Supporting Information for

Enantioselective tandem reaction over a site-isolated bifunctional catalyst

Jianyu Xu, Tanyu Cheng, Kun Zhang, Ziyun Wang and Guohua Liu*

Key Laboratory of Resource Chemistry of Ministry of Education, Shanghai Key Laboratory of Rare Earth Functional Materials, Shanghai Normal University, Shanghai 200234, P. R. China.

CONTENTS

Experimental part and Data of chiral products2
Figure S1. The FT-IR spectrum of catalyst 311
Figure S2. The solid-state ¹³ C CP MAS NMR spectra of of the fresh catalyst 5 and
the recycled catalyt 512
Figure S3. The solid-state ³¹ P CP MAS NMR spectrum of catalyst 513
Figure S4. The solid-state ²⁹ Si CP MAS NMR spectrum of catalyst 513
Figure S5. The small-angle powder XRD pattern of catalyst 514
Figure S6. The nitrogen adsorption-desorption isotherm of catalyst 515
Figure S7. The TEM imanges of the fresh catalyst 5 and the recycled catalyst 516
Table S1. Optimizing reaction conditions for the tandem Sonogashira coupling–ATH
of 4-iodoacetophenone and phenylacetylene17
Figure S8. Characterizations of chiral products18
Figure S9. HPLC analyses for chiral products
Table S2. Reusability of catalyst 5
Figure S10. Reusability of catalyst 5

Experimental

1). General. All experiments, which are sensitive to moisture or air, were carried out under an Ar atmosphere using the standard Schlenk techniques. Diphenyl(2-(triethoxysilyl)ethyl)phosphine, (TEOS), tetraethoxysilane 1,4-bis(triethyoxysilyl)ethane, cetyltrimethylammonium (CTAB), bromide fluorocarbon surfactant (FC-4:

 $[C_{3}F_{7}O(CF(CF_{3})CF_{2}O)_{2}CF(CF_{3})CONH(CH_{2})_{3}N^{+}(C_{2}H_{5})_{2}CH_{3}]I^{-}),$

4-(2-(trimethoxysilyl)ethyl)benzene-1-sulfonyl chloride, 4-(methylphenylsulfonyl)-1,2-diphenylethylenediamine [(S,S)-TsDPEN], surfactant P123 (CH₂-CH₂O)₂₀(CH₂(CH₃)CH₂O)₇₀(CH₂CH₂O)₂₀), [mesityleneRuCl₂]₂ were purchased from Sigma-Aldrich Company Ltd and used as received. Compound of (*S*,*S*)-4-(trimethoxysilyl)ethyl)phenylsulfonyl-1,2-diphenylethylenediamine [*J. Mater.*

Chem., **2010**, *20*, 1970.] and bis[(diphenylphosphino)ethyltriethoxysilane)]palladium dichloride [*J. Catal.* **1998**, *178*, 284] were synthesized according to the reported literature.

2). Characterization. Ru, Pd loading amounts in the catalyst was analyzed using an inductively coupled plasma optical emission spectrometer (ICP, Varian VISTA-MPX). Fourier transform infrared (FTIR) spectra were collected on a Nicolet Magna 550 spectrometer using KBr method. X-ray powder diffraction (XRD) was carried out on a Rigaku D/Max-RB diffractometer with CuKa radiation. Scanning electron microscopy (SEM) images were obtained using a JEOL JSM-6380LV microscope operating at 20 kV. Transmission electron microscopy (TEM) images were performed on a JEOL JEM2010 electron microscope at an acceleration voltage of 220 kV. Nitrogen adsorption isotherms were measured at 77 K with a Quantachrome Nova 4000 analyzer. The samples were measured after being outgassed at 423 K overnight. Pore size distributions were calculated by using the BJH model. The specific surface areas (SBET) of samples were determined from the linear parts of BET plots ($p/p_0 =$ 0.05-1.00). Solid state NMR experiments were explored on a Bruker AVANCE spectrometer at a magnetic field strength of 9.4 T with ¹H frequency of 400.1 MHz, ¹³C frequency of 100.5 MHz and ²⁹Si frequency of 79.4 MHz, and ³¹P frequency of 169.3 MHz with 4 mm rotor at two spinning frequency of 5.5 kHz and 8.0 kHz, TPPM decoupling is applied in the during acquisition period. ¹H cross polarization in all solid state NMR experiments was employed using a contact time of 2 ms and the pulse lengths of 4 μ s.

