

## Supplementary Information

# Calcium Phosphate-Based Organic-Inorganic Hybrid Nanocarriers with pH-Responsive On/Off Switch of Photodynamic Therapy

Takahiro Nomoto,<sup>a,b</sup> Shigeto Fukushima,<sup>c</sup> Michiaki Kumagai,<sup>c</sup> Kozo Miyazaki,<sup>d,e</sup> Aki Inoue,<sup>c</sup> Peng Mi,<sup>b</sup> Yoshinori Maeda,<sup>a,e</sup> Kazuko Toh,<sup>f</sup> Yu Matsumoto,<sup>e,g</sup> Yuji Morimoto,<sup>d</sup> Akihiro Kishimura,<sup>c,h,i</sup> Nobuhiro Nishiyama\*<sup>b</sup> and Kazunori Kataoka\*<sup>a,c,e,f</sup>

**a.** Department of Bioengineering, Graduate School of Engineering, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8656, Japan, **b.** Polymer Chemistry Division, Chemical Resources Laboratory, Tokyo Institute of Technology, R1-11, 4259 Nagatsuta, Midori-ku, Yokohama 226-8503, Japan, **c.** Department of Materials Engineering, Graduate School of Engineering, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8656, Japan, **d.** Department of Integrative Physiology and Bio-Nano Medicine, National Defense Medical College, 3-2 Namiki, Tokorozawa, Saitama 359-8513, Japan, **e.** Center for Disease Biology and Integrative Medicine, Graduate School of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033, Japan, **f.** Innovation Center of Nanomedicine, Kawasaki Institute of Industry Promotion, 66-20 Horikawa-cho, Saiwai-ku, Kawasaki 212-0013, Japan, **g.** Department of Otorhinolaryngology and Head and Neck Surgery, Graduate School of Medicine and Faculty of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033, Japan, **h.** Department of Applied Chemistry, Faculty of Engineering, Kyushu University, 744 Moto-oka, Nishi-ku, Fukuoka 819-0395, Japan, **i.** Center for Molecular Systems, Kyushu University, 744 Moto-oka, Nishi-ku, Fukuoka 819-0395, Japan

Table of contents

**Supplementary Methods**

**Supplementary Figure S1. XRD patterns of CaP incorporating Ce6**

**Supplementary Figure S2. FT-IR spectra of CaP incorporating Ce6**

**Supplementary Figure S3. <sup>1</sup>H NMR of Ce6 encapsulated in hydrothermally synthesized CaP**

**Captions for Supplementary Movies S1–2**

## Supplementary Methods

### **X-ray diffraction (XRD)**

To evaluate crystal phase of calcium phosphate (CaP) in the nanocarrier, CaP incorporating chlorin e6 (Ce6) was prepared in the absence of poly(ethylene glycol)-*b*-poly(L-aspartic acid) (PEG-PAsp). Solution of 2.5 M CaCl<sub>2</sub> was diluted in 10 mM Tris/HCl buffer (pH 7.6) (2 mL: 18 mL). This solution was added into the same volume of another solution containing Ce6 (100 μg/mL) in 50 mM HEPES buffer containing 6 mM Na<sub>2</sub>HPO<sub>4</sub> and 140 mM NaCl (pH 7.1) to prepare CaP incorporating Ce6. The mixed aqueous solution was then incubated at 37 °C for at least 30 min. For hydrothermal synthesis, the CaP incorporating Ce6 was autoclaved at 120 °C for 60 min, using LSX-700 (TOMY Seiko Co., Ltd., Tokyo, Japan). The CaP incorporating Ce6 with or without hydrothermal synthesis was spun down, and the supernatant was carefully removed. These precipitates were washed with ethanol, and centrifuged, followed by removal of the supernatant. The prepared CaP incorporating Ce6 was then put in an aluminum mold. After drying the samples by air, the X-ray diffraction was measured with Cu Kα radiation using XRD-DSC II (RIGAKU Corporation, Tokyo, Japan).

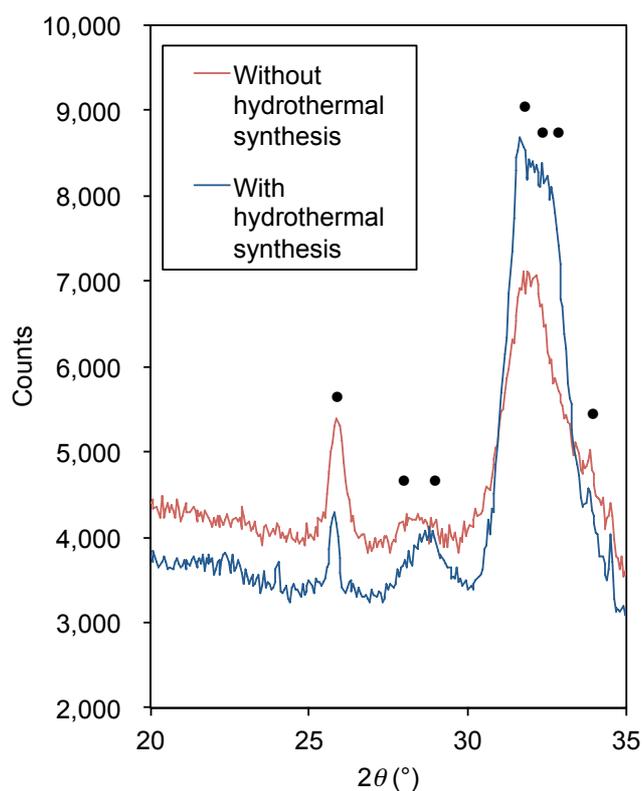
### **Fourier transform infrared (FT-IR) spectroscopy**

CaP incorporating Ce6 was prepared and purified in the same method for preparation of the CaP for XRD measurement. The CaP incorporating Ce6 was dried by air, and FT-IR spectrum was measured using FT/IR-4600 (JASCO Corporation, Tokyo, Japan).

