High Fidelity Sorting of Remarkably Similar Components via Metal-Mediated Assembly

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Electronic Supplementary Information

Table of Contents

General Information	2
Synthesis of Compounds	2
NMR Spectral Data	5
Mass Spectral Data	11
Assembly Mixing Experiments	12
Ligand Displacement Experiments	22
References	24

General Information:

¹H and ¹³C NMR spectra were recorded on a Varian Inova 400 MHz or Varian Inova 500 MHz NMR spectrometer. DOSY spectra were recorded on a Bruker Avance 600 MHz spectrometer. Proton (¹H) chemical shifts are reported in parts per million (δ) with respect to tetramethylsilane (TMS, δ =0), and referenced internally with respect to the protio solvent impurity. Deuterated NMR solvents were obtained from Cambridge Isotope Laboratories, Inc., Andover, MA, and used without further purification. Mass spectra were recorded on an Agilent 6210 LC TOF mass spectrometer using electrospray ionization with fragmentation voltage set at 115v and processed with an Agilent MassHunter Operating System. All other materials were obtained from Aldrich Chemical Company, St. Louis, MO, or TCI, Tokyo, Japan and were used as received. Solvents were dried through a commercial solvent purification system (Pure Process Technologies, Inc.). The synthesis and characterization of ligands **A** and **B**, as well as complexes **1** and **2**, are described in our previous report.¹ X-ray crystallographic data for homocomplexes **1** (CCDC # 951758) and **2** (CCDC # 951759) can be found in our previous report.¹ The minimized structures of cages **3-5** were obtained via density functional calculations, and were optimized using the dispersion-corrected B97-D density functional^{2,3} in the 6-31G(d) basis set.⁴

Synthesis of Compounds



3,7-Dinitrodibenzosuberenone (*S-1*):

3,7-Dinitrodibenzosuberone (100 mg, 0.33 mmol) was added to a 50 mL round bottom flask with stir bar, followed by the addition of benzene (25 mL). 1.1 equivalents N-bromosuccinimide (64.8 mg, 0.36 mmol) was slowly added to the flask, followed by 0.1 eq benzoyl peroxide (8.0 mg, 0.03 mmol). The reaction mixture was refluxed for 12 h, after which the solid was filtered and dried. The crude solid was placed into a 50mL round bottom flask followed by acetone (25 mL). Potassium iodide (60.4 mg, 0.33mmol) was added to the flask and the mixture was stirred at room temperature for 4 h. The reaction mixture was then filtered and the product collected as a light yellow solid (47 mg, 50 %). ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.70.25 (d, *J* = 2.5 Hz, 2H), 8.53 (dd, *J* = 8.5, 2.6 Hz, 2H), 7.99 (d, *J* = 8.5 Hz, 2H),

6.58 (s, 2H). ¹³C NMR (150 MHz, DMSO- d_6): δ 180.2, 148.4, 140.0, 138.4, 134.6, 134.1 127.7, 126.3. HRMS (ESI) m/z calcd for C₁₅H₈N₂O₅ (**S-1**⁺) 296.0418, found 296.0228.



3,7-Diaminodibenzosuberenone (C):

S-1 (400 mg, 1.34 mmol) was added to a 50 mL round bottom flask with stir bar, followed by addition of Raney© 2800 Ni suspension in water (1.0 mL) and MeOH (25 mL). The flask was fixed with a septum and purged with nitrogen gas. Hydrazine monohydrate (2.0 mL, 41.2 mmol) was slowly added. After the addition, the reaction was stirred at room temperature. After 24 h the reaction mixture was diluted with acetone (100 mL) followed by filtering through celite. After evaporating the solvent *in vacuo*, the residue was triturated in deionized water (200 mL) before being filtered using celite. The filter was rinsed clean using MeOH (150 mL) before evaporating the solvent *in vacuo* to give an orange-yellow solid. This was recrystallized from EtOH to give product as an orange solid (162 mg, 50 %). ¹H NMR (400 MHz; DMSO-*d*₆) δ 7.33 (s, 2H), 7.31 (d, *J* = 5.9 Hz, 2H), 6.89 (dd, *J* = 5.9, 2.2 Hz, 2H), 6.69 (d, *J* = 2.2 Hz, 2H). ¹³C NMR (150 MHz, DMSO-*d*₆): δ 191.4, 149.8, 138.7, 133.4, 127.6, 126.0, 119.7, 113.3. HRMS (ESI) m/z calcd. for C₁₅H₁₃N₂O ([**C**•H]⁺) 237.1178, found 237.1248.



