

**Electronic Supplementary Information (ESI)**

for

**Rapid detection of hydrazine in almost wholly water solution and in living cells with a new colorimetric and fluorescent turn-on probe**

Qisong Zhai, Weiyong Feng, and Guoqiang Feng\*

*Key Laboratory of Pesticide and Chemical Biology of Ministry of Education, College of Chemistry, Central China*

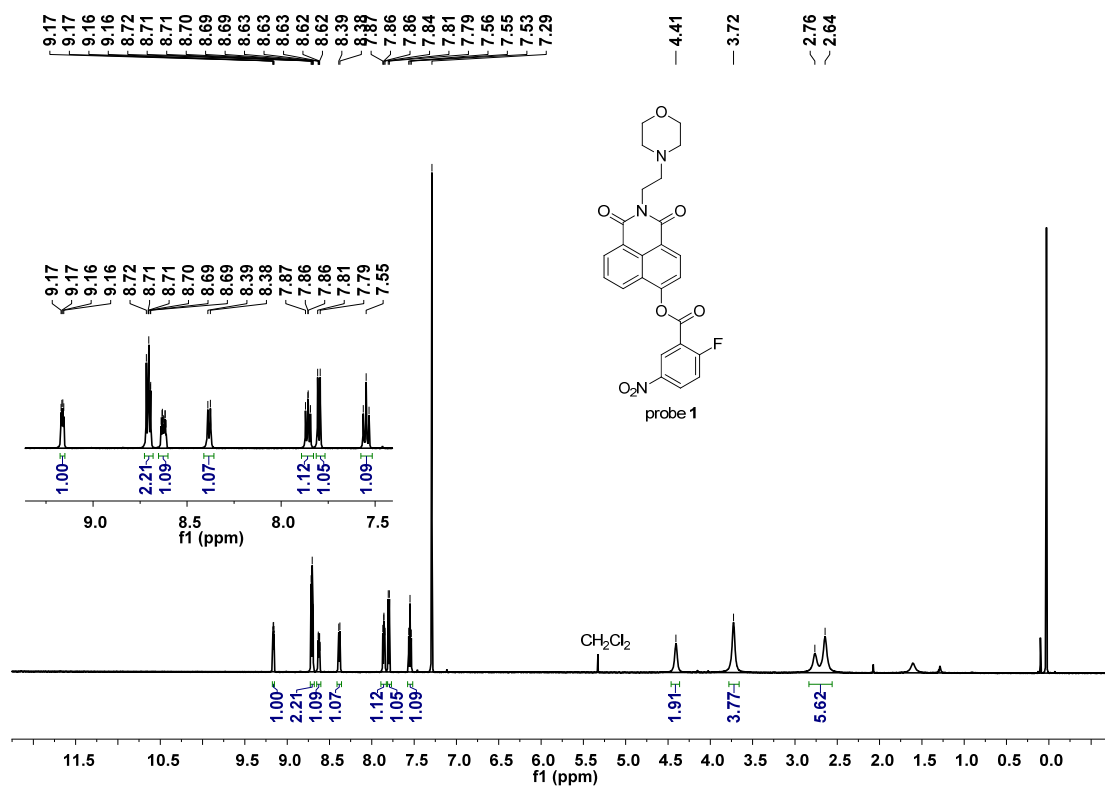
*Normal University, 152 Luoyu Road, Wuhan 430079, P. R. China. E-mail:*

[gf256@mail.ccnu.edu.cn](mailto:gf256@mail.ccnu.edu.cn)

