

Electronic Supplementary Information (ESI)

Redox-responsive Host-guest System Using Redox-active Pillar[5]arene Containing One Benzoquinone Unit

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Experimental section

Materials. All solvents and reagents were used as supplied. Per-ethylated pillar[5]arene (**H1**), **G**, quinone-**H2** and 2OH-**H2** were synthesized according to the previous papers.^{S1-S3}

Measurements. The ¹H NMR spectra were recorded at 500 MHz and ¹³C NMR spectra were recorded at 125 MHz with a JEOL-ECA500 spectrometer.

Determination of association constants.

In quinone-**H2** ⊃ **G** complex in CDCl₃, chemical exchange between uncomplexed and complexed species was fast on an NMR timescale. Thus, NMR titrations were done with solutions which had a constant concentration of **G** (2 mM) and varying concentrations of quinone-**H2**. By the non-linear curve-fitting method,^{S4} the association constant *K* for the host-guest complex between quinone-**H2** and **G** was found to be $K = 28 \pm 1.1 \text{ M}^{-1}$.

In 2OH-**H2** ⊃ **G** complex in CDCl₃, chemical exchange between complexed and uncomplexed species was slow on an NMR timescale. Thus, ¹H NMR spectra of mixtures of 2OH-**H2** and **G** in different ratio showed two sets of resonances for complexed and uncomplexed **G**. The association constant for 2OH-**H2** ⊃ **G** complex was $(4.6 \pm 1.4) \times 10^2 \text{ M}^{-1}$, calculated from integrations of complexed (Fig. 2b, pink peak a') and uncomplexed signals (Fig. 2b, pink peak b) of **G**.

Determination of rate constants of the host-guest exchange. The exchange rate constant (*k*) at a coalescence temperature was estimated by using the approximate expression^{S5,6}:

$$\Delta G^\ddagger = 8.314T_c[22.96 + \log(T_c/\delta\nu)]$$

where ΔG^\ddagger is the free energy of activation for the exchange, $\delta\nu$ is the chemical shift difference between the proton signals from complexed and uncomplexed proton protons (Fig. S6, signals g and g'). The coalescence temperature T_c was estimated on the coalescence signal in these proton signals. The extrapolated value of *k*, which represent rate constant, at 25 °C was obtained from the Eyring equation:

$$k = k_B T / h \exp(-\Delta G^\ddagger / RT)$$

where k_B is the Boltzmann constant, T is the absolute temperature, h is Plank's constant, ΔG^\ddagger is the free energy of activation, and R is the gas constant.

$$k = k_{in} + k_{out} \quad K = k_{in} / k_{out}$$

In 2OH-**H2**⊃**G** complex, k , k_{in} and k_{out} (25 °C) were 190 s⁻¹, 189 s⁻¹ and 1 s⁻¹, respectively.

Reversible redox reaction of H2⊃G complex by alternating addition of oxidant and reductant. To the mixture of 2OH-**H2** (2.5 mM) and **G** (2.5 mM) in a mixture of CDCl₃ and CD₃OD (CDCl₃ : CD₃OD = 9:1), oxidant, tetrabutylammonium periodate (2 equiv. to 2OH-**H3**, 5.0 mM) was added. After 72 h, we checked the complexation by ¹H NMR (Fig 4b). To the mixture, reductant, tetrabutylammonium tetrahydroborate (2.5 mM) was added. After 72 h, we checked the complexation by ¹H NMR (Fig 4c). This redox-switching process was repeated by alternating addition of the oxidant and reductant.

