

Supporting Informations

Palladium Catalyzed Hydroamidocarbonylation of 1,3-dienes

Haoquan Li, Xianjie Fang, Ralf Jackstell, Helfried Neumann and Matthias Beller*

Leibniz-Institut für Katalyse e.V., an der Universität Rostock, Albert-Einstein-Str. 29a, 18059 Rostock, Germany.

Contents

General Remarks.....	1
General Procedure for the optimization and the scope	1
General Procedure for Reactions With Butadiene	2
Characterization Data.....	2
Chiral HPLC Spectrum for Compound 3aq.....	9
NMR Spectrum	11

General Remarks

All chemicals were purchased from Sigma-Aldrich, Strem, Acros, TCI or Alfa Aesar and used as received unless stated otherwise. Solvents (Anhydrous and under inert atmosphere) were collected from the solvent purification system and used under standard schlenk technique. 1,3-Butadiene is obtained from Evonik Industry. Isoprene was purchased from Acros and stored under -28°C under Argon atmosphere. NMR spectra were recorded on Bruker Avance 300 and Bruker ARX 400 spectrometers. Chemical shifts (ppm) are given relative to solvent: references for CDCl₃ were 7.26 ppm (¹H NMR) and 77.00 ppm (¹³C NMR). Multiplets were assigned as s (singlet), d (doublet), t (triplet), q (quartet), p (pentet) dd (doublet of doublet), m (multiplet) and br. s (broad singlet). Unless otherwise stated, yields refer to isolated yields, estimated to be >95% pure according to ¹H-NMR spectroscopy or GC. GC-yields were calculated using isooctane as internal standard. All measurements were carried out at room temperature unless otherwise stated. Electron impact (EI) mass spectra were recorded on AMD 402 mass spectrometer (70 eV). High resolution mass spectra (HRMS) were recorded on Agilent 6210. The data are given as mass units per charge (m/z). Gas chromatography analysis was performed on an Agilent HP-7890A instrument with a FID detector and HP-5 capillary column (polydimethylsiloxane with 5% phenyl groups, 30 m, 0.32 mm i.d., 0.25 µm film thickness) using argon as carrier gas. The products were isolated from the reaction mixture by column chromatography on silica gel 60, 0.063-0.2 mm, 70-230 mesh (Merck).

General Procedure for the optimization and the scope

A 4 mL screw-cap glass vial was charged with palladium salt, ligand, solid substrates and an oven-dried stirring bar under air. The vial was closed by PTFE/white rubber septa and phenolic cap and connected with atmosphere with a needle. Then the vial was evacuated under vacuum and recharged with argon for three times. Then, toluene (2 ml) is added via syringe, followed by the addition of isoprene and amide (in case of liquid) by micro syringe. The vial was fixed in an alloy plate and put into Paar 4560 series autoclave (300 mL) under argon atmosphere. At room temperature, the fixed autoclave is washed with carbon monoxide for three times before 40 bar of carbon monoxide was charged. The autoclave was placed on a heating plate equipped with magnetic stirring and an aluminum block. The reaction is allowed to heat at 100°C for 20 hours. After the reaction finishes, the autoclave is cooled to room temperature. The CO pressure was carefully released. Isooctane (100 µL) was added in to the reaction mixture as internal standard. The mixture was analyzed by GC. After removal of solvent

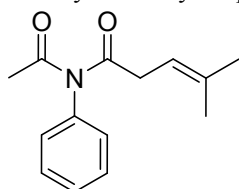
under reduced pressure, pure product was obtained by column chromatography on silica gel (eluent: pentane/ethyl acetate = 10:1).

General Procedure for Reactions With Butadiene

In a 25 mL HASTELLOY® autoclave equipped with thermometer sensor, a pressure sensor and a mechanical stirring bar, Palladium, Xantphos were added. Then the autoclave was closed. Next, the autoclave was evacuated under vacuum and recharged with argon under schlenk line for three times. Then autoclave is again opened and connected to argon flow. After toluene (10 ml) and amide were injected by syringe under argon flow, the autoclave is closed again. Afterwards, the autoclave is cooled down with an acetone-dry ice bath and about 1.1 g of 1,3-butadiene is slowly added into the autoclave while stirring (checked by balance). Further, after the autoclave is warmed up to room temperature, carbon monoxide is purged into the autoclave up to 40 bar. The reaction is allowed to be heated in a heating block to certain temperature for 20 hours. The gas consumption was monitored by Paar 4848 gas controller. After the reaction, the autoclave was cooled to room temperature and the pressure was carefully released. The mixture was diluted with acetone and analysed by gas chromatography. Pure product was obtained by column chromatography over silica gel (eluent: pentane/ethyl acetate = 10:1).

Characterization Data

N-acetyl-4-methyl-*N*-phenylpent-3-enamide

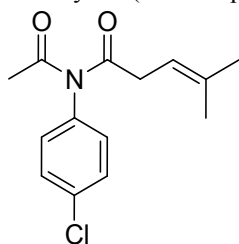


¹H NMR (300 MHz, CDCl₃) δ = 7.59 – 7.35 (m, 3H), 7.24 – 7.00 (m, 2H), 5.27 (tt, J = 6.9, 1.4 Hz, 1H), 3.18 (d, J = 6.9 Hz, 2H), 2.37 (s, 3H), 1.72 (d, J = 1.3 Hz, 3H), 1.48 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.6, 173.2, 139.1, 135.9, 129.8, 128.8, 115.8, 37.9, 27.2, 25.7, 18.1.

HRMS (EI): Calcd. for C₁₄H₁₇O₂N₁ [M]⁺: 231.12538, Found: 231.12593.

N-acetyl-*N*-(4-chlorophenyl)-4-methylpent-3-enamide

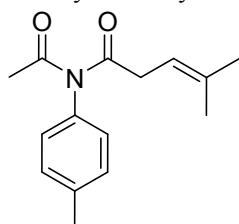


¹H NMR (300 MHz, CDCl₃) δ = 7.57 – 7.35 (m, 2H), 7.16 – 6.93 (m, 2H), 5.32 – 5.14 (m, 1H), 3.17 (d, J = 6.9 Hz, 2H), 2.37 (s, 3H), 1.85 – 1.64 (m, 3H), 1.49 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.26, 172.98, 137.53, 136.18, 134.88, 130.28, 130.00, 115.56, 37.89, 27.11, 25.72, 18.10.

HRMS (EI): Calcd. for C₁₄H₁₆O₂Cl₁N₁ [M]⁺: 265.08641; Found: 265.08674. Calcd. for C₁₄H₁₆O₂Cl₁N₁ [M]⁺: 267.08346; Found: 267.08412.

N-acetyl-4-methyl-*N*-(*p*-tolyl)pent-3-enamide

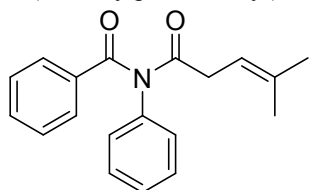


¹H NMR (300 MHz, CDCl₃) δ = 7.23 (d, J = 8.0 Hz, 2H), 7.05 – 6.90 (m, 2H), 5.26 (tt, J = 7.0, 1.5 Hz, 1H), 3.16 (d, J = 6.9 Hz, 2H), 2.36 (s, 3H), 2.34 (s, 3H), 1.72 – 1.67 (m, 3H), 1.47 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.6, 173.3, 138.7, 136.4, 135.6, 130.4, 128.5, 116.0, 37.8, 27.1, 25.7, 21.2, 18.1.

HRMS (EI): Calcd. for C₁₅H₁₉O₂N₁ [M]⁺: 245.14103; Found: 245.14145.

N-(4-methylpent-3-enoyl)-*N*-phenylbenzamide

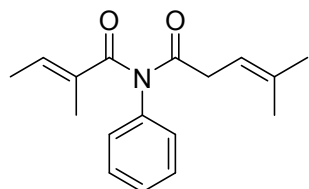


¹H NMR (300 MHz, CDCl₃) δ = 7.59 – 7.48 (m, 2H), 7.38 – 7.14 (m, 6H), 7.10 – 7.01 (m, 2H), 5.35 – 5.22 (m, 1H), 3.32 (ddt, *J* = 8.3, 2.2, 1.1 Hz, 2H), 1.63 (d, *J* = 1.4 Hz, 3H), 1.47 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 175.5, 173.0, 139.1, 136.4, 135.0, 132.1, 129.4, 129.1, 128.5, 128.3, 128.1, 116.2, 37.2, 25.7, 18.2.

HRMS (EI): Calcd. for C₁₉H₁₉O₂N₁ [M]⁺: 293.14103 ; Found: 293.14104 .

4-Methyl-*N*-(2-methylbut-2-enoyl)-*N*-phenylpent-3-enamide

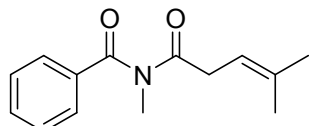


¹H NMR (300 MHz, CDCl₃) δ = 7.37 – 7.23 (m, 2H), 7.28 – 7.17 (m, 1H), 7.09 – 6.97 (m, 2H), 6.37 – 6.20 (m, 1H), 5.22 (dddt, *J* = 7.1, 5.7, 2.9, 1.4 Hz, 1H), 3.28 – 3.00 (m, 2H), 1.72 – 1.63 (m, 3H), 1.68 – 1.60 (m, 3H), 1.65 – 1.56 (m, 3H), 1.45 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 175.1, 174.9, 139.1, 136.02, 135.95, 133.8, 129.3, 128.1, 127.9, 116.3, 36.9, 25.7, 18.1, 14.2, 13.0.

HRMS (EI): Calcd. for C₁₇H₂₁O₂N₁ [M]⁺: 271.15668 ; Found: 271.15630.

N-methyl-*N*-(4-methylpent-3-enoyl)benzamide

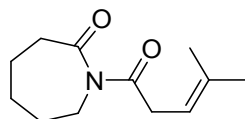


¹H NMR (400 MHz, CDCl₃) δ = 7.63 – 7.57 (m, 2H), 7.56 – 7.49 (m, 1H), 7.48 – 7.40 (m, 2H), 5.38 – 5.21 (m, 1H), 3.34 (d, *J* = 6.9 Hz, 2H), 3.18 (s, 3H), 1.68 (s, 3H), 1.52 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ = 175.6, 174.1, 135.7, 135.3, 132.4, 128.8, 128.5, 116.5, 37.3, 34.4, 25.7, 18.1.

HRMS (EI): Calcd. for C₁₄H₁₇O₂N₁ [M]⁺: 231.12538; Found: 231.12601.

1-(4-Methylpent-3-enoyl)azepan-2-one

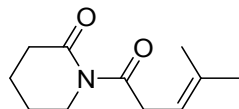


¹H NMR (300 MHz, CDCl₃) δ = 5.38 – 5.26 (m, 1H), 3.91 – 3.82 (m, 2H), 3.64 – 3.47 (m, 2H), 2.78 – 2.64 (m, 2H), 1.85 – 1.48 (m, 12H).

¹³C NMR (75 MHz, CDCl₃) δ = 177.7, 174.9, 135.0, 116.9, 43.4, 39.9, 38.5, 29.2, 28.5, 25.8, 23.7, 18.2.

HRMS (EI): Calcd. for C₁₁H₁₇O₂N₁ [M]⁺: 195.12527; Found: 195.12538.

1-(4-Methylpent-3-enoyl)piperidin-2-one

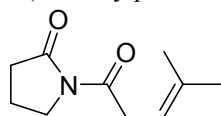


¹H NMR (300 MHz, CDCl₃) δ = 5.42 – 5.24 (m, 1H), 3.67 (t, 2H), 3.58 (d, *J* = 7.0 Hz, 2H), 2.60 – 2.44 (m, 2H), 1.85 – 1.72 (m, 4H), 1.72 – 1.65 (m, 3H), 1.59 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 175.6, 173.3, 134.9, 116.8, 44.0, 38.9, 34.9, 25.7, 22.4, 20.3, 18.1.

HRMS (EI): Calcd. for C₁₁H₁₇O₂N₁ [M]⁺:195.12538; Found:195.12491 .

1-(4-Methylpent-3-enoyl)pyrrolidin-2-one

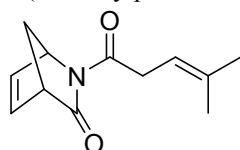


¹H NMR (300 MHz, CDCl₃) δ = 5.46 – 5.25 (m, 1H), 3.95 – 3.70 (m, 2H), 3.67 – 3.37 (m, 2H), 2.66 – 2.35 (m, 2H), 2.15 – 1.91 (m, 2H), 1.73 (s, 3H), 1.64 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 175.3, 172.9, 135.5, 115.7, 115.6, 45.6, 36.2, 33.7, 25.8, 25.7, 18.2, 18.2, 17.2.

HRMS (EI): Calcd. for C₁₀H₁₅O₂N₁ [M]⁺:181.10973; Found:181.10964.

2-(4-Methylpent-3-enoyl)-2-azabicyclo[2.2.1]hept-5-en-3-one

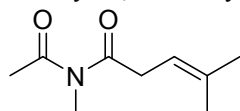


¹H NMR (300 MHz, CDCl₃) δ = 6.89 (ddd, *J* = 5.3, 2.4, 0.6 Hz, 1H), 6.65 (ddd, *J* = 5.2, 3.2, 1.5 Hz, 1H), 5.39 – 5.19 (m, 2H), 3.64 – 3.35 (m, 3H), 2.30 (dt, *J* = 8.6, 1.7 Hz, 1H), 2.19 (dt, *J* = 8.7, 1.5 Hz, 1H), 1.84 – 1.67 (m, 3H), 1.63 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 177.5, 171.6, 140.5, 138.1, 135.6, 115.6, 60.4, 54.7, 54.7, 35.4, 25.8, 18.2.

HRMS (EI): Calcd. for C₁₂H₁₅O₂N₁ [M]⁺: 205.10973; Found: 205.10955 .

N-acetyl-*N*,4-dimethylpent-3-enamide

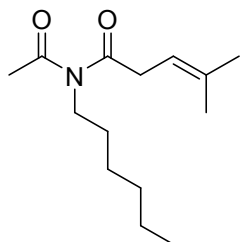


¹H NMR (300 MHz, CDCl₃) δ = 5.38 – 5.19 (m, 1H), 3.36 (d, *J* = 6.8 Hz, 2H), 3.18 (s, 3H), 2.38 (s, 3H), 1.72 (d, *J* = 1.4 Hz, 3H), 1.61 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.8, 173.4, 135.8, 115.8, 37.5, 31.5, 26.8, 25.7, 18.1.

HRMS (EI): Calcd. for C₉H₁₅O₂N₁ [M]⁺:169.10973; Found:169.10987.

N-acetyl-*N*-hexyl-4-methylpent-3-enamide

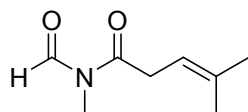


¹H NMR (300 MHz, CDCl₃) δ 5.30 (t, *J* = 6.9 Hz, 1H), 3.60 (t, *J* = 7.9 Hz, 2H), 3.37 (d, *J* = 6.9 Hz, 2H), 2.37 (s, 3H), 1.73 (s, 3H), 1.62 (s, 3H), 1.59 – 1.43 (m, 2H), 1.35 – 1.19 (m, 6H), 0.85 (t, *J* = 5.7 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.9, 173.3, 135.6, 116.2, 44.7, 37.4, 31.4, 29.3, 26.6, 26.6, 25.7, 22.5, 18.1, 14.0.

HRMS (EI): Calcd. for C₁₄H₂₅O₂N₁ [M]⁺: 239.18798 ; Found: 239.18811 .

N-formyl-*N*,4-dimethylpent-3-enamide

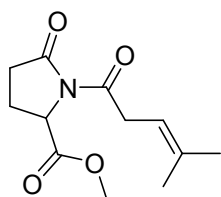


¹H NMR (400 MHz, CDCl₃) δ = 9.15 (s, 1H), 5.31 – 5.13 (m, 1H), 3.31 (d, *J* = 6.7 Hz, 1H), 3.04 (s, 3H), 1.70 (s, 3H), 1.61 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) δ = 173.3, 162.7, 137.0, 114.5, 35.0, 25.6, 18.1.

HRMS (EI): Calcd. for C₈H₁₃O₂N₁ [M]⁺:155.09408; Found:155.09387 .

N-formyl-*N*,4-dimethylpent-3-enamide

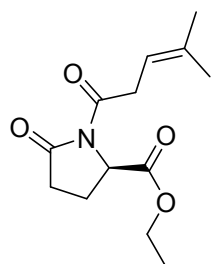


¹H NMR (300 MHz, CDCl₃) δ = 5.35 – 5.17 (m, 1H), 4.68 (dd, J = 9.5, 2.8 Hz, 1H), 3.69 (d, J = 2.9 Hz, 3H), 3.63 – 3.51 (m, 2H), 2.82 – 2.41 (m, 2H), 2.39 – 2.17 (m, 1H), 2.15 – 1.93 (m, 1H), 1.67 (s, 3H), 1.58 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.3, 172.6, 171.5, 135.7, 115.1, 57.9, 52.6, 36.0, 31.9, 25.7, 21.3, 18.1.

HRMS (EI): Calcd. for C₁₂H₁₇O₄N₁ [M]⁺: 239.11521; Found: 239.11477.

Ethyl (R)-1-(4-methylpent-3-enoyl)-5-oxopyrrolidine-2-carboxylate

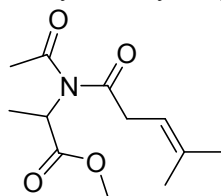


¹H NMR (300 MHz, CDCl₃) δ = 5.41 – 5.24 (m, 1H), 4.69 (dd, J = 9.5, 2.7 Hz, 1H), 4.17 (q, J = 7.1 Hz, 2H), 3.62 (d, J = 6.8 Hz, 2H), 2.69 (dt, J = 17.8, 10.0 Hz, 1H), 2.59 – 2.45 (m, 1H), 2.41 – 2.20 (m, 1H), 2.11 – 1.94 (m, 1H), 1.70 (s, 3H), 1.60 (s, 3H), 1.23 (t, J = 7.2 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.3, 172.6, 171.0, 135.8, 115.2, 61.6, 58.1, 36.0, 32.0, 25.7, 21.3, 18.2, 14.0.

HRMS (EI): Calcd. for C₁₃H₁₉O₄N₁ [M]⁺: 253.13086 ; Found: 253.13081 .

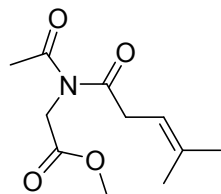
Methyl *N*-acetyl-*N*-(4-methylpent-3-enoyl)alaninate



¹H NMR (300 MHz, CDCl₃) δ = 5.38 – 5.18 (m, 1H), 4.51 (q, J = 6.9 Hz, 1H), 3.67 (s, 3H), 3.41 – 3.27 (m, 2H), 2.37 (s, 3H), 1.73 (d, J = 1.4 Hz, 3H), 1.61 (d, J = 1.2 Hz, 3H), 1.50 (d, J = 6.9 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.4, 172.8, 170.7, 136.3, 115.4, 54.2, 52.5, 37.7, 26.6, 25.7, 18.1, 15.5.

Methyl *N*-acetyl-*N*-(4-methylpent-3-enoyl)glycinate

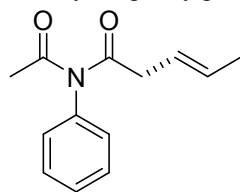


¹H NMR (300 MHz, CDCl₃) δ = 5.35 – 5.18 (m, 1H), 4.39 (s, 2H), 3.71 (s, 3H), 3.32 (d, J = 6.8 Hz, 2H), 2.38 (s, 3H), 1.70 (s, 3H), 1.58 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.1, 172.8, 169.1, 136.2, 115.4, 52.5, 45.7, 37.3, 26.3, 25.7, 18.0.

HRMS (EI): Calcd. for C₁₂H₁₇O₄N₁ [M]⁺: 239.11521 ; Found: 239.11577 .

N-acetyl-*N*-phenylpent-3-enamide



Trans isomer:

¹H NMR (300 MHz, CDCl₃) δ = 7.55 – 7.36 (m, 3H), 7.22 – 7.07 (m, 2H), 5.91 – 5.26 (m, 2H), 3.22 – 3.10 (m, 2H), 2.36 (s, 3H), 1.74 – 1.61 (m, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.5 , 173.2 , 139.0 , 129.8 , 128.9 (3C) 122.8 , 42.0 , 27.2 , 18.0 .

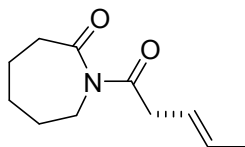
Cis isomer:

¹H NMR (300 MHz, CDCl₃) δ = 7.55 – 7.36 (m, 3H), 7.22 – 7.07 (m, 2H), 5.91 – 5.26 (m, 2H), 3.32 – 3.24 (m, 2H), 2.33 (s, 3H), 1.53 – 1.48 (m, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.1 , 173.2 , 139.0 , 129.8 , 128.9 (2C), 127.8 , 121.8, 36.7, 27.1, 13.1.

HRMS (EI): Calcd. for C₁₃H₁₅O₂N₁ [M]⁺: 217.10973; Found: 217.11032.

1-(Pent-3-enoyl)azepan-2-one



Trans isomer: (73%)

¹H NMR (300 MHz, CDCl₃) δ = 5.73 – 5.47 (m, 2H), 3.99 – 3.81 (m, 2H), 3.62 – 3.53 (m, 2H), 2.77 – 2.64 (m, 2H), 1.87 – 1.55 (m, 9H).

¹³C NMR (75 MHz, CDCl₃) δ = 177.6 , 174.8 , 128.8 , 124.0 , 43.3 , 42.7 , 39.8 , 29.2 , 28.5 , 23.7 , 18.0 .

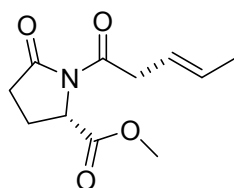
Cis isomer: (27%)

¹H NMR (300 MHz, CDCl₃) δ = 5.73 – 5.47 (m, 2H), 3.99 – 3.81 (m, 2H), 3.70 – 3.61 (m, 2H), 2.77 – 2.64 (m, 2H), 1.87 – 1.55 (m, 9H).

¹³C NMR (75 MHz, CDCl₃) δ = 177.6 , 174.5 , 127.1 , 122.9 , 43.3 , 39.8 , 37.4 , 29.2 , 28.5 , 23.7 , 13.1 .

HRMS (EI): Calcd. for C₁₁H₁₇O₂N₁ [M]⁺: 195.12538 Found: 195.12527.

Methyl (S,E)-5-oxo-1-(pent-3-enoyl)pyrrolidine-2-carboxylate



E/Z mixture

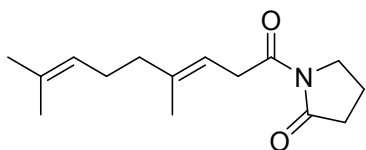
¹H NMR (400 MHz, CDCl₃) δ = 5.73 – 5.50 (m, 2H), 4.81 – 4.68 (m, 1H), 3.75 (d, *J* = 0.8 Hz, 3H), 3.73 – 3.60 (m, 2H), 2.80 – 2.64 (m, 1H), 2.63 – 2.48 (m, 1H), 2.41 – 2.21 (m, 1H), 2.15 – 1.96 (m, 1H), 1.71 – 1.61 (m, 3H).

Major isomer **¹³C NMR** (101 MHz, CDCl₃) δ = 174.3 , 172.7 , 171.6 , 129.8 , 122.2 , 57.9 , 52.7 , 40.2 , 32.0 , 21.4 , 18.1 .

Minor Isomer **¹³C NMR** (101 MHz, CDCl₃) δ = 174.3 , 172.2 , 171.6 , 127.9 , 121.1 , 57.9 , 52.7 , 34.9 , 32.0 , 21.4 , 13.2 .

HRMS (EI): Calcd. for C₁₁H₁₅O₄N₁ [M]⁺: 225.09956 Found: 225.09956.

1-(4,8-Dimethylnona-3,7-dienoyl)pyrrolidin-2-one



¹H NMR (300 MHz, CDCl₃) δ = 5.41 – 5.26 (m, 1H), 5.14 – 4.91 (m, 1H), 3.85 – 3.67 (m, 2H), 3.57 (d, J = 6.9 Hz, 2H), 2.54 (t, J = 8.1 Hz, 2H), 2.15 – 1.88 (m, 6H), 1.78 – 1.55 (m, 6H), 1.55 – 1.48 (m, 3H).

Major Isomer:

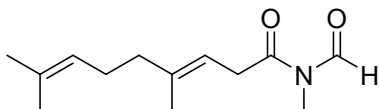
¹³C NMR (75 MHz, CDCl₃) δ = 175.2, 172.7, 138.9, 131.4, 124.0, 115.5, 45.5, 39.6, 36.1, 33.7, 26.5, 25.7, 17.6, 17.2, 16.6.

Minor Isomer:

¹³C NMR (75 MHz, CDCl₃) δ = 175.2, 172.8, 139.0, 131.7, 124.0, 116.3, 45.5, 39.6, 35.9, 32.3, 26.3, 23.5, 17.6, 17.2, 16.6.

HRMS (EI): Calcd. for C₁₅H₂₃O₂N₁ [M]⁺: 249.17233 ; Found: 249.17250 .

N-formyl-N,4,8-trimethylnona-3,7-dienamide

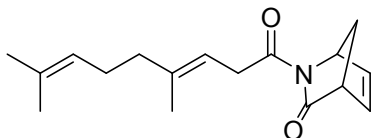


¹H NMR (300 MHz, CDCl₃) δ = 9.19 (s, 1H), 5.35 – 5.21 (m, 2H), 5.13 – 4.99 (m, 1H), 3.35 (d, J = 6.9 Hz, 3H), 3.18 – 3.02 (m, 5H), 2.05 (s, 3H), 1.75 (q, J = 1.3 Hz, 5H), 1.66 (s, 4H), 1.59 (s, 5H).

¹³C NMR (75 MHz, CDCl₃) δ = 173.3, 162.7, 140.7, 132.4, 123.5, 115.0, 34.8, 32.3, 26.6, 26.1, 25.7, 23.4, 17.7.

HRMS (EI): Calcd. for C₁₃H₂₁O₂N₁ [M]⁺: 223.15668 ; Found: 223.15705 .

2-(4,8-Dimethylnona-3,7-dienoyl)-2-azabicyclo[2.2.1]hept-5-en-3-one

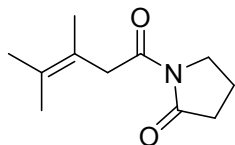


¹H NMR (300 MHz, CDCl₃) δ = 6.99 – 6.80 (m, 1H), 6.71 – 6.55 (m, 1H), 5.41 – 5.30 (m, 1H), 5.29 – 5.22 (m, 0H), 5.13 – 5.01 (m, 1H), 3.58 – 3.45 (m, 2H), 3.45 – 3.36 (m, 1H), 2.35 – 2.16 (m, 2H), 2.12 – 1.95 (m, 4H), 1.77 – 1.54 (m, 10H).

¹³C NMR (75 MHz, CDCl₃) δ = 177.4, 171.5, 140.5, 139.2, 139.2, 138.1, 131.8, 131.5, 124.1, 124.0, 116.2, 115.4, 60.4, 54.7, 54.7, 39.7, 35.4, 35.1, 32.3, 26.6, 26.4, 25.7, 23.5, 17.7, 17.7, 16.6. (mixture of cis 31% and trans 69%)

HRMS (EI): Calcd. for C₁₇H₂₃O₂N₁ [M]⁺: 273.17233; Found: 273.17228.

1-(3,4-Dimethylpent-3-enoyl)pyrrolidin-2-one

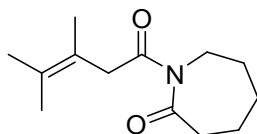


¹H NMR (300 MHz, CDCl₃) δ = ¹H NMR (300 MHz, Chloroform-*d*) δ 3.80 (t, J = 7.1 Hz, 2H), 3.65 (s, 2H), 2.59 (t, J = 8.1 Hz, 2H), 2.13 – 1.95 (m, 2H), 1.69 (s, 3H), 1.67 – 1.63 (m, 3H), 1.64 – 1.57 (m, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 175.5, 172.7, 128.4, 120.8, 45.6, 42.2, 33.8, 20.7, 20.6, 17.2.

HRMS (EI): Calcd. for C₁₁H₁₇O₂N₁ [M]⁺: 195.12538 ; Found: 195.12543 .

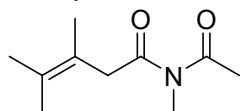
1-(3,4-Dimethylpent-3-enoyl)azepan-2-one



¹H NMR (300 MHz, CDCl₃) δ = 3.86 – 3.68 (m, 2H), 3.51 (s, 2H), 2.70 – 2.48 (m, 2H), 1.81 – 1.44 (m, 15H).

¹³C NMR (75 MHz, CDCl₃) δ = 177.7, 174.7, 127.7, 122.0, 44.3, 43.3, 39.7, 29.1, 28.6, 23.7, 20.6, 20.5, 19.2.

N-acetyl-N,3,4-trimethylpent-3-enamide

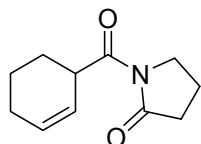


¹H NMR (300 MHz, CDCl₃) δ = 3.37 (s, 2H), 3.16 (s, 3H), 2.37 (s, 3H), 1.67 (s, 3H), 1.63 (s, 3H), 1.59 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ = 174.6, 173.4, 128.6, 120.8, 43.4, 31.4, 26.8, 20.6, 18.9.

HRMS (EI): Calcd. for C₁₀H₁₇O₂N [M]⁺: 183.12538; Found: 183.12483.

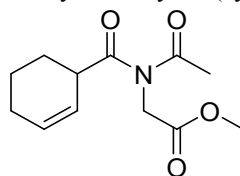
N-acetyl-N,3,4-trimethylpent-3-enamide



¹H NMR (300 MHz, CDCl₃) δ = 5.98 – 5.78 (m, 1H), 5.73 – 5.56 (m, 1H), 4.32 – 4.18 (m, 1H), 3.79 (t, *J* = 7.1 Hz, 2H), 2.59 (t, *J* = 8.1 Hz, 2H), 2.35 – 1.49 (m, 8H).

