Electronic Supplementary Information

Strongly Oxidizing Perylene-3,4-dicarboximides for Use in Water Oxidation Photoelectrochemical Cells

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I. Synthesis

¹H NMR and ¹³C NMR spectra were recorded at ambient temperature at 500 MHz and 126 MHz, respectively, using a Bruker Avance III spectrometer. Mass spectrometry was performed using a Bruker Autoflex III MALDI-TOF instrument. Unless otherwise noted, all reagents were obtained from commercial suppliers and used without further purification. Reagents P1,¹ P3,¹ P4,² P5,³ P6,⁴ 2-phenylpyridin-4-ylamine (NH₂-ppy),⁵ and [Cp*IrCl₂]₂⁶ were synthesized according to reported methods.



Scheme S1. Synthesis of 1A-4A and 1B-4B^a

^{*a*}Reaction conditions: (a, **P2**) (1) 4-*tert*-butylphenol, Cs₂CO₃, DMF, 160 °C, 75 min, (2) HOAc, RT, 1.5 h, 80%; (a, **P3**) previously reported.¹ (b) (1) KOH, (KF·2H₂O), *t*-BuOH, 70-105 °C, 35-120 min; (2) HOAc, 2 N HCl, RT, overnight, 7-68%. (c, **1B**, **2B**) 1-bromohexane, K₂CO₃, 1-hexanol, 90 °C, 19-29 h, 44-51%. (d) (1) KOH, *t*-BuOH, 100 °C, 20 min; (2) HOAc, 2 N HCl, RT, overnight; (3) 1-bromohexane, K₂CO₃, 1-hexanol, 90 °C, 13 h, 17%. (e) 5-bromoisophthalate, K₂CO₃, Pd(PPh₃)₄, toluene/H₂O/EtOH,80 °C, 13 h, 97%. (f) (1) KOH, THF, H₂O, 80 °C, 12 h; (2) 2 N HCl, 97%.



Compound P2.⁷ Compound **P1** (804 mg, 0.926 mmol), 4-*tert*-butylphenol (461 mg, 3.07 mmol), and Cs₂CO₃ (560 mg, 1.72 mmol), were combined in DMF (32 mL) and heated to 160 °C under N₂. After 75 min, the mixture was cooled, and 5 mL acetic acid was added and stirred for 1.5 h. Brine was added until a precipitate was formed, and the crude product was collected by filtration. Column chromatography on a silica column with 4:1 CH₂Cl₂/hexanes as the eluent yielded the product as a mixture of isomers with the 1,6:1,7 isomers in a 1:5.4 ratio (749 mg, 80%). For the 1,7 isomer (not isolated): ¹H NMR (500 MHz, CDCl₃) δ 9.66 (d, *J* = 8.3 Hz, 2H), 8.69 (d, *J* = 8.3 Hz, 2H), 8.45 (s, 2H), 7.50 – 7.43 (m, 6H), 7.32 (d, *J* = 7.8 Hz, 4H), 7.11 (d, *J* = 8.8 Hz, 4H), 2.72 (hept, *J* = 6.7 Hz, 4H), 1.35 (s, 18H), 1.17 – 1.12 (m, 24H). ¹³C NMR (125 MHz, CDCl₃): δ 163.51, 162.99, 155.31, 152.64, 148.02, 145.59, 133.81, 130.81, 130.44, 129.73, 129.65, 129.02, 127.46, 125.88, 124.78, 124.57, 124.07, 123.99, 122.28, 118.70, 34.55, 31.48, 29.18, 24.06, 24.02. HRMS (MALDI) m/z calcd for C₆₈H₆₇N₂O₆ [M + H]⁺ 1007.500; found 1007.507.



Compound 1A. KOH (35 mg, 0.62 mmol) was added to a solution of **P2** (57 mg, 0.056 mmol) in 5 mL *t*-BuOH and heated over 50 min. to 70 °C under N₂. The reaction mixture was then cooled, and 7 mL acetic acid and 3.5 mL 2 N HCl were added and stirred overnight. The crude product was collected by filtration then purified on a silica column with 1% acetic acid/15% hexanes/84% CH₂Cl₂ as eluent followed by HPLC to separate the 1,6 and 1,7 isomers with 40% hexanes/60% CH₂Cl₂ as eluent to yield the pure product (15 mg, 31%). ¹H NMR (500 MHz, CDCl₃) δ 9.74 – 9.62 (m, 2H), 8.70 (d, *J* = 8.3 Hz, 1H), 8.62 (d, *J* = 8.4 Hz, 1H), 8.43 (s, 1H), 8.37 (s, 1H), 7.52 – 7.43 (m, 5H), 7.31 (d, *J* = 7.8 Hz, 2H), 7.16 – 7.08 (m, 4H), 2.70 (hept, *J* = 6.7 Hz, 2H), 1.38 (s, 9H), 1.36 (s, 9H), 1.81 – 1.11 (m, 12H). ¹³C NMR (125 MHz, CDCl₃): δ 163.47, 162.97, 159.96, 156.27, 155.63, 152.50, 152.30, 148.77, 148.65, 145.67, 135.25, 133.25, 132.45, 130.78, 130.42, 129.96, 129.82, 129.79, 129.74, 129.07, 127.80, 127.71, 127.41, 125.83, 125.70, 125.56, 124.65, 124.20, 123.48, 122.90, 119.51, 119.29, 119.21, 117.95, 34.74, 34.72, 31.58, 29.30, 24.17, 24.11. MS (MALDI) m/z calcd for C₅₆H₅₀NO₇ [M + H]⁺ 848.36; found 848.28.



Compound 1B. To a solution of **1A** (11 mg, 0.013 mmol) and 1-bromohexane (0.03 mL, 0.2 mmol) in 2 mL 1-hexanol was added K₂CO₃ (13 mg, 0.094 mmol). The reaction flask was heated to 90 °C under N₂. After 29 h, the mixture was cooled to RT and chloroform was added. This mixture was extracted 4 \times H₂O, and solvent was removed under vacuum. Column chromatography on a silica column with 20% hexanes/80% CH₂Cl₂ as the eluent yielded the pure product (5.7 mg, 44%). ¹H NMR (500 MHz, CDCl₃) δ 9.41 – 9.37 (m, 2H), 8.62 (d, J = 8.3 Hz, 1H), 8.39 (s, 1H), 8.05 (d, J = 8.2 Hz, 1H), 7.79 (s, 1H), 7.48 – 7.38 (m, 5H), 7.30 (d, J = 7.8 Hz, 2H), 7.09 - 7.04 (m, 4H), 4.30 (t, J = 6.9 Hz, 2H), 4.24 (t, J = 6.9 Hz, 2H), 2.71 (hept, J = 6.8Hz, 2H), 1.81 - 1.73 (m, 2H), 1.73 - 1.65 (m, 2H), 1.49 - 1.39 (m, 4H), 1.37 - 1.25 (m, 8H), 1.35 (s, 9H), 1.33 (s, 9H), 1.16 – 1.11 (m, 12H), 0.95 - 0.81 (m, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 168.53, 167.79, 163.83, 163.29, 153.91, 153.23, 153.02, 147.69, 147.55, 145.74, 134.45, 132.80, 132.25, 131.04, 130.96, 130.89, 130.78, 129.63, 129.55, 129.33, 128.94, 128.54, 127.98, 127.39, 127.32, 125.98, 125.87, 125.38, 124.99, 124.12, 122.93, 121.63, 121.27, 118.66, 118.49, 66.00, 65.91, 34.60, 31.65, 31.59, 31.53, 29.24, 28.70, 28.44, 25.82, 25.64, 24.16, 24.11, 22.70, 22.64, 14.19, 14.16. HRMS (MALDI) m/z calcd for $C_{68}H_{76}NO_8$ [M + H]⁺ 1034.557; found 1034.555.



