

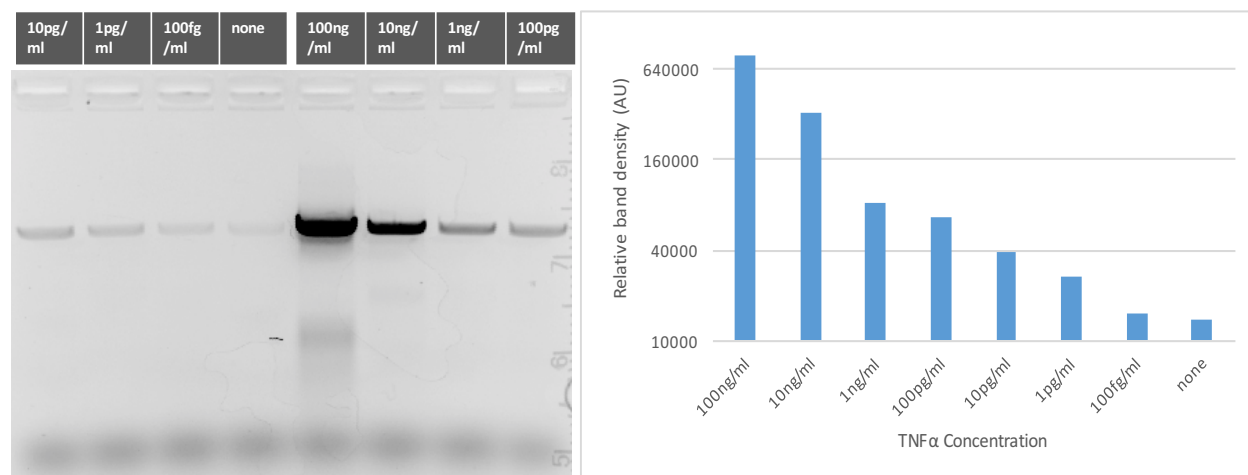
### Supporting information

Real-Time Femtomolar Detection of Cancer Biomarkers from Photoconjugated Antibody-Phage Constructs

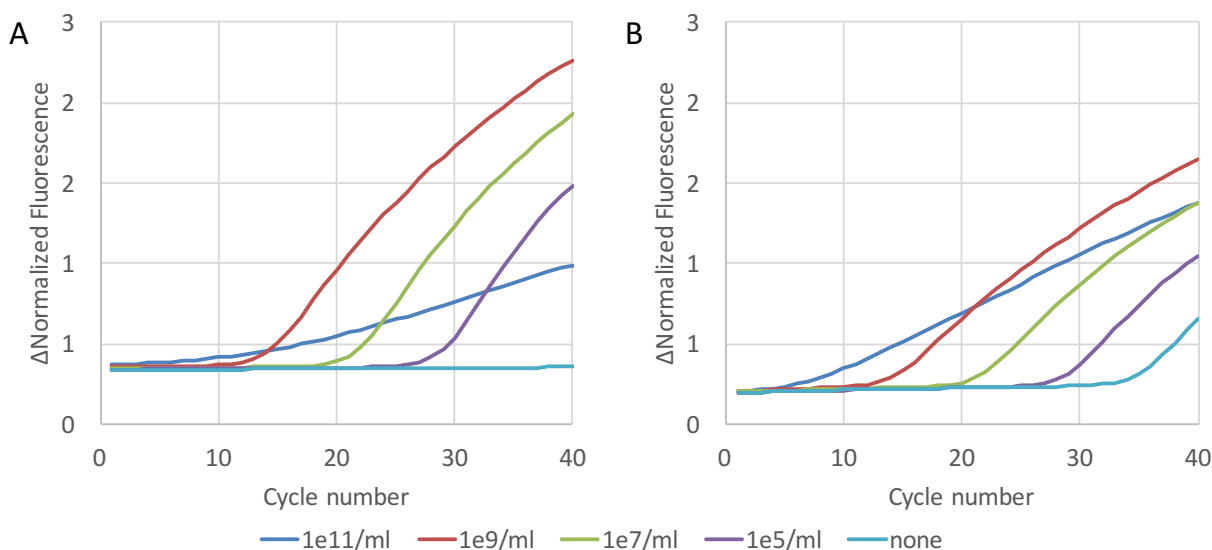
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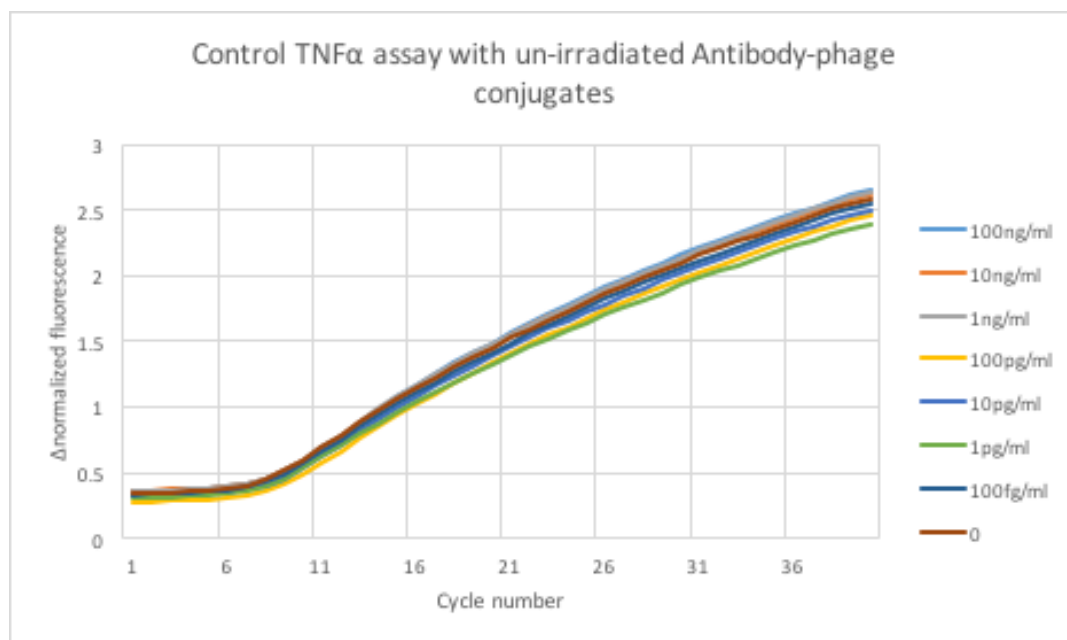
\*To whom correspondence is addressed: Jennifer.Cha@colorado.edu