¹³C NMR (75 MHz, CDCl₃) δ = 175.9, 174.9, 129.7, 124.3, 46.0, 41.6, 34.0, 25.4, 24.6, 20.8, 17.2.

Methyl *N*-acetyl-*N*-(cyclohex-2-ene-1-carbonyl)glycinate



¹H NMR (300 MHz, CDCl₃) δ = 5.94 – 5.76 (m, 1H), 5.66 – 5.45 (m, 1H), 4.39 (s, 2H), 3.70 (s, 3H), 3.75-3.65 (m, 1H), 2.33 (s, 3H), 2.22 – 1.48 (m, 6H).

¹³C NMR (75 MHz, CDCl₃) δ = 177.5, 172.9, 169.2, 130.4, 123.8, 52.6, 46.1, 42.7, 26.2, 25.7, 24.6, 20.6.

Chiral HPLC Spectrum for Compound 3aq

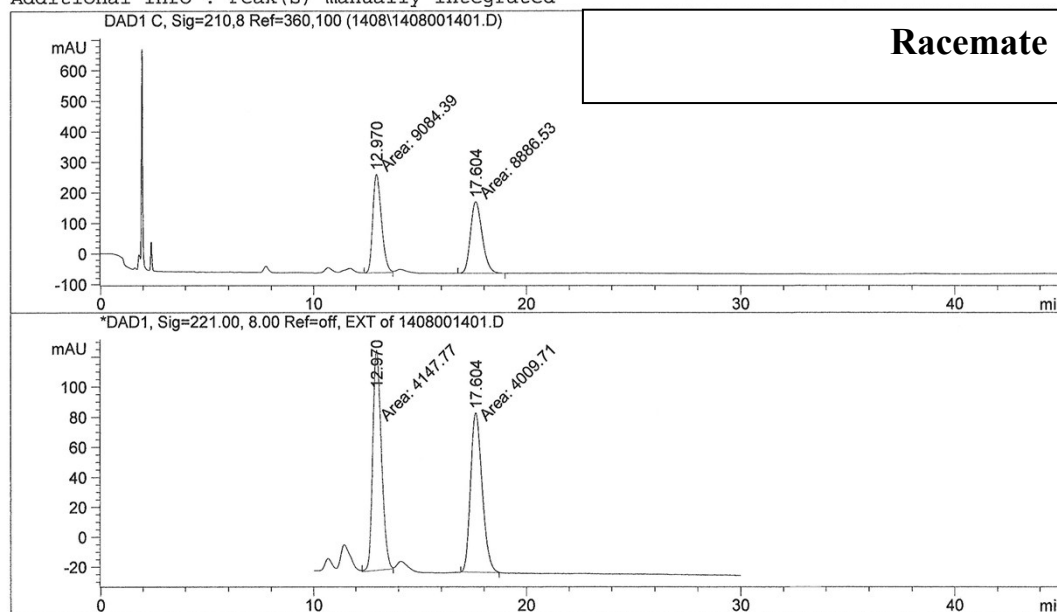
Data File D:\HPCHEM\2\DATA\1408\1408001401.D

Sample Name: LHQ784A

```
=====
Acq. Operator   :                               Seq. Line :    1
Acq. Instrument : LC 2                         Location  : Vial 11
Injection Date  : 8/14/2014 11:56:52 AM        Inj       :    1
                                           Inj Volume: 0.2 µl

Acq. Method     : C:\CHEM32\2\METHODS\FISCHER.M
Last changed    : 8/14/2014 12:42:11 PM
                  (modified after loading)
Analysis Method : C:\CHEM32\2\METHODS\FISCHER.M
Last changed    : 8/14/2014 12:46:13 PM
                  (modified after loading)
Method Info     : Cellulose 2, Heptan/Isoprop 95:5, Fluß: 1ml/min
=====
```

Additional Info : Peak(s) manually integrated



Area Percent Report

```
=====
Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
=====
```

Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.970	MM	0.4691	9084.39160	322.78085	50.5505
2	17.604	MM	0.6324	8886.52637	234.19997	49.4495
Totals :				1.79709e4	556.98082	

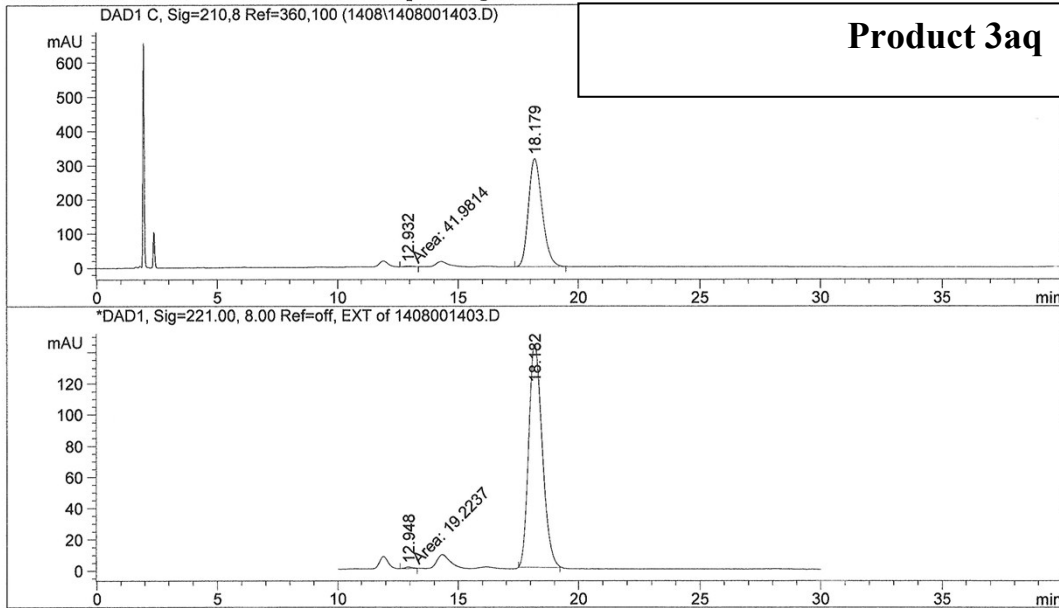
50.49
49.51

Data File D:\HPCHEM\2\DATA\1408\1408001403.D
Sample Name: LHQ784B

```
=====
Acq. Operator   :                               Seq. Line :    3
Acq. Instrument : LC 2                         Location  : Vial 12
Injection Date  : 8/14/2014 1:24:26 PM          Inj       :    1
                                                Inj Volume: 0.2 µl

Acq. Method     : C:\CHEM32\2\METHODS\FISCHER.M
Last changed    : 8/14/2014 12:42:11 PM
                  (modified after loading)
Analysis Method : C:\CHEM32\2\METHODS\FISCHER.M
Last changed    : 8/14/2014 2:31:35 PM
                  (modified after loading)
Method Info     : Cellulose 2, Heptan/Isoprop 95:5, Fluß: 1ml/min
=====
```

Additional Info : Peak(s) manually integrated



=====
Area Percent Report
=====

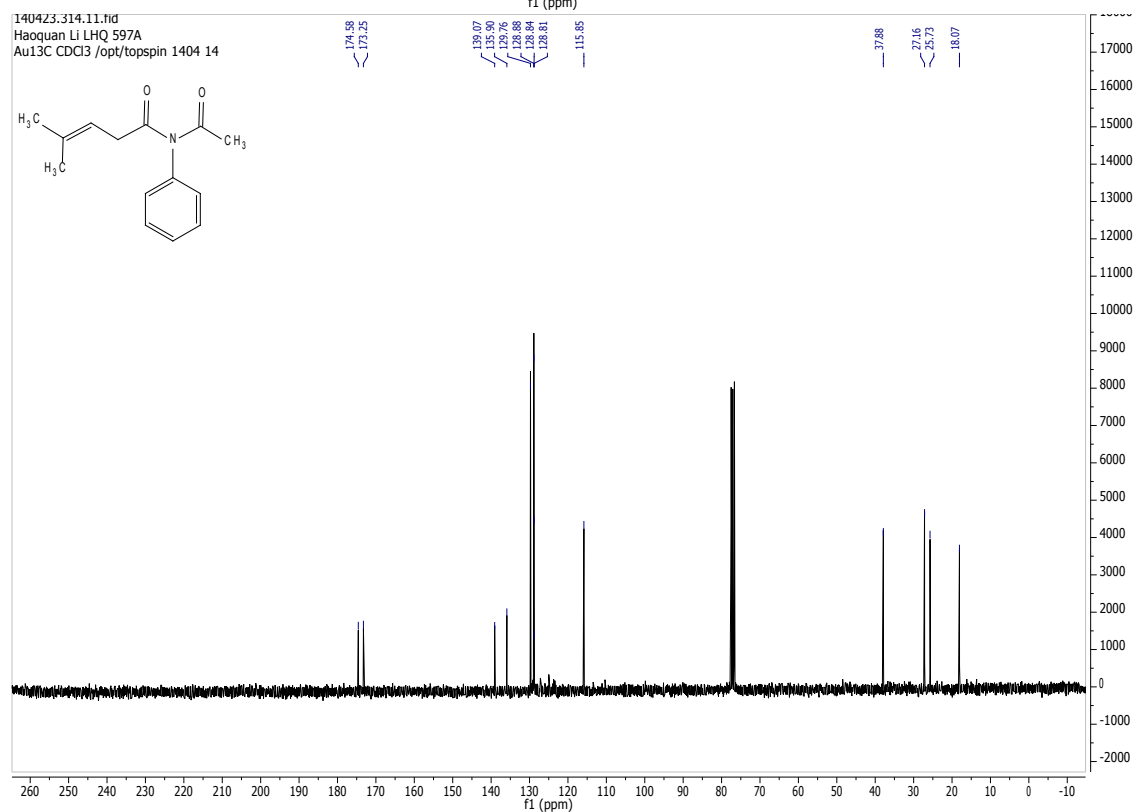
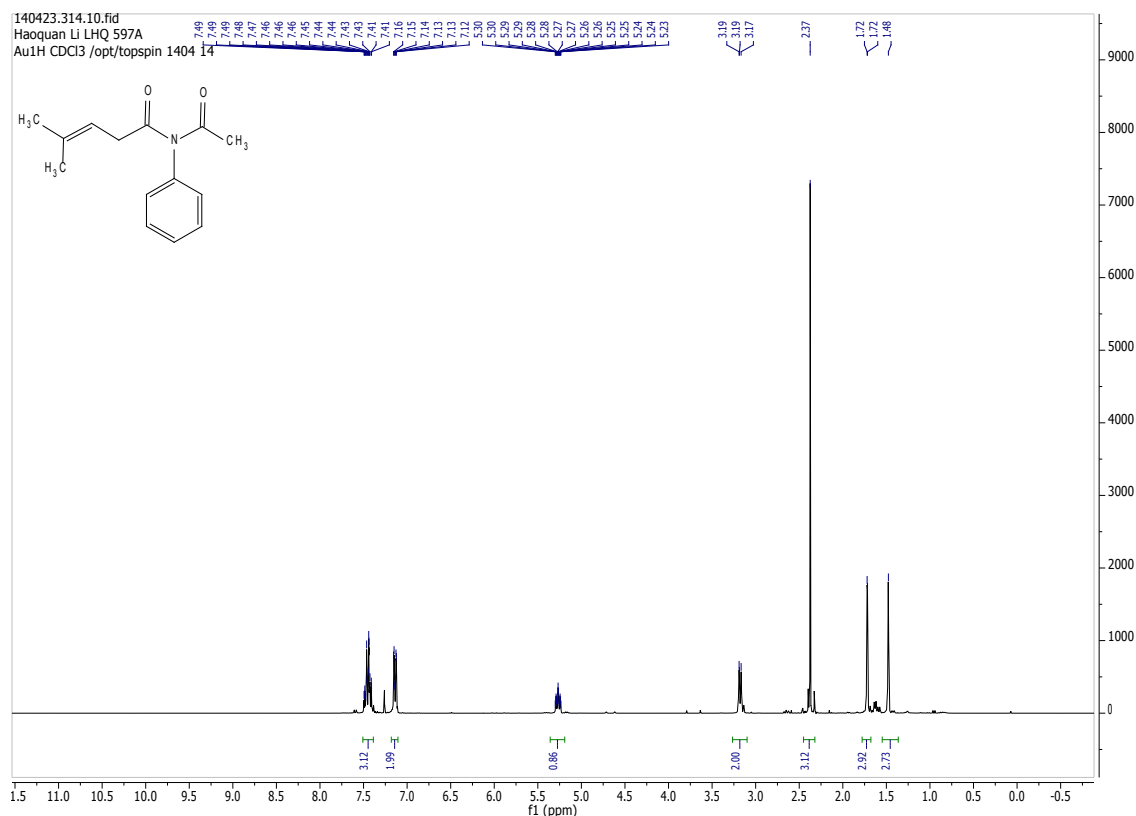
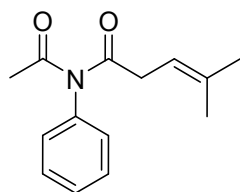
Sorted By : Signal
Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

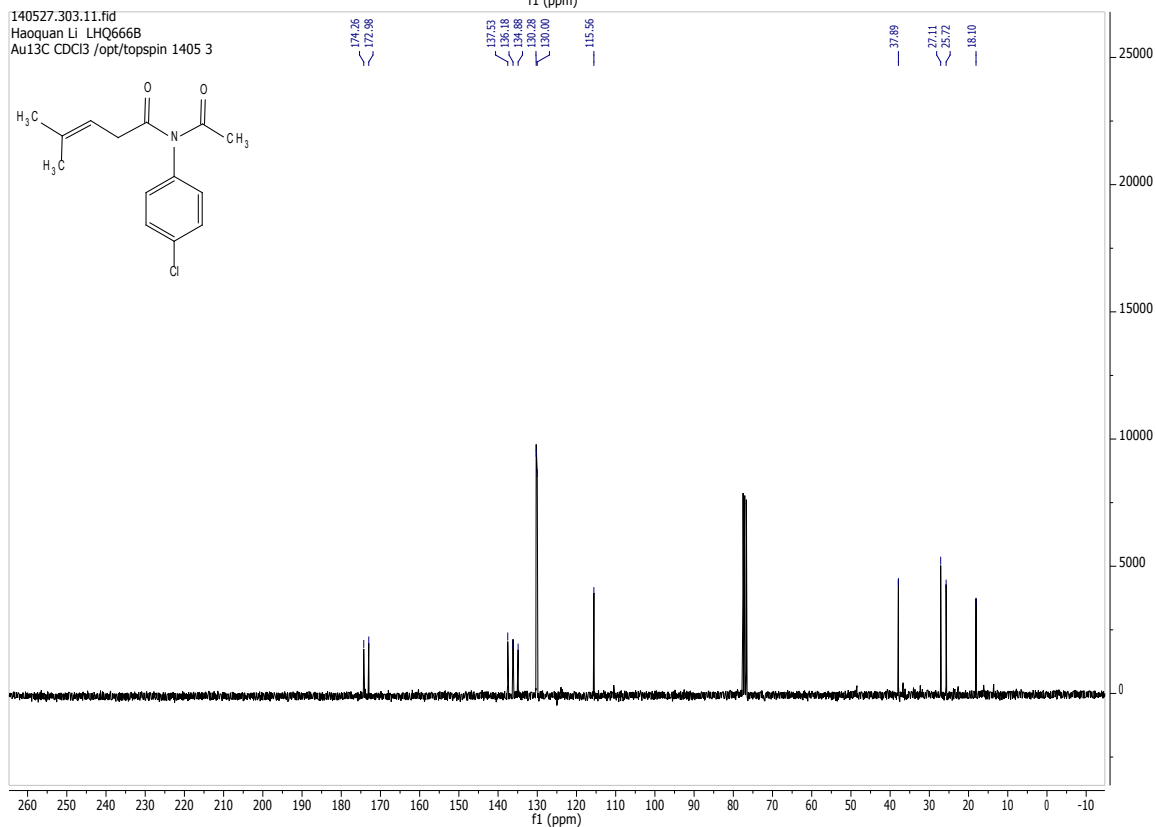
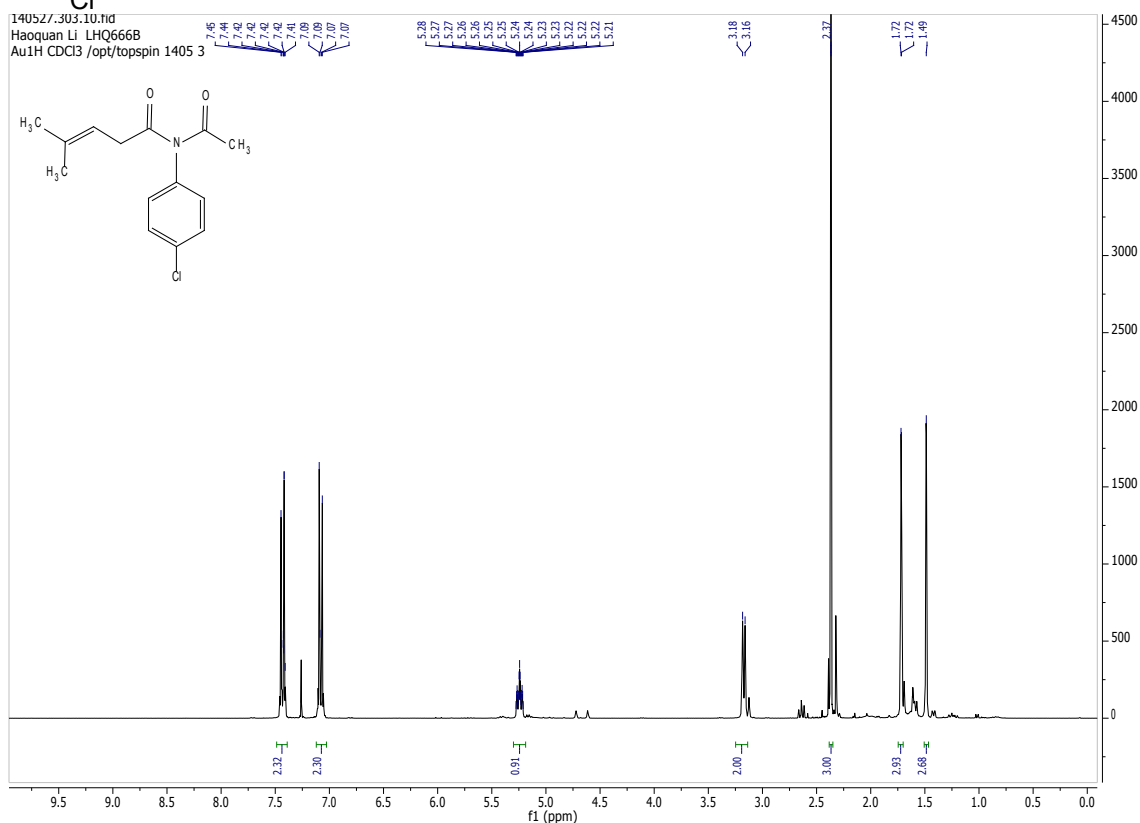
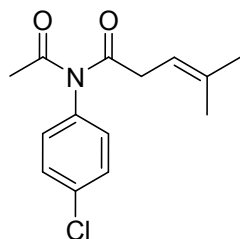
Signal 1: DAD1 C, Sig=210,8 Ref=360,100

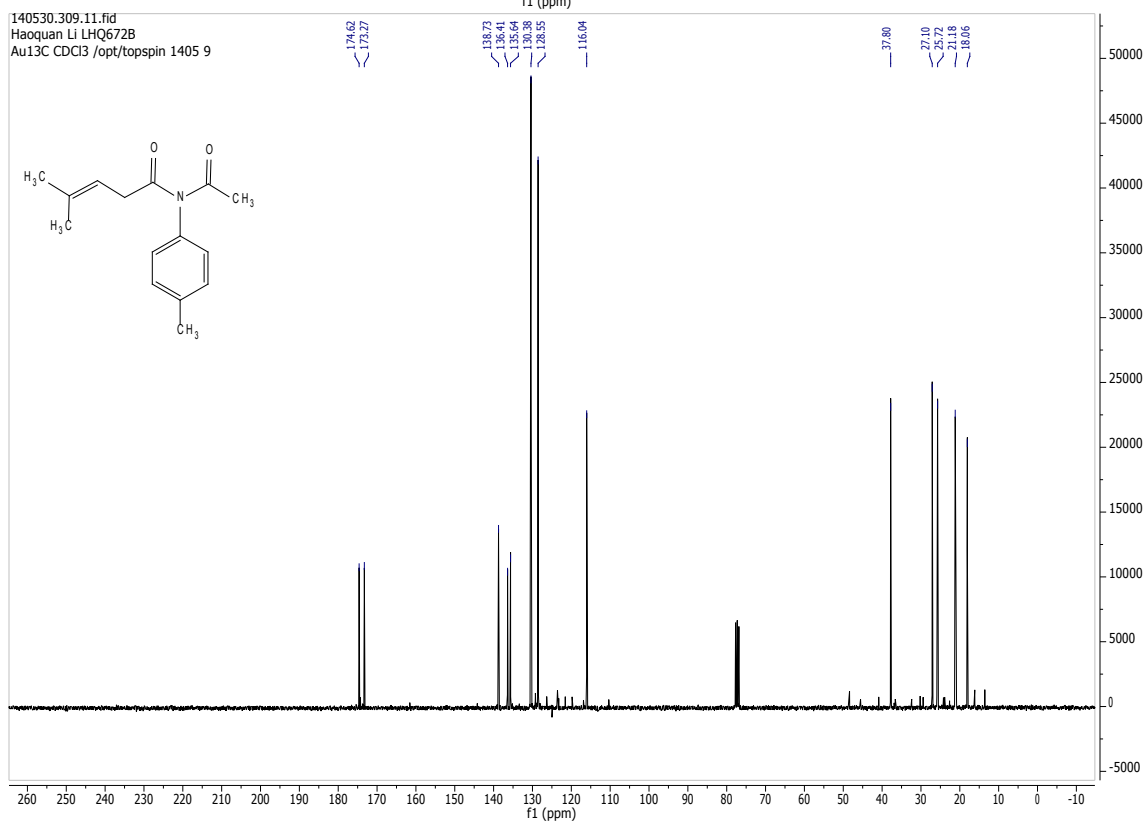
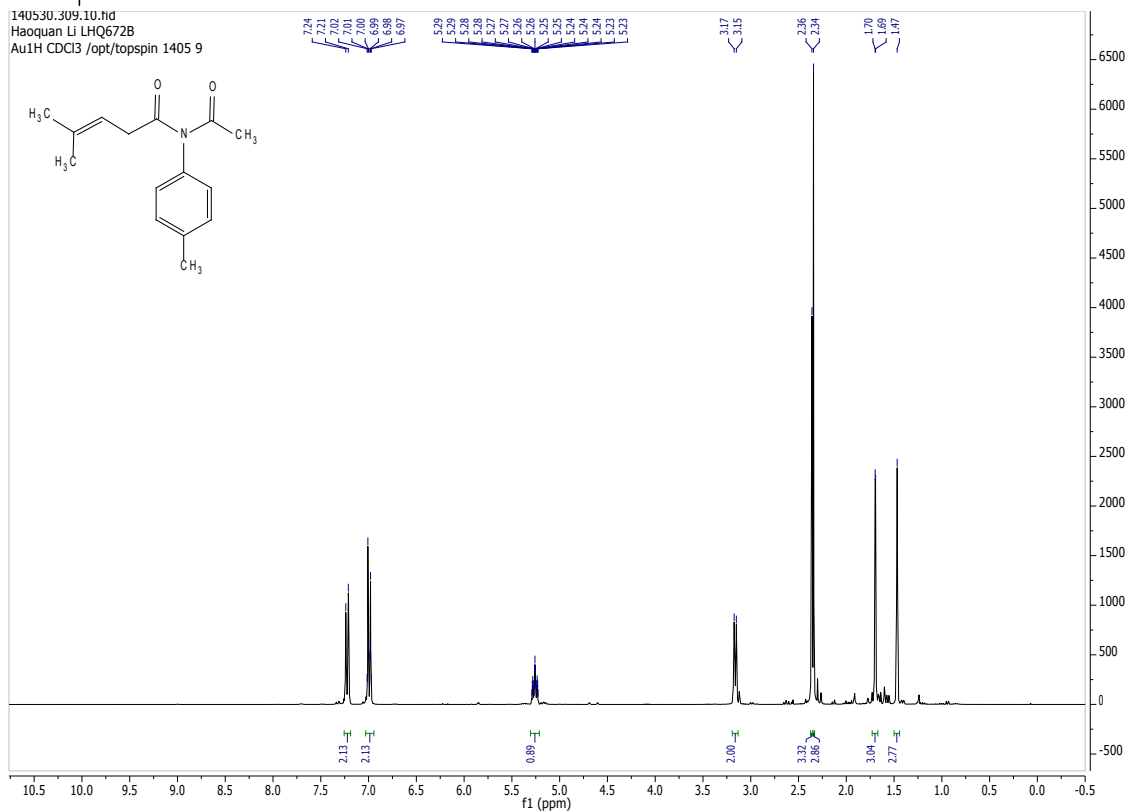
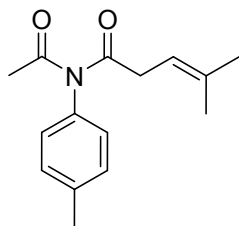
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.932	MM	0.3623	41.98138	1.93121	0.3366
2	18.179	BB	0.6130	1.24302e4	315.94415	99.6634
Totals :				1.24722e4	317.87536	

0,30
99,70

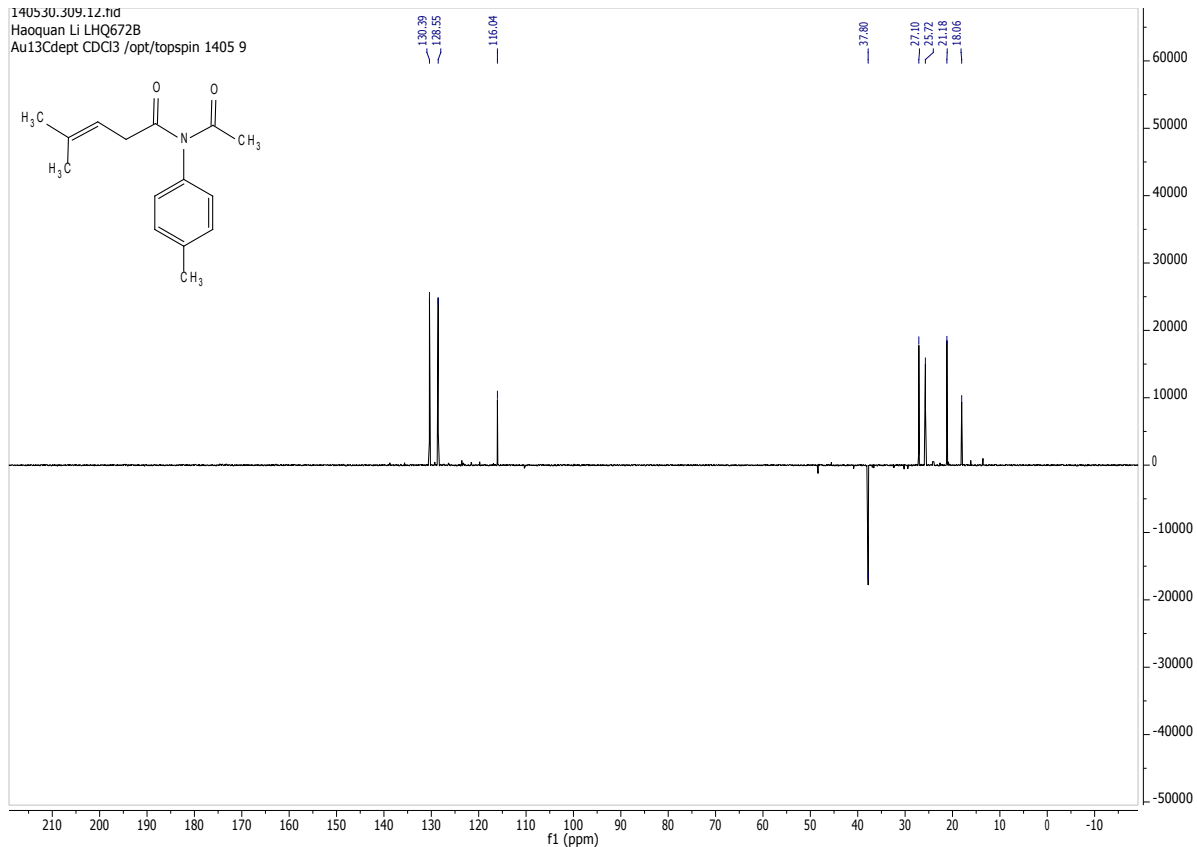
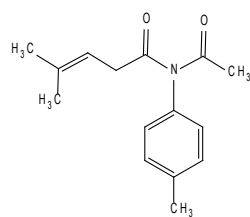
NMR Spectrum

