Electronic Supplementary Information (ESI)

# A dual-response BODIPY-based fluorescent probe for the discrimination of Glutathione from Cystein and Homocystein

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1. Time-dependent spectral changes of BODIPY-S in the presence of thiols.

**Fig. S1** Time-dependent spectral changes of **BODIPY-S** (5  $\mu$ M) in the absence and presence of 5 mM thiols in acetonitrile / HEPES buffer (1:1, v/v, 20 mM, pH 7.4) at 37 °C. GSH: (a) Absorption and (b) emission spectra,  $\lambda_{ex} = 565$  nm. Cys: (c) Absorption and (d) emission spectra,  $\lambda_{ex} = 480$  nm. Hcy: (e) Absorption and (f) emission spectra,  $\lambda_{ex} = 480$  nm.



# 2. MS of BODIPY-S with GSH, Cys and Hcy.

Fig. S2 MS spectra of the products from BODIPY-S with thiols.

3. Photophysical spectra of BODIPY 4 and BODIPY-S +Cys.



Fig. S3 Photophysical spectra of BODIPY 4 and BODIPY-S + Cys.



#### 4. Time dependent spectral changes of S-S-BODIPY in the presence of thiols.

**Fig. S4** Time-dependent spectral changes of **S-S-BODIPY** (5  $\mu$ M) in the absence and presence of 5 mM thiols in acetonitrile / HEPES buffer (1:1, v/v, 20 mM, pH 7.4) at 37 °C. Cys: (a) Absorption and (b) emission spectra,  $\lambda_{ex} = 487$  nm. Hcy: (c) Absorption and (d) emission spectra,  $\lambda_{ex} = 487$  nm. GSH: (e) absorption and (f) emission spectra,  $\lambda_{ex} = 487$  nm.



#### 5. <sup>1</sup>H NMR and HRMS of the product from S-S-BODIPY with thiols.



# 6. Reaction mechanism.



Scheme S1. Reaction mechanism of compound BODIPY-S toward GSH, Cys/Hcy.



Scheme S2. Reaction mechanism of compound S-S-BODIPY toward GSH, Cys/Hcy.

#### 7. Pseudo first-order kinetic plot.

Time-course kinetic measurements of **S-S-BODIPY-S** with GSH were performed using the fluorescence intensity at 605 nm. Data were collected under pseudo-first-order conditions. Spectra were acquired in acetonitrile / HEPES buffer (1:1, v/v, 20 mM, pH 7.4) at 37 °C. The pseudo-first-order rate constant for the reaction was determined by fitting the fluorescence intensity changes of the samples to the pseudo first-order equation:  $Ln((I_{max}-I_t)/I_{max})) = -k_{obs} t$ .

Where  $I_t$  and  $I_{max}$  represent the fluorescence intensities at times t and the maximum value obtained after the reaction was complete.  $k_{obs}$  is the observed rate constant.



Fig. S6 Pseudo first-order kinetic plots of the reaction of S-S-BODIPY-S (5 µM) with GSH (5 mM).

#### 8. HRMS of S-S-BODIPY-S with thiols.

#### **Elemental Composition Report**

Single Mass Analysis Tolerance = 30.0 mDa / DBE: min = -1.5, max = 100.0 Element prediction: Off Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions 556 formula(e) evaluated with 1 results within limits (up to 1 best isotopic matches for each mass) Elements Used: C: 0-30 H: 0-33 11B: 0-1 N: 0-5 O: 0-7 F: 0-2 S: 0-1





#### Elemental Composition Report

Single Mass Analysis Tolerance = 30.0 mDa / DBE: min = -1.5, max = 100.0 Element prediction: Off Number of isotope peaks used for i-FIT = 2

Monoisotopic Mass, Even Electron Ions 200 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass) Elements Used: C: 0-23 H: 0-150 11B: 0-1 N: 0-3 O: 0-3 F: 0-2 S: 0-1









Fig. S7 HRMS spectra of the products from S-S-BODIPY-S with thiols.