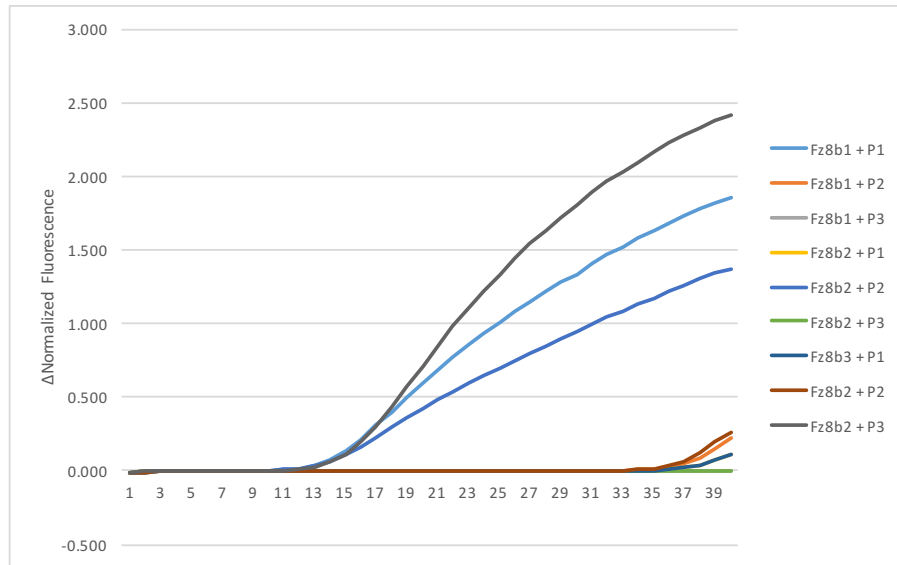
**Figure S1: Gel electrophoresis based assay for detecting TNFα:** Anti-TNFα-phage were used to detect various concentrations of TNFα diluted in 0.1%PBST solution with 5mg/ml BSA. Immunoassays were otherwise performed as reported in materials and methods. PCR was performed in a standard thermocycler for 25 cycles by adding assay beads directly to PCR reaction mixture containing primers, dNTPs and polymerase. After PCR reaction, reaction mixtures were added directly to a 0.75% agarose gels and electrophoresed for 30mins at 120V. Samples are placed in the gel out of order to avoid any gel effects.