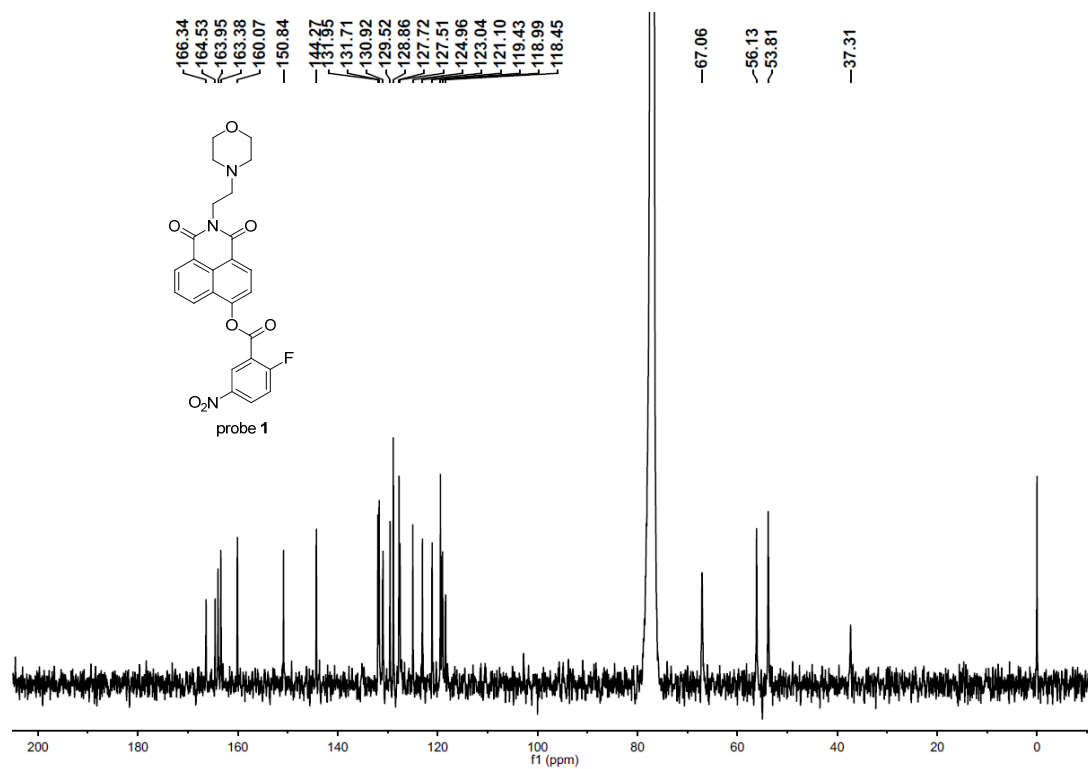
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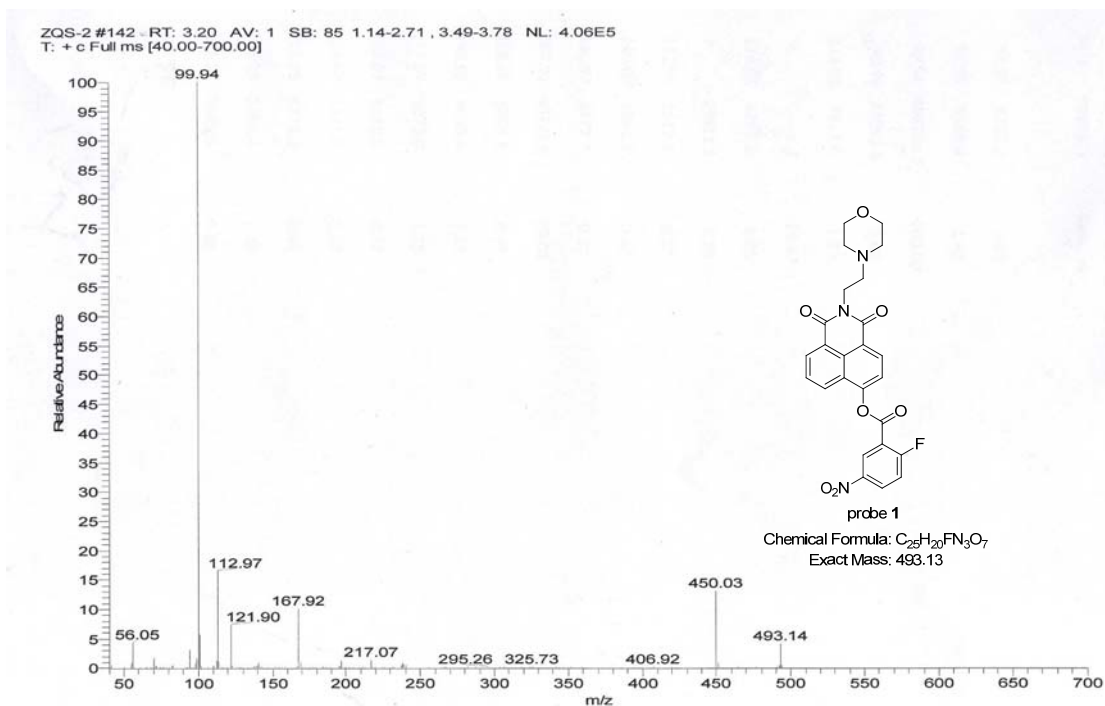
# 1. Structure characterizations of probe 1



