Supporting Information

Sweet Supramolecular Elastomers from α,ω-(β-Cyclodextrin terminated) PDMS

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Experimental Section

Materials

Octamethylcyclotetrasiloxane (D₄, 98%), and 1,3-bis(4-hydroxybutyl)tetramethyldisiloxane (95%) were obtained from Gelest. Propiolic acid (95%), *N*-(3-dimethylaminopropyl)-*N*'-ethyl-carbodiimide hydrochloride (EDC-HCl, commercial grade), 1,1,3,3-tetramethylguanidine (TMG, 99%) and dimethylaminopyridine (DMAP, 99%) were obtained from Sigma-Aldrich. Trifluoroacetic acid (analytical grade) (TFA) was obtained from Carlo Erba Reagents. Mono-6-deoxy-6-azido- β -cyclodextrin 1 was synthesized according to a published procedure,¹ using materials purchased from Sigma Aldrich. ESI-MS and FTIR spectra are given in Figures S1 and S2, respectively. Solvents DMSO-*d*₆, CDCl₃, THF, isopropanol, CH₂Cl₂ (DCM), and toluene were all obtained from Sigma Aldrich and used as received.

Characterization

¹H NMR spectra were recorded at room temperature on a Bruker AC-250 spectrometer (at 250 MHz for ¹H), a Bruker AVANCE 500MHz Spectrometer (500 MHz for ¹H and 125 MHz for ¹³C) or a Bruker AVANCE II spectrometer (at 400 MHz for ¹H and 100.6 MHz for ¹³C) using deuterated solvents (CDCl₃ or DMSO- d_6). ¹³C chemical shifts are reported with respect to CDCl₃ as an internal standard, set at 77.23 ppm. ¹H NMR chemical shifts are reported with respect to chloroform (CHCl₃) as an internal standard, set at 7.26 ppm or with respect to residual proton signal of DMSO- d_6 set at 2.50 ppm. Coupling constants (*J*) are recorded in Hertz (Hz). The abbreviations s=singlet, d=doublet, t=triplet, m=multiplet, are used to report spectra.

DOSY experiments were performed on a Bruker Avance III 400 spectrometer equipped with a 5 mm multinuclear broad band probe (BBFO+) with z-gradient coil. A double stimulated echo sequence (DSTE) incorporating bipolar gradient pulses was used for the measurement of self-diffusion coefficients. Gradients were incremented from 0.96 G cm⁻¹ to 47.19 G cm⁻¹ in 40 steps using an exponential ramp. 64 scans were acquired with 16k data points. A gradient pulse length $\delta/2$ of 1.5 ms was used with a

diffusion delay Δ set to 100 ms. Fourier transformation was applied in F2 with 2 Hz exponential broadening. The diffusion dimension of the 2D DOSY spectra was processed with TOPSPIN DOSY software.

Size exclusion chromatography (SEC) in THF eluent was measured on a Shimadzu apparatus equipped with a CTO-20A oven, set at 35°C, a RID 10A refractive index detector, and a Viscotek 270 Dual detector (viscometer and light scattering). A mixture of TFA (0.342 g, 0.33 mmol) and TMG (0.115 g, 0.11 mmol) was first weighed into a vial with 1 mL of THF and mixed vigorously. The rubber (15 mg) was then placed in a vial with 3 drops of the chaotropic salt solution and 4 mL of THF solvent and left for 24 h, at which time it was found to have dissolved.

Infrared spectra were measured on a Nicolet iS10 Thermo Scientific, using the ATR mode, at room temperature. Differential scanning calorimetry and thermogravimetric analyses were measured on TA Instruments Q500 and Q20, respectively. Rheological measurements were taken on an ARES SN Rheometer from TA Instruments. All specimens were tested as 1-mm thick disks, under a nitrogen atmosphere, using 25 mm parallel plate geometry. Oscillatory sweep measurements were taken as a function of angular frequency in the range 0.1-100 rad/s, at 3% strain, whereas temperature sweeps measurements were done between 30 and 150 °C, at constant frequency and strain (1 Hz, 3%).

Transmission electron microscopy was measured either on Philips CM120 TEM (Lyon) or on a Titan 1 FEI Titan 80-300 Cubed (Hamilton). Thin slices (~60 nm) of the elastomers were prepared by microtoming. Ruthenium tetraoxide vapor (popular for heterogeneous polymer systems) was used as a stain, as it has been shown to enhance electron density contrast for polymers containing ethers, alcohols and aromatics.² Given the nature of cyclodextrin and the fact that it is connected to the PDMS through an aromatic linkage, cyclodextrin rich areas in the material were expected to provide greater contrast.