2,7-Dinitroxanthone (S-2)

Fuming nitric acid (15 mL) and concentrated sulfuric acid (10 mL) were added to a 250 mL round bottom flask and chilled to 0 °C. Xanthone (2 g, 10.1 mmol) was slowly added to the solution in 200 mg portions with vigorous stirring over a period of 30 minutes. The yellow solution was allowed to stir for an additional hour at 0 °C. The mixture was then poured into a beaker containing 150 g ice with vigorous stirring and the precipitate was filtered. The pale yellow crude product was then recrystallized from nitromethane to yield an off white solid (1.98 g 68 %) ¹H NMR (400 MHz, DMSO-*d*₆): δ 9.25 (d, *J* = 2.8 Hz, 2H), 8.65 (dd, *J* = 9.2, 2.8 Hz 2H), 7.75 (d, *J* = 9.2 Hz, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆): δ 175.6, 159.5, 144.8, 131.0, 122.7, 121.8, 121.5. HRMS (ESI) m/z calcd. for C₁₃H₆N₂O₆ (**S**-2⁺) 286.0257, found 286.0228.



2,7-Diaminoxanthone (D)

S-2 (100 mg, 0.33 mmol) was placed in a 50 mL round bottom flask with a stir bar followed by 10 mL of concentrated hydrochloric acid. Then, 4.5 equivalents of tin(II)chloride dihydrate was added to the flask. The reaction was refluxed for 12 h with stirring, then allowed to cool to room temperature. The reaction mixture was diluted with 50mL deionized water and brought to a pH of 8.5 using 2 M NaOH. The yellow solution was then extracted using ethyl acetate until no color was present in the aqueous layer (3 x 15 mL). The solution was dried using anhydrous MgSO4, filtered and the solvent removed in vacou to yield an orange solid (57 mg, 72 %). ¹H NMR (400 MHz, DMSO-*d*₆): δ 7.33 (d, *J* = 8.8 Hz, 2H), 7.22 (d, *J* = 2.8 Hz, 2H), 7.07 (dd, *J* = 9.0 Hz, 2H), 5.32 (s, 4H). ¹³CNMR (100 MHz, DMSO-*d*₆): δ 176.8, 148.7, 145.1, 123.9, 122.1, 119.2, 106.8. HRMS (ESI) m/z calcd. for C₁₃H₁₁N₂O₂ ([**D**•H]⁺) 227.0713, found 227.0628.



Cage 3:

Dianiline **C** (38.3 mg, 0.161 mmol), 2-pyridine carboxaldehyde (27.0 µL, 0.32 mmol) and Fe(ClO₄)₂•xH₂O (35.1 mg) were combined in anhydrous MeCN (5 mL) in a 50 mL round-bottomed flask under a blanket of N₂ gas. The solution was then heated at 45 °C for 10 h with stirring. The purple solution was diluted with Et₂O (25 mL), cooled to -25 °C followed by filtration of the resulting precipitate. After drying, the product was isolated as a purple solid (65.0 mg, 92 %) ¹H NMR (400 MHz; CD₃CN) δ 8.74 (s, 2H), 8.50 (d, *J* = 7.4 Hz, 2H), 8.44 (t, *J* = 7.3 Hz, 2H), 7.8 (t, *J* = 5.2 Hz, 2H), 7.56 (d, *J* = 8.3 Hz, 2H), 7.43 (d, *J* = 5.3 Hz, 2H), 7.2 (s, 2H), 6.56 (d, *J* = 2.2 Hz, 2H), 5.62 (dd, *J* = 8.2, 2.2 Hz, 2H). ¹³C NMR (150 MHz, DMSO-*d*₆): δ 186.4, 176.4, 158.6, 156.4, 150.4, 140.4, 138.5, 135.8, 134.8, 132.7, 131.8, 130.4, 125.5, 122.9. HRMS (ESI) m/z calcd. for C₈₁H₆₀Fe₂N₁₂O₃ ([**3**]⁴⁺) 339.0897, found 339.0802