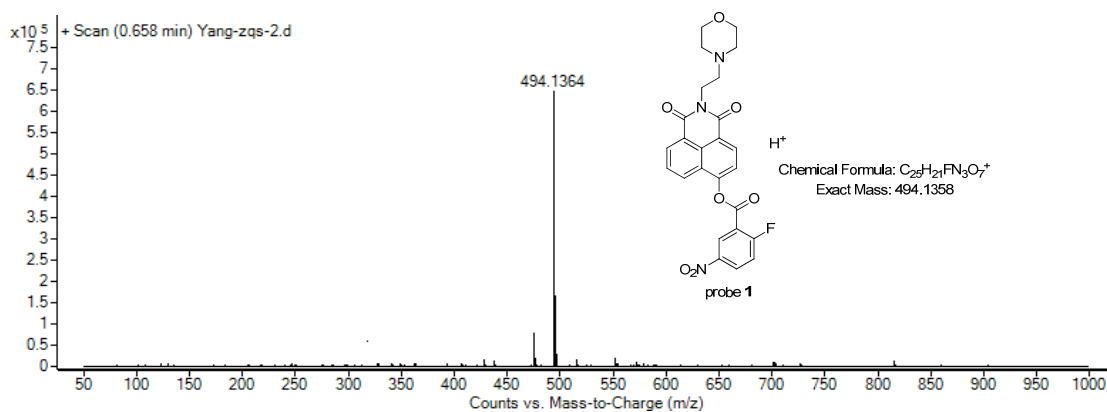
<sup>1</sup>H-NMR spectrum of probe 1 in CDCl<sub>3</sub>



