz
FILFD	
FILFD	

\\ECA\SharedDocs\data\Ken Muka\KMU-10-76-frac-7-10\KMU-10-76-frac7-10-2A.xls



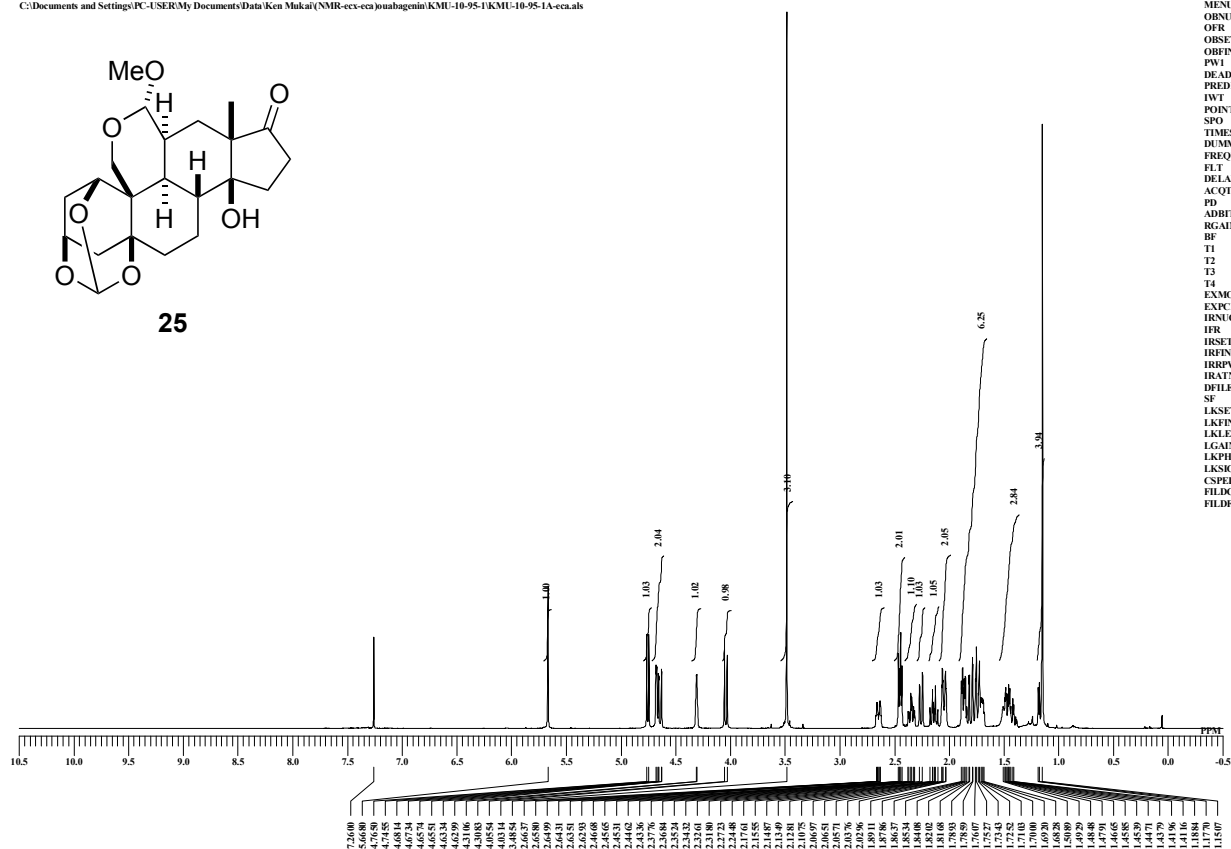
DATIM	02-02-2014 21:46:02
MENUP	OBUC 1H
OFNR	395.88 MHz
IRSET	6.28 KHz
OFBN	0.87 Hz
PW1	6.44 usec
DEADT	0.000 usec
PREDL	0.000 usec
IWT	1.000 sec
POINT	13107
SPO	13107
LMES	8
DUMMY	1
FREQU	5938.15 Hz
FLT	2.000 Hz
DELAY	16.000 usec
ACQTM	2.2073 sec
PD	10.0000 sec
ADBIT	16
RGAIN	24
BF	0.01 Hz
T1	0.00
T2	0.00
T3	100.00
T4	100.00
EXMOD	single_pulse.e2
EXPCM	OBUC 1H
IFR	395.88 MHz
IRSET	6.28 KHz
IRFN	0.87 Hz
IRBPW	5.79 usec
IRATN	7.57
DFILE	KMU-10-76-trac7-10-2A..S
LFKST	13.20 KHz
LFKFN	75.7 Hz
LFKEV	0
LGAIN	0
LKFHS	0
LKSGS	0
CSPED	0 Hz
FILDC	
FILDF	

\\ECA\SharedDocs\data\Ken Mukai\KMU-10-76-frac-7-10\KMU-10-76-frac7-10Carbon-1A.xls



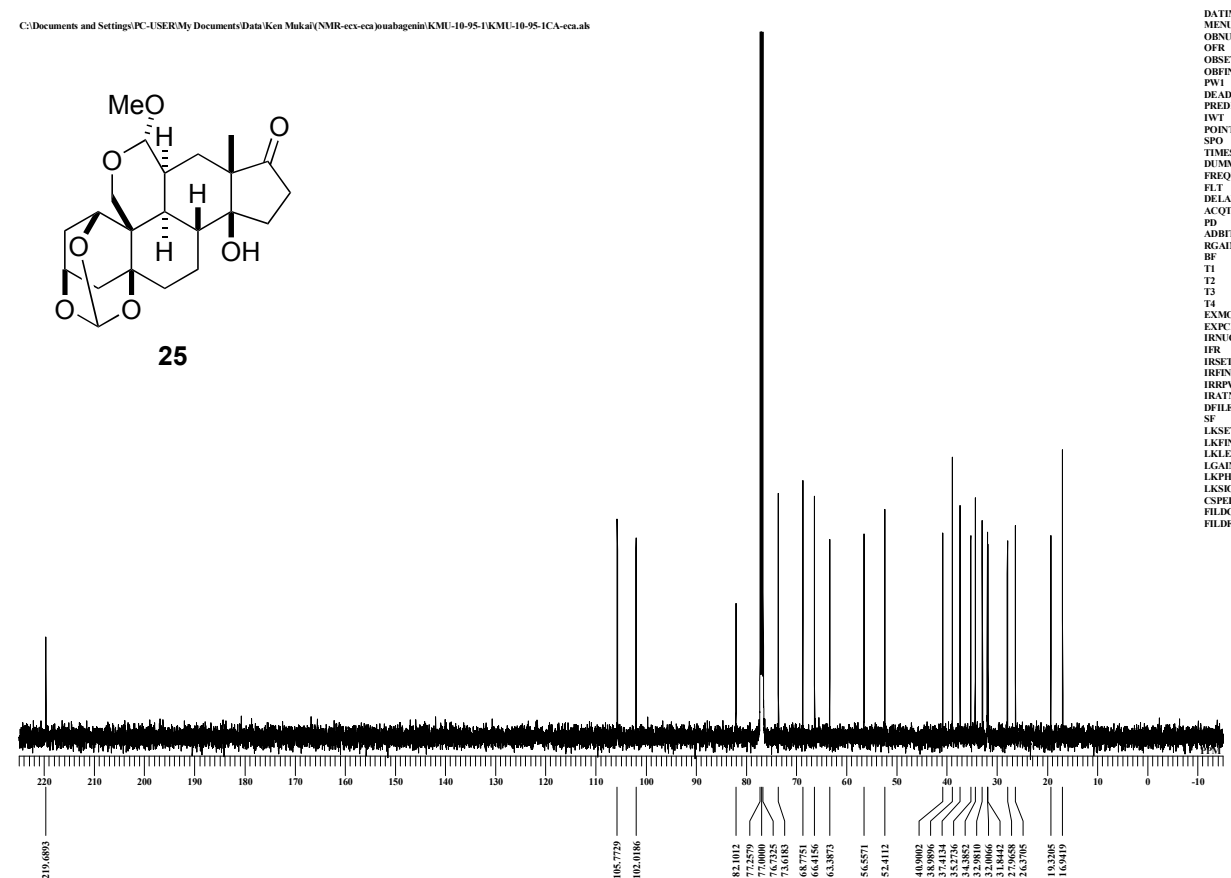
DATIM	03-02-2014 06:49:11
MENUP	
OBNU	13C
OFRC	99.55 MHz
ORSET	5.13 kHz
OFBN	0.98 Hz
PW1	3.03 usec
DEADT	
PREL1	0.000 usec
IWT	1.000 sec
POINT	26214
SPO	26214
LMES	1024
DUMMY	4
FREQU	24999.62 MHz
FLT	1.2500 Hz
DELAY	20.00 usec
ACQTM	14846 sec
PD	7.0000 sec
ADBIT	16
RGAIN	60
BF	1.00 Hz
T1	0.00
T2	0.00
T3	100.00
T4	100.00
EXMOD	single_pulse_dec
EXPCM	
IRNUC	IH
IFR	39.58 MHz
IRSET	62.82 kHz
IRFN	0.87 Hz
IRBPW	7.7
IRATN	5.7
DLFE	KMU-10.76-7rac-10Car
LFKST	13.20 kHz
LFKFN	75.7 Hz
LKLEV	0
LGAIN	0
LKFHS	0
LKSGS	0
CSPED	0 Hz
FILDC	
FILDF	

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DATIM	2014-04-07 20:50:08
MENUF	
OBNUC	IH
OFR	490.15 MHz
ORSET	9.16 KHz
ORBIN	7.60 Hz
PW1	6.00 msec
DE ADT	0.00 msec
PDEL1	13.52 msec
IWT	1.0000 sec
POINT	13107
SPO	13107
TIMES	16
DUMMY1	16
FREQU	7352.83 KHz
FLT	37000 Hz
DE LAY	13.52 msec
ACQTM	1.7826 sec
PD	10.0000 sec
ADBIT	16
RGAIN	36
BF	0.01 Hz
T1	0.00 msec
T3	0.00 msec
T4	100.00 msec
EXMOD	single_pulse
EXPCM	
IRNUC1	IH
IFR	490.15 MHz
IRSET	9.16 KHz
IRFIN	7.60 msec
IRBPW	6.00 msec
IRATN	4
DFILE	KML-10-95-1A-eacab5
LKSET	70.30 KHz
LKLET	32.5 Hz
LKLEV	0
LKGAIN	0
LKSPIN	0
LKSG	0
CSPD	0 Hz
FILDE	

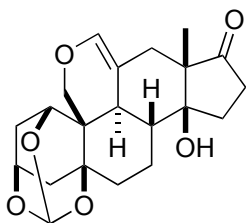
C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\NMR-ecx-eca\ouabagenin\KMU-10-95-1\KMU-10-95-1CA-eca.als