9. Time-dependent spectral changes of S-S-BODIPY-S in the presence of Cys/Hcy.

**Fig. S8** Time-dependent spectral changes of **S-S-BODIPY-S** (5  $\mu$ M) in the absence and presence of Cys (5 mM) or Hcy (5 mM) in acetonitrile / HEPES buffer (1:1, v/v, 20 mM, pH 7.4) at 37 °C. Cys: (a) absorption and (b) emission spectra; Hcy: (c) absorption and (d) emission spectra.  $\lambda_{ex} = 490$  nm.

## 10. The easy-to-monitor fluorescence color change.



Fig. S9 Fluorescence color changes in the presence of Cys, Hcy and GSH.

## 11. Fluorescent confocal image of cells.



**Fig. S10** Fluorescent confocal image of cells: (a-c) HeLa cells pretreated with 500  $\mu$ M *N*-methylmaleimide for 20 min, then incubated with **S-S-BODIPY-S** (10  $\mu$ M) for 4 h, the excitation wavelength was 561 nm and the emission was collected at 600-645 nm, (a) overlap field, (b) fluorescence image, (c) bright filed.

#### **12.** Experimental section

**Materials.** Unless other specified, all chemical reagents and solvents for synthesis were purchased from commercial suppliers and were used without further purification. Anhydrous dichloromethane was dried over  $CaH_2$  and distilled prior to use.

**Instruments.** NMR spectra were recorded on a Bruker AV-400 spectrometer with chemical shifts reported in ppm at room temperature. Mass spectra were obtained on a HP 1100 LC-MS spectrometer. UV-vis absorption spectra were recorded on a Varian Cary 100 spectrophotometer. Fluorescence spectra were measured with a Varian Cary Eclipse Fluorescence spectrophotometer. Spectral-grade solvents were used for measurements of UV-vis absorption and fluorescence. For absorption or fluorescence measurements, compounds were dissolved in  $CH_3CN$  to obtain stock solutions (5.0 mM), followed by diluting with aqueous buffer solutions to the desired concentrations.

**Cells culture and imaging.** HeLa cells and MKN-45 cells were cultured in Roswell Park Memorial Institute 1640 medium (RPMI-1640) supplemented with 10% fetal bovine serum (FBS) in a humidified atmosphere of 5/95 CO<sub>2</sub>/air incubator for 24 h at 37 °C. For confocal fluorescence imaging, cells were incubated in glass bottom dishes for 24 h. Cells were incubated at 37 °C with 10  $\mu$ M **S-BODIPY-S** for 4 h, washed with D-Hanks and fluorescence images were captured. In a control experiment, Hela cells were pre-treated with *N*-methylmaleimide (500  $\mu$ M) for 20 min at 37 °C, and then washed with D-Hanks. These cells were further loaded with 10  $\mu$ M **S-BODIPY-S** for 4 h, washed with D-Hanks. Samples were excited at 514 nm and observed between 600-620 nm for the red channel and 560-580 nm for the orange channel.



Synthesis of Compound 2. HO-BODIPY-Cl (100 mg, 0.3 mmol) and *p*-thiocresol (39 mg, 0.3 mmol) were dissolved in CH<sub>3</sub>CN (30 ml), and then DMAP (38 mg, 0.3 mmol) was added, the mixture was stirred for 1 h at room temperature under N<sub>2</sub> atmosphere. The solvent was then removed under vacuum, the resulting residue was purified by column chromatography on silica gel to afford the compound 2 (102 mg, yield: 83%). <sup>1</sup>H NMR (*d*<sub>6</sub>-DMSO, 400 MHz, ppm):  $\delta$  10.51 (s, 1H), 7.60-7.54 (m, 5H), 7.48-7.43 (m, 3H), 7.38-7.36 (d, *J* = 8.4 Hz, 2H), 6.90 (s, 1H), 6.63-6.60 (dd, *J*<sub>1</sub> = 2.0 Hz, *J*<sub>2</sub> = 8.8 Hz, 1H), 6.55-6.53 (d, *J* = 4.8 Hz, 1H), 5.87-5.86 (d, *J* = 4.8 Hz, 1H), 2.38 (s, 3H), 1.68 (s, 3H). HRMS (ESI, *m/z*): [M - H]<sup>-</sup> calcd for C<sub>27</sub>H<sub>20</sub>BF<sub>2</sub>N<sub>2</sub>OS: 469.1357, Found: 469.1364.

