

Directly linked hydroporphyrin dimers

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General experimental procedures

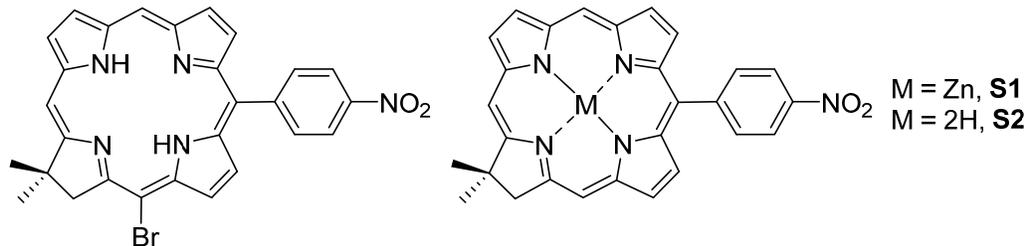
Synthesis. Microwave heating was performed using a Biotage Initiator microwave reactor. ^1H NMR (500 or 400 MHz) and ^{13}C NMR (125 or 100 MHz) spectra were recorded on a Bruker 500 MHz or a Bruker or a Varian 400 MHz instrument. Chemical shifts were referenced to residual solvent peaks and are given as follows: chemical shift (δ , ppm), multiplicity [s, singlet; br, broad; d, doublet, m, multiplet), coupling constant (Hz), integration]. LC-MS analysis was carried out using an Agilent 1100 and Waters micromass ZQ tandem system. HR-ESI-MS analyses were performed at the Organisch Chemisches Institut WWU Münster. All compounds displayed the expected isotope distribution pattern. Anhydrous CH_2Cl_2 was obtained by distillation from CaH_2 under an Ar atmosphere. Anhydrous THF was available from a VAC solvent purifier. **Chl**,¹ **Chl10^{Ph}**,¹ **Chl10^{Mes}**,¹ **Chl10^{PhNO2}-Zn**,² **Chl15^{Br}**,³ **Chl10^{Ph}15^{Br}**,³ **Chl10^{Mes}15^{Br}**,³ **Chl3^{Br}**,⁴ **Chl13^{Br}**,⁵ and **Chl15^{Ph}** (Ref 3) were synthesized following literature methods. All other chemicals were from commercial sources and used as received.

Chromatography. Preparative chromatography was performed using silica (230–400 mesh). Thin layer chromatography was performed on silica-coated aluminum plates. Samples were visualized by UV-light (254 and 356 nm), or staining with $\text{KMnO}_4/\text{K}_2\text{CO}_3$ or cerium ammonium molybdate.

Photophysical measurements. UV-Vis absorption spectroscopy was performed on a Varian Cary 300 instrument; (sh) denotes shoulder to a peak. Steady state and time-resolved emission spectra were collected at room temperature using a Horiba Scientific FluoroMax 4 instrument equipped with a flash lamp. Phosphorescence mission intensity changes were fitted to single exponential decays using the instrument's software (FluorEssence Version 3.5.1.20, based on Origin 8.1090). Fitting to double exponential was less suitable based on R^2 and χ .

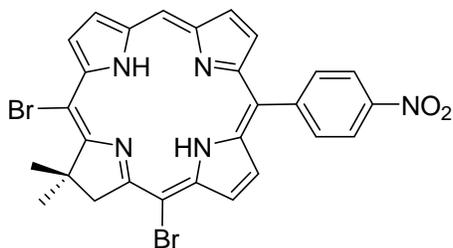
Electrochemistry. Cyclic voltammetry measurements were performed on 1 mM solutions of the analytes in dry CH_2Cl_2 using NBu_4PF_6 (0.1 M solution) as supporting electrolyte. Glassy C-electrode was used as the working electrode and Ag/AgNO_3 as the reference electrode. All measurements were done at $v = 100$ mV/s scanning rate. All solutions were degassed by 15 min bubbling of Ar through the electrochemical cell. At the end of the measurement a small amount of ferrocene was added to each solution as external reference. All potentials are given versus $\text{Fc}^{+/0}$.

Synthesis

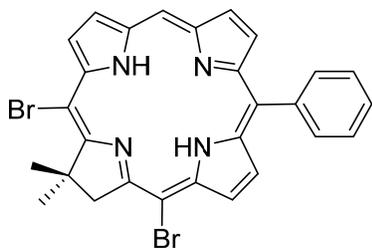


Chl10^{PhNO₂}**15**^{Br}. Zn(II)-chlorin (**S1**,² 31 mg, 0.059 mmol) was dissolved in CH₂Cl₂/TFA (20 mL, 1:1), and the solution was stirred at room temperature for 12 h. The TFA was neutralized by the dropwise addition of 2 M aqueous NaOH. The phases were separated, and the aqueous layer was extracted with CH₂Cl₂ twice. The combined organic phases were washed with saturated aqueous Na₂CO₃, dried over MgSO₄, filtered, and eluted with heptane:CH₂Cl₂ (1:1) through a short silica column. The solvent was evaporated, and the solid residue was dried briefly under vacuum (**S2**, 23.4 mg, 86%), and then used for the bromination. A solution of the free base chlorin obtained in the previous step (20.7 mg, 0.0449 mmol) was dissolved in a mixture of CH₂Cl₂ and TFA (10:1, 11 mL), and was treated with a single portion of NBS (7.2 mg, 0.040 mmol, 0.9 equiv.). The mixture was stirred at room temperature in the dark until TLC-analysis indicated the progress of the reaction [silica, heptane:CH₂Cl₂ (2:1)]. Aqueous NaOH (2 M) was added dropwise to quench the TFA. The phases were separated, and the aqueous phase was extracted with CH₂Cl₂. The combined organic phase was washed with saturated aqueous Na₂CO₃, and was dried over MgSO₄, filtered, and concentrated. Purification by column chromatography [silica, heptane:CH₂Cl₂ (2:1)] yielded first the dibrominated chlorin, and then the monobrominated product as a brown solid (18 mg, 83%): ¹H NMR (400 MHz, CDCl₃) -2.26 (s, 1H), -2.08 (s, 1H), 2.06 (s, 6H), 4.71 (s, 2H), 8.25 (d, *J* = 8.8 Hz, 2H), 8.45 (d, *J* = 4.4 Hz, 1H), 8.57 (d, *J* = 8.8 Hz, 2H), 8.61 (d, *J* = 4.8 Hz, 1H), 8.90–8.91 (m, 2H), 8.96 (d, *J* = 4.8 Hz, 1H), 9.21 (d, *J* = 4.4

Hz, 1H), 9.25 (d, $J = 4.8$ Hz, 1H), 9.76 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) 31.5, 46.3, 55.2, 95.5, 96.9, 106.7, 120.1, 121.8, 122.1, 124.7, 125.2, 127.4, 129.0, 132.4, 132.8, 133.9, 134.49, 134.54, 136.1, 138.1, 131.2, 147.7, 148.7, 150.8, 152.9, 162.8, 177.0; HR-ESI-MS calcd 540.1030 obsd 540.1039 $[(\text{M} + \text{H})^+]$, $\text{M} = \text{C}_{28}\text{H}_{22}\text{BrN}_5\text{O}_2$; $\lambda_{\text{abs}} = 280, 408, 505, 535, 590, 642$ (CH_2Cl_2); $\lambda_{\text{em}} = 646$ nm ($\lambda_{\text{ex}} = 408$ nm, CH_2Cl_2).

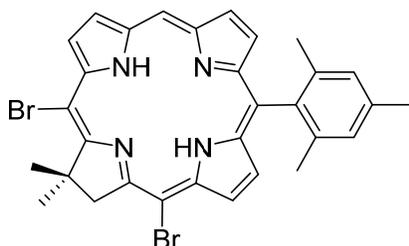


Chl10^{PhNO₂}-15^{Br}-20^{Br}. Characterisation data for the dibrominated side-product (trace amounts formed): ^1H NMR (500 MHz, CDCl_3) -2.22 (s, 1H), -1.72 (s, 1H), 2.31 (s, 6H), 4.82 (s, 2H), 8.29 (d, $J = 8.5$ Hz, 2H), 8.45 (d, $J = 4.1$ Hz, 1H), 8.62 (d, $J = 8.5$ Hz, 2H), 8.66 (d, $J = .2$ Hz, 1H), 8.93 (d, $J = 4.1$ Hz, 1H), 9.22 (dd, $J_1 = 4.6$ Hz, $J_2 = 1.7$ Hz, 1H), 9.34 (d, $J = 4.6$ Hz, 1H), 9.46 (dd, $J_1 = 4.6$ Hz, $J_2 = 1.7$ Hz, 1H), 9.81 (s, 1H); ESI-MS calcd 617.0 obsd 617.9 $[(\text{M} + \text{H})^+]$, $\text{M} = \text{C}_{28}\text{H}_{21}\text{Br}_2\text{N}_5\text{O}_2$ $\lambda_{\text{abs}} = 306, 370, 405, 514, 542, 598, 649$ nm (CH_2Cl_2); $\lambda_{\text{em}} = 652$ nm ($\lambda_{\text{ex}} = 405$ nm, CH_2Cl_2).



Chl10^{Ph}-15^{Br}-20^{Br}. Characterisation data for the dibrominated species: (4 mg, 7.5%): ^1H NMR (500 MHz, CDCl_3) -2.11 (s, 1H), -1.63 (s, 1H), 2.30 (s, 6H), 4.80 (s, 2H), 7.73–7.76 (m, 3H), 8.09–8.11 (m, 2H), 8.55 (d, $J = 4.6$ Hz, 1H), 8.75 (d, $J = 5.2$ Hz, 1H), 8.88 (d, $J = 4.1$ Hz, 1H),

9.18 (dd, $J_1 = 4.6$ Hz, $J_2 = 1.7$ Hz, 1H), 9.27 (d, $J = 4.6$ Hz, 1H), 9.42 (dd, $J_1 = 4.6$ Hz, $J_2 = 1.7$ Hz, 1H), 9.77 (s, 1H); ESI-MS calcd 572.0 obsd 572.9 [(M + H)⁺, M = C₂₈H₂₂ Br₂N₄]; $\lambda_{\text{abs}} = 308, 370, 405, 515, 542, 597, 649$ nm (CH₂Cl₂); $\lambda_{\text{em}} = 653$ nm ($\lambda_{\text{ex}} = 405$ nm, CH₂Cl₂).



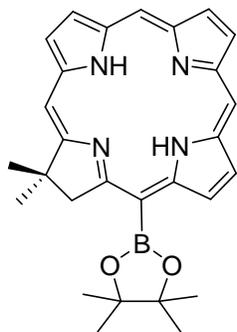
Chl10^{Mes}15^{Br}20^{Br}. Characterisation data for the dibrominated species: (10.8 mg, 14%): ¹H NMR (400 MHz, CDCl₃) -2.03 (s, 1H), -1.62 (s, 1H), 1.84 (s, 6H), 2.28 (s, 6H), 2.60 (s, 3H), 4.78 (s, 2H), 7.25 (s, 2H), 8.37 (d, $J = 4.4$ Hz, 1H), 8.58 (s, $J = 4.8$ Hz, 1H), 8.81 (d, $J = 4.4$ Hz, 1H), 9.13–9.14 (m, 1H), 9.21 (d, $J = 4.8$ Hz, 1H), 9.38–9.39 (m, 1H), 9.71 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) 21.2, 29.1, 31.3, 46.1, 48.8, 60.4, 95.8, 96.8, 109.0, 121.7, 124.9, 126.0, 126.6, n127.8, 128.8, 131.7, 133.7, 135.0, 136.1, 136.9, 137.8, 138.3, 138.9, 139.1, 140.5, 151.7, 153.9, 161.8, 172.2; ESI-MS calcd 614.0 obsd 614.9 [(M + H)⁺, M = C₃₁H₂₈Br₂N₄]; $\lambda_{\text{abs}} = 308, 369, 406, 514, 540, 597, 650$ nm (CH₂Cl₂); $\lambda_{\text{em}} = 653$ nm ($\lambda_{\text{ex}} = 406$ nm, CH₂Cl₂).

General procedure for chlorin borylation

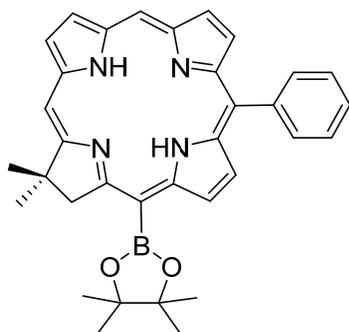
The reaction is based on a borylation described in reference 6.

A solution of bromochlorin (1.0 equiv.) in dichloroethane/Et₃N (6:1, 10 mM) was treated with PdCl₂(PPh₃)₂ (0.2 equiv.). The mixture was deoxygenated with three rounds of freeze-pump-thaw, the flask was back-filled with Ar, HBpin (10 equiv.) was added, and the flask was immersed into an oil bath heated at 90 °C. Heating was continued for 16 h. The reaction mixture was allowed to cool back to room temperature. Saturated aqueous NH₄Cl and CH₂Cl₂ were added. The phases were separated, and the aqueous phase was extracted with CH₂Cl₂ twice. The

combined organic phase was washed with saturated aqueous NH_4Cl , dried over MgSO_4 , filtered and concentrated. The residue was purified by column chromatography [silica, heptane/ CH_2Cl_2 (1:1)] yielding the borylated chlorin as a dark green-gray solid.

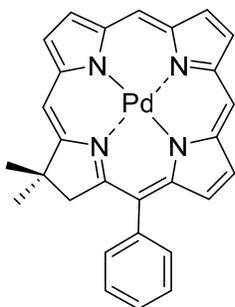


Chl15^{Bpin}, 21 mg, 70%; ^1H NMR (500 MHz, CDCl_3) -1.94 (br, 1H), -1.70 (br, 1H), 1.77 (s, 12H), 2.06 (s, 6H), 4.86 (s, 2H), 8.91 – 8.93 (m, 2H), 9.03 (s, 2H), 9.19 (d, $J = 4.5$ Hz, 1H), 9.23 – 9.25 (m, 1H), 9.56 – 9.58 (m, 1H), 9.79 (s, 1H), 9.84 (s, 1H); ^{13}C NMR (125 MHz, CDCl_3) 25.4 , 31.8 , 46.1 , 53.5 , 84.5 , 94.7 , 107.6 , 107.7 , 123.6 , 125.0 , 127.9 , 129.3 , 129.6 , 132.6 , 132.2 , 134.2 , 135.0 , 140.5 , 145.3 , 151.2 , 151.5 , 170.0 , 175.1 ; HR-ESI-MS calcd 467.2618 obsd 467.2622 $[(\text{M} + \text{H})^+]$, $\text{M} = \text{C}_{28}\text{H}_{31}\text{BN}_4\text{O}_2$, calcd 489.2437 obsd 489.2448 $(\text{M} + \text{Na})^+$; $\lambda_{\text{abs}} = 276$, 393 , 495 , 522 , 585 , 638 nm (CH_2Cl_2); $\lambda_{\text{em}} = 643$ nm ($\lambda_{\text{ex}} = 393$ nm, CH_2Cl_2).



Chl10^{Ph}15^{Bpin}, 125 mg, 86%; ^1H NMR (400 MHz, CDCl_3) -1.70 (s, 2H), 1.73 (s, 12H), 2.03 (s, 6H), 4.79 (s, 2H), 7.70 – 7.72 (m, 3H), 8.11 (dd, $J_1 = 7.4$ Hz, $J_2 = 1.2$ Hz, 2H), 8.52 (d, $J = 4.0$ Hz, 1H), 8.74 (d, $J = 4.8$ Hz, 1H), 8.84 (s, 1H), 8.86 – 8.89 (m, 2H), 9.16 (d, $J = 4.4$ Hz, 1H), 9.35 (d,

$J = 4.8$ Hz, 1H), 9.75 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) 25.2, 31.5, 46.0, 53.2, 84.4, 94.2, 108.0, 122.3, 123.2, 124.4, 126.6, 127.5, 128.1, 129.2, 131.8, 132.1, 133.9, 134.46, 134.52, 140.9, 142.3, 144.5, 151.0, 152.3, 169.0, 175.6; HR-ESI-MS calcd 543.2932 obsd 543.2930 [(M + H) $^+$, M = $\text{C}_{34}\text{H}_{35}\text{BN}_4\text{O}_2$], calcd 565.2751 obsd 565.2759 [(M + Na) $^+$]; $\lambda_{\text{abs}} = 279, 404, 502, 529, 589, 616, 641$ nm (CH_2Cl_2); $\lambda_{\text{em}} = 646$ nm ($\lambda_{\text{ex}} = 404$ nm, CH_2Cl_2).

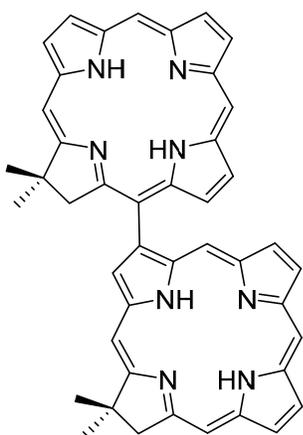


Chl15^{Ph}-Pd. A sample of **Chl15^{Ph}** (Ref 3) (6.2 mg, 0.012 mmol) in a microwave vial was dissolved in pyridine (1 mL), and the solution was treated with $\text{Pd}(\text{acac})_2$ (18 mg, 0.06 mmol). The vial was sealed, and the reaction mixture was heated in a microwave reactor at 180 °C for 30 min. The reaction mixture was allowed to cool to room temperature, and the solvent was evaporated at reduced pressure. The dark residue was dissolved in a small amount of CH_2Cl_2 , and the solution was loaded onto a silica gel chromatography column. Elution with Heptane/ CH_2Cl_2 (2:1) yielded a bright red solid (6.2 mg, quant.): ^1H NMR (500 MHz, CDCl_3) 1.93 (s, 6H), 4.27 (s, 2H), 7.64–7.68 (m, 3H), 7.84–7.86 (m, 2H), 8.16 (d, $J = 4.6$ Hz, 1H), 8.77 (d, $J = 4.6$ Hz, 1H), 8.85 (s, 1H), 8.88 (d, $J = 4.6$ Hz, 1H), 8.96 (s, 1H), 9.00 (d, $J = 4.6$ Hz, 1H), 9.79 (d, $J = 5.2$ Hz, 1H); ^{13}C NMR (125 MHz, CDCl_3) 31.0, 45.7, 51.2, 96.6, 109.8, 110.8, 113.4, 127.1, 128.2, 128.4, 128.5, 128.7, 132.0, 133.1, 138.5, 138.8, 139.3, 139.6, 143.6, 145.8, 146.5, 146.5, 149.4, 162.5; ESI-MS calcd 520.0 obsd 520.8 [(M) $^-$, M = $\text{C}_{28}\text{H}_{22}\text{N}_4\text{Pd}$]; $\lambda_{\text{abs}} = 275, 335, 391, 480, 546, 587$ nm (THF); $\lambda_{\text{abs}} = 269, 337, 392, 480, 548, 589$ nm (CH_2Cl_2); steady-

state emission: $\lambda_{em} = 597, 645 \text{ nm}$ ($\lambda_{ex} = 392 \text{ nm}$, CH_2Cl_2); time-resolved emission: $\lambda_{phos} = 762 \text{ nm}$ (77 K, $\lambda_{ex} = 391 \text{ nm}$, THF); $\tau = 406 \mu\text{s}$

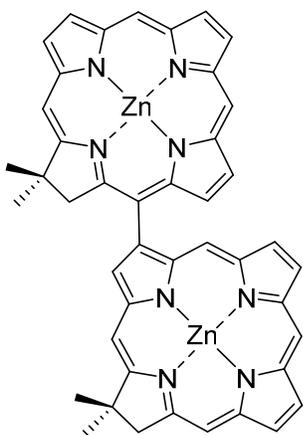
General procedure for chlorin Suzuki coupling

A solution of bromochlorin and chlorin-Bpin (1.0 equiv. each for **Chl15^{Bpin}**, 1.0 and 2.0 equiv., respectively, when **Chl10^{Ph}15^{Bpin}** was used) in toluene/DMF (4 mM and 40 mM for **Chl15^{Bpin}** and **Chl10^{Ph}15^{Bpin}**, respectively) was treated with K_2CO_3 or Cs_2CO_3 (6 or 3 equiv.) and $\text{Pd}(\text{PPh}_3)_4$ (0.25 equiv.). The mixture was deoxygenated with three rounds of freeze-pump-thaw, the flask was back-filled with Ar, and the mixture was immersed into a pre-heated oil bath heated at $85 \text{ }^\circ\text{C}$. Heating was continued for 24 h. The reaction mixture was allowed to cool back to room temperature. Saturated aqueous NH_4Cl and CH_2Cl_2 were added. The phases were separated, and the aqueous phase was extracted with CH_2Cl_2 twice. The combined organic phase was washed with saturated aqueous NH_4Cl , dried over MgSO_4 , filtered and concentrated. The residue was purified by column chromatography [silica, heptane/ CH_2Cl_2 (1:1), then, if required, heptane:EtOAc (9:1)] yielding first the dehalogenated and proto-deborylated chlorins, and finally the chlorin dimer as a dark green-brown solid.