3). Preparation of Catalyst 5. In a typical synthesis, (The first step for the synthesis of mesoporous yolk functionalized with chiral siloxane) 0.10 g (0.27 mmol) of cetyltrimethylammonium bromide (CTAB) was completely dissolved in 45.0 mL of aqueous sodium hydroxide (0.35 mL, 2.0 N). The mixture was stirred at room temperature for 0.5 h. Subsequently, 0.18 (0.50)mmol) of g 1,2-bis(triethoxysilyl)ethane (1), 0.125 g (0.25)mmol) of (S,S)-4-(trimethoxysilyl)ethyl)phenylsulfonyl-1,2-diphenylethylenediamine (2) and 0.43 g of (2.07 mmol) of tetraethoxysilane (TEOS) was added at room temperature under vigorous stirring. Finally, 0.40 mL of ethyl acetate was added and the mixture was stirred at 80 °C for 2 h. (The second step for the coating above yolk with a SiO₂-coated layer) After cooling the above mixture down to 38 $^{\circ}$ C, an aqueous solution (80 mL of water, 50 mL of ethanol, 0.30 g (0.82 mmol) of CTAB and 1.0 mL (25 wt%) of NH₃·H₂O) was added and the mixture was stirred 38 °C for 0.5 h. Subsequently, 0.5 mL, 0.47 g (2.26 mmol) of TEOS was added and the mixture was stirred at 38 °C for another 2 h. (The third step for the coating above SiO₂-coated yolk with an ethylene-bridged organopalladium-functionalized silica layer) An aqueous solution (3 mL of water containing 0.04 g (0.044 mmol) of FC-4 $([C_3F_7O(CF(CF_3)CF_2O)_2CF(CF_3)CONH(CH_2)_3N^+(C_2H_5)_2CH_3]\Gamma), 0.08 g (0.22 mmol)$ of CTAB and 0.20 mL (25 wt%) of NH₃·H₂O) was added and the mixture was stirred at 38 °C for 0.5 h. Then, 0.89 g of 1,2-bis(triethoxysilyl)ethane (2.50 mmol) in 2 mL of ethanol and 0.116 g (0.125)mmol) of bis[(diphenylphosphino)ethyltriethoxysilane)]palladium dichloride (3) (2 min later) were added subsequently under vigorous stirring for 1.5 h. Finally, the temperature was raised to 80 °C and the mixture was stirred at 80 °C for another 3 h. After cooling the above mixture down to room temperature, the solid was collected by filtration. (The fourth step for the selective etching) To remove the surfactant and form yolk-shell structured mesoporous nanoparticles, the collected solids (1.22 g) were dispersed in 120 mL of solution (80 mg (1.0 mmol) of ammonium nitrate in 120 mL (95%) of ethanol), and the mixture was stirred at 60 °C for 10 h. After cooling the above mixture down to room temperature, the solid was filtered and washed with excess water and ethanol, and dried at ambient temperature under vacuum overnight to afford Shell@SiO₂@Yolk as a light-yellow powder (0.86 g). (*The fifth step for the* complexation) 50.0 mg of [MesityleneRuCl₂]₂ (4) (0.086 mmol) was added to a suspension of Shell@SiO₂@Yolk (0.50 g) in 20.0 mL of dry CH₂Cl₂ at room temperature, and the resulting mixture was stirred at 25 °C for 12 h. The mixture was

filtered through filter paper and then rinsed with excess CH₂Cl₂. After Soxhlet extraction for 24 h in CH₂Cl₂ to remove homogeneous and unreacted starting materials, the solid was dried at ambient temperature under vacuum overnight to afford PdPPh₂@MesityleneRuArDPEN@MNPs (**5**) (0.52 g) as a red powder. ICP analysis showed that the Pd and Ru loadings were 5.38 mg (0.0508 mmol of Pd) and 10.12 mg (0.099 mmol of Ru) per gram of catalyst, respectively. IR (KBr) cm⁻¹: 3424.9 (s), 3061.8 (w), 2977.6 (w), 2921.9 (w), 1636.4 (m), 1496.6 (w), 1449.9 (w), 1412.4 (w), 1384.6 (w), 1077.1 (s), 787.7 (m), 695.2 (m), 573.4 (w), 545.6 (w), 462.1 (m). ¹³C CP/MAS NMR (161.9 MHz): 147.7, 138.9, 128.1 (<u>C</u> of Ph and Ar groups), 105.5, 102.5 (<u>C</u> of mesitylene groups), 76.5, 71.6 (<u>C</u>H of -NCHPh), 59.8 (*C*H₂ of -OCH₂CH₃), 53.4 (-NCH₂- or -NCH₃ of CTAB), 29.8 (<u>C</u>H₂ of -CH₂P), 22.9 (<u>C</u>H₂ of -CH₂Ar), 20.5 (<u>C</u>H₃ of mesitylene), 18.0 (<u>C</u>H₃ of -OCH₂CH₃, and -<u>C</u>H₃ or -*C*H₂- of CTAB), 4.8 (-<u>C</u>H₂Si-) ppm; ³¹P MASNMR (169.3 MHz): 39.2 (cis), 29.8 (trans) ppm. ²⁹Si MAS NMR (79.4 MHz): T² (δ = -58.7 ppm), T³ (δ = -66.4 ppm), Q² (δ = -96.3 ppm), Q³ (δ = -102.2 ppm), Q⁴ (δ = -111.8 ppm).