<sup>13</sup>C-NMR spectrum of probe 1 in CDCl<sub>3</sub>

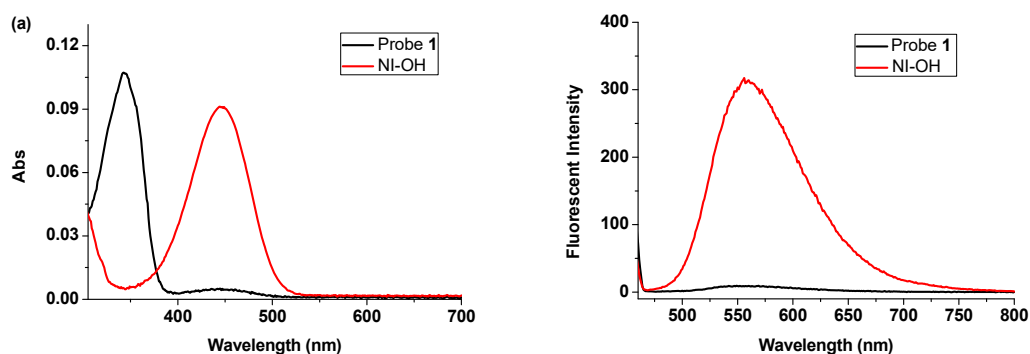


MS (EI) spectrum of probe 1

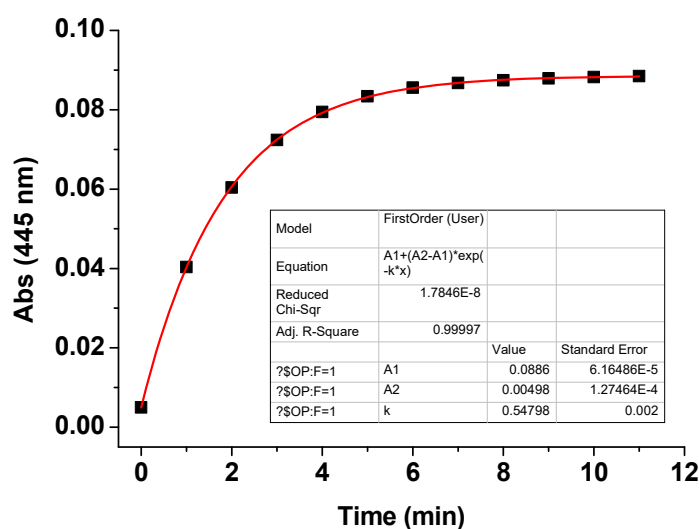


HR-MS spectrum of probe 1

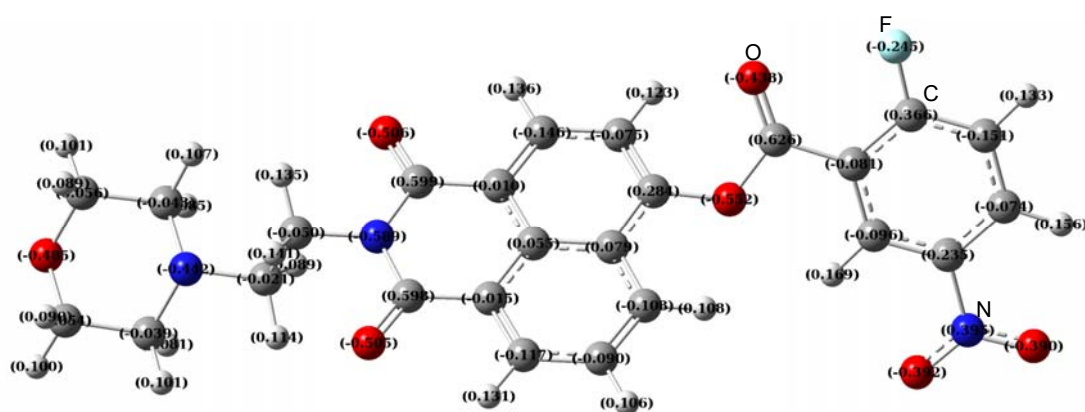
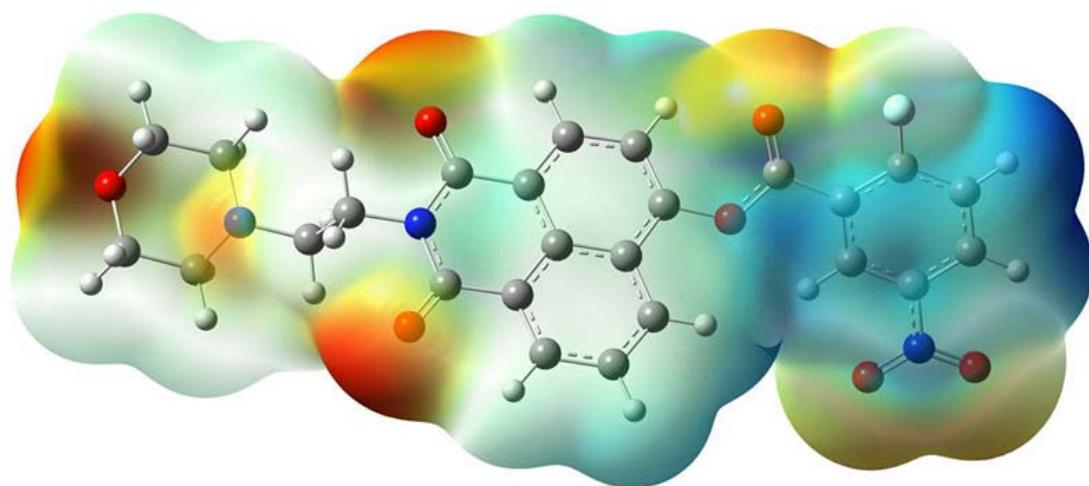
## 2. Additional spectra and data



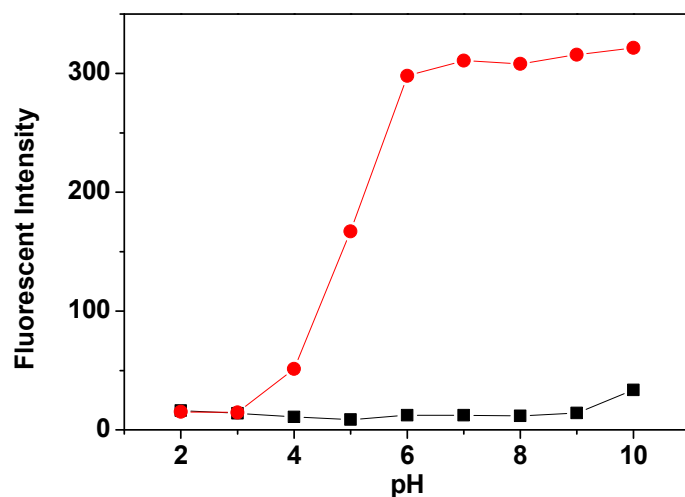
**Fig. S1.** (a) UV-Vis and (b) fluorescence spectra of probe 1 (10  $\mu\text{M}$ ) and NI-OH (10  $\mu\text{M}$ ) in PBS buffer (10 mM, pH 7.4) with 2%  $\text{CH}_3\text{CN}$  at 37  $^\circ\text{C}$ . For fluorescence,  $\lambda_{\text{ex}} = 450 \text{ nm}$ , slit width:  $d_{\text{ex}} = 5 \text{ nm}$ ,  $d_{\text{em}} = 10 \text{ nm}$ .