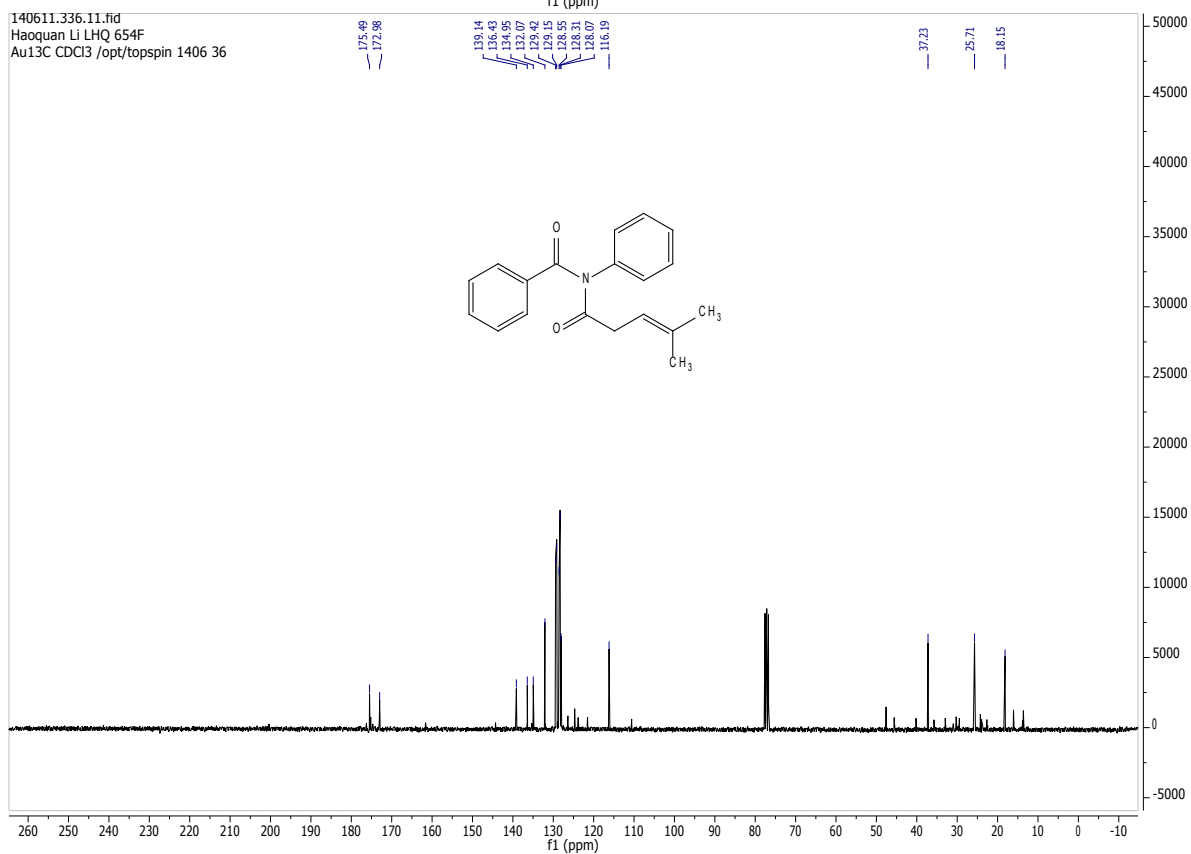
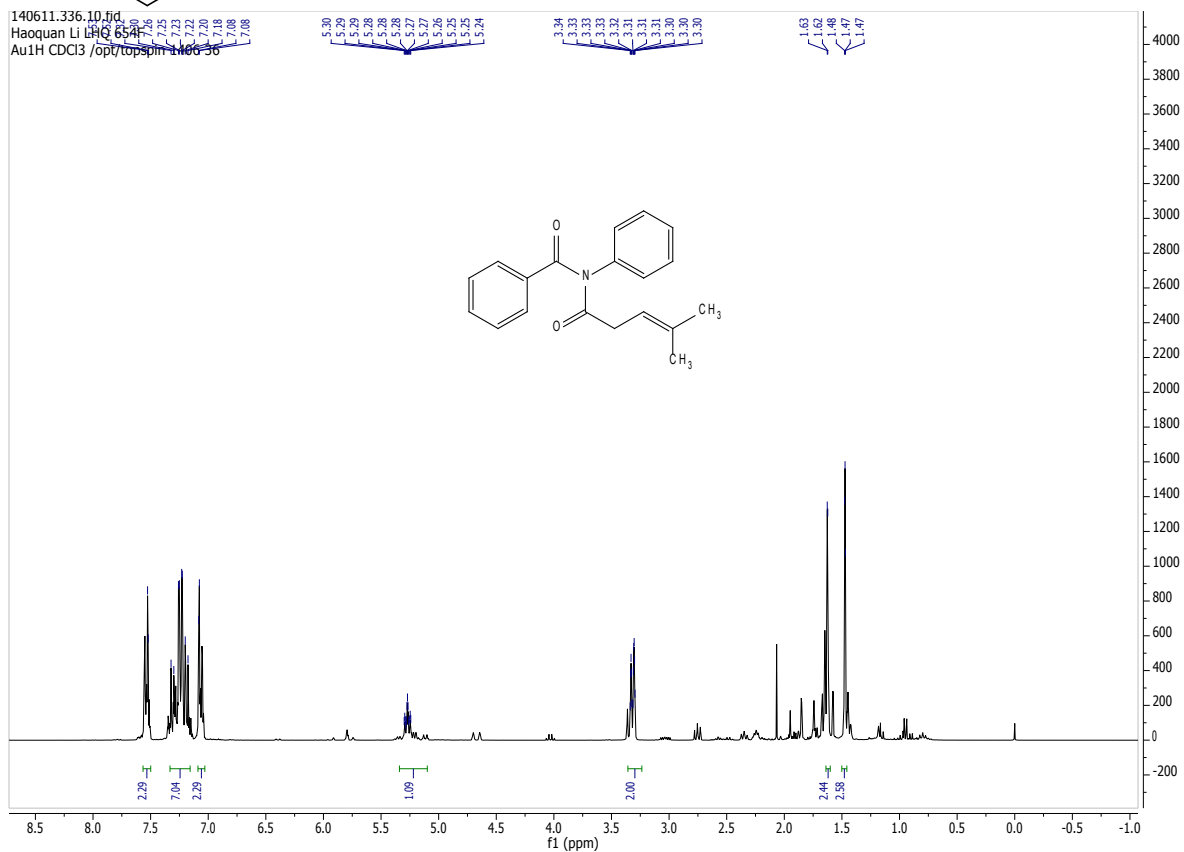
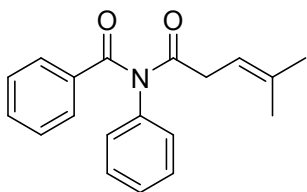


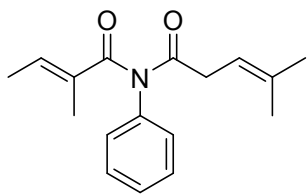




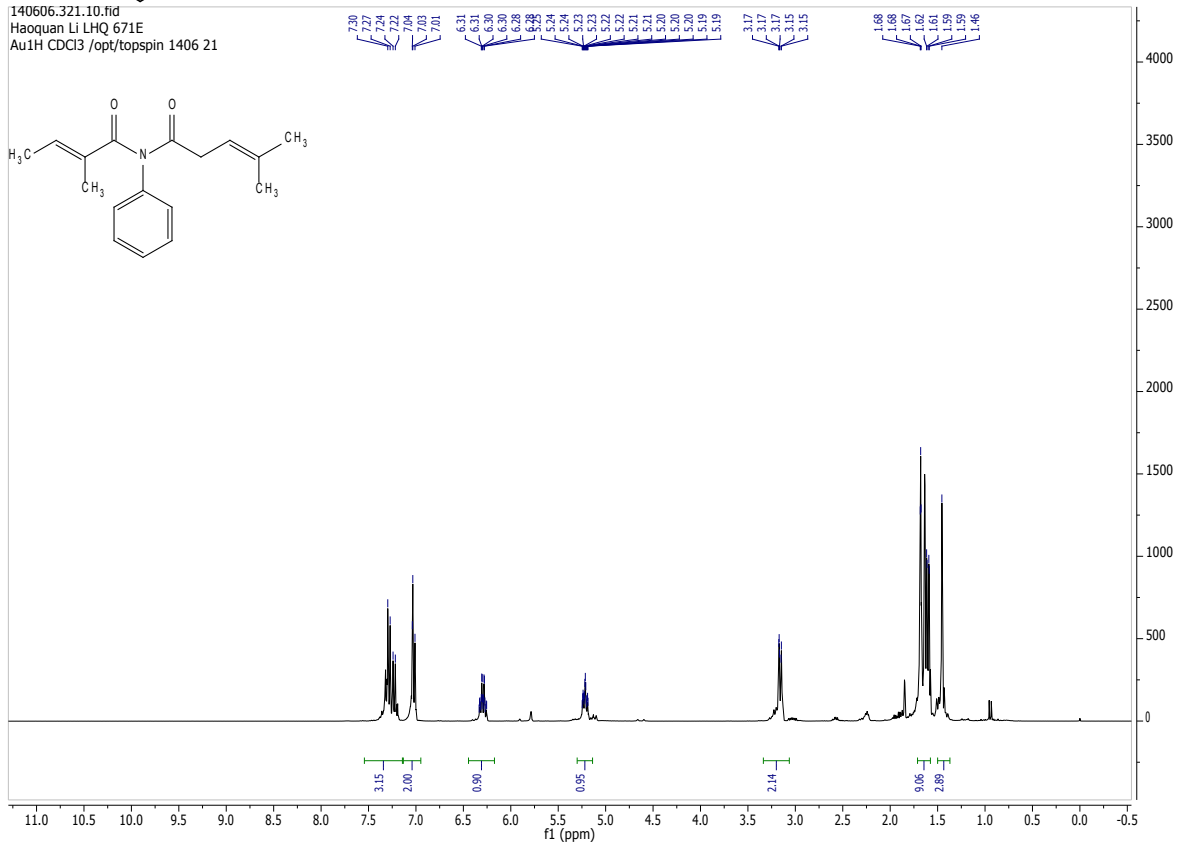
140530.309.12.hd
Haoquan Li LHQ672B
Au13Cdept CDCl3 /opt/topspin 1405 9



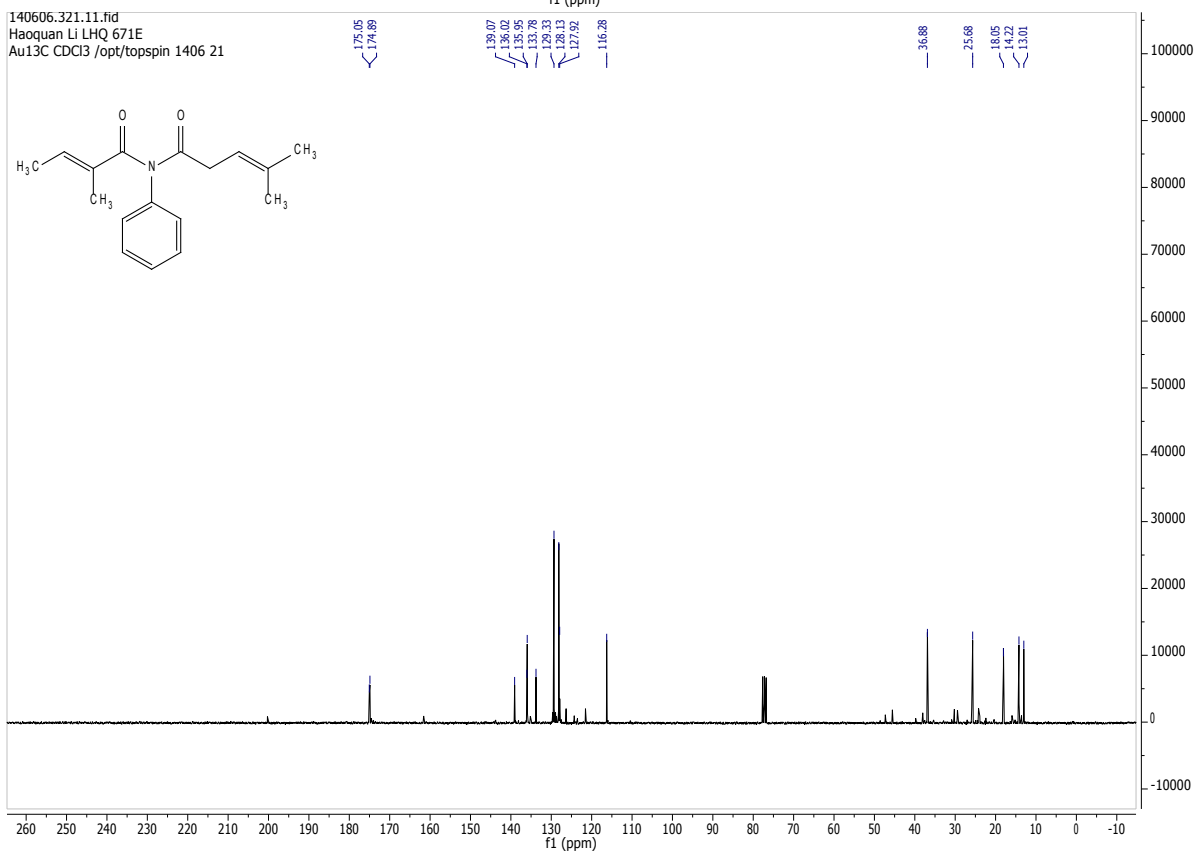


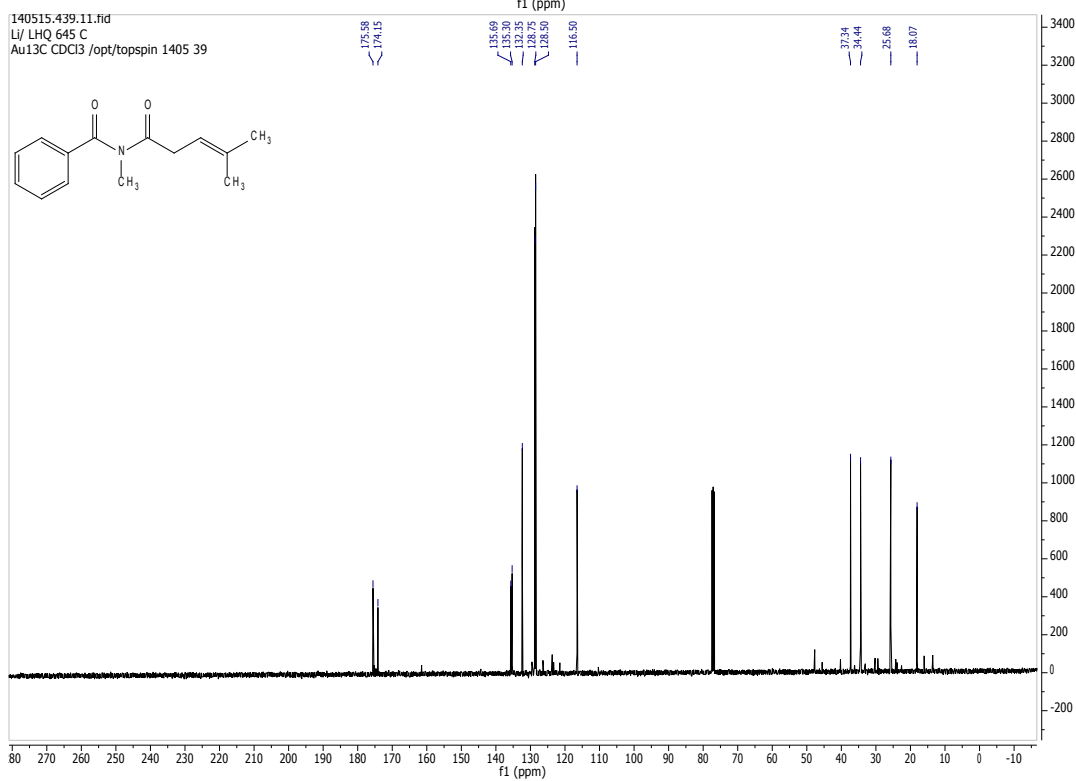
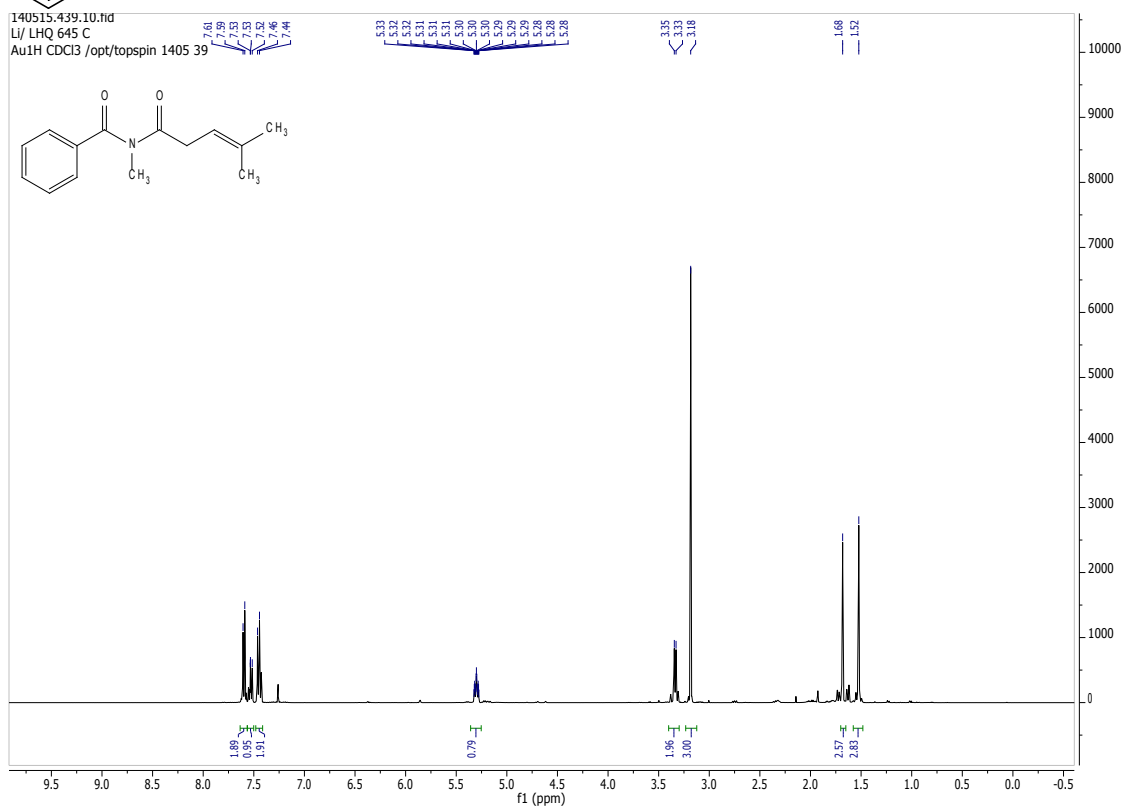
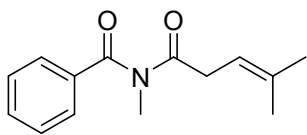


140606.321.10.fid
Haoquan Li LHQ 671E
Au1H CDCl3 /opt/topspin 1406 21

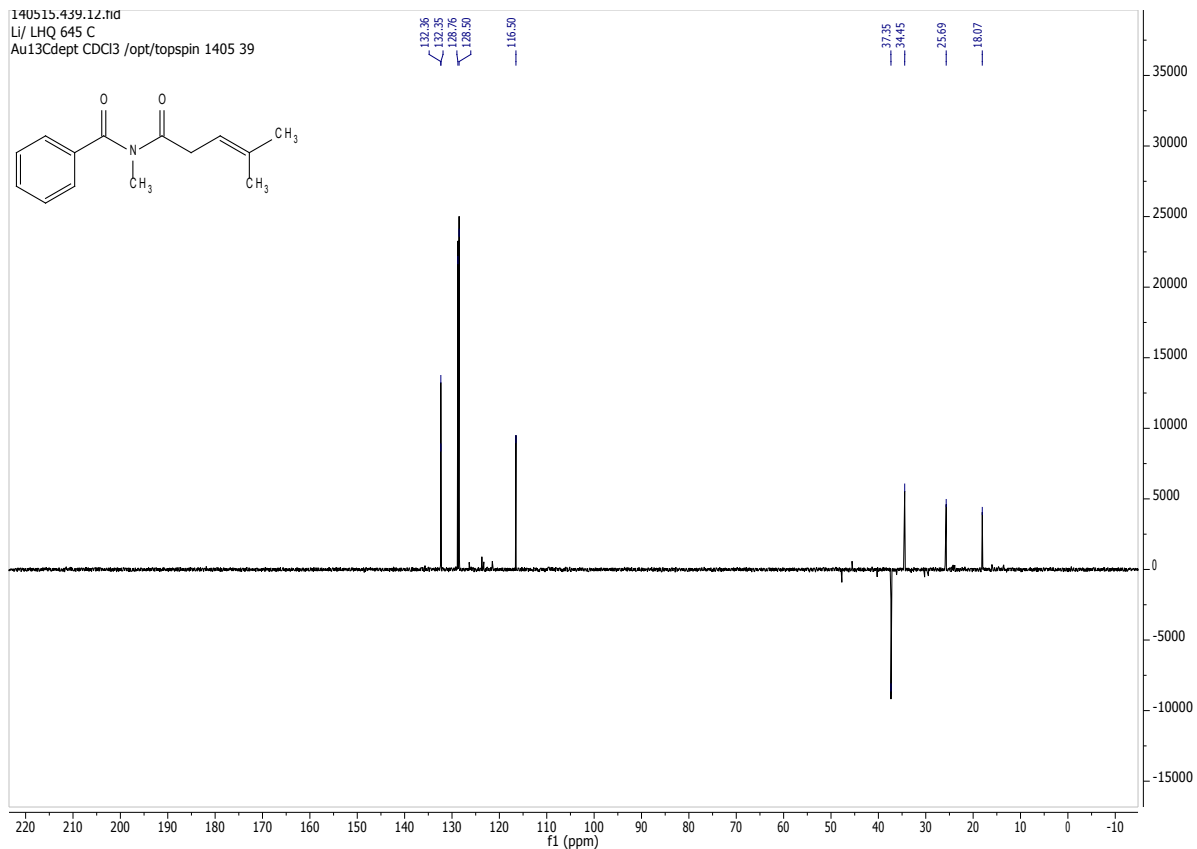
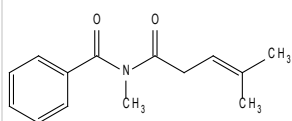


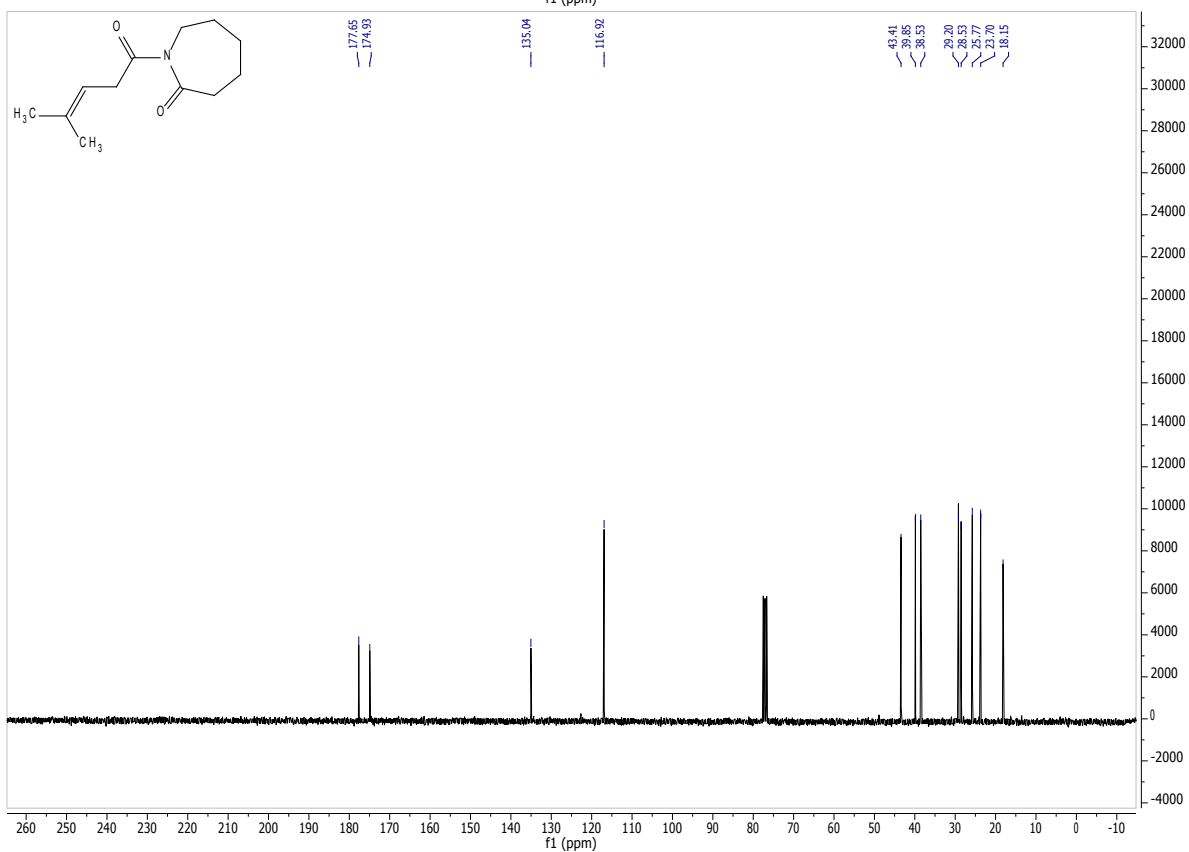
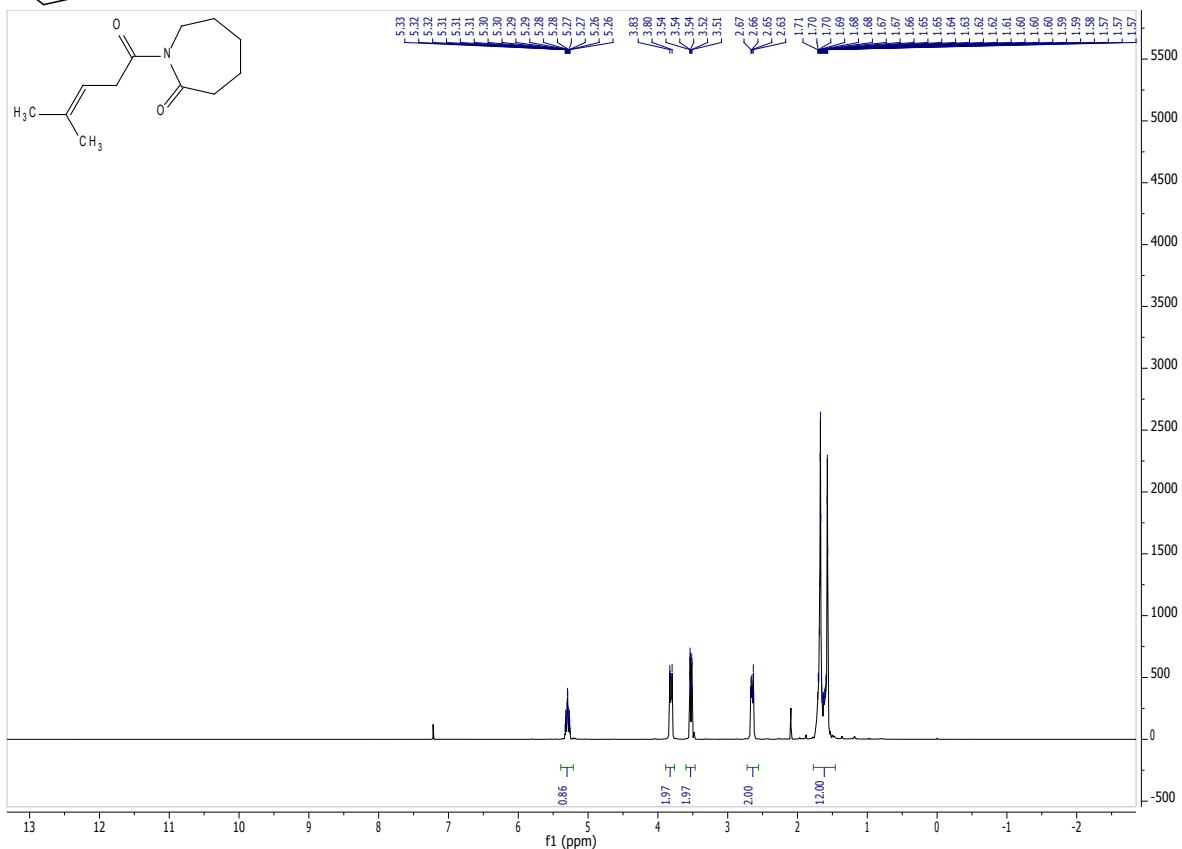
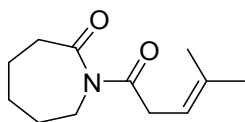
140606.321.11.fid
Haoquan Li LHQ 671E
Au13C CDCl3 /opt/topspin 1406 21

