

Supporting Information

Facile construction of pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-ones via a redox-amination-aromatization-Friedel-Crafts acylation cascade reaction and discovery of novel topoisomerase inhibitors

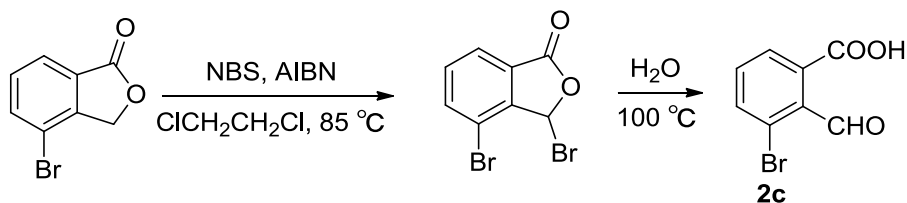
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and Wei Wang^{†,≠,*}

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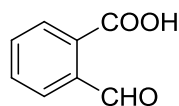
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General Information: All starting materials are commercially available and analytical pure. Microwave reactions were carried out in a Biotage (initiator-type) reactor. ^1H NMR and ^{13}C NMR spectra were recorded on Bruker AVANCE II 300 or AVANCE II 600 Nuclear Magnetic Resonance spectrometer (Bruker Company, Germany), using TMS as an internal standard and CDCl_3 or $\text{DMSO}-d_6$ as solvents. Chemical shifts (δ values) and coupling constants (J values) are given in ppm (δ) and Hz, respectively. The mass spectra were recorded on an Esquire 3000 LC-MS mass spectrometer. High resolution mass spectrometry (HRMS) was recorded on an Agilent 6538 UHD Accurate-Mass Q-TOF LC/MS spectrometer. Silica gel thin-layer chromatography was performed on precoated plates GF-254 (Qingdao Haiyang Chemical, China) and compounds were visualized with a UV light at 254 and 365 nm. Silica gel column chromatography was performed with Silica gel 60G (Qindao Haiyang Chemical, China). Structural determination of the products was determined by X-ray and NMR.

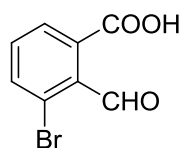
General procedure for the preparation of substrates 2c-e.



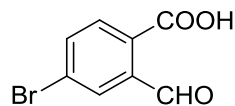
A mixture of 1,2-dichloroethane (25 mL), 4-bromoisobenzofuran-1(3H)-one (0.95 g, 4.5 mmol, 1.0 equiv), *N*-bromosuccinimide (0.89 g, 5.0 mmol, 1.1 equiv) and azobisisobutyronitrile (0.04 g, 0.2 mmol, 0.04 equiv) was refluxed for 1 h. The reaction mixture was kept in an ice bath for 2 h then filtered. The solvent was removed under reduced pressure. Water (10 mL) was added and the resulting mixture was refluxed for 1 h. The reaction mixture was cooled to room temperature and extracted with EtOAc. The combined organic phases were dried and concentrated under reduced pressure to give compound **2c** as a white solid (0.78 g, 76.2% yield). The synthetic method for substrates **2a** and **2d-e** were similar to the synthesis of substrate **2c**. Substrates **2b** and **2f** are commercially available.



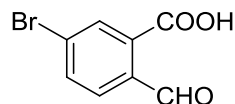
2-Formylbenzoic acid (2a): White solid (0.55g), yield: 82.1%. ^1H NMR (300 MHz, DMSO- d_6 , TMS): δ = 8.18 (s, 1H), 7.77-7.85 (m, 2H), 7.64-7.69 (m, 2H), 6.67 (s, 1H); ^{13}C NMR (75 MHz, DMSO- d_6 , TMS): δ = 168.92, 147.90, 134.97, 130.98, 127.04, 124.98, 124.19, 98.76; HRMS (ESI) calcd for $\text{C}_8\text{H}_5\text{O}_3$ $[\text{M}-\text{H}]^-$ = 149.0244, found 149.0243.



3-Bromo-2-formylbenzoic acid (2c): White solid (0.78 g), yield: 76.2%. ^1H NMR (300 MHz, DMSO- d_6 , TMS): δ = 8.28 (d, J = 8.6 Hz, 1H), 7.99 (d, J = 7.8 Hz, 1H), 7.85 (d, J = 7.4 Hz, 1H), 6.61 (d, J = 8.6 Hz, 1H); ^{13}C NMR (75 MHz, DMSO- d_6 , TMS): δ = 168.38, 147.05, 138.94, 133.75, 129.95, 125.03, 118.41, 99.23; HRMS (ESI) calcd for $\text{C}_8\text{H}_4\text{BrO}_3$ $[\text{M}-\text{H}]^-$ = 226.9349, found 226.9350.



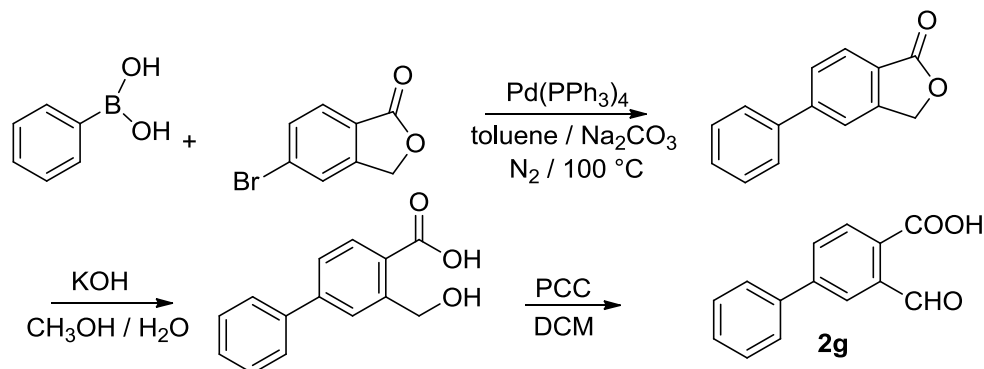
4-Bromo-2-formylbenzoic acid (2d): White solid (0.68 g), yield: 56.3%. ^1H NMR (300 MHz, DMSO- d_6 , TMS): δ = 7.90-7.91 (m, 1H), 7.84 (dd, J = 1.5, 8.1 Hz, 1H), 7.50 (d, J = 8.1 Hz, 1H), 7.63 (d, J = 8.0 Hz, 1H), 6.63 (s, 1H); ^{13}C NMR (75 MHz, DMSO- d_6 , TMS): δ = 168.38, 147.05, 138.94, 133.75, 129.95, 125.03, 118.41, 99.23; HRMS (ESI) calcd for $\text{C}_8\text{H}_4\text{BrO}_3$ $[\text{M}-\text{H}]^-$ = 226.9349, found 226.9348.



5-Bromo-2-formylbenzoic acid (2e): White solid (0.65 g), yield: 63.4%. ^1H NMR (600 MHz, DMSO- d_6 , TMS): δ = 8.27 (dd, J = 4.1 Hz, 1H), 7.96-8.00 (m, 2H), 7.63 (d, J = 7.7 Hz, 2H), 6.65 (d, J = 4.6 Hz, 1H); ^{13}C NMR (75 MHz, DMSO- d_6 , TMS): δ = 167.57, 149.41, 133.74, 128.45, 126.97, 126.42, 125.81, 97.66; HRMS (ESI) calcd

for $\text{C}_8\text{H}_4\text{BrO}_3$ $[\text{M}-\text{H}]^- = 226.9349$, found 226.9349.

General procedure for the preparation of substrates **2g-i**.



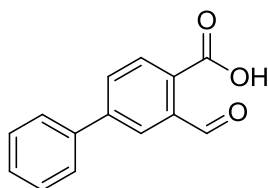
Phenylboronic acid (1.9 g, 15 mmol, 1.5 equiv), 5-bromoisobenzofuran-1(3H)-one (2.3 g, 10 mmol, 1.0 equiv) and tetrakis(triphenylphosphine)palladium(0) ($\text{Pd}(\text{PPh}_3)_4$) (1.0 g, 0.8 mmol, 0.08 equiv) were added to a mixed solvent of toluene (20 mL) and 2 M Na_2CO_3 solution (16 mL). The resulted solution was stirred at $100\text{ }^\circ\text{C}$ for 3 h under nitrogen atmosphere. Then the reaction mixture was extracted with EtOAc (3×30 mL). The combined EtOAc layer was washed with brine, dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The crude product was purified by column chromatography (silica gel, petroleum ether/EtOAc = 8:1, v/v) afforded 5-phenylisobenzofuran-1(3H)-one (2.0 g, 63.5% yield) as white solid.

5-Phenylisobenzofuran-1(3H)-one (1.5 g, 7.1 mmol, 1.0 equiv) and KOH (0.6 g, 11 mmol, 1.5 equiv) were added to the mixed solvent of methanol (17 mL) and water (3 mL) (methanol/water = 85:15, v/v). The resulted solution was stirred at $50\text{ }^\circ\text{C}$ for 2 h, and the pH value was adjusted to 4 by saturated KHSO_4 aqueous solution. After filtration, product **2g** was dried at $50\text{ }^\circ\text{C}$ as a white solid (1.62 g, 80.2% yield).