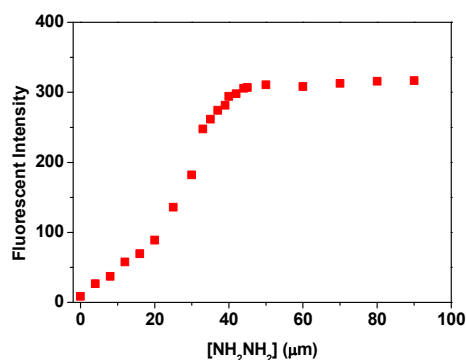
**Fig. S2.** Kinetic curve of probe 1 (10  $\mu\text{M}$ ) with hydrazine (50  $\mu\text{M}$ ) in PBS buffer (10 mM, pH 7.4) with 2%  $\text{CH}_3\text{CN}$  at 37  $^\circ\text{C}$ . The reaction was monitored by absorbance change at 445 nm and the data were fitted by a first-order reaction scheme as shown in the figure.



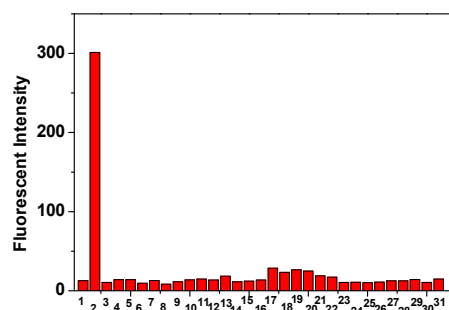
**Fig. S3.** Calculated charge distribution map of probe 1 by DFT (density functional theory).



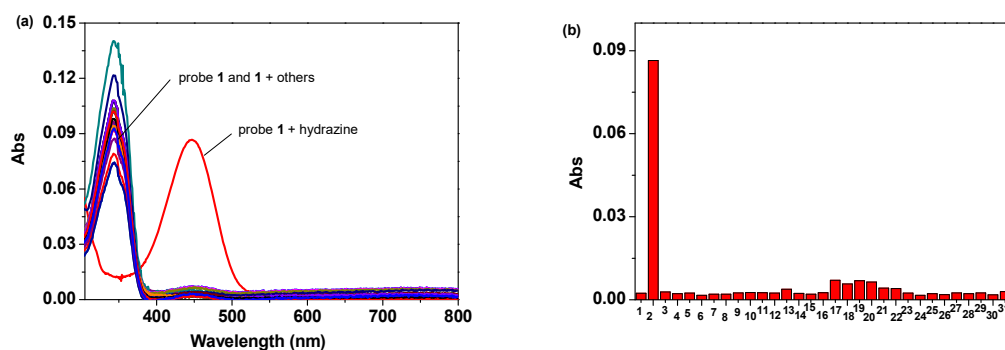
**Fig. S4.** The fluorescent responses of probe 1 (10  $\mu$ M, black) and probe 1 with hydrazine (50  $\mu$ M, red) at 558 nm under different pHs. All experiment was performed in PBS buffer (10 mM) with with 2%  $\text{CH}_3\text{CN}$  at 37  $^\circ\text{C}$  and each spectrum was obtained 10 min after mixing.  $\lambda_{\text{ex}} = 450$  nm, slit width:  $d_{\text{ex}} = 5$  nm,  $d_{\text{em}} = 10$  nm.



**Fig. S5.** Fluorescence responses of probe **1** (10  $\mu\text{M}$ ) at 558 nm upon addition of hydrazine (0-90  $\mu\text{M}$ ) in PBS buffer (10 mM, pH 7.4) with 2%  $\text{CH}_3\text{CN}$  (v/v) at 37  $^\circ\text{C}$ . Final concentration of hydrazine: 0, 4, 8, 12, 16, 20, 25, 30, 33, 35, 37, 39, 40, 42, 44, 45, 50, 60, 70, 80 and 90  $\mu\text{M}$ .

