**S**1

## **Supporting Information**

# A Fully Synthetic and Self-Adjuvanting Globo H-Based Anticancer Vaccine Elicited Strong T Cell-Mediated Antitumor Immunity

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#### I. Conjugate Synthesis and Analysis Experiments

General Experimental Methods. Chemicals and materials were obtained from commercial sources and were used as received without further purification unless otherwise noted. MS 4 Å was flame-dried under high vacuum and used immediately after cooling under a N<sub>2</sub> atmosphere. Analytical TLC was carried out on silica gel 60Å  $F_{254}$  plates with detection by a UV detector and/or by charring with 15% (v/v)  $H_2SO_4$  in EtOH. NMR spectra were recorded on a 400, 500, or 600 MHz machine with chemical shifts reported in ppm ( $\delta$ ) downfield from tetramethylsilane (TMS) that was used as an internal reference.



**Compound 6.** To a stirred solution of **4** (18 mg, 8  $\mu$ mol) and *p*-nitrophenol (5.9 mg, 42  $\mu$ mol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) was added EDC·HCl (8.2 mg, 42  $\mu$ mol) in an ice bath. After the mixture was stirred at rt for 5 h, it was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and condensed in vacuum. The residue was purified on a TLC plate (MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 1:20, v/v) to give the activated ester **6** as a white floppy solid (16 mg, 83.5%). Its spectroscopic data were identical to that reported in the literature (Z. Zhou, M. Mondal, G. Liao and Z. Guo, *Org. Biomol. Chem.*, 2014, **12**, 3238-3245).



**Compound S2.** A mixture of **S1** (10.0 mg, 4.46  $\mu$ mol, Z. Zhou, M. Mondal, G. Liao and Z. Guo, *Org. Biomol. Chem.*, 2014, **12**, 3238-3245) and 10% Pd-C (10.0 mg) in CH<sub>2</sub>Cl<sub>2</sub>-MeOH (3:1, 8 mL) was stirred under an atmosphere of H<sub>2</sub> at rt for 12 h. Thereafter, the catalyst was removed by filtration through a Celite pad, and the Celite pad was subsequently washed with CH<sub>2</sub>Cl<sub>2</sub>-MeOH-H<sub>2</sub>O (3:3:1). The combined filtrates were concentrated in vacuum, to give **S2** as a yellow solid (7.0 mg, 93.7%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O=3:3:1):  $\delta$  5.05-4.90 (br, 2H), 4.88-4.80 (br, 1H), 4.46-4.37 (br, 1H), 3.85-3.80 (br, 4H), 3.80-3.75 (m, 3H), 3.75-3.60 (br, 1H), 3.60-3.42 (m, 2H), 3.41-3.20 (m, 3H), 2.98 (br, 1H,  $\frac{1}{2}CH_2$ NH<sub>2</sub>), 2.90 (br, 1H,  $\frac{1}{2}CH_2$ NH<sub>2</sub>), 2.43-2.00 (m, 12 H, lipid), 1.50-0.95(br, 108 H, 54 x CH<sub>2</sub>, lipid), 0.81-0.60 (18 H, 6 x CH3, lipid). <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O=3:3:1):  $\delta$  173.71, 173.57, 172.34, 171.32, 171.00, 101.53, 75.18, 74.02, 71.26, 70.87, 68.60, 68.15, 60.03, 53.68, 52.87, 48.96, 48.81, 48.67, 48.52, 48.24, 48.10, 42.01, 41.60, 41.13, 40.98, 40.33, 37.27, 36.94, 34.27, 33.90, 33.80, 31.68, 31.14, 31.05,

29.93, 29.87, 29.82, 29.42, 29.32, 29.29, 29.25, 29.12, 28.98, 28.95, 25.34, 25.25, 25.05, 25.00, 24.86, 24.81, 22.42, 13.71; <sup>31</sup>P NMR (400 MHz, CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O=3:3:1):  $\delta$  0.405; HRMS (ESI, TOF): calcd. For C<sub>90</sub>H<sub>171</sub>N<sub>3</sub>O<sub>22</sub>P [M+H]<sup>+</sup> m/z, 1677.2091; found, 1677.2091. (**S2** was used as the capture reagent for ELISA analysis of MPLA-specific antibodies.)