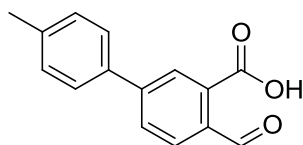
3-(Hydroxymethyl)-[1,1'-biphenyl]-4-carboxylic acid (0.60 g, 2.6 mmol, 1.0 equiv) was dissolved into dichloromethane (8 mL), pyridinium chlorochromate (PCC) (1.1 g, 5.2 mmol, 2.0 equiv) was added, and the solution was stirred for 2 h at room temperature. Then, the solvent was evaporated under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/EtOAc = 4: 1-1:1, v/v) to give the substrate **2g** (0.13 g, yield: 21.9%) as a white solid.

The synthetic method for substrates **2h** and **2i** was similar to the synthesis of

substrate **2g**. Substrate **2h** was obtained as crude compound by a simple purification procedure through flash column chromatography on silica gel.

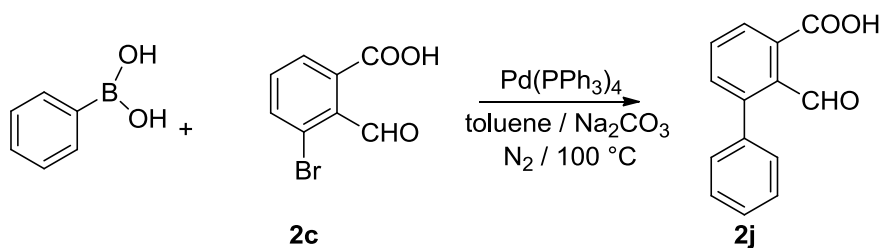


3-Formyl-[1,1'-biphenyl]-4-carboxylic acid (2g): White solid (0.13 g), yield: 21.9%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 7.97 (d, J = 7.6 Hz, 1H), 7.86 (d, J = 6.9 Hz, 2H), 7.66 (d, J = 7.6 Hz, 2H), 7.53 (t, J = 7.1 Hz, 2H), 7.47-7.49 (m, 1H), 6.71 (s, 1H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 168.45, 148.17, 147.07, 139.37, 130.20, 129.17, 128.82, 127.52, 125.88, 125.55, 121.92, 97.23; HRMS (ESI) calcd for $\text{C}_{14}\text{H}_9\text{O}_3$ $[\text{M}-\text{H}]^-$ = 225.0553, found 225.0552.

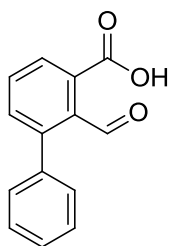


4-Formyl-4'-methyl-[1,1'-biphenyl]-3-carboxylic acid (2i): White solid (0.82 g), yield: 13.2 %. ^1H NMR (300 MHz, CDCl_3 , TMS): δ = 8.09 (d, J = 1.0 Hz, 1H), 7.96 (dd, J = 1.6, 7.9 Hz, 2H), 7.71 (d, J = 7.8 Hz, 1H), 7.54 (d, J = 8.1 Hz, 2H), 7.32-7.35 (m, 2H), 6.70 (s, 1H), 2.45 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3 , TMS): δ = 168.64, 144.60, 144.55, 138.48, 136.26, 133.47, 129.87, 127.68, 127.14, 123.66, 123.51, 97.36, 29.70; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{11}\text{O}_3$ $[\text{M}-\text{H}]^-$ = 239.0714, found 239.0715.

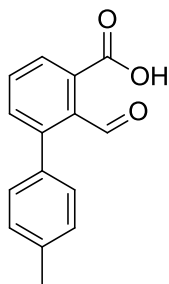
General procedure for the preparation of substrates 2j-t



Phenylboronic acid (0.26 g, 2.1 mmol, 1.2 equiv), 3-bromo-2-formylbenzoic acid **2c** (0.4g, 1.7 mmol, 1.0 equiv) and $\text{Pd(PPh}_3)_4$ (0.18g, 0.14 mmol, 0.08 equiv) were added to the mixed solvent of toluene (4.8 mL) and 2 M Na_2CO_3 solution (6.0 mL). The resulted solution was stirred at 100 °C for 3 h under nitrogen atmosphere. Then the reaction mixture was extracted with EtOAc (3×30 mL). The combined EtOAc layer was washed with brine, dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The crude product was purified by column chromatography (silica gel, petroleum ether/EtOAc = 8:1, v/v) afforded 2-formyl-[1,1'-biphenyl]-3-carboxylic acid **2j** (0.48 g, 47.1% yield) as a white solid. The synthetic method for substrates **2k-t** were similar to the synthesis of substrate **2j**.

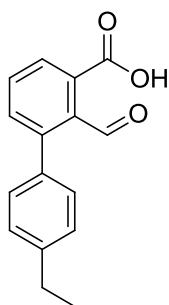


2-Formyl-[1,1'-biphenyl]-3-carboxylic acid (2j): White solid (0.18 g), yield: 47.1%. ^1H NMR (300 MHz, $\text{DMSO-}d_6$, TMS): δ = 7.91 (d, J = 8.2 Hz, 1H), 7.82 (dd, J = 0.9, 7.4 Hz, 1H), 7.80 (1.0, 7.6 Hz, 1H), 7.73 (t, J = 7.5 Hz, 1H), 7.63-7.65 (m, 2H), 7.46-7.49 (m, 2H), 7.40-7.42 (m, 1H), 7.04 (d, J = 8.1 Hz, 1H); ^{13}C NMR (75 MHz, $\text{DMSO-}d_6$, TMS): δ = 168.48, 144.41, 138.04, 137.36, 134.90, 131.16, 128.61, 128.48, 127.98, 127.33, 123.54, 98.26; HRMS (ESI) calcd for $\text{C}_{14}\text{H}_9\text{O}_3$ $[\text{M-H}]^-$ = 225.0557, found 225.0559.

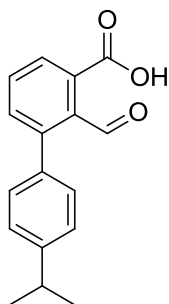


2-Formyl-4'-methyl-[1,1'-biphenyl]-3-carboxylic acid (2k): White solid (0.24 g), yield: 59.5%. ^1H NMR (300 MHz, $\text{DMSO-}d_6$, TMS): δ = 7.92 (s, 1H), 7.71-7.83 (m,

3H), 7.56 (d, $J = 7.9$ Hz, 2H), 7.31 (d, $J = 7.9$ Hz, 2H), 7.06 (s, 1H), 2.38 (s, 3H); ^{13}C NMR (75 MHz, $\text{DMSO}-d_6$, TMS): $\delta = 169.85, 145.59, 139.32, 138.70, 136.03, 135.76, 132.43, 130.52, 129.67, 128.61, 124.59, 99.65, 22.02$; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{11}\text{O}_3$ $[\text{M}-\text{H}]^- = 239.0714$, found 239.0715.

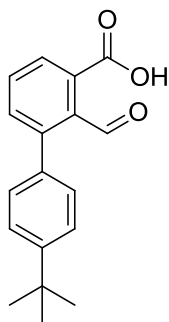


4'-Ethyl-2-formyl-[1,1'-biphenyl]-3-carboxylic acid (2l): White solid (0.11 g), yield: 26.1%. ^1H NMR (300 MHz, CDCl_3 , TMS): $\delta = 7.88$ (dd, $J = 1.3, 6.9$ Hz, 1H), 7.65-7.74 (m, 2H), 7.49 (d, $J = 8.1$ Hz, 2H), 7.33 (d, $J = 8.2$ Hz, 2H), 6.85 (d, $J = 7.4$ Hz, 1H), 2.74 (q, $J = 7.6$ Hz, 2H), 1.31 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3 , TMS): $\delta = 168.70, 144.65, 143.20, 139.04, 135.28, 134.54, 131.43, 128.52, 128.37, 127.39, 124.18, 97.14, 28.57, 12.38$; HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{13}\text{O}_3$ $[\text{M}-\text{H}]^- = 253.0870$, found 253.0870.

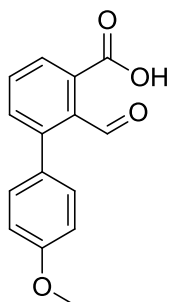


2-Formyl-4'-isopropyl-[1,1'-biphenyl]-3-carboxylic acid (2m): Yellow solid (0.31 g), yield: 60.3%. ^1H NMR (600 MHz, CDCl_3 , TMS): $\delta = 7.88$ (d, $J = 7.4$ Hz, 1H), 7.74 (dd, $J = 0.8, 7.6$ Hz, 1H), 7.69 (t, $J = 7.6$ Hz, 1H), 7.53 (d, $J = 8.1$ Hz, 2H), 7.38 (d, $J = 8.1$ Hz, 2H), 6.87 (s, 1H), 2.99-3.04 (m, 1H), 1.34 (d, $J = 6.9$ Hz, 6H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): $\delta = 168.86, 149.20, 143.25, 139.04, 135.29, 134.65, 131.37, 128.54, 127.37, 126.93, 124.13, 97.28, 33.85, 23.89$; HRMS (ESI) calcd for

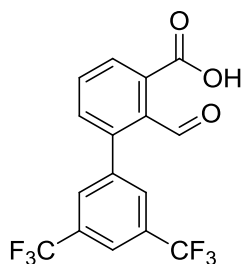
$C_{17}H_{15}O_3$ $[M-H]^- = 267.1027$, found 267.1029.