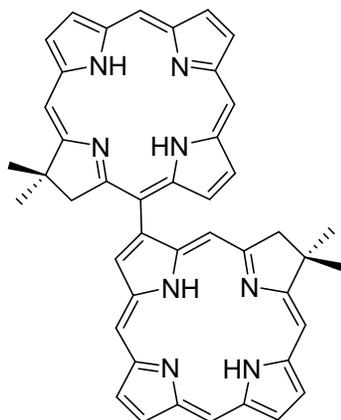
Chl_{23,15}, 12 mg, 52%; $^1\text{H NMR}$ (400 MHz, CDCl_3) -1.90 (s, 1H), -1.80 (s, 1H), -2.27 – (-1.72) (br, 2H), 1.85 (s, 3H), 1.97 (s, 3H), 2.22 (s, 3H), 2.24 (s, 3H), $4.24, 4.49$ (ABq, $J = 17.6 \text{ Hz}$, 4H),

4.79 (s, 2H), 8.40 (d, $J = 4.4$ Hz, 1H), 8.48 (d, $J = 4.0$ Hz, 1H), 8.89 (d, $J = 4.4$ Hz, 1H), 9.00 (d, $J = 4.4$ Hz, 1H), 9.05–9.08 (m, 3H), 9.16–9.17 (m, 3H), 9.27–9.28 (m, 2H), 9.31 (s, 1H), 9.34 (d, $J = 4.8$ Hz, 1H), 9.61 (s, 1H), 9.84 (s, 1H), 9.96 (s, 1H), 9.98 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3 , several peaks are doubled, all signals are reported) 29.4, 29.66, 29.70, 31.18, 31.25, 31.43, 31.46, 46.3, 46.8, 52.1, 52.5, 94.6, 95.0, 96.7, 103.9, 106.5, 106.6, 106.7, 107.8, 123.7, 124.1, 126.4, 127.8, 128.2, 132.2, 132.4, 132.6, 132.9, 134.5, 135.0, 135.3, 136.1, 139.0, 139.8, 140.4, 141.6, 142.3, 151.5, 151.9, 152.3, 163.4, 164.5, 175.1, 175.3; HR-ESI-MS calcd 701.3112 obsd 701.3125 $[(\text{M} + \text{Na})^+]$, $\text{M} = \text{C}_{44}\text{H}_{38}\text{N}_4$; $\lambda_{\text{abs}} = 277, 391, 413, 498, 523, 585, 642$ nm (CH_2Cl_2); $\lambda_{\text{em}} = 646$ nm ($\lambda_{\text{ex}} = 413$ nm, CH_2Cl_2).

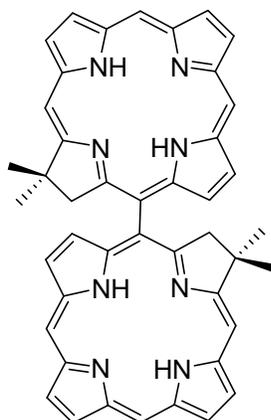


Chl_{23,15}-Zn₂, 4.2 mg, quant.; ^1H NMR (500 MHz, CDCl_3) 1.78 (s, 3H), 1.93 (s, 3H), 2.18 (s, 3H), 2.19 (s, 3H), 4.13, 4.44 (ABq, $J = 17.0$ Hz, 2H), 4.69 (s, 2H), 8.25 (d, $J = 4.0$ Hz, 1H), 8.27 (d, $J = 4.5$ Hz, 1H), 8.71 (d, $J = 4.0$ Hz, 1H), 8.73 (s, 1H), 8.75 (d, $J = 4.0$ Hz, 1H), 8.78 (s, 1H), 8.81 (d, $J = 4.0$ Hz, 1H), 8.83 (d, $J = 4.5$ Hz, 1H), 8.91 (s, 1H), 8.97–9.00 (m, 3H), 9.10 (d, $J = 4.0$ Hz, 1H), 9.13 (d, $J = 4.5$ Hz, 1H), 9.29 (s, 1H), 9.56 (s, 1H), 9.69 (s, 1H), 9.70 (s, 1H); ^{13}C NMR (125 MHz, CDCl_3) 31.5, 32.0, 32.2, 32.3, 32.4, 33.7, 46.5, 47.2, 52.4, 53.3, 95.3, 95.7, 97.5, 106.6, 109.7, 109.8, 110.7, 128.5, 128.7, 129.2, 129.4, 129.5, 129.75, 129.77, 131.9, 133.0, 134.0, 147.9, 148.06, 148.13, 148.29, 148.33, 148.6, 150.2, 151.4, 153.9, 155.0, 155.6, 156.7,

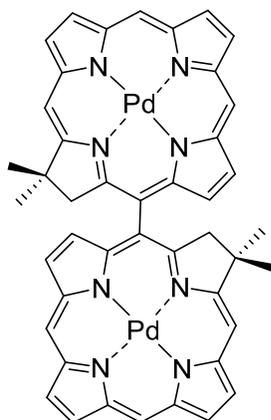
160.2, 160.8, 171.9, 172.2; HR-ESI-MS calcd 802.14948 obsd 802.14959 (M^+ , $M = C_{44}H_{34}N_4Zn_2$); $\lambda_{abs} = 281, 322, 401, 504, 575, 615$ nm (CH_2Cl_2); $\lambda_{em} = 619$ nm ($\lambda_{ex} = 401$ nm, CH_2Cl_2).



Chl₂13,15, 4.2 mg, 32%; 1H NMR (400 MHz, $CDCl_3$) -2.18 (br, 2H), -1.92 (s, 1H), -1.82 (s, 1H), 1.83 (s, 3H), 1.95 (s, 3H), 2.18 (s, 3H), 2.20 (s, 3H), 4.20, 4.46 (ABq, $J = 17.7$ Hz, 2H), 4.82 (s, 2H), 8.39 (d, $J = 4.7$ Hz, 1H), 8.47 (d, $J = 4.2$ Hz, 1H), 8.90 (d, $J = 4.2$ Hz, 1H), 9.05–9.08 (m, 5H), 9.15–9.16 (m, 2H), 9.27 (s, 1H), 9.29 (d, $J = 4.7$ Hz, 1H), 9.33 (d, $J = 4.7$ Hz, 1H), 9.36 (s, 1H), 9.56 (s, 1H), 9.87 (s, 1H), 9.95 (s, 1H), 9.98 (s, 1H); ^{13}C NMR (400 MHz, $CDCl_3$) 31.17, 31.23, 31.4, 31.9, 46.3, 46.7, 52.2, 52.5, 94.7, 95.0, 96.6, 103.9, 106.3, 106.8, 107.8, 123.6, 124.0, 124.1, 124.2, 126.4, 127.9, 128.0, 132.4, 132.9, 134.5, 134.7, 135.3, 136.4, 138.2, 140.4, 140.5, 141.5, 142.4, 151.5, 151.6, 151.8, 152.3, 163.7, 164.4, 175.0, 175.1; HR-ESI-MS calcd 679.3292 obsd 679.3267 [$(M + H)^+$, $M = C_{44}H_{38}N_4$], calcd 701.3112 obsd 701.3107 ($M + Na$) $^+$; $\lambda_{abs} = 277, 392, 413, 497, 584, 642$ nm (CH_2Cl_2); $\lambda_{em} = 645$ nm ($\lambda_{ex} = 413$ nm, CH_2Cl_2).

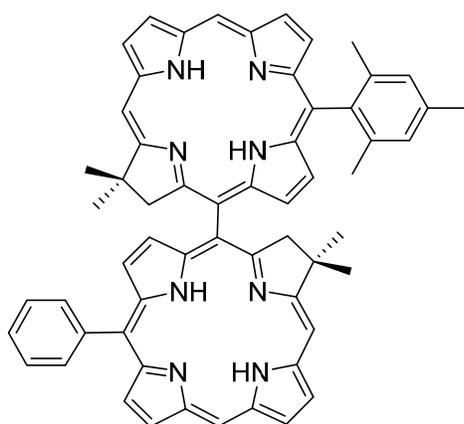


Chl₂15,15, 11 mg, 32%; ¹H NMR (400 MHz, CDCl₃) -1.95 (s, 2H), -1.81 (s, 2H), 1.82 (s, 6H), 1.85 (s, 6H), 3.78, 3.85 (ABq, *J* = 17.2 Hz, 4H), 7.92 (d, *J* = 4.8 Hz, 2H), 9.01 (d, *J* = 4.8 Hz, 2H), 9.06–9.08 (m, 4H), 9.16 (s, 4H), 9.34 (d, *J* = 4.8 Hz, 2H), 9.96 (s, 2H), 9.98 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) 30.8, 31.6, 46.2, 52.6, 95.0, 106.4, 107.8, 110.8, 123.5, 124.2, 127.9, 128.0, 132.4, 132.9, 134.4, 135.4, 140.5, 142.0, 151.5, 152.4, 165.2, 174.7; HR-ESI-MS calcd 679.3293 obsd 679.3292 [(M + H)⁺, M = C₄₄H₃₈N₄], calcd 701.3112 obsd 701.3113 (M + Na)⁺; λ_{abs} = 277, 413, 500, 591, 645 nm (CH₂Cl₂); λ_{em} = 648 nm (λ_{ex} = 413 nm, CH₂Cl₂).



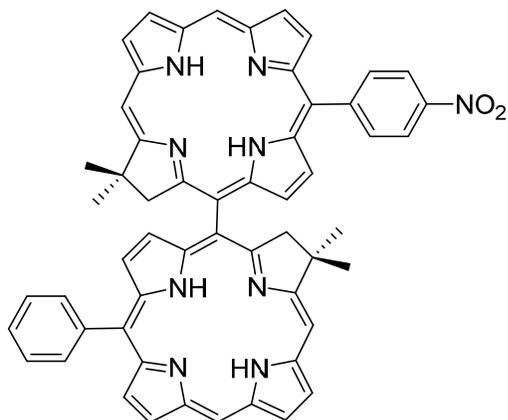
Chl₂15,15-Pd₂. A sample of **Chl₂15,15** (5.2 mg, 0.0077 mmol) in a microwave vial was dissolved in pyridine (1.5 mL), and the solution was treated with Pd(acac)₂ (49.7 mg, 0.163 mmol). The vial was sealed, and the reaction mixture was heated in a microwave reactor at 180 °C for 30 min. The reaction mixture was allowed to cool to room temperature, and the solvent

was evaporated at reduced pressure. The dark residue was dissolved in a small amount of CH_2Cl_2 , and the solution was loaded onto a silica gel chromatography column. Elution with CH_2Cl_2 yielded a dark blue solid (quant): due to the poor solubility of the compound in a range of solvents (CH_2Cl_2 , CHCl_3 , DMSO, THF, MeOH, EtOAc, toluene, acetone) we could not collect a good quality ^1H NMR-spectrum. ESI-MS calcd 443.1 obsd 442.8 $[(\text{M})^{2-}]$, $\text{M} = \text{C}_{44}\text{H}_{34}\text{N}_8\text{Pd}_2$]; $\lambda_{\text{abs}} = 334, 401, 482, 514, 560, 601$ nm (THF); $\lambda_{\text{abs}} = 401, 481, 560, 601$ nm (CH_2Cl_2); steady-state emission: $\lambda_{\text{em}} = 608, 644$ nm ($\lambda_{\text{ex}} = 401$ nm, CH_2Cl_2); time-resolved emission: $\lambda_{\text{phos}} = 776$ nm (77 K, $\lambda_{\text{ex}} = 401$ nm, THF); $\tau = 359$ μs .

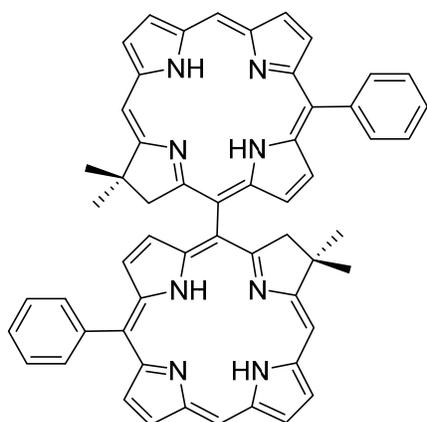


Chl_{215,15}-PhMes, 12.5 mg, 11%; ^1H NMR (50 MHz, CDCl_3) -1.81 (s, 1H), -1.70 (s, 1H), -1.38 (s, 1H), -1.35 (s, 1H), 1.80 (s, 3H), 1.84 (s, 3H), 1.88 (s, 3H), 1.91 (s, 3H), 2.52 (s, 3H), 3.75 – 3.83 (m, 4H), 7.18 (m, 2H), 7.64 – 7.72 (m, 3H), 7.71 (d, $J = 5.0$ Hz, 1H), 7.81 (d, $J = 5.0$ Hz, 1H), 8.12 – 8.13 (m, 1H), 8.17 – 8.19 (m, 1H), 8.35 (d, $J = 4.5$ Hz, 1H), 8.52 (d, $J = 5.0$ Hz, 1H), 8.53 (d, $J = 4.5$ Hz, 1H), 8.69 (d, $J = 4.5$ Hz, 1H), 8.95 – 8.98 (m, 3H), 9.00 – 9.01 (m, 1H), 9.03 (d, $J = 4.5$ Hz, 1H), 9.29 (d, $J = 5.0$ Hz, 1H), 9.31 (d, $J = 45$ Hz, 1H), 9.86 (s, 1H), 9.92 (s, 1H); ^{13}C NMR (125 MHz, CDCl_3 , several peaks multiplied, all signals reported) 21.5 , 21.54 , 21.57 , 22.9 , 29.5 , 29.8 , 29.9 , 30.4 , 30.8 , 30.9 , 31.4 , 31.6 , 31.7 , 31.8 , 32.1 , 46.2 , 52.8 , 52.9 , 94.8 , 94.9 , 106.8 , 107.1 , 110.1 , 111.5 , 121.5 , 123.0 , 123.7 , 123.8 , 124.0 , 126.8 , 127.2 , 127.7 , 128.3 ,

128.4, 128.5, 129.8, 131.7, 132.4, 132.82, 132.85, 134.0, 134.1, 134.5, 135.1, 135.23, 135.25, 137.6, 138.2, 139.18, 139.20, 141.1, 141.2, 142.1, 142.17, 142.21, 152.1, 152.6, 165.1, 165.3, 175.3, 175.5; HR-ESI-MS calcd 873.4388 obsd 873.4384 $[(M + H)^+]$, $M = C_{59}H_{52}N_4$, calcd 895.4207 obsd 895.4199 $(M + Na)^+$; $\lambda_{abs} = 283, 410, 421, 508, 534, 589, 648$ nm (CH_2Cl_2); $\lambda_{em} = 652$ nm ($\lambda_{ex} = 421$ nm, CH_2Cl_2).



Chl_{215,15}-PhNO₂, 6.2 mg, 32%; 1H NMR (500 MHz, $CDCl_3$) -1.92 (s, 1H), -1.80 (s, 1H), -1.46 (s, 1H), -1.34 (s, 1H), 1.80 (s x s, 2 x 3H), 1.85 (s, 3H), 1.86 (s, 3H), 3.79 (d, $J = 1.2$ Hz, 2H), 3.83 (d, $J = 1.2$ Hz, 2H), 7.63 – 7.68 (m, 3H), 7.74 (d, $J = 4.5$ Hz, 1H), 7.84 (d, $J = 5.0$ Hz, 1H), 8.11 – 8.14 (m, 1H), 8.17 – 8.19 (m, 1H), 8.32 (dd, $J_1 = 8.1$ Hz, 1H, $J_2 = 1.7$ Hz, 1H), 8.38 (dd, $J_1 = 8.7$ Hz, $J_2 = 1.7$ Hz, 1H), 8.43 (d, $J = 4.6$ Hz, 1H), 8.52 – 8.57 (m, 3H), 8.59 (d, $J = 4.1$ Hz, 1H), 8.69 (d, $J = 4.5$ Hz, 1H), 8.98 (s, 1H), 9.02 – 9.05 (m, 3H), 9.07 – 9.08 (m, 2H), 9.32 (d, $J = 4.5$ Hz, 1H), 9.35 (d, $J = 4.5$ Hz, 1H), 9.92 (s, 1H), 9.97 (s, 1H); ESI-MS calcd 875.4 obsd 876.3 $[(M + H)^+]$, $M = C_{56}H_{45}N_9O_2$; $\lambda_{abs} = 280, 420, 508, 534, 591, 650$ nm (CH_2Cl_2); $\lambda_{em} = 654$ nm ($\lambda_{ex} = 420$ nm, CH_2Cl_2).



Chl_{215,15}-Ph₂, 17.5 mg, 16%; ¹H NMR (500 MHz, CDCl₃) -1.79 (s, 2H), -1.34 (s, 2H), 1.80 (s, 6H), 1.86 (s, 6H), 3.80 (s, 2H), 3.81 (s, 2H), 7.64–7.68 (m, 6H), 7.79 (d, *J* = 5.0 Hz, 2H), 8.14 (d, *J* = 5.0 Hz, 2H), 8.19–8.20 (m, 2H), 8.53 (d, *J* = 5.0 Hz, 2H), 8.70 (d, *J* = 4.5 Hz, 2H), 8.99 (s, 2H), 9.03–9.04 (m, 4H), 9.32 (d, *J* = 4.5 Hz, 2H), 9.93 (s, 2H); ¹³C NMR (125 MHz, CDCl₃) 29.7, 31.6, 46.1, 52.7, 97.7, 107.0, 111.6, 122.9, 123.5, 123.9, 126.6, 126.7, 127.6, 128.2, 128.4, 132.2, 132.7, 133.86, 133.93, 135.0, 141.1, 141.9, 142.0, 152.0, 152.5, 165.1, 175.3; HR-ESI-MS obsd 831.3899 [(M + H)⁺, M = C₅₆H₄₆N₈]; λ_{abs} = 281, 411, 421, 507, 533, 590, 649 nm (CH₂Cl₂); λ_{em} = 653 nm (λ_{ex} = 421 nm, CH₂Cl₂).

Table S1. Photophysical properties of dimers and reference compounds.^a

Compound	$\lambda_{\text{Soret}}, \lambda_{\text{Q}}$	$I_{\text{B}}/I_{\text{Q}}$	$\lambda_{\text{em}} (\lambda_{\text{exc}})$	Φ^{c}
Chl₂3,15	391, 413, 642	2.1	646	0.19
Chl₂13,15	392, 413, 642	2.0	645	0.16
Chl₂15,15	413, 645	2.6	648	0.255
Chl₂3,15-Zn₂	401, 615	2.6	619	0.11
Chl₂15^{Ph},15^{Mes}	410, 421, 648	2.6	652	0.39
Chl₂15^{Ph},15^{PhNO₂}	420, 650	2.9	654	n.d.
Chl₂15^{Ph},15^{Ph}	411, 421, 649	2.9	653	0.28
Chl	398, 636 ^d	2.4 ^d	636 ^d	0.19 ^d
Chl15^{Bpin}	393, 638	4.3	643	0.21
Chl10^{Ph}15^{Bpin}	404, 641	5.6	646	n.d.
Chl15^{Ph}-Pd^b	391, 587	1.9	n.d.	n.d.
Chl₂15,15-Pd₂^b	401, 601	1.5	n.d.	n.d.

^a In CH₂Cl₂. ^b In THF. ^c Using *meso*-tetraphenylporphyrin in toluene as standard. ^d From reference 7, measured in toluene.