Analysis of the carbohydrate loading of glycoconjugates (R. E. Wrolstad, T. E. Acree and E. A. Decker, Current Protocols in Food Analytical Chemistry, John Wiley & Sons, Inc., 2001): The sugar calibration curve was prepared using a standard solution of fucose, galactose, Nacetylgalactosamine, and glucose (1.2 mg/mL in 1/3/1/1 molar ratio) in distilled water. Aliquots were transferred to 10 dry 10 ml tubes in 5  $\mu$ L increments ranging from 5 to 50  $\mu$ L. In another 10 mL test tube, accurately weighed samples of the glycoconjugates to be analyzed were placed. At this point, all the tubes should contain between 5 to 50 µg of sugar, and one should contain an unknown amount of sugar to be determined. To all of the tubes were sequentially added 500  $\mu$ L of 4% phenol and 2.5 mL of 96% sulfuric acid. The glycosyl linkages were cleaved and a colored complex was formed in this step. Solutions were transferred from the test tubes to cuvettes and measured at the wavelength of 490 nm. The calibration curve was obtained by plotting A490 against the weight (µg) of sugar in the standard samples. The amount of sugar present in each unknown sample was calculated based on the A490 of the unknown sample against the calibration curve, while the free proteins KLH and HSA were used as blank controls for conjugates 2 and 3, respectively. The carbohydrate loading of each glycoconjugate was calculated according to the following equation, and the results for KLH conjugate 2 and HSA conjugate **3** were 8.0% and 14.0%, respectively.

Carbohydrate loading % = sugar weight in a tested sample/total weight of the sample  $\times$  100%

**Procedure for SDS-PAGE analysis of glycoconjugates:** The mixture of a glycoconjugate (1  $\mu$ g) in 0.1 M PSB buffer (1  $\mu$ l), LDS NuPAGE® sample buffer (4 ×, 2.5  $\mu$ l), reducing agent (10×, 1  $\mu$ l), and deionized water (5.5  $\mu$ l) was heated at 70 °C for 10 min, and loaded onto the sodium dodecyl sulfate polyacrylamide gel. The upper buffer chamber was filled with 200 ml of 1 × SDS running buffer containing 500  $\mu$ l of NuPAGE® antioxidant, and the lower buffer chamber was filled with 600 ml of 1 × SDS running buffer. The gel was run at 200 V for 2 h, and stained with Coomassie brilliant blue R-250 for 8 h (Ref: NuPAGE® technical instructions of Invitrogen).



**Figure S1**. The SDS-PAGE results. Lane 0: molecular markers; Lane 1: KLH protein; Lane 2: KLH-Globo H conjugate **2**.

#### II. Size Analysis of the MPLA Conjugate Liposomes

The liposomal formulation of Globo H-MPLA conjugate **1** prepared for the immunization of mice was subjected dynamic light scattering (DLS). The sample was tested five times, and the results are listed in Table S1. It was concluded that the average diameter of the liposome was 1,429.2  $\pm$  249 (standard deviation) nm with the polydispersity index (PDI) around 0.5832 (Figure S2).

test	PDI	Size (d, nm)	% volume	Width (d, nm)
#1	0.54	1503	96.5	225.2
# 2	0.472	1816	92.1	402.7
# 3	0.704	1366	100	173.2
#4	0.521	1302	96	203.1
# 5	0.679	1159	94.1	173.7
Ave	0.5832	1429.2	95.74	
SD	0.1023	249.12	2.947	
			Size Distribution by Volume	
	40			
	€ 30			
	9 10			
		1 10	100 Size (d.nm)	1000 10000
		F	Record 3: Globo H MPLA liposo	ome 1
	L			

Table S1. DLS analysis results for the liposomal formulation of Globo H-MPLA conjugate 1

Figure S2. Size distribution of the liposomes of conjugate 1 based on DLS volume analysis

## **III.** Calculated Antibody Titers of ELISA Experiments

Table S2. The total antibody	v titers of po	ooled antisera induced by	conjugates 1	and <b>2</b> (Figure 2A)
-		2	50	

	1			2			
	Mean	SD	Ν	Mean	SD	Ν	
d0	0	0	3	0	0	3	
d21	47824	3211	3	3299	481	3	
d27	57648	2452	3	10937	2417	3	
d38	63038	1595	3	29128	3110	3	

Table S3. The IgG antibody titers of pooled antisera induced by conjugates 1 and 2 (Figure 2B)

	1			2			
	Mean	SD	Ν	Mean	SD	Ν	
d0	0	0	3	0	0	3	
d21	46449	7768	3	2783	655	3	
d27	65577	850	3	8273	2600	3	
d38	69406	1584	3	29383	3326	3	