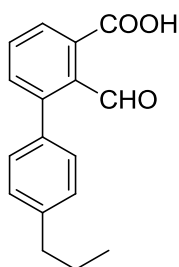
4'-(*Tert*-butyl)-2-formyl-[1,1'-biphenyl]-3-carboxylic acid (2n): Yellow oil (0.33 g), yield: 62%. 1H NMR (600 MHz, $CDCl_3$, TMS): δ = 7.87 (d, J = 7.4 Hz, 1H), 7.74 (d, J = 7.4 Hz, 2H), 7.68 (t, J = 7.8 Hz, 1H), 7.53 (s, 4H), 7.50 (d, J = 8.2 Hz, 2H), 1.41 (s, 9H); ^{13}C NMR (75 MHz, $DMSO-d_6$, TMS): δ = 170.80, 169.03, 150.87, 144.77, 138.42, 135.34, 134.97, 131.62, 128.69, 127.80, 125.92, 123.80, 98.77, 60.22, 34.81; HRMS (ESI) calcd for $C_{18}H_{17}O_3$ $[M-H]^- = 281.1183$, found 281.1182.



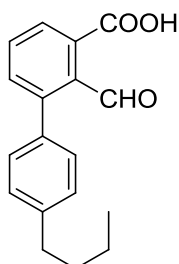
2-Formyl-4'-methoxy-[1,1'-biphenyl]-3-carboxylic acid (2o): White solid (0.11 g), yield: 25.3%. 1H NMR (300 MHz, $CDCl_3$, TMS): δ = 7.80 (dd, J = 1.3, 7.3 Hz, 1H), 7.59-7.69 (m, 2H), 7.47-7.52 (m, 2H), 7.09-7.18 (m, 1H), 6.96-7.01 (m, 2H), 6.80 (s, 1H), 3.84 (s, 3H); HRMS (ESI) calcd for $C_{15}H_{12}NaO_4$ $[M+Na]^+ = 279.0628$, found 279.0629.



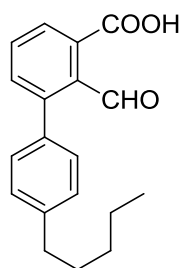
2-Formyl-3',5'-bis(trifluoromethyl)-[1,1'-biphenyl]-3-carboxylic acid (2p): White solid (0.18 g), yield: 28.7%. ^1H NMR (300 MHz, CD_3OD , TMS): δ = 8.38 (s, 2H), 8.21 (s, 1H), 8.14 (s, 1H), 7.95-8.02 (m, 2H), 7.83 (t, J = 7.6 Hz, 1H), 7.09 (s, 1H); ^{13}C NMR (75 MHz, CD_3OD , TMS): δ = 169.43, 146.19, 141.16, 136.80, 136.08, 132.75, 132.10, 131.67, 130.82, 128.73, 126.35, 99.47; HRMS (ESI) calcd for $\text{C}_{16}\text{H}_7\text{F}_6\text{O}_3$ $[\text{M}-\text{H}]^-$ = 361.0305, found 361.0307.



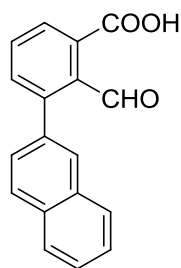
2-Formyl-4'-propyl-[1,1'-biphenyl]-3-carboxylic acid (2q): White solid (0.10 g), yield: 22.4%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 7.85 (dd, J = 1.0, 7.5 Hz, 1H), 7.69 (dd, J = 1.0, 7.6 Hz, 1H), 7.65 (t, J = 7.5 Hz, 1H), 7.45 (dt, J = 2.0, 4.0, 8.3 Hz, 2H), 7.28 (d, J = 8.3 Hz, 2H), 6.82 (d, J = 3.6 Hz, 1H), 2.64 (t, J = 7.6 Hz, 2H), 1.65-1.72 (m, 2H), 0.98 (t, J = 7.3 Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 168.68, 143.22, 139.08, 135.27, 134.56, 131.42, 128.96, 128.45, 127.41, 124.18, 97.14, 37.76, 29.71, 24.41, 13.90; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{15}\text{O}_3$ $[\text{M}-\text{H}]^-$ = 267.1027, found 267.1028.



4'-Butyl-2-formyl-[1,1'-biphenyl]-3-carboxylic acid (2r): White solid (0.21 g), yield: 43.5%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 7.87 (s, 1H), 7.77 (t, J = 6.9 Hz, 2H), 7.69 (t, J = 7.6 Hz, 1H), 7.54 (d, J = 8.0 Hz, 2H), 7.27 (d, J = 8.0 Hz, 2H), 7.01 (s, 1H), 2.61 (t, J = 7.6 Hz, 2H), 1.54-1.60 (m, 2H), 1.29-1.35 (m, 2H), 0.89 (t, J = 7.4 Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 169.03, 144.78, 142.74, 138.53, 135.27, 135.19, 131.62, 129.04, 128.88, 127.83, 123.78, 98.81, 34.97, 33.48, 22.31; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{17}\text{O}_3$ $[\text{M}-\text{H}]^-$ = 281.1183, found 281.1184.



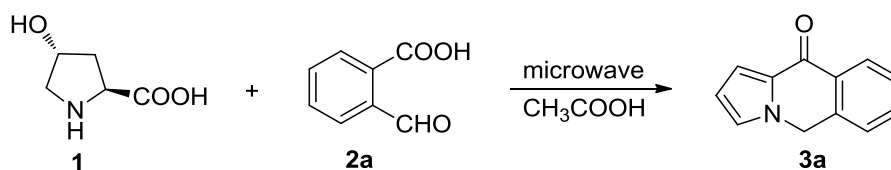
2-Formyl-4'-pentyl-[1,1'-biphenyl]-3-carboxylic acid (2s): White solid (0.26 g), yield: 52.5%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 7.86 (dd, J = 1.0, 7.4 Hz, 1H), 7.71 (dd, J = 1.0, 7.6 Hz, 1H), 7.66 (t, J = 7.5 Hz, 1H), 7.47 (dt, J = 2.0, 4.0, 8.3 Hz, 2H), 7.29 (dt, J = 2.0, 4.0, 8.3 Hz, 2H), 6.83 (d, J = 7.2 Hz, 1H), 3.76 (d, J = 7.7 Hz, 1H), 2.67 (t, J = 7.7 Hz, 2H), 1.65-1.69 (m, 2H), 1.35-1.39 (m, 4H), 0.91-0.94 (m, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 168.23, 142.93, 142.74, 138.58, 134.77, 134.02, 130.91, 128.39, 127.97, 126.91, 123.66, 96.69, 35.16, 31.08, 30.53, 22.05, 13.53; HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{19}\text{O}_3$ $[\text{M}-\text{H}]^-$ = 295.1340, found 295.1342.



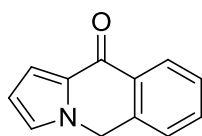
2-Formyl-3-(naphthalen-2-yl)benzoic acid (2t): Gray solid (0.10 g), yield: 22.4%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.03 (d, J = 1.5 Hz, 1H), 7.96 (d, J = 8.5 Hz, 1H), 7.89-7.93 (m, 3H), 7.82 (dd, J = 0.9, 7.5 Hz, 1H), 7.72 (t, J = 7.5 Hz, 1H), 7.65

(dd, $J = 1.8, 8.4$ Hz, 1H), 7.54-7.57 (m, 2H), 6.89 (d, $J = 6.3$ Hz, 1H), 3.69 (s, 1H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): $\delta = 168.05, 143.02, 138.56, 135.04, 134.19, 132.81, 132.43, 131.02, 128.09, 127.78, 127.48, 127.29, 127.06, 126.26, 125.71, 124.01, 96.57$; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{11}\text{O}_3$ $[\text{M}-\text{H}]^- = 275.0714$, found 275.0716.