VT-NMR experiments

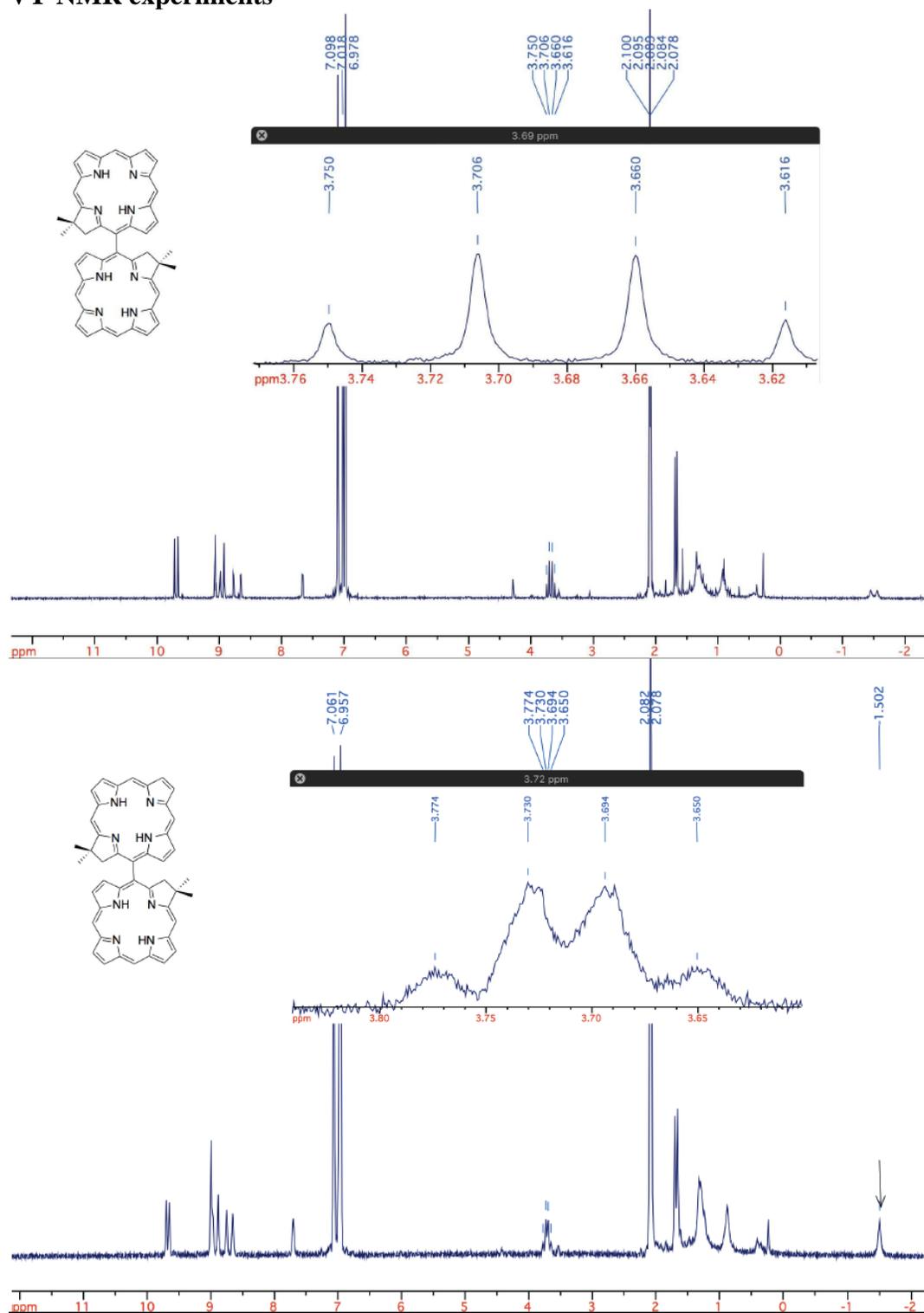


Figure S1. Spectra of Chl₂13,15 at room temperature (top) and at 90 °C (bottom) in toluene-*d*₈, with the CH₂ ABq-signal shown separately.

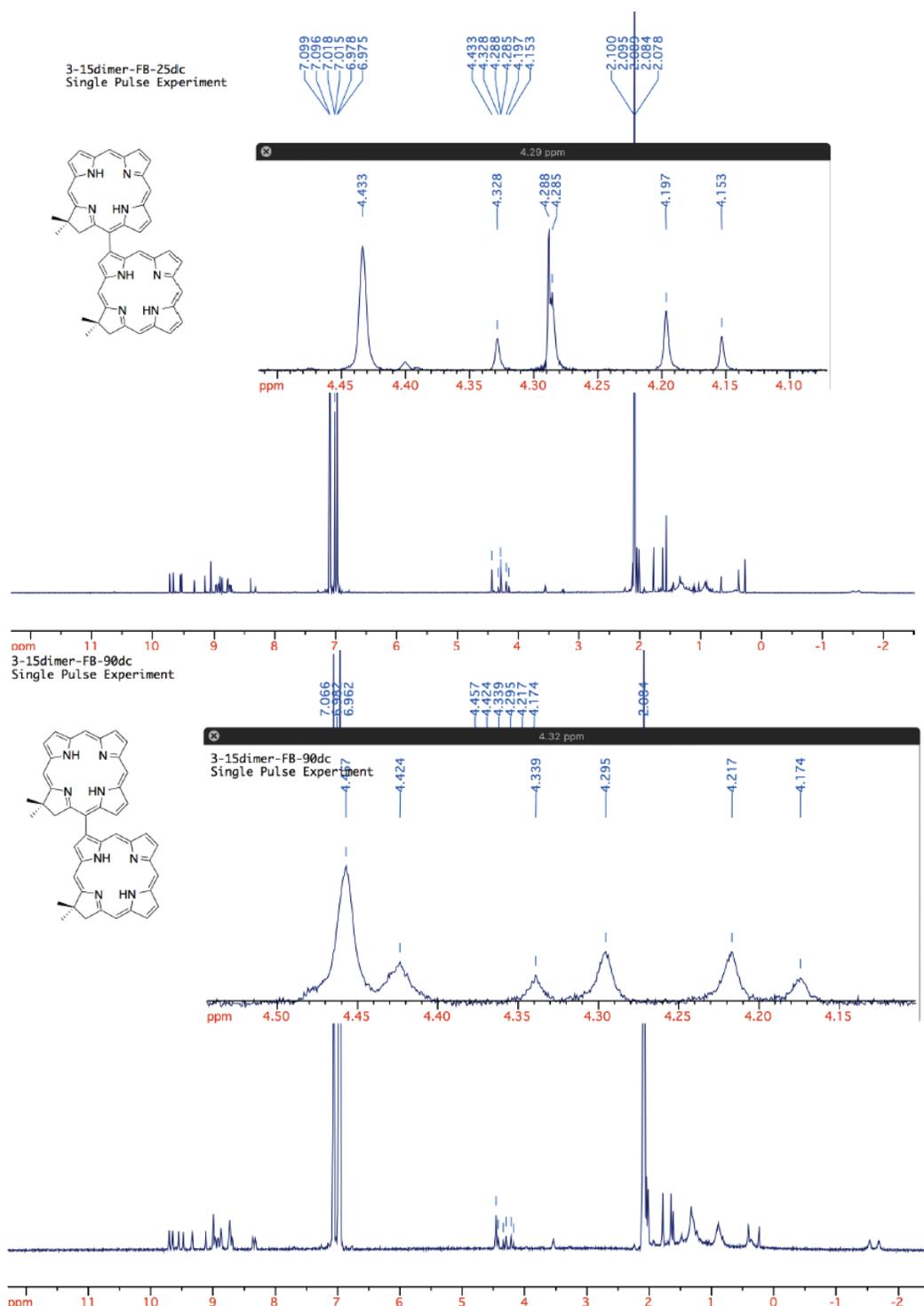


Figure S2. Partial spectra of **Chl₂15,15** at room temperature (top) and at 90 °C (bottom) in toluene-*d*₈, with the CH₂ ABQ-signal shown separately.

Phosphorescence spectra of Pd-complexes

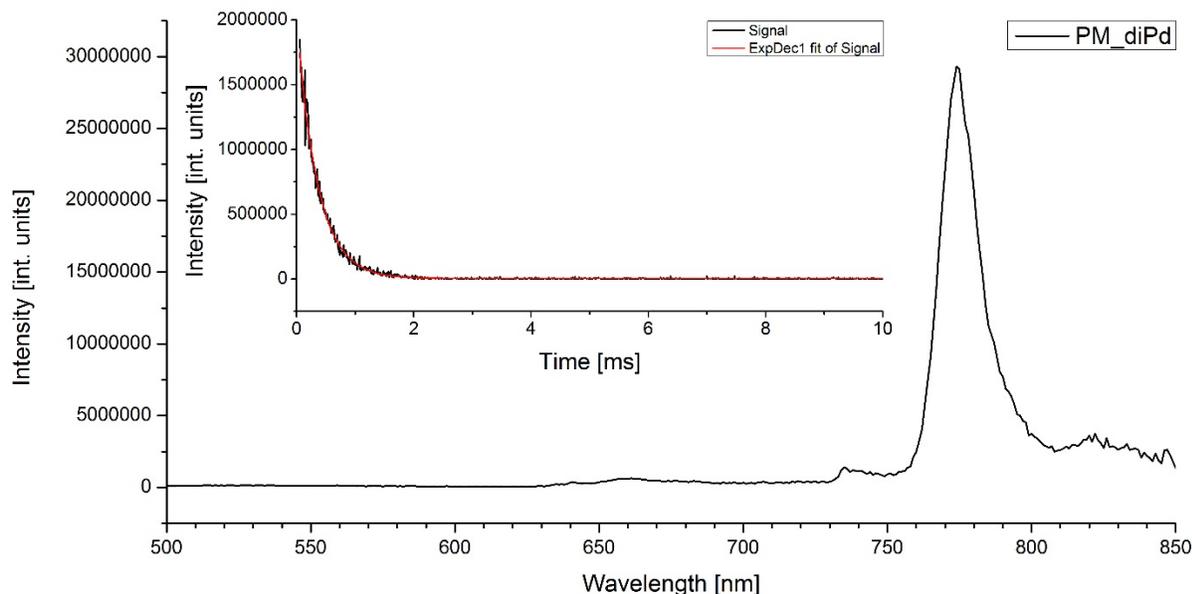


Figure S3. Time-resolved emission spectrum of **Chl₂15,15-Pd₂** at 77 K in THF ($\lambda_{\text{exc}} = 401$ nm, delay: 50 μs , integration time 10^5 μs). The inset shows the emission decay ($\lambda_{\text{exc}} = 401$ nm, $\lambda_{\text{em}} = 774$ nm, sample window 0.2 ms, integration time 100 ms) fitted with a monoexponential. Fitting with a biexponential did not improve the fit as judged by R^2 and χ , but yielded two lifetimes, 340 μs and 732 μs with 96% and 4% contribution, respectively.

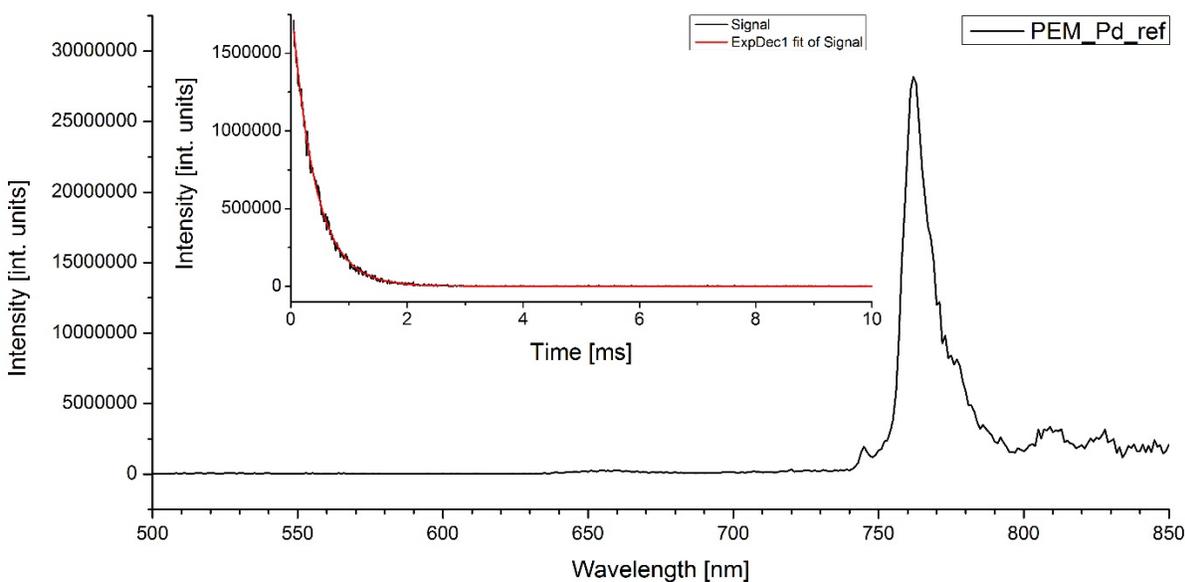


Figure S4. Time-resolved emission spectrum of **Chl₂15^{Ph}-Pd** at 77 K in THF ($\lambda_{\text{exc}} = 391$ nm, delay: 50 μs , integration time 10^5 μs). The inset shows the emission decay ($\lambda_{\text{exc}} = 391$ nm, $\lambda_{\text{em}} = 762$ nm, sample window 0.2 ms, integration time 100 ms) fitted with a monoexponential.

Electrochemistry

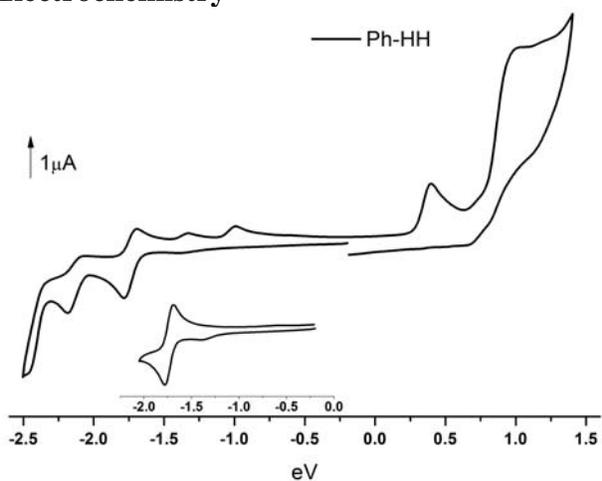


Figure S5. Measured for 1 mM solution of **Chl15^{Ph}** in CH₂Cl₂ (0.1 M NBu₄PF₆), glassy C-electrode, $\nu = 100$ mV/s. All potentials are given versus Fc⁺⁰.

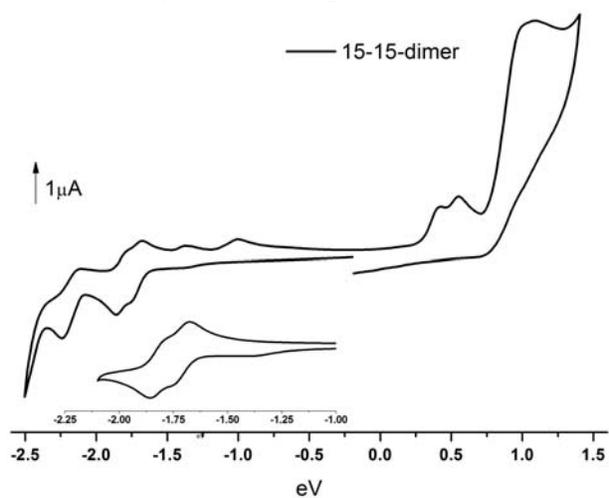


Figure S6. Measured for 1 mM solution of **Chl₂15,15** in CH₂Cl₂ (0.1 M NBu₄PF₆), glassy C-electrode, $\nu = 100$ mV/s. All potentials are given versus Fc⁺⁰.

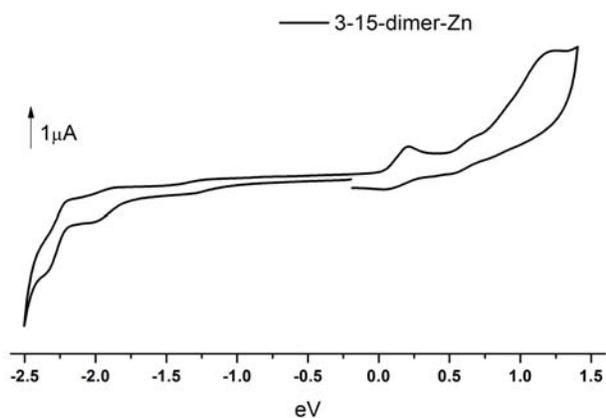


Figure S7. Measured for 1 mM solution of **Chl₂3,15-Zn₂** in CH₂Cl₂ (0.1 M NBu₄PF₆), glassy C-electrode, $\nu = 100$ mV/s. All potentials are given versus Fc⁺⁰.

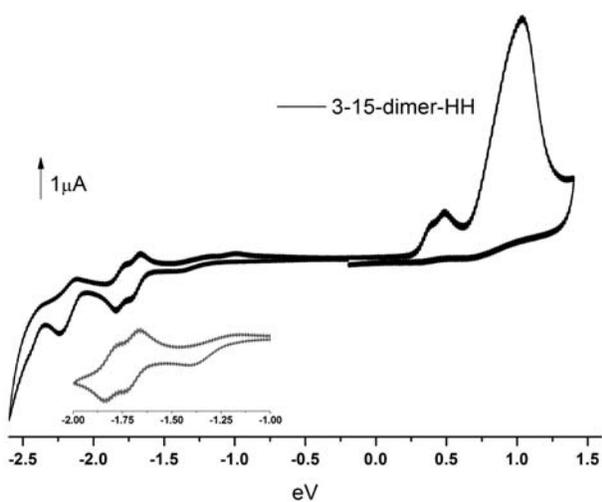


Figure S8. Measured for 1 mM solution of **Chl₂3,15** in CH₂Cl₂ (0.1 M NBu₄PF₆), glassy C-electrode, $\nu = 100$ mV/s. All potentials are given versus Fc⁺⁰.

X-ray diffraction data

All the measurements were performed using graphite-monochromatized Mo K_{α} radiation at 100K using a Bruker D8 APEX-II equipped with a CCD camera. The structure was solved by direct methods (SHELXS-2014) and refined by full-matrix least-squares techniques against F^2 (SHELXL-2014).⁸ The non-hydrogen atoms were refined with anisotropic displacement parameters. The H atoms of the CH₂ groups were refined with common isotropic displacement parameters for the H atoms of the same group and idealized geometry with approximately tetrahedral angles and C-H distances of 0.99Å.

Compound **Ch15^{Br}20^{Br}** crystallizes in the monoclinic space group C 2/c (No. 15) as dark brown blocks suitable for single crystal X-ray diffraction. The bond distances of **Ch15^{Br}20^{Br}** heterocycle are in agreement with related chlorin derivatives. The least squares planes (l.s.pl.) through the pyrrole units show significant twisting, i.e. torsion angles of adjacent range from 6.60(10)° to 12.74(11)°.

Table 2. Crystal data and structure refinement for **Ch115^{Br}20^{Br}**.

Crystal data	KEB36 095 F1
CCDC-No.	CCDC 1447589
Empirical formula	C ₂₈ H ₂₂ Br ₂ N ₄
Formula weight	574.31
Crystal description	block, dark brown
Crystal size	0.3 x 0.2 x 0.18
Crystal system, space group	monoclinic, C 2/c
Unit cell dimensions:	
a	26.108(2)
b	8.7687(7)
c	21.6124(18)
β	112.199(2)
Volume	4581.1(7)
Z	8
Calculated density	1.665 Mg/m ³
F(000)	2304
Linear absorption coefficient μ	3.565 mm ⁻¹
Absorption correction	multi-scan, SADABS 2008
Max. and min. transmission	0.7456 and 0.4920
Unit cell determination	2.0 < Θ < 25.2°
	4749 reflections used at 100K
Data collection	
Temperature	100(2)K
Diffractometer	Bruker APEX-II CCD
Radiation source	fine-focus sealed tube
Radiation and wavelength	MoK α , 0.71073Å
Monochromator	Graphite
Scan type	ω scans
Θ range for data collection	2.03 to 28.09°
Index ranges	-34 \leq h \leq 34, -11 \leq k \leq 11, -28 \leq l \leq 28
Reflections collected / unique	45290 / 5531
Significant unique reflections	4749 with I > 2 σ (I)
R(int), R(sigma)	0.0271, 0.0494
Completeness to Θ_{\max}	99.0%
Refinement	
Refinement method	Full-matrix least-squares on F ²
Data / parameters / restraints	5531/ 317/ 0
Goodness-of-fit on F ²	1.049
Final R indices [I > 2 σ (I)]	R1 = 0.0240, wR2 = 0.0538
R indices (all data)	R1 = 0.0321, wR2 = 0.0567
Weighting scheme	w=1/[$\sigma^2(F_o^2)+(aP)^2+bP$] where P=(F _o ² +2F _c ²)/3
Weighting scheme parameters a, b	0.0212, 5.9199
Largest Δ/σ in last cycle	0.003
Largest difference peak and hole	0.378 and -0.375 e/Å ³
Structure Solution Program	SHELXS-2014/7 (Sheldrick, 2008)
Structure Refinement Program	SHELXL-2014 (Sheldrick, 2008)