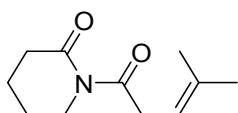




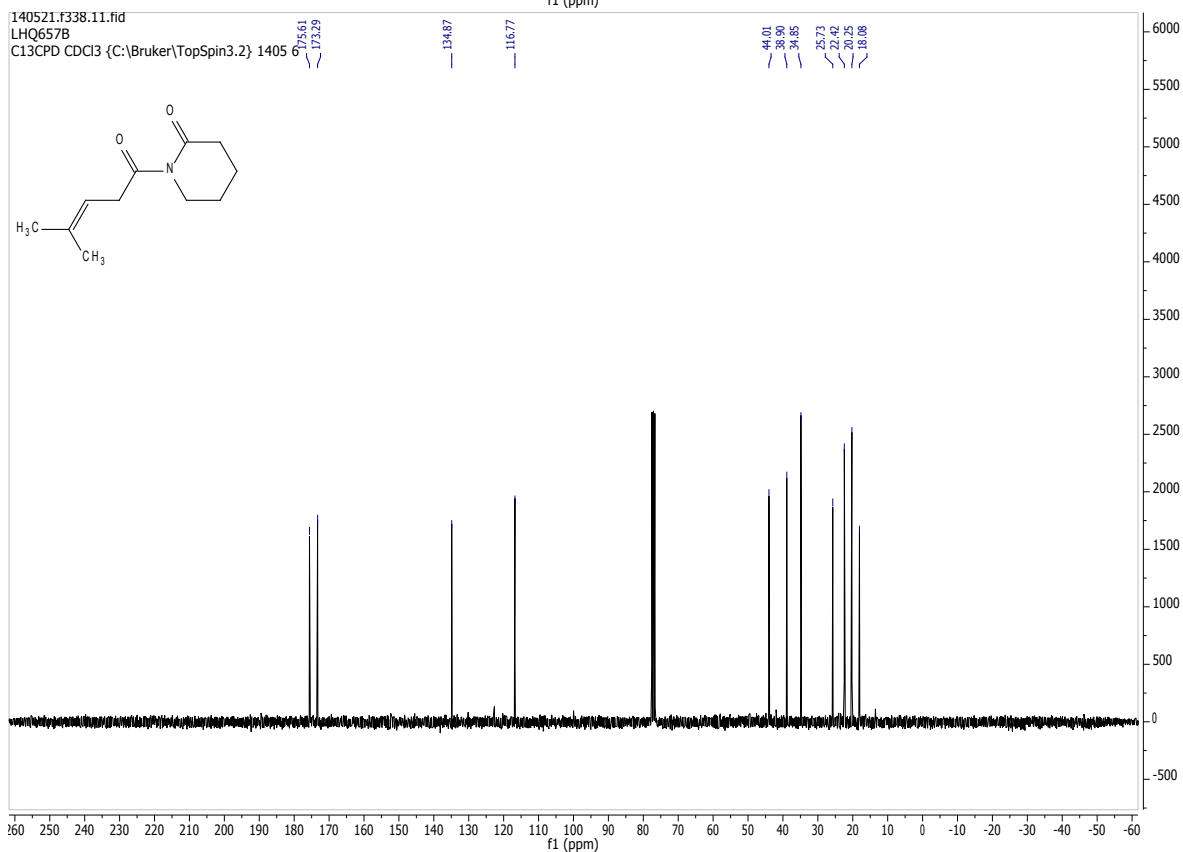
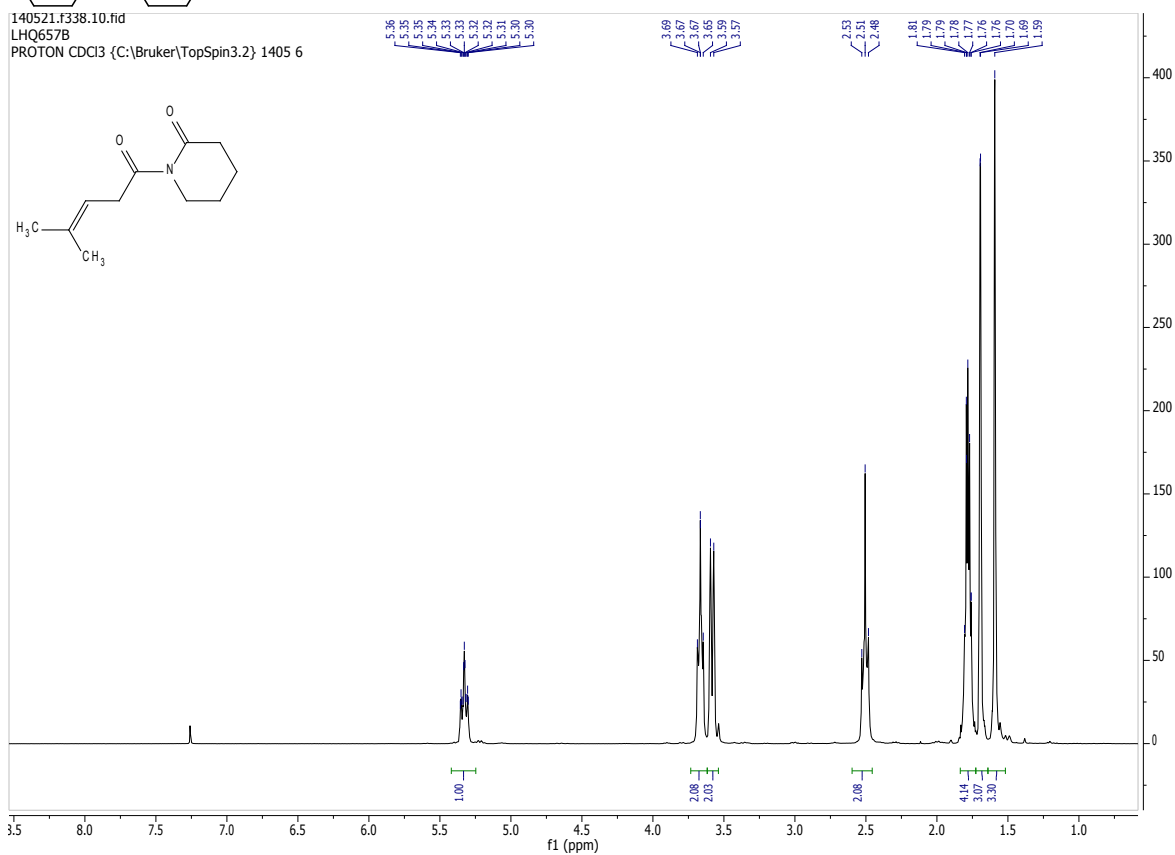
140515.439.12.hd
L/ LHQ 645 C
Au13Cdept CDCl3 /opt/topspin 1405 39

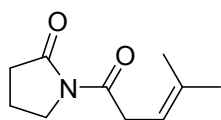




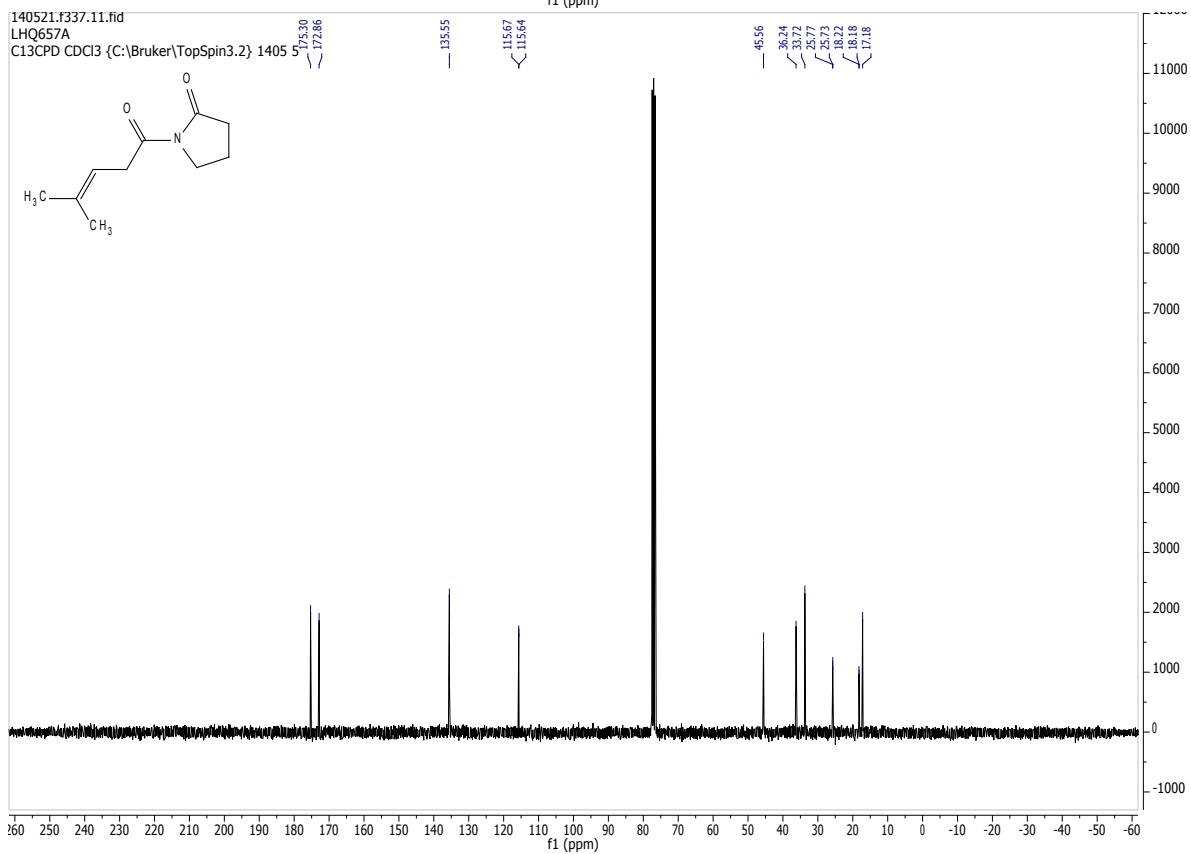
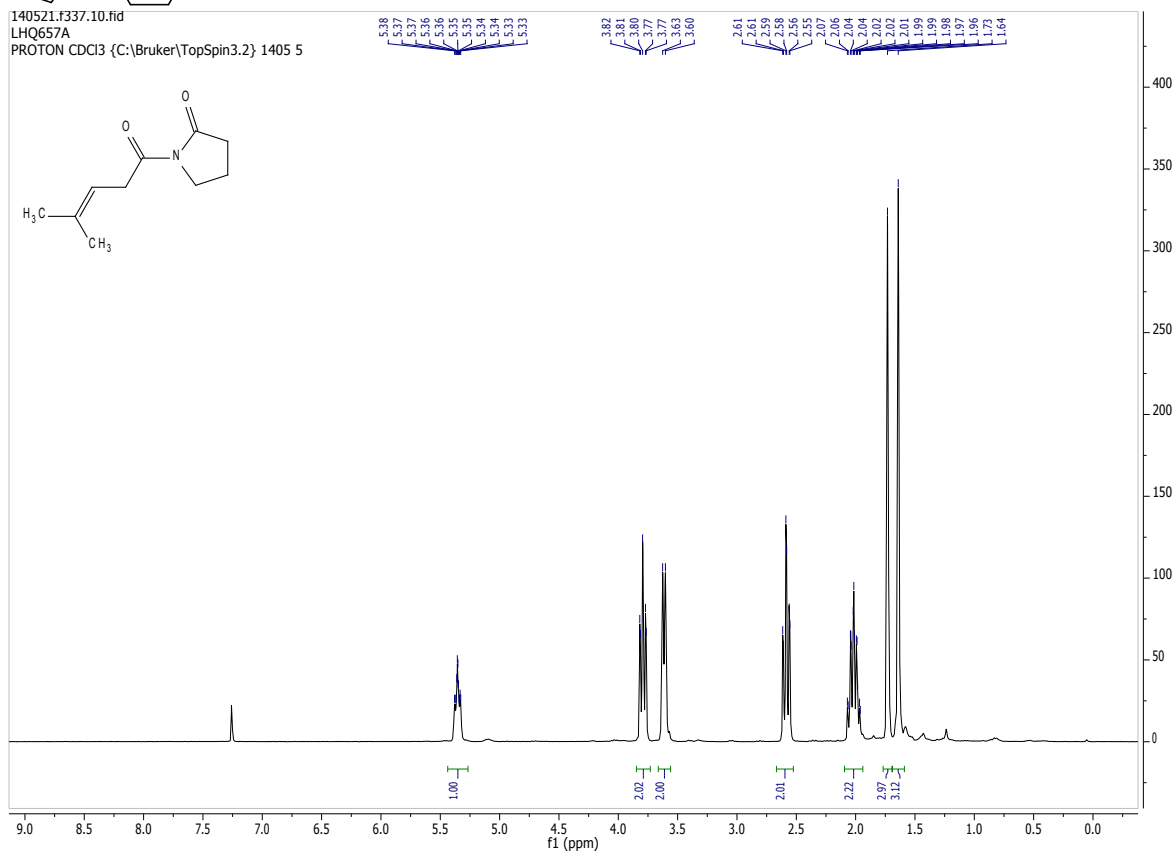


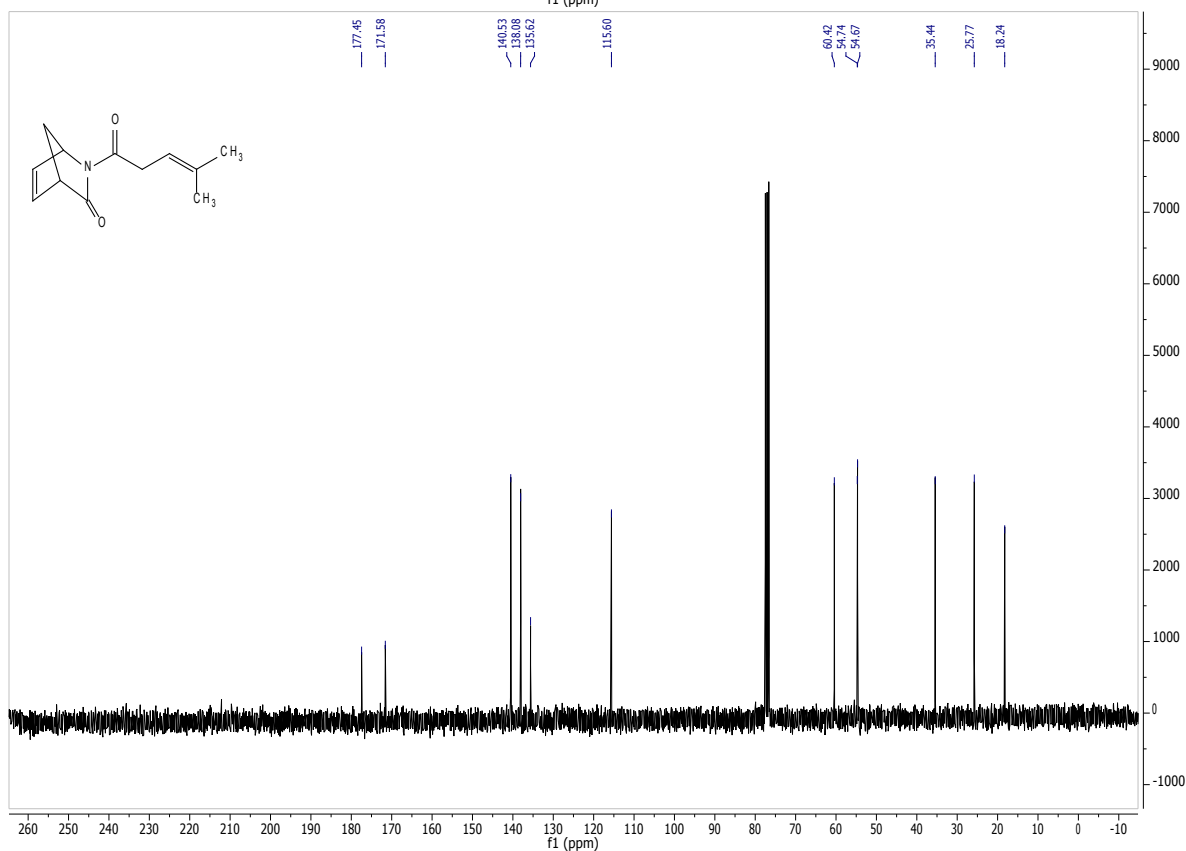
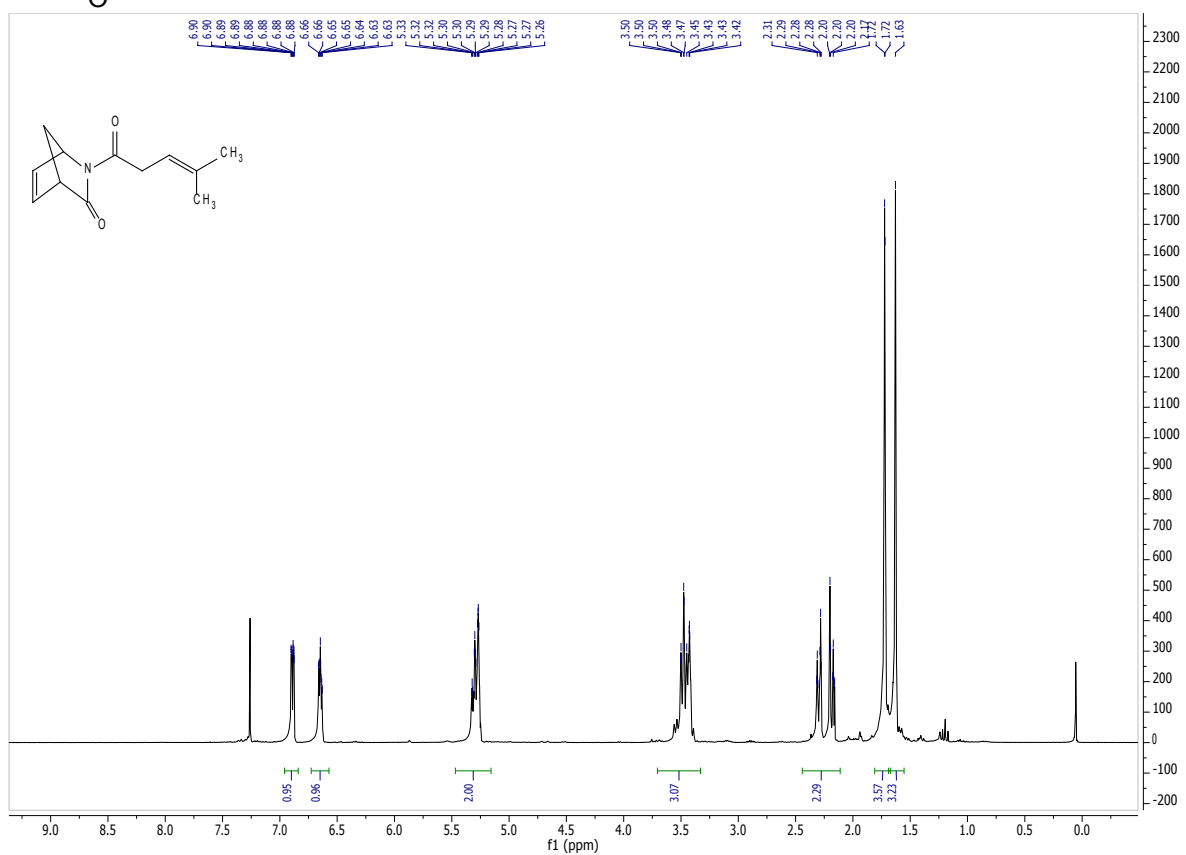
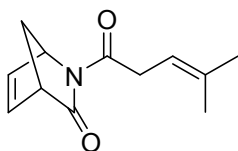
140521.f338.10.fid
LHQ657B
PROTON CDCl₃ {C:\Bruker\TopSpin3.2} 1405 6

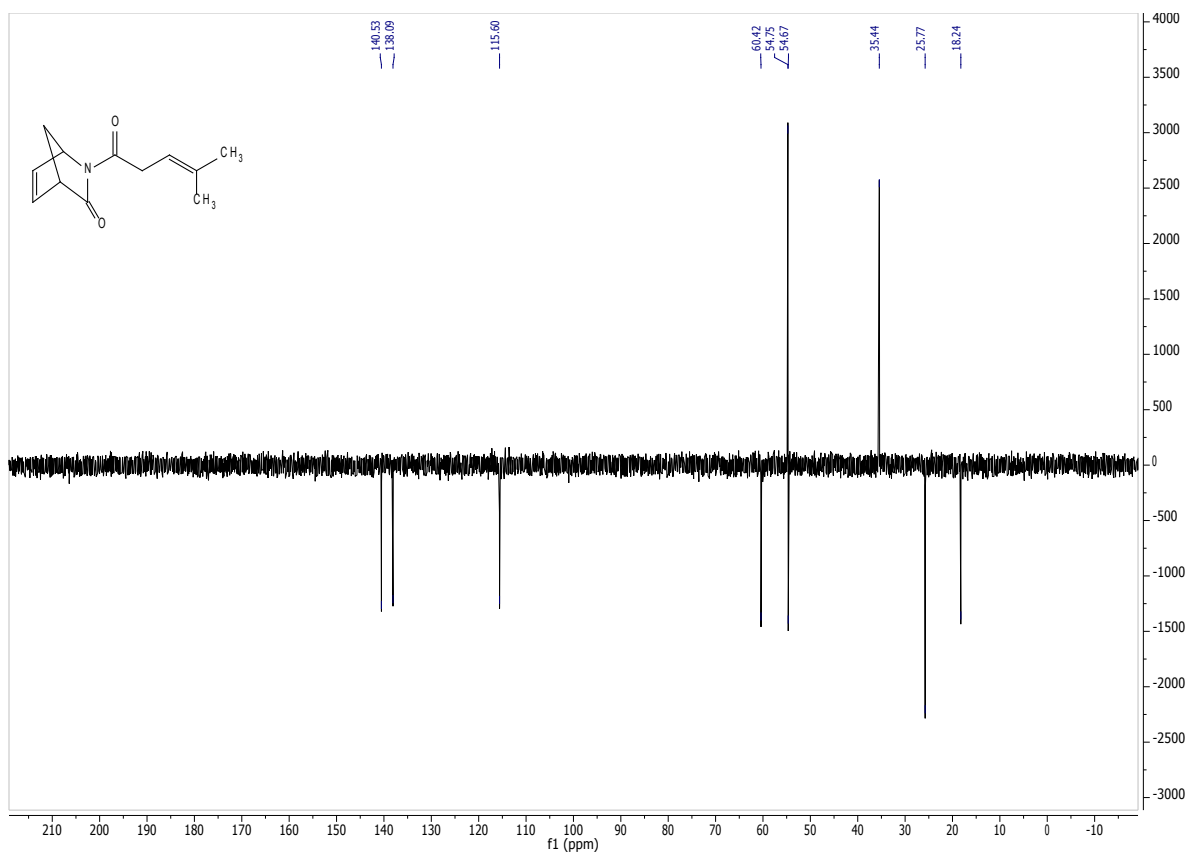


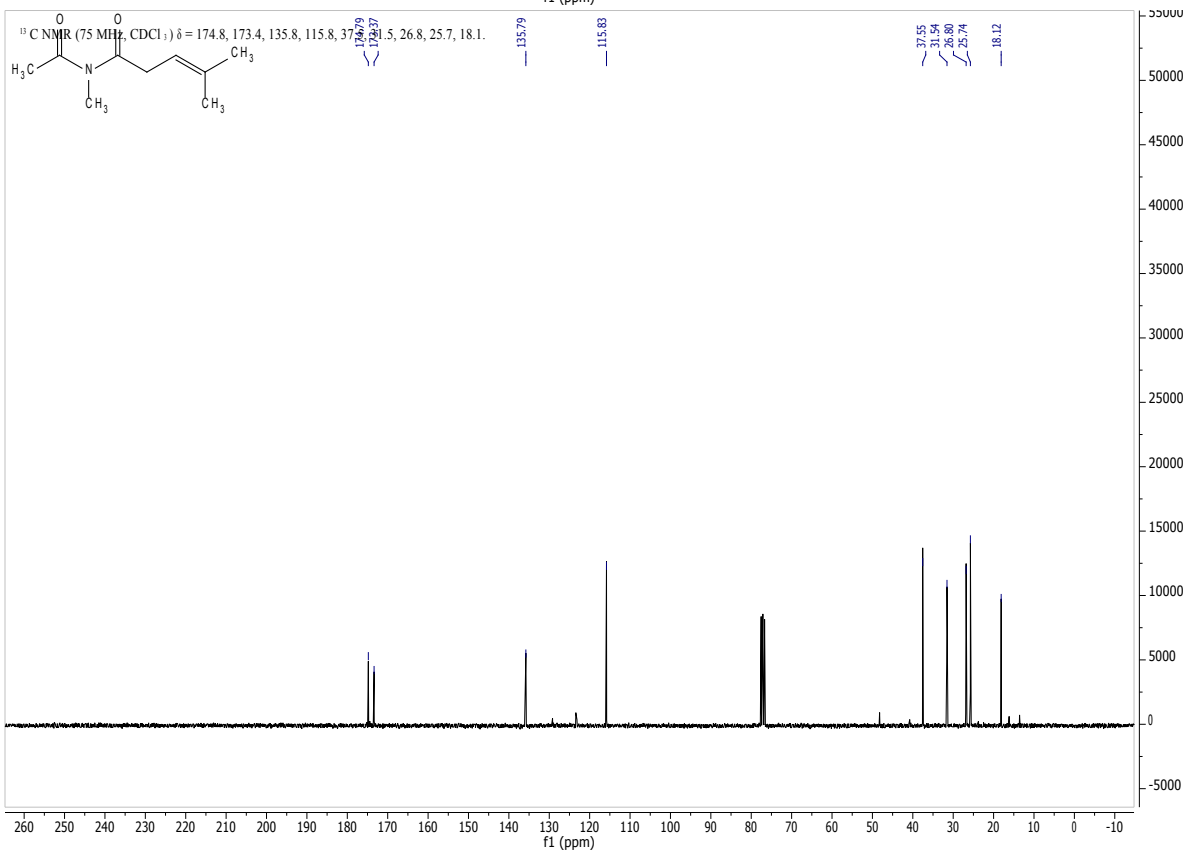
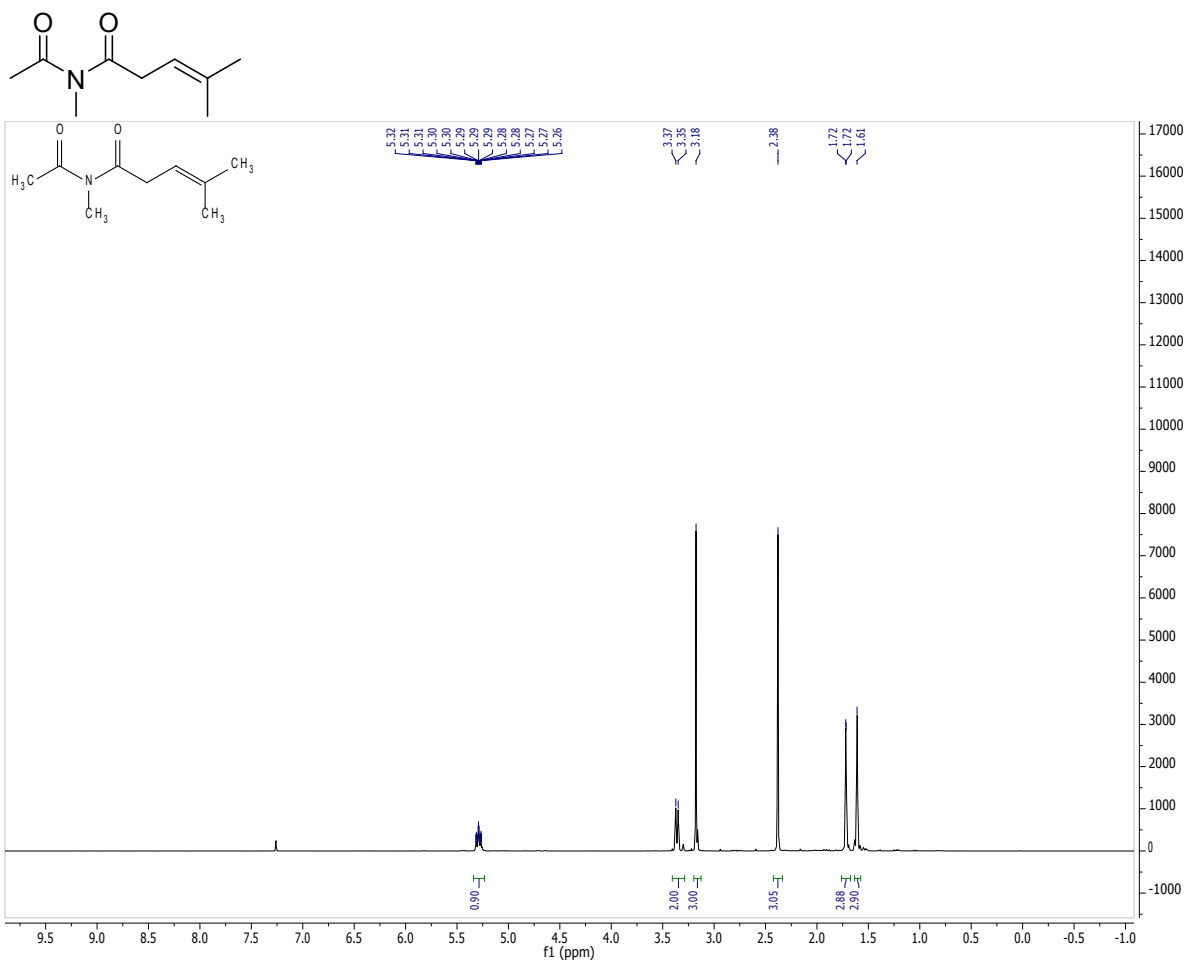


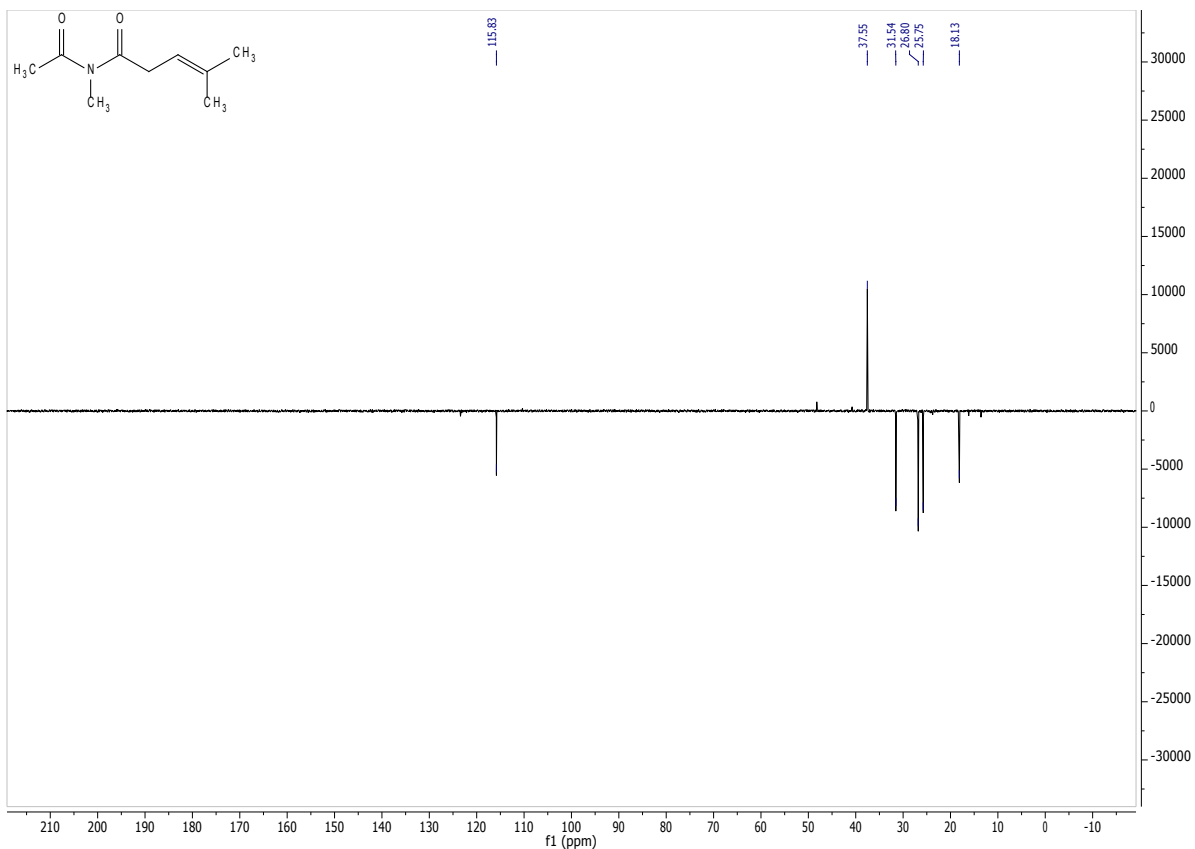
140521.f337.10.fid
LHQ657A
PROTON CDCl3 {C:\Bruker\TopSpin3.2} 1405 5