Table S4. The antibody titers of IgG subclasses in individual antiserum induced by 1 (Figure 3A)

mouse	1	2	3	4	5	6	Mean
IgG1	108952	3853	63146	12321	64011	130594	63813
IgG2b	10552	1	155	12	2430	14317	4578
IgG2c	6039	0	291	1751	55	5232	2228
IgG3	2177	0	2	14	7092	27667	6159

Table S4. The antibody titers of IgG subclasses in individual antiserum induced by 2 (Figure 3B)

mouse	1	2	3	4	5	6	Mean
IgG1	20952	22450	4333	22678	33973	65033	28237
IgG2b	21324	1772	2269	5963	17979	454	8294
IgG2c	22	2601	6299	15	1536	1	1746
IgG3	210	0	253	5148	2623	911	1524

Table S5. The antibody titers of IgM in individual antiserum induced by conjugate 1 and 2

mouse	1	2	3	4	5	6	Mean
Conjugate 1	2932	401	0	4866	39471	101153	24804
Conjugate 2	28681	20618	16195	8	2	19789	14216



#### IV. Raw Data for the Cytokine Release Assays

Figure S3. The expression levels of cytokines, including IL-3, IL-4, IL-9, IL-10, IL-12, IL-12p70, IL-13, IFN-y, MCP-1, MCP-5, and RANTES, have significantly increased in the antisera derived from mice immunized with Globo H-MPLA 1, compared to that of normal mouse sera.



Figure S4. The expression levels of cytokines, including GCSF, GM-CSF, IL-3, IL-4, IL-5, IL-6, IL-9, IL-10, IL-12, IL-12p70, IL-13, IFN-γ, MCP-1, MCP-5, RANTES, and TNF-α, have significantly increased in the antisera derived from mice immunized with Globo H-MPLA 1, compared to that of normal mouse sera.

Normal sera

#### **Globo H-KLH antisera**



	А	В	С	D	E	F	G	н
1	Pos	Pos	Neg	Neg	GCSF	GM-CSF	IL-2	IL-3
2	Pos	Pos	Neg	Neg	GCSF	GM-CSF	IL-2	IL-3
3	IL-4	IL-5	IL-6	IL-9	IL-10	IL-10 IL-12 p40p70		IL-13
4	IL-4	IL-5	IL-6	IL-9	IL-10 IL-12 p40p70		IL-12p70	IL-13
5	IL-17	IFN-y	MCP-1	MCP-5	RANTES	SCF	sTNFRI	TNF-α
6	IL-17	IFN-γ	MCP-1	MCP-5	RANTES	SCF	sTNFRI	TNF-α
7	Thrombopoietin	VEGF	BLANK	BLANK	BLANK	BLANK	BLANK	Pos
8	Thrombopoietin	VEGF	BLANK	BLANK	BLANK	BLANK	BLANK	Pos

Figure S5. The membrane map for experiments to obtain the results in Figures S3 and S4.

### V. Assays of MPLA- and KLH Specific Antibodies

MPLA-specific antibody titer was determined by ELISA, similar to that used to measure other antibodies but using MPLA as the capture reagent to coat the NUNC PolySorp<sup>TM</sup> 96-well plates. The MPLA derivative **S2** used for coating plates was dissolved in 0.2% triethylamine to get a final concentration of 0.03 mg/ml. After the solvent was evaporated, the plates were treated with blocking buffer following the normal protocol of ELISA. The MPLA-specific total antibody titer of the day 38 serum pooled from mice immunized with Globo H-MPLA conjugate **1** was 59,666, as compared to the Globo H-specific total antibody titer of 63,038 (Figure S6).



Figure S6. Titers of the Globo H- and MPLA-specific total antibodies in the pooled antiserum induced by conjugate 1. Data are presented as mean ± SEM. The difference of Globo H- and MPLA-specific total antibody titers is not statistically significant.

KLH-specific antibody titer was determined by ELISA, similar to that used to measure other antibodies but using KLH as the capture reagent to coat the plates. KLH powder was dissolved in the coating buffer (0.1 M bicarbonate, pH 9.6) to get a final concentration of 2  $\mu$ g/mL. Each well of the plates was treated with 100  $\mu$ L of KLH coating solution at 37 °C for 1 h and then with a blocking buffer following the normal protocol of ELISA. The KLH-specific antibody titer of the day 38 antiserum pooled from mice immunized with Globo H-KLH conjugate **2** was 293,919, and its Globo H-specific antibody titer was 23,177 (Figure S7). Evidently, the KLH conjugate provoked much stronger anti-KLH antibody response than the anti-globo H antibody response.