Cage 4:

Dianiline **D** (100 mg, 0.44 mmol), 2-pyridine carboxaldehyde (107 µL, 0.88 mmol) and Fe(ClO₄)₂•xH₂O (96.3 mg) were combined in anhydrous MeCN (10 mL) in a 50 mL round-bottomed flask under a blanket of N₂, followed by heating to 45 °C for 10 h. The solution was then cooled to room temperature, diluted with Et₂O (30 mL), and cooled to -25 °C followed by filtration of the resulting precipitate. Drying product *in vacuo* gave product as a purple solid (190 mg, 96 %). ¹H NMR (400 MHz; CD₃CN) δ 11.13 (m, 2H), 9.31 (s, 2H), 8.98 (m, 2H), 8.48 (s, 2H), 8.34 (s, 2H), 7.33 (m, 2H), 6.39 (s, 2H), 5.44 (d, J = 8.5 Hz, 2H). Cage **4** was not soluble enough to collect a ¹³C NMR spectrum within a reasonable amount of time. HRMS (ESI) m/z calcd for C₇₅H₅₄Fe₂N₁₂O₁₄Cl₂ ([**4**•(ClO₄)₂]²⁺) 761.7364, found 761.9462.

NMR Spectral Data



8.0 9.0 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 7.9 7.8 (ppm) 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.6 6.8 6.7 Figure S1: ¹H NMR spectrum of *S-1* (DMSO, 400 MHz, 298 K).



Figure S2: ¹³C NMR spectrum of *S-1* (DMSO, 150 MHz, 298 K).



Figure S3: ¹H NMR spectrum of C (DMSO, 400 MHz, 298 K).





8.9 8.7 9.3 8.5 8.3 7.9 7.7 7.5 (ppm) 9.1 8.1 7.1 6.9 6.7 6.5 6.3 6.1 5.9 5.7 **Figure S5:** ¹H NMR spectrum of *S*-2 (DMSO, 400 MHz, 298 K).



¹⁷⁸ 176 174 172 170 168 166 164 162 160 158 156 154 152 (ppm) 148 146 144 142 140 138 136 134 132 130 128 126 124 122 120 **Figure S6:** ¹³C NMR spectrum of *S-2* (DMSO, 150 MHz, 298 K).





¹⁹⁰ ¹⁸⁵ ¹⁸⁰ ¹⁷⁵ ¹⁷⁰ ¹⁶⁵ ¹⁶⁰ ¹⁵⁵ ¹⁵⁰ ¹⁴⁵ ¹⁴⁰ (ppm) ¹³⁰ ¹²⁵ ¹²⁰ ¹¹⁵ ¹¹⁰ ¹⁰⁵ ¹⁰⁰ ⁹⁵ ⁹⁰ ⁸⁵ ⁸⁰ **Figure S8:** ¹³C NMR spectrum of **D** (DMSO, 150 MHz, 298 K).





150 (ppm) 140 Figure S10: ¹³C NMR spectrum of Cage 3 (CD₃CN, 150 MHz, 298 K).



Figure S11: ¹H NMR scans of Cage 3 at various temperatures (CD₃CN, 600 MHz, 298-338 K).



^{11.0} 10.6 10.2 9.8 9.4 9.0 8.6 8.2(ppm) 7.8 7.4 7.0 6.6 6.2 5.8 5.4 **Figure S12:** ¹H NMR spectrum of Cage **4** (CD₃CN, 400 MHz, 298 K).



Mass Spectral Data





Assembly Mixing Experiments

General mixing procedure: All mixing experiments were performed in an NMR tube. One equivalent of dianiline **A** (3.5 mg, 0.015 mmol) and one equivalent of dianiline **B** (3.5 mg 0.015 mmol) were placed in an NMR tube. Deuterated acetonitrile (400 μ L) was added to the tube and a proton spectrum of the dianiline mixture obtained. 2 equivalents of 2-formylpyridine were added (5 μ L, 0.029 mmol) followed by 0.66 equivalents of iron perchlorate (100 μ L of 0.098 M Fe(ClO₄)₂•xH₂O in CD₃CN). A spectrum of the mixture was obtained. The tube was heated at 80 °C for 8 h. Another spectrum was taken after heating to show the favored cage and the unfavored dianiline ligand. A second 2 eq. of 2-formylpyridine and 0.66 eq. of iron perchlorate were added to the tube and a proton spectrum obtained. The tube was heated to 80 °C for 8 h. A final spectrum was obtained after heating to show both cages in solution.