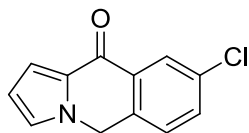
General procedure for the preparation of target compounds **3a-f**



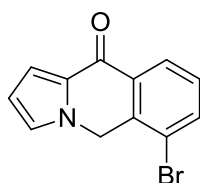
A 10 mL microwave reaction tube was charged with *trans*-4-hydroxy-*L*-proline **1** (0.16 g, 1.2 mmol, 1.2 equiv), 2-formylbenzoic acid **2a** (0.15 g, 1.0 mmol, 1.0 equiv), and CH_3COOH (2.0 mL). The reaction tube was sealed with a Teflon-lined snap cap and heated in the microwave reactor at 170 °C at very high absorption level for 20 min. After cooling, the crude reaction mixture was evaporated in vacuo and the crude product was purified by silica gel column chromatography (petroleum ether/EtOAc = 8: 1, v/v) to give **3a** (0.13 g, 71.7% yield) as a white solid. The synthetic method for compounds **3b-f** was similar to the synthesis of compound **3a**.



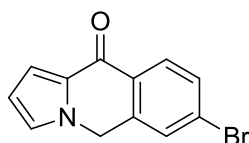
Pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3a**):** ^1H NMR (600 MHz, CDCl_3 , TMS): $\delta = 8.18$ (d, $J = 2.1$ Hz, 1H), 7.44 (dd, $J = 2.0, 8.1$ Hz, 1H), 7.22 (d, $J = 8.1$ Hz, 1H), 7.13-7.14 (m, 1H), 7.01-7.01 (m, 1H), 6.38 (dd, $J = 2.5, 4.2$ Hz, 1H), 5.28 (s, 2H); ^{13}C NMR (125 MHz, CDCl_3 , TMS): $\delta = 174.80, 135.60, 132.46, 130.69, 129.76, 127.99, 127.23, 125.68, 125.57, 113.77, 111.71, 46.98$; HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{10}\text{NO}$ $[\text{M}+\text{H}]^+ = 184.0758$, found 184.0759.



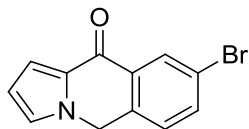
8-Chloropyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3b): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.27 (d, J = 2.2 Hz, 1H), 7.53 (dd, J = 2.3, 8.2 Hz, 1H), 7.30 (d, J = 8.2 Hz, 1H), 7.22 (dd, J = 1.3, 4.0 Hz, 1H), 7.10 (s, 1H), 6.46 (dd, J = 2.5, 4.0 Hz, 1H), 5.37 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 173.49, 134.55, 133.84, 132.61, 132.28, 129.40, 127.34, 127.21, 126.12, 114.46, 112.22, 46.77; HRMS (ESI) calcd for $\text{C}_{12}\text{H}_9\text{ClNO}$ $[\text{M}+\text{H}]^+ = 218.0367$, found 218.0369.



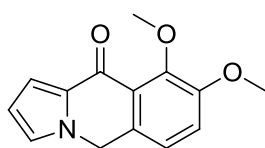
6-Bromopyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3c): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.35 (dd, J = 1.0, 7.6 Hz, 1H), 7.83 (dd, J = 1.0, 7.3 Hz, 1H), 7.43 (t, J = 8.0 Hz, 1H), 7.25 (dd, J = 1.4, 4.1 Hz, 1H), 7.20-7.21 (m, 1H), 6.52 (dd, J = 2.4, 4.1 Hz, 1H), 5.38 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 173.45, 136.30, 134.85, 132.87, 129.19, 128.58, 126.51, 125.17, 121.32, 114.12, 112.19, 48.27; HRMS (ESI) calcd for $\text{C}_{12}\text{H}_9\text{BrNO}$ $[\text{M}+\text{H}]^+ = 261.9862$, found 261.9863.



7-Bromopyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3d): ^1H -NMR (600 MHz, CDCl_3 , TMS): δ = 8.18 (d, J = 8.4 Hz, 1H), 7.63 (dd, J = 1.8, 8.4 Hz, 1H), 7.53 (s, 1H), 7.23 (dd, J = 1.4, 4.0 Hz, 1H), 7.10 (s, 1H), 6.47 (dd, J = 2.5, 4.0 Hz, 1H), 5.38 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 173.74, 139.23, 137.15, 131.44, 129.56, 128.90, 128.62, 127.39, 125.76, 114.18, 112.01, 46.45; HRMS (ESI) calcd for $\text{C}_{12}\text{H}_9\text{BrNO}$ $[\text{M}+\text{H}]^+ = 261.9862$, found 261.9863.

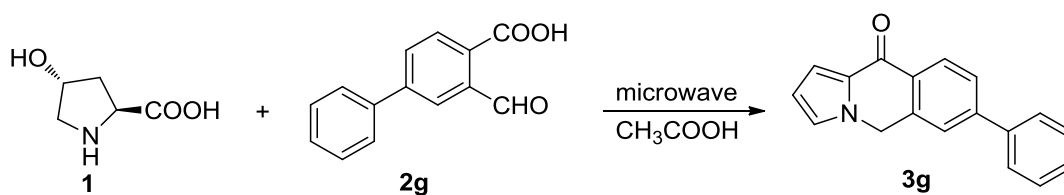


8-Bromopyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3e): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.43 (s, 1H), 7.68 (d, J = 8.1 Hz, 1H), 7.22-7.24 (m, 2H), 7.10 (s, 1H), 6.46 (s, 1H), 5.35 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 173.35, 135.44, 134.33, 132.45, 130.23, 129.36, 127.55, 126.12, 122.37, 114.46, 112.23, 46.80; HRMS (ESI) calcd for $\text{C}_{12}\text{H}_8\text{BrNNaO}$ $[\text{M}+\text{Na}]^+ = 261.9862$, found 261.9860.



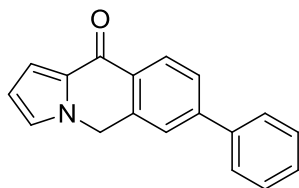
8,9-Dimethoxypyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3f): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 7.12-7.13 (m, 2H), 7.05 (dt, J = 0.9, 1.8, 8.4 Hz, 1H), 6.97-6.98 (m, 1H), 6.38 (dd, J = 2.5, 4.0 Hz, 1H), 5.28 (s, 1H), 3.96 (s, 3H), 3.90 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.77, 153.38, 150.75, 130.96, 129.07, 125.12, 124.38, 121.42, 116.60, 113.48, 111.50, 61.49, 56.44, 46.73; HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{14}\text{NO}_3$ $[\text{M}+\text{H}]^+ = 244.0968$, found 244.0969.

General procedure for the preparation of target compounds 3g-t

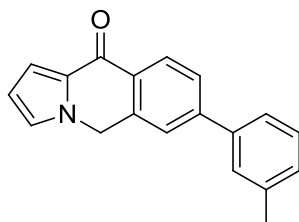


A 10 mL microwave reaction tube was charged with *trans*-4-hydroxy-*L*-proline **1** (0.13 g, 1.0 mmol, 1.0 equiv), 3-formyl-[1,1'-biphenyl]-4-carboxylic acid **2g** (0.27 g, 1.2 mmol, 1.2 equiv), and CH_3COOH (2.0 mL). The reaction tube was sealed with a Teflon-lined snap cap and heated in the microwave reactor at 170 °C at very high absorption level for 20 min. After cooling, the crude reaction mixture was evaporated in vacuo and the crude product was purified by silica gel column chromatography (petroleum ether/EtOAc = 16:1-14:1, v/v) to give **3g** (0.078 g, 30.0% yield) as a white

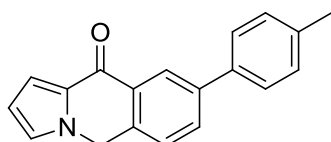
solid. The synthetic method for compounds **3h-t** were similar to the synthesis of compound **3g**.



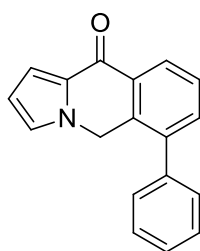
7-Phenylpyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3g): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.38 (d, J = 8.1 Hz, 1H), 7.71 (dt, J = 0.9, 1.7, 8.1 Hz, 1H), 7.63-7.65 (m, 2H), 7.55 (d, J = 0.9 Hz, 1H), 7.47-7.50 (m, 2H), 7.42 (tt, J = 1.2, 2.4, 6.9 Hz, 1H), 7.23 (dd, J = 1.5, 3.9 Hz, 3H), 7.10-7.11 (m, 1H), 6.46 (dd, J = 2.4, 4.0 Hz, 1H), 5.47 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.60, 145.33, 139.66, 136.14, 129.88, 129.56, 129.04, 128.43, 127.85, 127.30, 126.89, 125.63, 124.19, 113.17, 111.75, 47.16; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{14}\text{NO}$ $[\text{M}+\text{H}]^+ = 260.1070$, found 260.1071.