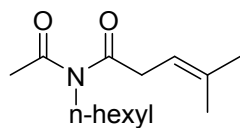




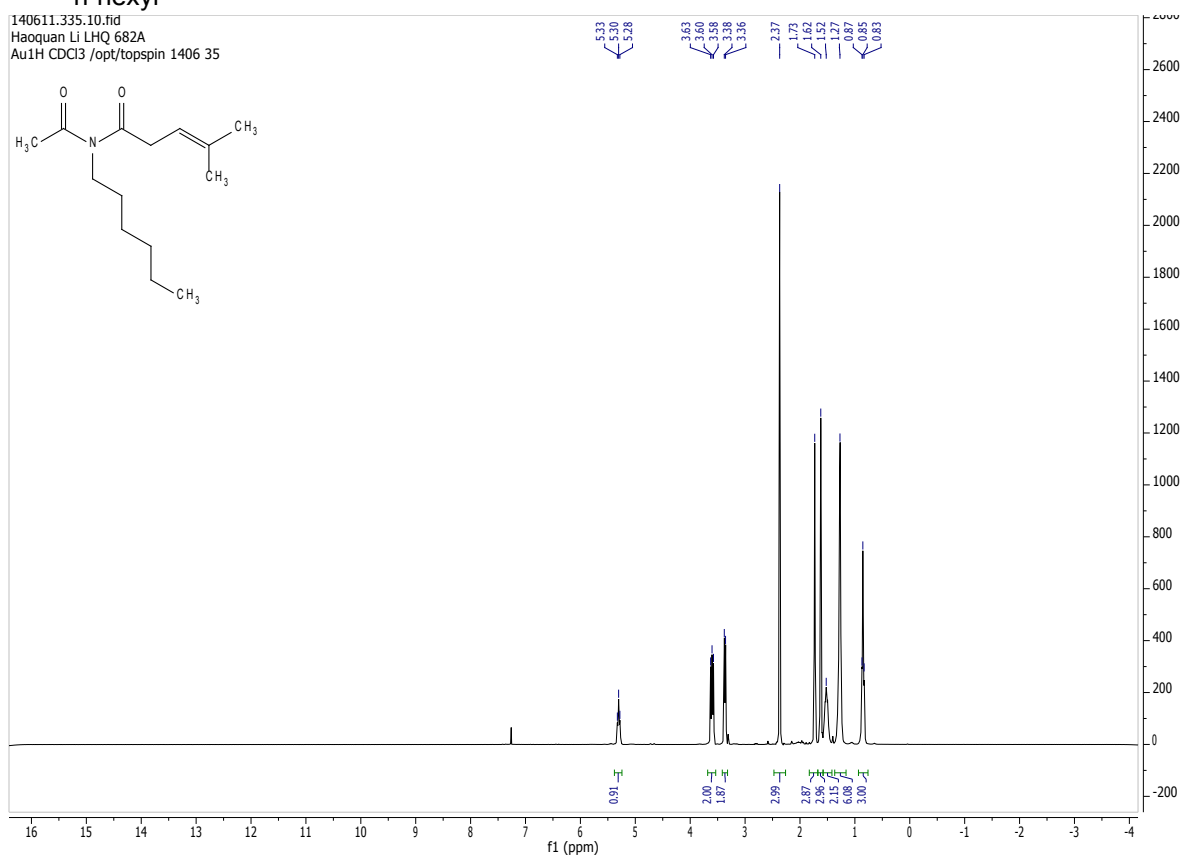




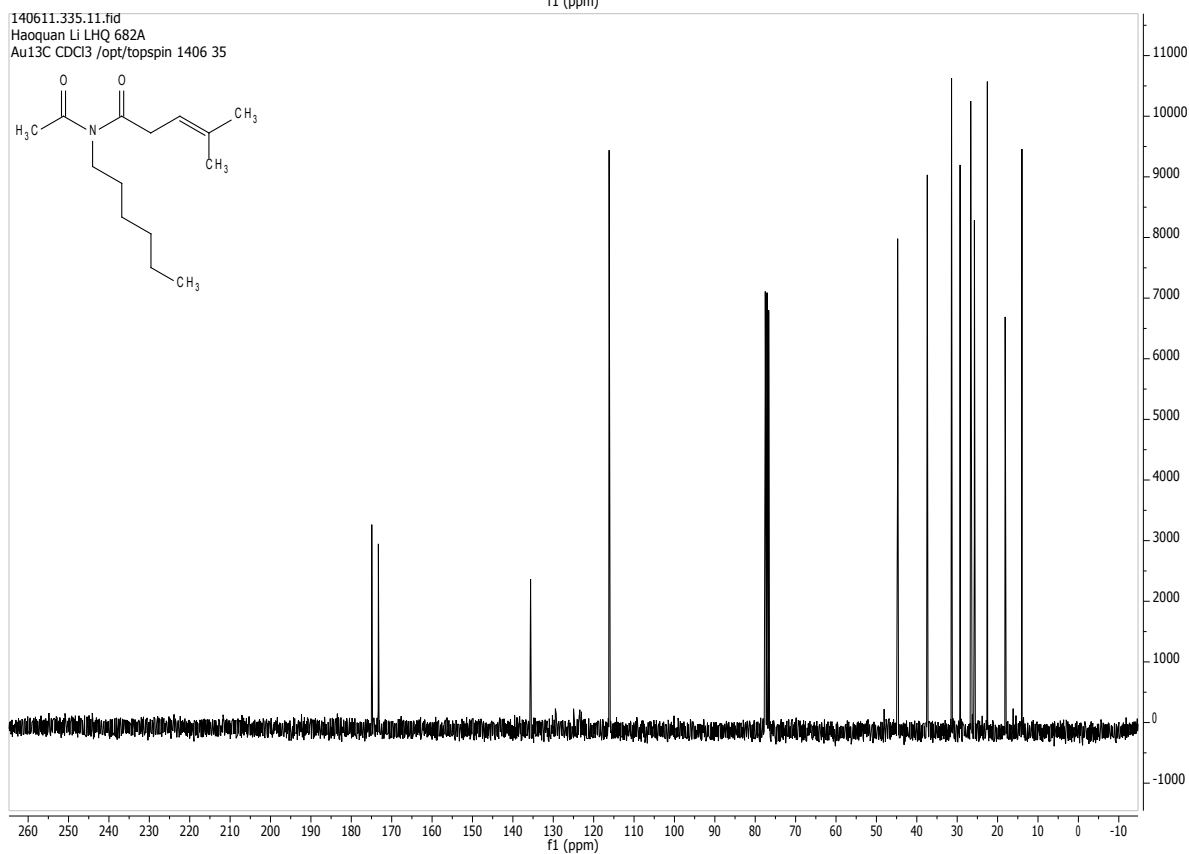




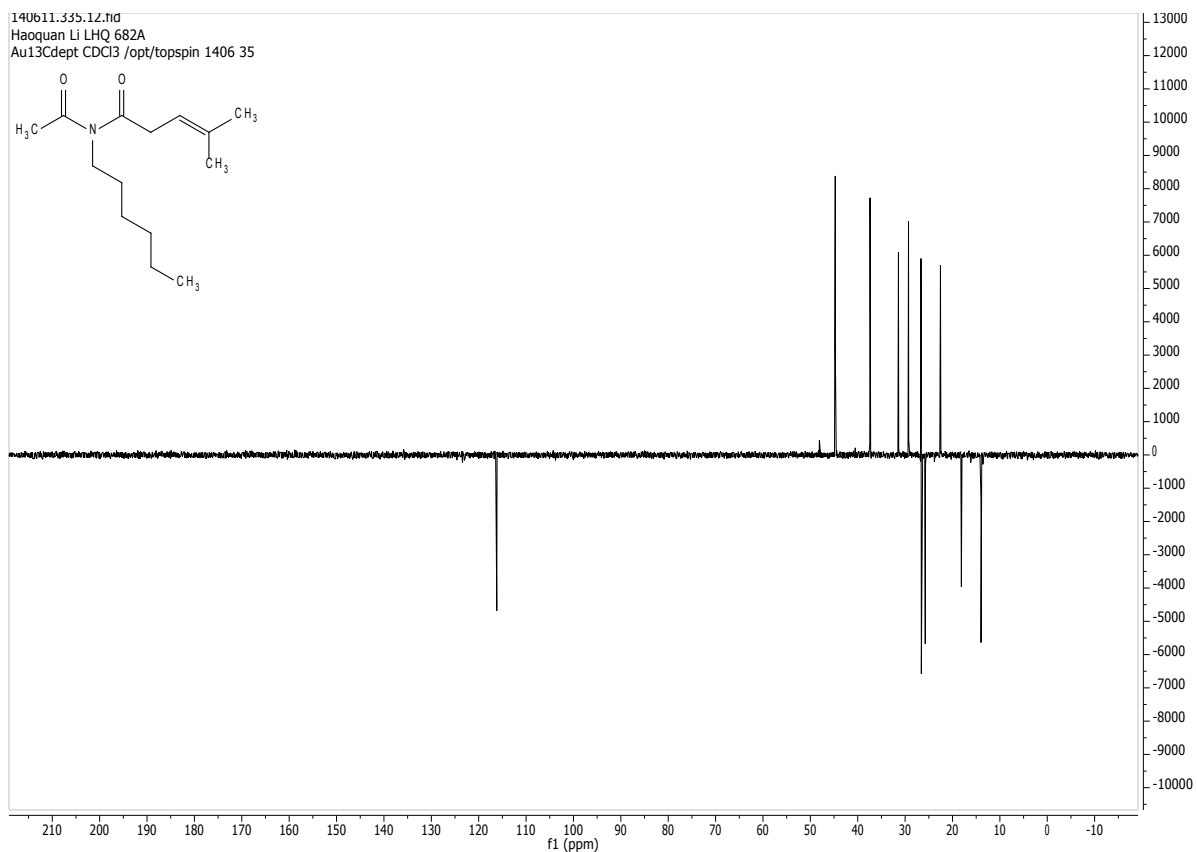
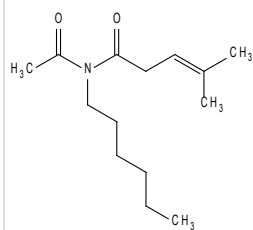
140611.335.10.fid
Haoquan Li LHQ 682A
Au1H CDCl3 /opt/topspin 1406 35

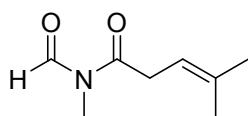


140611.335.11.fid
Haoquan Li LHQ 682A
Au13C CDCl3 /opt/topspin 1406 35

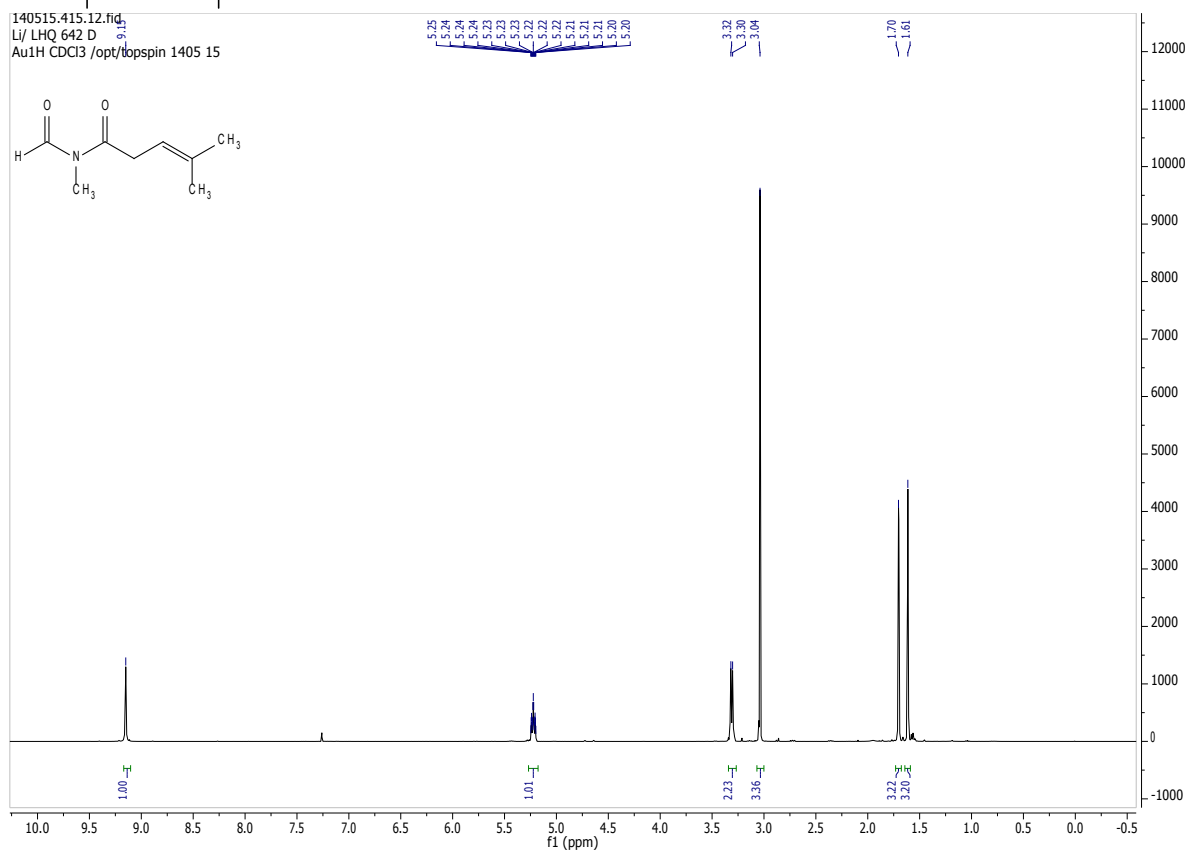
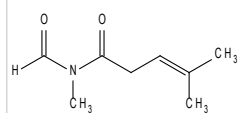


140611.335.12.nd
Haoquan Li LHQ 682A
Au13Cdept CDCl3 /opt/topspin 1406 35

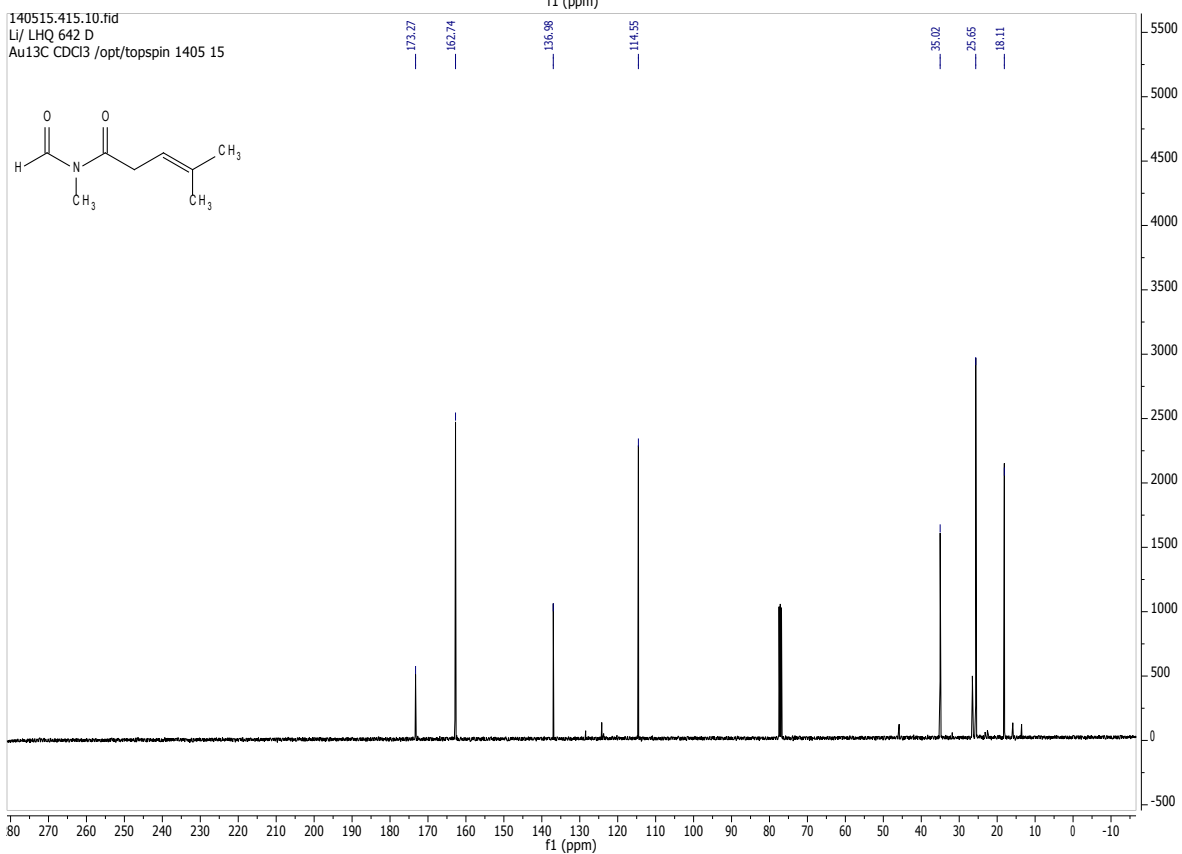
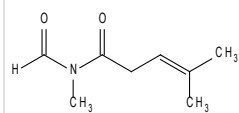


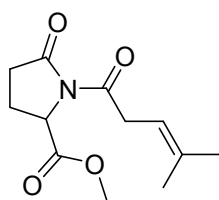


140515.415.12.fid
Li/ LHQ 642 D
Au1H CDCl3 /opt/topspin 1405 15

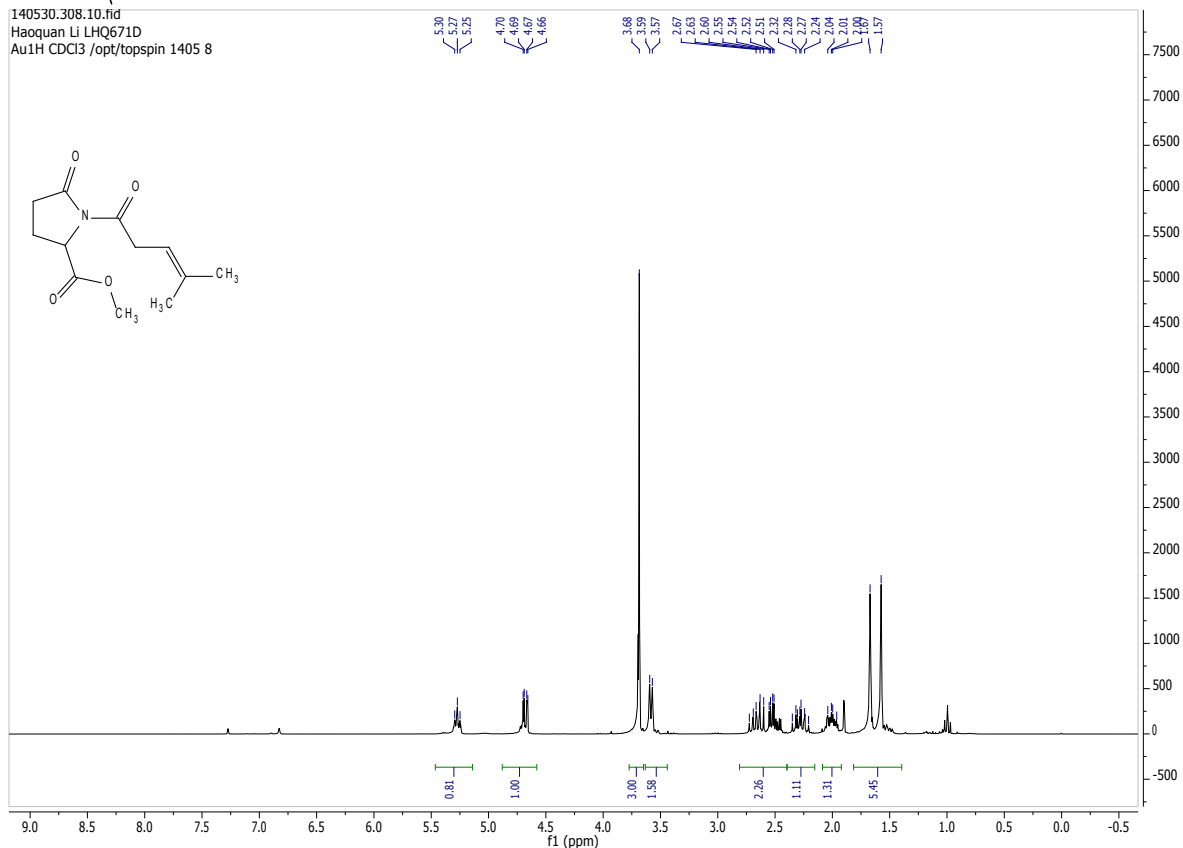


140515.415.10.fid
Li/ LHQ 642 D
Au13C CDCl3 /opt/topspin 1405 15

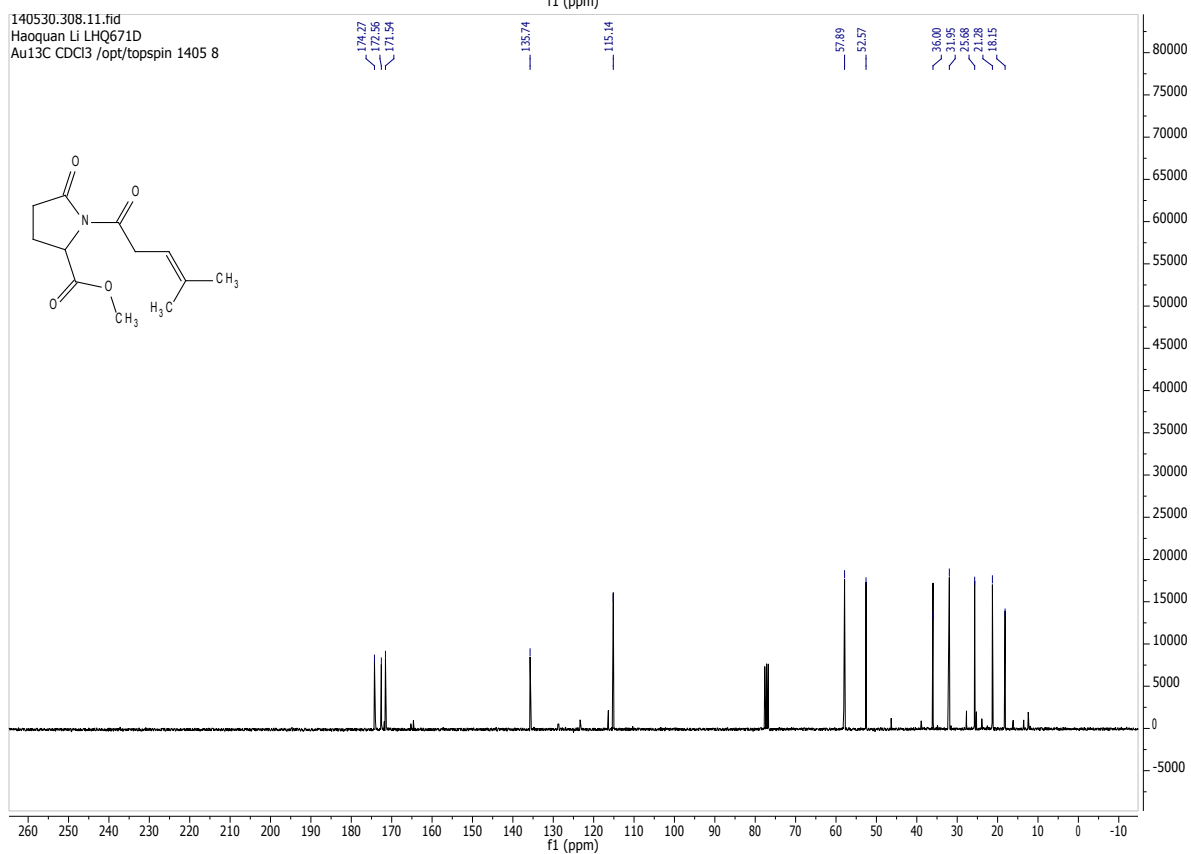


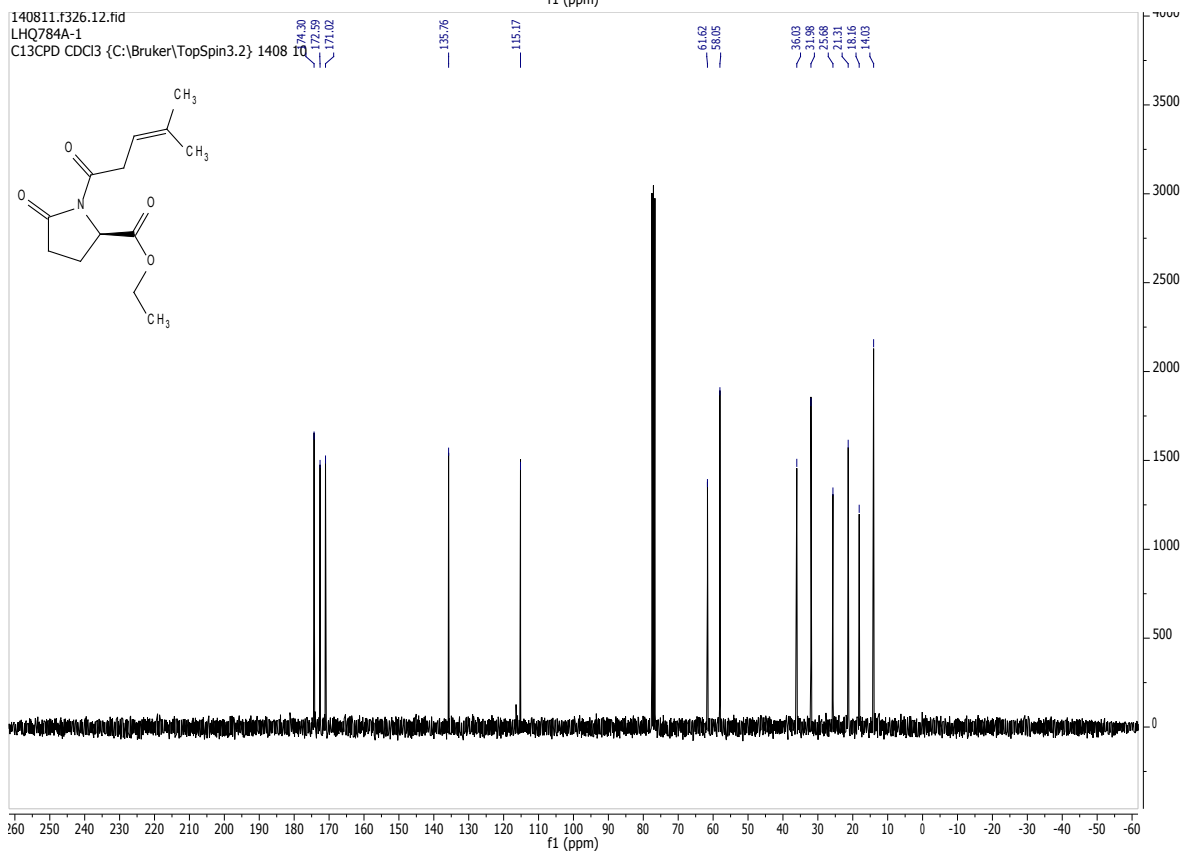
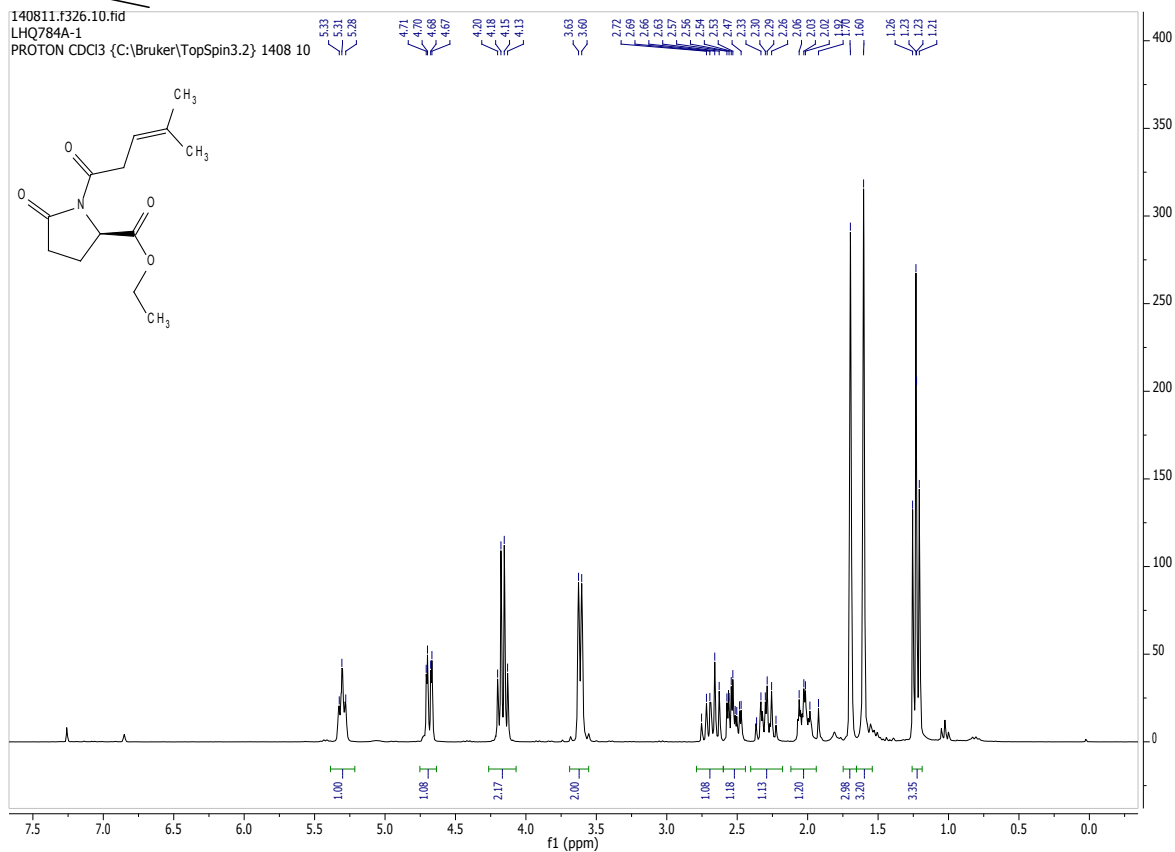
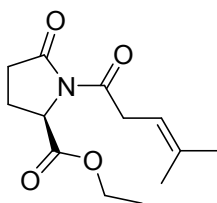