Mixing Experiment Between Ligands A and C



Figure S16: ¹H NMR spectra of mixing experiment between ligands **A** and **C** (CD₃CN, 400 MHz, 298K). i) Cage **3** (red), h) Cage **1** (blue), g) Dianiline **C** (orange), f) Dianiline **A** (green), e) Dianiline mixture, d) Mixture with first addition of 2-formylpyridine and iron with no heat, c) Mixture with first addition after 8 h at 80 °C, b) Mixture after second addition of 2-formylpyridine and iron with no heat, a) mixture after second addition and 8 h heat at 80 °C.

Mixing Experiment Between Ligands A and B



Figure S17: ¹H NMR spectra of mixing experiment between ligands **A** and **B** (CD₃CN, 400 MHz, 298K). i) Cage **2** (red), h) Cage **1** (blue), g) Dianiline **A** (green), f) Dianiline **B** (orange), e) Dianiline mixture, d) Mixture with first addition of 2-formylpyridine and iron with no heat, c) Mixture with first addition after 8 h at 80 °C, b) Mixture after second addition of 2-formylpyridine and iron with no heat, a) mixture after second addition and 8 h heat at 80 °C.

Mixing Experiment Between Ligands B and C



Figure S18: ¹H NMR spectra of mixing experiment between ligands **B** and **C** (CD₃CN, 400 MHz, 298K). i) Cage **3** (red), h) Cage **2** (blue), g) Dianiline **C** (orange), f) Dianiline **B** (green), e) Dianiline mixture, d) Mixture with first addition of 2-formylpyridine and iron with no heat, c) Mixture with first addition after 8 h at 80 °C, b) Mixture after second addition of 2-formylpyridine and iron with no heat, a) mixture after second addition and 8 h heat at 80 °C.





Figure S19: ¹H NMR spectra of mixing experiment between ligands **A** and **D** (CD₃CN, 400 MHz, 298K). i) Cage **1** (red), h) Cage **4** (blue), g) Dianiline **A** (orange), f) Dianiline **D** (green), e) Dianiline mixture, d) Mixture with first addition of 2-formylpyridine and iron with no heat, c) Mixture with first addition after 8 h at 80 °C, b) Mixture after second addition of 2-formylpyridine and iron with no heat, a) mixture after second addition and 8 h heat at 80 °C.

Mixing Experiment Between Ligands C and D



Figure S20: ¹H NMR spectra of mixing experiment between ligands C and D (CD₃CN, 400 MHz, 298K). i) Cage 3 (red), h) Cage 4 (blue), g) Dianiline C (orange), f) Dianiline D (green), e) Dianiline mixture, d) Mixture with first addition of 2-formylpyridine and iron with no heat, c) Mixture with first addition after 8 h at 80 °C, b) Mixture after second addition of 2-formylpyridine and iron with no heat, a) mixture after second addition and 8 h heat at 80 °C.

Mixing Experiment Between Ligands B and D



Figure S21: ¹H NMR spectra of mixing experiment between ligands **B** and **D** (CD₃CN, 400 MHz, 298K). i) Cage **2** (red), h) Cage **4** (blue), g) Dianiline **B** (orange), f) Dianiline **D** (green), e) Dianiline mixture, d) Mixture with first addition of 2-formylpyridine and iron with no heat, c) Mixture with first addition after 8 h at 80 °C, b) Mixture after second addition of 2-formylpyridine and iron with no heat, a) mixture after second addition and 8 h heat at 80 °C.