### **Validation of the integrity of Ce6 using <sup>1</sup>H nuclear magnetic resonance (NMR) spectroscopy**

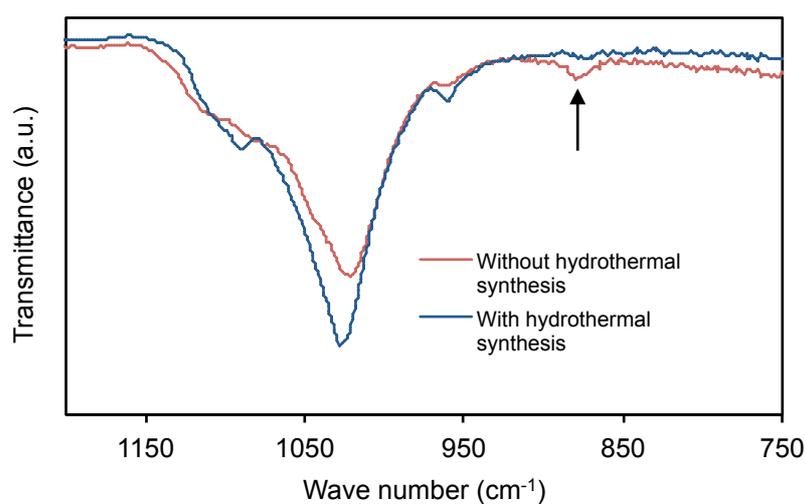
CaP incorporating Ce6 was prepared and purified in the same method for preparation of the CaP for XRD measurement. After vacuum-drying overnight, the hydrothermally synthesized CaP incorporating Ce6 was dissolved in the mixture of 0.5 N deuterium chloride and dimethyl sulfoxide-d<sub>6</sub> (v/v=1), and characterized by <sup>1</sup>H NMR spectroscopy using AVANCE III 400 (Bruker BioSpin K.K., Kanagawa, Japan).

## Supplementary Figures S1–S3



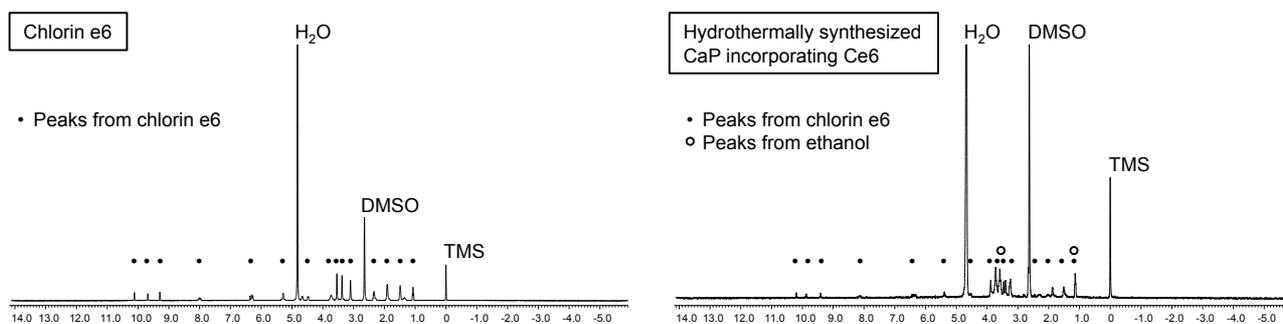
### Supplementary Figure S1. XRD patterns of CaP incorporating Ce6.

XRD patterns of CaP incorporating Ce6 with/without hydrothermal synthesis were measured on aluminum molds. Closed circles highlight typical peaks from hydroxyapatite.



### Supplementary Figure S2. FT-IR spectra of CaP incorporating Ce6.

FT-IR spectra of CaP incorporating Ce6 with/without hydrothermal synthesis were measured in solid form. The arrow indicates the peak corresponding to CO<sub>3</sub><sup>2-</sup>.



### Supplementary Figure S3. <sup>1</sup>H NMR of Ce6 encapsulated in hydrothermally synthesized CaP.

<sup>1</sup>H NMR spectra of Ce6 and Ce6 encapsulated in hydrothermally synthesized CaP were measured in the mixture of 0.5 N deuterium chloride and dimethyl sulfoxide-d<sub>6</sub> (v/v=1). Closed circles and open circles highlight the chemical shifts of chlorin e6 and ethanol that was used to wash the CaP, respectively.

### Captions for Supplementary Movies

#### Supplementary Movie S1. Intravital microscopic imaging of the bloodstream in the mouse injected with CaPCe6.

CaPCe6 was intravenously injected into a mouse and its blood vessels in the ear lobe were monitored using IVRTCLSM at 30 fps. Fluorescence of Ce6 and Dio-labeled red blood cells is shown in red and green, respectively.

#### Supplementary Movie S2. Intravital microscopic imaging of the bloodstream in the mouse injected with Ce6.

Ce6 was intravenously injected into a mouse and its blood vessels in the ear lobe were monitored using IVRTCLSM at 30 fps. Fluorescence of Ce6 and Dio-labeled red blood cells is shown in red and green, respectively.