General Syntheses

Synthesis of propiolate-terminated polydimethylsiloxane 2, 3

Two 1,3-bis(propiolatobutyl)-terminated polydimethylsiloxanes **2** and **3** (see Figure 1 in main text), which differ in molar mass, were synthesized following the procedure by Rambarran et al.³ The polysiloxane chain **2** was constituted from ~52 dimethylsiloxane units, corresponding to an average molar mass of 4,100 g·mol⁻¹ as determined by ¹H NMR and confirmed by GPC (THF: M_n = 4,100 g·mol⁻¹; PDI = 2.05.). Polysiloxane **3** has ~114 dimethylsiloxane units, corresponding to an average molar mass of 8,600 g·mol⁻¹ as determined by ¹H NMR and confirmed by GPC (THF: M_n = 7,800 g·mol⁻¹; PDI = 1.95).

Compound **2**: ¹H NMR (CDCl₃, 500 MHz, δ): 0.07 (s, 312H, SiC*H*₃); 0.52 (t, 4H, *J*=8.3 Hz, SiC*H*₂CH₂CH₂CH₂CH₂); 1.38 (m, 4H, SiCH₂CH₂CH₂CH₂); 1.69 (m, 4H, SiCH₂CH₂CH₂CH₂); 2.87 (s, 2H, C=C*H*); 4.19 (t, 4H, *J*=6.5 Hz, SiCH₂CH₂CH₂CH₂). See FTIR and ¹H NMR spectra in Figures S2 and S3, respectively.

Compound **3**: ¹H NMR (CDCl₃, 500 MHz, δ): 0.07 (s, 684H, SiC*H*₃); 0.52 (t, 4H, *J*=8.3 Hz, SiC*H*₂CH₂CH₂CH₂); 1.38 (m, 4H, SiCH₂CH₂CH₂CH₂); 1.69 (m, 4H, SiCH₂CH₂CH₂); 2.87 (s, 2H, C=C*H*); 4.20 (t, 4H, *J*=6.5 Hz, SiCH₂CH₂CH₂CH₂).

Synthesis of β -CD functional PDMSs 4, 5

Compounds 1 (0.17 g, 0.15 mmol per azide) and 2 (0.31g, 0.075 mmol, 0.15 mmol per alkyne) were mixed in a glass vial with 8 mL of 1:1 water: isopropanol at 80 °C. The reaction was initially turbid. After 5 d, the solution became transparent and a monolithic yellow elastomer 4 (0.40 g, 82% of the original starting mass) precipitated out of the solution. 4 could be swollen in dichloromethane after a few hours but did not dissolve. Partially reacted PDMS remaining in solution was removed simply by decanting. Any material not tightly bound within the network could be extracted with DCM, reducing the mass of the rubber to 0.21 g, 43% of the original mass of the reagents. The un-extracted material was characterized by IR, ¹H NMR (HRMAS), SEC (THF eluent), TGA, DSC, TEM and rheology.

This process was repeated starting from **3** (0.32 g 0.037 mmol, 0.074 mmol per alkyne) to create the elastomer **5** (0.28 g, 70% of the original mass of the reagents) taking 5 days to reach completion. After extraction with DCM, the mass was reduced to 0.25 g, 63% of the original mass of the reagents.

Compound **4**: ¹H NMR (CDCl₃, 400 MHz, δ): 0.07 (s, 342 H, SiC*H*₃); 0.62 (m, 4H, SiC*H*₂CH₂CH₂CH₂CH₂); 1.49 (m, 4H, SiCH₂CH₂CH₂CH₂); 1.83 (m, 4H, SiCH₂CH₂CH₂CH₂); 2.86 (s, 0.6H, C=C*H*); 4.37 (m, 4H, SiCH₂CH₂CH₂CH₂CH₂), 4.66 (broad s, H₂O), 8.18, 8.83 (s, 1.4H, *C*-*H* aromatic). Note that some of the silicone polymers reacted at only one end, such that residual SiCH₂CH₂CH₂CH₂CH₂ groups were present, with additional signals at 1.37, 1.69 and 4.23 ppm. SEC (after chaotropic salt treatment): M_n=28,700 g/mol, PDI=2.80.