Figure S22: ¹H-DOSY NMR spectrum of mixing between dianilines **A** and **C** after 1 addition of Fe(ClO₄)₂ and 2-formylpyridine and 8 h heat at 80°C (CD₃CN, 600 MHz, 298 K, $\Delta = 100$ ms, $\delta = 2.6$ µs, Diffusion Coefficient = 8.39 x10⁻¹⁰ m²/s for cage **1** vs. 2.15 x10⁻⁹ m²/s for dianiline **C** vs. 4.92 x10⁻⁹ m²/s for solvent).



Figure S23: ¹H-DOSY NMR spectrum of mixing between dianilines **A** and **B** after 1 addition of Fe(ClO₄)₂ and 2-formylpyridine and 8 h heat at 80°C (CD₃CN, 600 MHz, 298 K, $\Delta = 100$ ms, $\delta = 2.6$ µs, Diffusion Coefficient = 7.21 x10⁻¹⁰ m²/s for cage **1** vs. 1.91 x10⁻⁹ m²/s for dianiline **B** vs. 4.16 x10⁻⁹ m²/s for solvent).



8.8 8.6 8.2 8.0 7.8 7.2 7.0 6.8 6.6 (ppm) 6.2 6.0 5.8 3.2 3.0 2.8 2.6 2.4 2.2 2.0 8.4 7.6 7.4 1.8 Figure S24: ¹H-DOSY NMR spectrum of mixing between dianilines **B** and **C** after 1 addition of Fe(ClO₄)₂ and 2-formylpyridine and 8 h heat at 80°C (CD₃CN, 600 MHz, 298 K, $\Delta = 100$ ms, $\delta = 2.6$ μ s, Diffusion Coefficient = 9.23 x10⁻¹⁰ m²/s for cage **3** vs. 1.66 x10⁻⁹ m²/s for dianiline **B** vs. 5.57 x10⁻⁹ m^2/s for solvent).





Figure S26: ESI-MS of mixing experiment between dianilines A and B after 1 addition of $Fe(ClO_4)_2$ and 2-formylpyridine and 8h heat at 80°C. Only Cage 1 and ligand B



Figure S27: ESI-MS of mixing experiment between dianilines **B** and **C** after 1 addition of $Fe(ClO_4)_2$ and 2-formylpyridine and 8 h heat at 80°C. Only Cage **3** and ligand **B** were detected.

Ligand Displacement Experiments

General displacement procedure: All displacement experiments were performed in an NMR tube. One equivalent of preformed Cage **1** (8.6 mg, 0.005 mmol) and three equivalents of dianiline **B** (3.5 mg 0.015 mmol) were placed in an NMR tube. Dry deuterated acetonitrile (400 μ L) was added to the tube and a proton spectrum of the starting mixture obtained. The tube was heated at 80°C for 8 h. A second spectrum was obtained after heating to verify whether the preformed cage was displaced by the free dianiline ligand. Experiments were repeated with the addition of 6 molar equivalents of water and heated at 55°C for 1 hour.



Figure S28: ¹H NMR spectra of anhydrous displacement experiment between cage **3** and dianiline **A** (CD₃CN, 400 MHz, 298K). Top: Cage **1** (blue) and displaced dianiline **C** (orange) after heating at 80 °C for 8 h. Bottom: cage **3** (red) and dianiline **A** (green) prior to heating. The reverse experiment (cage **1** ligand displacement by dianiline **C**) showed no change in the spectrum even after heating for 24 h.



Figure S29: ¹H NMR spectra of displacement reaction between cage **3** and dianiline **A** (CD₃CN, 600 MHz, 343 K).



Figure S30: ¹H NMR spectra of displacement experiment between cage **2** and dianiline **A** (CD₃CN, 400 MHz, 298K). Top: cage **1** and displaced Dianiline **B** after heating at 80 °C for 8 h. Bottom: cage **2** and dianiline **A** prior to heating. The reverse experiment (cage **1** ligand displacement by dianiline **B**) showed no change in the spectrum even after heating for 24 h.



Figure S31: ¹H NMR spectra of anhydrous displacement experiment between cage **2** and dianiline **C** (CD₃CN, 400 MHz, 298K). Top: cage **3** and displaced dianiline **B** after heating at 80 °C for 8 h. Bottom: cage **2** and dianiline **C** prior to heating. The reverse experiment (cage **3** displacement by dianiline **B**) showed no change in the spectrum even after heating for 24 h.

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