

## Electronic Supporting Information

### **A novel scattering switch-on detection technique for target-induced plasmon-coupling based sensing by single-particle optical anisotropy imaging**

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#### **Experimental Section**

##### **Materials**

Hydrogen tetrachloroaurate (III) trihydrate ( $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$ ), sodium citrate, sodium chloride ( $\text{NaCl}$ ), dipotassium phosphate ( $\text{K}_2\text{HPO}_4$ ), monopotassium phosphate ( $\text{KH}_2\text{PO}_4$ ), hydroxylammonium chloride, glutathione (GSH, reduced), polyethylene glycol (PEG 400) were purchased from Sinopharm Chemical (Shanghai, China). Sodium hydrosulfide ( $\text{NaHS}$ ) was obtained from Sigma-Aldrich. All reagents were AR grade and used without further purification.

##### **Nanoprobe preparation and characterization**

The 18 nm AuNPs were synthesized according to the classical Frens method. The obtained 18 nm AuNPs were then used as seeds for the preparation of 50 nm AuNPs using a seed-mediated growth method. In brief, 1.432 mL 18 nm AuNPs and 255  $\mu\text{L}$  hydroxylammonium chloride (400 mM) were sequentially added into 18 mL DI water. After thoroughly mixed, 386  $\mu\text{L}$  of 24.28 mM  $\text{HAuCl}_4$  was gradually added into the mixture drop by drop in 30 min. The GSH@AuNPs were prepared by adding 5  $\mu\text{L}$  GSH (1 mM) to 1 mL freshly prepared AuNPs under stirring at 600 rpm for 15 min. The prepared AuNPs and GSH@AuNPs were characterized with UV-visible spectroscopy (Shimadzu, UV-1800, Japan) and

transmission electron microscopy (JEM 1230, JEOL, Japan). The hydrodynamic radius and surface charge of the nanoparticles before and after modification were measured using a Zetasizer Nano ZS instrument (Malvern, UK).

### **Imaging apparatus**

The imaging experiments were performed on a Nikon 80i upright microscope consisting of an oil immersion dark-field condenser (NA: 1.20~1.43) and a 40× objective (NA: 0.75). The original 100 W halogen tungsten lamp was replaced with a 1000 W professional studio flashlight (PN1000, JinBei Co., Shanghai, China), which features a flash duration time of 1/2000 s and a recharging time of less than 2 s. The polarizing optics is a DIC rotatable polarizer (Nikon, Japan) consists of a rotatable polarizer before the condenser and an analyzer before the detector. The detectors used in our experiment include a Neo sCMOS Camera (Andor, UK) and an Olympus DP72 color CCD camera (Japan).

### **Concept verification for H<sub>2</sub>S sensing by colorimetric assay**

Typically, the concentration of NaCl solution for H<sub>2</sub>S detection was optimized by testing the stability of the probes in salt solutions with various concentrations. By incubating the probes with a set of NaCl solutions with concentration ranging from 0 to 128 mM for 15 min, the nanoprobe were characterized with UV-visible spectroscopy. We also tested the effect of solution pH value (from 5 to 9) on probe stability. The detection time was selected according to reaction dynamics by monitoring the absorbance spectra of GSH@AuNPs incubated with 8 μM of NaHS. For H<sub>2</sub>S sensing in bulk solution, the concentration of NaHS ranged from 2 μM, 4 μM, to 20 μM. To further confirm the aggregation of nanoparticles induced by H<sub>2</sub>S addition, the nanoprobe with or without the incubation of 10 nM H<sub>2</sub>S were characterized with FLPDM and TEM.

### **H<sub>2</sub>S sensing with FLPDM**

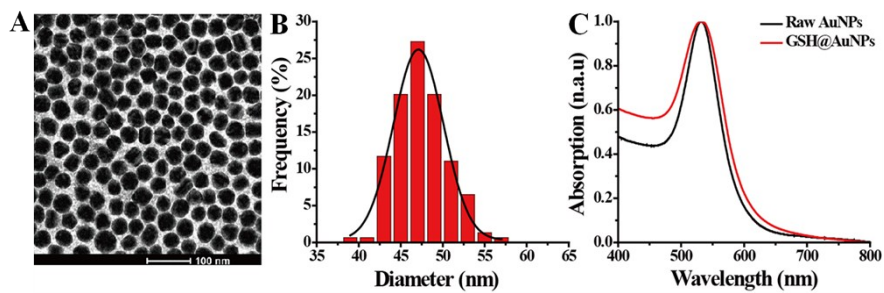
For H<sub>2</sub>S detection under FLPDM, the coverslips were carefully cleaned by using the piranha solution (H<sub>2</sub>SO<sub>4</sub>: H<sub>2</sub>O<sub>2</sub> = 3:1) to remove organic residues, followed by sonication in ultrapure water for at least three times to remove excess dusts. Then the coverslips were incubated with 1 wt% PEG 400 for 30 min to prevent the nonspecific absorption of nanoparticles during the detection experiments. To investigate the selectivity, the following anions and thiols including Cl<sup>-</sup>, Br<sup>-</sup>, I<sup>-</sup>, CO<sub>3</sub><sup>2-</sup>, NO<sub>3</sub><sup>-</sup>, S<sub>2</sub>O<sub>3</sub><sup>2-</sup>, SO<sub>4</sub><sup>2-</sup> and cysteine with a same concentration of 10 μM were incubated with GSH@AuNPs and followed by the particle counting with FLPDM. For the quantitative detection of H<sub>2</sub>S, the probe solution was incubated with NaHS