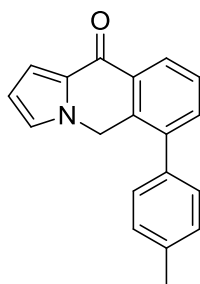
7-(*M*-tolyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3h): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.37 (d, J = 8.0 Hz, 1H), 7.69-7.71 (m, 1H), 7.54 (s, 1H), 7.42-7.45 (m, 2H), 7.37 (t, J = 7.6 Hz, 1H), 7.23-7.24 (m, 2H), 7.11 (s, 1H), 6.46 (dd, J = 2.4, 3.9 Hz, 1H), 5.47 (s, 2H), 2.44 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.16, 144.98, 138.22, 135.58, 129.38, 128.96, 128.68, 128.45, 127.55, 127.29, 126.41, 125.12, 123.91, 123.68, 123.02, 115.42, 113.25, 111.24, 46.66, 31.12; HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{16}\text{NO}$ $[\text{M}+\text{H}]^+ = 274.1226$, found 274.1228.



8-(*P*-tolyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3i): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.58 (s, 1H), 7.83 (dd, J = 1.3, 7.5 Hz, 1H), 7.62 (d, J = 8.0 Hz, 2H), 7.43 (d, J = 7.8 Hz, 1H), 7.32 (d, J = 7.8 Hz, 2H), 7.14 (s, 1H), 6.49-6.50 (m, 1H), 5.46 (s, 2H), 2.45 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.80, 140.87, 137.77, 136.72, 134.11, 130.93, 129.80, 129.65, 126.92, 126.21, 125.67, 125.12, 113.85, 111.75, 46.85, 21.13; HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{16}\text{NO}$ $[\text{M}+\text{H}]^+ = 274.1226$, found 274.1228.

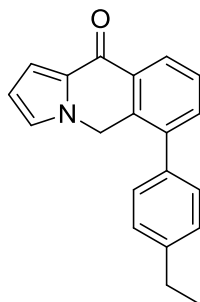


6-Phenylpyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3j): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.39 (dd, J = 1.0, 7.7 Hz, 1H), 7.56 (t, J = 7.6 Hz, 1H), 7.50-7.53 (m, 2H), 7.47-7.49 (m, 2H), 7.34-7.35 (m, 2H), 7.21 (dd, J = 1.1, 3.9 Hz, 1H), 6.96-6.97 (m, 1H), 6.42 (dd, J = 2.5, 4.0 Hz, 1H), 5.20 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 176.26, 141.45, 140.76, 135.20, 134.61, 132.44, 130.58, 130.29, 130.11, 129.46, 129.11, 127.94, 127.28, 115.03, 113.17, 47.70; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{14}\text{NO}$ $[\text{M}+\text{H}]^+ = 260.1070$, found 260.1068.

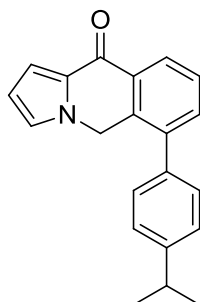


6-(*P*-tolyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3k): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.41 (dd, J = 1.2, 8.0 Hz, 1H), 7.59 (t, J = 7.4 Hz, 1H), 7.50 (dd, J = 1.5, 7.7 Hz, 1H), 7.37 (d, J = 7.7 Hz, 2H), 7.28 (d, J = 8.0 Hz, 2H), 7.25 (dd, J = 1.5, 3.9 Hz, 1H), 7.01-7.02 (m, 1H), 6.46 (dd, J = 2.4, 4.0 Hz, 1H), 5.26 (s, 2H), 2.52 (s, 3H);

^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.92, 140.06, 137.91, 136.40, 133.90, 133.32, 131.03, 129.54, 129.21, 128.58, 127.67, 126.38, 125.81, 113.57, 11.72, 46.32, 29.71, 21.27; HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{16}\text{NO}$ $[\text{M}+\text{H}]^+ = 274.1226$, found 274.1227.

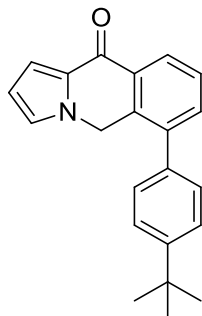


6-(4-Ethylphenyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3l): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.39 (dd, J = 0.9, 7.7 Hz, 1H), 7.57 (t, J = 7.5 Hz, 1H), 7.49 (dd, J = 1.2, 7.4 Hz, 1H), 7.38 (d, J = 7.9 Hz, 2H), 7.28-7.30 (m, 2H), 7.23 (dd, J = 1.3, 3.9 Hz, 1H), 7.00 (t, J = 1.8 Hz, 1H), 6.43 (dd, J = 2.4, 3.9 Hz, 1H), 5.25 (s, 2H), 2.80 (q, J = 7.6 Hz, 2H), 1.38 (t, J = 7.6 Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.92, 144.15, 140.07, 136.56, 133.92, 130.97, 129.16, 128.62, 128.31, 127.64, 126.32, 125.85, 113.55, 111.70, 46.32, 28.61, 15.45; HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{18}\text{NO}$ $[\text{M}+\text{H}]^+ = 288.1383$, found 288.1385.

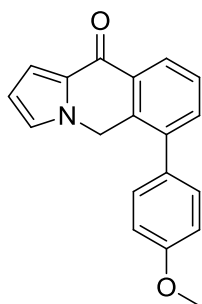


6-(4-Isopropylphenyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3m): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.40 (dd, J = 1.0, 7.8 Hz, 1H), 7.58 (t, J = 7.6 Hz, 1H), 7.50 (dd, J = 1.2, 7.4 Hz, 1H), 7.40 (d, J = 7.9 Hz, 2H), 7.30 (s, 1H), 7.29 (s, 1H), 7.24 (dd, J = 1.3, 3.9 Hz, 1H), 7.02 (t, J = 1.88 Hz, 1H), 6.44 (dd, J = 2.4, 3.9 Hz, 1H), 5.26 (s, 2H), 3.03-3.07 (m, 1H), 1.38 (d, J = 6.9 Hz, 6H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 175.58, 149.39, 140.73, 137.33, 14.59, 133.95, 131.68, 129.86, 129.25, 128.29,

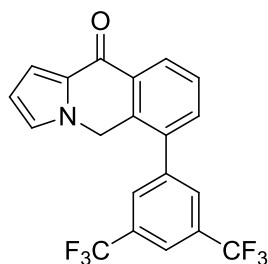
127.53, 126.99, 126.44, 114.19, 112.33, 46.98, 34.55, 24.62; HRMS (ESI) calcd for $C_{21}H_{20}NO$ $[M+H]^+ = 302.1539$, found 302.1540.



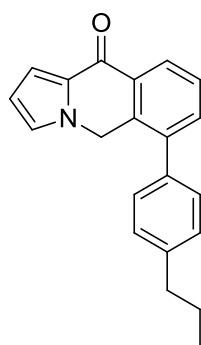
6-(4-(*Tert*-butyl)phenyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3n): 1H NMR (600 MHz, $CDCl_3$, TMS): δ = 8.40 (dd, J = 1.1, 7.8 Hz, 1H), 7.54-7.59 (m, 3H), 7.49 (dd, J = 1.2, 7.4 Hz, 1H), 7.31 (t, J = 2.2 Hz, 1H), 7.23 (dd, J = 1.3, 3.9 Hz, 1H), 7.02 (t, J = 1.8 Hz, 1H), 6.44 (dd, J = 2.5, 4.4 Hz, 1H), 5.27 (s, 2H), 1.45 (s, 9H); ^{13}C NMR (150 MHz, $CDCl_3$, TMS): δ = 174.96, 151.07, 140.04, 136.32, 133.98, 133.34, 131.05, 129.24, 128.38, 127.66, 126.36, 125.84, 125.76, 113.56, 111.70, 46.37, 34.73, 31.40; HRMS (ESI) calcd for $C_{21}H_{22}NO$ $[M+H]^+ = 316.1696$, found 316.1697.



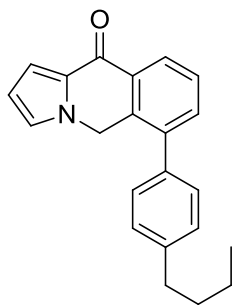
6-(4-Methoxyphenyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3o): 1H NMR (600 MHz, $CDCl_3$, TMS): δ = 8.34 (dd, J = 1.3, 7.7 Hz, 1H), 7.52 (t, J = 7.3 Hz, 1H), 7.44 (dd, J = 1.4, 7.4 Hz, 1H), 7.26 (t, J = 2.9 Hz, 1H), 7.24 (t, J = 2.8 Hz, 1H), 7.19 (dd, J = 1.5, 4.0 Hz, 1H), 7.03 (t, J = 2.9 Hz, 1H), 7.02 (t, J = 2.9 Hz, 1H), 6.95-6.96 (m, 1H), 6.40 (dd, J = 2.4, 3.9 Hz, 1H), 5.19 (s, 2H), 3.89 (s, 3H); ^{13}C NMR (150 MHz, $CDCl_3$, TMS): δ = 174.45, 158.93, 139.26, 133.54, 133.01, 130.54, 129.34, 128.69, 127.15, 125.81, 125.33, 113.76, 113.08, 111.23, 54.89, 45.85; HRMS (ESI) calcd for $C_{19}H_{16}NO_2$ $[M+H]^+ = 290.1176$, found 290.1178.