Compound 2A. Compound **P3** (20 mg, 0.019 mmol), KOH (34 mg, 0.61 mmol), and KF·2H₂O (51 mg, 0.54 mmol) in 2 mL *t*-BuOH were heated to 105°C under N₂. After 2 h, the mixture was cooled to RT and 3 mL acetic acid and 1.5 mL 2 N HCl were added. The reaction mixture was stirred overnight, and the crude product was collected by filtration. Column chromatography on a silica column with 1% ethyl acetate/ 29% hexanes/ 70% CH₂Cl₂ as the eluent yielded the product as a mixture of isomers (12 mg, 68%). For the 1,7 isomer: ¹H NMR (500 MHz, CDCl₃) δ 8.74 (s, 1H), 8.71 (s, 1H), 8.17 (d, *J* = 8.1 Hz, 1H), 8.13 (d, *J* = 8.2 Hz, 1H), 7.82 (d, *J* = 8.4 Hz, 1H), 7.80 (d, *J* = 8.4 Hz, 1H), 7.58 – 7.55 (m, 2H), 7.48 (t, *J* = 7.8 Hz, 1H), 7.39 (d, *J* = 1.4 Hz, 2H), 7.33 (d, *J* = 7.8 Hz, 2H), 2.71 (hept, *J* = 7.0 Hz, 2H), 1.33 (s, 36H), 1.18 (d, *J* = 6.8 Hz, 6H), 1.12 (d, *J* = 6.8 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 163.52, 163.48, 160.41, 160.12, 153.49, 145.72, 143.45, 142.80, 141.02, 140.76, 137.38, 137.09, 135.78, 134.97, 134.55, 132.42, 131.22, 130.89, 130.67, 130.31, 130.09, 129.81, 129.76, 129.70, 128.31, 124.21, 123.41, 123.34, 122.97, 122.78, 122.33, 118.24, 117.57, 35.29, 35.27, 31.60, 31.52, 29.39, 24.06, 24.04. HRMS (MALDI) m/z calcd for C₆₄H₆₆NO₅ [M + H]⁺ 928.494; found 928.496.



Compound 2B. To a solution of **2A** (15 mg, 0.016 mmol) in 2 mL 1-hexanol was added 1bromohexane (0.03 mL, 0.2 mmol) and K₂CO₃ (17 mg, 0.12 mmol). The reaction flask was heated to 90 °C under N₂. After 19 h, the mixture was cooled to RT and dried under vacuum. Column chromatography on a silica column with 30% hexanes/70% CH₂Cl₂ as the eluent yielded the product as a mixture of isomers (9.4 mg, 51%). For the 1,7 isomer: ¹H NMR (500 MHz, $CDCl_3$) δ 8.69 (s, 1H), 8.15 (s, 1H), 8.09 (d, J = 8.1 Hz, 1H), 7.64 (d, J = 8.0 Hz, 1H), 7.60 (d, J = 8= 8.2 Hz, 1H), 7.57 (d, J = 8.1 Hz, 1H), 7.51 - 7.44 (m, 3H), 7.37 (d, J = 1.4 Hz, 2H), 7.34 (d, J = 1.4 Hz, 1H), 7.34 (d, J = 1.4= 1.4 Hz, 2H), 7.32 (d, J = 7.8 Hz, 2H), 4.32 (t, J = 6.8 Hz, 2H), 4.28 (t, J = 6.9 Hz, 2H), 2.72 (hept, J = 6.6 Hz, 2H), 1.82 - 1.68 (m, 4H), 1.48 - 1.36 (m, 4H), 1.36 - 1.26 (m, 8H), 1.30 (s, 36H), 1.18 (d, J = 6.8 Hz, 6H), 1.11 (d, J = 6.8 Hz, 6H), 0.95 – 0.78 (m, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 168.54, 168.41, 163.93, 163.89, 152.84, 145.78, 141.57, 141.52, 141.44, 141.22, 136.45, 135.60, 134.33, 134.20, 132.56, 131.88, 131.05, 131.02, 130.88, 130.35, 130.30, 130.16, 129.95, 129.67, 129.60, 129.56, 128.67, 128.09, 127.28, 124.12, 123.86, 123.76, 122.05, 121.97, 121.47, 120.57, 65.95, 65.76, 35.16, 31.65, 31.60, 31.56, 31.51, 31.47, 29.30, 28.66, 28.64, 25.85, 25.74, 24.05, 22.70, 22.69, 14.18. MS (MALDI) m/z calcd for $C_{76}H_{92}NO_6$ [M + H]⁺ 1114.69; found 1114.73.



Compound 3A. Compound **P4** (200 mg, 0.376 mmol) and KOH (63 mg, 1.1 mmol) were dissolved in 30 mL *t*-BuOH and heated to 100 °C for 35 min. 20 mL acetic acid and 10 mL 2N HCl were added to the cooled reaction mixture, and it was stirred at room temperature overnight. The precipitated crude product was then collected by filtration, rinsed with H₂O, and dried on the filter paper with vacuum. The dried precipitate was then rinsed thoroughly with CH₂Cl₂. After solvent removal, the filtrate was subjected to column chromatography on a silica column with 2% acetone/98% CH₂Cl₂ \rightarrow 5% acetone/10% acetic acid/85% CH₂Cl₂. The fractions were extracted 4×H₂O before solvent removal under vacuum. The isolated material was further purified by dissolution in CH₂Cl₂ and addition of hexanes until precipitate began to form. Isolation of the precipitate by filtration yielded pure product (13 mg, 7%). ¹H NMR (500 MHz, CDCl₃) δ 8.79 – 8.63 (m, 8H), 5.12 – 5.01 (m, 1H), 2.33 – 2.20 (m, 2H), 2.01 – 1.88 (m, 2H), 0.93 (t, *J* = 7.5 Hz, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 160.18, 136.60, 133.86, 133.78, 124.11, 123.37, 119.18, 57.98, 25.13, 11.49. MS (MALDI) m/z calcd for C₂₉H₁₉NO₅ [M]⁺ 461.13; found 461.07.