with final concentrations ranging from 0.1 nM to 100  $\mu$ M. After reaction, 10  $\mu$ L diluted sample was placed between two coverslips and detected under FLPDM. The images were processed with ImageJ software and the nanoparticle counts were obtained by using the particle analysis module.

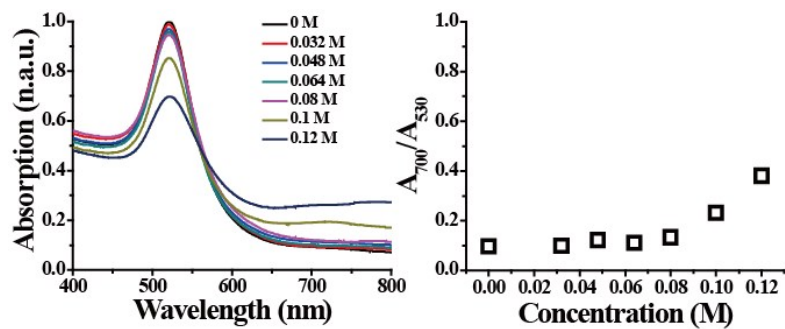
### Additional Tables and Figures

	Surface charge (mV)	Hydrodynamic diameters (nm)
Au NPs	-18.6	53.1
GSH-Au NPs	-28.0	63.3

**Table S1.** Surface charge and hydrodynamic size changes of the AuNPs before and after surface modification.



**Fig. S1.** Characterizations of the prepared GSH@AuNPs. Representative TEM image (A) and the size distribution (B) of the synthesized AuNPs. (C) Absorption spectra of raw AuNPs and GSH modified AuNPs.



**Fig. S2.** Optimization of NaCl concentration used in the detection of H<sub>2</sub>S. The GSH@AuNPs display well stability at a concentration of NaCl lower than 0.1 M.

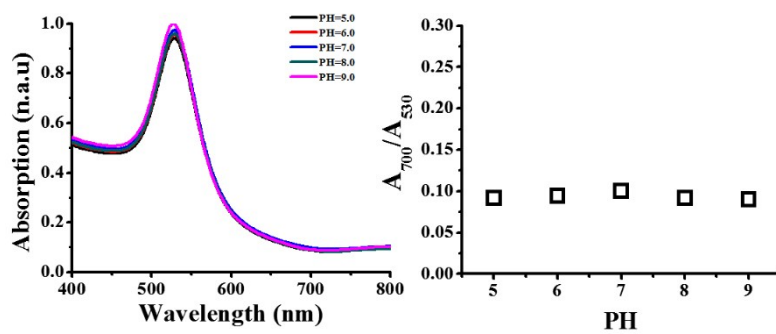
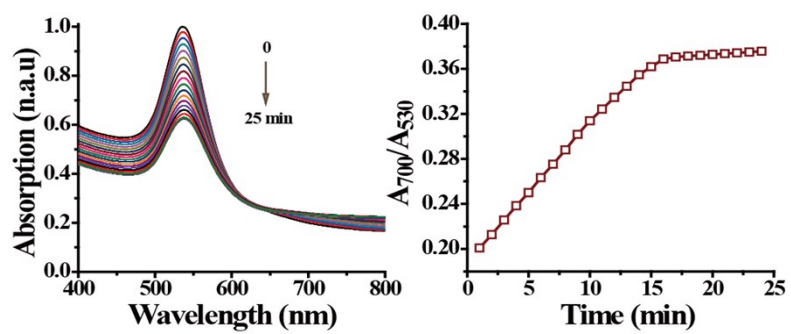
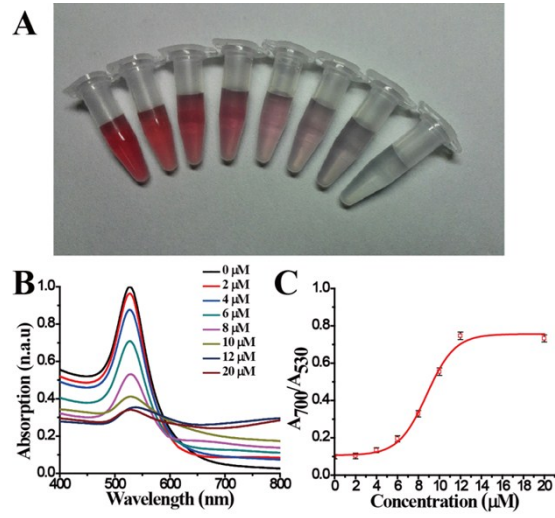


