## **Supporting Information**

## A rapid SERS method for label-free bacteria detection using polyethylenimine-modified Au-coated magnetic microspheres and Au@Ag nanoparticles

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**Fig. S1** (A) The SERS spectra of PATP measured with different concentrations on the Fe<sub>3</sub>O<sub>4</sub>@Au microspheres. (B) Calibration curve for PATP at a concentration range of  $10^{-10}$  to  $10^{-6}$  M obtained by using SERS intensity at 1078 cm<sup>-1</sup>. The error bars represent the standard deviations from 5 measurements.



Fig. S2 SERS spectra of different concentrations of *E. coli BL21* obtained with the  $Fe_3O_4@Au@PEI$  microspheres as SERS substrates.



**Fig. S3** Capture kinetics of the Fe<sub>3</sub>O<sub>4</sub>@Au@PEI microspheres for *E.coli BL21* at a regular shaking incubation of 250 rpm. The original concentrations of the bacteria in PBS (10 mM, pH 7.4) have an  $OD_{600}$  of 0.5.



**Fig. S4** TEM images of the synthesized enhanced nanoparticles: (A) 50 nm Au NPs, (B) 60 nm Au NRs, and (C) 60 nm Au@Ag NPs. The insets are the corresponding optical images. (D) UV– visible spectra of the synthesized enhanced nanoparticles. (E) Raman intensity of *E. coil* adsorbed on the three enhanced nanoparticles under the same conditions.



**Fig. S5** True color photos of (A) Au@Ag NPs in aqueous solution, (B) Au@Ag NPs in ethanol, and (C) the concentrated Au@Ag NPs in ethanol.



**Fig. S6** Effect of the Au@Ag NPs concentrations in the CEE three-step method for bacteria *E. coil* detection. (A) SERS spectra of Au@Ag NPs by increasing their concentration from 0-fold to 40-fold. (B) Plot of 729 cm<sup>-1</sup> band intensities versus the particle concentration of Au@Ag NPs from 0-fold to 40-fold. The error bars represent the standard deviation from five measurements.

Raman Shift (cm-1)	Assignments
563	carbonhydrates
624	aromatic ring skeletal
655	δ(COO-)
729	adenine, glycosidic ring mode
958	υ(CN)
1093	amide III, adenine, polyadenine, DNA
1250-1310	amide III
1268	δ(CH2) amide III
1328	υ(NH2) adenine, polyadenine, DNA
1310-1440	υ(COO-) symmetric
1370	υ(COO-) and δ(C-H) proteins
1440-1460	δ(CH2) saturated lipids
1540-1645	amide II, υ(CN), γ(NH)
1640-1680	amide I

Table. S1 Raman peaks of E. coli BL21 and corresponding assignments.



**Fig. S7** SERS spectra of *E. coli BL21* in different systems: (a) *E. coli BL21* (10<sup>6</sup> cells/mL) obtained with the Fe<sub>3</sub>O<sub>4</sub>@PEI microspheres and Au@Ag NPs, (b) *E. coli BL21* (10<sup>4</sup> cells/mL) obtained with the Fe<sub>3</sub>O<sub>4</sub>@Au@PEI microspheres and Au@Ag NPs, and (c) *E. coli BL21* (10<sup>4</sup> cells/mL) obtained with the Fe<sub>3</sub>O<sub>4</sub>@PEI microspheres and Au@Ag NPs. All the Raman spectra were shifted for clarity.



**Fig. S8** SERS spectra collected from 20 randomly selected spots on the (A) Fe<sub>3</sub>O<sub>4</sub>@Au@PEI-E. coli/Au@Ag complexes and (B) Fe<sub>3</sub>O<sub>4</sub>@Au@PEI-S. aureus/Au@Ag complexes substrates.