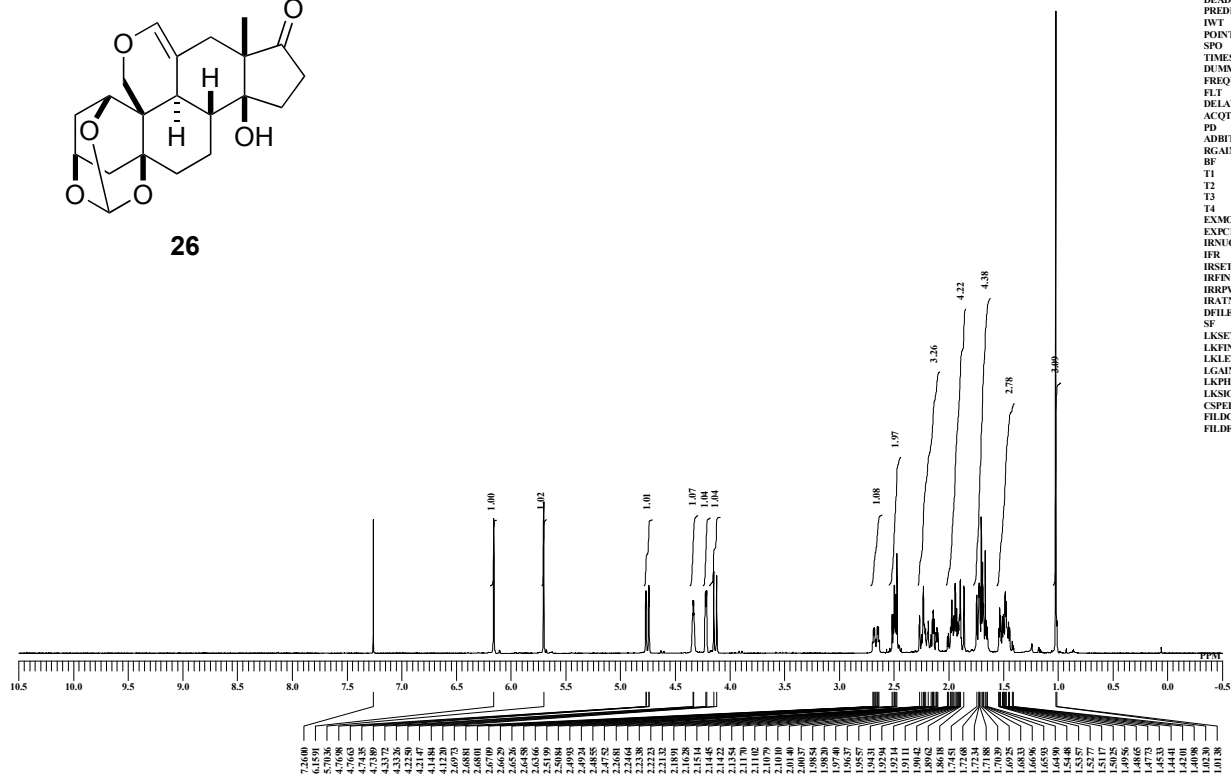
DATIM	2014-04-07 23:57:51
MENUP	
OBNUC	132.16 Hz
OFB	123.21 MHz
OBSET	6.71 Hz
OFBFIN	6.71 Hz
PW1	3.88 usec
DE-BIT	0.000 usec
PREDL	0.0000 usec
IWT	1.0000 usec
POINT	2612
SPO	2624
TIMES	5124
DUMMY4	
FREQU	30863.73 Hz
FLT	155000 Hz
DELAY	2.100 usec
ACQTM	0.8493 sec
PDB	7.0000 sec
ADIT	
RGAIN	60
BF	1.00 Hz
T1	0.00 Hz
T2	0.00 Hz
T3	90.00 Hz
T4	100.00 Hz
EXMCD	single_pulse_dec
INRUC	III
IFR	490.15 MHz
IRSET	9.916 KHz
IFR1	2.60 Hz
IRRPW	92. usec
IRATN	4
SFE	KMU-10-95-ICA-eaakb
DL	
LKSET	70.30 KHz
LKFIN	32.5 Hz
LKLEV	0
LKGIN	0 KHz
LKPHS	0
LKSGS	0
CPSPD	0 Hz
FILDC	
FILDF	

KMU-10-96-1-ecs

\\ECS\SharedDocs\data\Ken Makai\KMU-10-96-1\KMU-10-96-1A-ecs-2ab



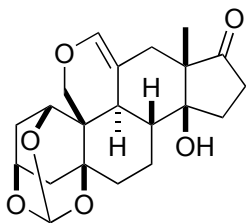
26



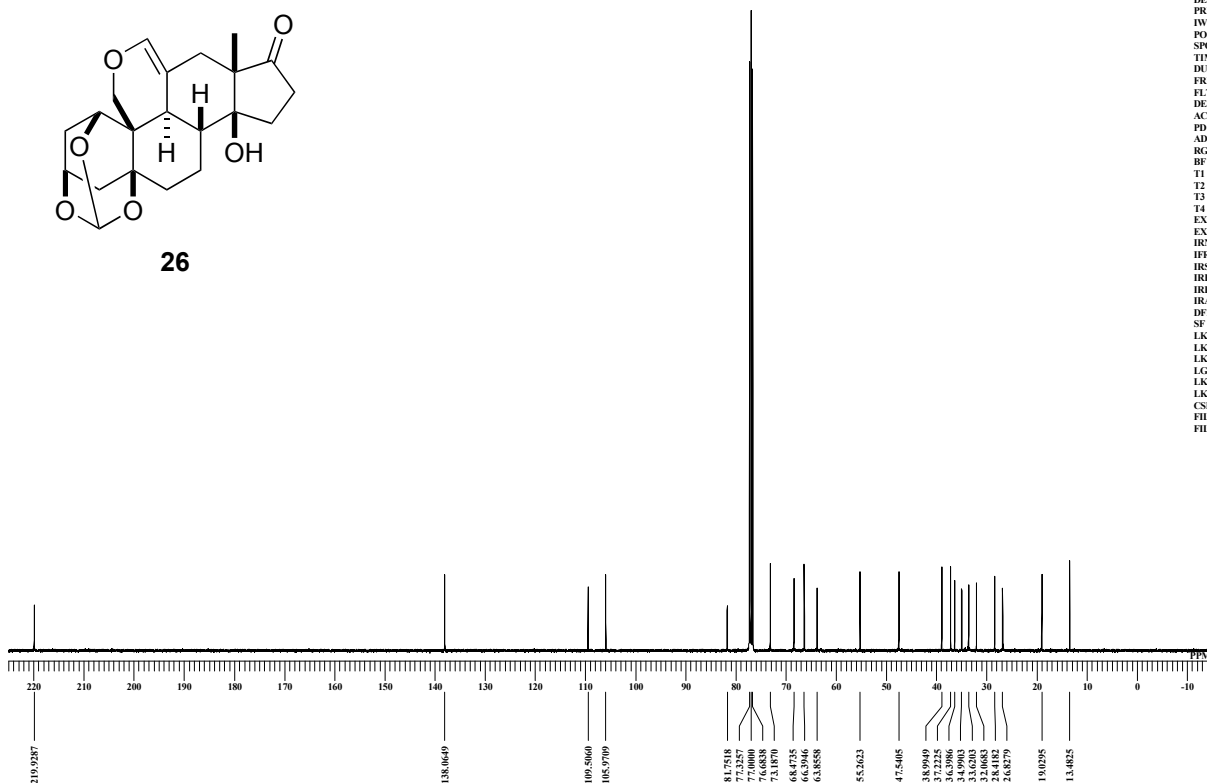
DATIM 11-04-2014 21:05:14
 MENUF
 OBNUC 1H
 OFR 395.88 MHz
 OBSET 6.28 KHz
 OBFIN 0.87 Hz
 PW1 6.44 usec
 DEADT 0.00 usec
 PREDL 0.00000 msec
 IWT 1.0000 sec
 POINT 13107
 SFO 13107
 TIMES 16
 DUMMY 1
 FREQU 5938.15 Hz
 FLT 30000 Hz
 DELAY 16.68 usec
 ACQTM 2.2073 sec
 PD 10.0000 sec
 ADBIT 16
 RGAIN 34
 BF 6.10 Hz
 T1 0.00
 T2 0.00
 T3 100.00
 T4 100.00
 EXMOD single_pulse.ex2
 EXPCM
 RNUC 1H
 FR 395.88 MHz
 IRSET 6.28 KHz
 IRFIN 0.87 Hz
 IRRPW 115 usec
 IRATN 79
 DFIL KMU-10-96-1A-ecs-2ab
 SF
 LKSET 13.20 KHz
 LKFIN 75.7 Hz
 LKLEV 0
 LGAIN 0
 LKFIN 0
 LKSIG 0
 CSPED 0 Hz
 FILDC
 FILDF

KMU-10-96-1C-ecs

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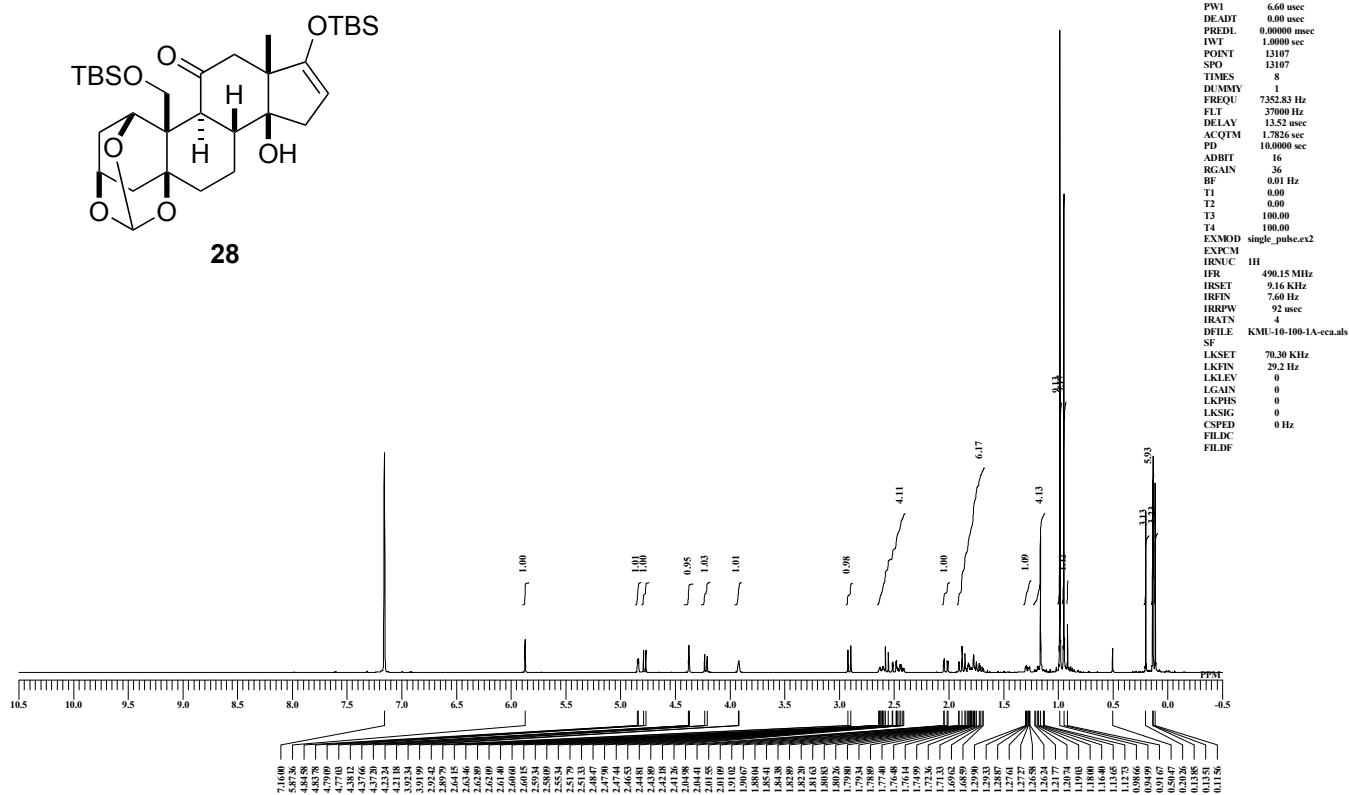
26