4). General procedure for one-pot tandem reaction. A typical procedure was as follows. Catalyst **5** (19.68 mg, 1.00 μ mol of Pd and 1.95 μ mol of Ru, based on ICP analysis), K₂CO₃ (13.80 mg, 0.10 mmol), HCO₂Na (68.0 mg, 1.0 mmol), iodoacetophenones (0.10 mmol), aryne (0.11 mmol), and 4.0 mL of the mixed solvents (H₂O/MeOH, v/v = 1/3) were added sequentially to a 10.0 mL round-bottom flask. The mixture was then stirred at 60 °C for 10-16 h. During this period, the reaction was monitored constantly by TLC. After completion of the reaction, the catalyst was separated by centrifugation (10,000 rpm) for the recycling experiment. The aqueous solution was extracted with ethyl ether (3 × 3.0 mL). The combined ethyl ether extracts were washed with brine twice and then dehydrated with Na₂SO₄. After evaporation of ethyl ether, the residue was purified by silica gel flash column chromatography to afford the desired products.

5). Data of chiral products

6a. (*S*)-1-(4-(phenylethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃): δ 7.57–7.53 (m, 4H), 7.40–7.35 (m, 5H), 4.95–4.90 (q, J = 8.0 Hz, 1H), 2.09 (s, 1H), 1.53–1.51 (d, J = 8.0 Hz, 3H); ¹³C{¹H} NMR (101 MHz, CDCl₃): δ 146.30, 141.99, 131.88, 128.63, 128.52, 125.71, 123.6, 122.49, 89.61, 70.21, 25.34; GC/MS

(m/z): 222; HPLC (OD-H, elute: Hexanes/i-PrOH = 97/3, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 16.3 \text{ min (major)}, t_2 = 20.7 \text{ min}.$

6b. (*S*)-1-(4-((4-fluorophenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃):



δ 7.54–7.04 (m, 8H), 4.96–4.91 (q, J = 8.0 Hz, 1H), 1.90 (s, 1H), 1.53–1.51 (d, J = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 162.72 (d, J = 250 Hz), 146.29, 133.67 (d, J = 8.3 Hz), 131.93, 125.67, 122.34, 119.58 (d,

J = 3.6 Hz), 115.87 (d, J = 22 Hz), 89.15, 88.45, 70.30, 25.38; GC/MS (m/z): 240; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t₁ = 16.5 min (major), t₂ = 18.9 min.

6c. (S)-1-(4-((3-fluorophenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃): δ 7.52–7.20 (m, 8H), 4.95–4.90 (q, J = 8.0 Hz, 1H), 1.78 (s, δ 7.52–7.20 (m, 8H), 4.95–4.90 (q, J = 8.0 Hz, 1H), 1.78 (s, 1H), 1.51–1.49 (d, J = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 162.85(d, J = 250 Hz), 146.59, 133.67, 132.08, 130.17 (d, J = 7.7 Hz), 115.76 (d, J = 21 Hz), 124.19 (d, J = 3.5 Hz), 122.12, (d, J = 15 Hz), 94.55, 82.83, 70.29, 25.37; GC/MS (m/z): 240; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t₁ = 19.9 min (major), t₂ = 23.1 min.

6d. (S)-1-(4-((2-fluorophenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃): δ 7.57–7.12 (m, 8H), 4.96–4.91 (q, J = 8.0 Hz, 1H), 1.93 (s, ¹H), 1.53–1.51 (d, J = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 162.87 (d, J = 250 Hz), 146.34, 133.68, 130.24 (d, J = 7.8 Hz), 128.95, 128.80, 125.97, 124.20 (d, J

130.24 (d, J = 7.8 Hz), 128.95, 128.80, 125.97, 124.20 (d, J = 13 Hz), 123.23, 115.76 (d, J = 21 Hz), 112.09 (d, J = 15

Hz), 94.88, 83.00, 70.25, 25.41; GC/MS (m/z): 240; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 21.9$ min (major), $t_2 = 26.2$ min.