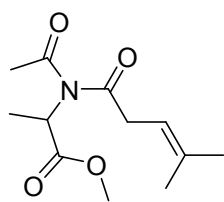
140530.308.10.fid
Haoquan Li LHQ671D
Au1H CDCl3 /opt/topspin 1405 8



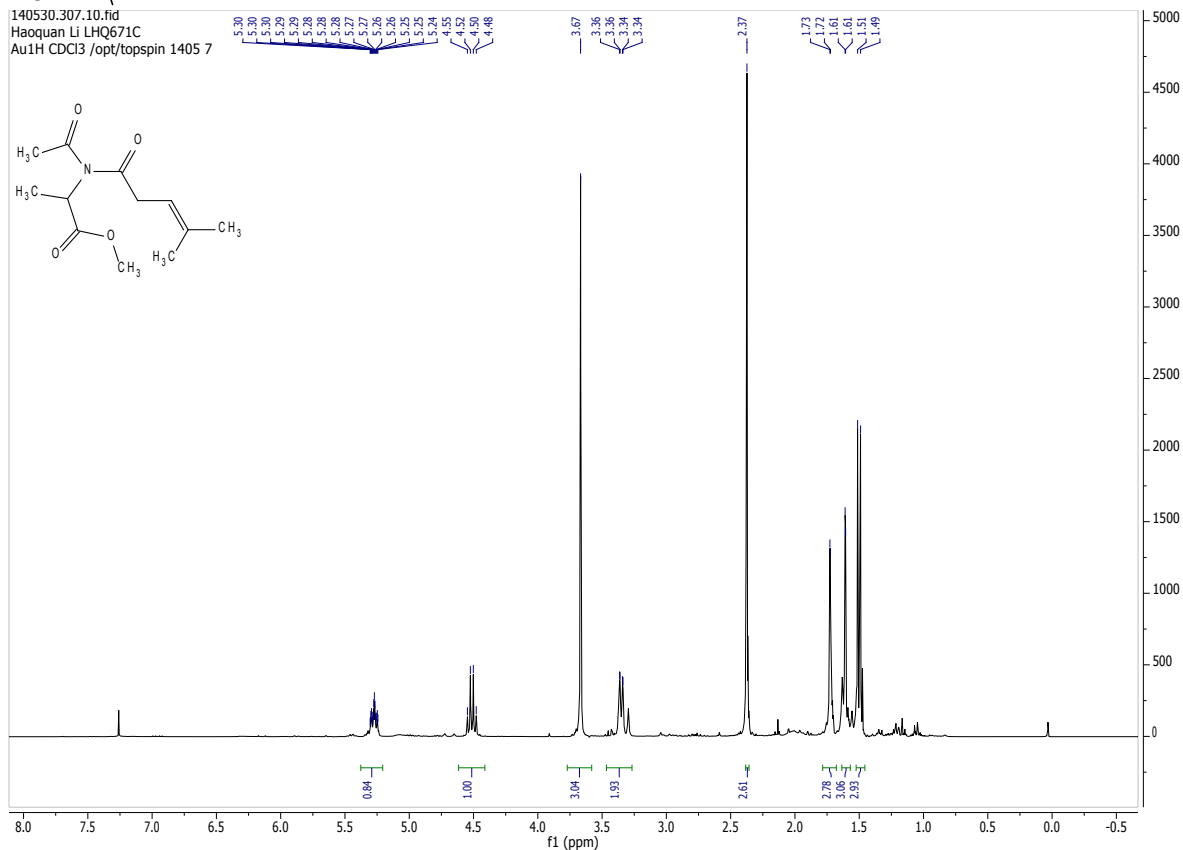
140530.308.11.fid
Haoquan Li LHQ671D
Au13C CDCl3 /opt/topspin 1405 8



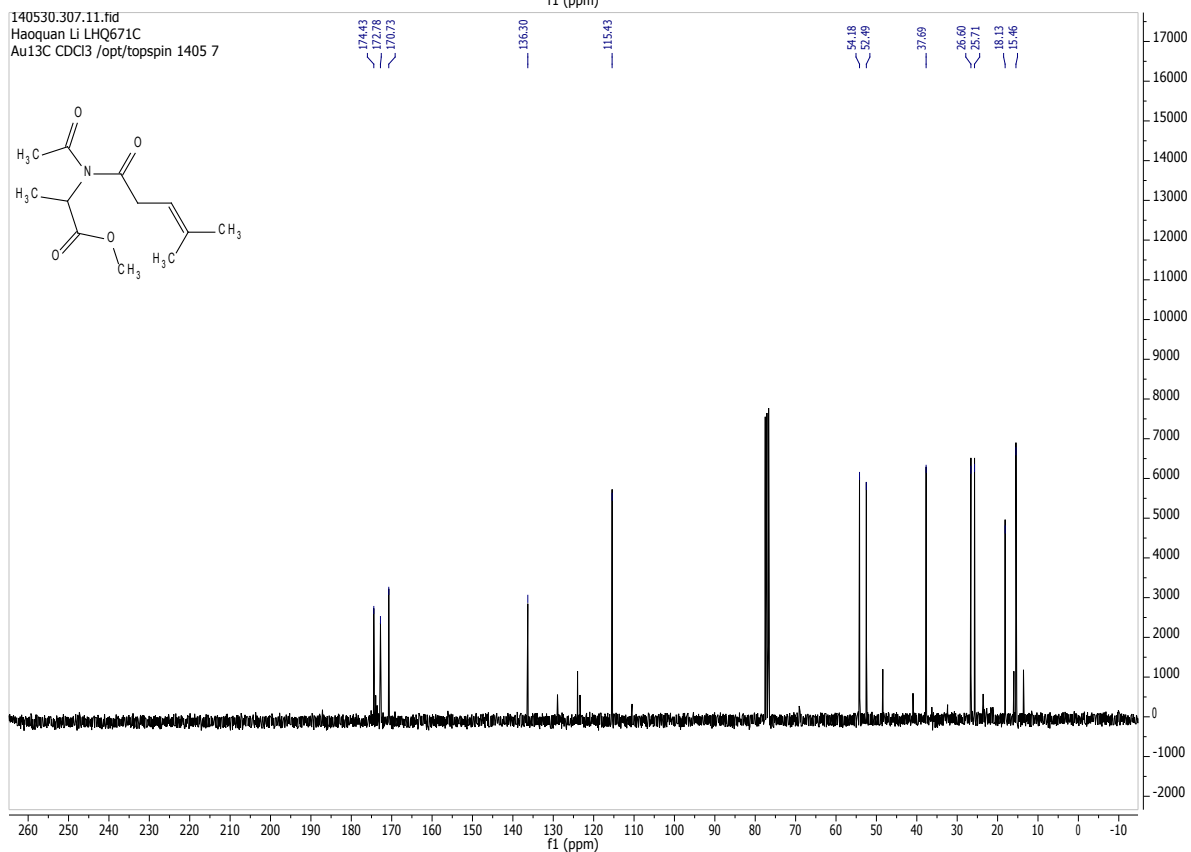


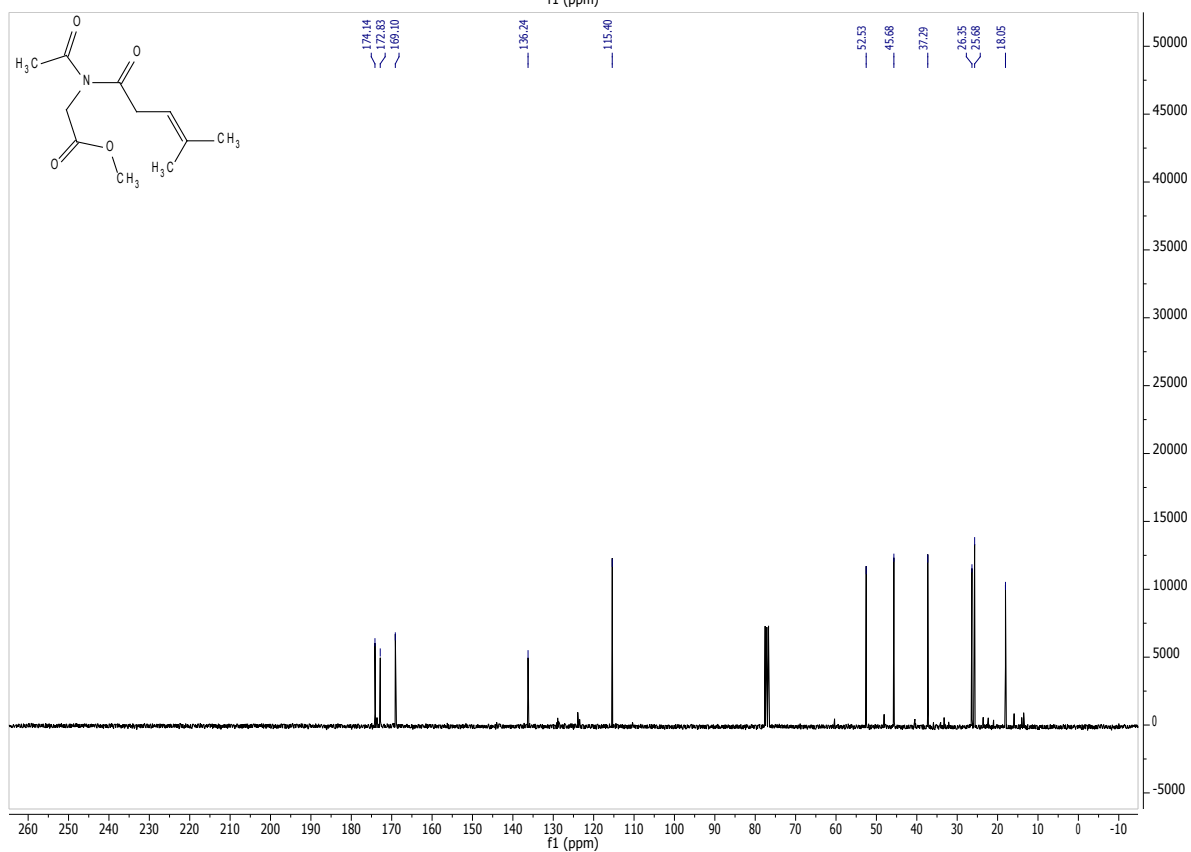
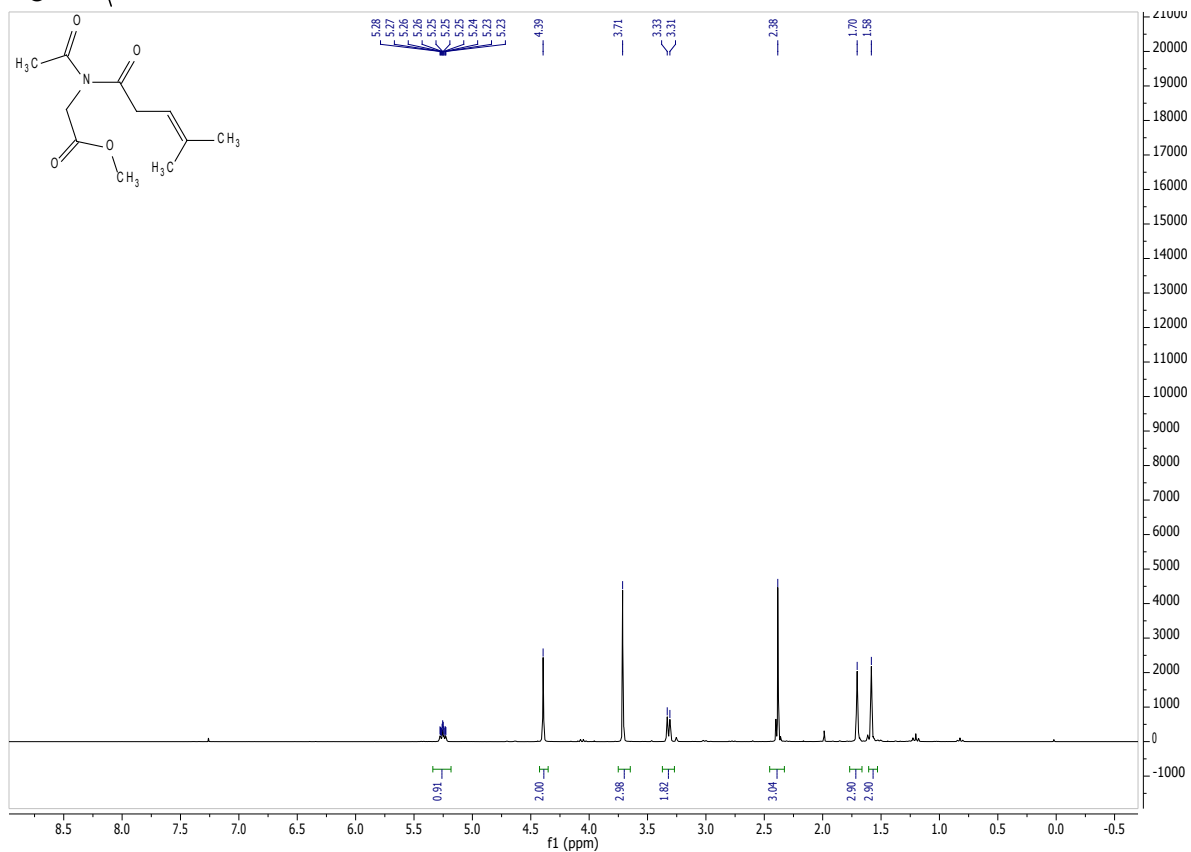
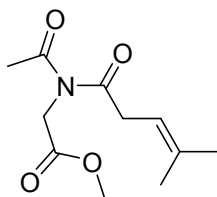


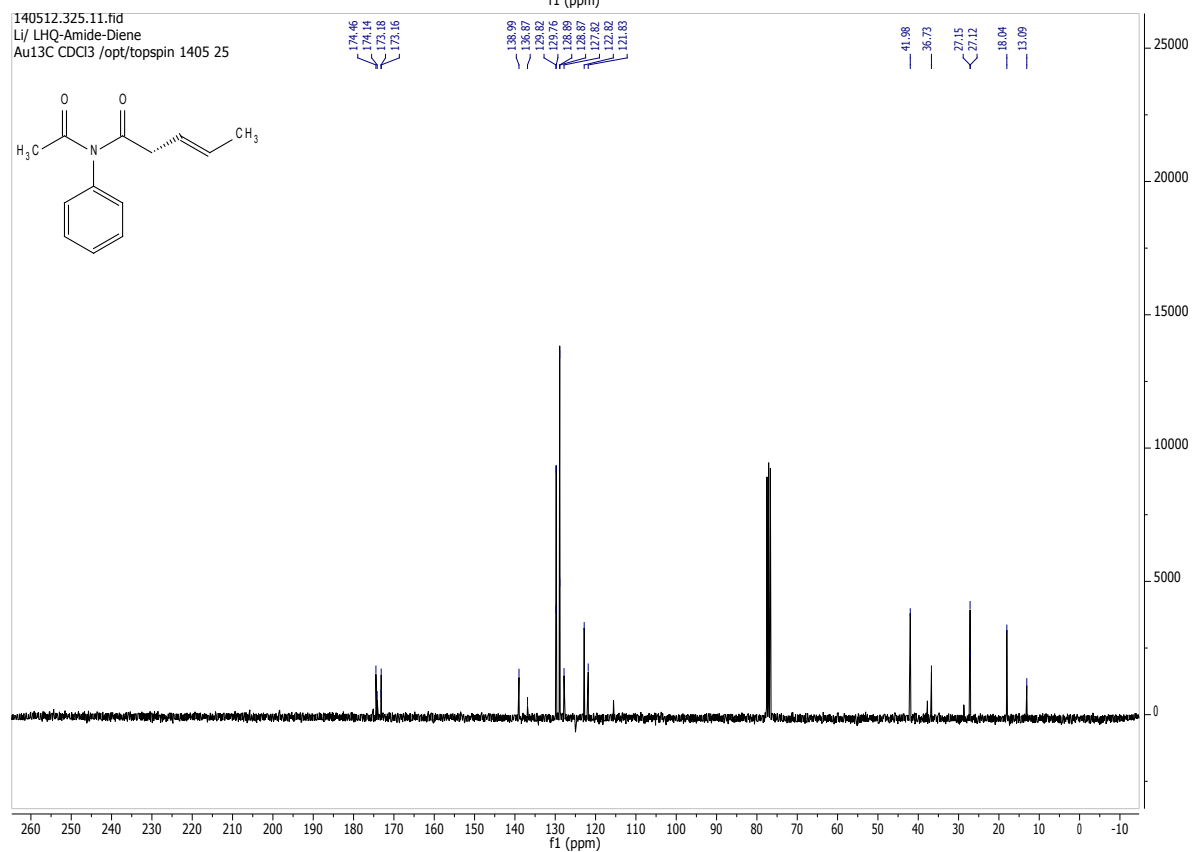
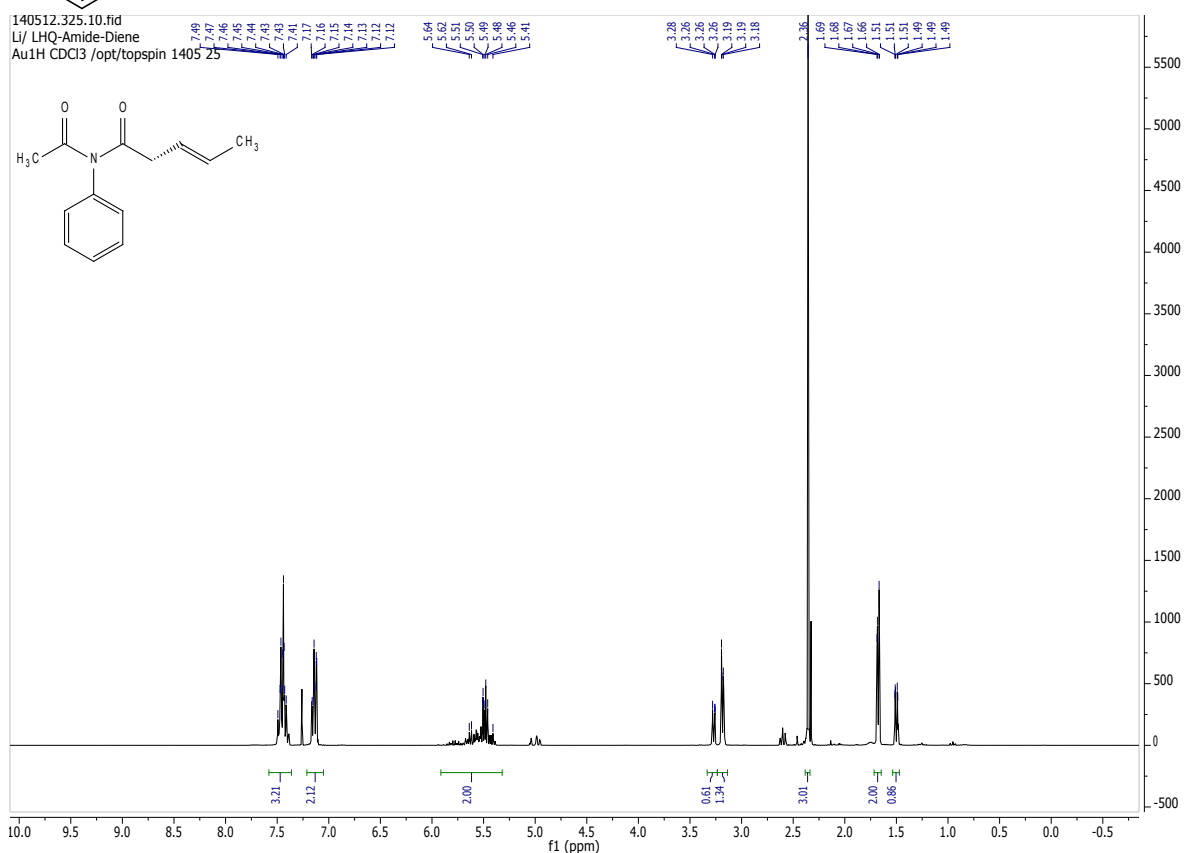
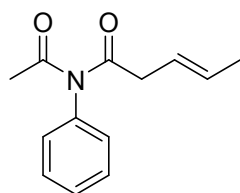
140530.307.10.fid
Haoquan Li LHQ671C
Au1H CDCl3 /opt/topspin 1405 7

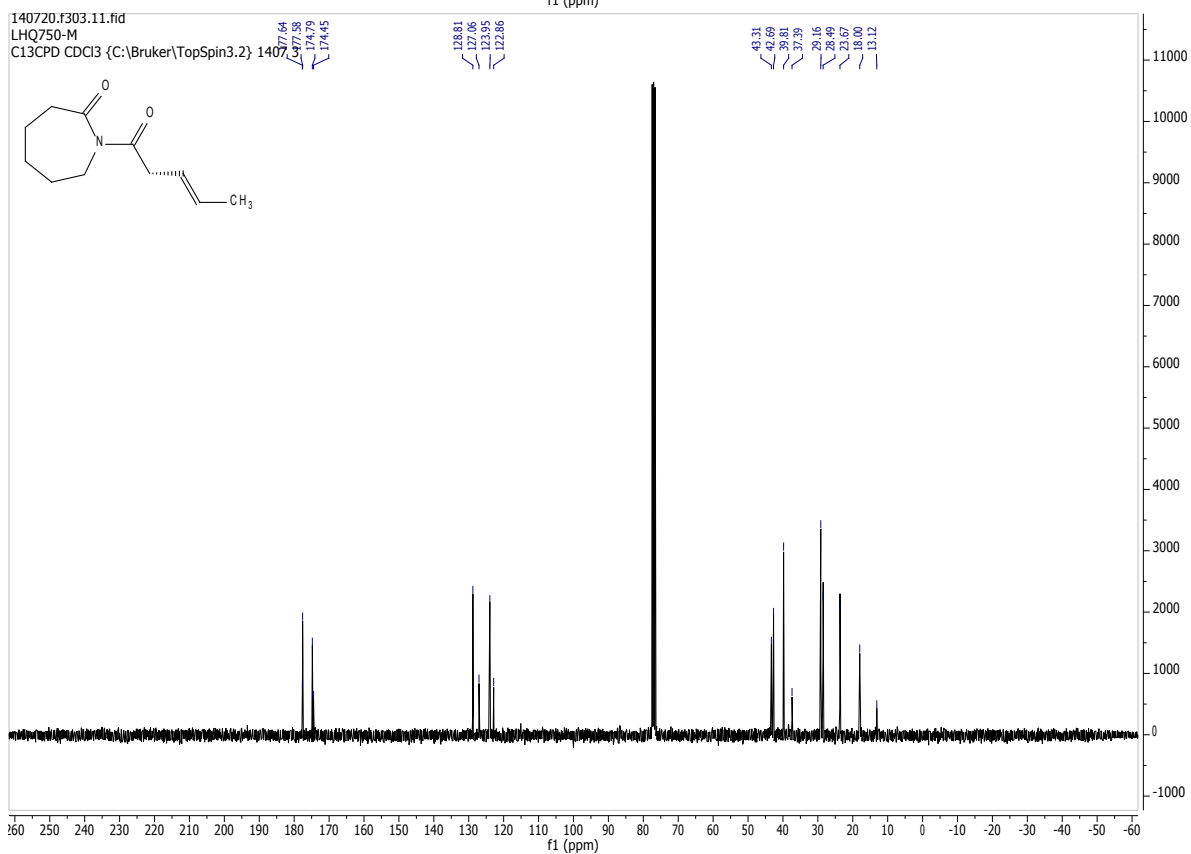
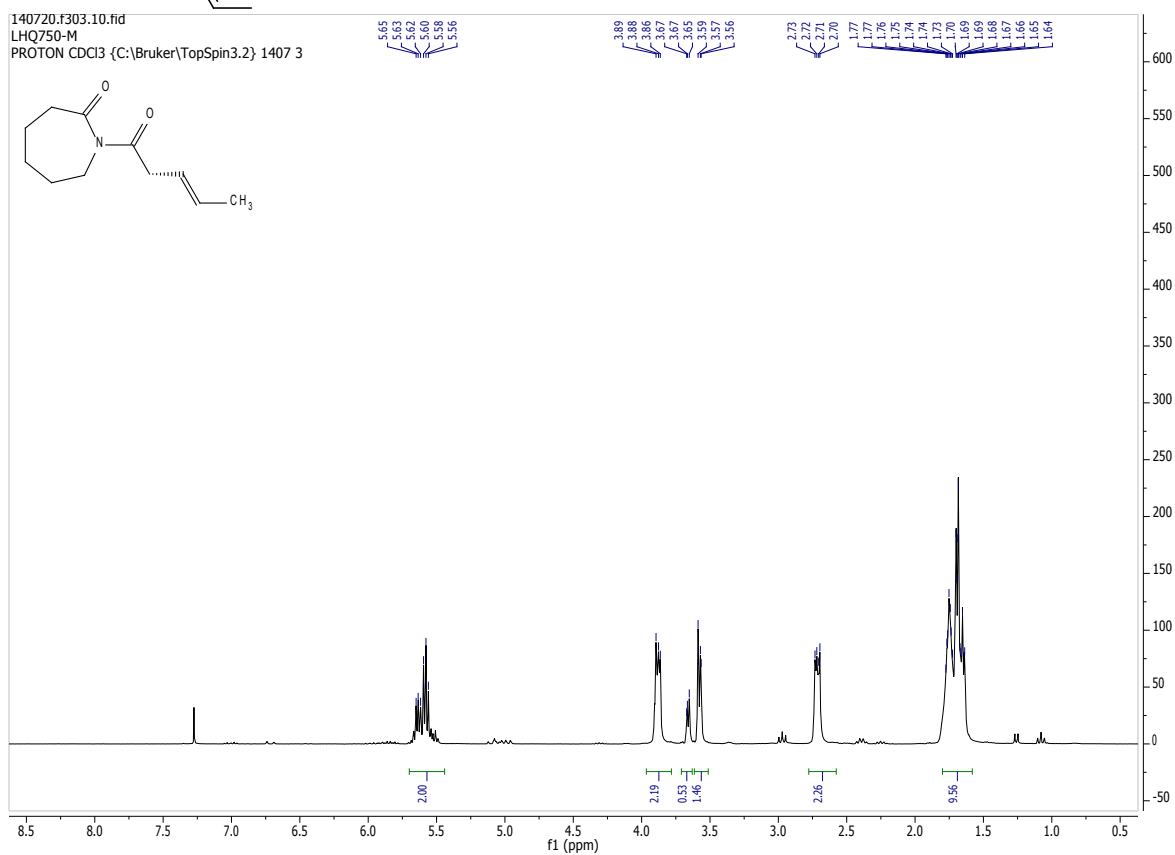
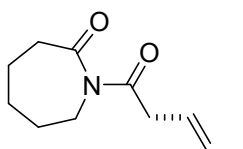


