

1 **Supporting information**

2 **Facile preparation of porous magnetic polydopamine microspheres through an**
3 **inverse replication strategy for efficient enzyme immobilization**

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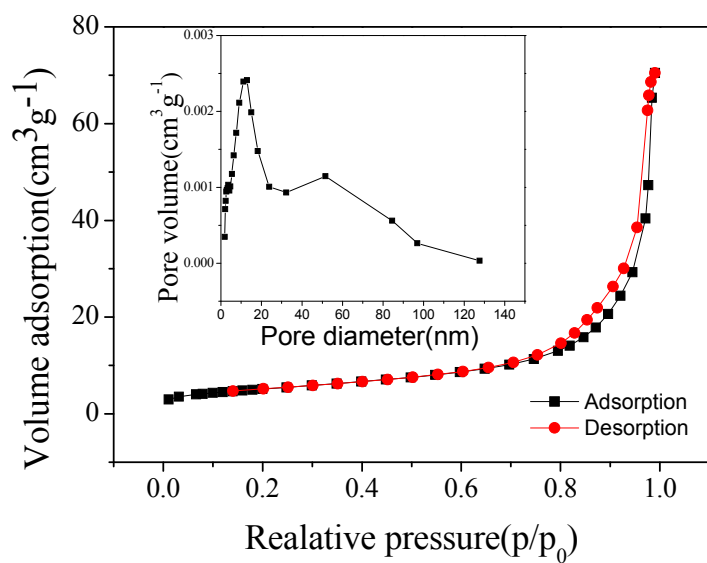
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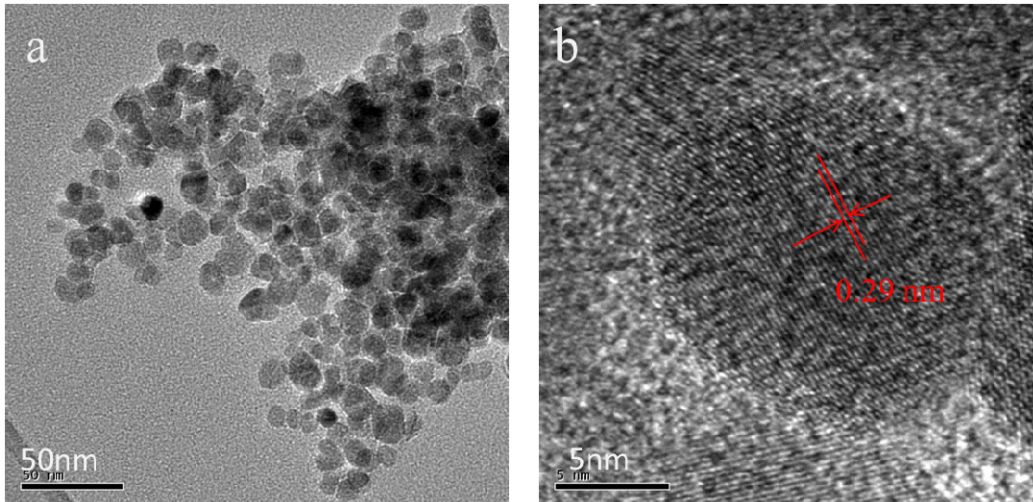
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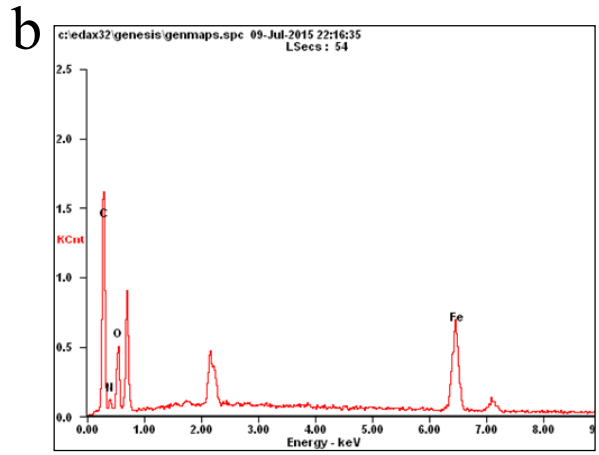
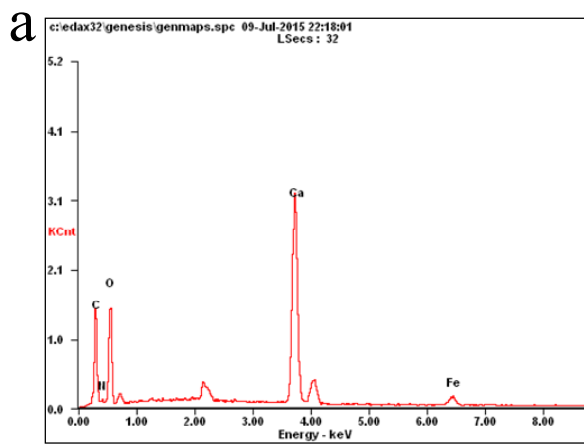
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3 **Fig.S1.** Pore size distribution curve by the BJH method and nitrogen adsorption-desorption isotherm
4 of Fe₃O₄@CaCO₃ microspheres with different diameters of 4.5 μm.