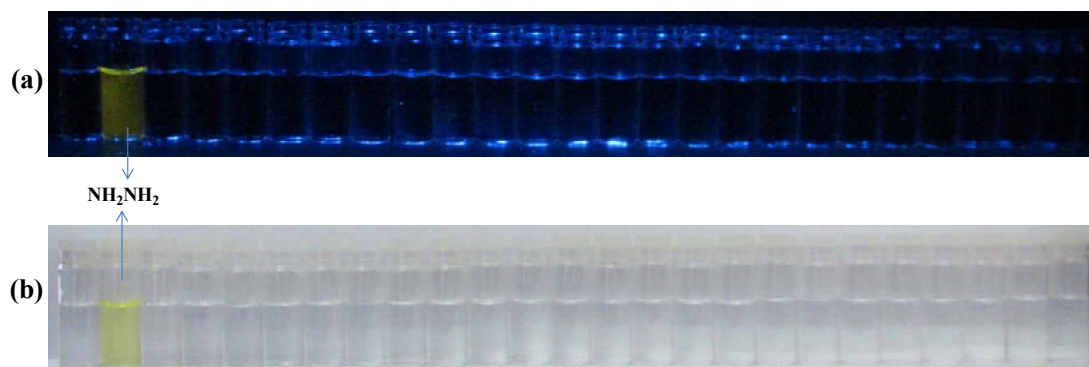


**Fig. S6.** Fluorescence intensity changes of probe **1** (10  $\mu\text{M}$ ) at 558 nm toward 100  $\mu\text{M}$  (except: hydrazine 50  $\mu\text{M}$ ) of various analytes including: 1. none, 2. hydrazine, 3. Cys, 4. Hcy, 5. GSH, 6. NaF, 7. NaCl, 8. NaBr, 9. NaI, 10.  $\text{Na}_2\text{S}_2\text{O}_7$ , 11.  $\text{Na}_2\text{SO}_3$ , 12.  $\text{Na}_2\text{CO}_3$ , 13. NaHS, 14.  $\text{NaNO}_3$ , 15. NaSCN, 16. NaAc, 17.  $\text{NH}_3 \cdot \text{H}_2\text{O}$ , 18.  $\text{NH}_2\text{OH} \cdot \text{HCl}$ , 19.  $\text{H}_2\text{NCH}_2\text{CH}_2\text{NH}_2$ , 20.  $\text{HOCH}_2\text{CH}_2\text{NH}_2$ , 21.  $\text{C}_6\text{H}_5\text{NH}_2$ , 22.  $\text{C}_6\text{H}_5\text{CH}_2\text{NH}_2$ , 23. NaClO, 24.  $\text{H}_2\text{O}_2$ , 25.  $\text{NaNO}_2$ , 26.  $^t\text{BuOO}^\bullet$ , 27.  $^\bullet\text{OH}$ , 28.  $\text{NaN}_3$ , 29.  $\text{Na}_3\text{PO}_4$ , 30.  $\text{Na}_2\text{SO}_4$ , 31.  $\text{Na}_2\text{S}$ . All experiments were performed in PBS buffer (10 mM, pH 7.4) with 2%  $\text{CH}_3\text{CN}$  at 37  $^\circ\text{C}$ , and each spectrum was obtained 10 min after addition of an analyte.  $\lambda_{\text{ex}} = 450 \text{ nm}$ , slit width:  $d_{\text{ex}} = 5 \text{ nm}$ ,  $d_{\text{em}} = 10 \text{ nm}$ .

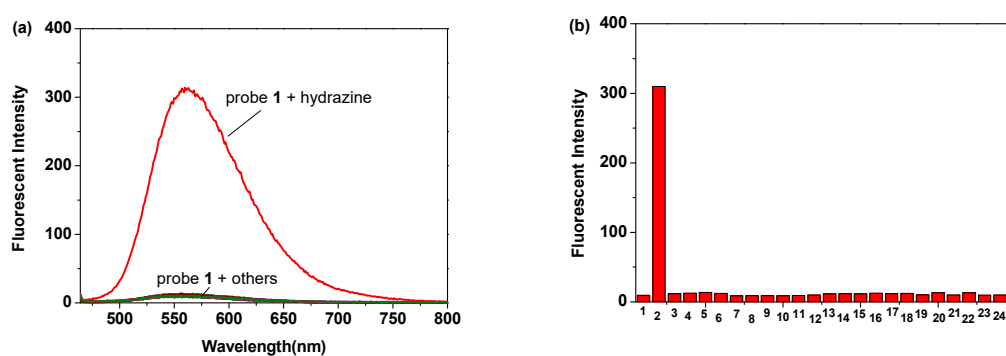


**Fig. S7.** (a) Absorption spectra changes of probe **1** (10  $\mu\text{M}$ ) toward various analytes (100  $\mu\text{M}$ , except hydrazine 50  $\mu\text{M}$ ). (b) Absorbance intensity changes of probe **1** (10  $\mu\text{M}$ ) at 445 nm toward 100  $\mu\text{M}$  of various analytes including: 1. none, 2. Hydrazine (50  $\mu\text{M}$ ), 3. Cys, 4. Hcy, 5. GSH,