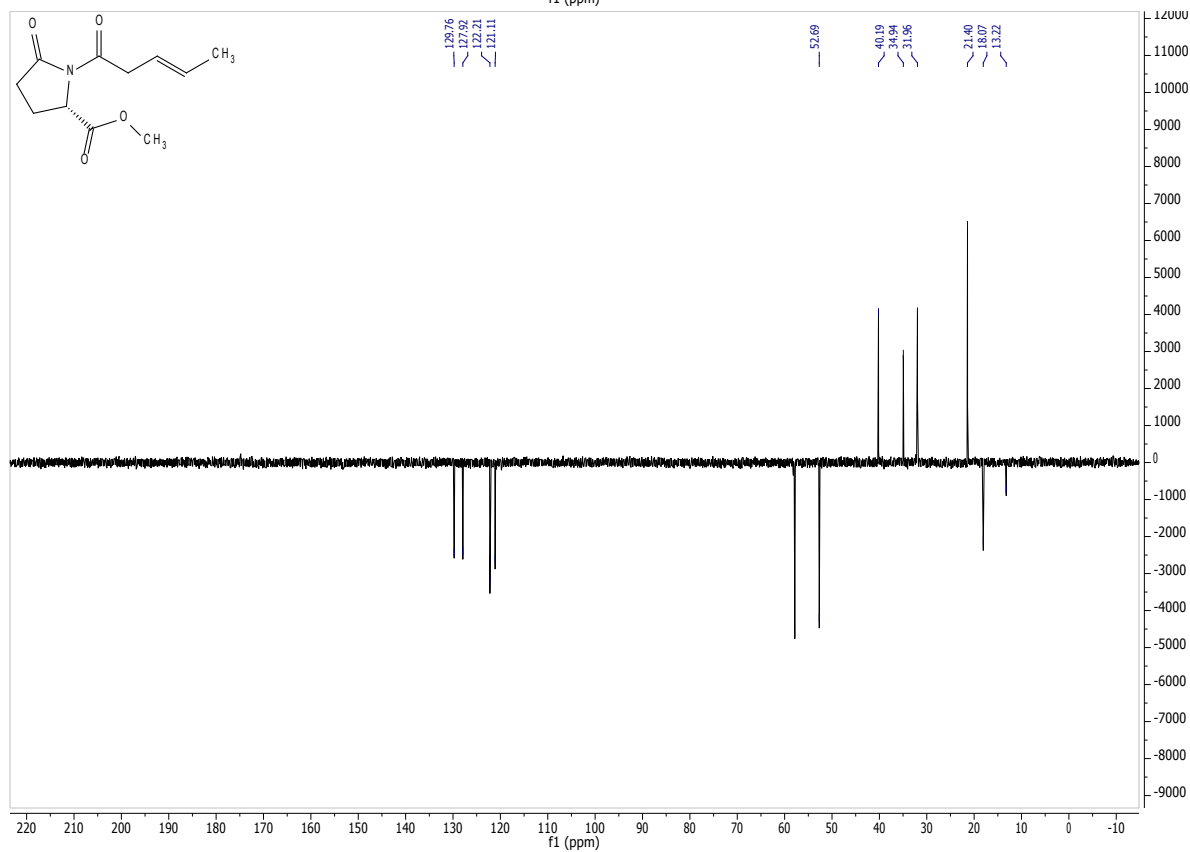
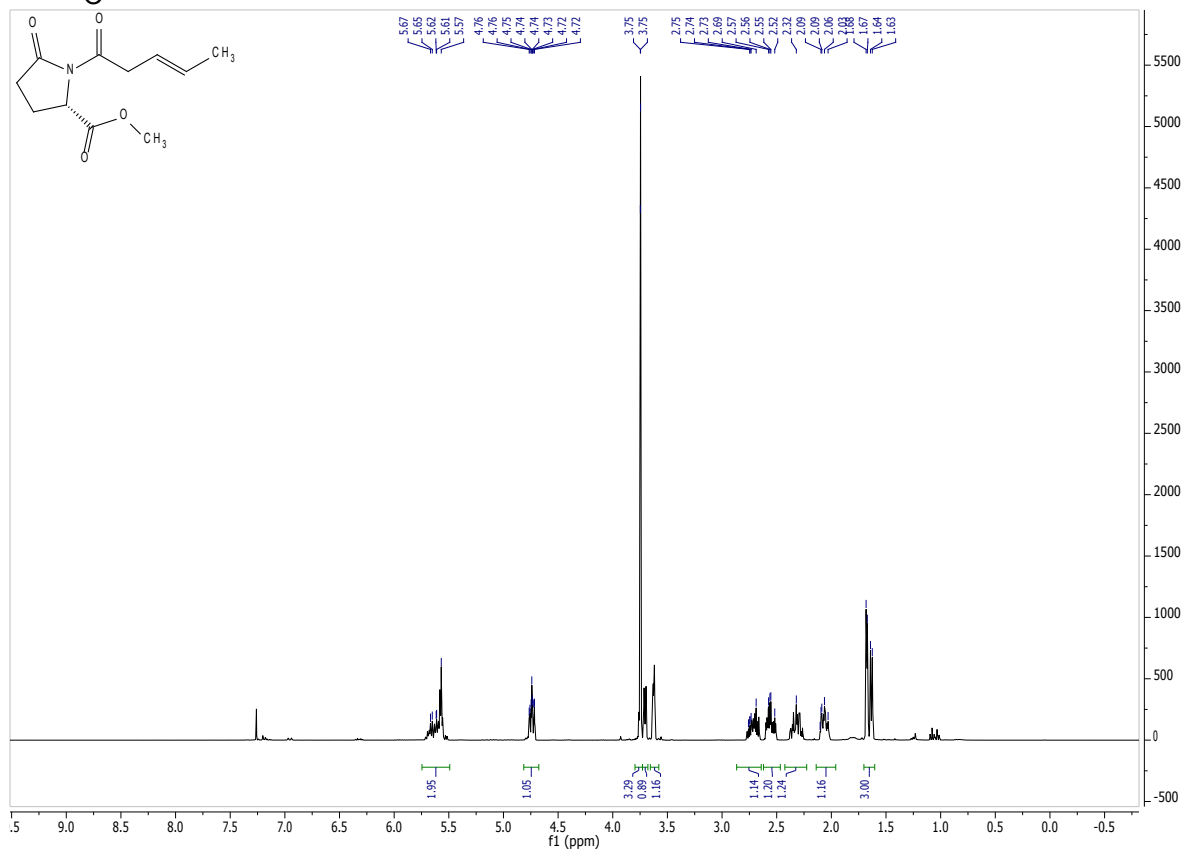
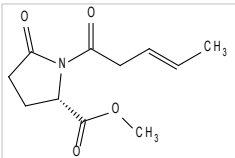
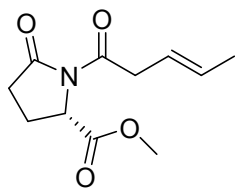
140530.307.11.fid
Haoquan Li LHQ671C
Au13C CDCl3 /opt/topspin 1405 7

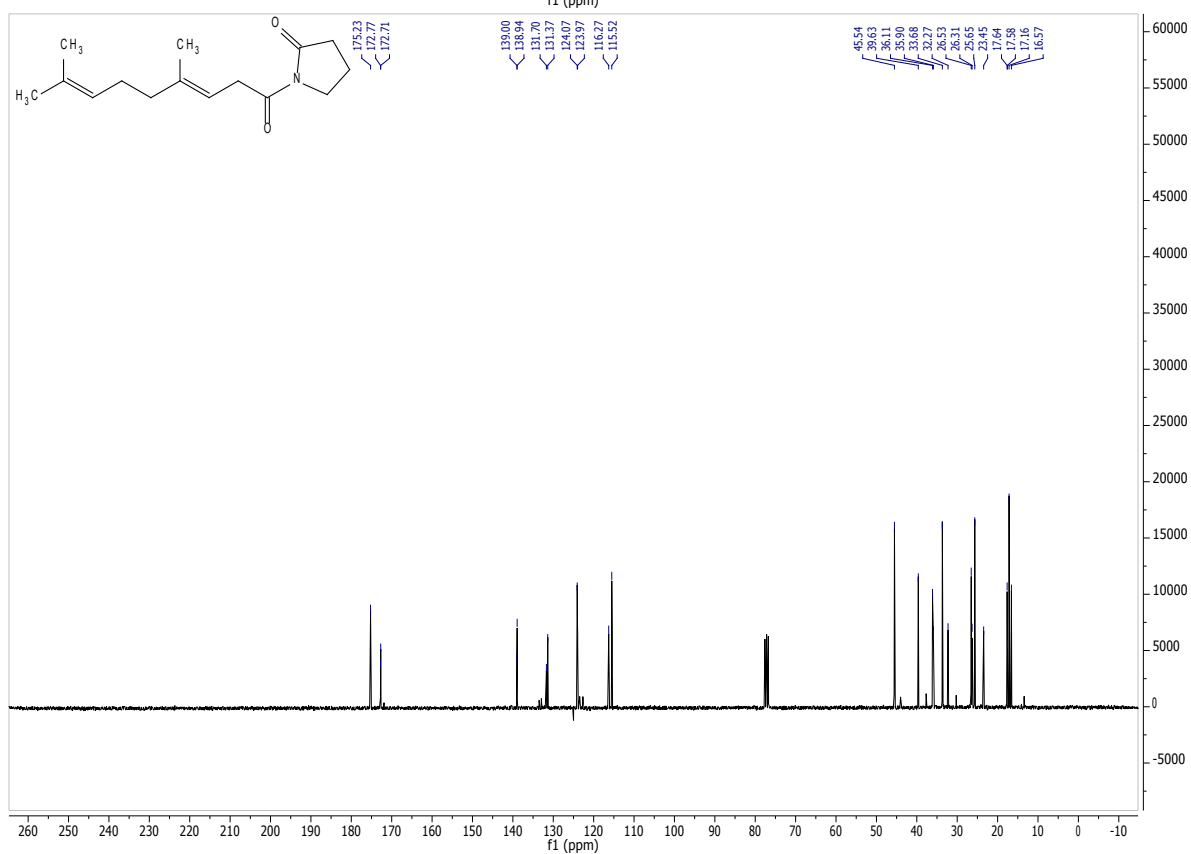
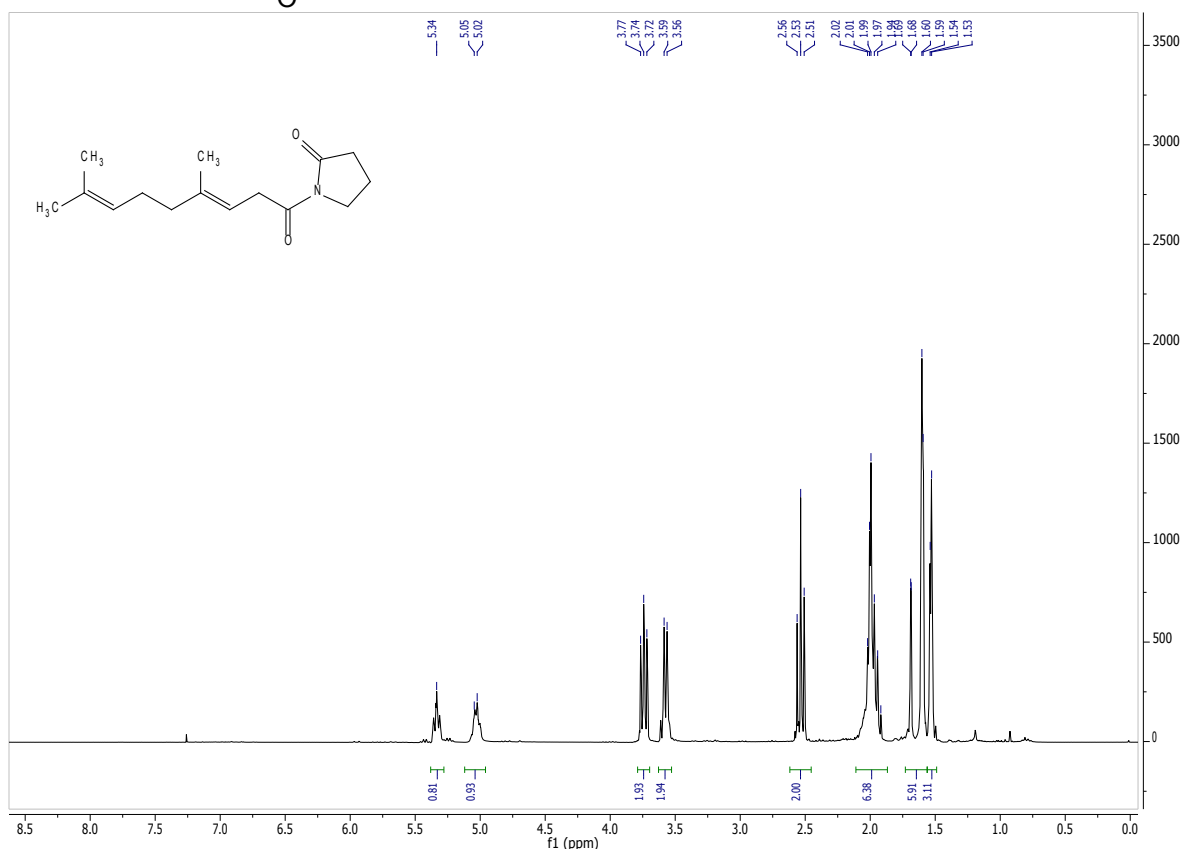
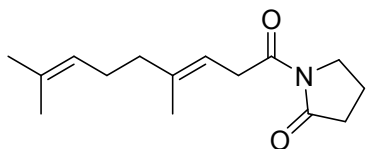


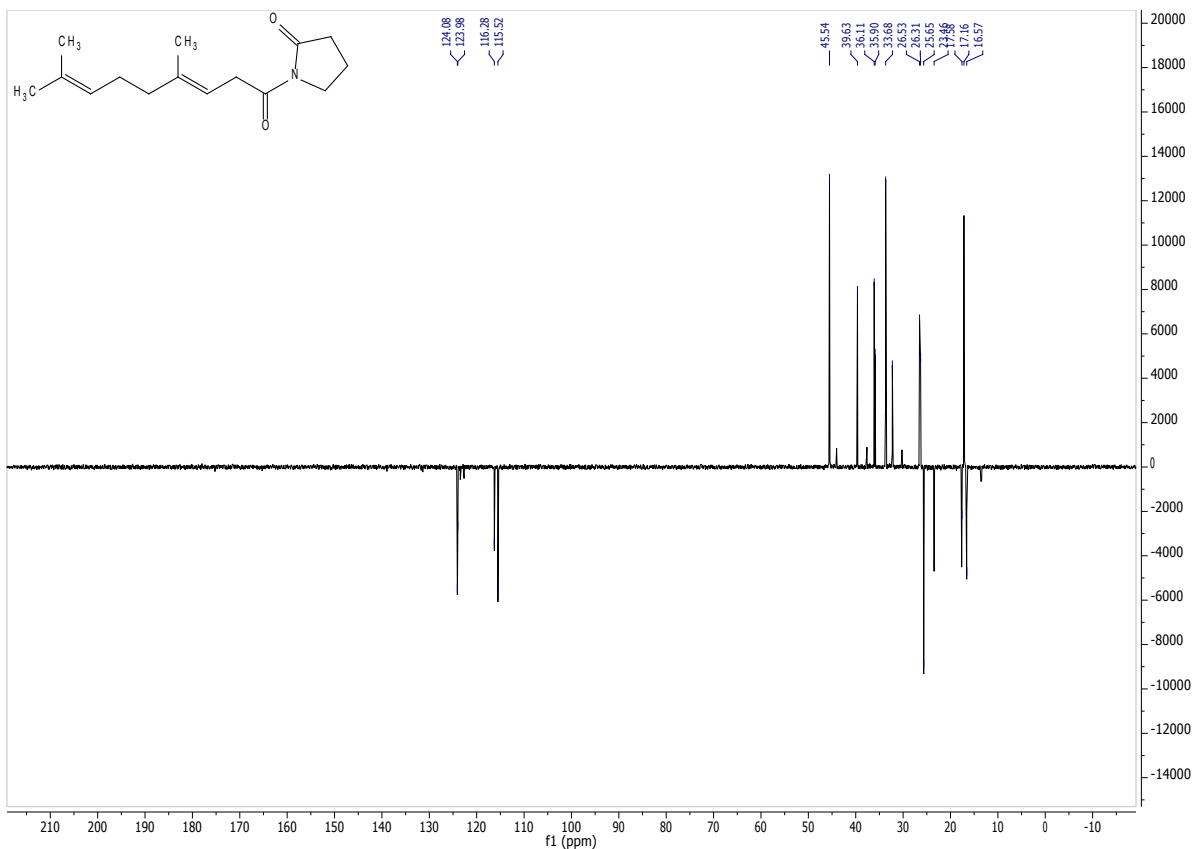


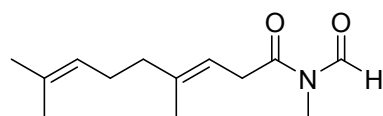




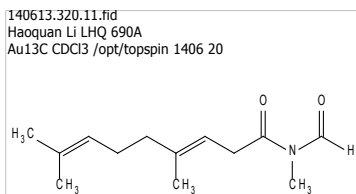
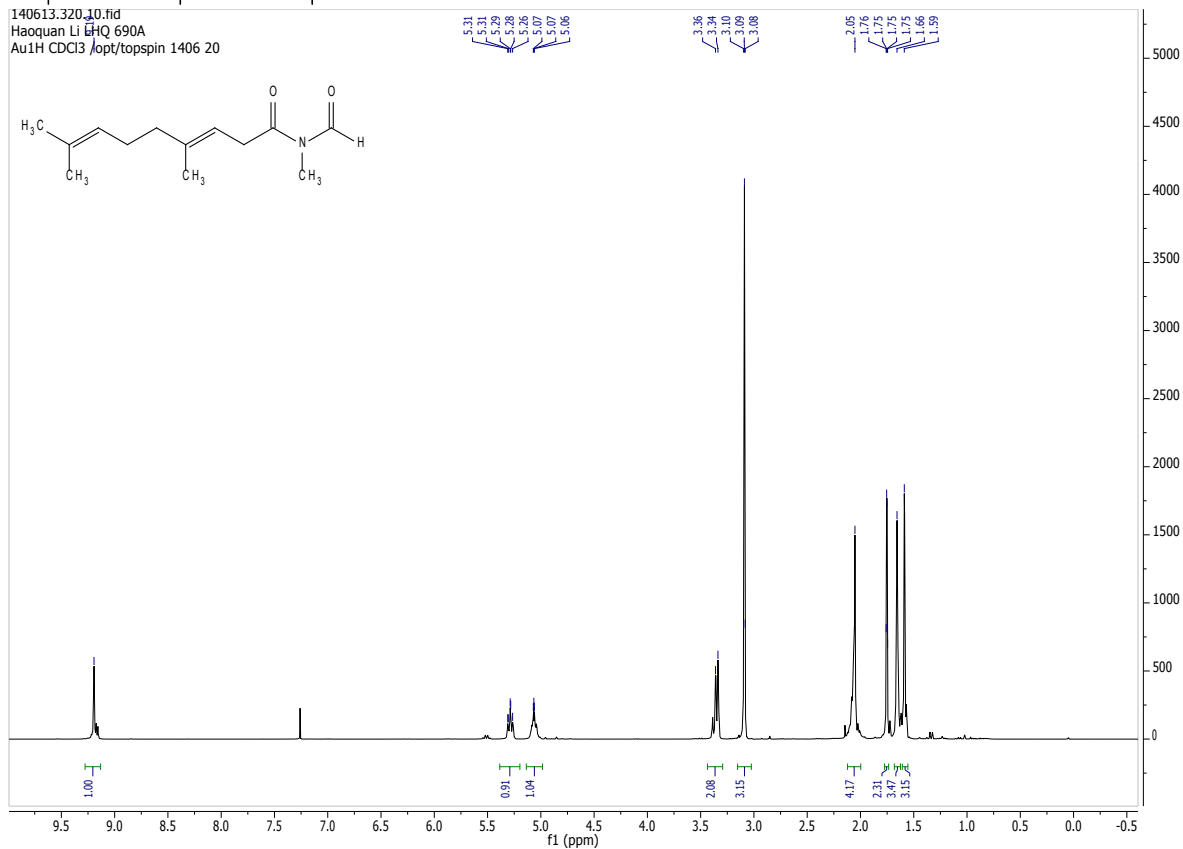
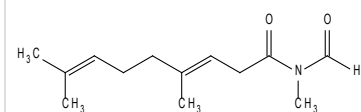




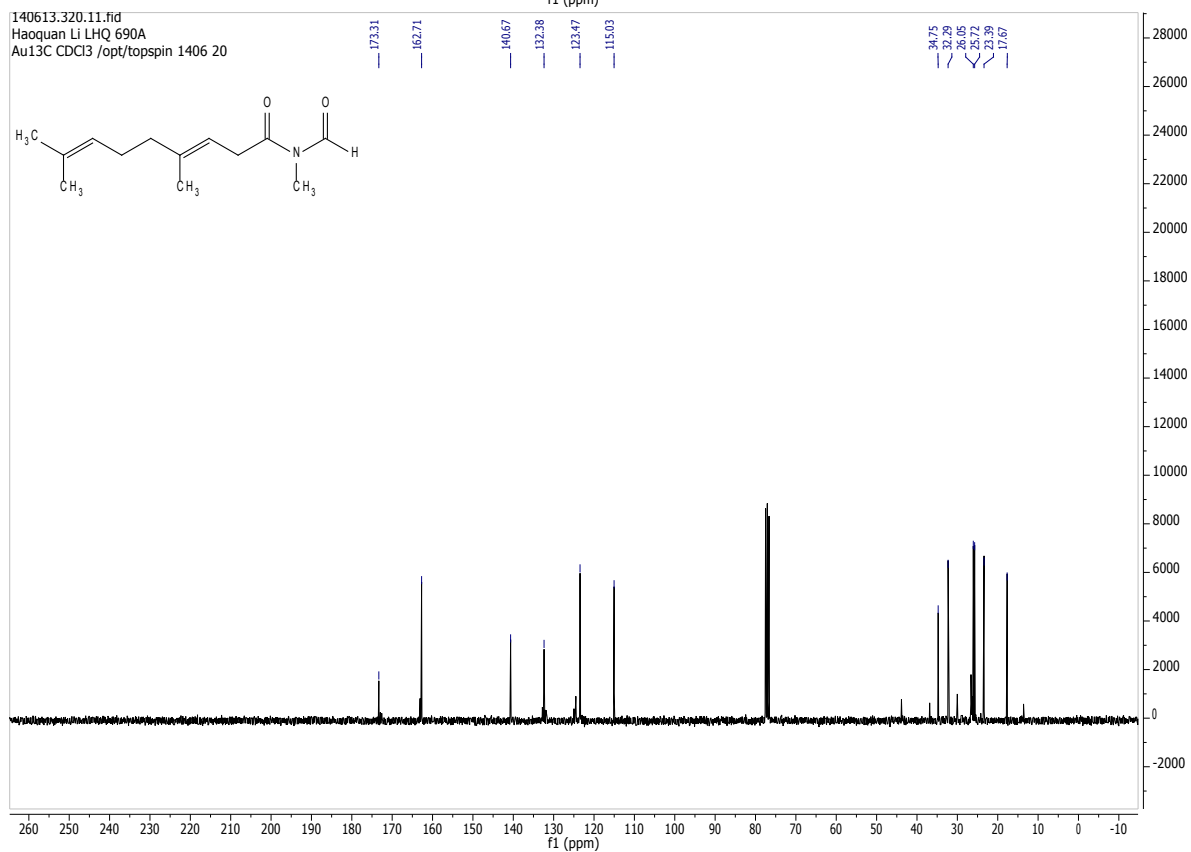


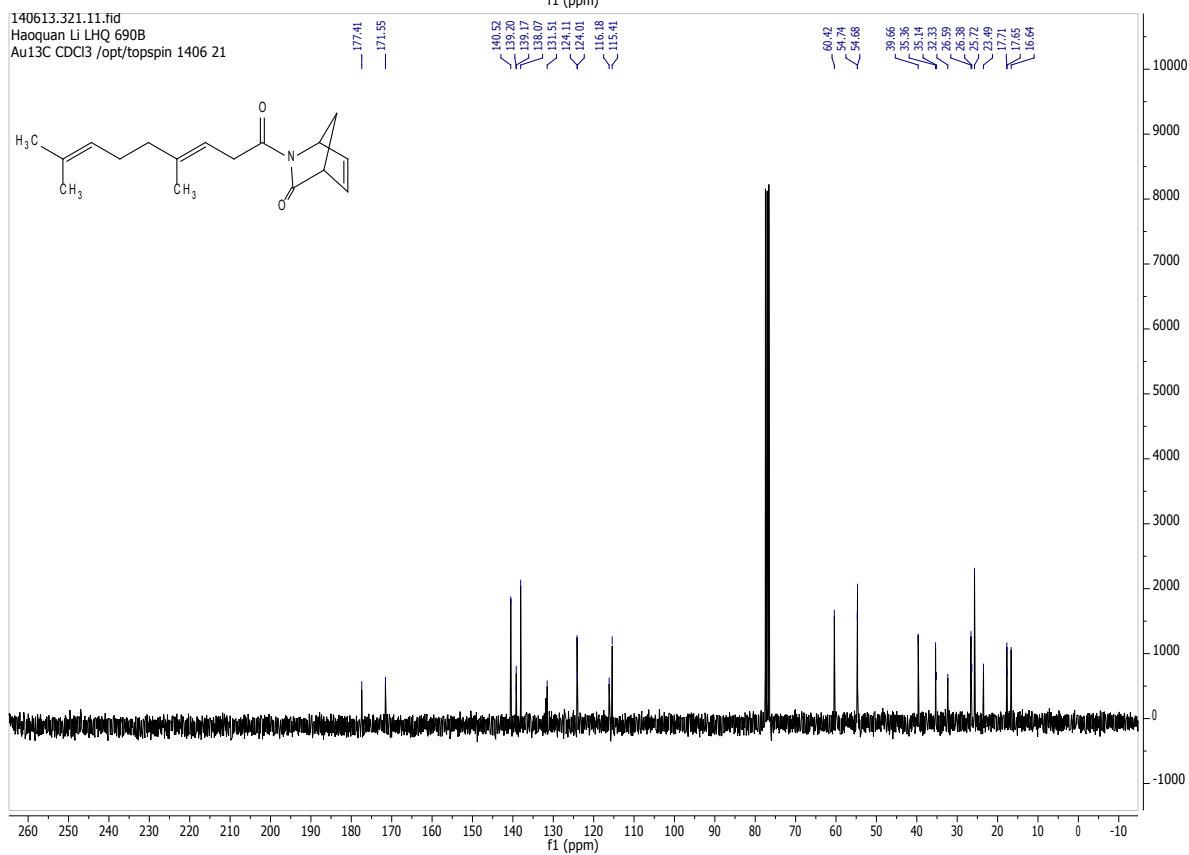
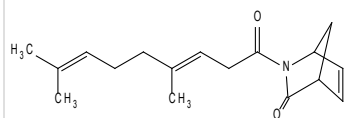
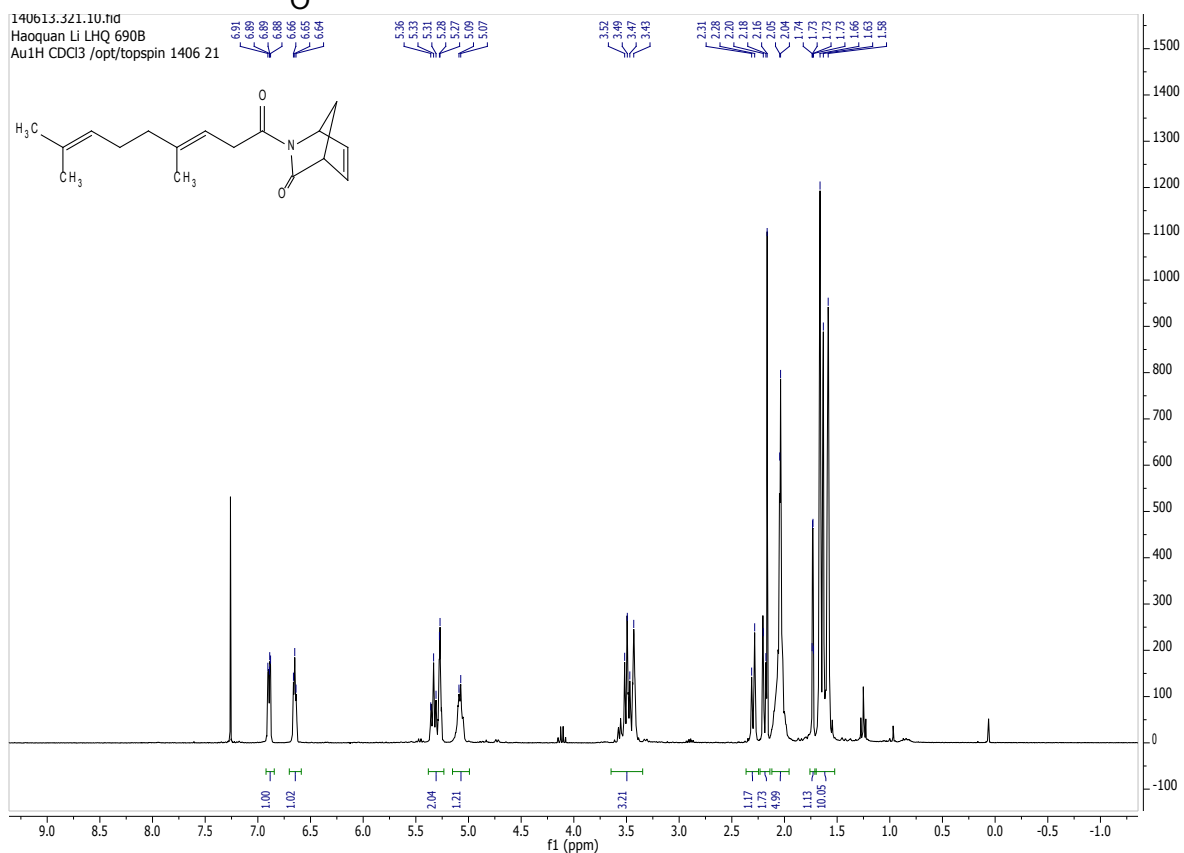
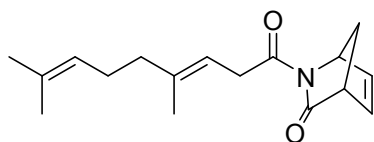


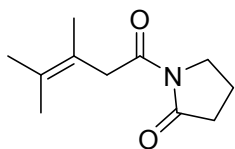
140613.320.10.fid
Haoquan Li LHQ 690A
Au1H CDCl3 /opt/topspin 1406 20