25.40; GC/MS (m/z): 300; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 19.2 \text{ min (major)}, t_2 = 21.6 \text{ min}.$

6f. (**S**)-1-(4-((4-bromophenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃):



δ 7.54–7.04 (m, 8H), 4.96–4.91 (q, *J* = 8.0 Hz, 1H), 1.90 (s, 1H), 1.53–1.51 (d, *J* = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 145.20, 131.99, 130.73, 130.60, 124.43, 121.44, 12.22, 120.94, 89.34, 87.21, 69.07,

24.15; GC/MS (m/z): 300; HPLC (OD-H, elute:Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 20.4$ min (major), $t_2 = 23.4$ min.

6g. (S)-1-(4-(p-tolylethynyl)phenyl)ethanol. ¹H NMR (400 MHz, $CDCl_3$): δ



7.51–7.49 (m, 2H), 7.48–7.43 (m, 2H), 7.41–7.36 (m, 2H), 7.33–7.14 (m, 2H), 4.93–4.88 (q, J = 8.0 Hz, 1H), 2.37 (s, 3H), 1.87 (s, H), 1.50–1.49 (d, J = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 146.13, 138.25, 132.43,

131.98, 129.40, 128.92, 128.49, 125.63, 123.30, 122.66, 89.75, 89.16, 70.32, 25.36, 21.47; GC/MS (m/z): 236; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 19.9$ min (major), $t_2 = 22.5$ min.

6h. (**S**)-1-(4-(**m-tolylethynyl**)**phenyl**)**ethanol.** ¹H NMR (400 MHz, CDCl₃): δ 7.54–7.52 (m, 2H), 7.39–7.35 (m, 2H), 7.28–7.16 (m, 2H)

6h (entry 8 in Table 1)

7.54–7.52 (m, 2H), 7.39–7.35 (m, 2H), 7.28–7.16 (m, 2H) 4.96–4.91 (q, J = 8.0 Hz, 1H), 2.38 (s, 3H), 1.53–1.51 (d, J = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 146.13,

 $138.25, 132.43, 131.98, 129.40, 128.92, 128.49, 125.63, 123.30, 122.66, 89.75, 89.16, 70.32, 25.36, 21.47; GC/MS (m/z): 236; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t_1 = 19.6 min (major), t_2 = 24.1 min.$



89.53, 88.16, 70.34, 55.52, 25.34; GC/MS (m/z): 252; HPLC (OD-H, elute: Hexanes/i-PrOH = 95/5, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 16.1$ min (major), $t_2 = 20.7$ min.

6j. (S)-1-(4-((3-methoxyphenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz,



6j (entry 10 in Table 1)

CDCl₃): δ 7.55-6.90 (m, 8H), 4.91–4.86 (q, *J* = 8.0 Hz, 1H), 3.83 (s, 3H), 2.43 (s, 1H), 1.59–1.48 (d, *J* = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl3): δ 159.58, 146.41, 132.01, 129.66, 126.20, 125.65, 116.60, 115.17,

89.46, 89.32, 70.30, 55.52, 25.37; GC/MS (m/z): 252; HPLC (OD-H, elute: Hexanes/i-PrOH = 95/5, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 14.5$ min (major), $t_2 = 17.3$ min.

6k. (S)-4-((4-(1-hydroxyethyl)phenyl)ethynyl)benzonitrile. 1 H NMR (400 MHz,NC-OHCDCl_3): δ 7.69-7.18 (m, 8H), 4.92-4.87 (q, J = 8.0 Hz,6k (entry 11 in Table 1)NMR (101 MHz, CDCl_3): δ 146.97, 132.06, 131.97,128.27, 125.57, 121.22, 118.55, 111.43, 90.70, 87.66,

70.02, 25.24; GC/MS (m/z): 247; HPLC (AD-H, elute: Hexanes/i-PrOH = 95/5, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 33.8 \text{ min}$ (major), $t_2 = 4.2 \text{ min}$.