Compound 3B. Compound P4 (99 mg, 0.19 mmol) and KOH (162 mg, 2.88 mmol) were added to 15 mL t-BuOH at 110 °C and stirred for 20 minutes under nitrogen. The reaction mixture was cooled, 20 mL acetic acid and 10 mL 2 N HCl were added, and it was stirred overnight. The precipitate was collected by filtration with water and once dry rinsed with 50 mL CH₂Cl₂. The filtrate was collected and the volume reduced to 5 mL. 30 mL hexanes was added, and the precipitate was collected by filtration. It was dissolved again in CH₂Cl₂ and the process was repeated three more times. This resulting mixture enriched in 3A (62 mg) was then dissolved in 10 mL pyridine. K₂CO₃ (186 mg, 1.35 mmol), 1-bromohexane (0.24 mL, 1.71 mmol), and 1hexanol (10 mL) were added and the reaction mixture was purged with nitrogen. It was heated to 90 °C for 13 hours then cooled, filtered, and the precipitate rinsed thoroughly with CH₂Cl₂. The filtrate was extracted 4×H₂O and the solvent removed under vacuum. Column chromatography on a silica column with 5% ethyl acetate/50% hexanes/45% CH₂Cl₂ as the eluent yielded the pure product (21 mg, 17%). ¹H NMR (500 MHz, CDCl₃) δ 8.49 (d, J = 8.0 Hz, 2H), 8.26 (t, J = 8.2 Hz, 4H), 8.00 (d, J = 7.9 Hz, 2H), 5.12 – 5.02 (m, 1H), 4.35 (t, J = 6.9 Hz, 4H), 2.34 – 2.21 (m, 2H), 2.03 - 1.89 (m, 2H), 1.87 - 1.77 (m, 4H), 1.53 - 1.42 (m, 4H), 1.41 - 1.29 (m, 8H),0.98 – 0.87 (m, 12H). ¹³C NMR (126 MHz, CDCl₃) δ 168.39, 135.20, 132.15, 131.90, 130.35, 129.41, 129.27, 129.13, 125.90, 122.52, 121.85, 66.00, 57.63, 31.67, 28.70, 25.83, 25.17, 22.71, 14.17, 11.54. MS (MALDI) m/z calcd for C₄₁H₄₆NO₆ [M]⁺ 647.32; found 647.26.



Compound 4B. K₂CO₃ (881 mg, 6.37 mmol) in 3 mL H₂O was added to a mixture of **P7** (166 mg, 0.273 mmol) and dimethyl 5-bromoisophthalate (50 mg, 0.18 mmol) in 7.7 mL toluene and 0.6 mL ethanol. The mixture was purged with nitrogen and Pd(PPh₃)₄ (15 mg, 0.013 mg) was added. The mixture was stirred at 80 °C under nitrogen for 13 hours and then cooled to room temperature. Chloroform was added to the cooled reaction mixture, it was extracted $3 \times H_2O$, and solvent was removed under vacuum. Column chromatography on a silica column with 1% acetone/30% hexanes/69% CH₂Cl₂ as the eluent yielded the pure product (119 mg, 97%). ¹H NMR (500 MHz, CDCl₃) δ 8.82 (t, *J* = 1.6 Hz, 1H), 8.73 – 8.66 (m, 2H), 8.58 – 8.48 (m, 4H), 8.43 (d, *J* = 1.6 Hz, 2H), 7.84 (d, *J* = 8.4 Hz, 1H), 7.67 – 7.59 (m, 2H), 7.49 (dd, *J* = 7.8 Hz, 1H), 7.35 (d, *J* = 7.8 Hz, 2H), 4.00 (s, 6H), 2.78 (hept, *J* = 6.8 Hz, 2H), 1.19 (d, *J* = 6.8 Hz, 12H). ¹³C NMR (126 MHz, CDCl₃) δ 166.18, 164.10, 145.82, 140.96, 140.79, 137.63, 137.34, 135.26, 132.54, 132.27, 132.25, 131.28, 131.11, 130.66, 130.28, 129.75, 129.61, 129.55, 128.64, 128.52, 127.73, 127.09, 124.23, 124.18, 123.53, 121.35, 121.30, 120.73, 120.58, 52.78, 29.28, 24.16. HRMS (MALDI) m/z calcd for C₄₄H₃₆NO₆ [M + H]⁺ 674.254; found 674.256.



Compound 4A. To **4B** (41 mg, 0.061 mmol) in THF was added KOH (38 mg, 0.67 mmol) in 0.5 mL H₂O. After stirring at 80 °C for 12 h under nitrogen, the reaction mixture was cooled and the 10 mL 2 N HCl was added. The crude product was collected by filtration and rinsed with H₂O. The product was purified on a silica column with 50% acetone/50% CH₂Cl₂ \rightarrow 5% acetic acid/50% acetone/45% CH₂Cl₂ as the eluent. Fractions were extracted 4 × H₂O before solvent removal under vacuum to yield the product (33 mg, 97%). ¹H NMR (500 MHz, THF-d8) δ 8.85 – 8.71 (m, 5H), 8.65 – 8.61 (m, 2H), 8.42 (d, *J* = 1.6 Hz, 2H), 7.91 (d, *J* = 8.4 Hz, 1H), 7.71 (d, *J* = 7.7 Hz, 1H), 7.68 (t, *J* = 8.0 Hz, 1H), 7.43 – 7.36 (m, 1H), 7.29 (d, *J* = 7.7 Hz, 2H), 2.79 (hept, *J* = 6.8 Hz, 2H), 1.14 (d, *J* = 6.8 Hz, 12H). ¹³C NMR (126 MHz, THF-d8) δ 166.87, 164.35, 146.86, 142.28, 141.54, 138.43, 138.19, 135.73, 133.53, 132.96, 132.73, 132.53, 131.49, 131.09, 130.70, 130.37, 129.66, 129.55, 129.40, 129.38, 128.54, 127.97, 125.35, 124.83, 124.35, 122.48, 121.85, 121.76, 30.01, 25.86. HRMS (MALDI) m/z calcd for C₄₂H₃₂NO₆ [M + H]⁺ 646.223; found 646.226.





^{*a*}Reaction conditions: (a) 2-phenylpyridin-4-ylamine, $Zn(OAc)_2$, pyridine, 100 °C, 48 h, 73%. (b) (1) KOH, *t*-BuOH, 70-80 °C, 70 min; (2) HOAc, 2 N HCl, RT, overnight, 17%. (c) $[Cp*IrCl_2]_2$, NaOAc·3H₂O, CH₂Cl₂, RT, 36 h, 58%. (d) 1-bromohexane, K₂CO₃, 1-hexanol, 90 °C, 50 h, 23%. (e) $[Cp*IrCl_2]_2$, NaOAc·3H₂O, CH₂Cl₂, RT, 24 h, 90%.



Compound P7. Compound **P6** (50 mg, 0.073 mmol) was dissolved in 10 mL pyridine and heated to 100 °C. **NH₂-ppy** (95 mg, 0.56 mmol) and Zn(OAc)₂ (64 mg, 0.35 mmol) were added and the reaction mixture was stirred for 48 hours. After solvent removal under vacuum, column chromatography on a silica column with 50% ethyl acetate/50% CH₂Cl₂ as the eluent yielded the pure product (53 mg, 73%). ¹H NMR (500 MHz, CDCl₃) δ 9.56 (d, *J* = 8.4 Hz, 2H), 8.80 (dd, *J* = 5.1, 0.7 Hz, 2H), 8.57 (d, *J* = 8.4 Hz, 2H), 8.31 (s, 2H), 7.96 – 7.87 (m, 4H), 7.78 – 7.76 (m, 2H), 7.52 – 7.45 (m, 4H), 7.41 – 7.37 (m, 6H), 7.21 (dd, *J* = 5.2, 1.9 Hz, 2H), 7.15 – 7.07 (m, 4H), 1.35 (s, 18H). ¹³C NMR (126 MHz, CDCl₃) δ 162.78, 162.45, 159.46, 155.95, 152.31, 150.97, 148.88, 143.60, 138.60, 133.89, 130.56, 129.43, 129.30, 129.11, 128.81, 127.80, 127.14, 125.08, 124.06, 123.96, 123.47, 122.03, 121.89, 120.97, 119.38, 34.73, 31.56. HRMS (MALDI) m/z calcd for C₆₆H₄₉N₄O₆ [M + H]⁺ 993.365; found 993.373.