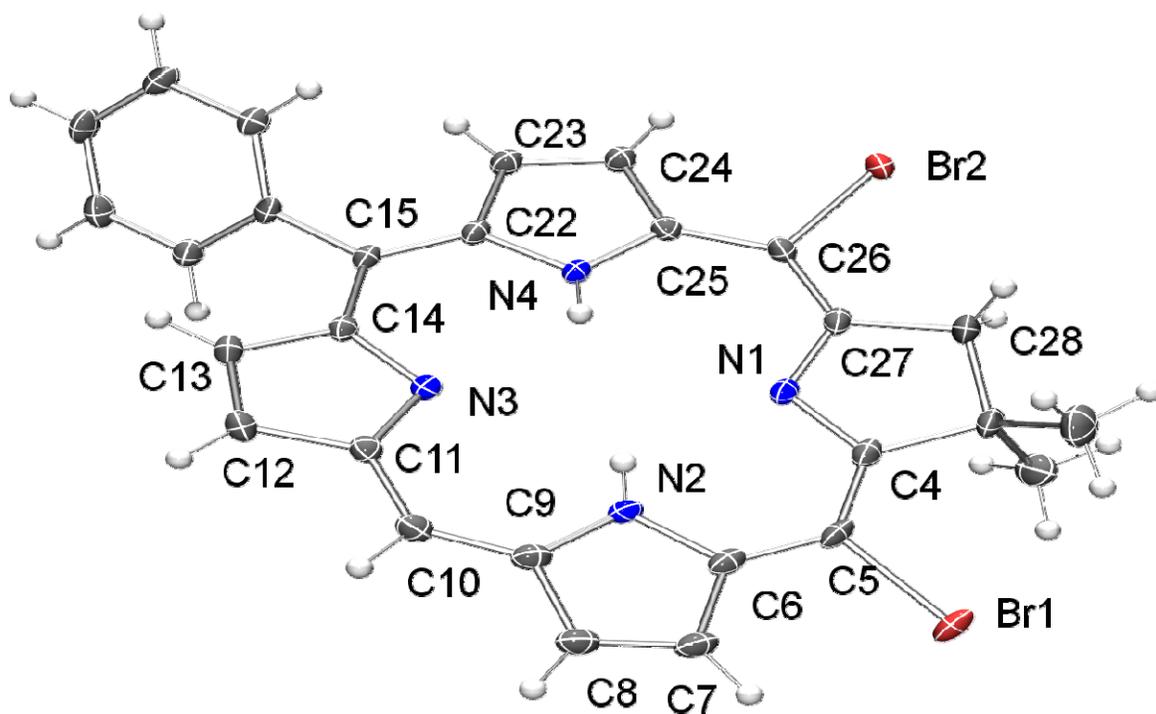
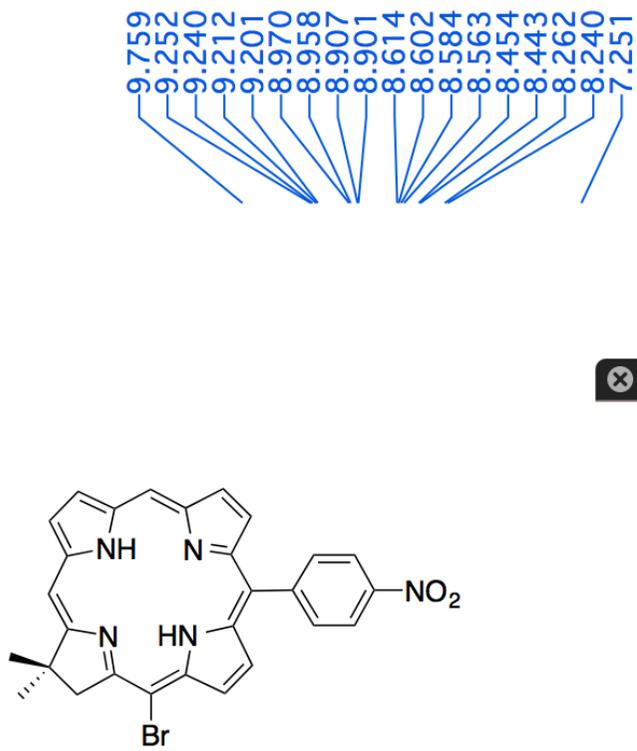


Figure S9: ORTEP representation of **Chl15^{Br}20^{Br}** with thermal ellipsoids at a probability level of 50%. Selected parameters (angles [°]): Angle between least squares planes of the pyrrole heterocycles: Plane N1 – Plane N2 12.74(11), Plane N2 –Plane N3 6.60(10), Plane N3 Plane N4 10.07(10), Plane N4 - Plane N1 12.59(10).

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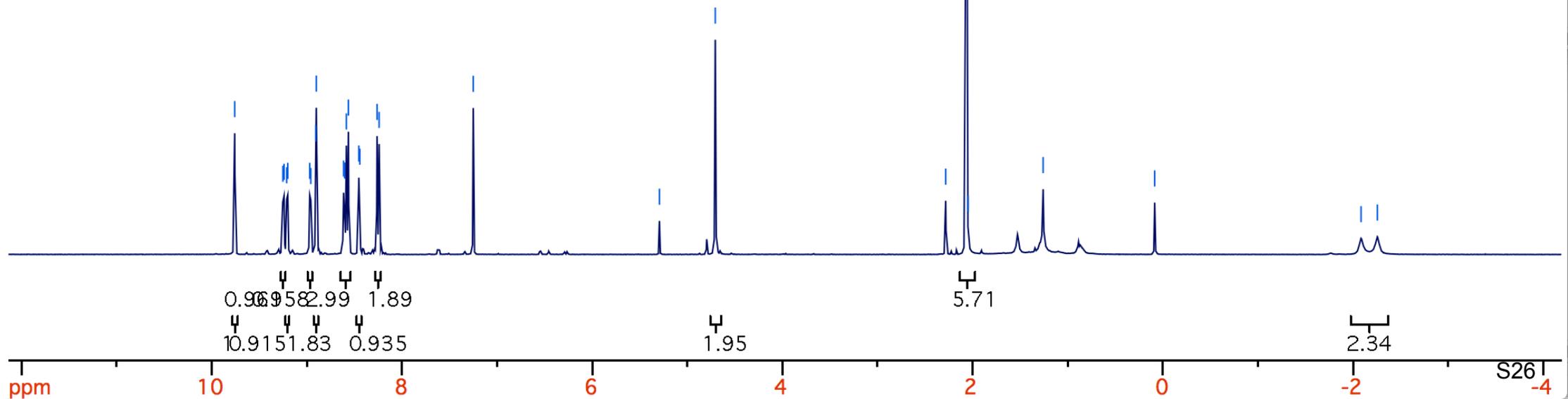
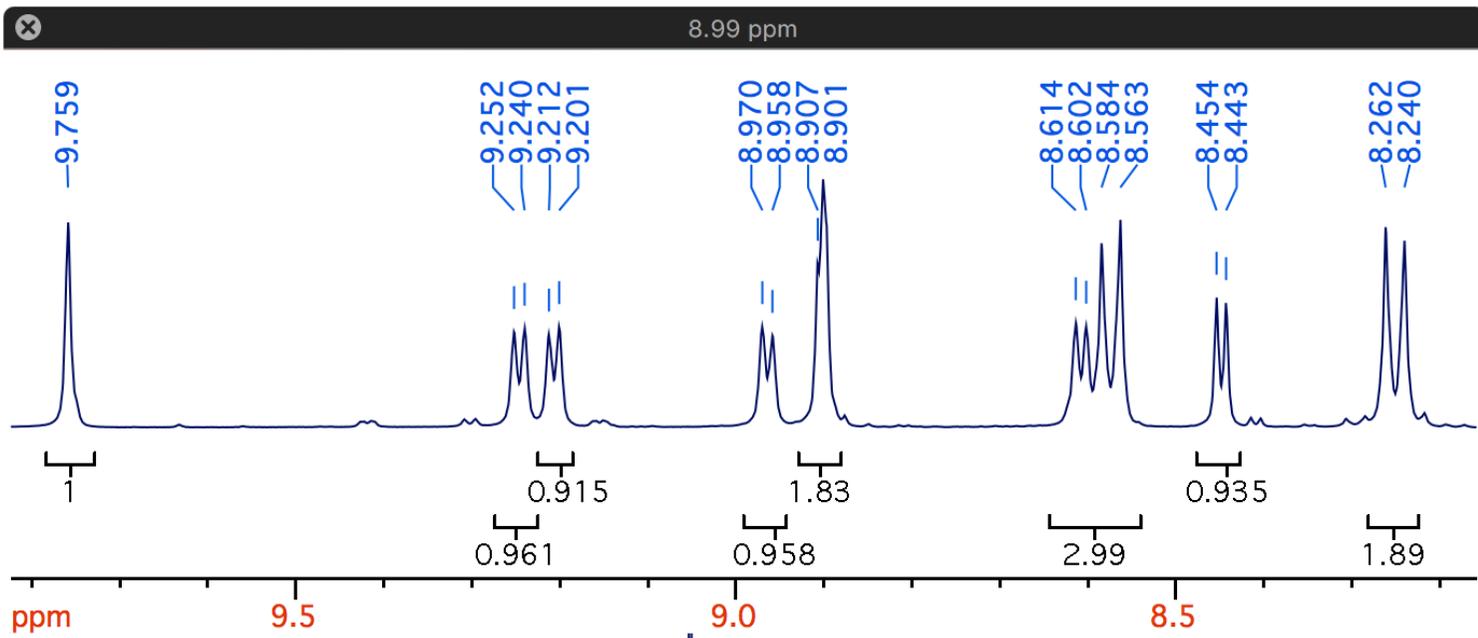
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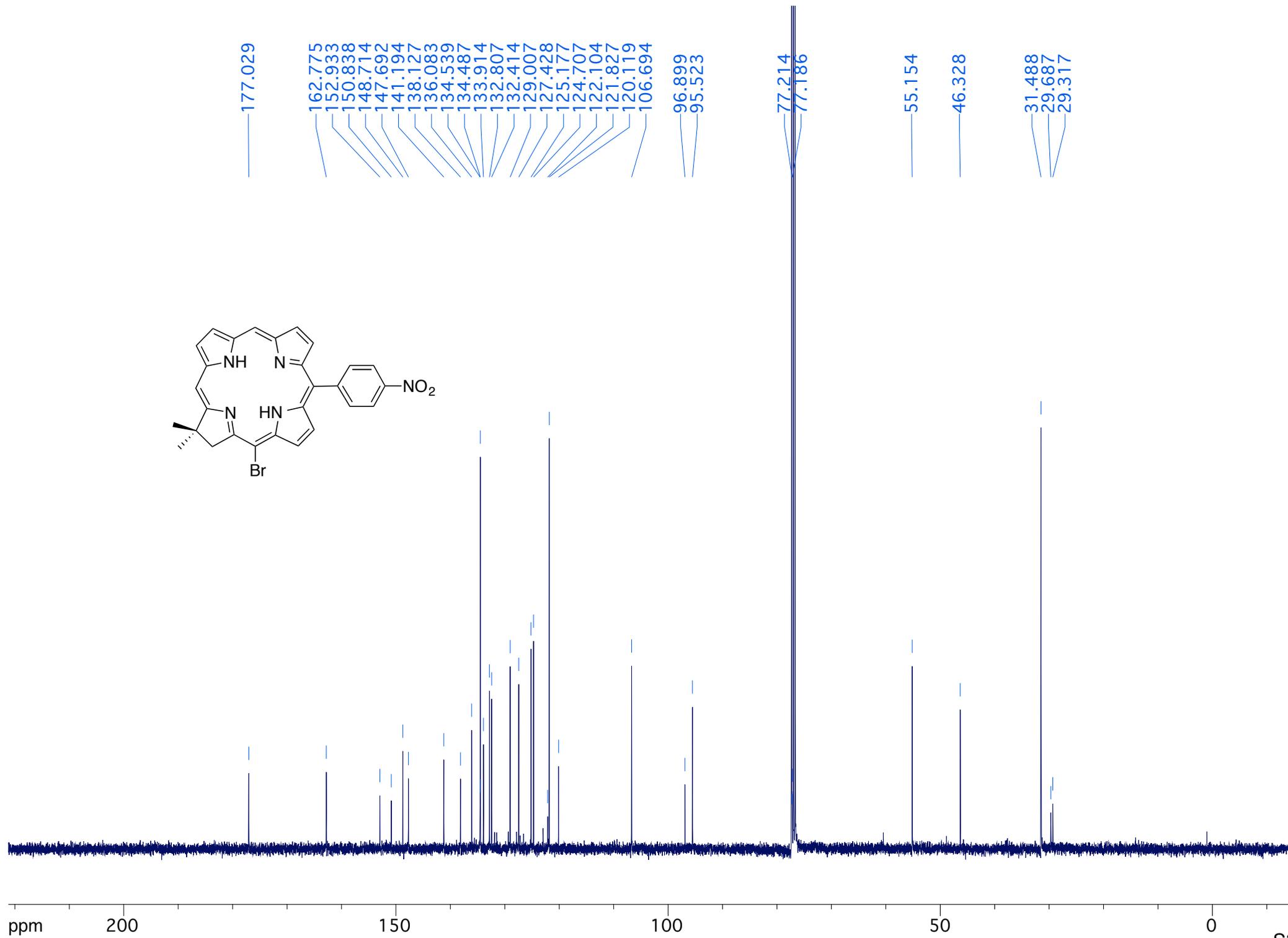
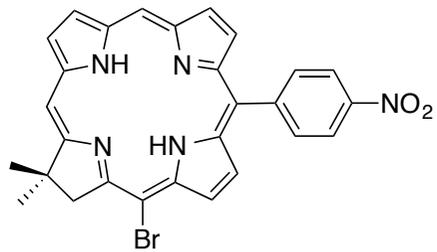
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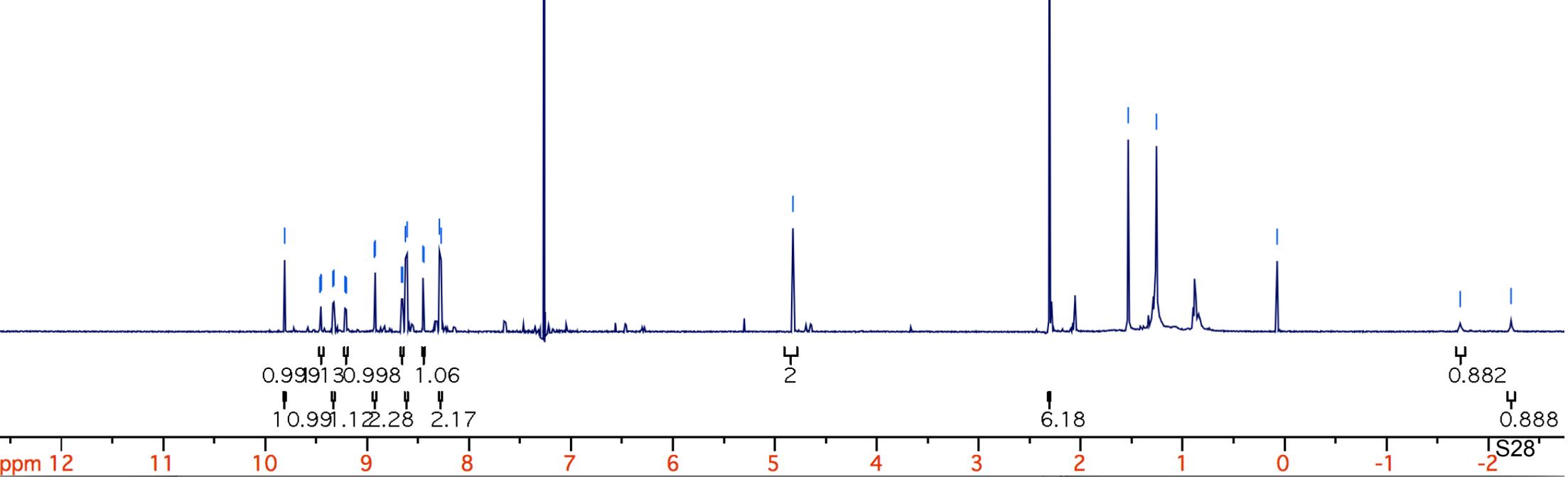
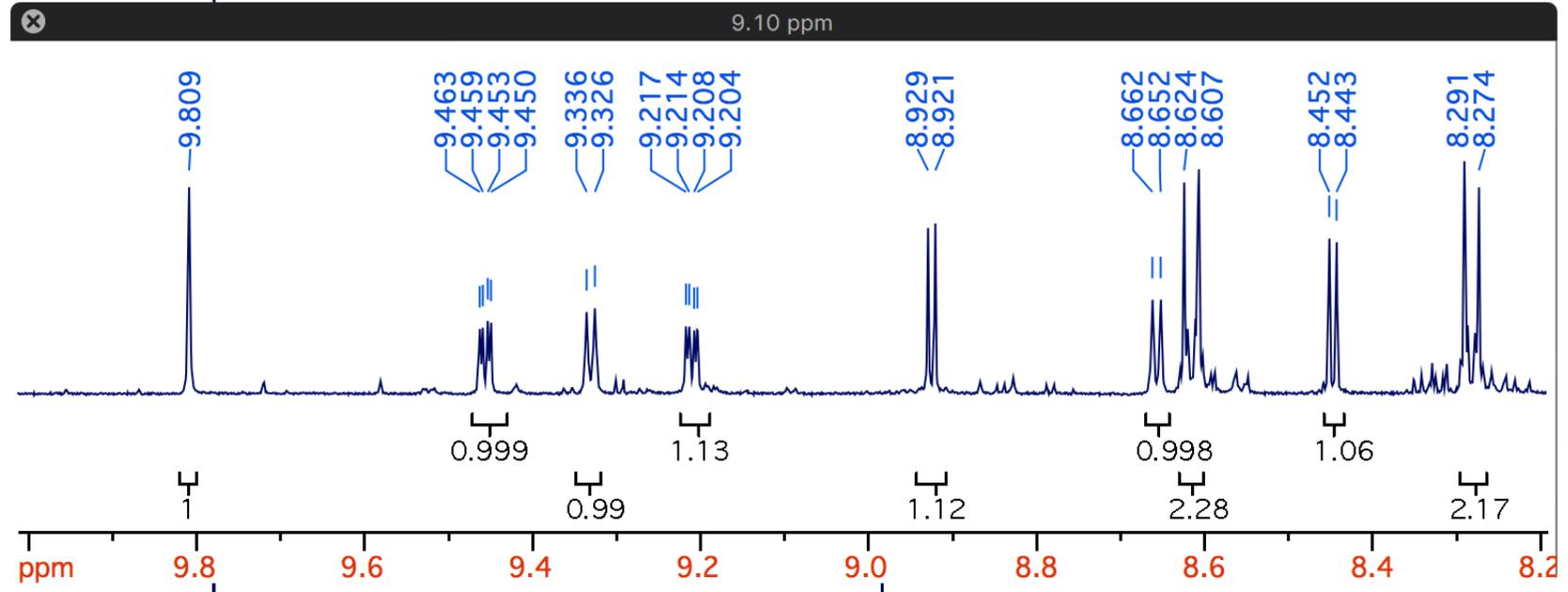
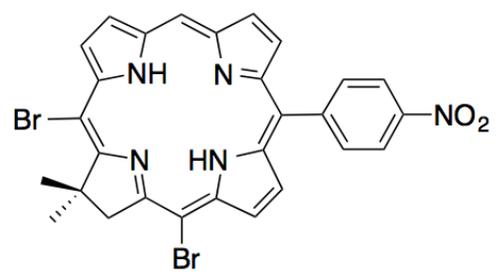
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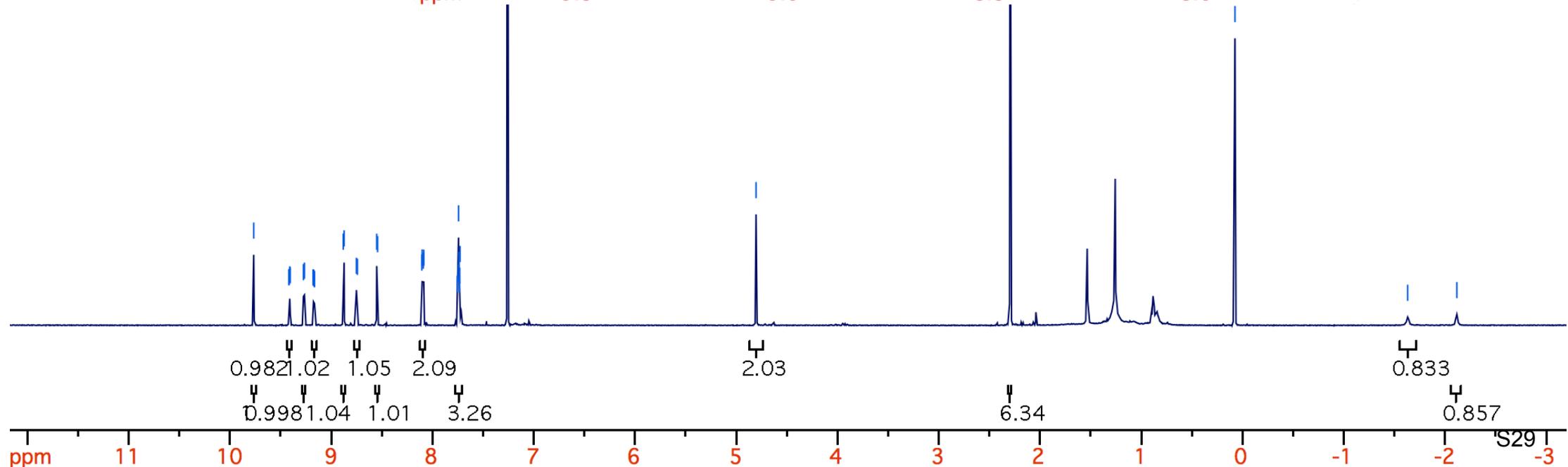
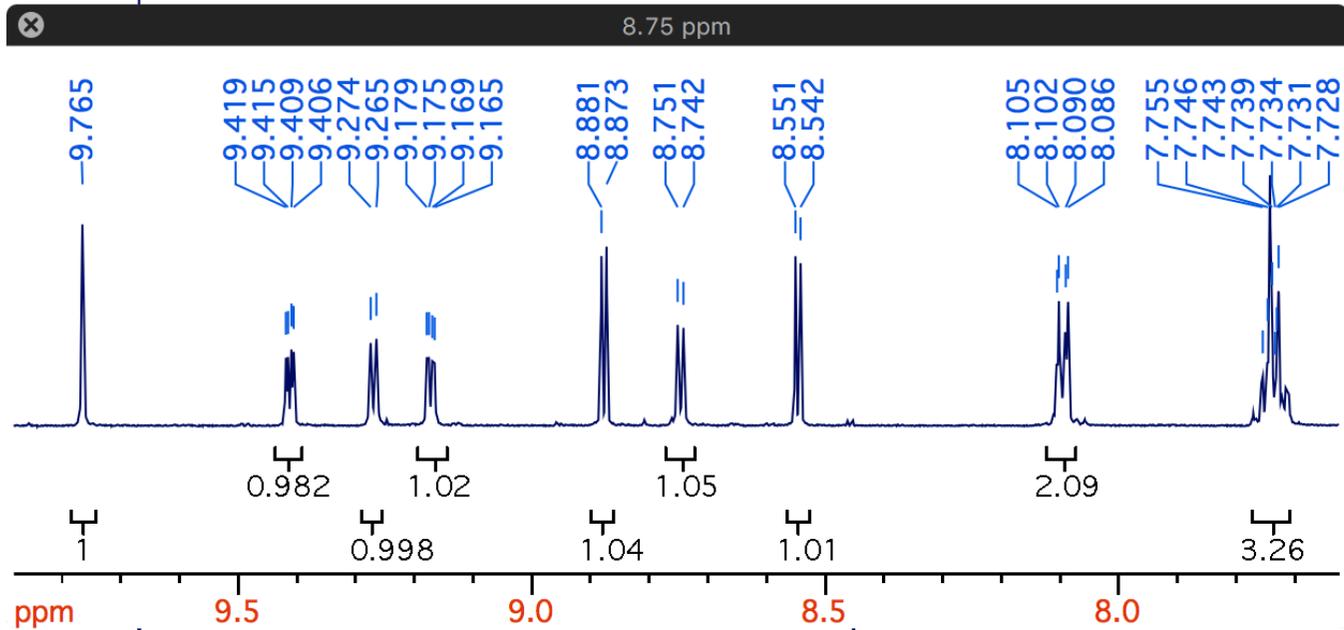
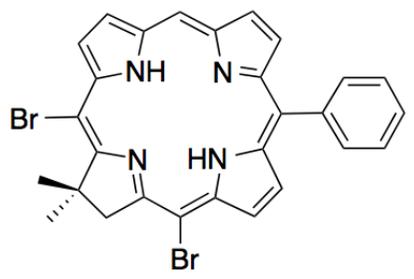
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2.117



