Electronic Supplementary information:

Physicochemical Insights of Irradiation-Enhanced Hydroxyl Radical Generation from ZnO Nanoparticles

Qingbo Yang, ¹ Tien-Sung Lin, ² Casey Burton, ¹ Sung-Ho Park, ² Yinfa Ma^{1*}

¹Department of Chemistry and Center for Single Nanoparticle, Single Cell, and Single

Molecule Monitoring, Missouri University of Science and Technology, Rolla, MO 65409,

United States

²Department of Chemistry, Washington University in St. Louis, St. Louis, MO 63130, United

States

*Corresponding Author

Address: Department of Chemistry

Missouri University of Science and Technology

400 West 11th Street

Rolla, MO65409

Phone: 573-341-6220

Fax: 573-341-6033

E-mail: <u>yinfa@mst.edu</u>

Table of contents

Page S-1 - S-8, Figure S1: Representative raw data of ESR analysis of free hydroxyl radical production under irradiation by 50-70 nm ZnO NPs with different dosages;

Page S-9, Figure S2: UV/Vis measurement and band gap energy (E_g) calculation of ZnO aqueous suspension for two hours under irradiation;

Page S-10 – **S-13, Figure S3:** Non-ultrasonication influences on ESR signal intensity of ZnO NPs;

Page S-14, Figure S4: Representative ESR signals of superior free hydroxyl radical production of 10 nm ZnO NPs in MQ water suspension;

Page S-15: **Figure S5**: a) Influence of ultrasonication on ZnO suspension pH value in PBS buffer, b) pH influence on UV/Vis absorption of ZnO NPs suspension, and c) hydroxyl radicals production rate plotting by compare ESR signal intensity and first four-minute's irradiation time;

Page S-16, Figure S6: Representative ESR spectra of ZnO NPs suspended in glycerol and ethanol;

Page S-17 – **S-19, Figure S7:** Representative ESR spectra of TiO_2 , SiO_2 or CeO_2 NPs with similar sizes under irradication;

Page S-20, Figure S8: ICP-MS analysis of the trace amount of impurity elements in 10 nm and 50-70 nm sized ZnO NPs;

Page S-21, Figure S9: Proposed cytotoxic pathways that ZnO NPs may be involved during early-stage cell-NPs interaction;

Page S-21: Reference.

Figure S1-a

0.1mg/mL nano-ZnO



Figure S1-b



0.5 mg/mL nano-ZnO

Figure S1-c



S-5

Figure S1-d



Figure S1-e



Figure S1-f



Figure S1 Electron spin resonance analysis of free hydroxyl radicals production under irradiation by 50-70 nm ZnO nanoparticles in aqueous suspension with different dosages of 0.1, 0.5, 1, 3, 10 and 30 mg/mL as a function of time.



Figure S2 UV/Vis absorption measurement and calculated band gap energy (E_g) plotting (inset) of 1 mg/mL 50-70 nm ZnO aqueous suspension for two hours under irradiation.

Figure S3-a



Figure S3-b



Figure S3-c







Figure S3 Effect of ultrasonication on ESR signal intensity of 10, 1 and 0.1 mg/mL 50-70 nm ZnO NPs.



Figure S4 ESR signals of irradiating 10 mg/mL ZnO NPs (10 nm) in aqueous suspension for

10 minutes.



Figure S5 a) Influence of ultrasonication on the pH of freshly made ZnO NPs suspension. Again, 1 mg/mL 50-70 nm ZnO NPs was used combined with the PBS solution. The PBS buffer pH had been adjusted to 7.4 before mixing with NPs. **b**) UV/Vis absorption behavior of ZnO NP suspension under varied pH conditions. Inset: Linear correlation plot of ZnO NP suspension absorption vs. basic pH conditions. Acidic condition (> pH 3) also support a linear Abs./pH correlation, but it is apparently different from basic side. **c**) The initial •OH generation rate plotted based on ESR signal intensity vs. the first four-minute's irradiation time. Triplicated data was acquired for statistical analysis.



Figure S6 ESR spectra of the DMPO-trapped radicals obtained through irradiation of 50-70 nm ZnO in glycerol or ethanol (EtOH).

Figure S7-a







S-18





Figure S7 ESR spectrum of DMPO-trapped-radicals generated by irradiation on 1 mg/mL of TiO_2 , SiO_2 or CeO_2 NPs with similar fabricated sizes. Labelled positions denote hydroxyl (blue arrow) and carbon-based (dark asterisk) free radicals.¹



Figure S8 ICP-MS analysis of the trace amount of impurity elements within 1mg/mL 10 nm or 50-70 nm sized ZnO particles that were used in this study. Particle suspensions were freshly prepared and ultrasonicated, followed by 1 hour of microwave digestion using 3 M nitric acid. ICP-MS analysis was conducted thereafter with properly dilution of dissolved samples.



Figure S9 Proposed cytotoxic intracellular pathway by ZnO NPs. Compared to a relatively slow zinc ion releasing process, a much quicker way of hydroxyl radicals production and toxicating process would be expected with the assistance of: oxygen abundance, basic local pH environment as well as irradiation.

References

1. Mazumdar, A.; Adak, S.; Chatterjee, R.; Banerjee, R. Mechanism-based inactivation of lacrimal-gland peroxidase by phenylhydrazine: a suicidal substrate to probe the active site. *Biochem. J* **1997**, *324*, 713-719.