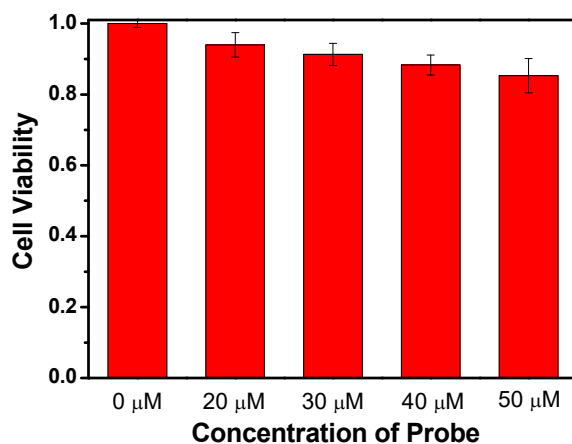
6. NaF, 7. NaCl, 8. NaBr, 9. NaI, 10. Na<sub>2</sub>S<sub>2</sub>O<sub>7</sub>, 11. Na<sub>2</sub>SO<sub>3</sub>, 12. Na<sub>2</sub>CO<sub>3</sub>, 13. NaHS, 14. NaNO<sub>3</sub>, 15. NaSCN, 16. NaAc, 17. NH<sub>3</sub>·H<sub>2</sub>O, 18. NH<sub>2</sub>OH·HCl, 19. H<sub>2</sub>NCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>, 20. HOCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>, 21. C<sub>6</sub>H<sub>5</sub>NH<sub>2</sub>, 22. C<sub>6</sub>H<sub>5</sub>CH<sub>2</sub>NH<sub>2</sub>, 23. NaClO, 24. H<sub>2</sub>O<sub>2</sub>, 25. NaNO<sub>2</sub>, 26. <sup>t</sup>BuOO<sup>•</sup>, 27. <sup>•</sup>OH, 28. NaN<sub>3</sub>, 29. Na<sub>3</sub>PO<sub>4</sub>, 30. Na<sub>2</sub>SO<sub>4</sub>, 31. Na<sub>2</sub>S. All experiments were performed in PBS buffer (10 mM, pH 7.4) with 2% CH<sub>3</sub>CN at 37 °C, and each spectrum was obtained 10 min after addition of an analyte.



**Fig. S8.** (a) Emission color changes and (b) color changes of probe **1** (10 μM) upon addition of 100 μM (except hydrazine 50 μM) of different analytes in PBS buffer (10 mM, pH 7.4,) with 2% CH<sub>3</sub>CN (v/v) at room temperature. Each vial from left to right: blank, hydrazine, Cys, Hcy, GSH, NaF, NaCl, NaBr, NaI, Na<sub>2</sub>S<sub>2</sub>O<sub>7</sub>, Na<sub>2</sub>SO<sub>3</sub>, NaAc, NaHS, NaSCN, NH<sub>3</sub>·H<sub>2</sub>O, NH<sub>2</sub>OH·HCl, H<sub>2</sub>NCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>, HOCH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>, C<sub>6</sub>H<sub>5</sub>NH<sub>2</sub>, C<sub>6</sub>H<sub>5</sub>CH<sub>2</sub>NH<sub>2</sub>, NaClO, H<sub>2</sub>O<sub>2</sub>, NaNO<sub>2</sub>, <sup>t</sup>BuOO<sup>•</sup> and <sup>•</sup>OH). The fluorescent color changes were observed under a portable 365 nm UV lamp. Visual color changes were observed under ambient light.

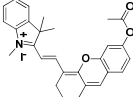
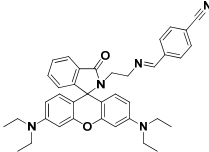
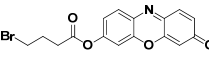
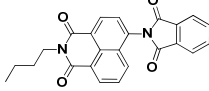
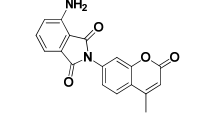
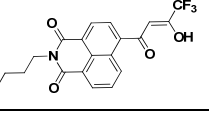
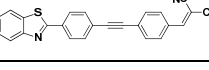
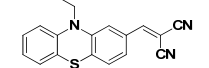


**Fig. S9.** (a) Fluorescence spectra changes of probe **1** (10 μM) toward various analytes (100 μM, except hydrazine, 50 μM). (b) Fluorescence intensity changes of probe **1** (10 μM) at 558 nm toward 100 μM (except: hydrazine 50 μM) of various analytes including: 1. none, 2. hydrazine, 3. K<sup>+</sup>, 4. Ca<sup>2+</sup>, 5. Na<sup>+</sup>, 6. Mg<sup>2+</sup>, 7. Zn<sup>2+</sup>, 8. Cu<sup>2+</sup>, 9. Trp, 10. Phe, 11. Gln, 12. Ala, 13. Leu, 14. Thr, 15. Ser, 16. Asp, 17. Ile, 18. Met, 19. Lys, 20. Gly, 21. Arg, 22. Tyr, 23. Pyr, 24. His. All experiments were performed in PBS buffer (10 mM, pH 7.4) with 2% CH<sub>3</sub>CN at 37 °C, and each spectrum was obtained 10 min after addition of an analyte. λ<sub>ex</sub> = 450 nm, slit width: d<sub>ex</sub> = 5 nm, d<sub>em</sub> = 10 nm.