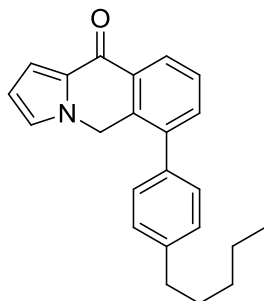
6-(3,5-Bis(trifluoromethyl)phenyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3p): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.49 (dd, J = 1.1, 7.8 Hz, 1H), 8.07 (s, 1H), 7.90 (s, 2H), 7.66 (t, J = 7.5 Hz, 1H), 7.52 (dd, J = 1.1, 7.3 Hz, 1H), 7.26 (dd, J = 1.3, 3.9 Hz, 1H), 7.05 (s, 1H), 6.49 (t, J = 2.5, 4.0 Hz, 1H), 5.19 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.05, 141.42, 136.79, 133.59, 132.81, 132.60, 132.38, 131.46, 129.02, 128.96, 128.22, 127.91, 126.11, 123.99, 122.24, 122.18, 114.15, 114.06, 112.15, 46.00; ^{19}F NMR (282 MHz, CDCl_3 , TMS): δ = -62.72 (s, 6F); HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{12}\text{F}_6\text{NO}$ $[\text{M}+\text{H}]^+ = 396.0818$, found 396.0819.



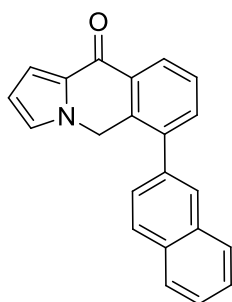
6-(4-Propylphenyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3q): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.39 (dd, J = 1.1, 7.8 Hz, 1H), 7.58 (t, J = 7.2 Hz, 1H), 7.50 (dd, J = 1.3, 7.4 Hz, 1H), 7.35 (d, J = 7.9 Hz, 2H), 7.27-7.29 (m, 2H), 7.23 (dd, J = 1.4, 3.9 Hz, 1H), 7.01 (s, 1H), 6.44 (dd, J = 2.4, 3.9 Hz, 1H), 5.25 (s, 2H), 2.73 (t, J = 7.6 Hz, 2H), 1.74-1.81 (m, 2H), 1.06 (t, J = 7.3 Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.94, 142.65, 140.08, 136.57, 133.94, 133.30, 130.99, 129.18, 128.88, 128.53, 127.64, 126.34, 125.84, 113.57, 111.70, 46.31, 37.80, 24.49, 13.94; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{20}\text{NO}$ $[\text{M}+\text{H}]^+ = 302.1539$, found 302.1540.



6-(4-Butylphenyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3r): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.36 (dd, J = 1.2, 7.8 Hz, 1H), 7.54 (t, J = 7.6 Hz, 1H), 7.46 (dd, J = 1.3, 7.4 Hz, 1H), 7.32 (d, J = 8.0 Hz, 2H), 7.24 (d, J = 8.3 Hz, 2H), 7.20 (dd, J = 1.5, 4.0 Hz, 1H), 6.97 (t, J = 1.6 Hz, 1H), 6.41 (dd, J = 2.4, 3.9 Hz, 1H), 5.21 (s, 2H), 2.72 (t, J = 7.8 Hz, 2H), 1.67-1.72 (m, 2H), 1.42-1.43 (m, 2H), 0.99 (t, J = 7.3 Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.44, 142.40, 139.61, 136.06, 133.44, 132.82, 130.54, 128.73, 128.35, 128.06, 127.16, 125.86, 125.32, 113.06, 111.20, 45.84, 34.94, 33.09, 29.21, 21.99, 13.50; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{21}\text{NO}$ $[\text{M}+\text{H}]^+ = 316.1696$, found 316.1697.

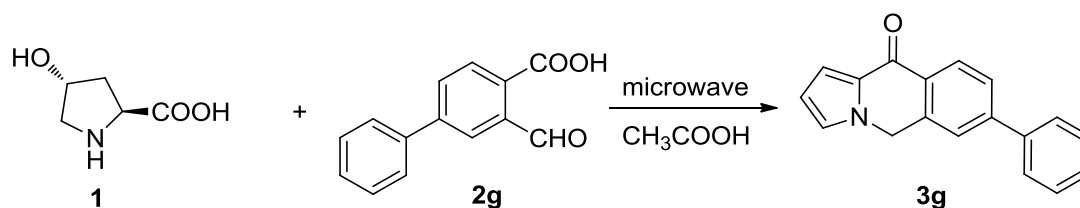


6-(4-Pentylphenyl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3s): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.36 (dt, J = 1.1, 2.3, 7.8 Hz, 1H), 7.52-7.55 (m, 1H), 7.46 (dd, J = 0.7, 7.4 Hz, 1H), 7.30-7.33 (m, 2H), 7.24 (dt, J = 1.7, 3.6, 8.0 Hz, 2H), 7.20 (dt, J = 1.1, 2.4, 4.0 Hz, 1H), 6.96-6.97 (m, 1H), 6.40-6.41 (m, 1H), 5.21 (s, 2H), 2.70 (t, J = 7.7 Hz, 2), 1.69-1.74 (m, 2H), 1.38-1.43 (m, 4H), 0.93-0.96 (m, 3H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.44, 142.45, 139.61, 136.05, 133.44, 132.82, 130.54, 128.72, 128.33, 128.06, 127.15, 125.86, 125.32, 113.06, 111.20, 45.84, 35.23, 31.15, 30.63, 29.21, 22.08, 13.57; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{24}\text{NO}$ $[\text{M}+\text{H}]^+ = 330.1852$, found 330.1853.



6-(Naphthalen-2-yl)pyrrolo[1,2-*b*]isoquinolin-10(5*H*)-one (3t): ^1H NMR (600 MHz, CDCl_3 , TMS): δ = 8.42 (dd, J = 1.5, 7.7 Hz, 1H), 7.99 (d, J = 8.4 Hz, 1H), 7.96 (dd, J = 3.1, 5.9 Hz, 1H), 7.90 (dd, J = 3.4, 6.1 Hz, 1H), 7.82 (d, J = 0.7 Hz, 1H), 7.55-7.61 (m, 4H), 7.46 (dd, J = 1.7, 8.3 Hz, 1H), 7.21 (dd, J = 1.5, 4.0 Hz, 1H), 6.91 (dd, J = 1.6, 2.1 Hz, 1H), 6.40 (dd, J = 2.4, 4.0 Hz, 1H), 5.24 (s, 2H); ^{13}C NMR (150 MHz, CDCl_3 , TMS): δ = 174.35, 139.50, 136.27, 133.42, 132.95, 132.83, 132.30, 130.62, 128.71, 128.14, 127.56, 127.38, 127.27, 127.22, 126.34, 126.16, 126.09, 125.35, 113.17, 111.28, 45.91, 29.21; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{16}\text{NO}$ $[\text{M}+\text{H}]^+ = 310.1226$, found 310.1228.

Table S1. Further exploration of the optimized reaction condition of diphenyl substituted substrate 2g.^a



Entry	Product	1 : 2 (equivalence ratio)	Time (min)	Yield ^b (%)
1	3g	1.2:1	20	20%
2	3g	1:1	20	18%
3	3g	1:1.2	20	25%
4	3g	1:1.2	40	30%

^aMicrowave reaction conditions (unless otherwise specified): solvent (2.0 mL), reaction mixture was heated in the microwave reactor at 170 °C for the appropriate time. ^b Yield of isolated product after column chromatography.

X-ray structure of **3j**

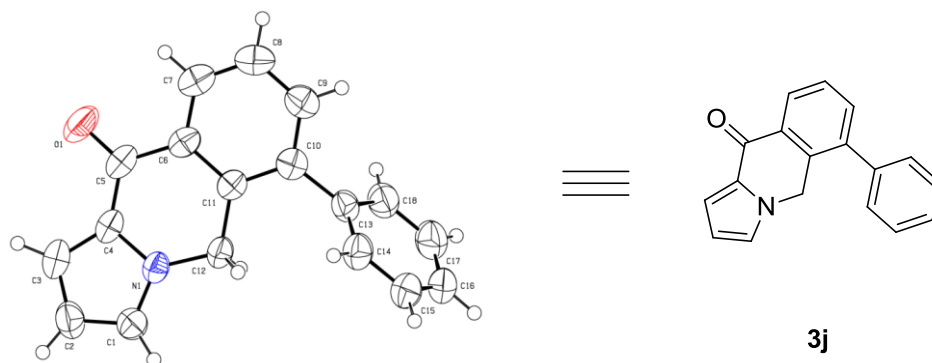


Table 1. Crystal data and structure refinement for a51209b (compound **3j**).