Compound P8. Compound P7 (303 mg, 0.305 mmol) and KOH (31 mg, 0.56 mmol) were dissolved in 45 mL t-BuOH at 70 °C. The mixture was then heated to 80 °C over 70 min and then cooled slightly before adding 30 mL acetic acid and 15 mL 2 N HCl. The mixture was stirred at room temperature overnight. Water was added until all the colored material was precipitated, and the precipitate was collected by filtration. Column chromatography on a silica column with 1% acetic acid/5% ethyl acetate/95% CH₂Cl₂ as the eluent yielded the product, and access starting material was recovered by eluting with 50% ethyl acetate/CH₂Cl₂. The fractions were extracted $3 \times H_2O$ before solvent removal under vacuum to yield the pure product (43 mg, 17%). ¹H NMR (500 MHz, CD₂Cl₂) δ 9.74 – 9.64 (m, 2H), 8.85 (dd, J = 5.1, 0.7 Hz, 1H), 8.64 (d, J = 8.4 Hz, 1H), 8.57 (d, J = 8.3 Hz, 1H), 8.35 (s, 1H), 8.30 (s, 1H), 8.07 - 7.99 (m, 2H), 7.75 (dd, J = 1.9, 0.7 Hz, 1H), 7.59 - 7.39 (m, 7H), 7.25 (dd, J = 5.2, 1.9 Hz, 1H), 7.21 - 7.12 (m, 7H)4H), 1.38 (s, 9H), 1.36 (s, 9H). ¹³C NMR (126 MHz, CD₂Cl₂) δ 163.22, 162.85, 160.37, 160.26, 159.46, 156.68, 156.26, 153.07, 152.96, 151.38, 149.32, 149.23, 144.45, 139.12, 135.42, 134.09, 132.51, 130.89, 130.31, 130.15, 130.03, 129.91, 129.68, 129.30, 128.18, 128.09, 127.66, 127.48, 125.89, 125.79, 125.63, 124.62, 124.54, 124.26, 123.01, 122.73, 121.11, 120.15, 119.73, 118.57, 35.06, 35.03, 31.73, 31.72. HRMS (MALDI) m/z calcd for $C_{55}H_{41}N_2O_7$ [M + H]⁺ 841.291; found 841.291.



Compound 5A. Compound P8 (10 mg, 0.012 mmol) was dissolved in 4 mL CH₂Cl₂ and purged with nitrogen. [Cp*IrCl₂]₂ (10 mg, 0.012 mmol) and NaOAc·3H₂O (5 mg, 0.03 mmol) were added, and the reaction mixture was stirred at room temperature in the dark under nitrogen for 36 hours. The crude product was purified on a silica column with $CH_2Cl_2 \rightarrow 20\% \ CH_2Cl_2/80\%$ acetone as the eluent, and the solvent was removed under vacuum. Hexanes were added to a nearly saturated solution of the isolated material in CH₂Cl₂ until a precipitate formed, which was collected by filtration to yield the pure product (8 mg, 58%). ¹H NMR (500 MHz, CDCl₃) δ 9.69 -9.63 (m, 2H), 8.83 (d, J = 6.1 Hz, 1H), 8.66 (d, J = 8.4 Hz, 1H), 8.57 (d, J = 8.4 Hz, 1H), 8.40 (s, 1H), 8.32 (s, 1H), 7.83 (dd, J = 7.6, 1.1 Hz, 1H), 7.82 (d, J = 2.1 Hz, 1H), 7.62 (dd, J = 8.0, 1.3 Hz, 1H), 7.57 - 7.45 (m, 4H), 7.21 (ddd, J = 7.4, 1.3 Hz, 1H), 7.15 - 7.08 (m, 5H), 7.04 - 7.046.96 (m, 1H), 1.70 (s, 15H), 1.39 (s, 9H), 1.36 (s, 9H). ¹³C NMR (126 MHz, CDCl₃) δ 169.18, 164.23, 162.57, 162.28, 159.80, 156.35, 156.01, 152.27, 152.24, 152.07, 149.06, 148.97, 143.74, 143.70, 136.00, 134.90, 133.71, 132.26, 131.52, 130.89, 129.79, 129.65, 129.33, 127.90, 127.83, 127.83, 127.12, 125.43, 125.25, 124.93, 124.42, 124.27, 123.88, 123.70, 122.60, 122.30, 119.66, 119.44, 119.41, 118.08, 88.93, 34.78, 34.75, 31.57, 9.09. HRMS (MALDI) m/z calcd for C₆₅H₅₅ClIrN₂O₇ [M - Cl]⁺ 1167.36; found 1167.33.



Compound P9. Compound P8 (37 mg, 0.044 mmol), 1-bromohexane (0.40 mL, 2.8 mmol), and K₂CO₃ (206 mg, 1.49 mmol) were dissolved in 8 mL 1-hexanol and heated to 90 °C for 50 hours. To the cooled reaction mixture was added CHCl₃ and 2 mL HOAc to re-dissolve the material. The mixture was filtered, the filtrate was extracted $4 \times H_2O_2$, and the solvent was removed under vacuum. Preparative TLC on silica with 1% ethyl acetate/99% CH₂Cl₂ yielded the pure product (10 mg, 23%). ¹H NMR (500 MHz, CDCl₃) δ 9.41 – 9.37 (m, 2H), 8.87 (d, J = 5.2 Hz, 1H), 8.59 (d, J = 8.4 Hz, 1H), 8.35 (s, 1H), 8.05 (d, J = 8.2 Hz, 1H), 8.02 - 7.98 (m, 2H), 7.79 (s, 1H), 7.73(d, J = 1.8 Hz, 1H), 7.48 - 7.39 (m, 7H), 7.25 (dd, J = 5.4, 2.0 Hz, 1H), 7.10 - 7.07 (m, 2H), 7.06-7.02 (m, 2H), 4.31 (t, J = 6.8 Hz, 2H), 4.24 (t, J = 6.9 Hz, 2H), 1.82 -1.73 (m, 2H), 1.73 -1.64 (m, 2H), 1.64 - 1.53 (m, 2H), 1.50 - 1.40 (m, 2H), 1.36 (s, 9H), 1.35 - 1.20 (m, 8H), 1.33(s, 9H), 0.94 - 0.79 (m, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 168.43, 167.70, 163.23, 162.88, 159.42, 154.37, 154.18, 153.13, 152.79, 150.98, 147.94, 144.08, 138.84, 134.99, 133.10, 132.15, 131.22, 130.95, 130.72, 129.43, 129.42, 129.23, 128.88, 128.84, 127.91, 127.47, 127.38, 127.25, 125.96, 125.51, 125.01, 124.87, 123.83, 122.30, 122.17, 121.12, 120.68, 118.93, 118.88, 66.02, 65.94, 34.64, 34.62, 31.64, 31.59, 31.57, 31.52, 28.70, 28.43, 25.82, 25.64, 22.70, 22.63, 14.19, 14.15. MS (MALDI) m/z calcd for $C_{67}H_{66}N_2O_8$ [M]⁻ 1026.48; found 1026.15.