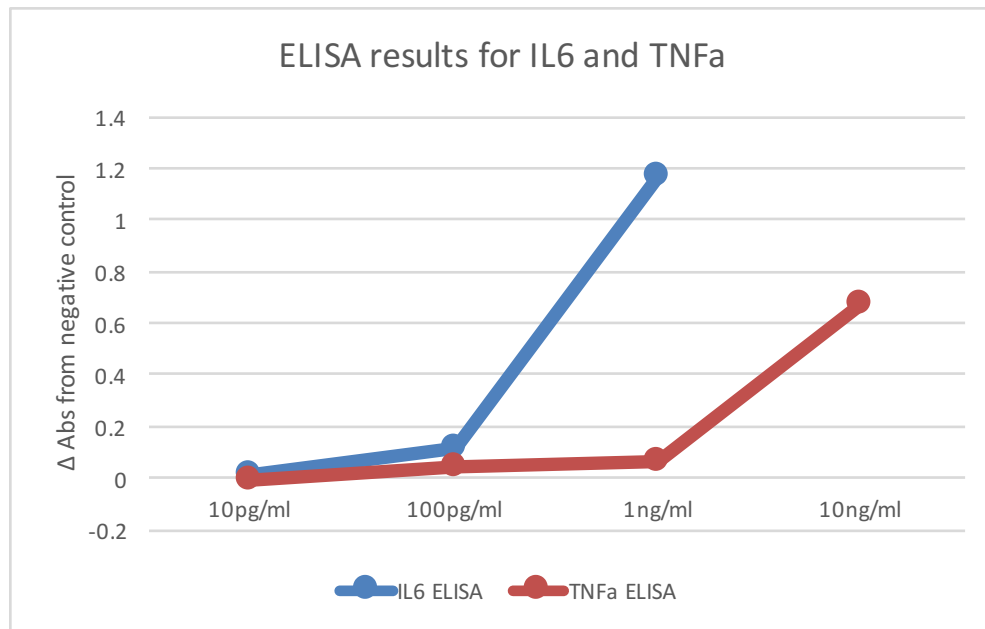
**Figure S2: Validation of Taqman probes for real time PCR detection of phage in the presence of immunoassay magnetic beads:** **A)** Indicated concentrations of phage are added to a real time PCR reaction mixture containing Taqman universal mastermix II (Life Technologies) along with phage specific primers (600nM) and Taqman probe (250nM) (Integrated DNA Technologies). **B)** Reactions were supplemented with 5ug of streptavidin functionalized magnetic beads (Dynabeads myONE streptavidin T1, Life Technologies).



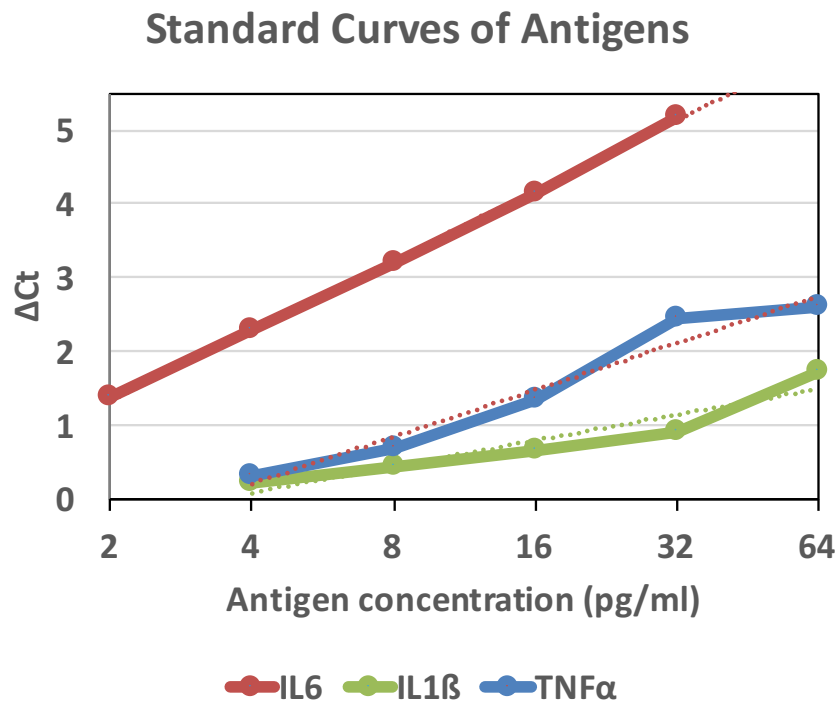
**Figure S3: Control qPCR assay for TNF $\alpha$  using phage antibody conjugates without UV irradiation.** Amplification curves for various TNF $\alpha$  concentrations in a TNF $\alpha$  assay performed as in Figure 3 but without irradiating the antibody-phage mixture prior to using it in the assay. Because of the lack of irradiation, phage in the mixture bind to the capture antibody present on the assay beads, causing an amplification at early cycles for all TNF $\alpha$  concentrations.