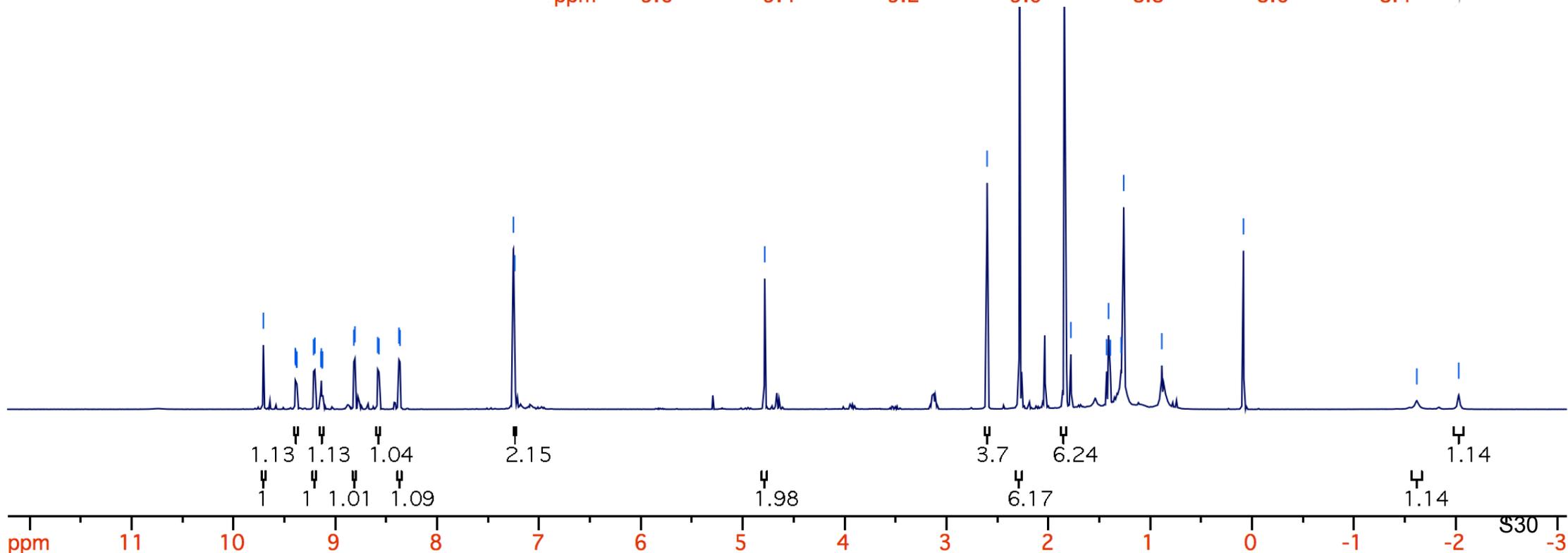
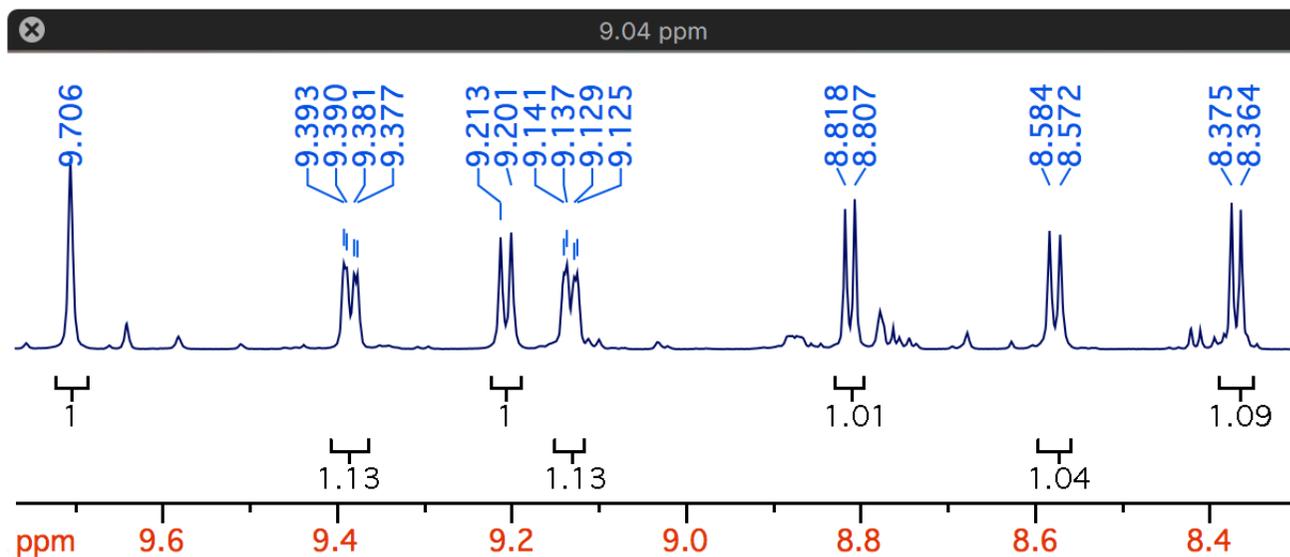
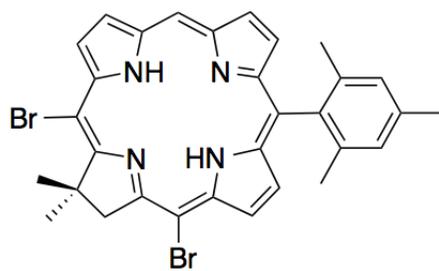
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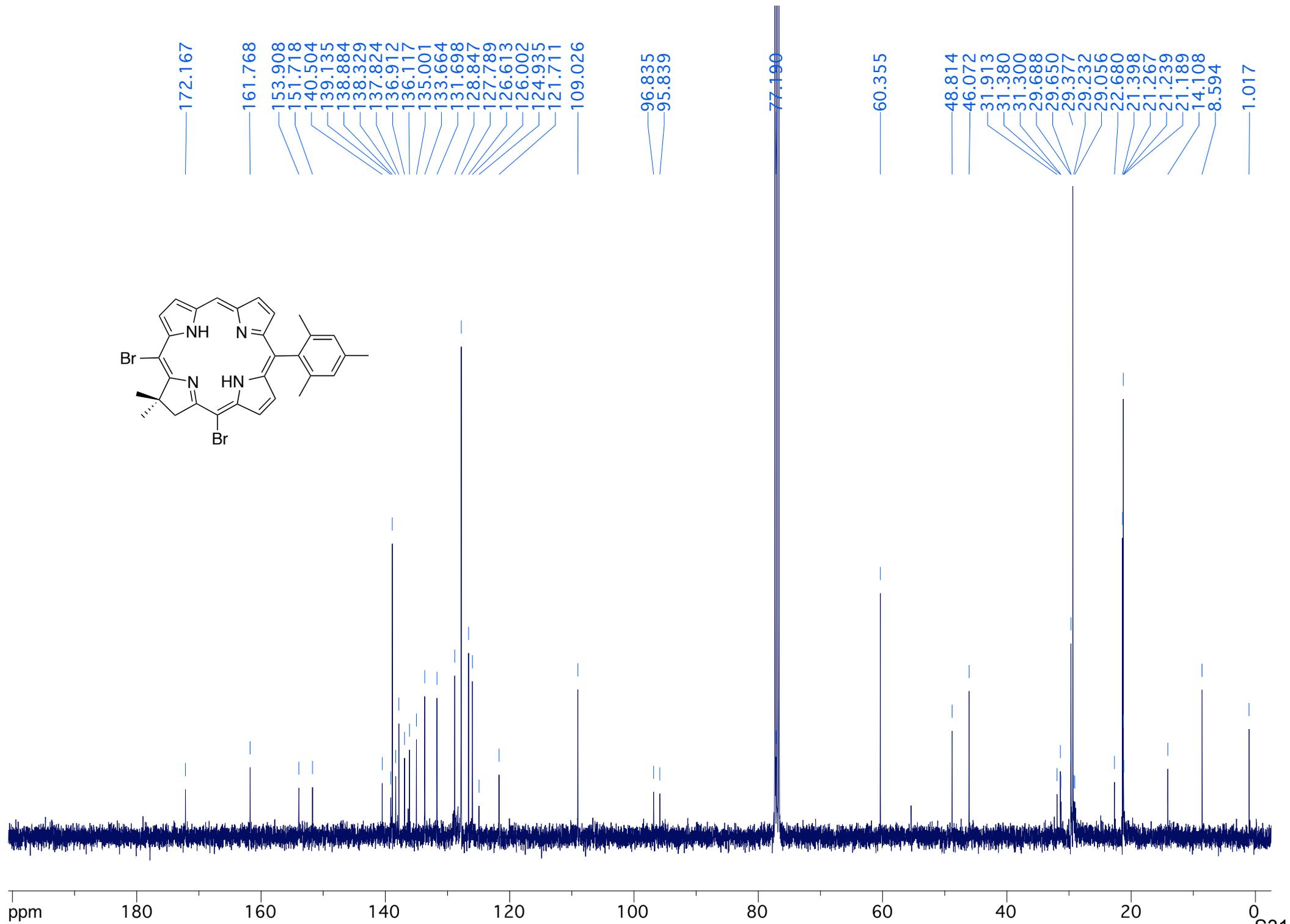
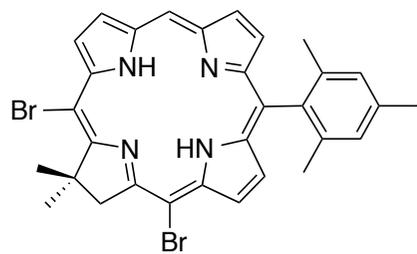
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0.083

1.618
2.030





9.836
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9.775
9.572
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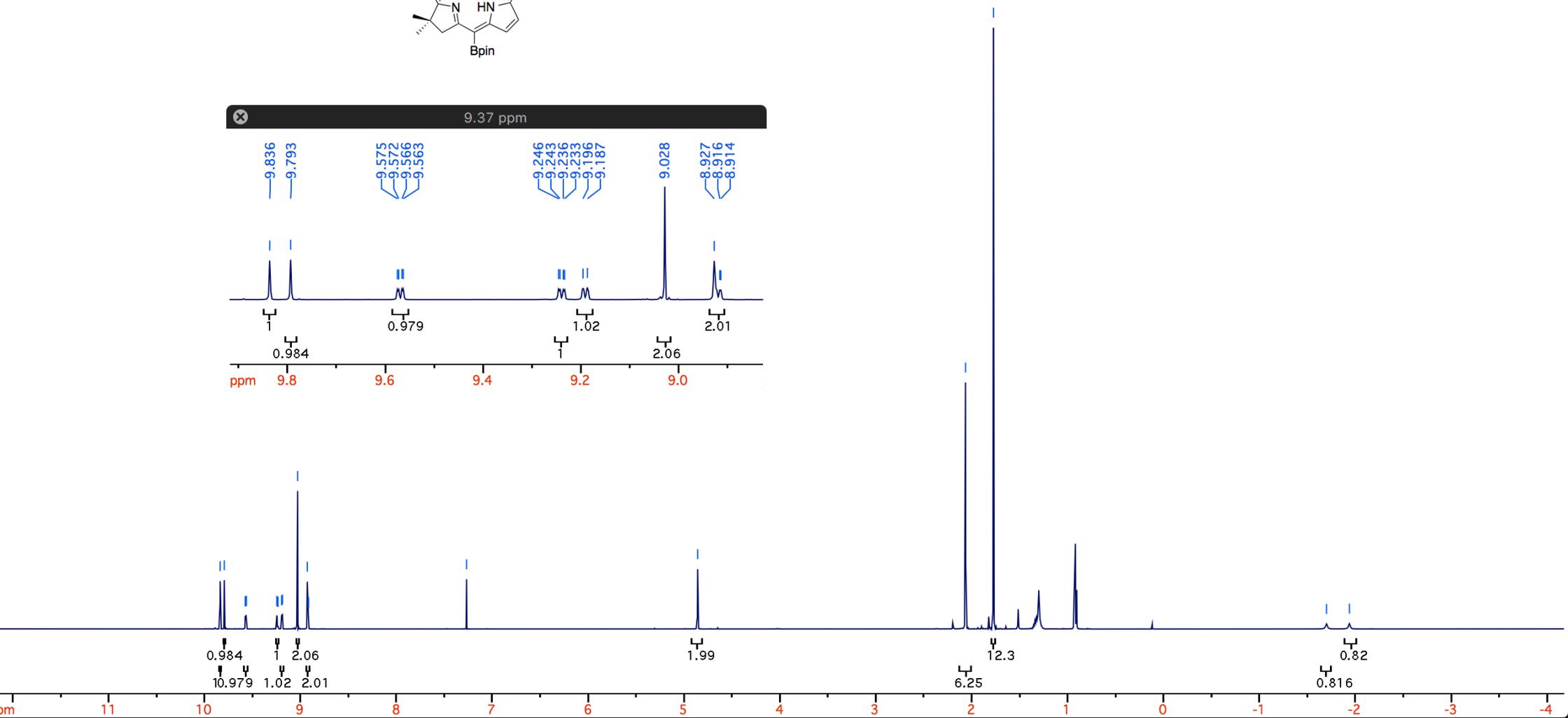
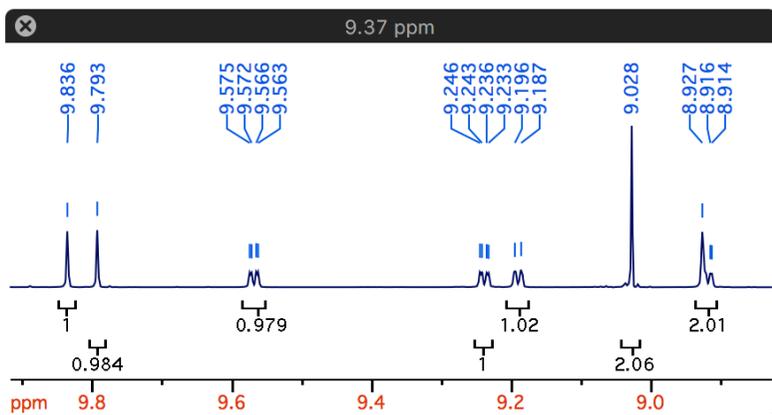
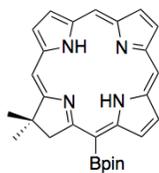
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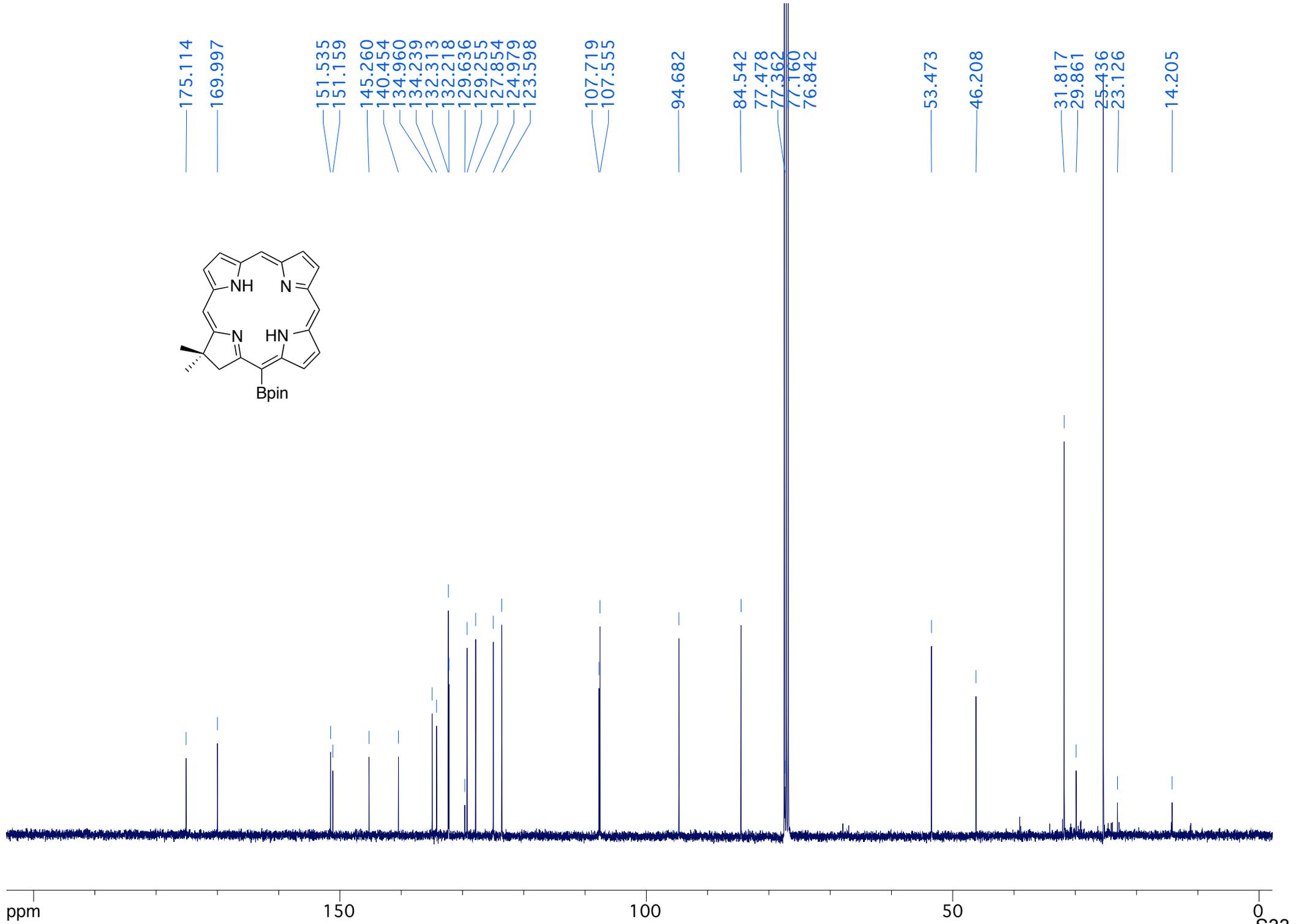
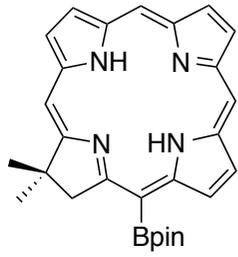
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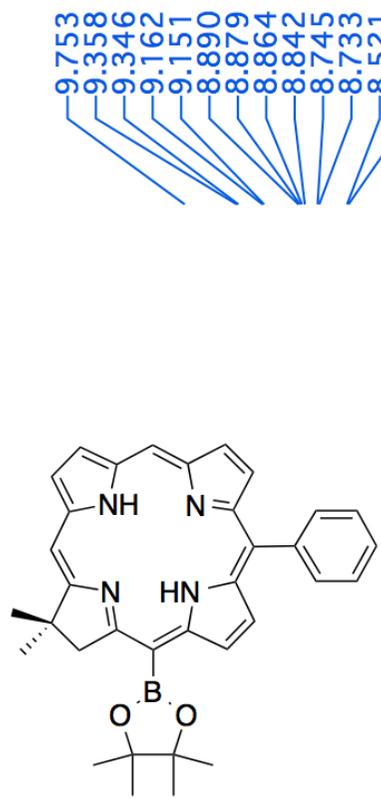
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1.700