Compound 5B. Compound P9 (9 mg, 0.009 mmol), [Cp*IrCl₂]₂ (8 mg, 0.009 mmol), and NaOAc 3H₂O (4 mg, 0.03 mmol) were dissolved in 2 mL CH₂Cl₂ and purged with nitrogen. The reaction mixture was stirred at room temperature in the dark for 24 hours. Column chromatography on a silica column with 5% ethyl acetate/95% CH₂Cl₂→10% acetone/90% CH₂Cl₂ as the eluent yielded the pure product (11 mg, 90%). ¹H NMR (500 MHz, CDCl₃) δ 9.43 -9.36 (m, 2H), 8.81 (d, J = 6.1 Hz, 1H), 8.59 (d, J = 8.4 Hz, 1H), 8.37 (s, 1H), 8.05 (d, J = 8.2Hz, 1H), 7.82 (d, J = 7.8 Hz, 1H), 7.81 (d, J = 2.2 Hz, 1H), 7.79 (s, 1H), 7.62 (dd, J = 7.9, 1.4Hz, 1H), 7.49 – 7.38 (m, 4H), 7.24 – 7.18 (m, 1H), 7.12 – 7.07 (m, 3H), 7.07 – 7.02 (m, 2H), 7.02 - 6.97 (m, 1H), 4.31 (t, J = 6.9 Hz, 2H), 4.24 (t, J = 6.9 Hz, 2H), 1.83 - 1.73 (m, 2H), 1.72-1.62 (m, 2H), 1.70 (s, 15H), 1.49 -1.41 (m, 2H), 1.36 (s, 9H), 1.35 -1.21 (m, 10H), 1.34 (s, 2H), 1.35 -1.21 (m, 2H), 1.35 -1.21 (m, 10H), 1.34 (s, 2H), 1.35 -1.21 (m, 2H), 1. 9H), 0.94 – 0.82 (m, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 169.09, 168.41, 167.68, 164.12, 163.01, 162.67, 154.43, 154.16, 153.22, 152.81, 151.96, 147.98, 147.94, 144.20, 143.84, 135.97, 135.08, 133.18, 132.19, 131.41, 131.31, 131.04, 130.71, 129.47, 129.24, 128.89, 127.94, 127.50, 127.40, 126.14, 125.56, 125.26, 124.90, 124.43, 123.82, 122.78, 122.28, 122.12, 121.11, 120.61, 119.56, 118.96, 118.81, 88.88, 66.01, 65.93, 34.65, 34.63, 31.65, 31.59, 31.53, 28.72, 28.45, 25.82, 25.64, 22.69, 22.63, 14.17, 14.14, 9.08. MS (MALDI) m/z calcd for C₇₇H₈₀ClIrN₂O₈ [M -Cl]⁺ 1353.55; found 1353.93.

II. Experimental details

Film Preparation.

Fluorine-doped tin oxide (FTO) coated glass plates (Hartford Glass, TEC 15) were sonicated in an Alconox solution for 15 min., rinsed with water, sonicated in ethanol for 15 min., sonicated in methanol for 15 min., then dried under a nitrogen stream. TiO₂ films were prepared by the doctor blade method using a commercially available TiO₂ paste (avg. 20 nm, Dyesol, DSL-90-T) and two parallel strips of adhesive tape as a mask. SnO₂ films were prepared according to a method reported by Bergeron et al.⁸ Poly(ethylene glycol) (0.6 g, $M_n \sim 2,000$) and 0.5 mL of a pH 11 NaHCO3(aq)/NaOH(aq) solution were combined with 10 mL of a 15% aqueous colloidal SnO2 solution (Alfa Aesar) and stirred for two hours. This paste was spin coated onto FTO films masked with tape at 2,000 rpm for 10 s. To prepare Al_2O_3 nanoparticle pastes, 1.0 g acetic acid and 800 mg Al₂O₃ nanopowder (13 nm, Sigma Aldrich) were combined and sonicated for 5 minutes followed by the addition of 4.0 g H₂O and sonication for another 5 minutes. Poly(ethylene glycol) (M_n ~20,000, 0.9 g; and M_n ~100,000, 0.4 g) was added, and the mixture was stirred vigorously overnight. Films were prepared by the doctor blade method on glass with an adhesive tape mask. All films were dried in an oven for 1 h then heated to 450 °C over 2 hours, held at 450 °C for 30 minutes, and cooled to 110 °C over 2 hours.

Steady-State Optical Spectroscopy.

Steady-state absorption spectra were collected at room temperature using a Shimadzu 1800 spectrometer. Solution samples were measured in a 2 mm path length cuvette in dichloromethane (optical density 0.3–0.7 at the maximum, which precludes aggregation). Fluorescence measurements were made on a PTI Quanta-Master 1 single-photon-counting fluorimeter in a right-angle configuration with a 10 mm quartz cuvette in dichloromethane with $\lambda_{ex} = 450$ nm.

Electrochemistry.

Electrochemical measurements for **1B-4B** were performed using a CH Instruments Model 660A electrochemical workstation. All measurements were performed under argon using a 1.0 mm diameter platinum disk working electrode, a platinum wire counter electrode, a silver wire pseudoreference electrode, and 0.1 M tetrabutylammonium hexafluorophosphate in acetonitrile. The ferrocene/ferrocenium redox couple (0.40 V vs SCE)⁹ was used as an internal standard. TBAPF₆ was recrystallized twice from ethanol prior to use.

nsTA and µsTA.

Nanosecond transient absorption (nsTA) experiments were performed by exciting samples translated in one dimension with 7 ns, 1.5 mJ, 490 nm pulses as previously described.¹⁰ Kinetic traces were collected from 390 to 800 nm at 5 nm intervals, omitting 520–540 nm. Spectra were constructed by merging the kinetic traces (binned to 3 ns steps, 100 laser shots per kinetic trace).

Transient absorption decays in the microsecond-second timescale were measured using a Nd:YAG laser (Big Sky Laser Technologies Ultra CFR Nd:YAG laser system, 6 ns pulse width). The third harmonic of the pulsed laser (532 nm), at a frequency of 0.8 Hz, was used as the excitation source. The laser intensity was adjusted to 20 µJ cm⁻² using neutral density filters. A liquid light guide with a diameter of 0.5 cm was used to transmit the laser pulse to the sample. A 100 W Bentham IL1 tungsten lamp was used as the probe light source, and the probing wavelength was selected using a monochromator (OBB-2001, Photon Technology International) placed prior to the sample. Several high pass and neutral density filters (Comar Instruments) were used to decrease the light intensity arriving to the detector. Transient absorption data were collected with a Si photodiode (Hamamatsu S3071). The information was passed through an amplifier box (Costronics) and recorded using a Tektronics TDS 2012c oscilloscope (microsecond to millisecond timescale) and a National Instruments (NI USB-6211) DAQ card

(millisecond to second timescale). The decays observed are the average of \sim 500 laser pulses. The data were processed using home-built software based on LabVIEW.¹¹



III. Steady-State Optical Characterization

Figure S1. Normalized absorption spectra of **1A** (red), **2A**, (orange), **3A** (green), and **4A** (blue) on TiO₂ under nitrogen after subtraction of the TiO₂ background.

Table S1. UV-Vis absorption maxima
for 1A-4A on Al ₂ O ₃ and TiO ₂

	Abs _{max}	Abs _{max}
	Al_2O_3	TiO ₂
1A	507 nm	513 nm
2A	522 nm	535 nm
3 A	481 nm	484 nm
4 A	490 nm	491 nm



Figure S2. (A, C, E) fsTA spectra and (B, D, F) decay-associated spectra obtained by SVD and global analysis of the fsTA spectra of **2A-4A** on Al₂O₃.