6l. (S)-1-(4-((4-(trifluoromethyl)phenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl3): δ 7.66–7.38 (m, 8H), 4.96–4.91 (q, J = 8.0 Hz, 1H), 2.07 (s, 1H), 1.53–1.51 (d, J = 8.0 Hz, 3H); ^{6l} (entry 12 in Table 1) ¹³C {1H} NMR (101 MHz, CDCl3): δ 146.62, 131.19, 131.80, 130.05(q, J = 32.7 Hz), 127.12, 125.51, 125.28

(q, J = 3.7 Hz), 122.61, 121.57, 91.65, 87.92, 70.05, 25.18; GC/MS (m/z): 300; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t₁ = 20.8 min (major), t₂ = 23.1 min.



flow rate: 1 mL/min, 25 °C), $t_1 = 16.5 \text{ min (major)}$, $t_2 = 22.7 \text{ min}$.

6n. (S)-1-(3-((4-fluorophenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl3):



 δ 7.46–7.04 (m, 8H), 4.95–4.90 (q, *J* = 8.0 Hz, 1H), 2.01 (s, 1H), 1.52–1.50 (d, *J* = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl3): δ 162.86 (d, *J* = 250 Hz), 146.59, 133.67, 132.08, 130.17 (d, *J* = 7.7 Hz), 125.64, 124.19 (d, *J* = 3.5 Hz), 122.16 (d, *J* = 15 Hz), 115.75 (d, J=21Hz), 112.16 (d, *J*

= 15 Hz), 94.51, 82.83, 70.29, 25.37; GC/MS(m/z): 240; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 16.5$ min (major), $t_2 = 18.9$ min.

60. (**S**)-1-(3-((3-fluorophenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃):



 δ 7.58–7.06 (m, 8H), 4.94–4.89 (q, *J* = 8.0 Hz, 1H), 2.19 (s, 1H), 1.53–1.51 (d, *J* = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 162.63 (d, *J* = 250 Hz), 146.37, 130.91, 130.17 (d, *J* = 8.6 Hz), 128.93, 128.84, 127.73 (d, *J* = 3.1 Hz), 125.98, 125.29 (d, *J* = 9.8 Hz), 123.12, 118.59 (d, *J* = 2.3 Hz), 94.44,

88.32, 70.23, 25.45; GC/MS (m/z): 240; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 20.4 \text{ min (major)}, t_2 = 25.1 \text{ min}.$

6p. (S)-1-(3-((2-fluorophenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃): HO δ 7.39–7.10 (m, 8H), 4.94–4.89 (q, J = 6.40 Hz, 1H), 2.15 (s,



6p (entry 16 in Table 1)

 $\delta7.39-7.10$ (m, 8H), 4.94–4.89 (q, J = 6.40 Hz, 1H), 2.15 (s, 1H), 1.55–1.51 (d, J = 6.40 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 164.12, 161.62, 146.34, 133.69, 130.94, 130.28, 130.20, 128.94, 128.80, 124.18, 115.43, 94.58, 82.90, 70.24, 25.41; GC/MS(m/z): 240; HPLC (OD-H, elute:

Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 oC), t_1 =24.7 min (major), t_2 = 26.7 min.



Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t1 = 23.8 min (major), t2 = 26.6 min.

6r. (**S**)-**1**-(**3**-((**4**-bromophenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃):



Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t_1 = 24.0 min (major), t_2 =27.3 min.

6s. (S)-1-(3-(p-tolylethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃): δ



7.51–7.14 (m, 8H), 4.93–4.88 (q, J = 4.0 Hz, 1H), 2.37 (s, 3H), 1.87 (s, 1H), 1.50–1.49 (d, J = 4.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 146.32, 138.69, 131.77, 130.77, 129.40, 128.83, 128.77, 125.50, 123.80, 123.80, 89.86, 89.00, 70.25, 25.39, 21.75; GC/MS (m/z): 236; HPLC (OD-H, elute:

Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t_1 = 22.7 min (major), t_2 = 24.0 min.

6t. (S)-1-(3-(m-tolylethynyl)phenyl)ethanol. ¹H NMR (400 MHz, CDCl₃): δ HO 3H),1.88 (s, 1H), 1.53–1.51 (d, J = 8.0 Hz, 1H), 2.38 (s, 3H),1.88 (s, 1H), 1.53–1.51 (d, J = 8.0 Hz, 3H); ¹³C {1H} NMR (101 MHz, CDCl₃): δ 146.28, 138.26, 132.44, 130.83, 129.46, 128.94, 128.49, 125.55, 123.73, 123.24, 89.84, 89.23, 70.28, 25.39, 21.46; GC/MS (m/z): 236; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t₁ = 20.9 min (major), t₂ = 27.3 min.



6v. (S)-1-(3-((3-methoxyphenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 MHz, MeO MeOMeO

(major), $t_2 = 16.6$ min.