1.939







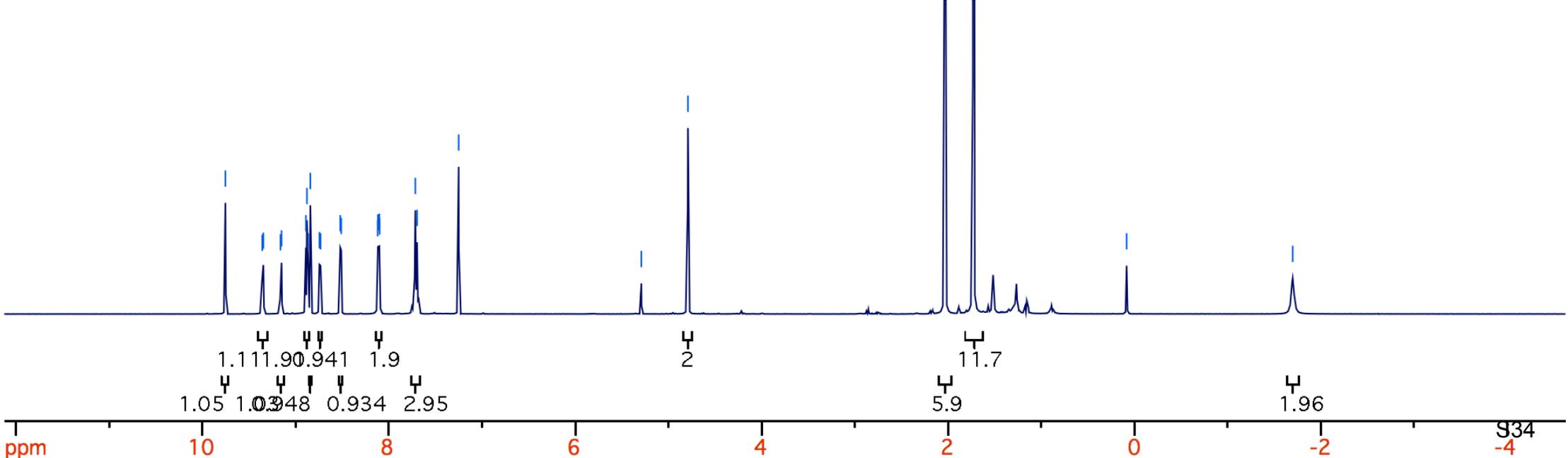
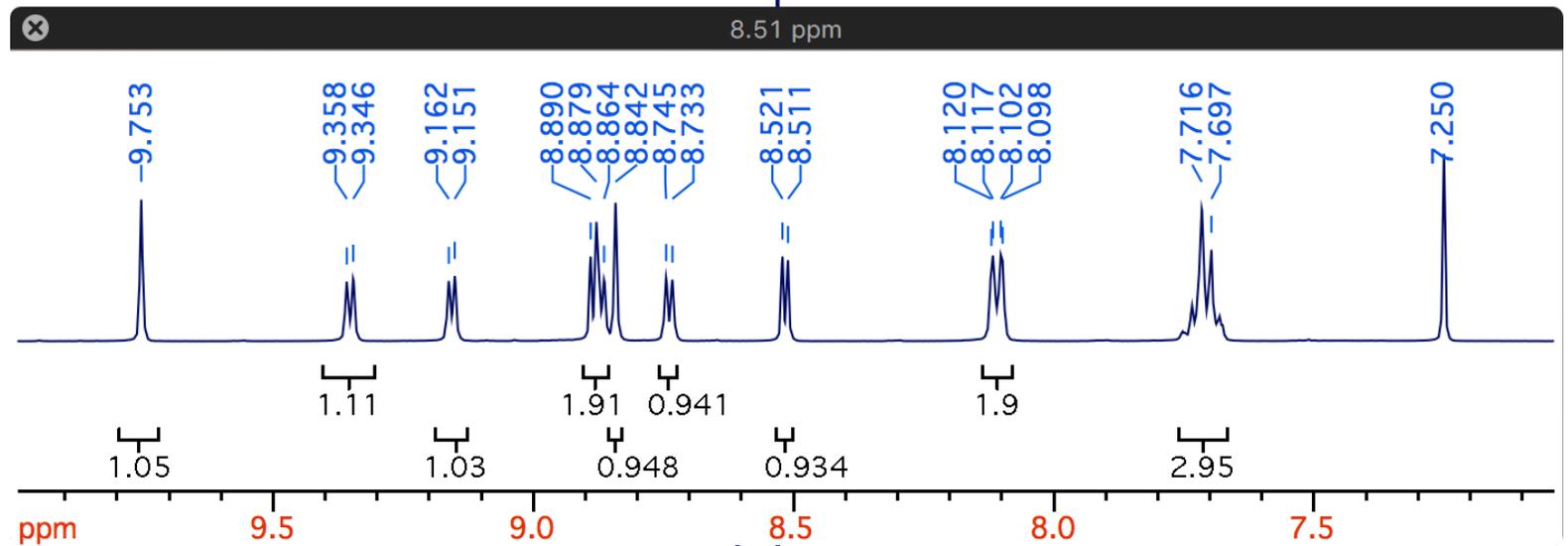
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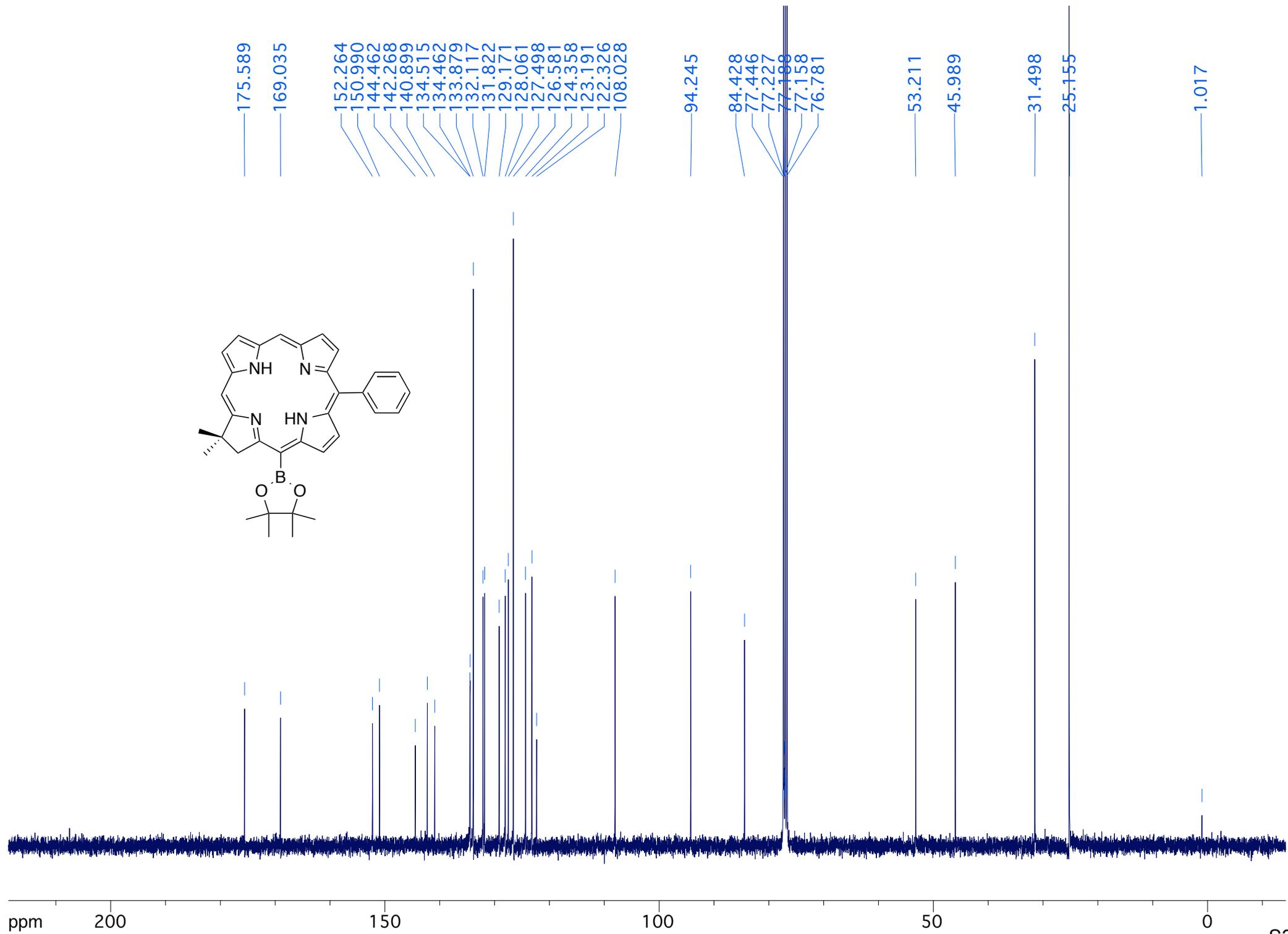
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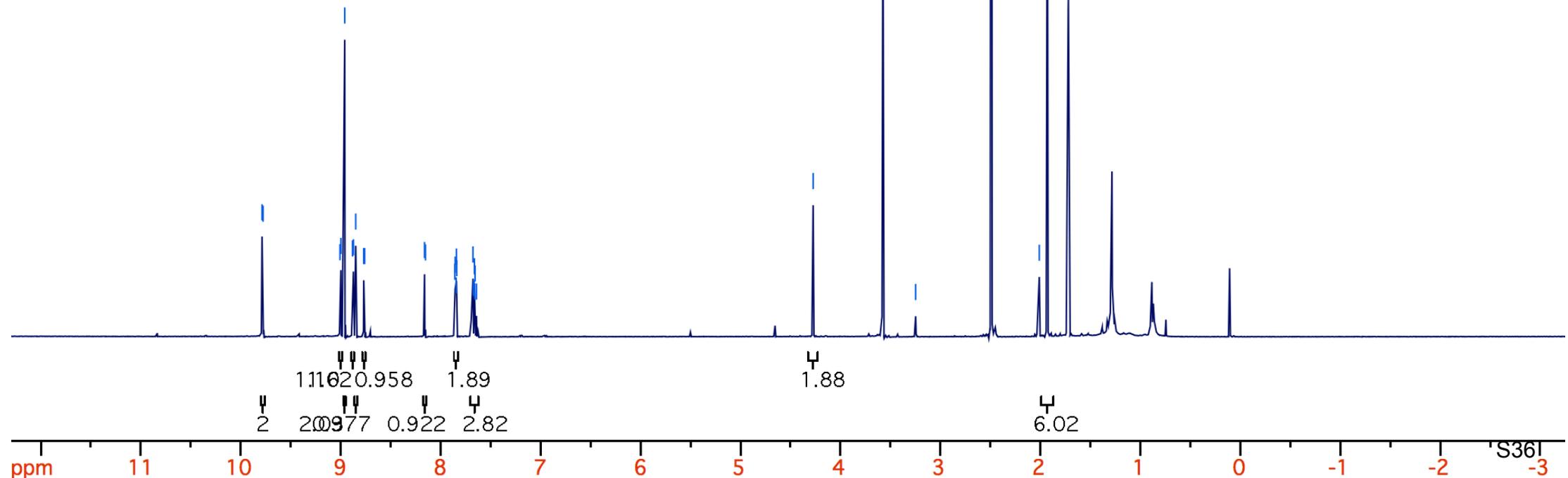
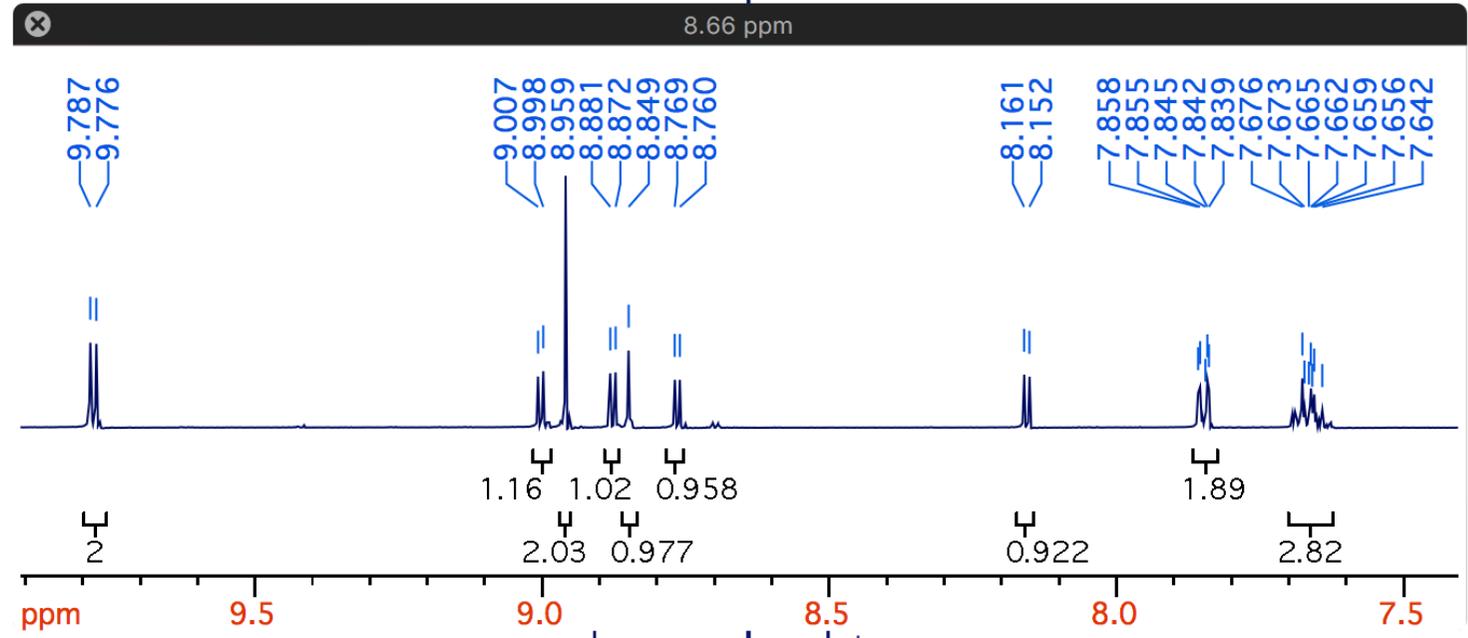
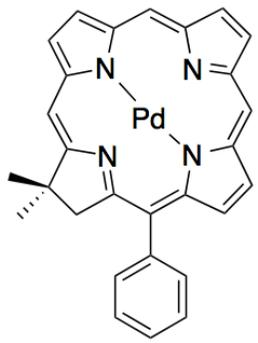
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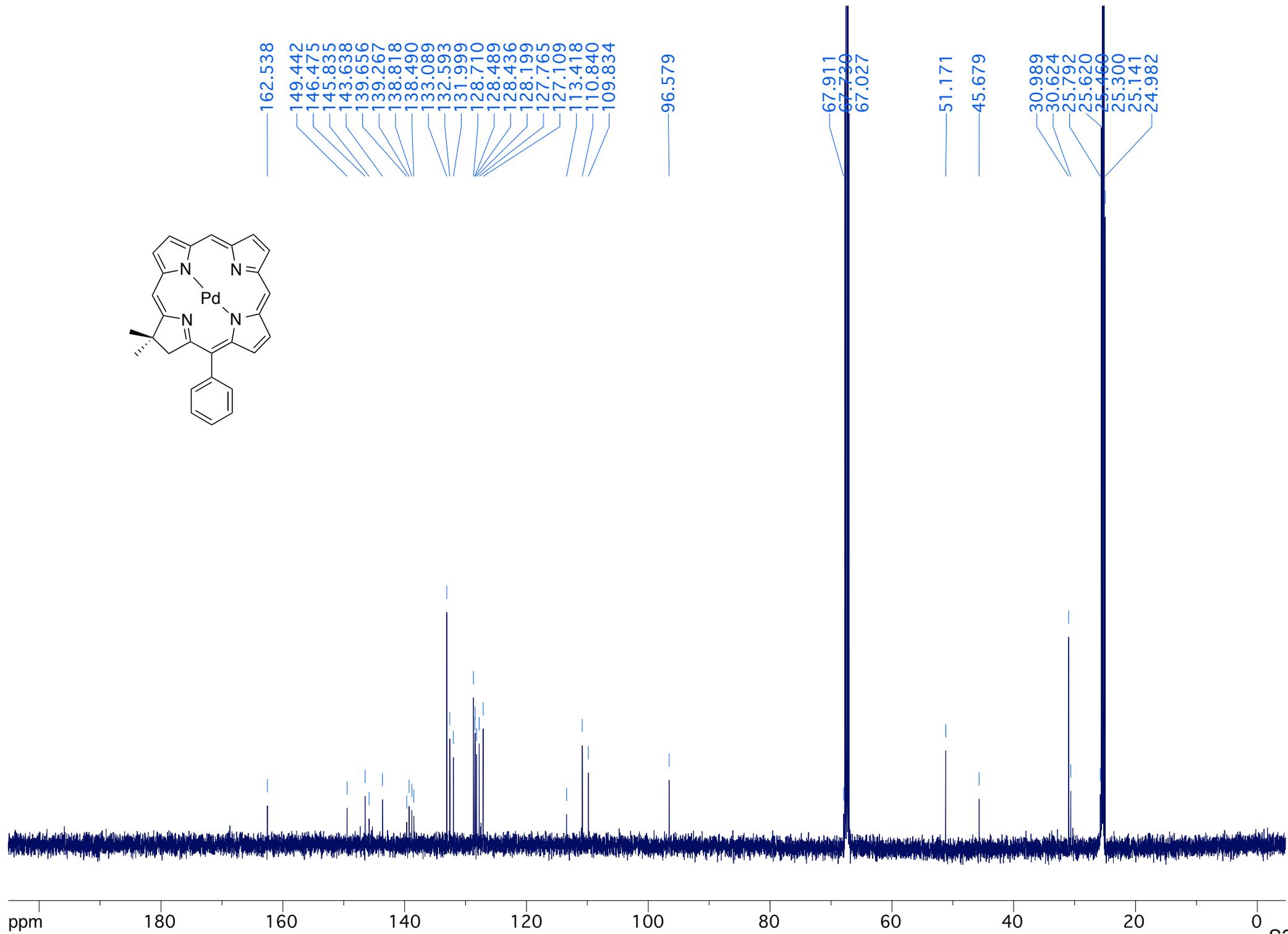
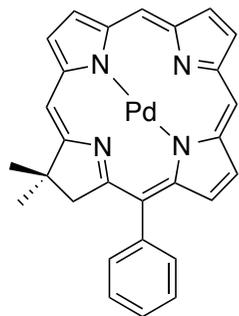