Compound **5**: ¹H NMR (CDCl₃, 400 MHz, δ): 0.07 (s, 654 H, SiC*H*₃); 0.62 (m, 4H, SiC*H*₂CH₂CH₂CH₂CH₂); 1.49 (m, 4H, SiCH₂CH₂CH₂CH₂); 1.83 (m, 4H, SiCH₂CH₂CH₂CH₂); 2.86 (s, 0.7H, C=C*H*); 4.37 (t, 4H, SiCH₂CH₂CH₂CH₂), 4.66 (broad s, H₂O), 8.18, 8.83 (s, 1.3H, *C*-*H* aromatic). As noted above, peaks from residual SiCH₂CH₂CH₂CH₂CH₂ groups were present at 1.37, 1.69 and 4.23 ppm. SEC (after chaotropic salt treatment): M_n=30,300 g/mol, PDI=2.60.

Click reaction with copper catalyst

1 (0.114 g, 56 μ mol), 2 (0.059 g, 51 μ mol), copper sulfate pentahydrate (99.99%, Sigma-Aldrich, 0.003g 12 μ mol) and sodium ascorbate (>98%, Sigma, 0.006 g, 28 μ mol) were mixed with 3mL of 1:1 H₂O:isopropanol. The cloudy dispersion was stirred at room temperature. After 45 min, the solution turned transparent and small drops of gels formed and stuck to the sides of the glass. The final elastomer was comparable to the thermally prepared counterpart in terms of its supramolecular state (it dissolves in THF/chaotropic salt solution, but only swells, and does not dissolve in good solvents for silicones).

Reaction of 2 with 3-azido-1-propanol

3-Azido-1-propanol was synthesized according to a previously described procedure. ⁴ Propiolateterminated PDMS (0.44 g, 0.21 mmol of alkyne) and 3-azido-1-propanol (0.09 g, 0.29 mmol) were added to a scintillation vial with 7 mL of 1:1 H₂O:IPA and left to stir at 80 °C for 5 days at which time a viscous oil was observed that was comprised of triazole-modified silicone, as confirmed by ¹H NMR.

¹H NMR (CDCl₃, 250 MHz, δ): 0.05 (s, 320H, SiC*H*₃); 0.59 (t, 4H, *J*=8.3 Hz, SiC*H*₂CH₂CH₂CH₂CH₂); 1.44 (m, 4H, SiCH₂CH₂CH₂CH₂); 1.81 (m, 4H, HOCH₂CH₂CH₂); 2.15 (m, 4H, SiCH₂CH₂CH₂CH₂); 3,65 (m, 4H, HOCH₂CH₂CH₂); 4.33 (t, 4H, *J*=6.5 Hz, SiCH₂CH₂CH₂CH₂); 4.58 (t, 4H, *J*=6.9 Hz, HOCH₂CH₂CH₂CH₂); 8.10, 8.11 (s, 2H, CH aromatic).

Synthesis of mono-(6-deoxy-6-azido)-peracylated- β -cyclodextrin 6

Mono-6-deoxy-6-azido- β -cyclodextrin was synthesized according to the previously published procedure: ⁵ an adaptation of the procedure by Noomen et al. ⁶ was used to synthesize the azido peracylated- β -cyclodextrin. Azido- β -cyclodextrin **1** (0.45 g, 0.38 mmol), DMAP (cat, 3.50 mg), acetic anhydride (3 ml ~3.24 g, 31.74 mmol) and pyridine (5 mL) were added to a 20 mL scintillation vial, sealed and stirred at 80 °C for 96 h (the mixture turned transparent upon mixing and heating). The reaction was removed from heat and allowed to cool to room temperature. Upon cooling in an ice bath, no product precipitated. The mixture was precipitated in 100 mL of cold water, vacuum filtered and allowed to dry on the filter paper and placed in a vacuum desiccator for 48 h, at which time 0.55 g of product (73% yield) was recovered.

¹H NMR (600 MHz, see figure for labeling): 2.10 (63 H, CH₃CO); 3.72 (m, 9H, H₄, H₆,^a, H₆,^b) 4.13-4.58 (19H, H₅, H₆^a, H₆); 4.81 (m, 7H, H₃); 5.09 (m, 7H, H₁). LRMS (ESI positive): m/z [M⁺ :(NH₄)] calculated = 2017.6, found = 2017.7, m/z [M⁺⁺:(NH₄)(K⁺)] calculated = 1028.3, found = 1028.4.