DATIM 12-04-2014 07:26:57
 MENUF
 OBNUC 13C
 OFR 99.55 MHz
 OBSET 5.13 KHz
 OBFIN 0.98 Hz
 PW1 3.03 usec
 DEADT 0.00 usec
 PREDL 0.00000 msec
 IWT 1.0000 sec
 POINT 26214
 SFO 26214
 TIMES 1024
 DUMMY 4
 FREQU 24999.62 Hz
 FLT 125000 Hz
 DELAY 20.50 usec
 ACQTM 1.0486 sec
 PD 7.0000 sec
 ADBIT 16
 RGAIN 60
 BF 1.00 Hz
 T1 0.00
 T2 0.00
 T3 100.00
 T4 100.00
 EXMOD single_pulse_dec
 EXPCM
 RNUC 1H
 FR 395.88 MHz
 IRSET 6.28 KHz
 IRFIN 0.87 Hz
 IRRPW 115 usec
 IRATN 79
 DFIL KMU-10-96-1C-ecs-Lab
 SF
 LKSET 13.20 KHz
 LKFIN 75.7 Hz
 LKLEV 0
 LGAIN 0
 LKFIN 0
 LKSIG 0
 CSPED 0 Hz
 FILDC
 FILDF

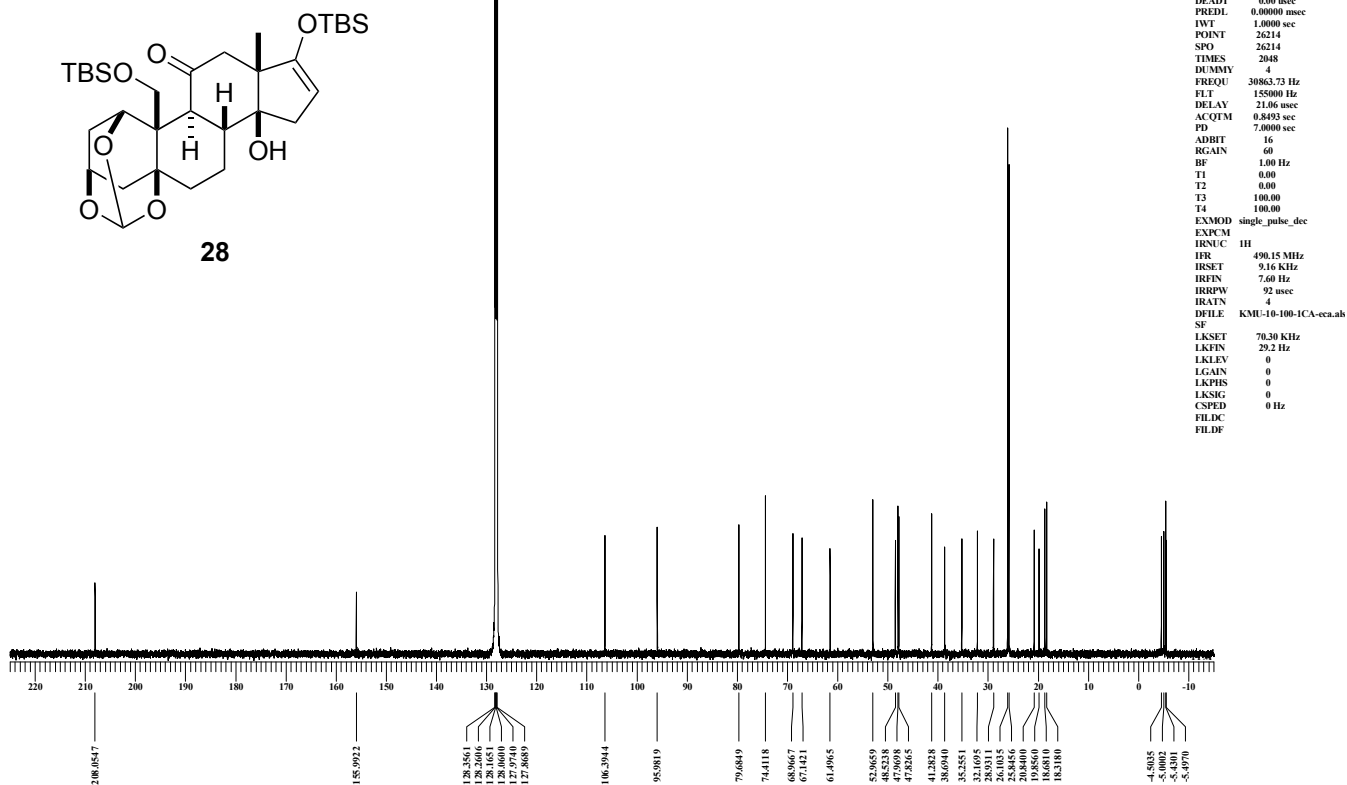
KMU-10-99-1-eca

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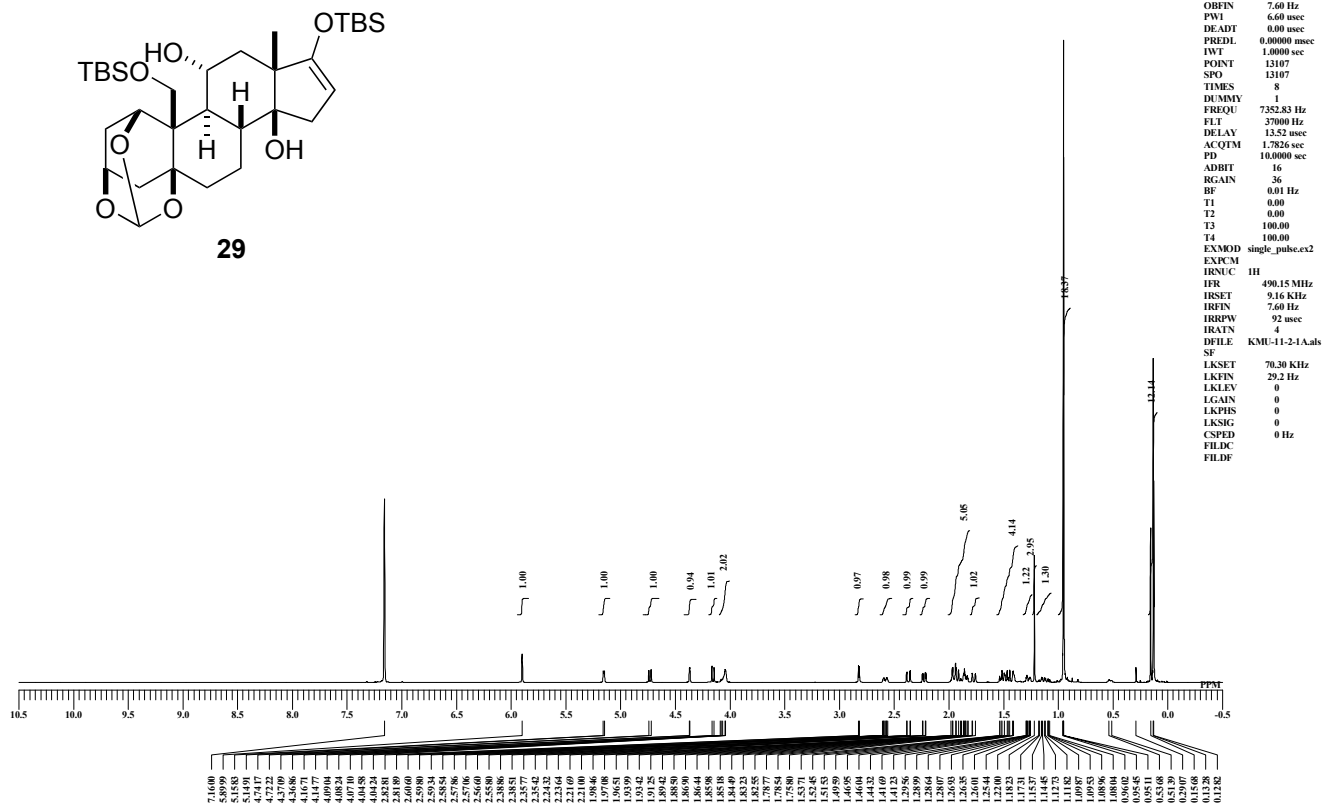
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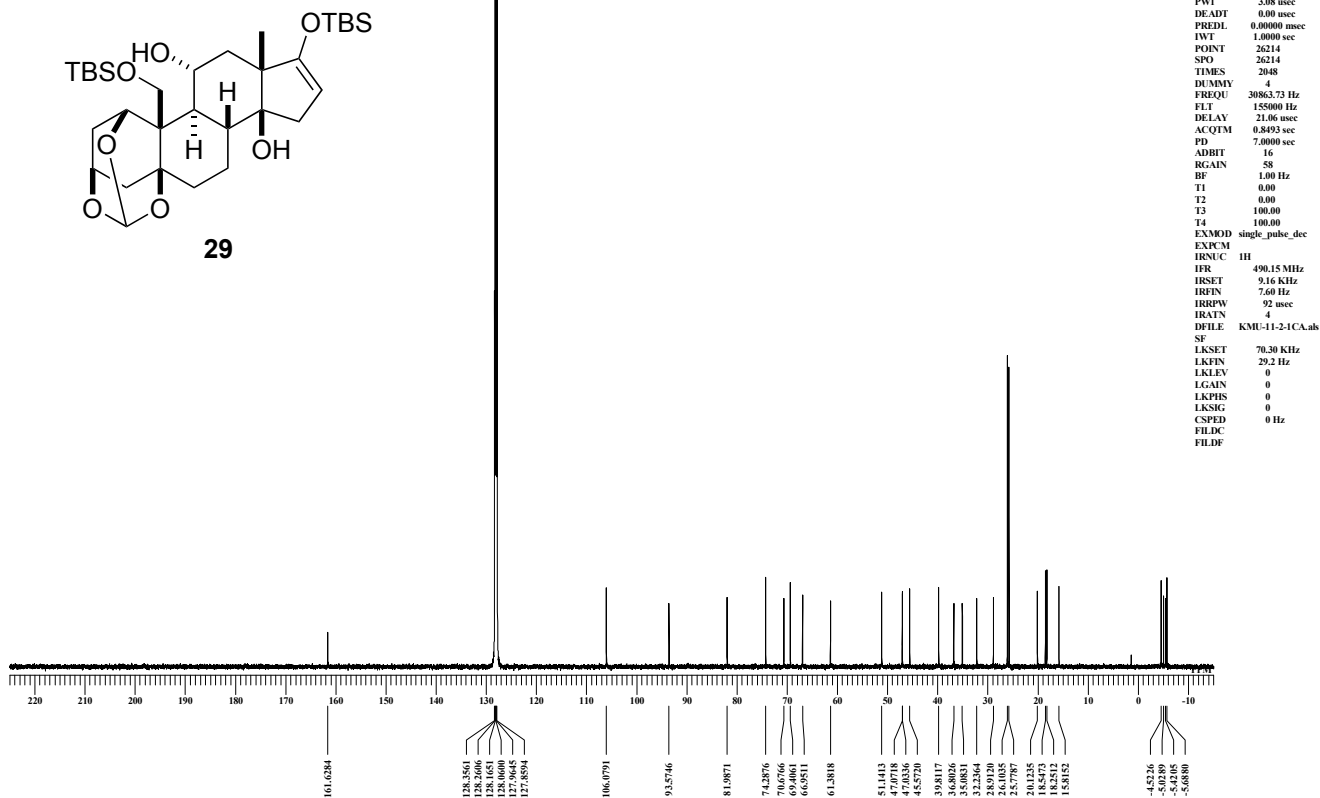
KMU-11-2-1

