# **Supporting Information for:**

# A fully disposable paper-based electrophoresis microchip with integrated pencil-drawn electrodes for contactless conductivity detection

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## Experimental

#### 1. Chemicals

All chemicals were used without further purification, in analytical grade, and purchased from Sigma-Aldrich, St. Louis, Missouri, USA. The artificial serum was purchased from Doles Reagents (Goiânia, GO, Brazil).

#### 2. Instrumentation for Electrophoresis and Contactless Conductivity Detection

For the experiments with paper-based microchip, it was used a bipolar two-channel high voltage sequencer from eDAQ (model ER230, Denistone East, NSW, Australia) and a labmade system developed based on the electronics described by da Silva and co-workers.<sup>1</sup> A function generator (model GV-2002, ICEL, Manaus, Brazil) was used to apply the high-frequency sinusoidal wave. The measured current was converted to voltage, rectified, filtered, amplified, and monitored using an A/D interface (model NI USB-6009) from National Instruments. The data acquisition was performed with 1ms of resolution by using a software written in LabVIEW<sup>TM</sup> v. 2010.

Experiments of zone electrophoresis on glass microchips were performed using a Quad HV microchip electrophoresis system (model ER455) supplied by eDAQ (Denistone East, NSW, Australia). The system comprises two high-voltage sequencers model ER230 and a C<sup>4</sup>D model ER225. This system is coupled with microfluidic platform model ET-225 (Micronit Microfluids Enschede, Netherlands). Electrophoretic separations were performed using commercial glass microchips model ET190 with integrated sensing electrodes from Micronit Microfluids (Enschede, Netherlands). Glass microchips comprise two channels (100  $\mu$ m width  $\times$  10  $\mu$ m depth) arranged in double-T geometry with gap of 100  $\mu$ m and the electrophoresis chip consisted of a 40 mm long separation channel, 0.7 cm long side arms and effective length equal to 33 mm. Two parallel electrodes measuring 200  $\mu$ m wide  $\times$  500  $\mu$ m long  $\times$  200 nm thick spaced by 250  $\mu$ m were used for C<sup>4</sup>D

measurements. PowerChrom software (version 2.7.9) was used for the acquisition, display, and analysis of electropherograms. All the sequences of high voltages were controlled using either the eDAQ Sequencer software (version 1.3.3).

## 3. Results

3.1 Ohm plot



*Figure S1.* Plot of current versus voltage for paper-based microchannels filled with running buffer composed of MES/His (20 mM, each), pH 6.1.

## 3.2. Evaluation of different electrode arrangements and injection-to-injection repeatability



*Figure S2.* Electropherograms showing five consecutive injections of a standard solution containing 500  $\mu$ mol L<sup>-1</sup> Na<sup>+</sup> for electrode cell attached on (a) top, (b) bottom, and (c) top-bottom sides.

**Table S1.** Comparison of the analytical performance for the different electrodes arrangements (n = 5).

Electrode Arrangement	Intensity (V)	Area (V.s)	Migration Time (s)	Efficiency (N.m <sup>-1</sup> )	Asymmetry
Тор	0.48 ± 0.06	9.89 ± 1.24	134.2 ± 1.6	3959 ± 167	0.52 ± 0.04
Bottom	0.62 ± 0.03	16.64 ± 1.05	153.6 ± 2.1	9161 ± 419	0.30 ± 0.02
Top-Bottom	0.52 ± 0.04	17.93 ± 1.80	167.8 ± 1.0	3871 ± 300	0.25 ± 0.02



*Figure S3.* Electropherograms of ten consecutive injections for a Na<sup>+</sup> standard solution (250  $\mu$ mol L<sup>-1</sup>).

## 3.3. Analytical Curves



*Figure S4.* Calibration curves for (a) BSA and (b) creatinine on paper-based electrophoresis microchips.

## 3.4. Detection of BSA and creatinine in artificial serum samples



*Figure S5.* Electropherograms showing five analysis of (a) BSA and (b) creatinine in artificial serum sample using paper-based electrophoresis microchips.

Table S2. Chemical composition of artificial serum sample.

Analyte	Average Concentration	
Uric Acid	4.08 mg/dL	
Albumin	2.06 g/dL	
Alanine aminotransferase / Glutamic-pyruvate transaminase	55 IU/L	
Amylase	234 U/dL	
Aspartate aminotransferase/Glutamic-oxaloacetic	63 IU/L	
transaminase		
Billirubin	0.79 mg/dL	
Calcium	8.96 mg/dL	
Total Iron Binding Capacity	228 μg/dL	
Chloride	91 mmol/L	
Total Cholesterol	125 mg/dL	
HDL Cholesterol	64 mg/dL	
LDL Cholesterol	39 mg/dL	
Creatinine	1.04 mg/dL	
Lactate dehydrogenase	260 IU/L	
Iron	84 μg/dL	
Alkaline Phosphatase	144 U/L	
Phosphate	3.71 mg/dL	
γ-glutamiltranspeptidase	56 IU/L	
Glucose	90 mg/dL	
Magnesium	1.62 mg/dL	
Potassium	3.46 mmol/L	
Total protein	3.69 g/dL	
Triglycerides	89 mg/dL	
Urea	33 mg/dL	

## 3.5. Detection of BSA and creatinine on commercial glass electrophoresis chips



*Figure S6.* Electropherograms showing the individual analysis of (a) BSA and (b) creatinine solutions as well as a mixture of both analytes on commercial glass electrophoresis devices.

### 4. References

<sup>(1)</sup> da Silva, J. A. F.; Guzman, N.; do Lago, C. L. J. Chromatogr. A 2002, 942, 249-258.