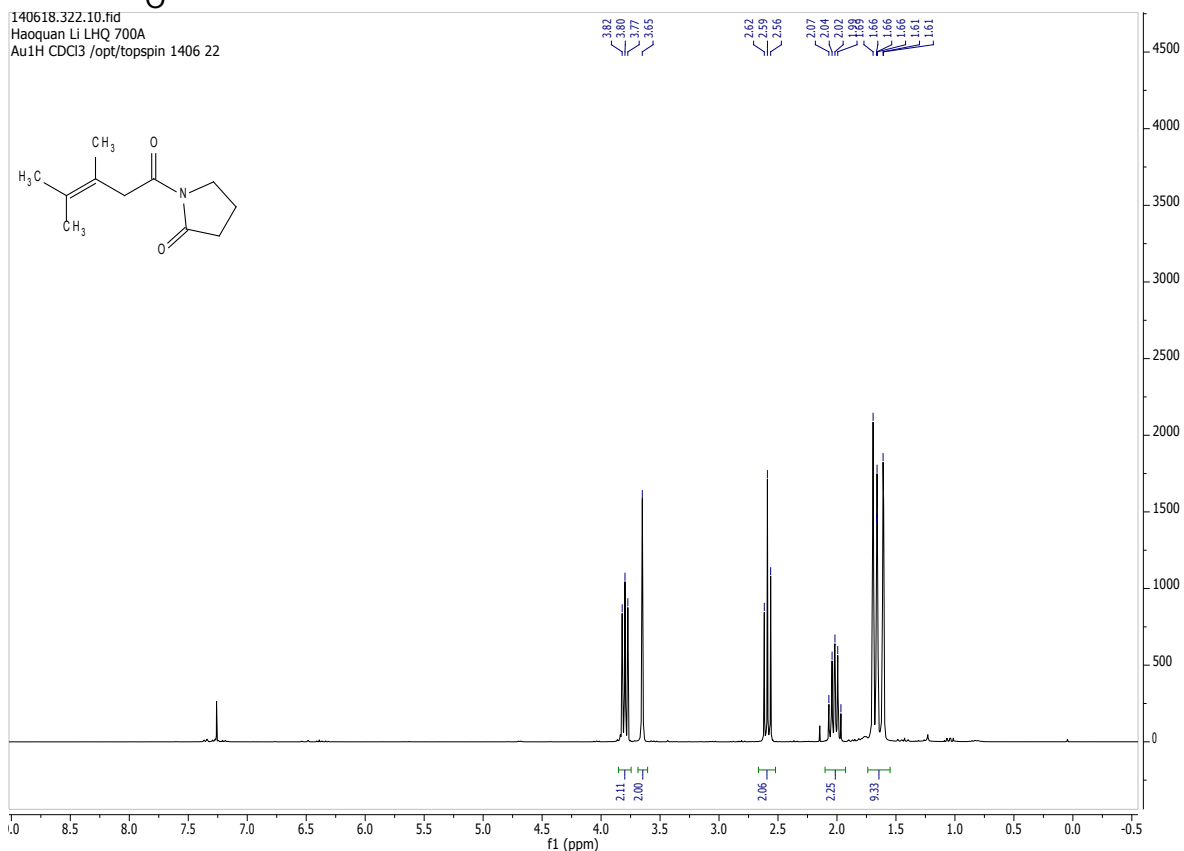
140613.320.11.fid
Haoquan Li LHQ 690A
Au13C CDCl3 /opt/topspin 1406 20



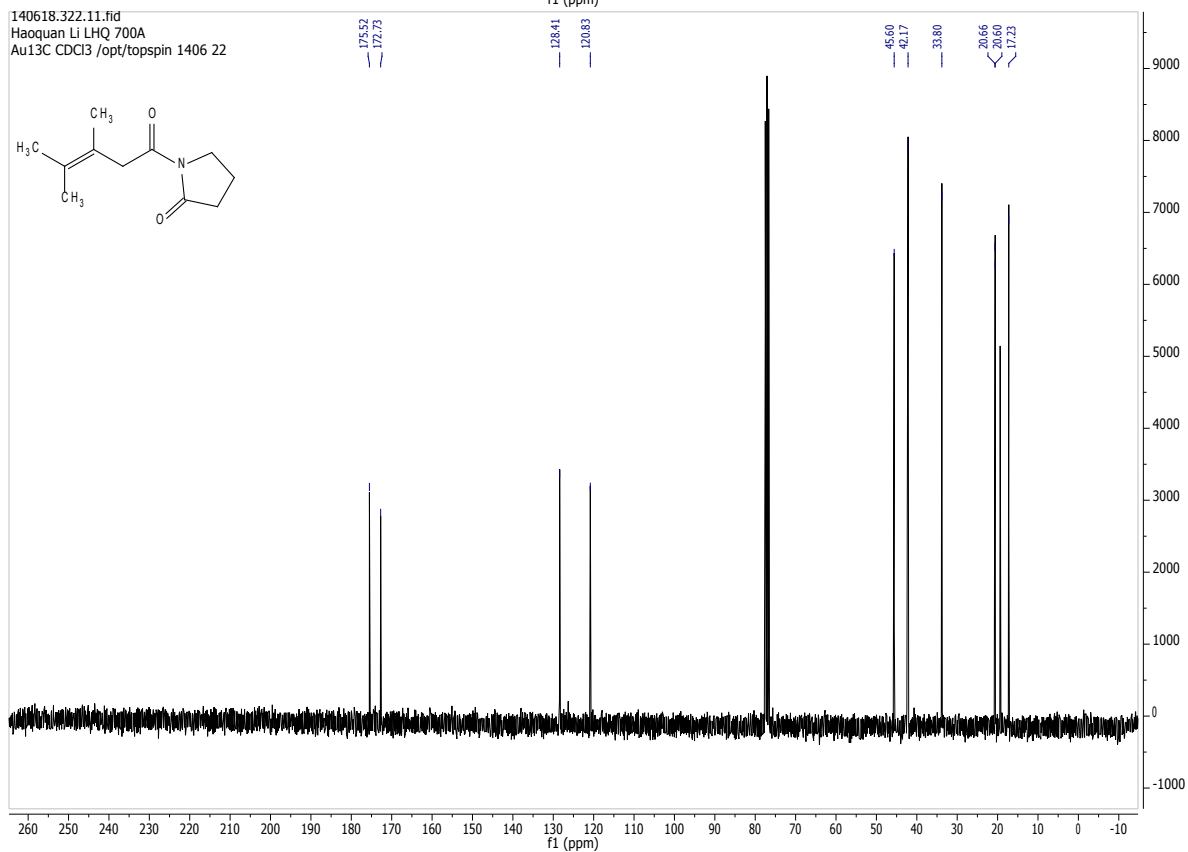


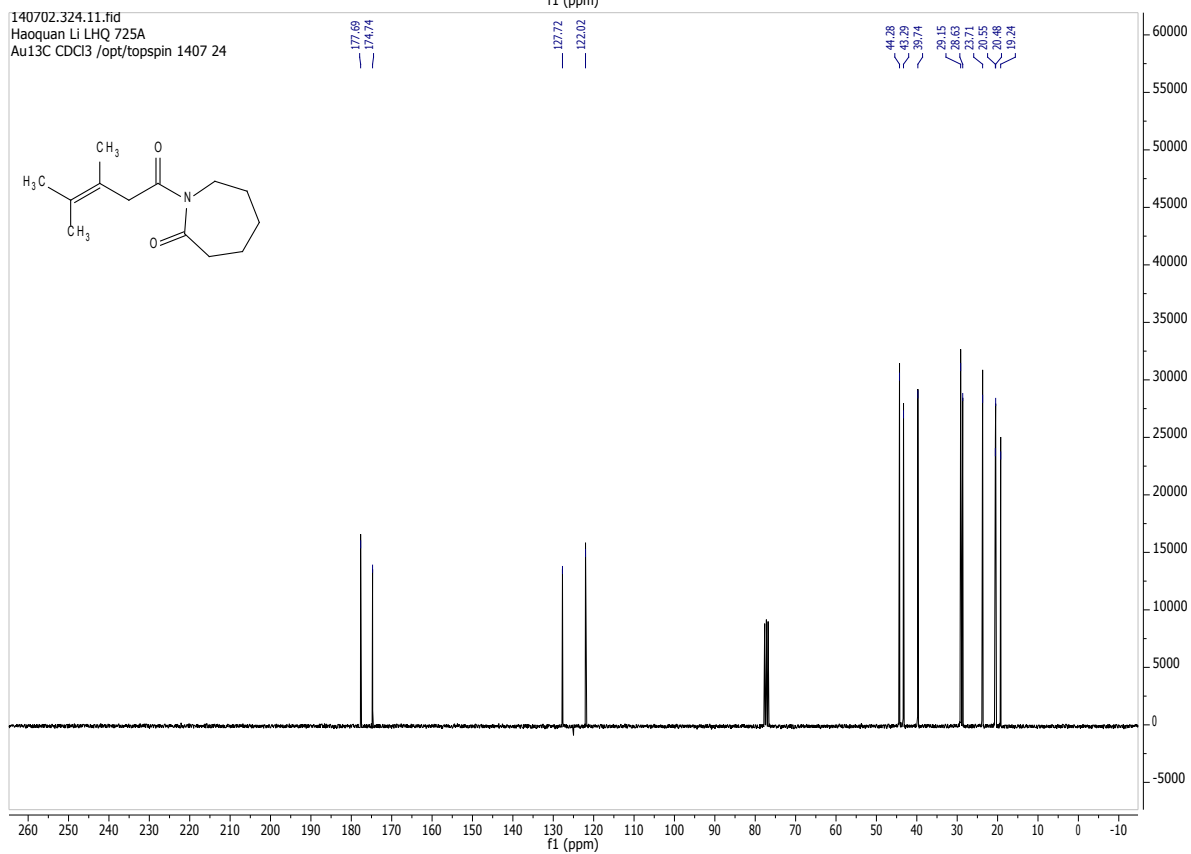
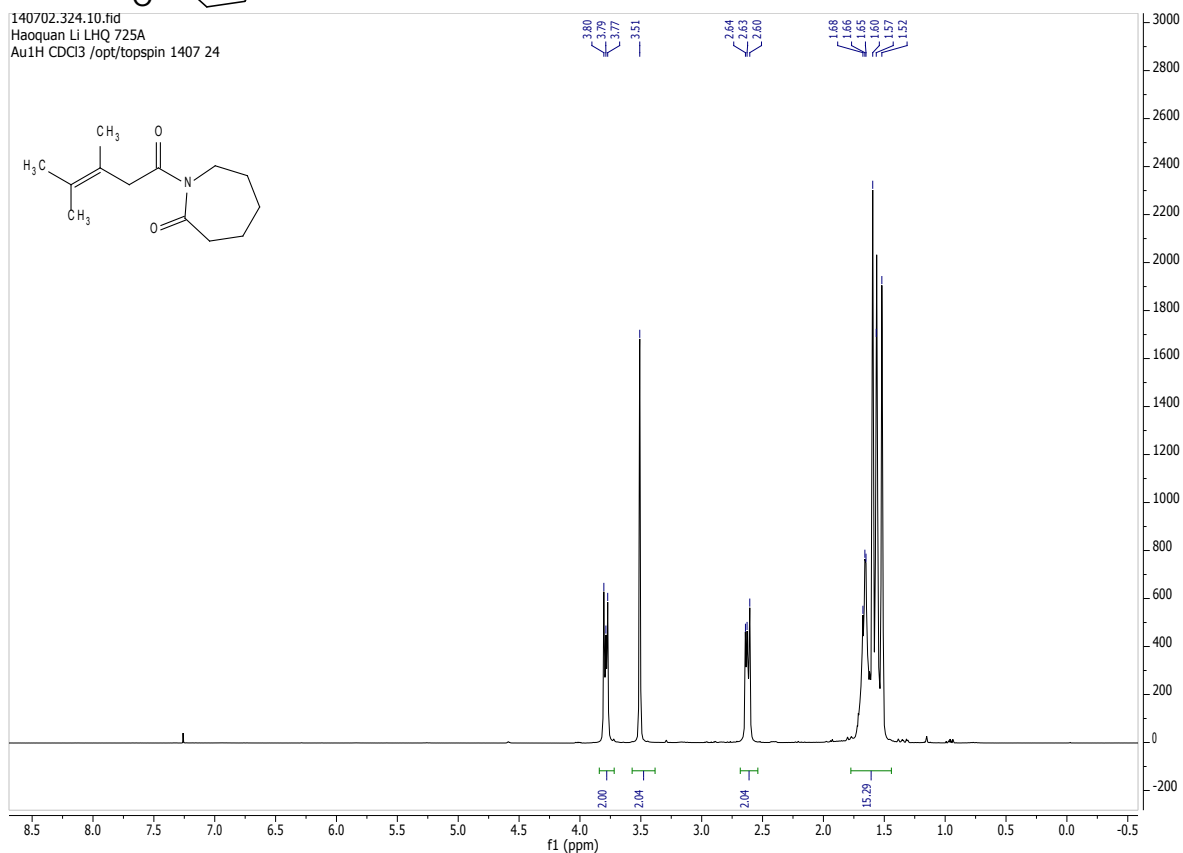
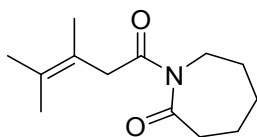


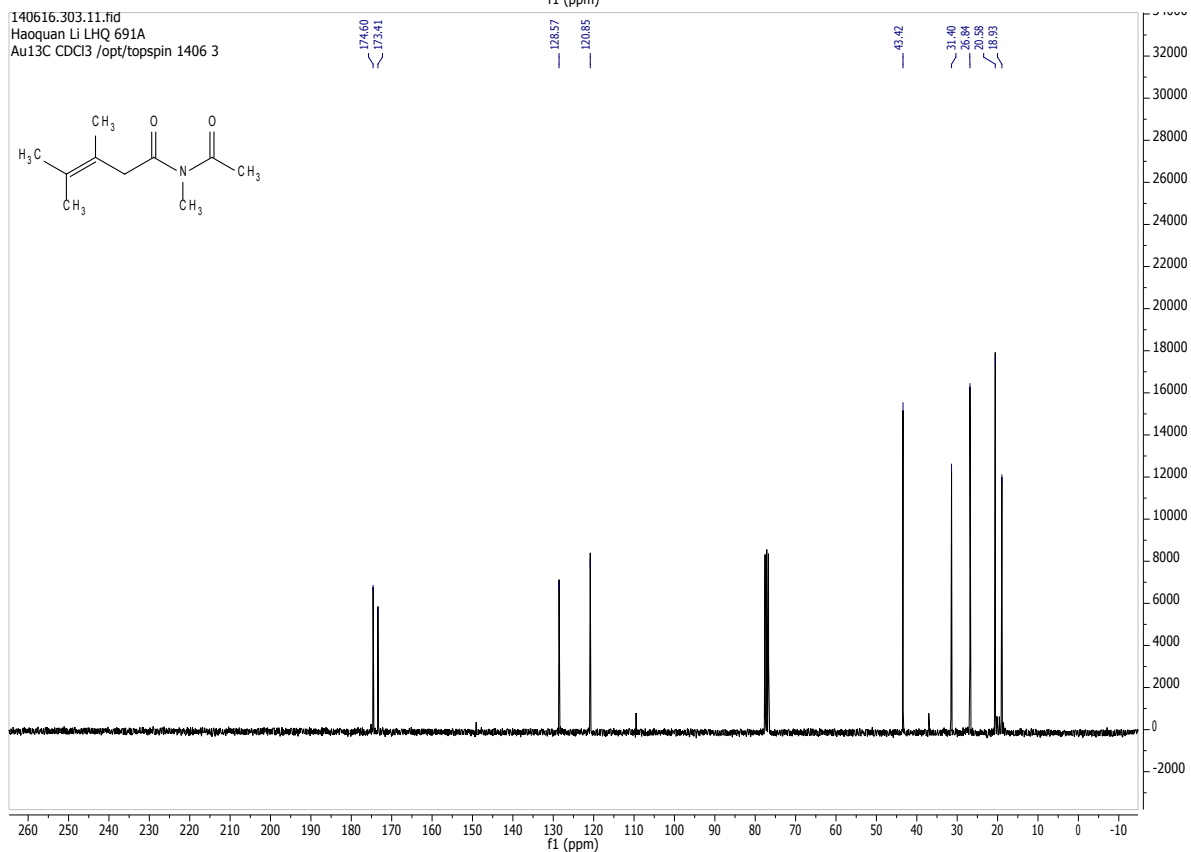
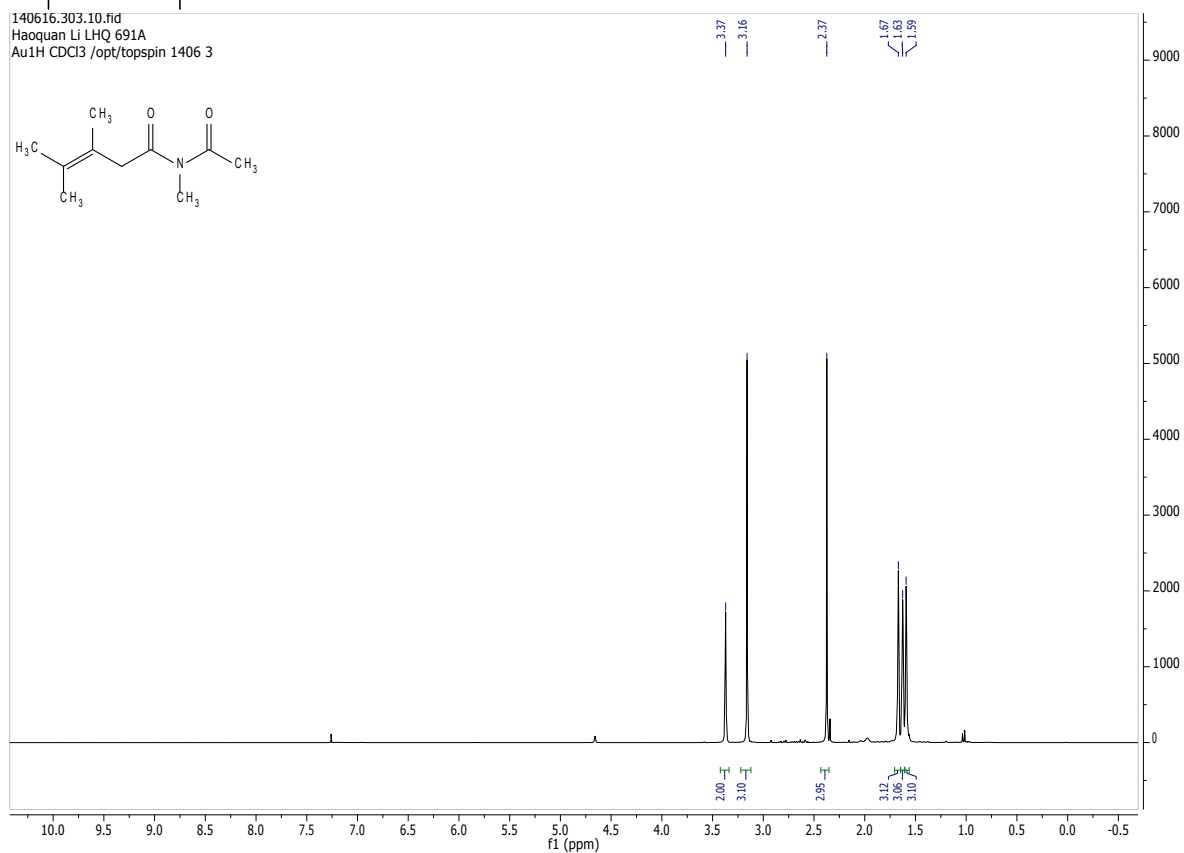
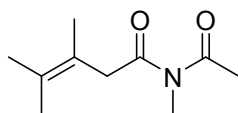
140618.322.10.fid
Haoquan Li LHQ 700A
Au1H CDCl3 /opt/topspin 1406 22

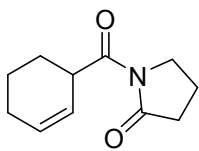


140618.322.11.fid
Haoquan Li LHQ 700A
Au13C CDCl3 /opt/topspin 1406 22

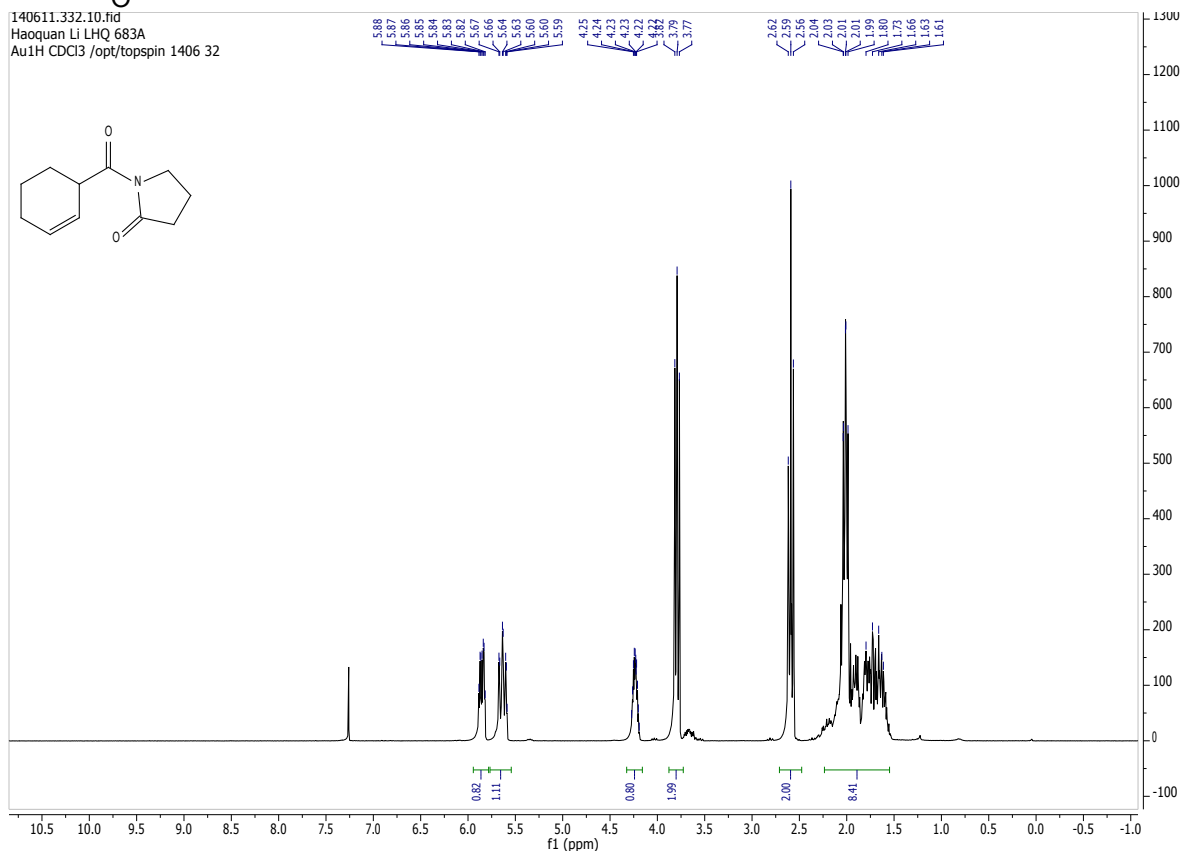




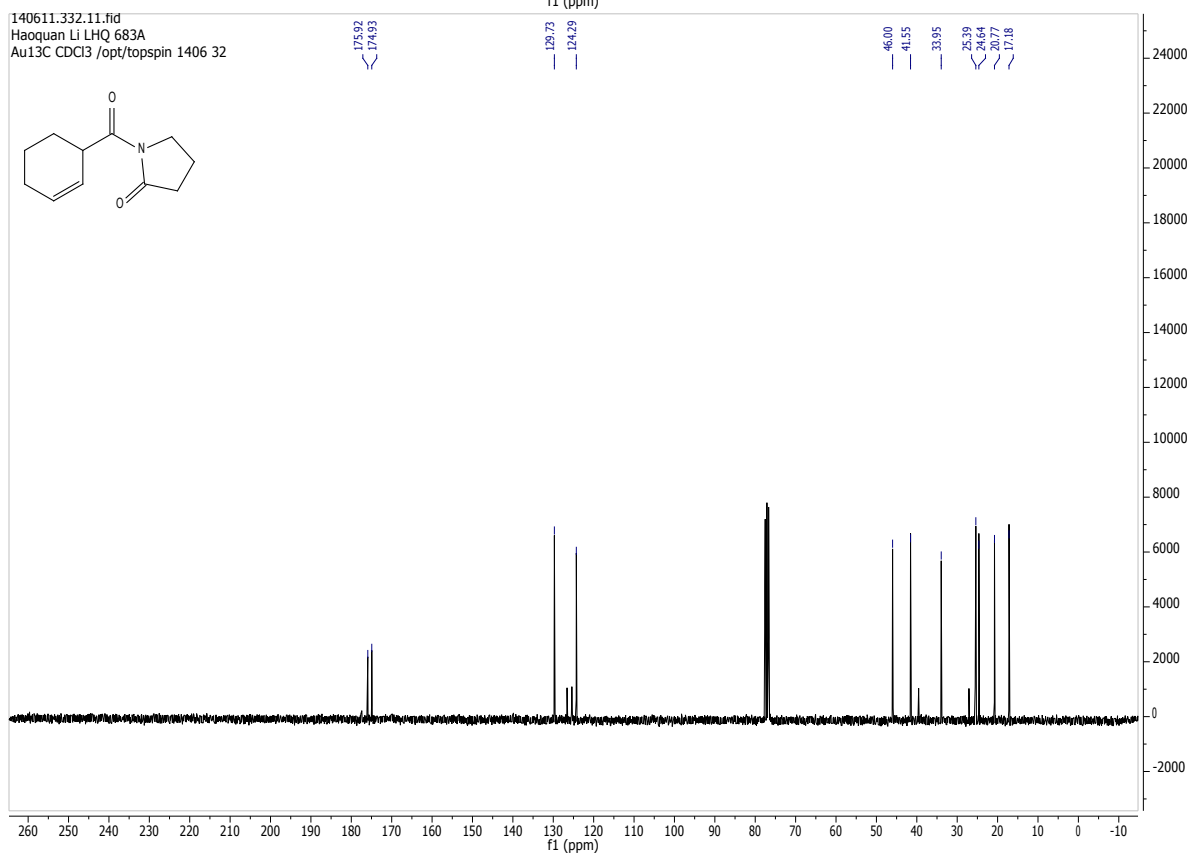


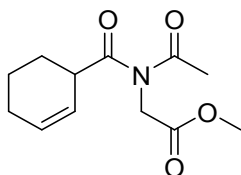


140611.332.10.fid
Haoquan Li LHQ 683A
Au1H CDCl₃ /opt/topspin 1406 32

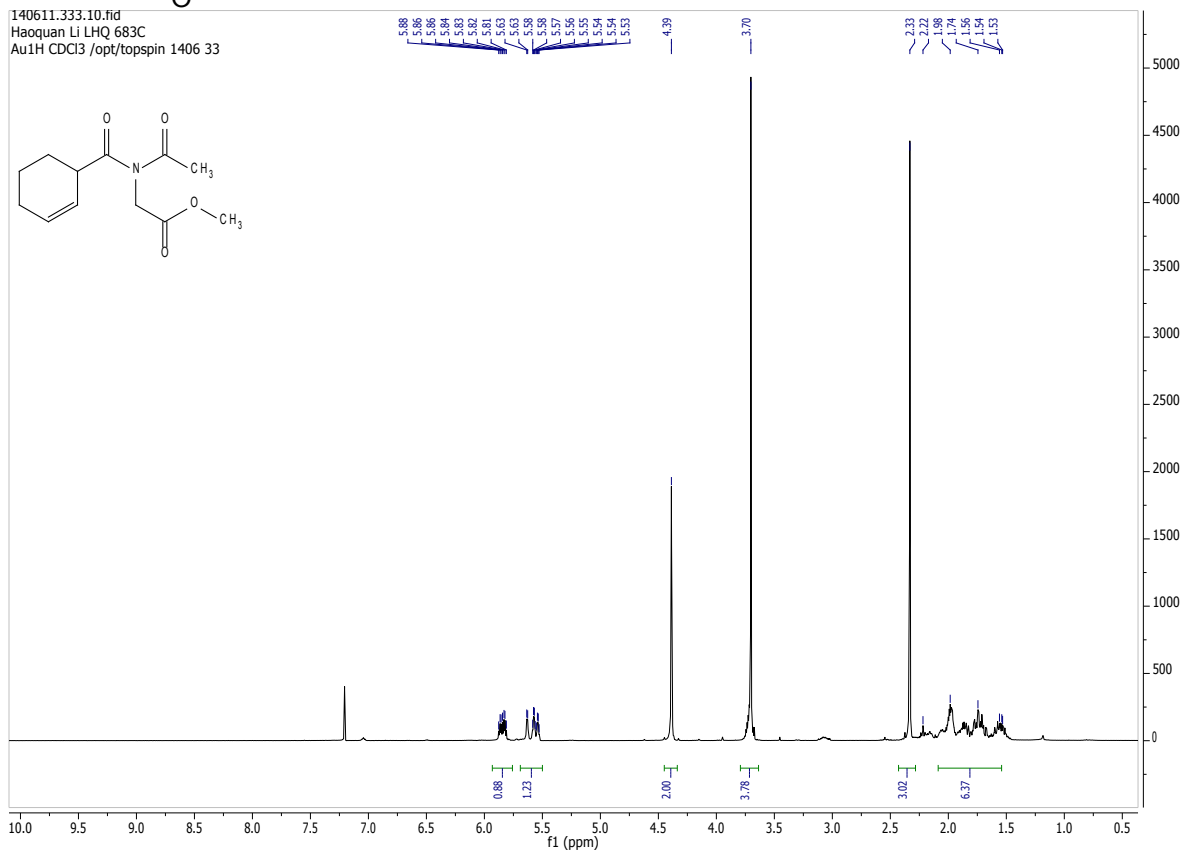


140611.332.11.fid
Haoquan Li LHQ 683A
Au13C CDCl₃ /opt/topspin 1406 32





140611.333.10.fid
Haoquan Li LHQ 683C
Au1H CDCl₃ /opt/topspin 1406 33



140611.333.11.fid
Haoquan Li LHQ 683C
Au13C CDCl₃ /opt/topspin 1406 33