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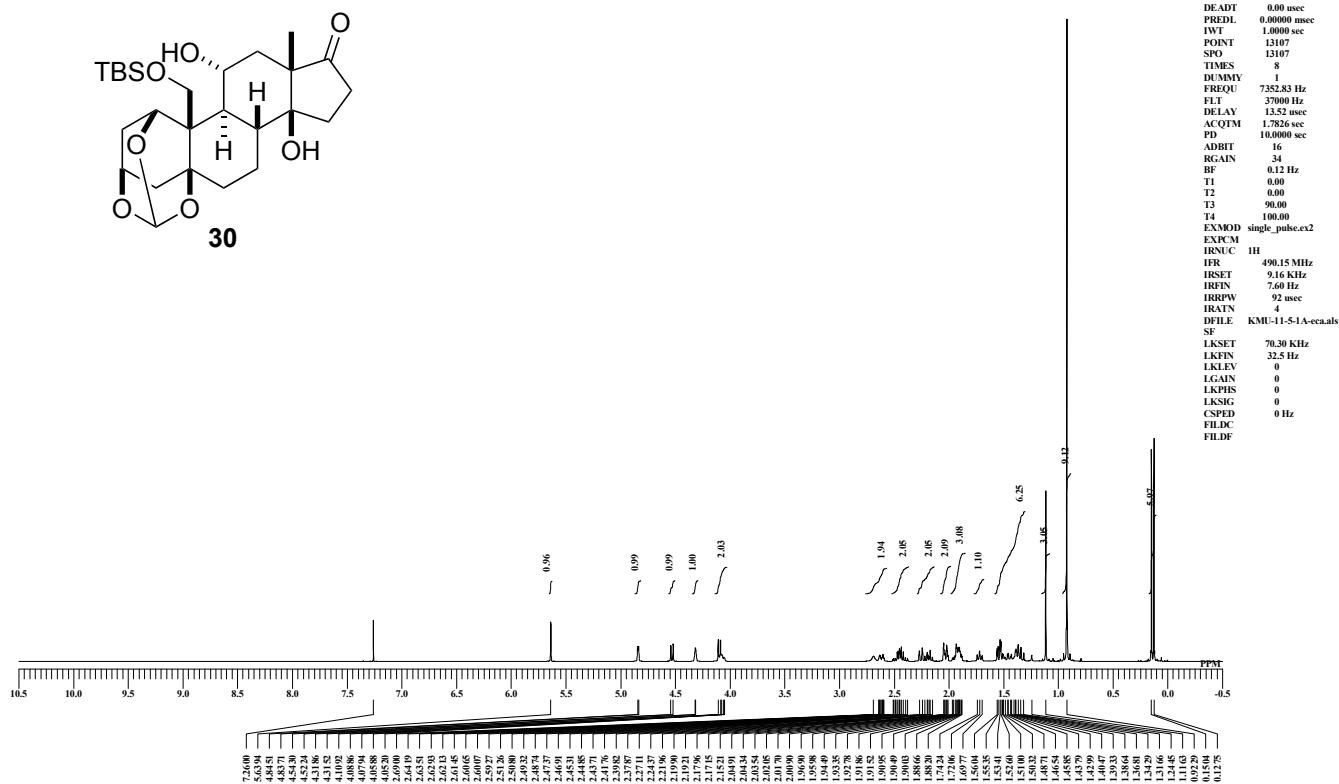
KMU-11-2-1C

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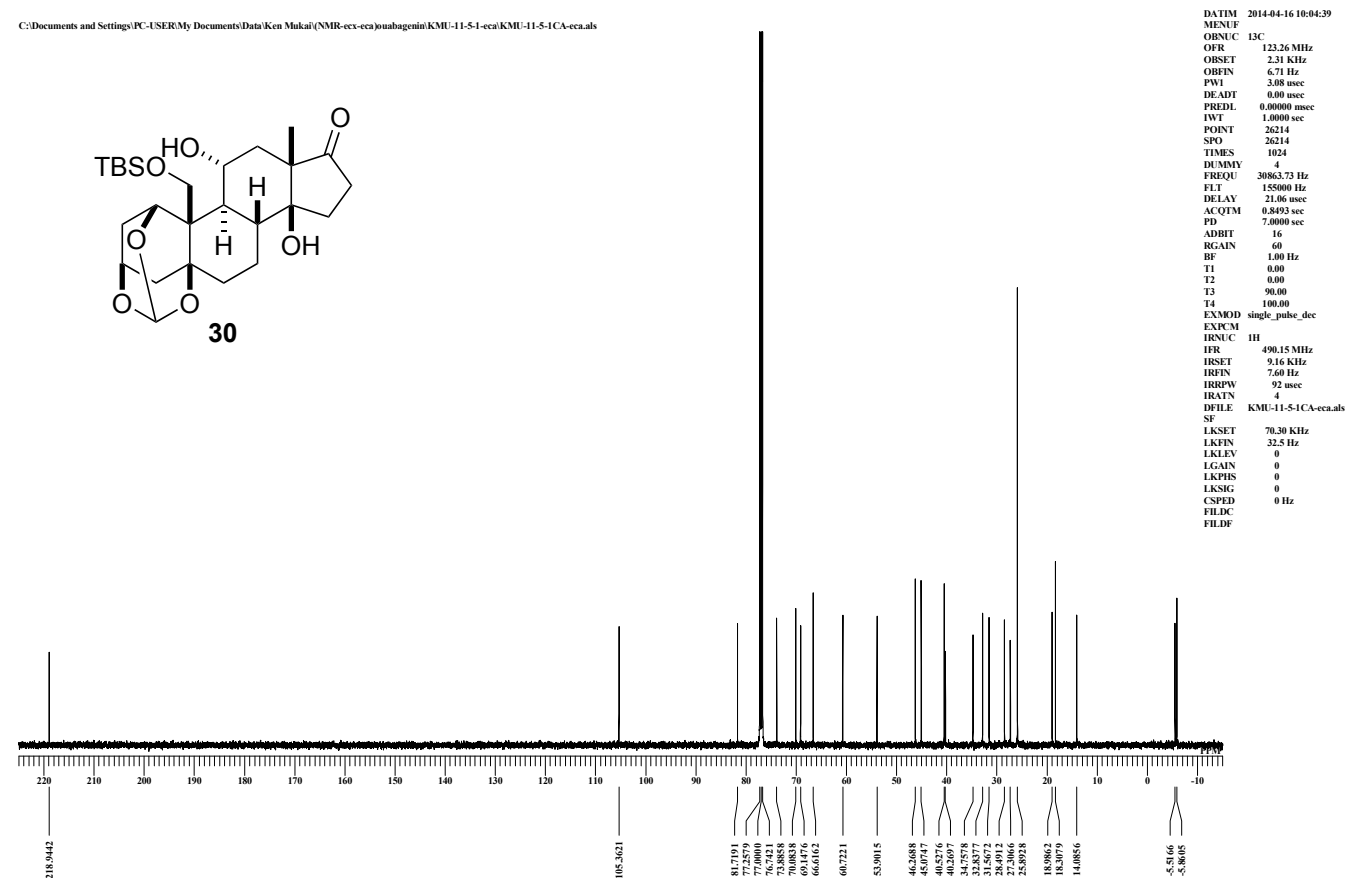
KMU-11-5-1-eca

C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\NMR-ecx-eca\joubagenio\KMU-11-5-1-eca\KMU-11-5-1-A-eca.als



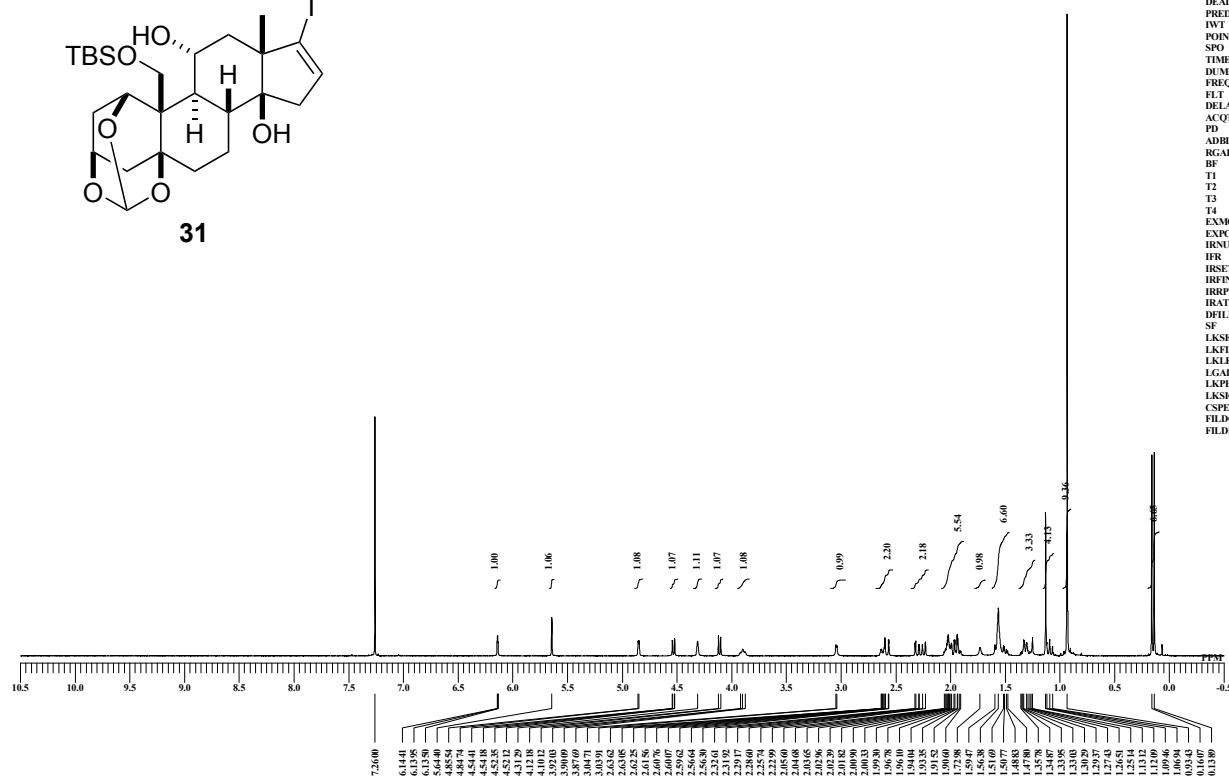
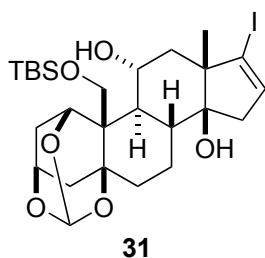
KMU-11-5-1C-eca