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2 **Fig.S2.** HRTEM images of Fe₃O₄ nanoparticles. (Diameter: 10~20 nm.)
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2 **Fig. S3.** EDS analysis of PMMSs (a) before and (b) after the template removal.

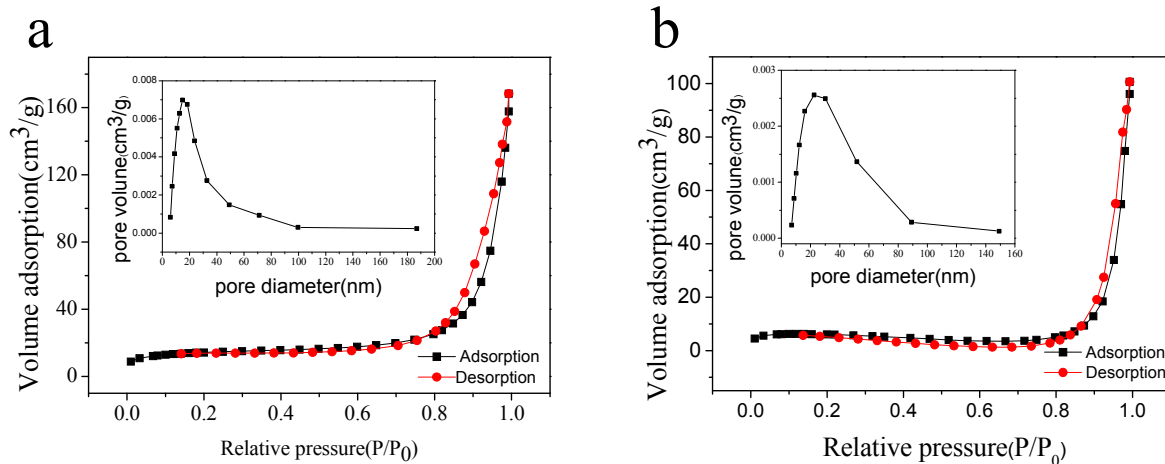
1 **Table S1.** The performance comparison for Fe₃O₄@CaCO₃ microspheres and PMMSs

	Fe ₃ O ₄ @CaCO ₃ microspheres	4.5 μm PMMSs
Immobilization efficiency (%)	2.1±0.1	74.9±2.5
Loading capability (mg/g)	9±0.5	274.5±3.4
Specific activity (U·mg ⁻¹ enzyme)	163.2±5.9	162.3±5.2

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3 In detail, the immobilization efficiency, loading capacity and specific activity for Fe₃O₄@CaCO₃
 4 microspheres were 2.1%, 9 mg/g and 163.2 U·mg⁻¹ enzyme, respectively. In comparison, the
 5 immobilization efficiency and loading capacity of Fe₃O₄@CaCO₃ microspheres were lower than
 6 those of PMMSs. This was probably owing to the rather weak interaction (physical adsorption)
 7 between YADH and Fe₃O₄@CaCO₃ microspheres. Once conducting the reaction with an equal
 8 amount of enzyme, the specific activity of Fe₃O₄@CaCO₃ microspheres (163.2 U·mg⁻¹ enzyme) was
 9 nearly equal to that of PMMSs (162.3 U·mg⁻¹ enzyme).