Synthesis of Compound 3. To a solution of HO-BODIPY-Cl (0.10 g, 0.3 mmol) in 30 mL CH<sub>3</sub>CN was added morpholine (30 mg, 0.3 mmol), the mixture was then stirred for 30 min at room temperature, washed with water, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The solvent was evaporated in vacuo. The residue was purified by flash chromatography (silica gel, eluent: ethyl acetate/petroleum 1:5) to afford compound 3 (0.100 g, yield: 88%). <sup>1</sup>H NMR (*d*<sub>6</sub>-DMSO, 400 MHz, ppm):  $\delta$  9.41 (s, 1H), 7.55-7.54 (m, 3H), 7.40-7.37 (m, 2H), 7.29-7.26 (d, *J* = 8.8 Hz, 1H), 6.96 (s, 1H), 6.84-6.83 (d, *J* = 5.2 Hz, 1H), 6.68-6.67 (d, *J* = 5.2 Hz, 1H), 6.54-6.51 (dd, *J*<sub>1</sub> = 2.0 Hz, *J*<sub>2</sub> = 8.8 Hz, 1H), 4.07-4.05 (t, *J* = 4.4 Hz, 4H), 3.82-3.80 (t, *J* = 4.4 Hz, 4H), 1.56 (s, 3H). HRMS (ESI, *m/z*): [M + H]<sup>+</sup> calcd for C<sub>24</sub>H<sub>23</sub>BF<sub>2</sub>N<sub>3</sub>O<sub>2</sub>: 434.1851, Found: 434.1846.

**Compound BODIPY-S.** To a solution of Compound **2** (50 mg, 0.1 mmol) and DMAP (20 mg, 0.2 mmol) in 30 mL anhydrous DCM was added acetic anhydride (16 mg, 0.2 mmol) at room temperature, the reaction mixture was stirred for 1 h under argon. Then the reaction

mixture was washed with water. The organic layer was dried with Na<sub>2</sub>SO<sub>4</sub>, and evaporated under vacuum. The resulting residue was purified by column chromatography on silica gel to get compound **BODIPY-S** (45 mg, yield 83%). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz, ppm):  $\delta$  7.58-7.56 (d, *J* = 8.4 Hz, 2H), 7.52-7.50 (m, 4H), 7.46-7.44 (d, *J* = 8.8 Hz, 1H), 7.37-7.35 (m, 2H), 7.29-7.27 (d, *J* = 8.0 Hz, 2H), 6.81-6.78 (dd, *J*<sub>1</sub> = 2.0 Hz, *J*<sub>2</sub> = 8.8 Hz, 1H), 6.58-6.57 (d, *J* = 4.8 Hz, 1H), 5.94-5.92 (d, *J* = 4.8 Hz, 1H), 2.42 (s, 3H), 2.33 (s, 3H), 1.74 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz, ppm):  $\delta$  169.43, 152.14, 145.89, 141.25, 141.09, 141.02, 135.40, 134.09, 132.35, 132.18, 130.75, 129.64, 129.54, 129.29, 128.60, 124.62,121.91, 121.36, 116.16, 107.49, 29.71, 21.18, 11.85. HRMS (ESI, *m/z*): [M + Na]<sup>+</sup> calcd for C<sub>29</sub>H<sub>23</sub>BF<sub>2</sub>N<sub>2</sub>NaO<sub>2</sub>S: 535.1434, Found: 535.1439.