Identification code	a51209b
Empirical formula	C ₁₈ H ₁₃ NO
Formula weight	259.29
Temperature	298(2) K
Wavelength	0.71073 Å
Crystal system	Monoclinic
Space group	P 2 ₁ /c
Unit cell dimensions	a = 10.016(4) Å α = 90°. b = 9.223(3) Å β = 94.662(4)°. c = 13.978(5) Å γ = 90°.
Volume	1287.0(8) Å ³
Z	4
Density (calculated)	1.338 Mg/m ³
Absorption coefficient	0.083 mm ⁻¹

F(000)	544
Crystal size	0.200 x 0.120 x 0.100 mm ³
Theta range for data collection	2.040 to 25.998°.
Index ranges	-10<=h<=12, -11<=k<=9, -17<=l<=16
Reflections collected	5708
Independent reflections	2525 [R(int) = 0.0566]
Completeness to theta = 25.242°	99.8 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.000 and 0.328
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	2525 / 0 / 182
Goodness-of-fit on F ²	0.927
Final R indices [I>2sigma(I)]	R1 = 0.0488, wR2 = 0.1156
R indices (all data)	R1 = 0.0716, wR2 = 0.1226
Extinction coefficient	0.018(3)
Largest diff. peak and hole	0.174 and -0.166 e.Å ⁻³

Table 2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for a51209b (compound **3j**). U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
N(1)	5859(1)	1675(1)	4833(1)	43(1)
O(1)	6071(1)	3067(2)	7222(1)	76(1)
C(1)	6750(2)	877(2)	4390(1)	53(1)
C(2)	7821(2)	580(2)	5037(1)	60(1)
C(3)	7566(2)	1208(2)	5901(1)	57(1)
C(4)	6350(2)	1905(2)	5772(1)	46(1)
C(5)	5666(2)	2864(2)	6376(1)	51(1)
C(6)	4467(2)	3625(2)	5935(1)	46(1)
C(7)	3858(2)	4670(2)	6465(1)	58(1)
C(8)	2711(2)	5358(2)	6099(1)	63(1)
C(9)	2122(2)	4932(2)	5215(1)	58(1)
C(10)	2685(2)	3875(2)	4669(1)	47(1)
C(11)	3906(2)	3250(2)	5015(1)	42(1)
C(12)	4569(2)	2169(2)	4406(1)	45(1)
C(13)	1927(2)	3355(2)	3767(1)	47(1)
C(14)	2385(2)	3536(2)	2867(1)	56(1)
C(15)	1676(2)	2980(2)	2062(1)	66(1)
C(16)	507(2)	2228(2)	2143(2)	70(1)
C(17)	25(2)	2065(2)	3030(2)	71(1)
C(18)	728(2)	2626(2)	3834(1)	58(1)

Table 3. Bond lengths [Å] and angles [°] for a51209b (compound **3j**).

N(1)-C(1)	1.345(2)
N(1)-C(4)	1.3805(19)
N(1)-C(12)	1.4515(19)
O(1)-C(5)	1.2336(19)
C(1)-C(2)	1.374(2)
C(2)-C(3)	1.383(3)
C(3)-C(4)	1.376(2)
C(4)-C(5)	1.435(2)
C(5)-C(6)	1.481(2)
C(6)-C(7)	1.387(2)
C(6)-C(11)	1.404(2)
C(7)-C(8)	1.374(3)
C(8)-C(9)	1.383(2)
C(9)-C(10)	1.386(2)
C(10)-C(11)	1.402(2)
C(10)-C(13)	1.498(2)
C(11)-C(12)	1.500(2)
C(13)-C(14)	1.383(2)
C(13)-C(18)	1.387(2)
C(14)-C(15)	1.381(3)
C(15)-C(16)	1.373(3)
C(16)-C(17)	1.375(3)
C(17)-C(18)	1.377(3)
C(1)-N(1)-C(4)	108.89(13)
C(1)-N(1)-C(12)	125.72(13)
C(4)-N(1)-C(12)	125.35(13)
N(1)-C(1)-C(2)	108.53(15)

C(1)-C(2)-C(3)	107.56(16)
C(4)-C(3)-C(2)	107.81(15)
C(3)-C(4)-N(1)	107.21(15)
C(3)-C(4)-C(5)	132.14(15)
N(1)-C(4)-C(5)	120.26(14)
O(1)-C(5)-C(4)	121.46(16)
O(1)-C(5)-C(6)	121.32(16)
C(4)-C(5)-C(6)	117.21(13)
C(7)-C(6)-C(11)	120.02(16)
C(7)-C(6)-C(5)	118.92(14)
C(11)-C(6)-C(5)	120.99(14)
C(8)-C(7)-C(6)	120.83(16)
C(7)-C(8)-C(9)	119.01(17)
C(8)-C(9)-C(10)	121.88(16)
C(9)-C(10)-C(11)	118.84(15)
C(9)-C(10)-C(13)	119.16(15)
C(11)-C(10)-C(13)	121.84(14)
C(10)-C(11)-C(6)	119.17(14)
C(10)-C(11)-C(12)	119.44(13)
C(6)-C(11)-C(12)	121.38(14)
N(1)-C(12)-C(11)	113.33(12)
C(14)-C(13)-C(18)	118.25(16)
C(14)-C(13)-C(10)	122.99(15)
C(18)-C(13)-C(10)	118.74(15)
C(15)-C(14)-C(13)	120.67(18)
C(16)-C(15)-C(14)	120.35(19)
C(15)-C(16)-C(17)	119.66(18)
C(16)-C(17)-C(18)	120.04(18)
C(17)-C(18)-C(13)	120.99(17)

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for a51209b (compound **3j**).

The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U^{11}	U^{22}	U^{33}	U^{23}	U^{13}	U^{12}
N(1)	43(1)	50(1)	36(1)	4(1)	-3(1)	-1(1)
O(1)	92(1)	90(1)	42(1)	-13(1)	-16(1)	1(1)
C(1)	53(1)	63(1)	44(1)	1(1)	3(1)	8(1)
C(2)	48(1)	72(1)	59(1)	10(1)	-2(1)	10(1)
C(3)	51(1)	66(1)	52(1)	8(1)	-14(1)	-8(1)
C(4)	48(1)	53(1)	37(1)	3(1)	-6(1)	-9(1)
C(5)	59(1)	54(1)	37(1)	-1(1)	-5(1)	-14(1)
C(6)	52(1)	46(1)	40(1)	-2(1)	6(1)	-11(1)
C(7)	70(1)	56(1)	47(1)	-7(1)	7(1)	-11(1)
C(8)	74(1)	53(1)	64(1)	-10(1)	18(1)	0(1)
C(9)	58(1)	54(1)	62(1)	2(1)	9(1)	6(1)
C(10)	50(1)	45(1)	46(1)	6(1)	7(1)	-3(1)
C(11)	46(1)	42(1)	38(1)	3(1)	4(1)	-6(1)
C(12)	44(1)	52(1)	37(1)	2(1)	-5(1)	0(1)
C(13)	44(1)	46(1)	50(1)	4(1)	-2(1)	8(1)
C(14)	51(1)	65(1)	52(1)	10(1)	-2(1)	0(1)
C(15)	66(1)	79(1)	50(1)	2(1)	-6(1)	14(1)
C(16)	56(1)	80(1)	70(1)	-17(1)	-18(1)	14(1)
C(17)	47(1)	81(1)	85(2)	-9(1)	-4(1)	-2(1)
C(18)	48(1)	66(1)	60(1)	2(1)	3(1)	4(1)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for a51209b (compound **3j**).

	x	y	z	U(eq)
H(1)	6656	577	3753	64
H(2)	8580	51	4915	72
H(3)	8118	1168	6468	69
H(7)	4231	4909	7076	69
H(8)	2337	6099	6441	76
H(9)	1324	5368	4979	69
H(12A)	3983	1338	4297	54
H(12B)	4691	2604	3788	54
H(14)	3179	4036	2805	67
H(15)	1991	3115	1460	79
H(16)	45	1831	1602	84
H(17)	-777	1575	3087	85
H(18)	393	2513	4431	70

Table 6. Torsion angles [°] for a51209b (compound **3j**).