Figure S3. fsTA single wavelength traces (circles) and fits (lines) of 1A-4A on Al_2O_3 . See the main text for descriptions of fitting procedures.



Figure S4. fsTA single wavelength traces (circles) and fits (lines) of **1A** (red), **2A** (orange), **3A** (green), and **4A** (blue) on Al₂O₃.

V. fsTA data of 1A-4A on TiO₂



Figure S5. Decay-associated spectra obtained by SVD and global analysis of the fsTA spectra of **1A-4A** on TiO₂ (Figure 7).

Sample	Decay- associated global fits τ (ps)	Single wavelength fits (% amplitude) ^{<i>a</i>} τ (ps)				% Al ₂ O ₃ ^b
1A		530 nm	600 nm	645 nm	680 nm	
Al ₂ O ₃	6.0 ± 0.5 160 ± 20 4500 ± 300	$\begin{array}{c} 6.1 \pm 0.2 \; (35\%) \\ 141 \pm 6 \; (32\%) \\ 3720 \pm 90 \; (33\%) \end{array}$	$\begin{array}{c} 6.8 \pm 0.8 \; (\text{-}42\%) \\ 56 \pm 5 \; (\text{-}50\%) \\ 6000 \pm 100 \; (100\%) \end{array}$	$\begin{array}{c} 13.1\pm0.7\ (18\%)\\ 310\pm10\ (32\%)\\ 5.2\pm0.1\ \mathrm{ns}\ (50\%) \end{array}$	4.5 ± 0.2 (36%) 119 ± 5 (33%) 3.7 ± 0.2 ns (31%)	100%
TiO ₂	1.2 ± 0.2 11 ± 2 200 ± 100 Inf.	$\begin{array}{c} Al_2O_3 \ (11\%) \\ 1.2 \pm 0.1 \ (46\%) \\ 11 \pm 2 \ (16\%) \\ 160 \pm 30 \ (10\%) \\ Inf. \ (18\%) \end{array}$	$\begin{array}{l} Al_2O_3 \ (18\%) \\ 1.1 \pm 0.1 \ (-64\%) \\ 150 \pm 10 \ (38\%) \\ Inf. \ (44\%) \end{array}$	$\begin{array}{c} Al_2O_3 \ (16\%) \\ 1.6 \pm 0.3 \ (25\%) \\ 11 \pm 2 \ (21\%) \\ 110 \pm 10 \ (19\%) \\ Inf. \ (19\%) \end{array}$	$\begin{array}{c} Al_2O_3 \ (11\%) \\ 1.0 \pm 0.1 \ (45\%) \\ 8.3 \pm 0.7 \ (24\%) \\ 110 \pm 10 \ (10\%) \\ Inf. \ (9\%) \end{array}$	14%
2A		550 nm	640 nm	670 nm	700 nm	
Al ₂ O ₃	8 ± 1 170 ± 20 2900 ± 200	$3.5 \pm 0.2 (21\%)$ $168 \pm 8 (29\%)$ $3020 \pm 60 (50\%)$	$\begin{array}{c} 17 \pm 2 \; (\text{-46\%}) \\ 150 \pm 10 \; (\text{-48\%}) \\ 3600 \pm 80 \; (100\%) \end{array}$	$\begin{array}{c} 1.09 \pm 0.07 \; (25\%) \\ 280 \pm 20 \; (14\%) \\ 3320 \pm 50 \; (61\%) \end{array}$	$\begin{array}{c} 4.7 \pm 0.2 \; (29\%) \\ 136 \pm 8 \; (28\%) \\ 2860 \pm 70 \; (44\%) \end{array}$	100%
TiO ₂	11 ± 2 200 ± 100 Infinite	$\begin{array}{c} Al_2O_3 \ (7\%) \\ 1.0 \pm 0.3 \ (20\%) \\ 12.9 \pm 0.8 \ (53\%) \\ 280 \pm 60 \ (11\%) \\ Inf. \ (9\%) \end{array}$	Al ₂ O ₃ (16%) 6.4 ± 0.5 (-85%) 81 ± 9 (-56%) Inf. (84%)	$\begin{array}{c} Al_2O_3 \ (11\%) \\ 0.8 \pm 0.2 \ (24\%) \\ 12 \pm 1 \ (43\%) \\ 180 \pm 50 \ (10\%) \\ Inf. \ (12\%) \end{array}$	$\begin{array}{c} Al_2O_3 \ (10\%) \\ 0.9 \pm 0.2 \ (22\%) \\ 13.1 \pm 0.9 \ (50\%) \\ 210 \pm 40 \ (13\%) \\ Inf. \ (4\%) \end{array}$	11%
3A		515 nm	600 nm	645 nm	680 nm	
Al ₂ O ₃	5 ± 1 300 ± 1000 5000 ± 3000	$\begin{array}{c} 2.21 \pm 0.09 \; (41\%) \\ 131 \pm 7 \; (28\%) \\ 3800 \pm 100 \; (31\%) \end{array}$	$28 \pm 2 (16\%) 460 \pm 30 (37\%) 6100 \pm 200 (47\%)$	$\begin{array}{c} 6.9\pm0.4~(21\%)\\ 290\pm10~(35\%)\\ 5500\pm100~(45\%)\end{array}$	$3.6 \pm 0.2 (26\%)$ $240 \pm 10 (32\%)$ $5300 \pm 200 (41\%)$	100%
TiO ₂	1.6 ± 0.2 30 ± 30 500 ± 200 Inf.	$\begin{array}{c} Al_2O_3 \ (21\%) \\ 1.8 \pm 0.2 \ (45\%) \\ 21 \pm 3 \ (25\%) \\ 250 \pm 60 \ (10\%) \end{array}$	$\begin{array}{c} Al_2O_3 \ (12\%) \\ 1.9 \pm 0.2 \ (23\%) \\ 44 \pm 3 \ (33\%) \\ 500 \pm 70 \ (21\%) \\ Inf. \ (11\%) \end{array}$	$\begin{array}{c} Al_2O_3 \ (7\%) \\ 1.7 \pm 0.1 \ (37\%) \\ 33 \pm 2 \ (32\%) \\ 460 \pm 50 \ (15\%) \\ Inf. \ (9\%) \end{array}$	$\begin{array}{c} Al_2O_3 \ (7\%) \\ 1.5 \pm 0.1 \ (45\%) \\ 26 \pm 2 \ (28\%) \\ 370 \pm 40 \ (13\%) \\ Inf. \ (7\%) \end{array}$	7%
4A		515 nm	600 nm	645 nm	680 nm	
Al ₂ O ₃	3.7 ± 0.7 100 ± 100 4000 ± 1000	$\begin{array}{c} 2.3 \pm 0.1 \; (41\%) \\ 90 \pm 4 \; (31\%) \\ 3400 \pm 100 \; (28\%) \end{array}$	$\begin{array}{c} 9.0 \pm 0.5 \; (27\%) \\ 200 \pm 10 \; (35\%) \\ 4900 \pm 200 \; (37\%) \end{array}$	$\begin{array}{c} 4.5 \pm 0.2 \; (32\%) \\ 141 \pm 6 \; (34\%) \\ 4000 \pm 100 \; (34\%) \end{array}$	$\begin{array}{c} 2.8 \pm 0.1 \; (40\%) \\ 88 \pm 5 \; (32\%) \\ 3000 \pm 100 \; (28\%) \end{array}$	100%
TiO ₂	1.0 ± 0.2 20 ± 4 400 ± 300 Inf.	$\begin{array}{c} Al_2O_3 \ (10\%) \\ 0.99 \pm 0.05 \ (48\%) \\ 16.6 \pm 0.9 \ (24\%) \\ 250 \pm 10 \ (17\%) \end{array}$	$\begin{array}{l} Al_2O_3 \ (36\%) \\ 46 \pm 3 \ (24\%) \\ 500 \pm 40 \ (18\%) \\ Inf. \ (22\%) \end{array}$	$\begin{array}{c} Al_2O_3 \ (21\%) \\ 1.0 \pm 0.1 \ (36\%) \\ 19 \pm 1 \ (22\%) \\ 260 \pm 20 \ (12\%) \\ Inf. \ (9\%) \end{array}$	$\begin{array}{c} Al_2O_3 \ (19\%) \\ 0.69 \pm 0.04 \ (42\%) \\ 12.0 \pm 0.7 \ (21\%) \\ 190 \pm 20 \ (10\%) \\ Inf. \ (8\%) \end{array}$	20%