Figure S7. Titers of the Globo H- and KLH-specific antibodies in the pooled antiserum induced by conjugate 2. Data are presented as mean  $\pm$  SEM. \* Compared to the Globo H-specific total antibody titer, the difference is statistically very significant (*P* < 0.0001).



VI. NMR and MS Spectra of Synthesized Compounds



<sup>1</sup>H NMR Spectrum of compound **6** (CDCl<sub>3</sub>, 600 MHz)

<sup>1</sup>H Spectrum of compound 7 [CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O (3:3:1) 600 MHz]



<sup>1</sup>H-<sup>1</sup>H COSY Spectrum of compound **7** [CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O (3:3:1) 600 MHz]



 $^{1}$ H- $^{13}$ C HMQC NMR Spectrum of compound 7 [CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O (3:3:1) 600/150 MHz]



 $^{31}\text{P}$  NMR Spectrum of compound 7 [CDCl\_3:CD\_3OD:D\_2O (3:3:1) 400 MHz]



Acquisition Parameter D:Methods\1User\_Methods\Guo\RP\_3147\_PepMixDHB\_June 1-2011.par



#### MALTI-TOF MS spectrum of compound 7

HRMS (ESI MS) spectrum of compound 7



<sup>1</sup>H NMR Spectrum of compound **1** [CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O (5:3:1) 600 MHz]



 $^{31}\text{P}$  NMR Spectrum of compound 1 [CDCl\_3:CD\_3OD:D\_2O (5:3:1) 400 MHz]



MS (ESI MS) spectrum of compound 1



MALTI-TOF MS spectrum of HSA protein

S18





MALTI-TOF MS spectrum of compound 3

S19



<sup>1</sup>H NMR Spectrum of compound S2 [CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O (3:3:1) 600 MHz]



 $^{13}\text{C}$  NMR Spectrum of compound **S2** [CDCl\_3:CD\_3OD:D\_2O (3:3:1) 600 MHz]



 $^{1}$ H- $^{1}$ H COSY NMR Spectrum of compound **S2** [CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O (3:3:1) 600 MHz]



<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound **S2** [CDCl<sub>3</sub>:CD<sub>3</sub>OD:D<sub>2</sub>O (3:3:1) 600 MHz]



Mercury 400 spectrometer

 $^{31}\text{P}\,\text{NMR}$  Spectrum of compound S2 [CDCl\_3:CD\_3OD:D\_2O (3:3:1) 400 MHz]

#### **Elemental Composition Report**

Page 1

Single Mas Tolerance = Element pred Number of is	s Analysis 5.0 PPM / Di diction: Off otope peaks us	BE: min = - sed for i-FIT	1.5, max = 1 ' = 6	100.0								
Monoisotopic 312 formula(e Elements Use C: 90-90 H G.Liao liao-3-1 1503023_51 11	Mass, Odd and I ) evaluated with d: 1: 0-200 N: 0- 26 inMeOH+DCM- 103 (20.378) Cm (1	Even Electro 2 results with 5 O: 0-25 +Water Cone( 099:1234-193	n lons hin limits (all P: 0-1 V)60 :296x2.000)	results (u LCT Prem	ip to 10 hier XE I	000) for ea	ach mass)			14:28:3	36 23- TOF 3	Mar-2015 MS ES+ .83e+003
100			1677.2	091 1678.2	2097							
%-					1679.2	167						
1662	7415				1	680,2323	1682.3810			168	8.6688	m/z
1667.5	1670.0	1672.5	1675.0	1677.5		1680.0	1682.5	1685.0		1687.5	1	1690.0
Minimum: Maximum:		3.0	5.0	-1.5 100	5 .0							
Mass	Calc. Mass	mDa	PPM	DBE	ā	i-FIT	i-FIT	(Norm)	Form	ula		
1677.2091	1677.2092	-0.1	-0.1	7.5	į.	54.1	0.1		C90 P	H171	N3	022 <b>4 y</b>
	1677.2174	-8.3	-4.9	7.5	8	56.9	2.8		C90	H170	N3	024

HRMS (ESI MS) spectrum of compound  $\mathbf{S2}$ 

S25