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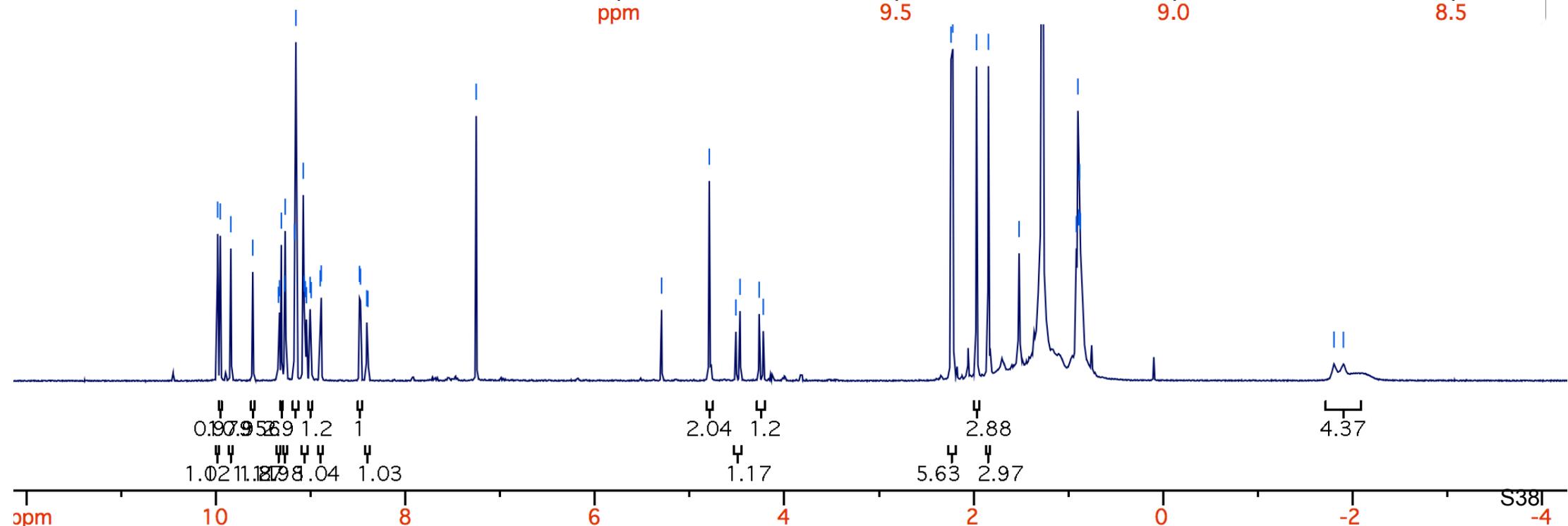
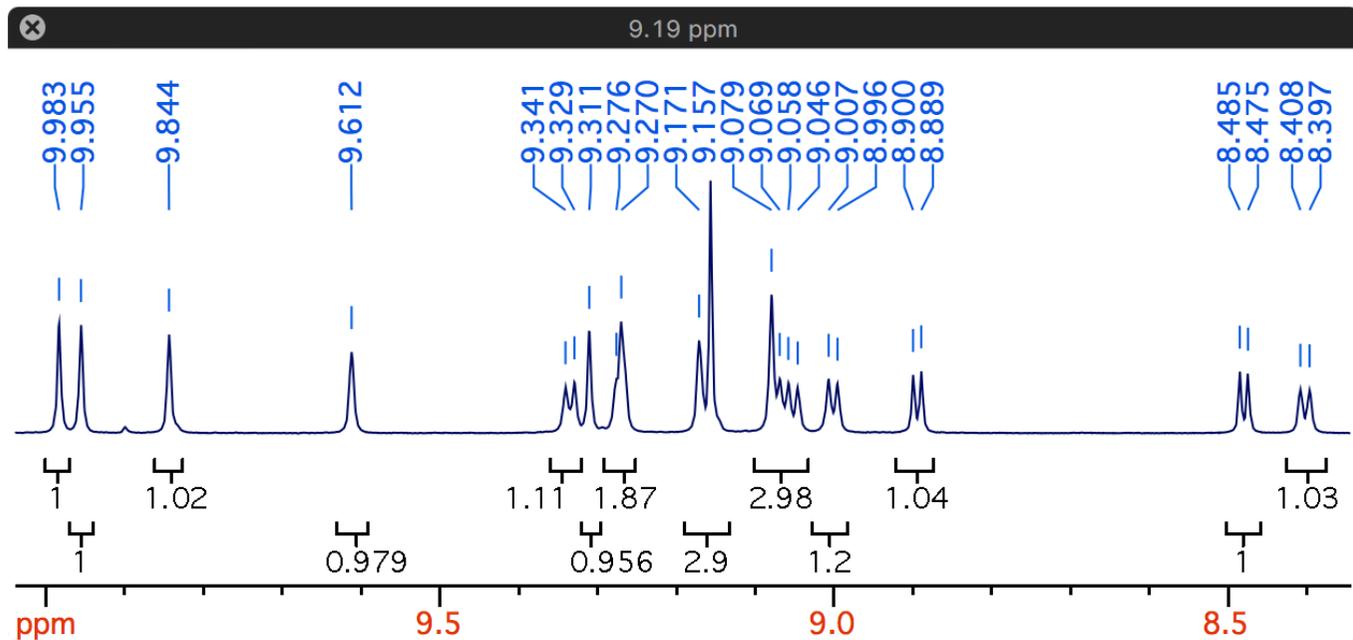
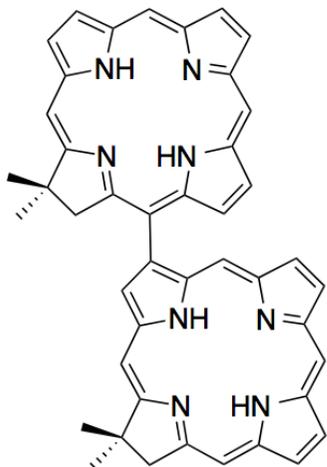


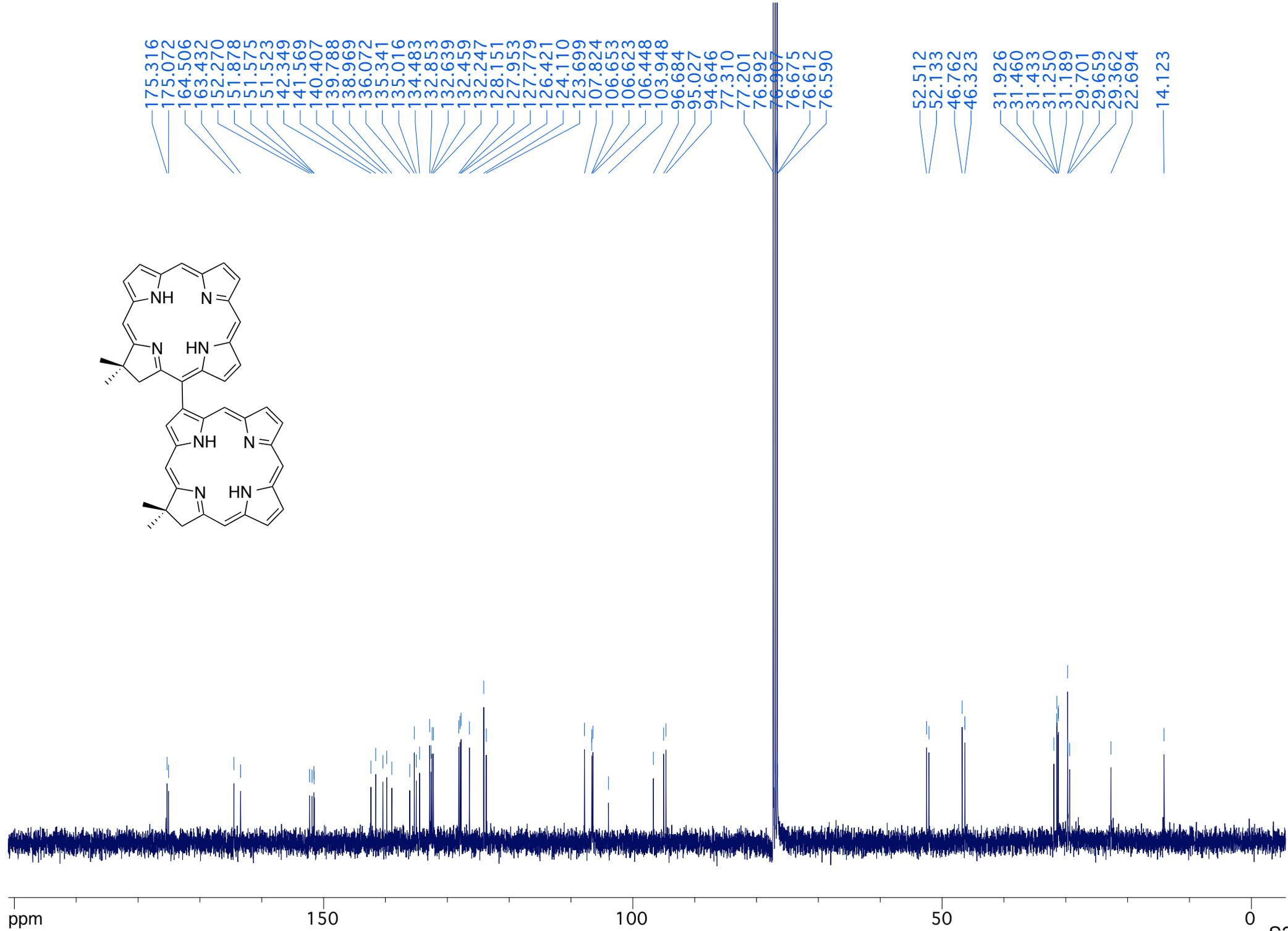
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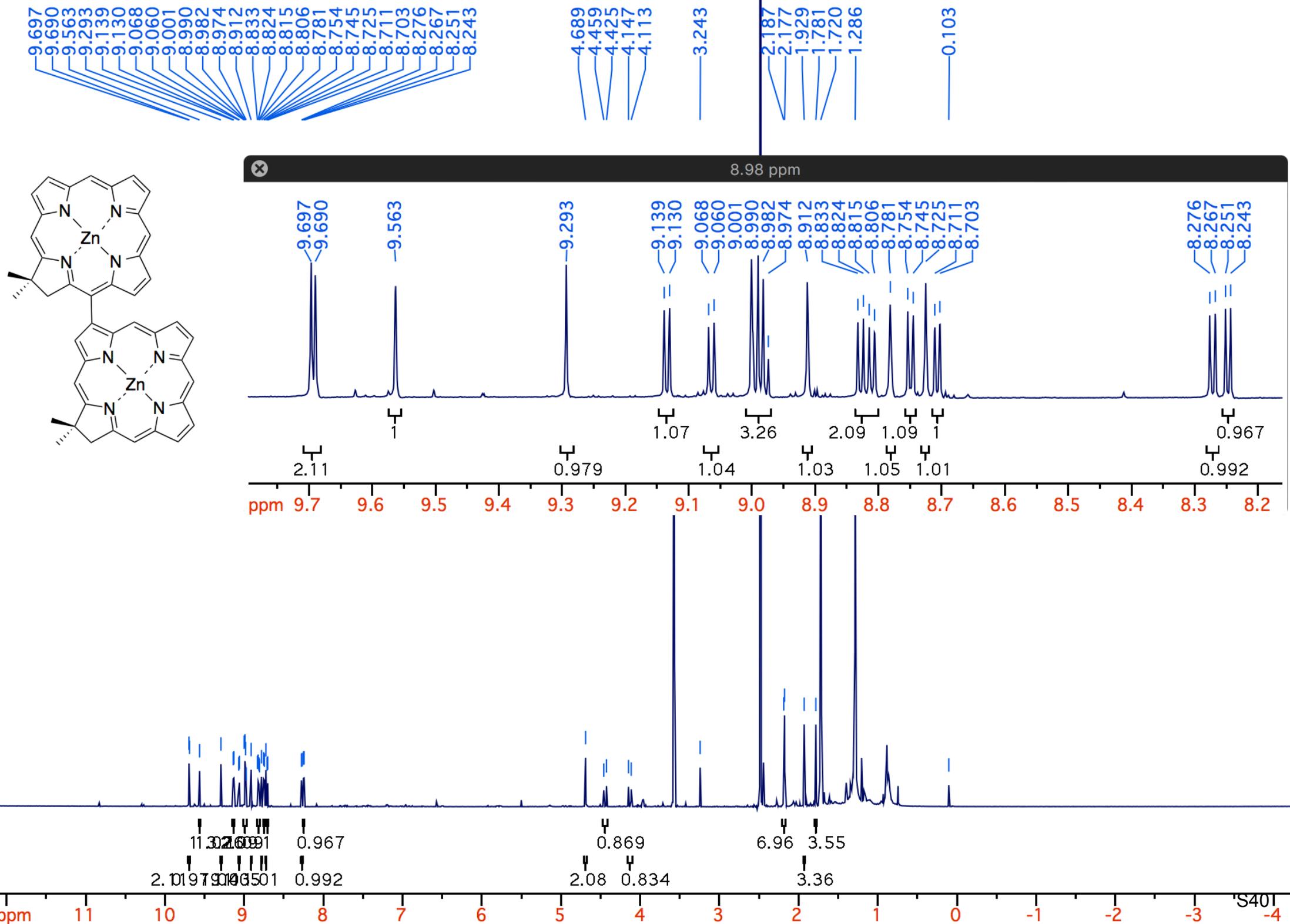
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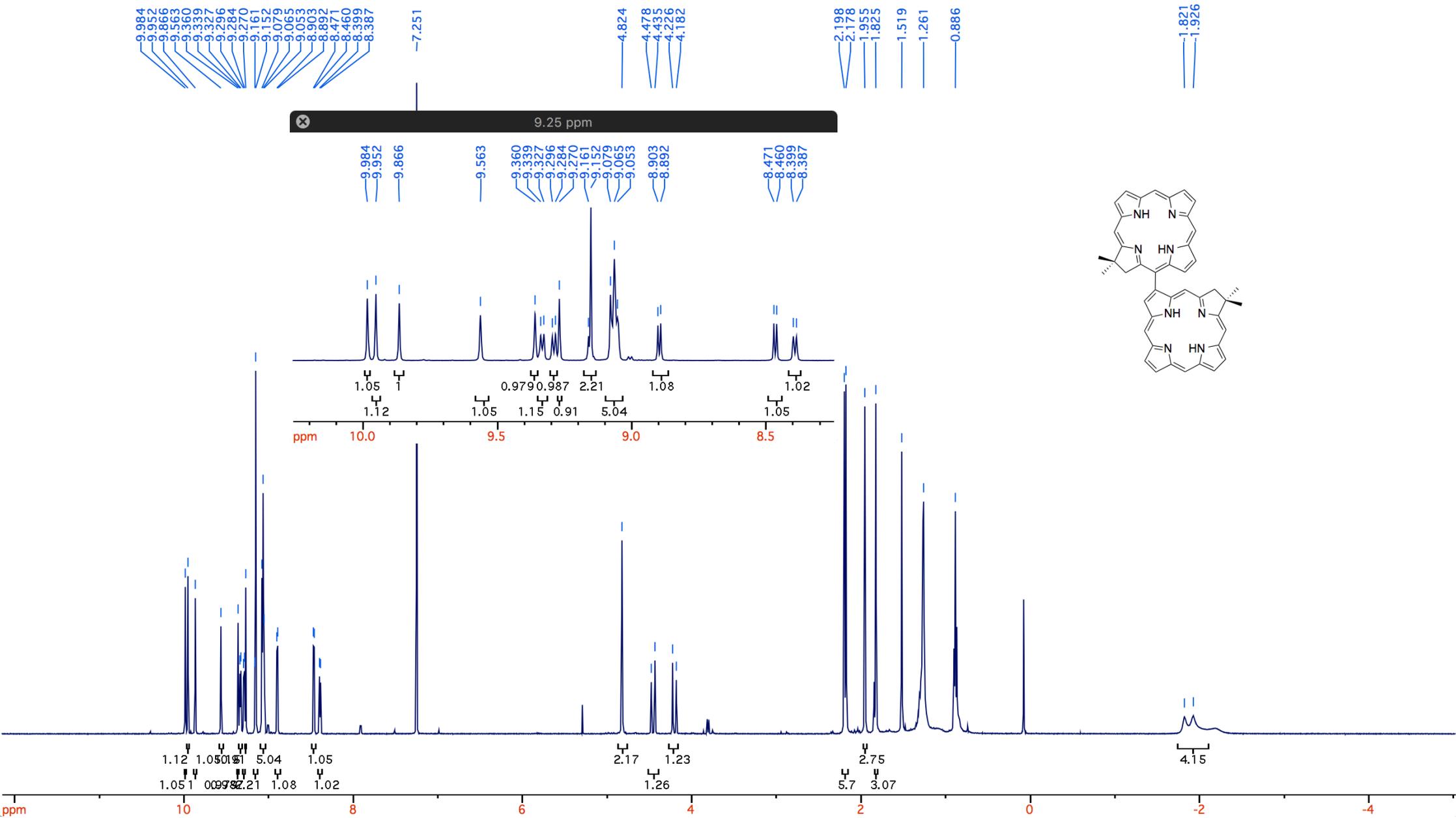
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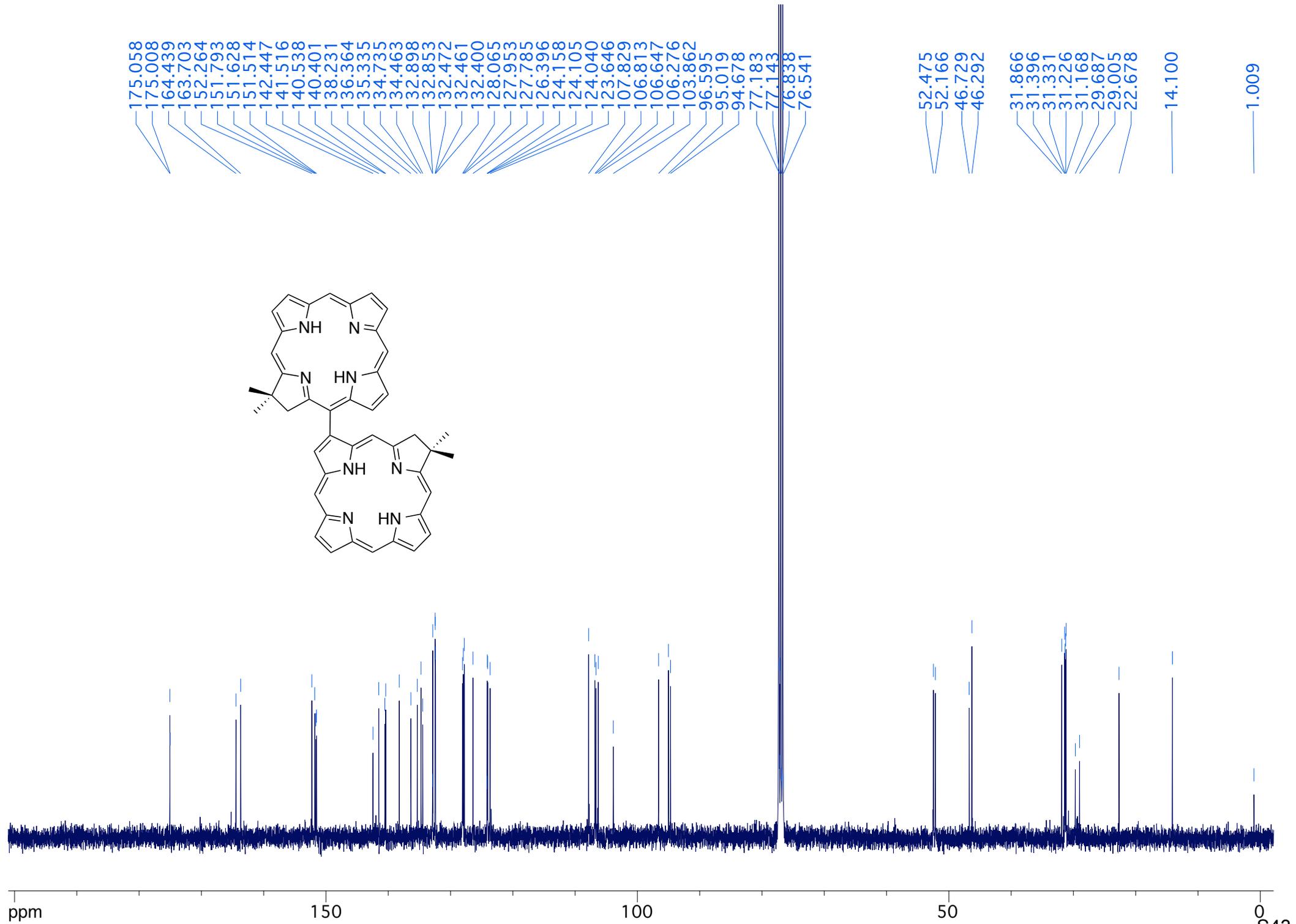
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1.904