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KMU-11-6-1-eca

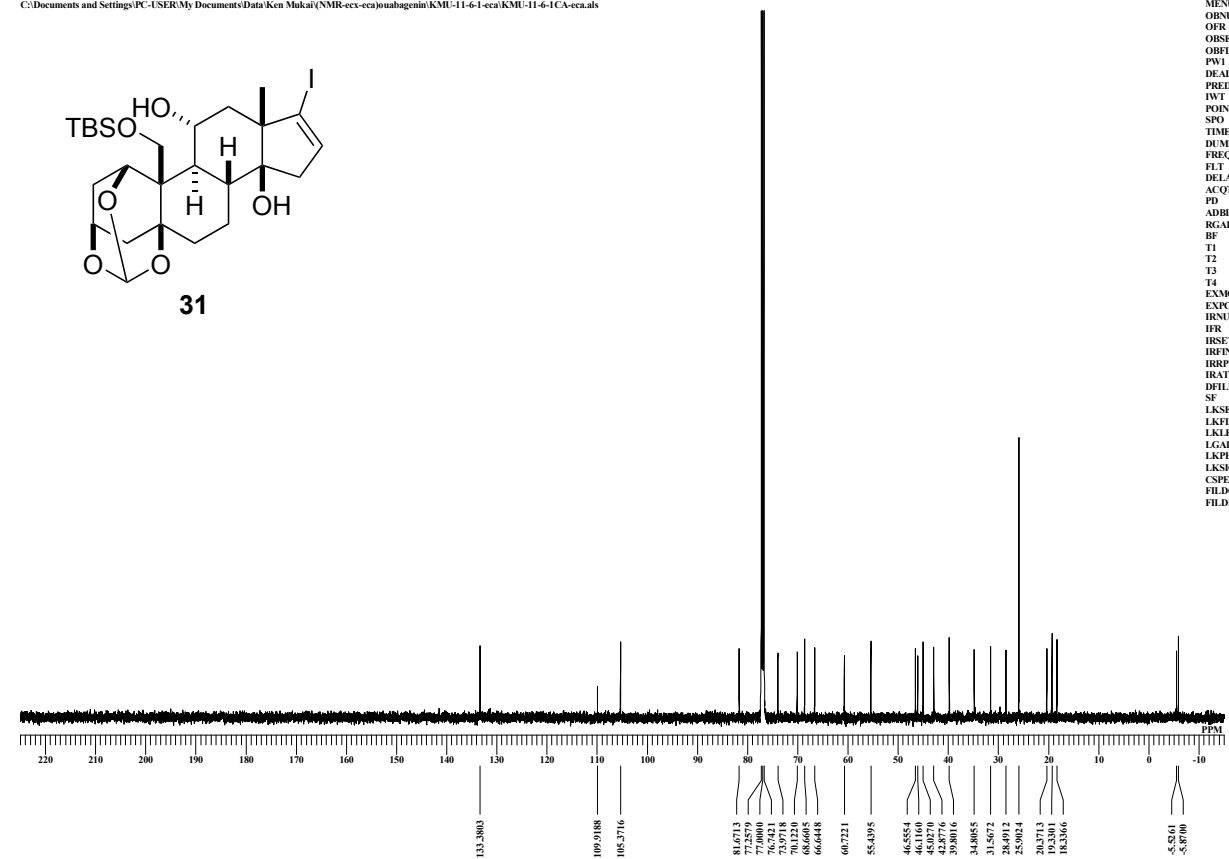
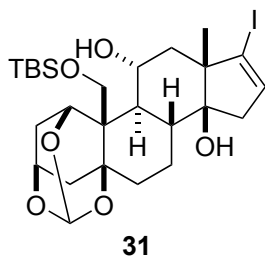
C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\NMR-ecx-eca\ouahagenin\KMU-11-6-1-eca\KMU-11-6-1-ecaA-for-data.als



DATIM 17-04-2014 15:04:11
 MENUF
 OBNUC 1H
 OFR 490.15 MHz
 OBSET 9.16 KHz
 OBFN 7.60 Hz
 PW1 6.60 usec
 DEADT 0.00 usec
 PREDL 0.00000 msec
 IWT 1.0000 sec
 POINT 13107
 SPO 13107
 TIMES 16
 DUMMY 1
 FREQU 7352.83 Hz
 FLT 37000 Hz
 DELAY 13.52 usec
 ACQTM 1.7826 sec
 PD 10.0000 sec
 ADBT 16
 RGAIN 40
 BF 0.01 Hz
 T1 0.00
 T2 0.00
 T3 90.00
 T4 100.00
 EXMOD single_pulse.ex2
 EXPCM
 IENUC 1H
 IR 490.15 MHz
 IRSET 9.16 KHz
 IRFN 7.60 Hz
 IRFPW 118 usec
 IRATN 79
 DFLE KMU-11-6-1-ecaA-for-da
 SF
 LKSET 70.30 KHz
 LKFN 32.5 Hz
 LKLEV 0
 LGAIN 0
 LKPS 0
 LKSG 0
 CSPED 0 Hz
 FLDC
 FILDF

KMU-11-6-1C-eca

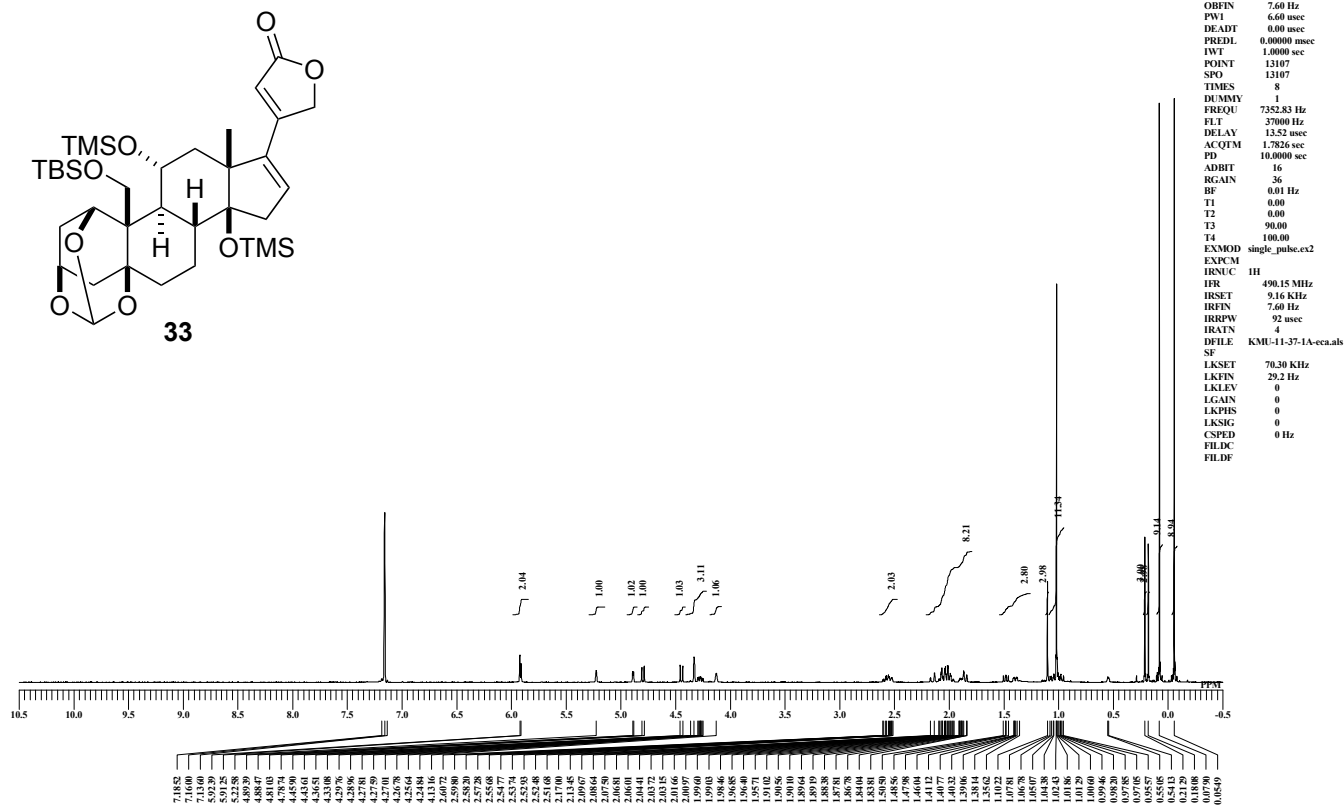
C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\NMR-ecx-eca\ouahagenin\KMU-11-6-1-eca\KMU-11-6-1CA-eca.als



DATIM 2014-04-17 01:46:22
 MENUF
 OBNUC 13C
 OFR 123.26 MHz
 OBSET 2.31 Hz
 OBFN 6.71 Hz
 PW1 3.08 usec
 DEADT 0.00 usec
 PREDL 0.00000 msec
 IWT 1.0000 sec
 POINT 26214
 SPO 26214
 TIMES 1024
 DUMMY 4
 FREQU 30863.73 Hz
 FLT 155000 Hz
 DELAY 21.06 usec
 ACQTM 0.8493 sec
 PD 7.0000 sec
 ADBT 16
 RGAIN 56
 BF 1.90 Hz
 T1 0.00
 T2 0.00
 T3 90.00
 T4 100.00
 EXMOD single_pulse_dec
 EXPCM
 IENUC 1H
 IR 490.15 MHz
 IRSET 9.16 KHz
 IRFN 7.60 Hz
 IRFPW 92 usec
 IRATN 4
 DFLE KMU-11-6-1CA-eca.als
 SF
 LKSET 70.30 KHz
 LKFN 32.5 Hz
 LKLEV 0
 LGAIN 0
 LKPS 0
 LKSG 0
 CSPED 0 Hz
 FLDC
 FILDF