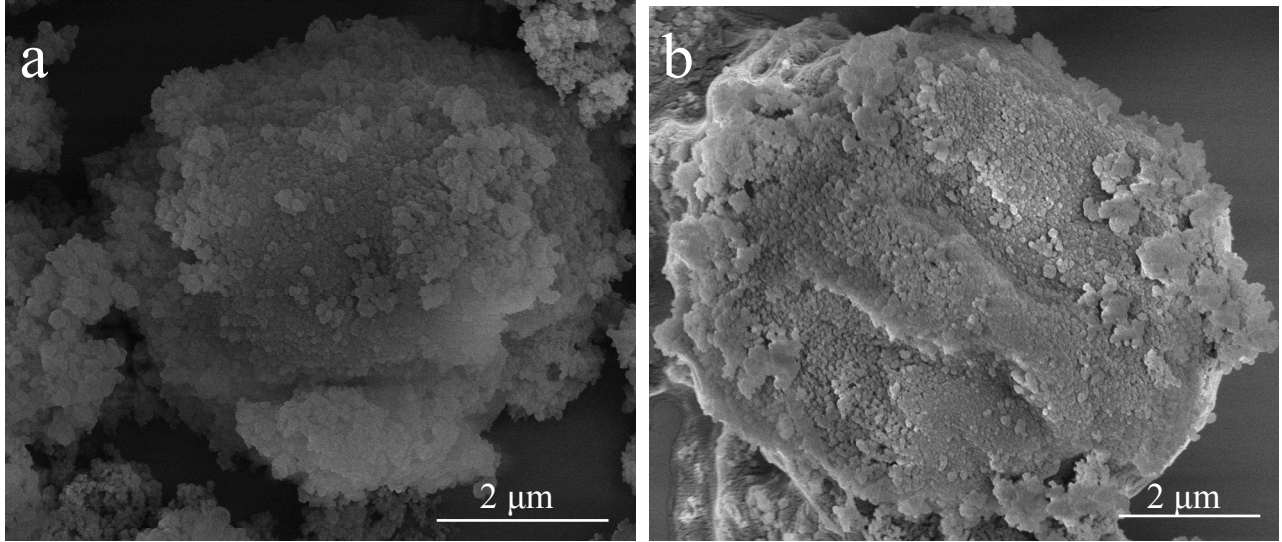
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2 **Fig. S4.** Pore size distribution curve by the BJH method and nitrogen adsorption-desorption isotherm
 3 of PMMSs with different diameters a) 4.5 μm and b) 3.0 μm .

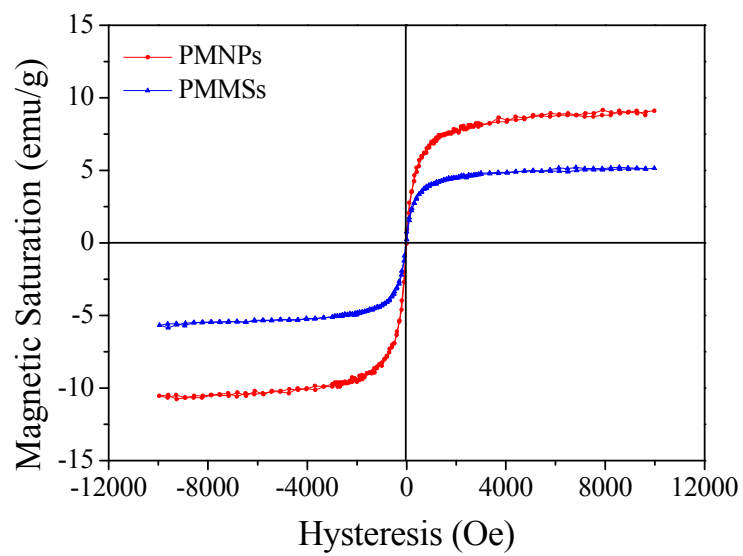
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2 **Fig. S5.** SEM images of YADH immobilized PMMSs (a) before and (b) after reaction.

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2 **Fig. S6.** Magnetic hysteresis loops of PMNPs and PMMSs.

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