Job plot for a mixture of G and quinone-H2

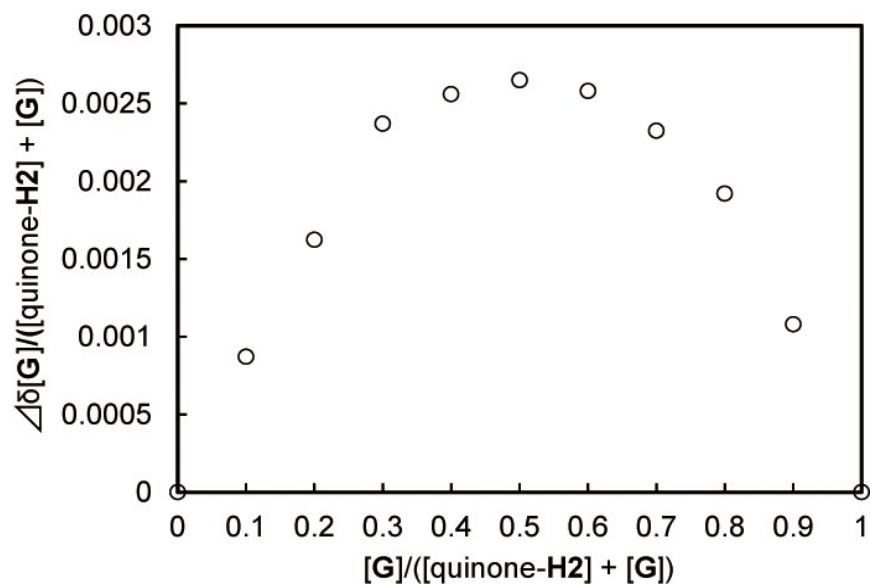
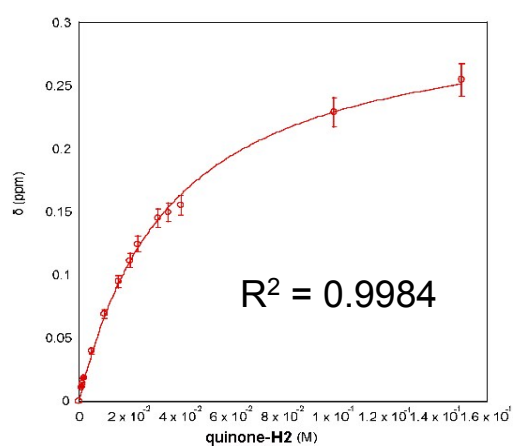
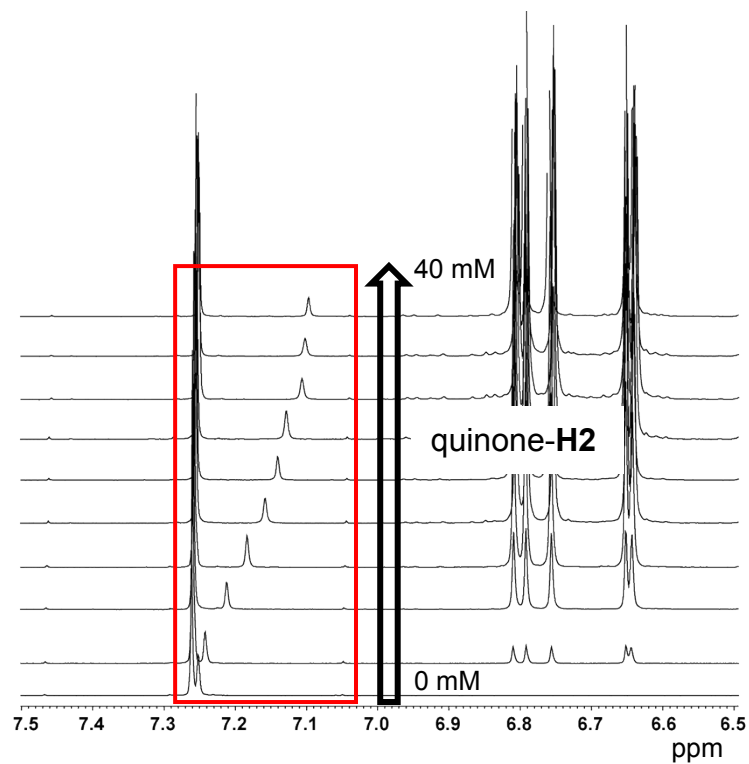


Fig. S1 Job plot between **G** (guest) and quinone-**H2** (host) was collected by plotting the $\Delta\delta$ in chemical shift of the triazole proton signal (Fig. 2d, yellow peak c). Concentration: $[G] + [quinone-H2] = 1$ mM. The plot indicates a 1:1 binding between the host and guest.

^1H NMR titration of G with quinone-H2 in CDCl_3



$$K_a = (28 \pm 1.1) \text{ M}^{-1} (25^\circ\text{C})$$

Fig. S2 ^1H NMR titration of G (Fig. 2d, proton peak c) with quinone-H2 in CDCl_3 at 25 $^\circ\text{C}$.