6w. (S)-1-(3-((4-(trifluoromethyl)phenyl)ethynyl)phenyl)ethanol. ¹H NMR (400

F₃C - **6w** (entry 23 in Table 1) MHz, CDCl₃): δ 7.66-7.37 (m, 8H), 4.95–4.90 (q, J = 8.0 Hz, 1H), 2.13 (s, 1H), 1.53–1.51 (d, J = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 146.24, 131.82, 130.73, 130.02 (q, J = 32.5 Hz), 128.78, 128.66, 127.07, 126.01, 125.30 (q, J = 3.3 Hz), 122.67, 91.74, 87.98, 69.95,

55.52, 25.21; GC/MS (m/z): 290; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 18.9 \text{ min (major)}$, $t_2 = 23.9 \text{ min}$.

6x. (S)-1-(4-(hex-1-yn-1-yl)phenyl)ethanol ¹H NMR (400 MHz, CDCl₃): δ 7.40-7.28 (dd, J = 8.0 Hz, 4H), 4.92–4.87 (q, J = 8.0 Hz, 1H), 2.14–2.11 (t, J = 8.0 Hz, 1H), 1.84 (s, 1H), 1.64–1.45 (m, 7H), 0.98–0.95 (t, J = 8.0 Hz, 3H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 145.28, 131.89, 125.16, 123.44, 90.59, 80.55, 70.33, 31.07, 25.32, 22.25, 19.33, 13.87; GC/MS (m/z): 266; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), t₁ = 12.4min (major), t₂ = 17.5 min.



266; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 33.8 \text{ min}$ (major), $t_2 = 53.6 \text{ min}$.

6z. (S)-1-(3-((4-((S)-1-hydroxyethyl)phenyl)ethynyl)phenyl)ethanol. ¹H NMR (400 HO HO HO HO HO HO HZ, 2H), 2.25 (s, 2H), 1.53–1.48 (d, J = 4.0 Hz, 6H); ¹³C{1H} NMR (101 MHz, CDCl₃): δ 146.09, 146.02, 131.77, 130.61, 128.65, 128.56, 125.45, 125.42, 123.37, 122.22, 89.30, 89.27, 70.07, 70.01, 25.17, 25.12; GC/MS

(m/z): 266; HPLC (OD-H, elute: Hexanes/i-PrOH = 98/2, detector: 254 nm, flow rate: 1 mL/min, 25 °C), $t_1 = 33.6$ min (major), $t_2 = 37.6$ min.

Figure S1. The FT-IR spectrum catalyst 5.









Figure S3. The solid-state ³¹P CP MAS NMR spectrum of catalyst **5**.



Figure S4. The solid-state ²⁹Si CP MAS NMR spectrum of catalyst 5.



Figure S5. The small-angle powder XRD pattern catalyst 5.





Figure S6. The nitrogen adsorption-desorption isotherms of catalyst 5.

Figure S7. The TEM imanges of the fresh catalyst **5** and the recycled catalyst **5**. TEM images of the fresh catalyst **5**



TEM of the recycled catalyst **5**



Table S1. Optimizing reaction conditions for the tandem Sonogashira coupling–ATH of 4-iodoacetophenone and phenylacetylene.^a

ŌН

			Catalyst 5			
		Γ K ₂	₂ CO ₃ , HCOONa			
Entry	Solvents	Τ (°C)	Time (h)	Yield (%)	ee (%)	TOF ^b
1	H ₂ O	60	14	82	83	5.9
2	i-PrOH	60	12	90	87	7.5
3	EtOH	60	12	92	91	7.7
4	CH ₃ OH	60	7	93	95	13.3
5	H ₂ O/CH ₃ OH (1:1)	60	7	88	93	12.6
6	H ₂ O/CH ₃ OH (1:2)	60	7	91	95	13.0
7	H ₂ O/CH ₃ OH (1:3)	60	7	93	97	13.3
8	H ₂ O/CH ₃ OH (1:3)	70	7	96	95	13.7
9	H ₂ O/CH ₃ OH (1:3)	50	7	85	97	12.1

^a Reaction conditions: catalyst **5** (19.68 mg, 1.00 μ mol of Pd and 1.95 μ mol of Ru, based on ICP analysis), K₂CO₃ (13.80 mg, 0.10 mmol), HCO₂Na (68.0 mg, 1.0 mmol), iodoacetophenones (0.10 mmol), phenylacetylene (0.11 mmol), and 4.0 mL of solvents. ^b TOF (TOF = number of moles of substrate converted per mole of catalyst per hour).