Fig. S3. The GSH@AuNPs display well stability in buffer solutions with pH values ranging from 5 to 9.

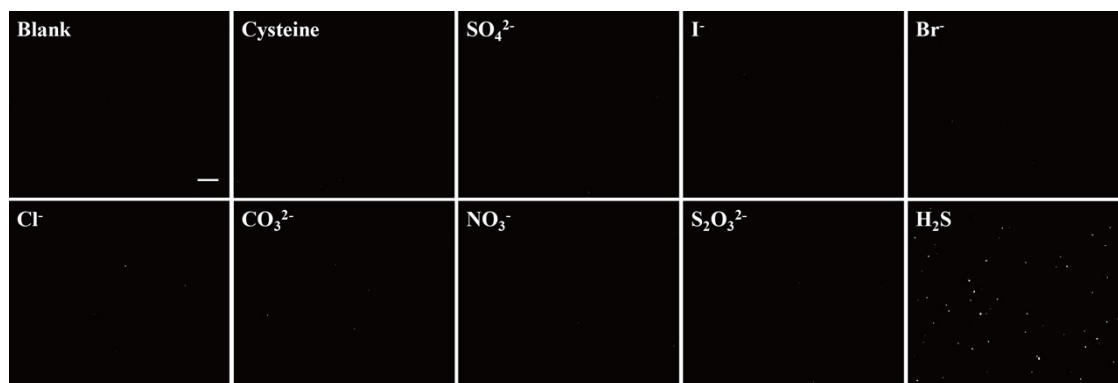


**Fig. S4.** Time course of absorption spectrum and relative absorption ratio changes of the GSH@AuNPs with the addition of 8  $\mu\text{M}$  of NaHS.

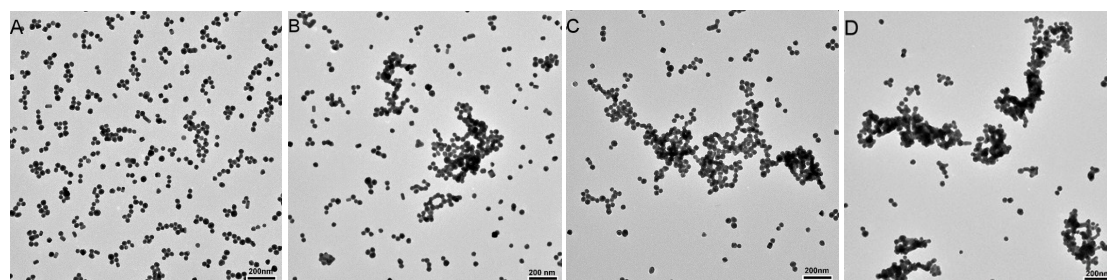




**Fig. S5.** Detection of  $\text{H}_2\text{S}$  with GSH@AuNPs as probes in bulk solutions. Digital photo (A), absorption spectra (B) and relative absorption ratio (C) of the GSH@AuNPs in the presence of  $\text{H}_2\text{S}$  at different concentrations from 0 to 20  $\mu\text{M}$ .



**Fig. S6.** Typical FLPDM images for studying the selectivity of GSH@AuNPs towards H<sub>2</sub>S. The scale bar is 20  $\mu$ m.



**Fig. S7.** Representative TEM images of GSH@AuNPs with the addition of H<sub>2</sub>S at various concentrations. (A) 0.1 nM, (B) 10 nM, (C) 1  $\mu$ M, (D) 100  $\mu$ M.

Table S2. Comparison of this work with other available methods for particle-aggregation based sensing strategies.

Method	Principle	Background interference	Sensitivity	Ref.
Latex agglutination test	Latex agglutination	low	low	1
Colorimetric assays	Aggregation induced color change	high	middle	2
Dynamic light scattering	Aggregation induced size change	middle	high	3
Single particle counting	Aggregation induced number change	high	high	4
Scattering switch on	Aggregation induced symmetry change	free	high	This study

Reference.

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