Variable temperature ^1H NMR spectra of a mixture of quinone-H2 and G in CDCl_3

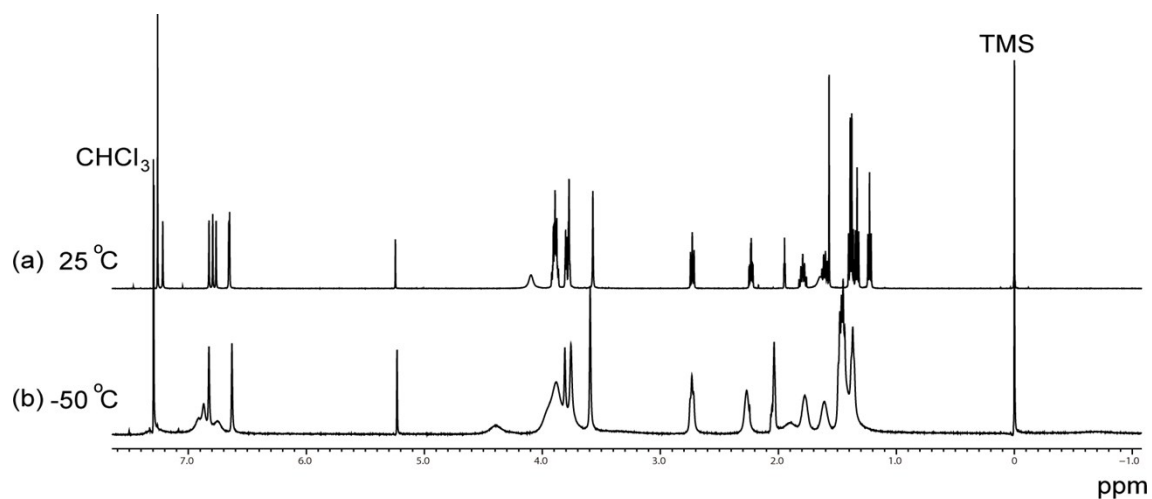


Fig. S3 Variable temperature ^1H NMR spectra (5 mM) of a mixture of quinone-H2 and G in CDCl_3 . The averaged proton signals were observed at $-50\text{ }^\circ\text{C}$, while the signals were largely broadening, indicating that the exchange was still fast on the NMR timescale even at $-50\text{ }^\circ\text{C}$.

Job plot for a mixture of G and 2OH-H2

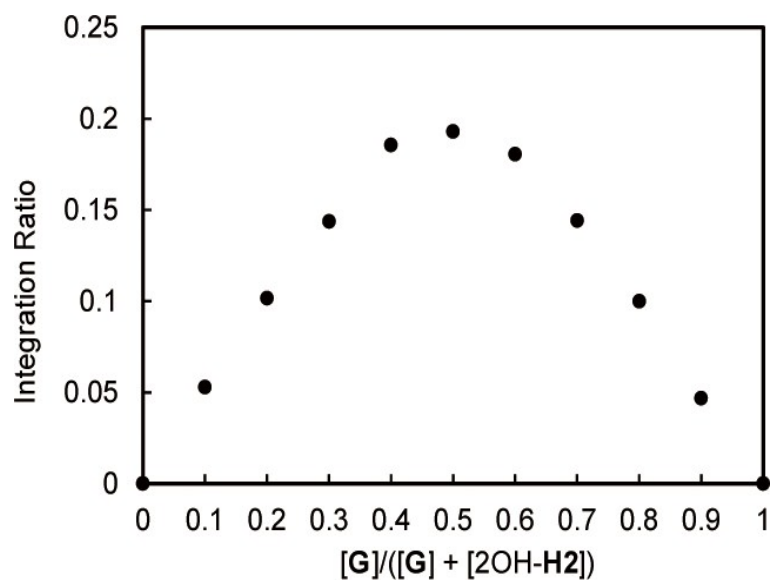


Fig. S4 Job plot between **G** (guest) and **2OH-H2** (host). The job plot was conducted by varying the mole fractions of the guest and host. Integration ratios between uncomplexed and complexed proton signals (Fig. 2b, pink peaks b and a') were utilized. Concentration: $[G] + [2OH-H2] = 1$ mM. The plot indicates a 1:1 binding between the host and guest.