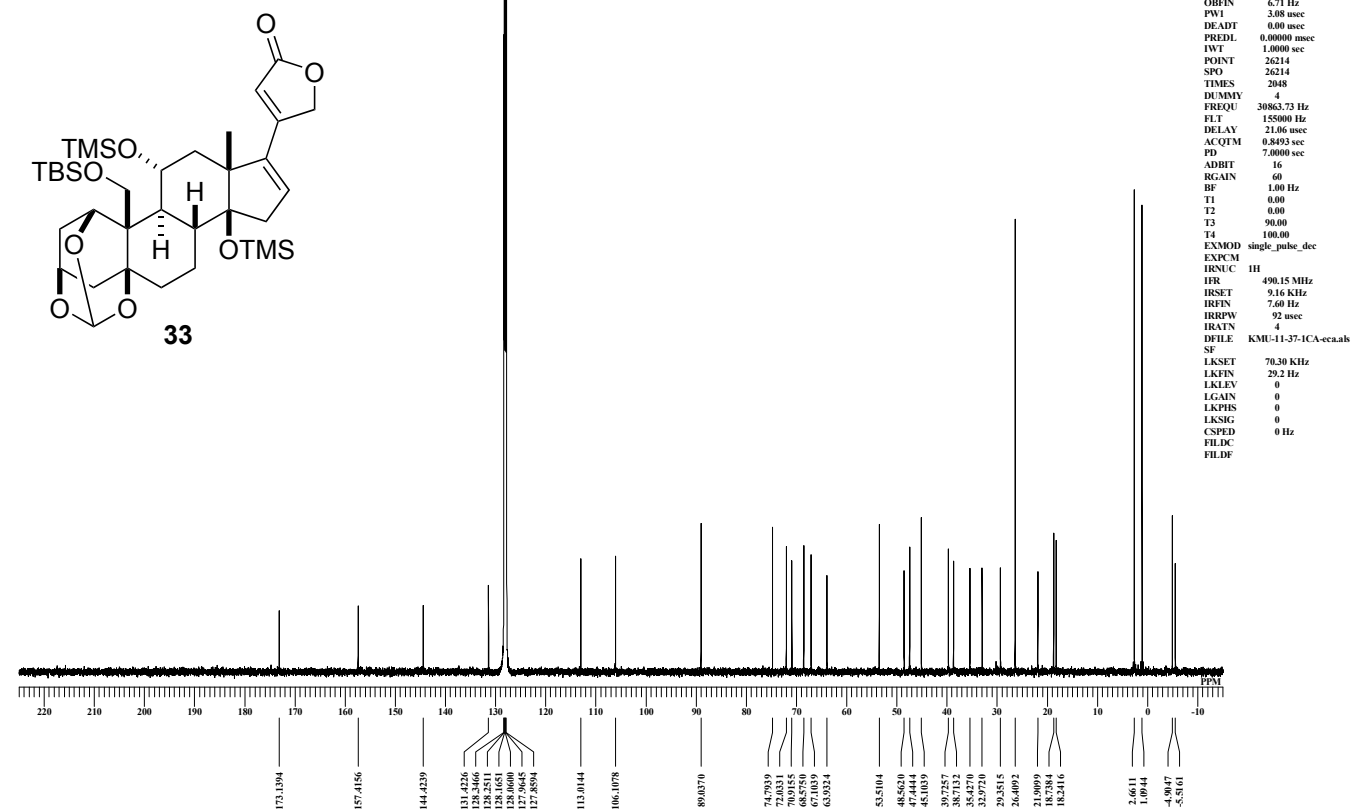
KMU-11-37-1-eca

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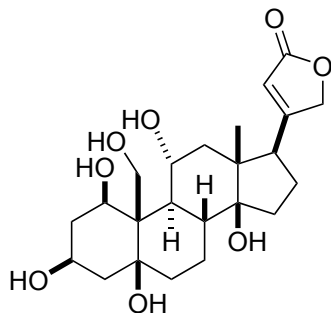
KMU-11-37-1C-eca

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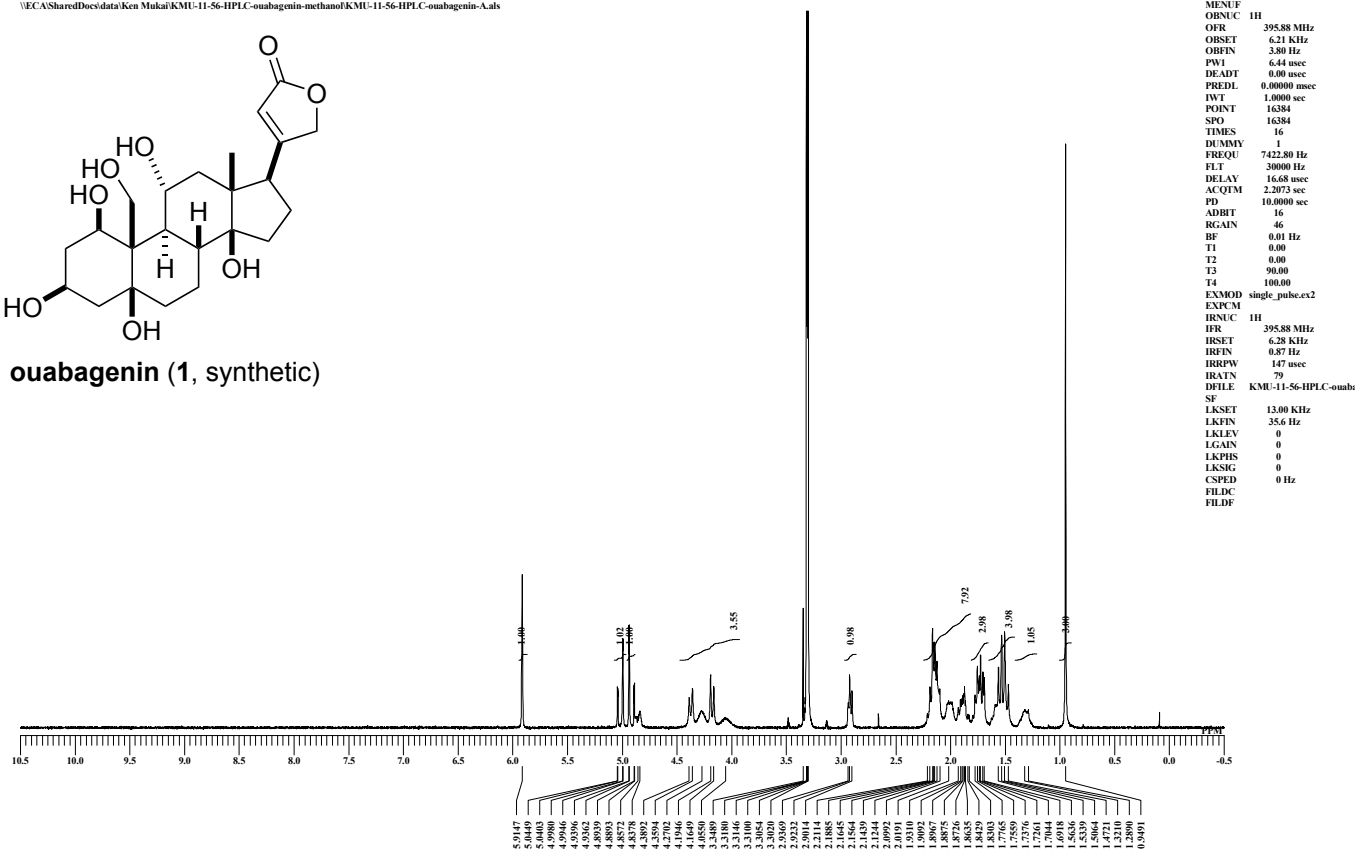


KMU-11-56-HPLC-ouabagenin

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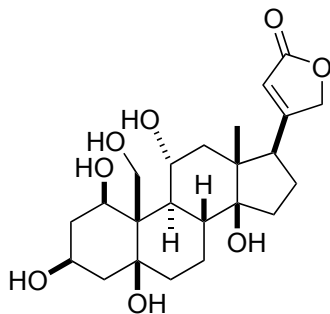


ouabagenin (1, synthetic)

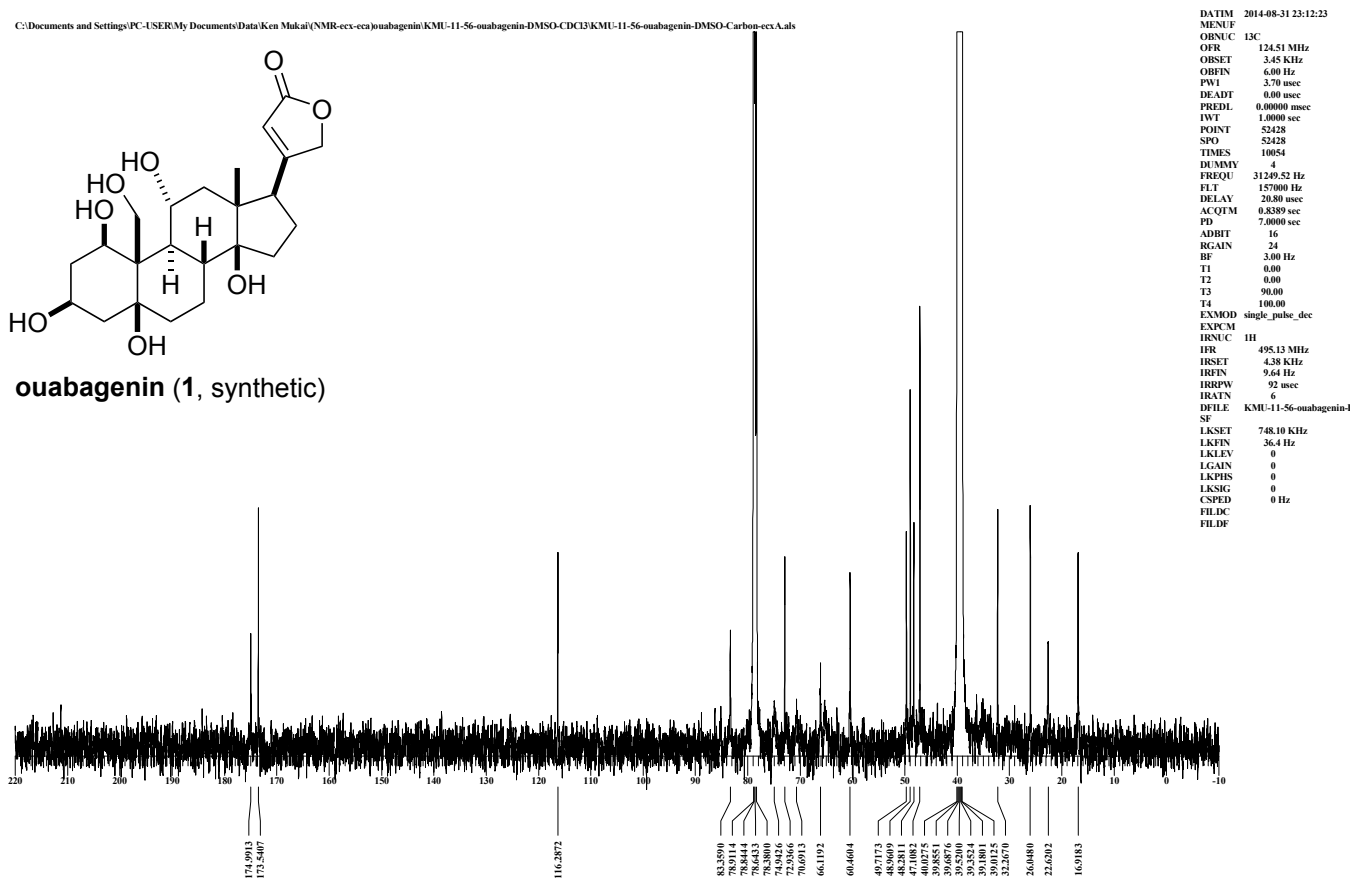


KMU-11-56-ouabagenin-DMSO-Carbon-cex

C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\NMR-cex-cex\ouabagenin\KMU-11-56-ouabagenin-DMSO-CDCl3\KMU-11-56-ouabagenin-DMSO-Carbon-cex-A.xls