Table S2. fsTA fits of 1A-4A on TiO₂

^{*a*}For "Al₂O₃ (%)", this is fixed as the triexponential decay for the Al₂O₃ sample at the corresponding wavelength. ^{*b*}The average of the weights at the two reddest wavelengths.



Figure S6. fsTA single wavelength traces (circles) and fits (lines) of 1A-4A on TiO₂.



Figure S7. fsTA single wavelength traces (circles) and fits (lines) of 1A (red), 2A (orange), 3A (green), and 4A (blue) on TiO₂.



Figure S8. fsTA single wavelength traces (circles) and fits (lines) of 2A-4A on Al₂O₃ (red) and TiO₂ (orange).



Figure S9. fsTA (A) spectra and (B) decay-associated spectra obtained by SVD and global analysis of the fsTA spectra of **1A** on TiO₂ loaded from acetone. (C) and (D) Normalized single wavelength traces for **1A** on Al₂O₃ loaded from CH₂Cl₂ (black), TiO₂ loaded from CH₂Cl₂ (red), and TiO₂ loaded from acetone (orange) at 530 nm and 680 nm.

VI. fsTA data of 1A on TiO_2 with Li^+ and on SnO_2



Figure S10. Decay-associated spectra obtained by SVD and global analysis of the fsTA spectra (Figure 8) of 1A on TiO_2 with Li^+ and on SnO_2 .

1A	Decay- associated	Single wavelength fits (% amplitude) ^{<i>a</i>} τ (ps)				% Al ₂ O ₂ ^b
sample	global fits τ (ps)	530 nm	600 nm	645 nm	680 nm	, , , , , , , , , , , , , , , , , , , ,
TiO ₂ with Li ⁺		Al ₂ O ₃ (12%)	Al ₂ O ₃ (23%)	Al ₂ O ₃ (26%)	Al ₂ O ₃ (24%)	25%
	0.9 ± 0.2	0.70 ± 0.02 (61%)	0.44 ± 0.04 (-62%)	1.4 ± 0.1 (26%)	0.81 ± 0.03 (57%)	
	300 ± 300	15 ± 5 (2%)	63 ± 3 (32%)	44 ± 7 (8%)	Inf. (19%)	
	Inf.	Inf. (24%)	Inf. (46%)	Inf. (40%)		
SnO_2		Al ₂ O ₃ (8%)	Al ₂ O ₃ (22%)	Al ₂ O ₃ (20%)	Al ₂ O ₃ (19%)	19%
	3.5 ± 0.4	2.4 ± 0.1 (42%)	2.0 ± 0.1 (-58%)	$2.7 \pm 0.2 \ (26\%)$	1.8 ± 0.1 (44%)	
	100 ± 200	27 ± 2 (30%)	97 ± 9 (41%)	40 ± 4 (33%)	21 ± 1 (27%)	
	6000 ± 200	230 ± 20 (20%)	530 ± 70 (33%)	290 ± 30 (22%)	230 ± 20 (10%)	
		Inf. (-9%)	Inf. (5%)			

Table S3. fsTA fits of 1A or	1 TiO2 with Li	⁺ and on SnO ₂
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^{*a*}For "Al₂O₃ (%)", this is fixed as the triexponential decay for the Al₂O₃ sample at the corresponding wavelength. ^{*b*}The average of the weights at the two reddest wavelengths.



Figure S11. fsTA single wavelength traces (circles) and fits (lines) of 1A on TiO₂ with Li⁺ and on SnO₂.



Figure S12. fsTA single wavelength traces (circles) and fits (solid lines) at 600 nm and 680 nm of **1A** on Al_2O_3 (black), TiO_2 (red), TiO_2 with Li^+ (orange), and SnO_2 (green).



VII. fsTA data after subtraction of non-injecting signal

Figure S13. fsTA spectra of 1A-4A on TiO₂ after subtraction of the non-injecting signal.



Figure S14. fsTA single wavelength traces (circles) and fits (lines) of 1A (red), 2A (orange), 3A (green), and 4A (blue) on TiO_2 after subtraction of the non-injecting signal.



Figure S15. Decay-associated spectra obtained by SVD and global fitting of the fsTA spectra after subtraction of the non-injecting signal of **1A-4A** on TiO₂ (Figure S12).



Figure S16. SVD/global fit of the fsTA spectra after subtraction of the non-injecting signal of **1A** on TiO_2 (Figure S12) to an A to B to C model (Figure 11).



Figure S17. (A, C, E) Species-associated spectra obtained by (B, D, F) SVD/global fit of the fsTA spectra after subtraction of the non-injecting signal of **2A** on TiO₂ to an A to B model and of **3A** and **4A** on TiO₂ to A to B to C models (Figure S12).



Figure S18. fsTA spectra of 1A on TiO₂ with Li⁺ and on SnO₂ after subtraction of the non-injecting signal.



Figure S19. fsTA single wavelength traces (circles) and fits (lines) of **1A** on TiO_2 (red), TiO_2 with Li^+ (orange), and SnO_2 (green) after subtraction of the non-injecting signal.



Figure S20. Decay-associated spectra obtained by SVD and global fitting of the fsTA spectra after subtraction of the non-injecting signal of **1A** on TiO₂ with Li⁺ and on SnO₂ (Figure S17).



Figure S21. (A) SVD/global fit of the fsTA spectra after subtraction of the non-injecting signal of **1A** on TiO_2 with Li⁺ (Figure S17) to an A to B to C model (Figure 11). (B) Species-associated spectra obtained by (C) SVD/global fit of the fsTA spectra after subtraction of the non-injecting signal of **1A** on SnO₂ (Figure S17) to an A to B to C to ground state model.