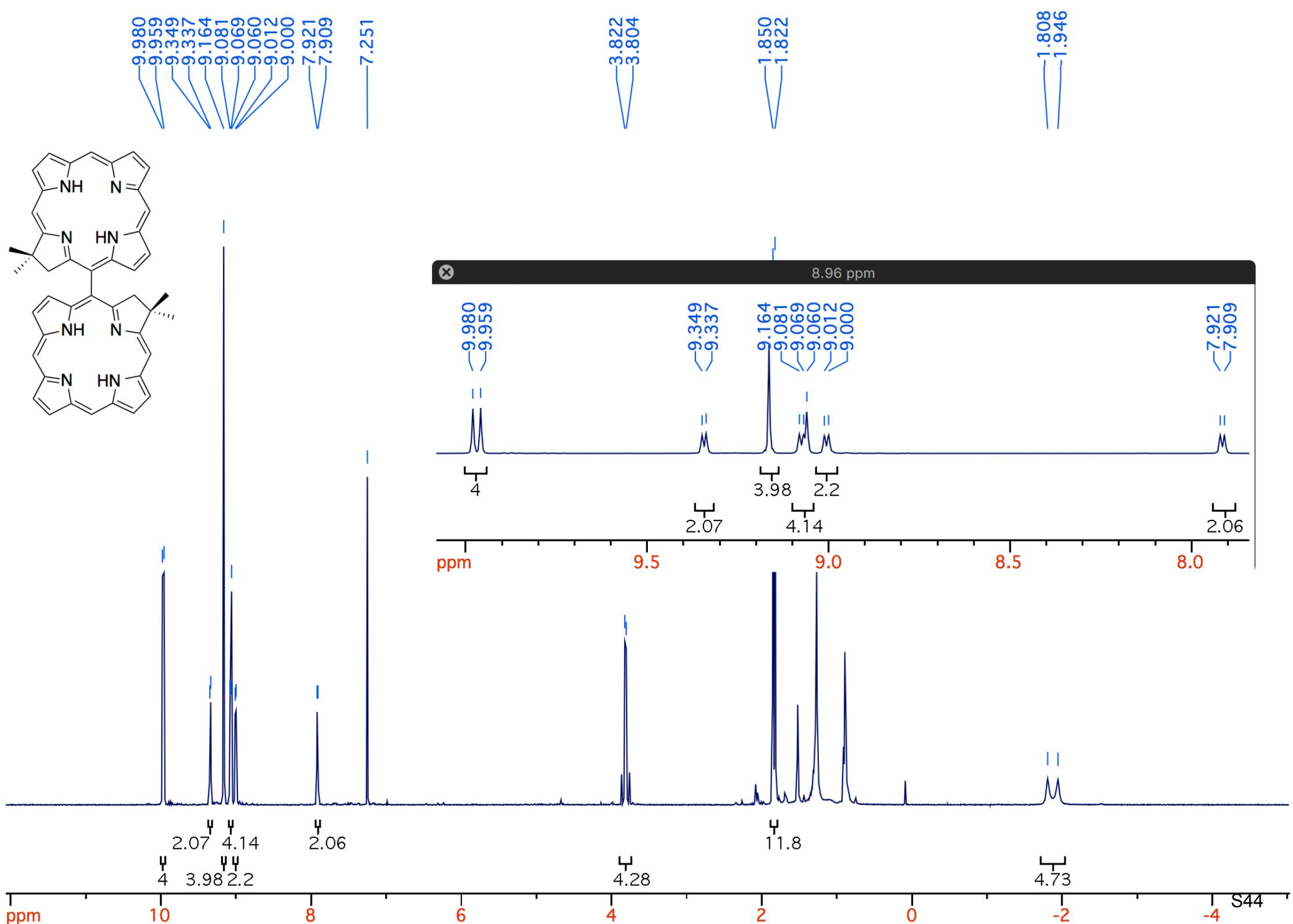


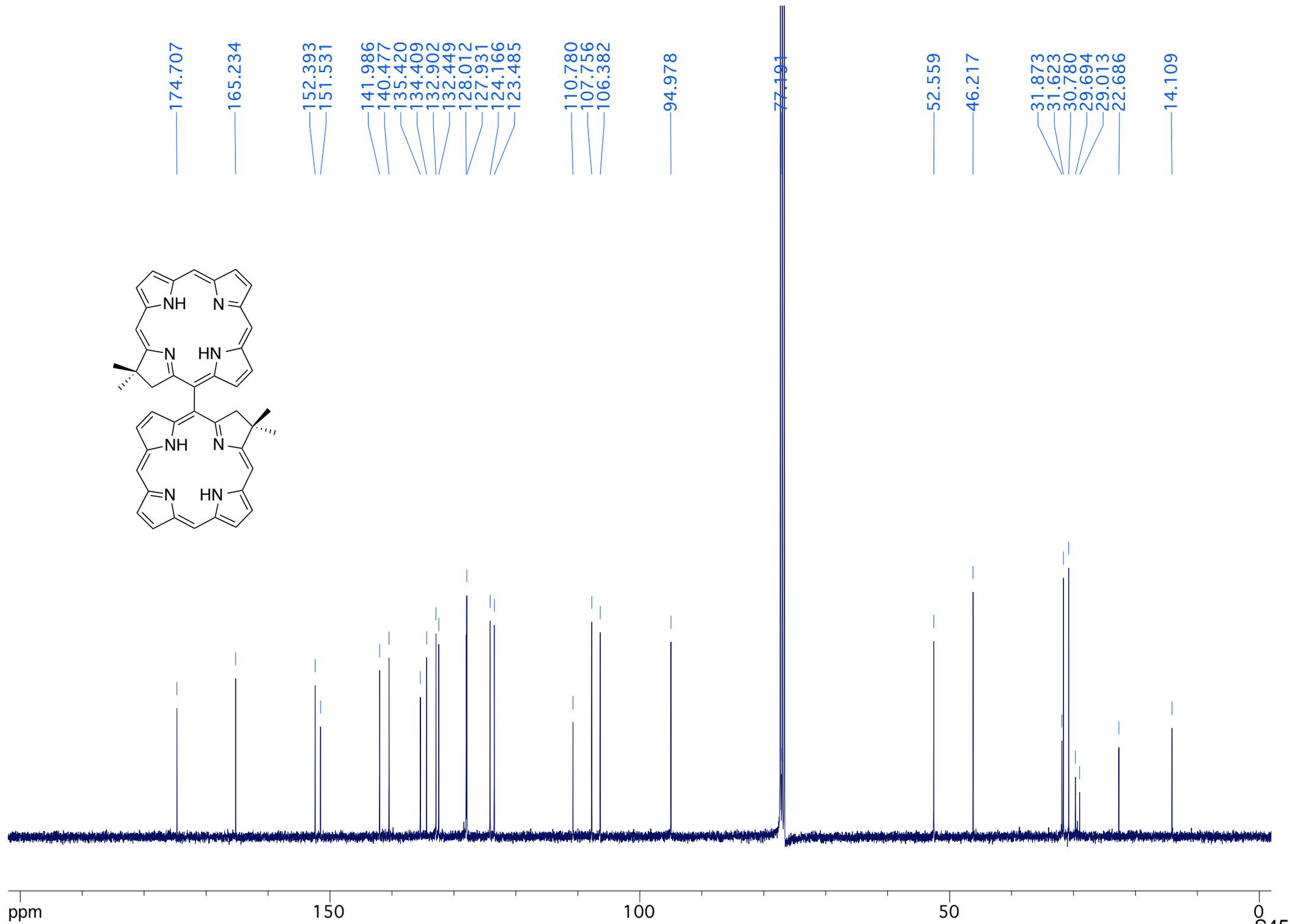
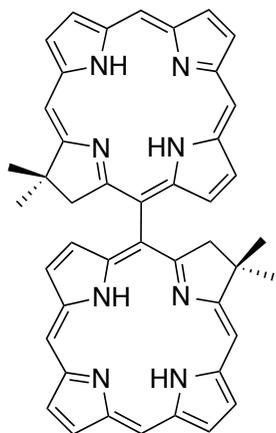


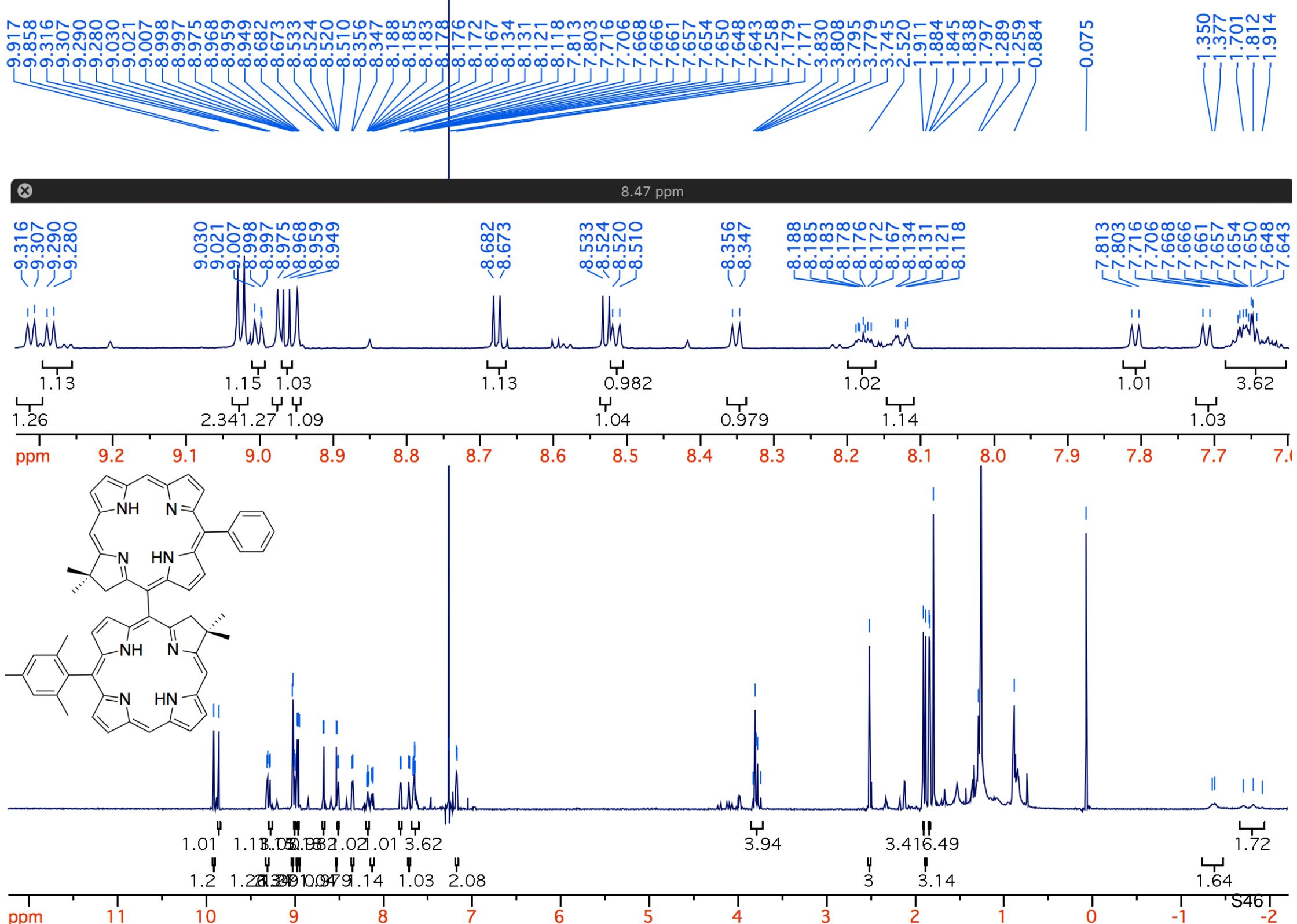


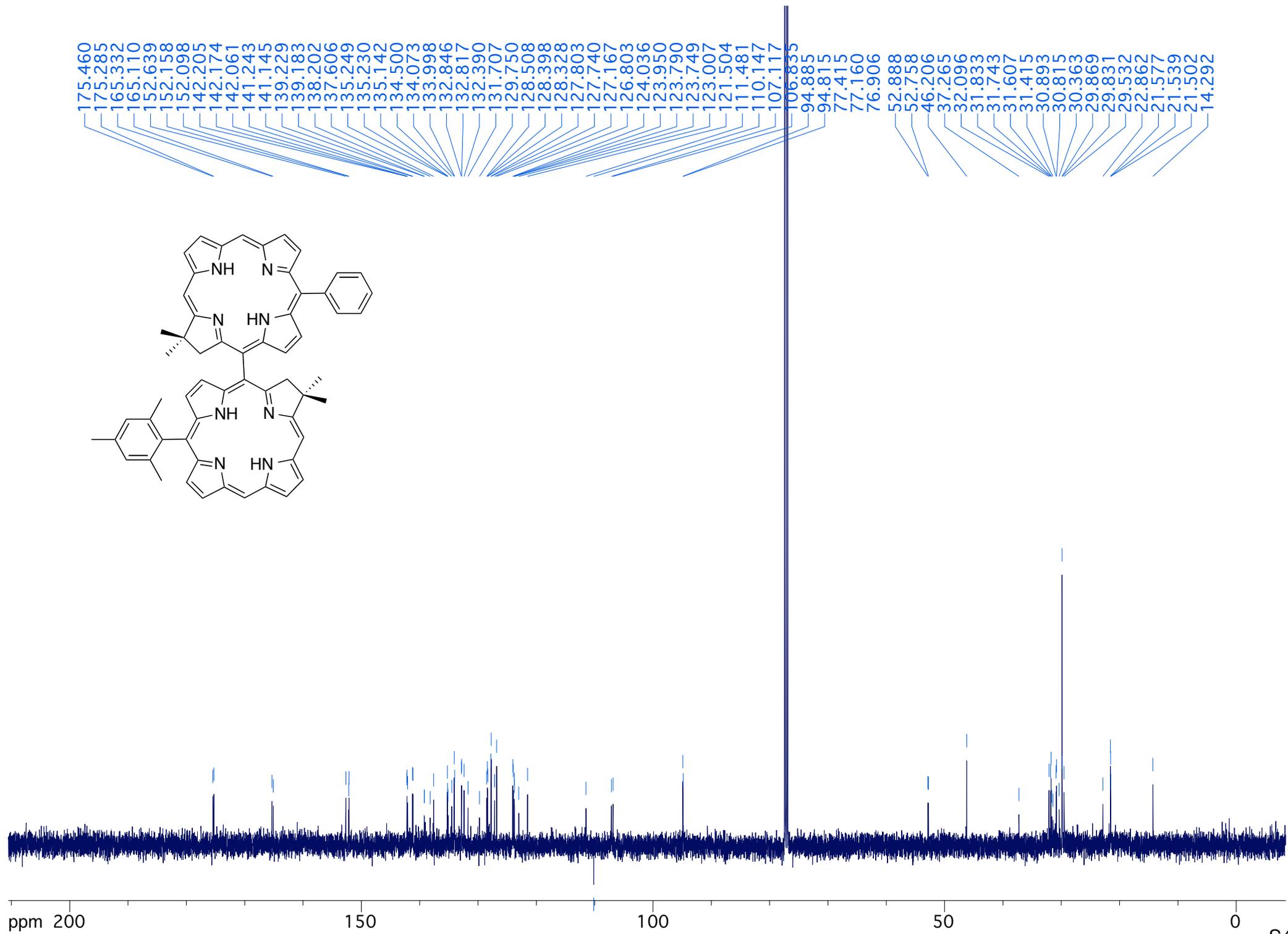


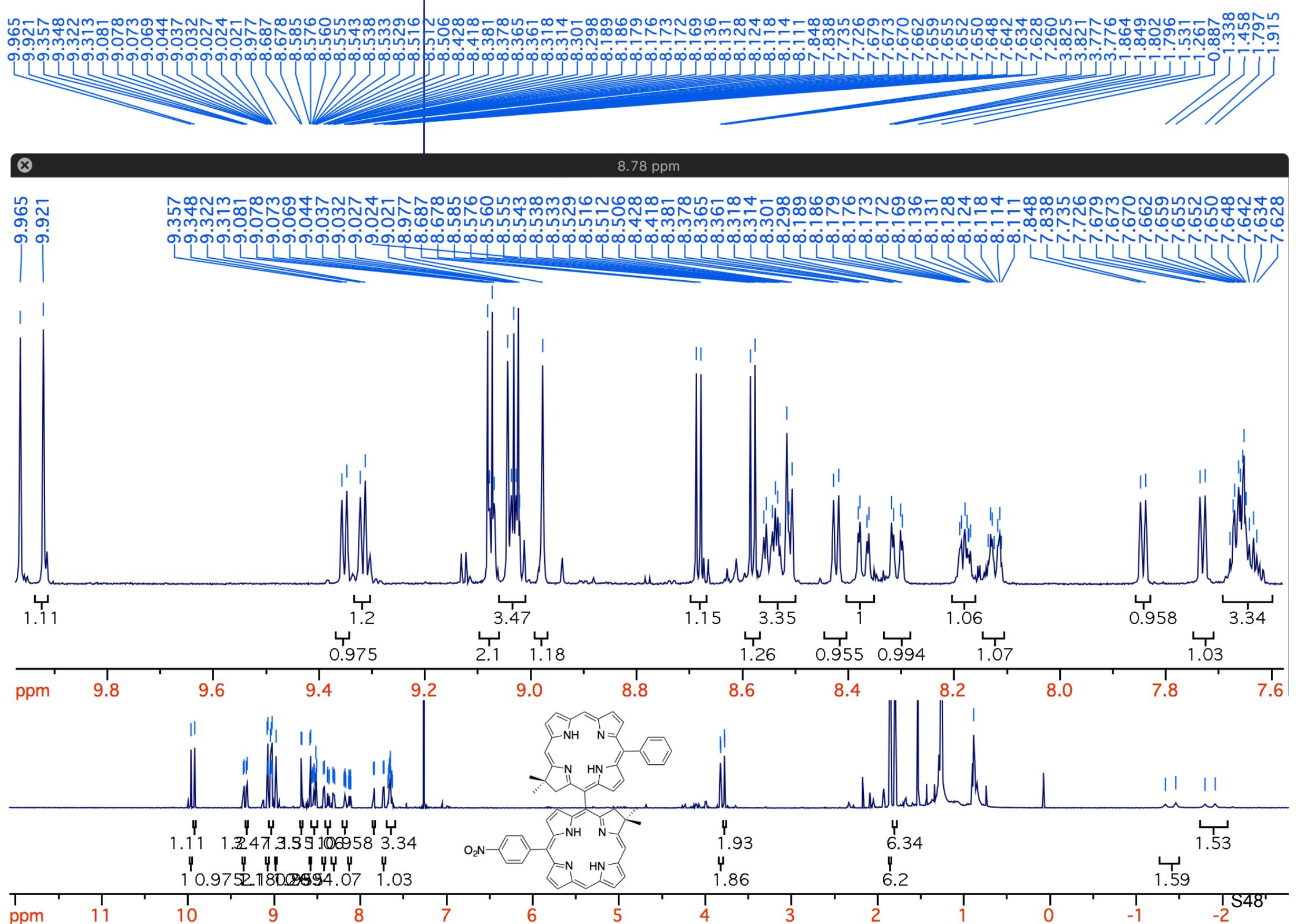


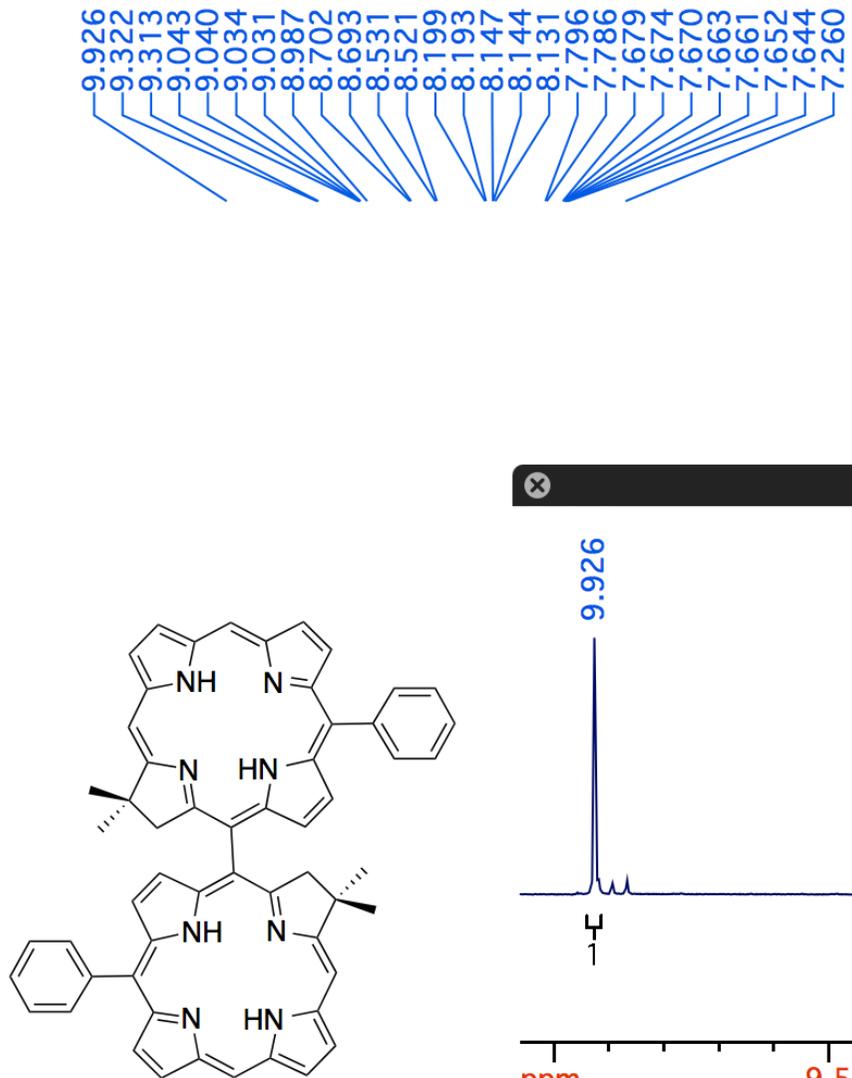












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1.789