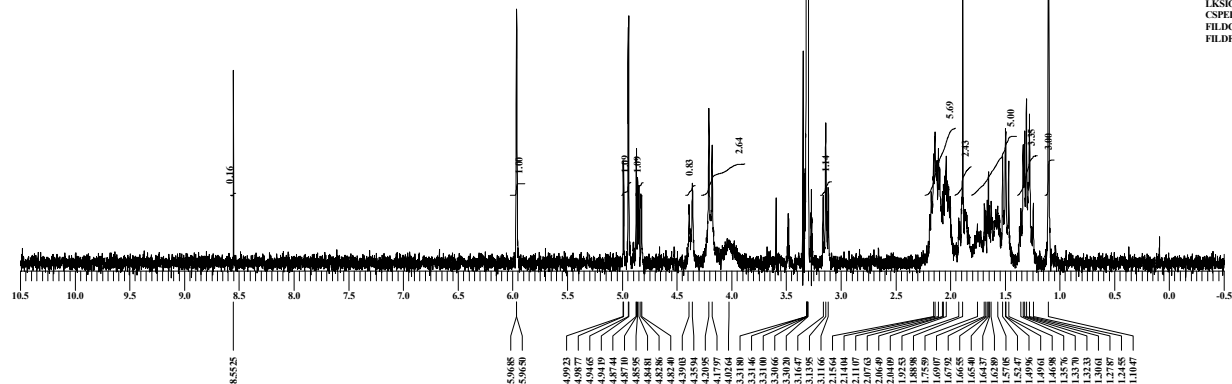
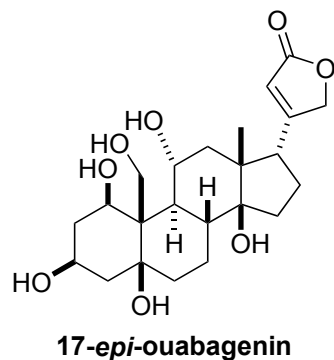


ouabagenin (1, synthetic)

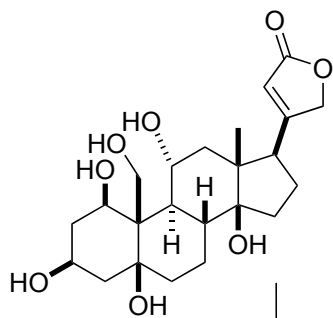


KMU-11-56-minor-product-dant-H2O

C:\Documents and Settings\PC-USER\My Documents\Data\Ken Mukai\NMR-ecs-eca\ouabagenin\KMU-11-56-minor-product-dant-H2O\KMU-11-56-minor-product-dant-H2O-Aals



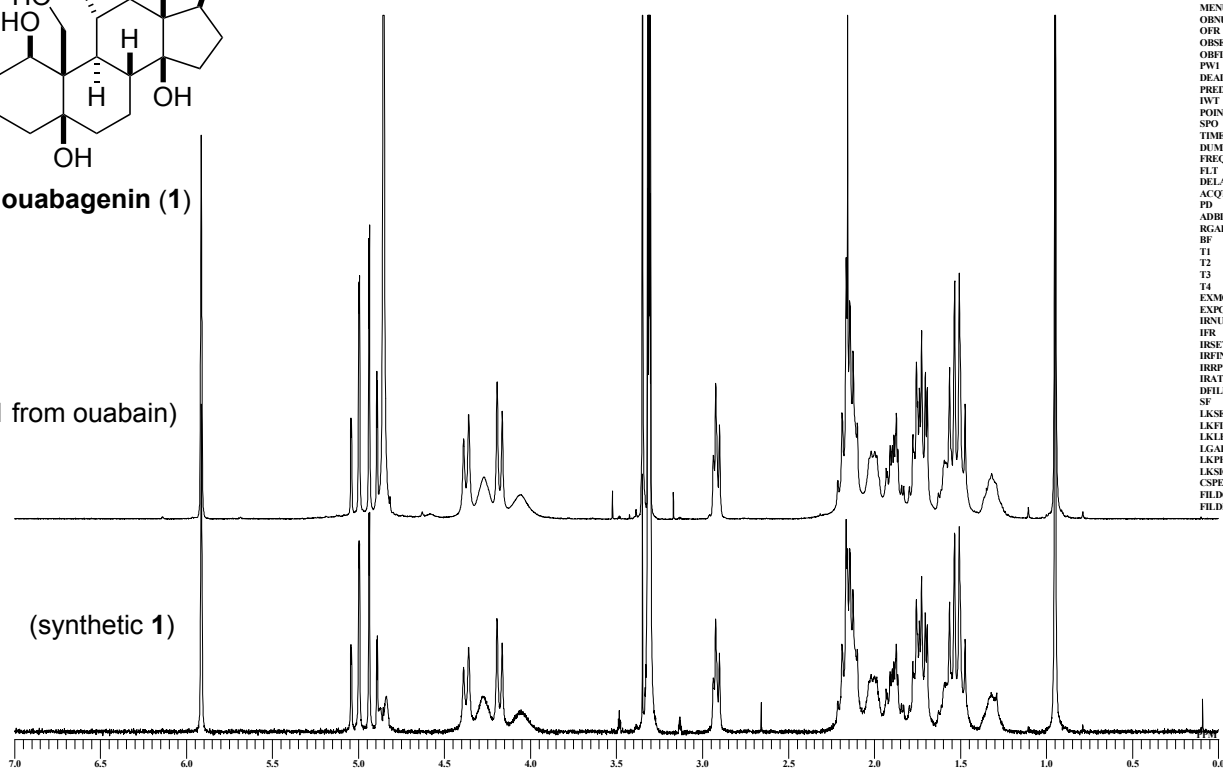
DATIM 28-08-2014 14:38:47
 MENUF
 OBNUC 1H
 OFR 395.88 MHz
 OBSET 6.21 KHz
 OFBIN 0.51 Hz
 PW1 6.44 usec
 DEADT 0.00 usec
 PREDL 0.00000 usec
 IWT 1.0000 sec
 POINT 16384
 SFO 16384
 TIMES 8
 DUMMY 1
 FREQU 7422.80 Hz
 FLT 30000 Hz
 DELAY 16.68 usec
 ACQTM 2.2073 sec
 PD 10.0000 sec
 ADBIT 16
 RGAIN 50
 BF 0.01 Hz
 T1 0.00
 T2 0.00
 T3 100.00
 T4 100.00
 EXMOD single_pulse.cx2
 EXPCM
 IRNUC 1H
 FR 395.88 MHz
 IRSET 6.28 KHz
 IRFIN 0.87 Hz
 IRRPW 115 usec
 IRATN 79
 DFILE KMU-11-56-minor-prodn
 SF 13.00 KHz
 LKSET
 LKLEV 0
 LKFIN 35.6 Hz
 LKFIN 0
 LKFIN 0
 LKSG 0
 CSPED 0 Hz
 FILDC
 FILDF



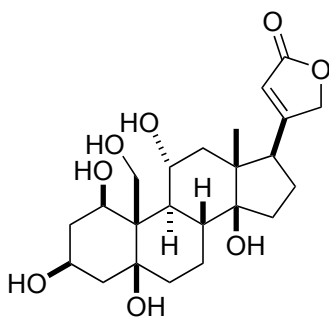
ouabagenin (1)

(1 from ouabain)

(synthetic 1)



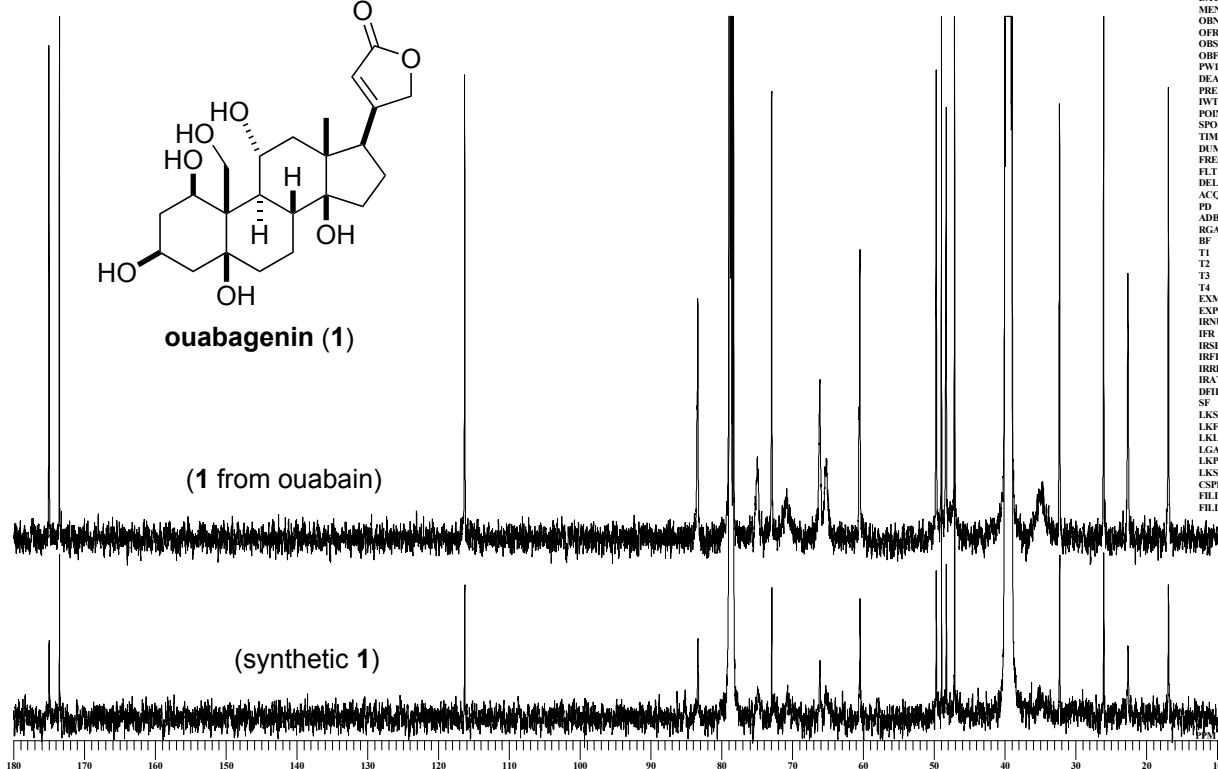
DATIM 26-06-2014 10:05:07
 MENUF
 ORNUC 1H
 OFR 395.88 MHz
 ORSET 6.21 KHz
 OFPIN 3.80 Hz
 PW1 6.44 usec
 DEADT 0.00 usec
 PREDL 0.00000 msec
 IWT 1.0000 sec
 POINT 13107
 SPO 13107
 TIMES 16
 DUMMY 1
 FREQU 5938.15 Hz
 FLT 30000 Hz
 DELAY 16.08 usec
 ACQTM 2.2073 sec
 PD 10.0000 sec
 ADBIT 16
 RGAIN 46
 BF 0.01 Hz
 T1 0.00
 T2 0.00
 T3 90.00
 T4 100.00
 EXMOD single_pulse.cx2
 EXPCM
 IRNUC 1H
 IFR 395.88 MHz
 IRSET 6.28 KHz
 IRFIN 0.87 Hz
 IRPW 1.47 usec
 IRATN 79
 DFIL KMU-11-56-HPLC-ouaba
 SF
 LKSET 13.00 KHz
 LKFIN 35.6 Hz
 LKLEV 0
 LGAIN 0
 LKPHS 0
 LKSG 0
 CSPED 0 Hz
 FILDC
 FILDF



ouabagenin (1)

(1 from ouabain)