Acylated-β-Cyclodextrin-functional PDMS **7** (*Thermal Cycloaddition*)

6 (0.41 g, 0.20 mmol) and **2** (0.40 g, 0.20 mmol of alkynyl groups) were weighed into a 10 mL roundbottomed flask fit with a condenser. The reagents were homogenized with 1 mL of toluene + 1 mL of CHCl₃ and stirred at 90 °C. The initial reaction mixture was colorless and transparent. As the reaction progressed, the mixture turned to yellow and finally to a light brown. At 144 h, the reaction was found to be complete by proton NMR (75% conversion) as demonstrated by the disappearance of the C-H signal from the propiolate (2.87 ppm), the appearance of the C-H aromatic signals from triazole of the product (8.15 - 8.24 ppm) and the splitting pattern of the CH₂ α Si. Two signals were observed, one corresponding to the product integrating to 1.5 and one corresponding to the starting material integrating to 0.5. The reaction was stopped at this time to mirror the reaction progress with the non-acylated cyclodextrin. Upon removal of the solvent, the product was transparent brown solid of mass 0.74 g (91% yield). The product could be dissolved in various organic solvents.

¹H NMR (600 MHz): 0.07 (s, 288 H, SiCH₃); 0.58 (t, 4H, J=9 Hz, SiCH₂CH₂CH₂CH₂CH₂); 1.45 (m, 4H, SiCH₂CH₂CH₂CH₂CH₂); 1.81 (m, 4H, SiCH₂CH₂CH₂CH₂) 2.10 (126 H, CH₃CO); 3.72 (m, 18H, H₄, H₆, ^a, H₆, ^b) 4.13-4.58 (42H, H₅, H₆, ^a, H₆, SiCH₂CH₂CH₂CH₂CH₂); 4.81 (m, 14H, H₃); 5.09 (m, 14H, H₁); 8.05, 8.16 (s, 2H, C-*H aromatic*). Note that some of the silicone polymers reacted at only one end, such that residual SiCH₂CH₂CH₂CH₂CH₂OCOCH groups were present with additional signals at 0.53, 1.38, 1.68 and 2.87 ppm along with a small amount of unreacted compound **6** whose signals overlapped with the product.

Characterization of synthons



Figure S1. Electrospray-mass spectrum of N_3 - β -CD protonated by Na^+ (main peak at 1183.4 g/mol).



Figure S2. FTIR spectra of starting material: (top) N_3 - β -CD 1; (bottom) Propiolate-PDMS 2.



Figure S3. ¹H NMR spectra of propiolate-PDMS 2 in CDCl₃:



Figuree S4. TGA of N_3 - β -CD 1 and and propiolate-PDMS 2.

Characterization of α, ω - (β -CD functionalized) -PDMS



Figure S5. Zoom on the FTIR region of OH stretch for propriolate PDMS 2 (top) and elastomer 4 (bottom).



Figure S6. ¹H HRMAS spectrum of the elastomer 5.



Figure S7. DSC cycles of the elastomer 4.



Figure S8: TGA of 4 (thin line) and 5 (thick line) elastomers.

Rheology study



Figure S9. Frequency sweeps at constant strain for propriolate-PDMSs 2 (thick line) and 3 (thin line).



Figure S10. Temperature sweeps at constant strain for elastomer 4.



Figure S11. Complex viscosity versus angular frequency for elastomers 4 (thick line) and 5 (thin line).

Microscopy study



Figure S12: SEM images on a conventional TEM (a,b,e,f) and High Resolution TEM (c,d,g,h) of elastomers 4 (upper row) and 5 (bottom row).

Complementary analyses



Figure S13. ¹H NMR spectrum in CDCl₃ of acetylated β -CD **6**.



Figure S14: ¹³C NMR spectrum in CDCl₃ of acetylated β -CD 6.



Figure S15: SEC analyses in THF of the original propriolate polymers **2**, **3** and final elastomers **4** and **5** after the click reaction with **1**, and solubilization in a chaotropic salt/THF solution. The fact that the molar masses of the complexes show an increased mass suggests that some aggregation is occurring owing to the poor THF solubility of the CD end chains.