6b (*Entry 2 in Table 1*): (S)-1-(4-((4-fluorophenyl)ethynyl)phenyl)ethanol







6c (*Entry 3 in Table 1*): (S)-1-(4-((3-fluorophenyl)ethynyl)phenyl)ethanol



6d (*Entry 4 in Table 1*): (S)-1-(4-((2-fluorophenyl)ethynyl)phenyl)ethanol







6e (*Entry 5 in Table 1*): (S)-1-(4-((4-chlorophenyl)ethynyl)phenyl)ethanol



6f (*Entry 6 in Table 1*): (S)-1-(4-((4-bromophenyl)ethynyl)phenyl)ethanol







6g (Entry 7 in Table 1): (S)-1-(4-(p-tolylethynyl)phenyl)ethanol



6h (*Entry 8 in Table 1*): (S)-1-(4-(m-tolylethynyl)phenyl)ethanol (5h)







6<u>i (Entry 9 in Table 1):</u> (S)-1-(4-((4-methoxyphenyl)ethynyl)phenyl)ethanol



6j (Entry 10 in Table 1): (S)-1-(4-((3-methoxyphenyl)ethynyl)phenyl)ethanol







6<u>k (Entry 11 in Table 1):</u> (S)-4-((4-(1-hydroxyethyl)phenyl)ethynyl)benzonitrile



6] (Entry 12 in Table 1): (S)-1-(4-((4-(trifluoromethyl)phenyl)ethynyl)phenyl)ethanol









6m (Entry 13 in Table 1): (S)-1-(3-(phenylethynyl)phenyl)ethanol


6<u>n (Entry 14 in Table 1):</u> (S)-1-(3-((4-fluorophenyl)ethynyl)phenyl)ethanol







6<u>o (Entry 15 in Table 1):</u> (S)-1-(3-((3-fluorophenyl)ethynyl)phenyl)ethanol



6<u>p (Entry 16 in Table 1):</u> (S)-1-(3-((2-fluorophenyl)ethynyl)phenyl)ethanol





S41



6<u>q (Entry 17 in Table 1):</u> (S)-1-(3-((4-chlorophenyl)ethynyl)phenyl)ethanol



6<u>r (Entry 18 in Table 1):</u> (S)-1-(3-((4-bromophenyl)ethynyl)phenyl)ethanol









6s (Entry 19 in Table 1): (S)-1-(4-(p-tolylethynyl)phenyl)ethanol



6<u>t (Entry 20 in Table 1):</u> (S)-1-(4-(m-tolylethynyl)phenyl)ethanol









6<u>u (Entry 21 in Table 1):</u> (S)-1-(4-((4-methoxyphenyl)ethynyl)phenyl)ethanol



6<u>v (Entry 22 in Table 1):</u> (S)-1-(4-((3-methoxyphenyl)ethynyl)phenyl)ethanol







6<u>w (Entry 23 in Table 1):</u> (S)-1-(3-((4-(trifluoromethyl)phenyl)ethynyl)phenyl)ethanol



6x (Entry 24 in Table 1): (S)-1-(4-(hex-1-yn-1-yl)phenyl)ethanol











6<u>z (Scheme 3): (S)-1-(3-((4-((S)-1-hydroxyethyl)phenyl)ethynyl)phenyl)</u>ethanol





S56

Figure 9. HPLC analyses for chiral products



6a (Entry 1 in Table 1): (S)-1-(4-(phenylethynyl)phenyl)ethanol



6b (Entry 2 in Table 1): (S)-1-(4-((4-fluorophenyl)ethynyl)phenyl)ethanol





6c (Entry 3 in Table 1): (S)-1-(4-((3-fluorophenyl)ethynyl)phenyl)ethanol





6d (Entry 4 in Table 1): (S)-1-(4-((2-fluorophenyl)ethynyl)phenyl)ethanol 5(d)





6e (Entry 5 in Table 1): (S)-1-(4-((4-chlorophenyl)ethynyl)phenyl)ethanol





<u>6f (Entry 6 in Table 1):</u> (S)-1-(4-((4-bromophenyl)ethynyl)phenyl)ethanol

6g (Entry 7 in Table 1): (S)-1-(4-(p-tolylethynyl)phenyl)ethanol







ОH

18.00

含量 单位

20.00

峰类型

未知

未知

分钟

22.00

峰代码

(S)

16.00

积分类型

14.00

% 面积

98.15

1.85

高度 (微伏)

568484 bb

12371 bb

(R)