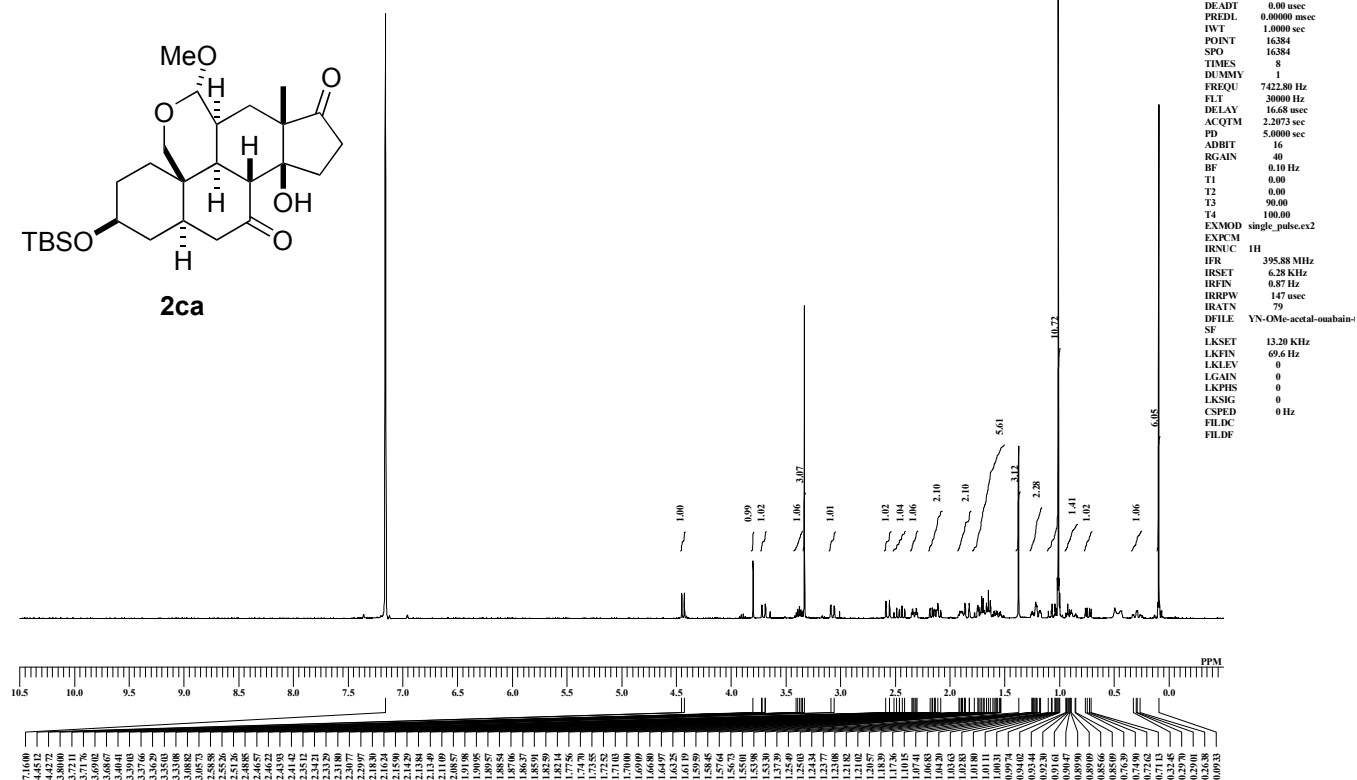
(synthetic 1)



DATIM 2014-08-31 23:12:23
 MENUF
 ORNUC 13C
 OFR 124.51 MHz
 ORSET 3.45 KHz
 OFPIN 6.00 Hz
 PW1 3.70 usec
 DEADT 0.00 usec
 PREDL 0.00000 msec
 IWT 1.0000 sec
 POINT 52428
 SPO 52428
 TIMES 10054
 DUMMY 4
 FREQU 31240.52 Hz
 FLT 157000 Hz
 DELAY 20.80 usec
 ACQTM 0.8389 sec
 PD 7.0000 sec
 ADBIT 16
 RGAIN 24
 BF 3.00 Hz
 T1 0.00
 T2 0.00
 T3 90.00
 T4 100.00
 EXMOD single_pulse_dec
 EXPCM
 IRNUC 1H
 IFR 495.13 MHz
 IRSET 4.38 KHz
 IRFIN 9.64 Hz
 IRPW 92 usec
 IRATN 6
 DFIL KMU-11-56-ouabagenin-1
 SF
 LKSET 748.10 KHz
 LKFIN 36.4 Hz
 LKLEV 0
 LGAIN 0
 LKPHS 0
 LKSG 0
 CSPED 0 Hz
 FILDC
 FILDF

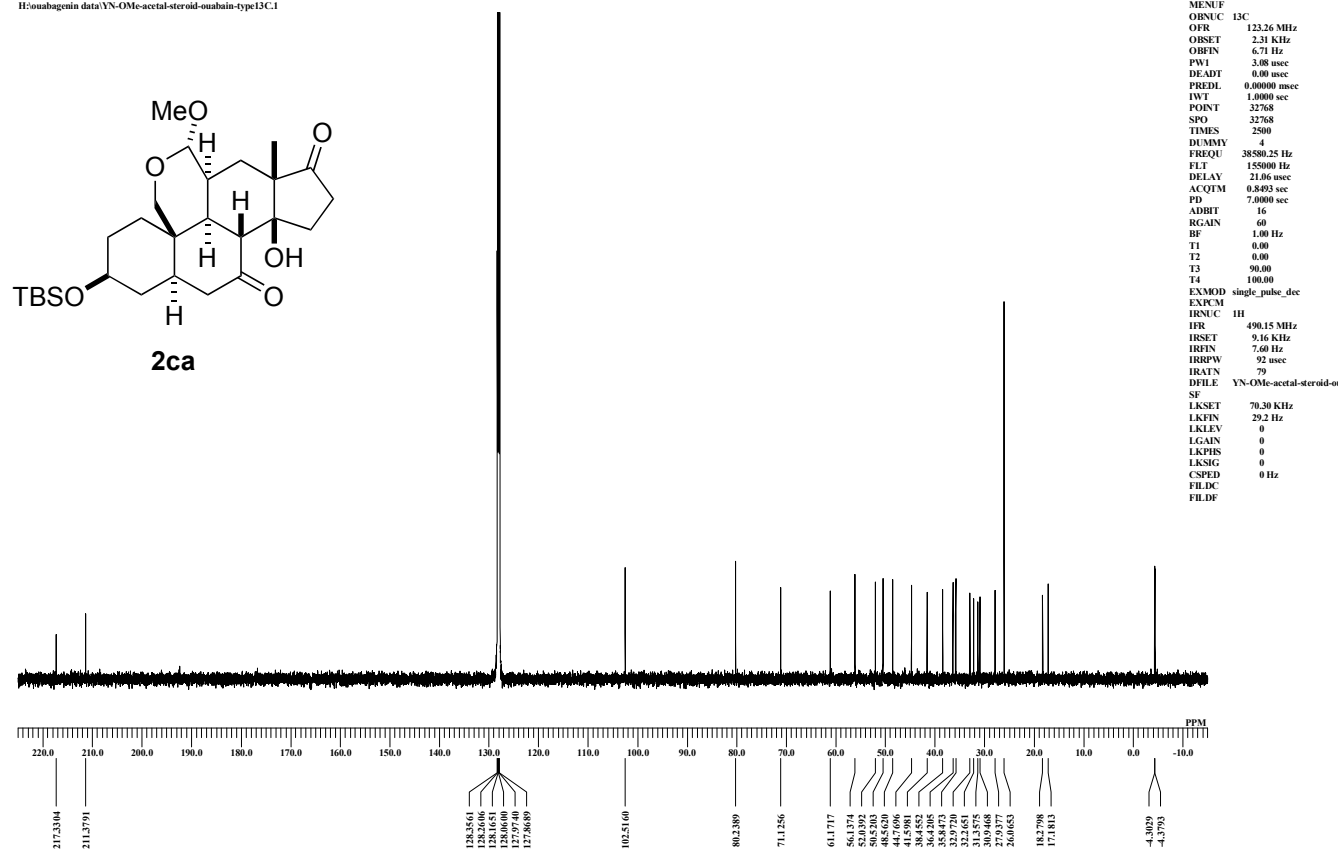
YN-OMe-acetal-ouabain-type

C:\Documents and Settings\PC-USER\My Documents\Data\Nakagawa\ouabagenin data\YN-OMe-acetal-ouabain-type-Lals



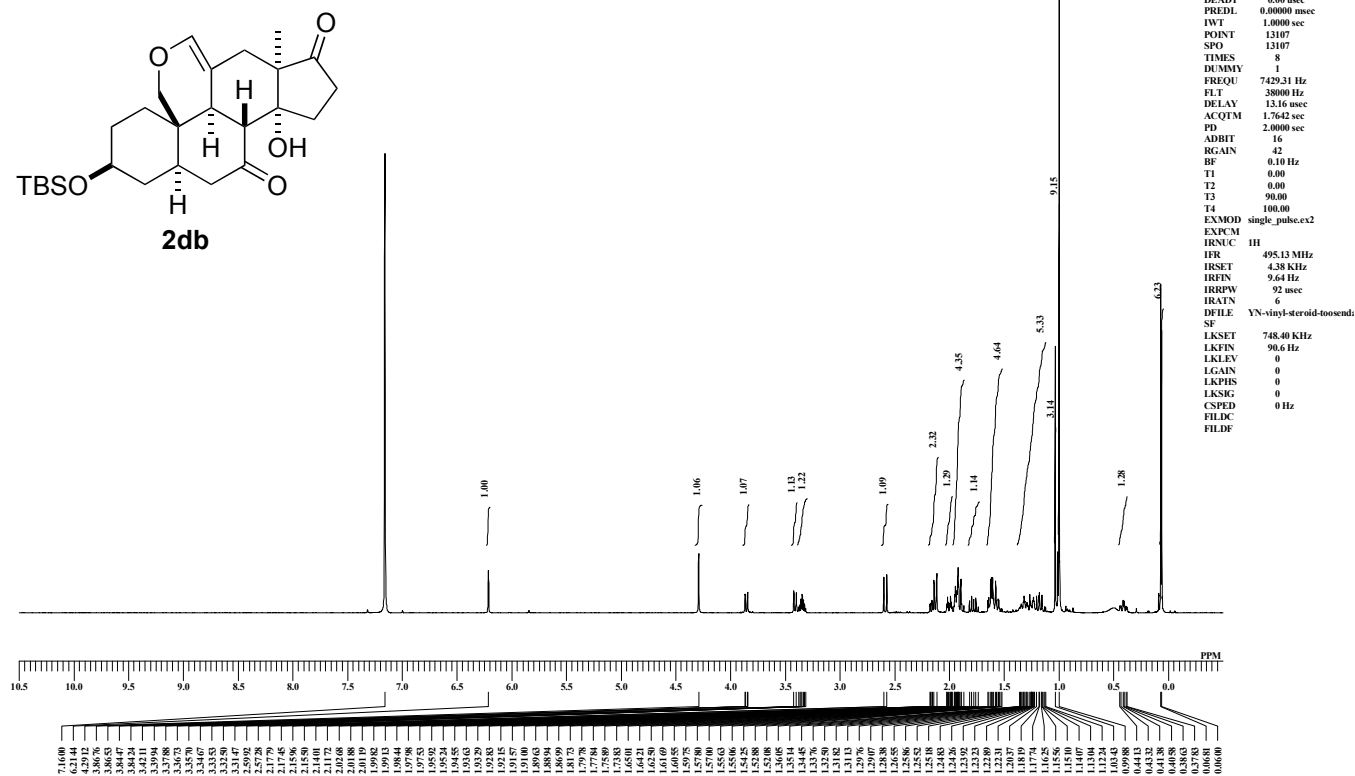
YN-OMe-acetal-steroid-ouabain-type13C

H:\ouabagenin data\YN-OMe-acetal-steroid-ouabain-type13C.1



YN-vinyl-steroid-toosendanin-type

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YN-vinyl-steroid-toosendanin-type-13C

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