DOSY measurements and interpretation

A specific study on the interactions between acetylated cyclodextrin **6** and PDMS propriolate **2** was first carried out to check whether the cyclodextrin cage could accommodate alkyne groups or PDMS chains in its core. The mixture was first prepared in CDCl₃, introducing 26.6 mg of **6** and 26.5 mg of **2** in 1.15 g of deuterated solvent (1/1 molar ratio alkyne/CD). First comparison of the ¹H NMR spectra revealed no shifting of the protons inside the cavities, as expected for a complexed cage (signals H3 and H5, see ref. 7, for instance). The proton of the acetylene function around 3 ppm is slightly shifted, but this is due to its inherent lability (also observed with dilution). DOSY analysis was done on this mixture and compared to the polymer **7**, obtained by click reaction between **6** and **2**. 2D DOSY maps are shown in Figure S16. In the mixture, the diffusion coefficients of **6** and terminal groups of **2** are close (380 vs. 290 $\mu m^2/s$, respectively). When considering the *D* (Me₂SiO) units of the backbone, the diffusion coefficient is slightly different (190 $\mu m^2/s$, not shown), in better agreement with the molar mass differences (2007 and 4100 g/mol for **6** and **2**, respectively). Such variation of *D* coefficients of both entities are clearly separated, it is concluded that under these conditions, no tight bounding between PDMS and CD through an inclusion complex was observed.

In the case of polymer 7, all the signals of PDMS and β -CD line to a smaller value of D corresponding to the polymer ($D = 185 \ \mu m^2/s$). Viscosity effects were excluded as the diffusion coefficient of CHCl₃ was found constant for both solutions. The ratio of D coefficients of free versus clicked β -CD can be expressed as:

$$D_{free}/D_{clicked} \approx (M_{free}/M_{clicked})^{-lpha}$$

where M_i are the average molar mass of the considered products, and $\alpha = 1/dF$ with dF the fractal dimension of the products.⁹ Assuming that the fractal dimension of both free and clicked β -CD is the same and estimated at 2 for a polymer in a theta solvent (α value of 0.5), very close to the value reported for simple β -CD at 0.49.¹⁰ With a calculated D_{cliked}/D_{free} ratio of 0.49, one finds a $M_{free}/M_{clicked}$ ratio of 0.24, which corresponds to a polymer of one PDMS chain coupled with two β -CD molecules. One can see also traces of free PDMS and free CD signals.



Figure S16: 2D DOSY maps of (left) the acetylated β -CD **6** and the propiolate PDMS **2** mixed together in CDCl₃ and (right) the polymer **7** from click reaction in CDCl₃.

References for supporting information

- 1. R. C. Petter, J. S. Salek, C. T. Sikorski, G. Kumaravel, F. T. Lin J. Am. Chem. Soc. 1990, 112, 3860.
- 2. J. S. Trent, J. I. Scheinbeim, P. R. Couchman Macromolecules 1983, 16, 589.
- 3. T. Rambarran, F. Gonzaga, M. A. Brook *Macromolecules* 2012, 45, 2276.
- 4. S. El Habnouni , V. Darcos , X. Garric , J.-P. Lavigne , B. Nottelet, J. Couda *Adv. Funct. Mater.* 2011, **21**, 3321.
- 5. W. Tang, S. C. Ng, *Nature Protocols* 2008, 3, 691.
- 6. A. Noomen, A. Penciu, S. Hbaieb, H. Parrot-Lopez et al. Mat. Sci. Eng. C 2008, 28, 705.

7. A. Calderini, F. B. T. Pessine, M. H. Martins, *J. Incl. Phenom. Macrocycl. Chem.* 2013, **75**, 77; R. Ferrazza, B. Rossi, G. Guella, *J. Phys. Chem. B* 2014, **118**, 7147.

8. J. Viéville, M. Tanty, M.-A. Delsuc, *J. Magnet. Res.* 2011, **212**, 169. See also a recent paper showing the interest of using PDMS in DOSY experiments: S. Huan, J. Gao, R. Wu, S. Li, Z. Bai, *Angew. Chem. Int. Ed.* 2014, **53**, 11592.

S. Augé, P.-O. Schmit, C. A. Crutchfield, M. T. Islam et al. *J. Phys. Chem. B* 2009, **113**, 1914.
S. Viel, D. Capitani, L. Mannina, A. Segre, *Biomacromolecules* 2003, **4**, 1843.