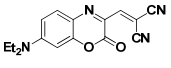
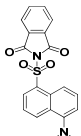
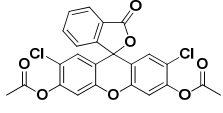
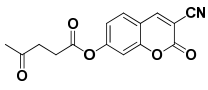
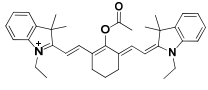
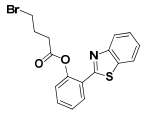
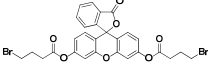
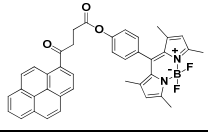
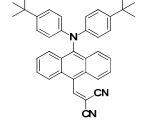
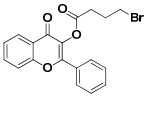
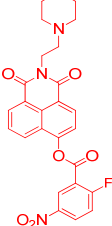


**Fig. S10.** Percentage of viable HeLa cells after treatment with indicated concentrations of probe after 24 hours. The cell viability was observed via MTT assay.

### 3. Table S1. Comparison of some representative fluorescent hydrazine probes

Probes	Detection system	Range of linear correlation / $\mu\text{M}$	LOD	Time / min	Ref.
	DMSO-HEPES buffer (1:4, v/v)	0-50	0.17 $\mu\text{M}$ (5.4 ppb).	10	Anal. Chem. 2015, 87, 9101
	$\text{CH}_3\text{CN}$ -HEPES buffer (9:1, v/v)	1-150	58 nM	--	Anal. Chim. Acta 2015, 893, 84
	$\text{CH}_3\text{CN}$ -HEPES buffer (1:9, v/v)	10-200	2 $\mu\text{M}$	60	Biosens. Bioelectron. 2014, 58, 282
	$\text{H}_2\text{O}$ -DMSO (4:6, v/v)	1-50	0.3 ppb	2	Chem. Commun., 2014, 50, 1485
	$\text{H}_2\text{O}$ -DMSO (3:7, v/v)	0.1-1.0	3.2 ppb	0.5	Anal. Chem., 2014, 86, 4611
	$\text{H}_2\text{O}$ - $\text{CH}_3\text{CN}$ (1:9, v/v)	0-5	3.2 ppb	60	Chem. Sci. 2013, 4, 4121
	$\text{H}_2\text{O}$ -THF (1:1, v/v)	0.06-0.12	0.11 ppb	3	Dyes Pigm. 2013, 99, 966
	Tris.HCl buffer-DMF (3/7, v/v)	5-20	12.191 nM	--	J. Mater. Chem. B, 2014, 2, 1846



	DMSO-acetate buffer (9:1, v/v)	0.5-3.5	13.4 ppb	20	Chem. Commun., 2012,48, 8117
	DMSO-HEPES buffer (9:1, v/v)	0-5	6.01 ppb	60	J. Mater. Chem. B, 2014,2, 7344
	DMSO- Tris buffer solution (1 : 1, v/v)	0-25	2.9 ppb	10	Org. Biomol. Chem. 2013, 11, 2961
	DMSO- acetate buffer (7:3, v/v)	0-14	2.46 μM	15	Org. Lett., 2011,13,5260
	DMSO-acetate buffer (9:1, v/v)	10.0-80.0	0.81 ppb	40	Org. Lett., 2013, 15, 4022
	CH <sub>3</sub> CN-HEPES buffer (2:3, v/v)	1-9.5	2.2 ppb	15	Org. Lett., 2013, 15, 5412
	CH <sub>3</sub> OH-H <sub>2</sub> O (1 : 1, v/v,)	2-39	38.81nM	15	RSC Adv., 2014, 4, 14210
	DMSO-HEPES buffer (7:3, v/v)	0-10	1.87 μM	--	RSC Adv., 2015, 5, 58228
	CH <sub>3</sub> CN	0-5.5	0.2 μM (7 ppb) in CH <sub>3</sub> CN	--	Sens. Actuators, B 2014,199,93
	DMSO-PBS buffer (9:1, v/v)	0-50	0.15 μM (4.8 ppb)	30	Sens. Actuators, B 2015,216,141
	PBS buffer (2% CH <sub>3</sub> CN, v/v)	0-20	0.10 μM (~3 ppb)	10	This work