**Compound S-S-BODIPY**. Compound **3** (100 mg, 0.2 mmol) was dissolved in 1,2dichloroethane. 1,1'-Carbonyldiimidazole (75 mg, 0.5 mmol) was added to the above solution, the reaction mixture was stirred for 12 h at 40 °C under argon. To the reaction mixture was added bis(2-hydroxyethyl)disulfide (106 mg, 0.7 mmol), the resulting reaction mixture was then refluxed for 24 h. After cooling to room temperature, the solvent was removed under vacuum. The resulting residue was purified by column chromatography on silica gel ( ethyl acetate / petroleum = 1: 7) to afford the compound **S-S-BODIPY** as a red solid (28 mg, yield 20%). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz, ppm):  $\delta$  7.56 (s, 1H), 7.49-7.47 (m, 3H), 7.45-7.42 (d, *J* = 8.8 Hz, 1H), 7.35-7.33 (m, 2H), 6.87-6.84 (dd, *J*<sub>1</sub> = 2.0 Hz, *J*<sub>2</sub> = 8.8 Hz, 1H), 6.75-6.74 (d, *J* = 5.2 Hz, 1H), 6.34-6.33 (d, *J* = 5.6 Hz, 1H), 4.54-4.50 (t, *J* = 6.8 Hz, 2H), 4.08-4.06 (m, 4H), 3.93-3.89 (m, 6H), 3.06-3.03 (t, *J* = 6.8 Hz, 2H), 2.93-2.90 (t, *J* = 5.6 Hz, 2H), 1.67 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz, ppm):  $\delta$  153.77, 149.53, 141.94, 139.22, 135.90, 135.07, 132.22, 130.02, 129.75, 128.83, 128.40, 122.00, 120.46, 115.12, 113.83, 106.41, 66.87, 66.21, 60.23, 50.82, 41.60, 36.83, 29.71, 10.79. HRMS (ESI, *m/z*): [M + Na]<sup>+</sup> calcd for C<sub>29</sub>H<sub>30</sub>BF<sub>2</sub>N<sub>3</sub>NaO<sub>5</sub>S<sub>2</sub>: 636.1580, Found: 636.1589.

Compound S-S-BODIPY-S. The solution of compound 2 (100 mg, 0.2 mmol) and 1,1carbonyldiimidazole (45 mg, 0.3 mmol) in 1, 2-dichloroethane (30 mL) was stirred for 24 h at 40 °C under argon. The progress of the reaction was monitored by TLC, when the starting material disappeared and a new intermediate appeared, the bis(2-hydroxyethyl)disulfide (98 mg, 0.6 mmol) was then added to the reaction mixture, the resulting mixture was refluxed for 48 h. After cooling to room temperature, the reaction mixture was diluted with  $CH_2Cl_2$  and washed with water for 3 times, dried over Na<sub>2</sub>SO<sub>4</sub>, and the solvent was removed under vacuo. The residue was purified by flash chromatography (silica gel, eluent: ethyl acetat / petroleum 1:10) to afford S-S-BODIPY-S (20 mg, yield 14%); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz, ppm):  $\delta$ 7.61 (s, 1H), 7.58-7.56 (d, J = 8.4 Hz, 2H), 7.52-7.50 (m, 3H), 7.48-7.45 (d, J = 8.8 Hz, 1H), 7.37-7.35 (m, 2H), 7.29-7.27 (d, J = 8.0 Hz, 2H), 8.87-8.85 (dd,  $J_1 = 2.0$  Hz,  $J_2 = 8.8$  Hz, 1H), 6.60-6.58 (d, J = 4.4 Hz, 1H), 5.95-5.94 (d, J = 4.4 Hz, 1H), 4.56-4.53 (t, J = 6.8 Hz, 2H), 3.94-3.91 (t, J = 5.6 Hz, 2H), 3.08-3.05 (t, J = 6.8 Hz, 2H), 2.95-2.92 (t, J = 5.6 Hz, 2H), 2.42 (s, 3H), 1.74 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz, ppm): δ 153.32, 152.10, 145.52, 141.39, 141.17, 140.98, 135.40, 134.01, 132.54, 131.85, 130.78, 129.75, 129.60, 129.28, 128.63, 124.49, 122.08, 121.57, 115.29, 107.09, 66.46, 60.26, 41.59, 36.73, 21.41, 11.84; HRMS (ESI, m/z):  $[M + Na]^+$  calcd for  $C_{32}H_{29}N_2BS_3F_2O_4Na$ : 673.1248, Found: 673.1245.

### 13. NMR and HRMS Spectra





S17















S21