26.00

结构1

分子量

结构1

说明

28.00

结构 1 公式 30.0

•

结构1

结构

O

Q

24.066

24.00

结构1 名

6h (Entry 8 in Table 1): (S)-1-(4-(m-tolylethynyl)phenyl)ethanol

₹ 0.30

0.20-

0.10-

0.00

10.5691 分钟, 0.5999 AU

10.00

▲

(分钟)

19.607

24.066

12.00

面积

微伏*秒)

25065496

471846

Ŧ

名称



6i (Entry 9 in Table 1): (S)-1-(4-((4-methoxyphenyl)ethynyl)phenyl)ethanol 5(i)





6j (Entry 10 in Table 1): (S)-1-(4-((3-methoxyphenyl)ethynyl)phenyl)ethanol



6k (Entry 11 in Table 1): (S)-4-((4-(1-hydroxyethyl)phenyl)ethynyl)benzonitrile



<u>61 (Entry 12 in Table 1):</u>

(S)-1-(4-((4-(trifluoromethyl)phenyl)ethynyl)phenyl)ethanol







6m (Entry 13 in Table 1): (S)-1-(3-(phenylethynyl)phenyl)ethanol





6n (Entry 14 in Table 1): (S)-1-(3-((4-fluorophenyl)ethynyl)phenyl)ethanol



_															_	
0.40																
L		0.30				Ę			HO >		8		숾			
	₹ 0.20-															
0.10- (±)																
																0
2.00 4.00 5.00 10.00 12.00 14.00 16.00 18.00 20.00 22.00 24.00 26.00 28.00 30.00 10.3837 分钟, 0.3257 AU 分钟																
	名称	保留时间 (分钟)	面积 (微伏*秒)	% 面积	高度 (微伏)	积分类型	含量	单位	峰类型	峰代码	结构1 名	结构 1 说明	结构1 分子量	结构1 公式	结构1 结构	
1		20.442	13105077	49.47	401115	BB			未知						0	
2		25.052	13385641	50.53	346810	BV			未知						0	
	-							_								6

60 (Entry 15 in Table 1): (S)-1-(3-((3-fluorophenyl)ethynyl)phenyl)ethanol 5(0)





6p (Entry 16 in Table 1): (S)-1-(3-((2-fluorophenyl)ethynyl)phenyl)ethanol




6q (Entry 17 in Table 1): (S)-1-(3-((4-chlorophenyl)ethynyl)phenyl)ethanol





6r (Entry 18 in Table 1): (S)-1-(3-((4-bromophenyl)ethynyl)phenyl)ethanol











6t (Entry 20 in Table 1): (S)-1-(3-(m-tolylethynyl)phenyl)ethanol









未知

19.815

30333504

99.16 1043712 BB



6v (Entry 22 in Table 1): (S)-1-(3-((3-methoxyphenyl)ethynyl)phenyl)ethanol

6w (Entry 23 in Table 1):

(S)-1-(3-((4-(trifluoromethyl)phenyl)ethynyl)phenyl)ethanol





6x (Entry 24 in Table 1):

(S)-1-(4-(hex-1-yn-1-yl)phenyl)ethanol





ID#	名称	保留时间	峰非	面积	高度	面积%	
1	RT12.443	12.443	1	62183092	1013690	98, 3400	
2	RT17.495	17.495	2	1049643	18591	1.6600	



6y (Scheme 2): (1S,1'S)-1,1'-(ethyne-1,2-diylbis(4,1-phenylene))diethanol





6z (Scheme 2): (S)-1-(3-((4-(trifluoromethyl)phenyl)ethynyl)phenyl)ethanol



Entry	1	2	3	4	5	6	7	8	9
Yield [%]	93	93	93	92	91	91	91	90	88
ee [%]	97	96	95	95	94	94	94	94	94

Table S1. Reusability of catalyst **5** for the enantioselective tandem Sonogashira coupling-ATH reaction of 4-iodoacetophenone and phenylacetylene.^[a]

[a] Reaction conditions: catalyst 5 (196.80 mg), K_2CO_3 (138.0 mg, 1.0 mmol), HCO_2Na (680.0 mg, 10.0 mmol), iodoacetophenones (1.0 mmol), aryne (1.10 mmol), and 40.0 mL of the mixed solvents ($H_2O/MeOH v/v = 1/3$), reaction temperature (60 °C), reaction time (16 h).

Figure 10. Reusability of catalyst **5** for the Sonogashira coupling-ATH reaction of 4-iodoacetophenone and phenylacetylene.

Recycle 2







Recycle 4



Recycle 5



Recycle 6







Recycle 8



Recycle 9