Van't Hoff plot of a mixture of G and 2OH-H2 in CDCl₃

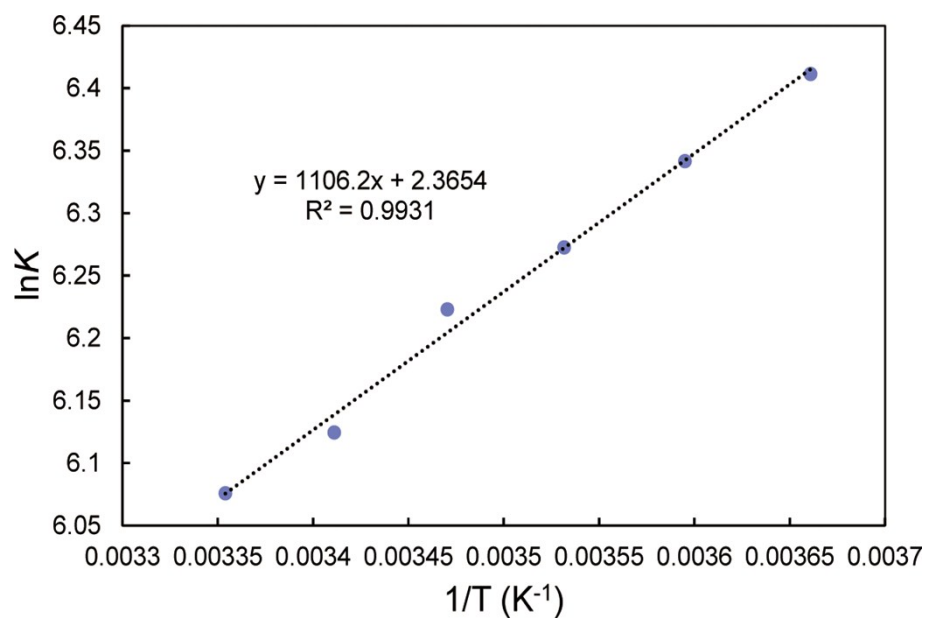


Fig. S5 van't Hoff plots for a mixture of **G** and **2OH-H2**. From the van't Hoff plots, ΔH , ΔS and ΔG_{298} were calculated to be -9.20 kJmol^{-1} , $19.7 \text{ JK}^{-1}\text{mol}^{-1}$, and -14.6 kJmol^{-1} , respectively.

Variable temperature ^1H NMR spectra of a mixture of G and 2OH-H2 in CDCl_3

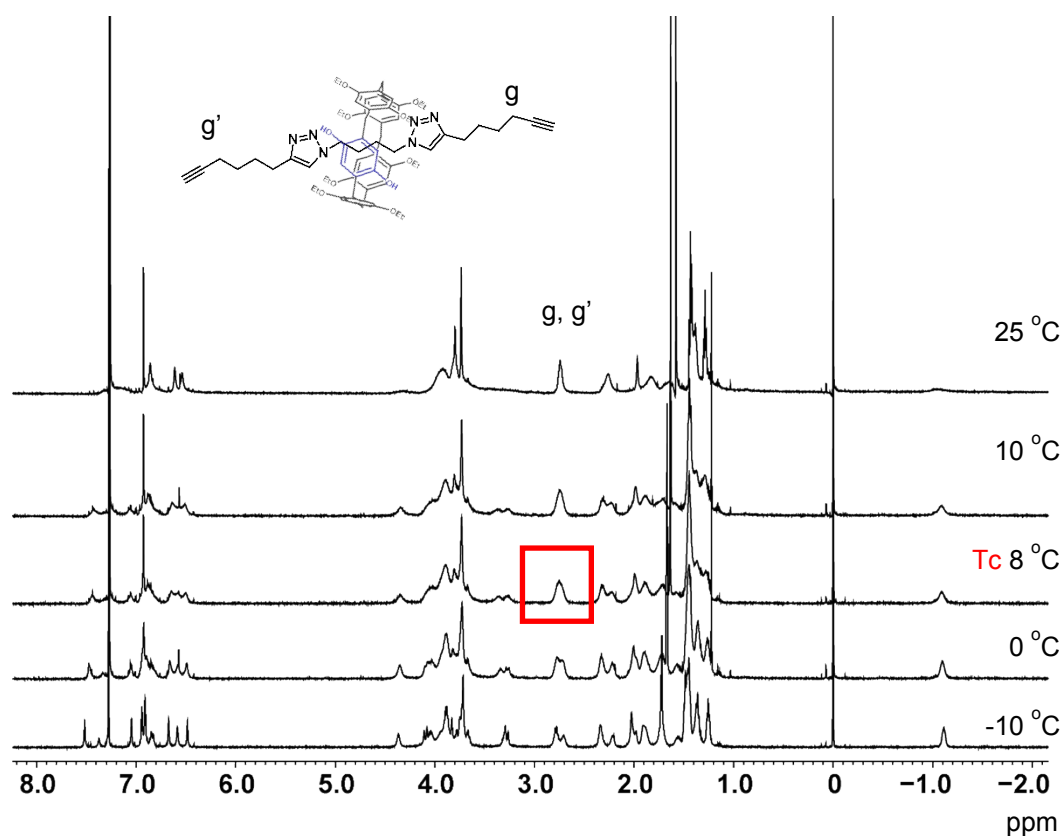


Fig. S6 Variable-temperature ^1H NMR spectra (CDCl_3) of a mixture of G and 2OH-H2 in CDCl_3 . The coalescence of the sets of the signals of from G was observed at 8 °C. From the coalescence temperature ($T_c = 8\text{ °C}$), the rate constant (k) at 298 K was calculated to be $1.9 \times 10^2\text{ s}^{-1}$.

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