Sample	Decay- associated global fits τ (ps)	Species- associated global fits to models au (ps)	Single wavelength fits (% amplitude) τ (ps)		
1A on			600 nm	645 nm	680 nm
TiO ₂	1.0 ± 0.2 9 ± 2 170 ± 60 Inf.	$a1.35 \pm 0.06$ 28 ± 1 Inf.	1.4 ± 0.1 (-62%) 230 ± 10 (42%) Inf. (58%)	3.9 ± 0.2 (44%) 84 ± 5 (31%) Inf. (25%)	1.91 ± 0.07 (63%) 41 ± 2 (24%) Inf. (12%)
1A on			600 nm	645 nm	680 nm
TiO ₂ with Li ⁺	0.7 ± 0.3 40 ± 30 1000 ± 400 Inf.	$a0.68 \pm 0.02$ 1380 ± 60 Inf.	$\begin{array}{l} 0.50 \pm 0.04 \ (\text{-}68\%) \\ 58 \pm 5 \ (21\%) \\ 1200 \pm 200 \ (16\%) \\ \text{Inf.} \ (63\%) \end{array}$	$\begin{array}{c} 1.3 \pm 0.2 \; (35\%) \\ 27 \pm 6 \; (9\%) \\ \text{Inf.} \; (56\%) \end{array}$	0.95 ± 0.03 (73%) Inf. (27%)
1A on			600 nm	645 nm	680 nm
SnO ₂	2.4 ± 0.2 40 ± 20 490 ± 80	$b2.27 \pm 0.08$ 84 ± 4 $10,000 \pm 1000$	$\begin{array}{l} 1.89 \pm 0.08 \; (-69\%) \\ 180 \pm 5 \; (76\%) \\ 4300 \pm 300 \; (24\%) \end{array}$	7.1 ± 0.4 (46%) 145 ± 5 (54%)	3.07 ± 0.09 (66%) 76 ± 3 (34%)
2A on			640 nm	670 nm	700 nm
TiO ₂	10 ± 2 150 ± 80 Inf.	^c 18.5 ± 0.5 Inf.	$7.1 \pm 0.6 (-104\%)$ $100 \pm 10 (-63\%)$ Inf. (100%)	$1.4 \pm 0.3 (32\%) 22 \pm 1 (52\%) Inf. (16\%)$	9.6 ± 0.5 (71%) 160 ± 20 (22%) Inf. (6%)
3A on			600 nm	645 nm	680 nm
TiO ₂	1.6 ± 0.3 30 ± 30 400 ± 200 Inf.	^{<i>a</i>} 1.76 ± 0.06 97 ± 5 Inf.	$\begin{array}{l} 1.9 \pm 0.2 \ (25\%) \\ 43 \pm 3 \ (37\%) \\ 510 \pm 40 \ (25\%) \\ \mathrm{Inf.} \ (13\%) \end{array}$	$3.3 \pm 0.2 (47\%)$ $100 \pm 4 (41\%)$ Inf. (12%)	2.5 ± 0.1 (54%) 79 ± 4 (36%) Inf. (10%)
4A on			600 nm	645 nm	680 nm
TiO ₂	2.4 ± 0.4 80 ± 30 Inf.	$a1.05 \pm 0.05$ 126 ± 8 Inf.	$\begin{array}{c} 0.47 \pm 0.03 \; (-98\%) \\ 38 \pm 4 \; (33\%) \\ 330 \pm 40 \; (31\%) \\ \mathrm{Inf.} \; (36\%) \end{array}$	$\begin{array}{c} 1.6 \pm 0.2 \; (39\%) \\ 19 \pm 3 \; (29\%) \\ 190 \pm 30 \; (19\%) \\ \text{Inf.} \; (13\%) \end{array}$	0.75 ± 0.06 (49%) 10.8 ± 0.8 (26%) 170 ± 10 (15%) Inf. (10%)

Table S4. fsTA fits after subtraction of non-injecting signal

^{*a*}A to B to C

^bA to B to C to ground state

^cA to B



Figure S22. nsTA (A, E) spectra and (B-D, F) 560 nm traces (circles) and fits (lines) of 1A-4A on Al₂O₃.



Figure S23. nsTA (A) spectra and (B-E) 610 nm traces (circles) and fits (lines) of 1A-4A on TiO₂.



Figure S24. µsTA (A, C, E) spectra and (B, D, F) single wavelength traces of 2A-4A on TiO₂.



Figure S25. Normalized μ sTA single wavelength traces of 1A (black), 2A (red), 3A (orange), and 4A (yellow) at 650 and 750 nm.

X. fsTA data of 5A and 5B



Figure S26. fsTA spectra of 5B in CH₂Cl₂.



Figure S27. (A) Species-associated spectra obtained by (B) SVD/global fit of the fsTA spectra of **5B** in CH_2Cl_2 (Figure S24) to an A to B to C model.



Figure S28. fsTA (A) spectra and (B) decay-associated spectra obtained by SVD and global analysis of the fsTA spectra of **5A** on Al_2O_3 .

Differences in the spectral shape for this fsTA data set compared to all the others can be attributed to the addition of a reference channel in the fsTA apparatus.



Figure S29. Decay-associated spectra obtained by SVD and global analysis of the fsTA spectra of 5A on TiO₂.



Figure S30. fsTA single wavelength traces (circles) and fits (lines) of 5A on Al₂O₃ and TiO₂.



Figure S31. fsTA single wavelength traces (circles) and fits (lines) of **1A** on Al_2O_3 (red) and TiO_2 (orange) and **5A** on Al_2O_3 (green) and TiO_2 (blue) at 600 nm.

5A	Decay- associated		Single wavelength fits (% of amplitude) ^{<i>a</i>} τ (ps)			% Al ₂ O ₃ b
sample	global fits τ (ps)	530 nm	600 nm	645 nm	680 nm	
Al ₂ O ₃	$\begin{array}{c} 0.93 \pm 0.09 \\ 11 \pm 3 \\ 200 \pm 100 \\ 7000 \pm 2000 \end{array}$	$\begin{array}{l} \textbf{1A/Al}_2O_3 \ (26\%) \\ 0.84 \pm 0.03 \ (55\%) \\ 18.6 \pm 0.7 \ (19\%) \end{array}$	$\begin{array}{c} \textbf{1A/Al}_2O_3 \ (19\%) \\ 11.3 \pm 0.3 \ (42\%) \\ 142 \pm 5 \ (36\%) \\ \text{Inf.} \ (3\%) \end{array}$	$\begin{array}{c} \textbf{1A/Al}_2O_3 \ (30\%) \\ 2.30 \pm 0.09 \\ (42\%) \\ 44 \pm 2 \ (28\%) \end{array}$	$\begin{array}{c} \mathbf{1A}/\mathrm{Al_2O_3}~(40\%) \\ 0.92 \pm 0.04~(42\%) \\ 16.8 \pm 0.8~(18\%) \end{array}$	35%
TiO ₂	1.9 ± 0.2 21 ± 3 400 ± 400 Inf.	$\begin{array}{l} 1A/Al_{2}O_{3}\ (28\%)\\ 1.66\pm0.05\ (44\%)\\ 26.9\pm0.7\ (28\%) \end{array}$	$\begin{array}{c} 1A/Al_2O_3\ (16\%)\\ 0.90\pm 0.04\ (-55\%)\\ 39\pm 2\ (28\%)\\ 430\pm 40\ (21\%)\\ Inf.\ (35\%) \end{array}$	$\begin{array}{l} 1 \text{A}/\text{Al}_2\text{O}_3 \ (27\%) \\ 4.0 \pm 0.2 \ (34\%) \\ 40 \pm 1 \ (39\%) \end{array}$	$\begin{array}{l} \textbf{1A}/\text{Al}_2\text{O}_3 \ (28\%) \\ 1.9 \pm 0.1 \ (36\%) \\ 27.0 \pm 0.8 \ (36\%) \end{array}$	27%

Table S5. fsTA fits of 5A on Al_2O_3 and TiO_2

^{*a*}For "Al₂O₃ (%)", this is fixed as the triexponential decay for the Al₂O₃ sample at the corresponding wavelength. ^{*b*}The average of the weights at the two reddest wavelengths.

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