Electronic Supplementary Material (ESI) for Journal of Materials Chemistry B. This journal is © The Royal Society of Chemistry 2015

(Electronic Supplementary Information for Journal of Materials Chemistry B)

Evaluation of AgHAP-containing polyurethane foam dressing for

wound healing: Synthesis, characterization, in vitro and in vivo studies

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Fig. S1. The GPC chromatogram of MeOH end-capped PU copolymer prepared at different NCO/OH ratio.



Fig. S2. Brookfield viscosity on free isocyanate content (NCO %) of PU copolymer.



Fig. S3. TEM image of AgHAP.



Fig. S4. SEM images and EDS analysis of AgHAP/PUFs before and after release experiments in deionized water; (A) AgHAP(2.5)/PUF, (B) AgHAP(5.0)/PUF, (C) AgHAP(7.5)/PUF (magnification 2,000 x).



Fig. S5. Inhibitory effect of AgHAP/PUFs against *Escherichia coli* and *Staphylococcus aureus*; (A) AgHAP(0)/PUF, (B) AgHAP(2.5)/PUF, (C) AgHAP(5.0)/PUF, (D) AgHAP(7.5)/PUF.



10²

10¹

10⁰

 12 14 Time (hr)

י 18 20 22 24

10⁴ 10³

10²

10¹

10⁰

 12 14 Time (hr)



AgHAP/PUFs against four micro-organisms during a 24 h period; (A) MRSA (ATCC 33591), (B) *E. coli* (ATCC 35218), (C) *P. aeruginosa* (ATCC 27853), (D) *S. aureus* (ATCC 25923).

Fig. S7. Wound size reduction (%) in infected wound treated with AgHAP(0)/PUF and AgHAP(5.0)/PUF (n = 5).

Table S1. Average molecular weights and polydispersity index (PDI) of PU prepolymers with different NCO/OH ratio.

NCO %	Retention time (min)	Mn	Mw	MP	Polydispersity
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4	25.952	8,822	10,698	7,344	1.212681
5	25.940	8,327	9,797	7,382	1.176435
6	25.911	8,003	9,144	7,477	1.142612

Table S2. Inhibition zone length (mm). The size of each foam sample was 15mm diameter.

Micro-	Inhibition zone length diameter (mm)				
organisms	(A)AgHAP(0)	(B)AgHAP(2.5)	(C)AgHAP(5.0)	(D)AgHAP(7.5)	
organishis	/PUF	/PUF	/PUF	/PUF	
E. coli	0	0	0.75	1.5	
S. aureus	0	0.5	2.0	2.3	

Table S3. Antimicrobial activity of AgHAP/PUFs ; The colony-forming unit (CFU) of MRSA, E. coli, P. aeruginosa and S. aureus after treatment with AgHAP/PUFs after (A) 1 h and (B) 3 h.

(A) after 1 h

		MRSA (ATCC 33591)		E. coli (ATCC 35218)	
Ma(untreated, 0 contact time)		2.2 x 10 ⁵ CFU	Reduction of	2.1 x 10 ⁵ CFU	Reduction of
Sample code		Number of surviving bacteria (CFU/ml)	bacteria (%)	Number of surviving bacteria (CFU/ml)	bacteria (%)
	Control	3.3 x 10 ⁵	0	2.9 x 10 ⁵	0
Мс	AgHAP(0)/PUF	2.4 x 10 ⁵	27.3	2.0 x 10 ⁵	31.0
	AgHAP(2.5)/PUF	1.7 x 10 ³	99.5	1.7 x 10 ³	99.4
	AgHAP(5.0)/PUF	< 10	99.9	< 10	99.9
	AgHAP(7.5)/PUF	< 10	99.9	< 10	99.9

		P. aeruginosa (ATCC 27853)		S. aureus (ATCC 25923)	
Ma(untreated, 0 contact time)		2.3 x 10 ⁵ CFU	Reduction of	2.2 x 10 ⁵ CFU	Reduction of
Sample code		Number of surviving bacteria (CFU/ml)	ng bacteria (%)	Number of surviving bacteria (CFU/ml)	bacteria (%)
	Control	2.6 x 10 ⁵	0	3.0 x 10 ⁵	0
Мс	AgHAP(0)/PUF	1.8 x 10 ⁵	30.8	2.2 x 10 ⁵	26.6
	AgHAP(2.5)/PUF	1.3 x 10 ³	99.5	6.0 x 10 ³	98.0
	AgHAP(5.0)/PUF	< 10	99.9	< 10	99.9
	AgHAP(7.5)/PUF	< 10	99.9	< 10	99.9

(B) after 3 h

MRSA (ATCC 33591)	E. coli (ATCC 35218)
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Ma(untreated, 0 contact time)		2.2 x 10 ⁵ CFU	Reduction of	2.1 x 10 ⁵ CFU	Reduction of
Sample code		Number of surviving bacteria (CFU/ml)	bacteria (%)	Number of surviving bacteria (CFU/ml)	bacteria (%)
	Control	4.2 x 10 ⁵	0	5.6 x 10 ⁵	0
Mc	AgHAP(0)/PUF	3.0 x 10 ⁵	28.6	4.0 x 10 ⁵	28.6
	AgHAP(2.5)/PUF	< 10	99.9	< 10	99.9
	AgHAP(5.0)/PUF	< 10	99.9	< 10	99.9
	AgHAP(7.5)/PUF	< 10	99.9	< 10	99.9

		P. aeruginosa (ATCC 27853)		S. aureus (ATCC 25923)	
Ma(untreated, 0 contact time)		2.3 x 10 ⁵ CFU	Reduction of	2.2 x 10 ⁵ CFU	Reduction of
Sample code		Number of surviving bacteria (CFU/ml)	ving bacteria (%) ml)	Number of surviving bacteria (CFU/ml)	bacteria (%)
Mc	Control	5.6 x 10 ⁵	0	5.1 x 10 ⁵	0
	AgHAP(0)/PUF	4.2 x 10 ⁵	25.0	3.8 x 10 ⁵	25.5
	AgHAP(2.5)/PUF	< 10	99.9	< 10	99.9
	AgHAP(5.0)/PUF	< 10	99.9	< 10	99.9
	AgHAP(7.5)/PUF	< 10	99.9	< 10	99.9