**Figure S4: Sequence specific real time PCR amplification of various phage via various primers:** Taqman probe based real time amplification curves of the three phage types (Fz8b1,2,3) used for multiplex immunoassays diluted to 1e9/ml in reactions with primer sets specific to each phage type (P1,2,3). In multiplex immunoassays, Fz8b1 was conjugated with TNF $\alpha$ , Fz8b2 with IL6 and Fz8b3 with IL-1 $\beta$



**Figure S5: Comparative phage ELISAs of IL6 and TNF $\alpha$ :** Antibody-phage conjugates prepared identically used to detect listed concentrations of IL-6 or TNF $\alpha$  immobilized on magnetic beads via Enzyme linked immunosorbent assay and an Anti-M13-HRP conjugate.



**Figure S6: Standard curves at low concentrations of three antigens in multiplex assay format:** Each antigen listed is diluted to the indicated concentrations and tested in multiplex immunoassay format. Resulting  $\Delta C_t$  values for each dilution series are analyzed for lines of best fit through the least square method. Resulting logarithmic formula are used to calculate the indicated concentration from  $\Delta C_t$  values attained in later multiplex assays testing multiple antigens at once.

**Table S1: Sequence of primers used for Taqman probe based qPCR.**

| Name/description         | Sequence                             |
|--------------------------|--------------------------------------|
| fwd primer 1             | Tacccatgaacacaagttgcg                |
| fwd primer 2             | TcttgaaacaggttctagcggAA              |
| fwd primer 3             | CTTtcatgtgaacttgaggtacgAA            |
| rev primer for all phage | AAgccgcggtaaatagcaataa               |
| Universal Taqman probe   | FAM-ATGCGAAAAACCTAAAAGAGCTTGCCGA-ZEN |

### Phage construct Fz8b-1 used in this study

Mon Mar 28, 2016 10:14 MDT

GenBank Fz8b-1.ape

Text Map

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[illegible]

2176 **gaaccccgctaaatcctaataccttctcttctgaggagctctcagccctctttaactttcatgtttcagaataaataggttc** 2250  
pIII

2251 **cgaaataggcagggtgcattaaactgtttatcacgggcactgttactcaaggcaactgaccccgtaaaacttattac** 2325  
pIII

2326 **cagtacactcctgtatcatcaaaagccatgtatgacgcttactggaacggtaaattcagagactgcgctttccat** 2400  
pIII

2401 **tctggctttaatgaagatccattcgtttgtgaatatcaaggccaatcgtctgacctgcctcaacctcctgtcaat** 2475  
pIII

2476 **gctggcggcggtcctggtggtggttctggtggcggtcctgaggggtggcggtcctgaggggtggcggttctgaggg** 2550  
pIII

2551 **ggcggtcctgaggggtggcggttccgggtggcggtcctgggttccgggtgattttgattatgaaaaaatggcaaacgct** 2625  
pIII

2626 **aataagggggctatgaccgaaaaatgccgatgaaaaacgcgtacagctcgtacgctaaaggcgaacttgattctgtc** 2700  
pIII

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pIII

2776 **ggtgattttgctggctctaattcccaaatggctcaagtcggtgacggtgataattcacctttaatgaataatttc** 2850  
pIII

2851 **cgtaaatatttaccttctttgcctcagtcggttgaaatgtcgcccttatgtctttggcgctggtaaaccatatgaa** 2925  
pIII

2926 **ttttctatttgattgtgacaaaaataaaacttattccgtggtgtcttttcggtttcttttatatgttgccacctttatg** 3000  
pIII

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pIII

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3226 ctgatattagcgcacaaattaccctctgattttgttcaggcggttcagtttaattctcccgtctaatgcgcttccct 3300

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3376 attgggataaataaatatggctgtttattttgtaactggcaaattaggctctggaagacgctcggttagcggttg 3450

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