C(4)-N(1)-C(1)-C(2)	0.16(19)
C(12)-N(1)-C(1)-C(2)	-177.68(15)
N(1)-C(1)-C(2)-C(3)	0.5(2)
C(1)-C(2)-C(3)-C(4)	-0.9(2)
C(2)-C(3)-C(4)-N(1)	1.02(19)
C(2)-C(3)-C(4)-C(5)	-171.60(17)
C(1)-N(1)-C(4)-C(3)	-0.74(18)
C(12)-N(1)-C(4)-C(3)	177.12(14)
C(1)-N(1)-C(4)-C(5)	172.93(14)
C(12)-N(1)-C(4)-C(5)	-9.2(2)
C(3)-C(4)-C(5)-O(1)	-9.5(3)
N(1)-C(4)-C(5)-O(1)	178.68(15)
C(3)-C(4)-C(5)-C(6)	169.33(17)
N(1)-C(4)-C(5)-C(6)	-2.5(2)
O(1)-C(5)-C(6)-C(7)	5.3(2)
C(4)-C(5)-C(6)-C(7)	-173.53(14)
O(1)-C(5)-C(6)-C(11)	-171.45(15)
C(4)-C(5)-C(6)-C(11)	9.7(2)
C(11)-C(6)-C(7)-C(8)	-0.4(3)
C(5)-C(6)-C(7)-C(8)	-177.22(15)
C(6)-C(7)-C(8)-C(9)	3.9(3)
C(7)-C(8)-C(9)-C(10)	-2.8(3)
C(8)-C(9)-C(10)-C(11)	-1.7(3)
C(8)-C(9)-C(10)-C(13)	173.75(16)
C(9)-C(10)-C(11)-C(6)	5.1(2)
C(13)-C(10)-C(11)-C(6)	-170.25(14)
C(9)-C(10)-C(11)-C(12)	-176.39(14)
C(13)-C(10)-C(11)-C(12)	8.3(2)

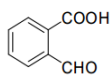
C(7)-C(6)-C(11)-C(10)	-4.1(2)
C(5)-C(6)-C(11)-C(10)	172.62(14)
C(7)-C(6)-C(11)-C(12)	177.44(14)
C(5)-C(6)-C(11)-C(12)	-5.8(2)
C(1)-N(1)-C(12)-C(11)	-169.85(14)
C(4)-N(1)-C(12)-C(11)	12.6(2)
C(10)-C(11)-C(12)-N(1)	176.78(13)
C(6)-C(11)-C(12)-N(1)	-4.8(2)
C(9)-C(10)-C(13)-C(14)	116.81(18)
C(11)-C(10)-C(13)-C(14)	-67.8(2)
C(9)-C(10)-C(13)-C(18)	-65.0(2)
C(11)-C(10)-C(13)-C(18)	110.39(18)
C(18)-C(13)-C(14)-C(15)	-1.1(3)
C(10)-C(13)-C(14)-C(15)	177.13(15)
C(13)-C(14)-C(15)-C(16)	-0.5(3)
C(14)-C(15)-C(16)-C(17)	1.8(3)
C(15)-C(16)-C(17)-C(18)	-1.4(3)
C(16)-C(17)-C(18)-C(13)	-0.2(3)
C(14)-C(13)-C(18)-C(17)	1.5(3)
C(10)-C(13)-C(18)-C(17)	-176.83(16)

Symmetry transformations used to generate equivalent atoms:

NMR Spectra

NMR of compound 2a

wsc-0802-1a H

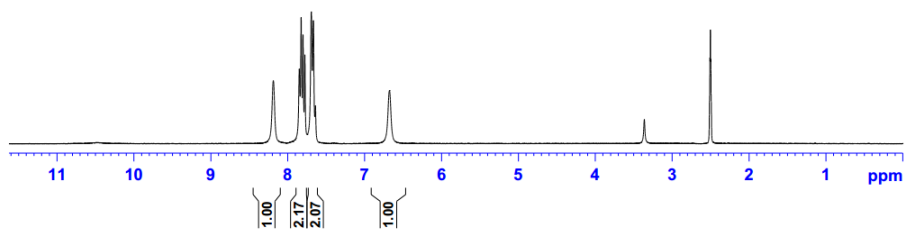


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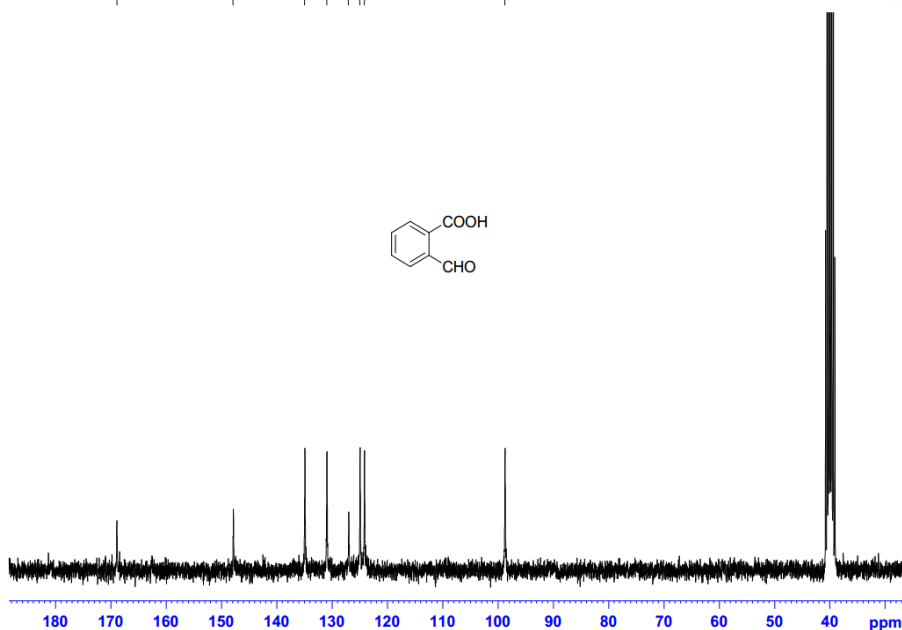
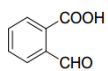
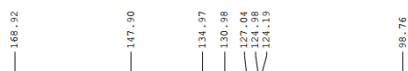
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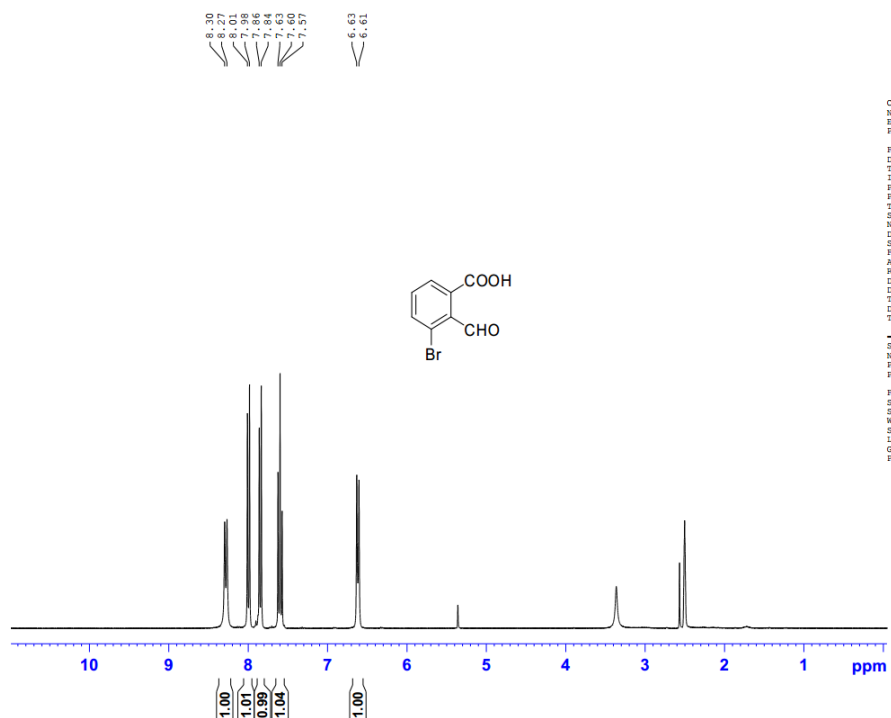


wsc-0802-1a C



NMR of compound **2c**

WSC-0827 H



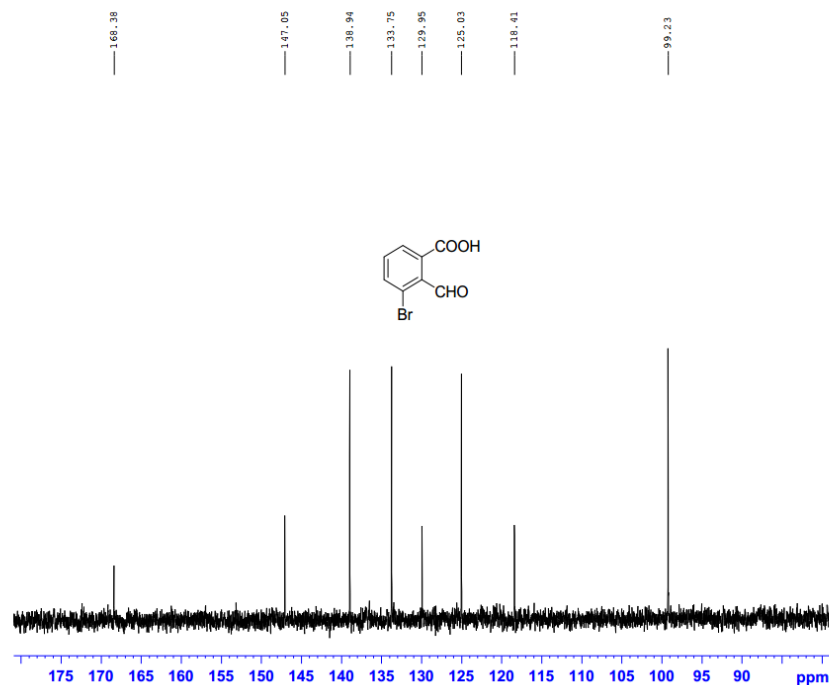
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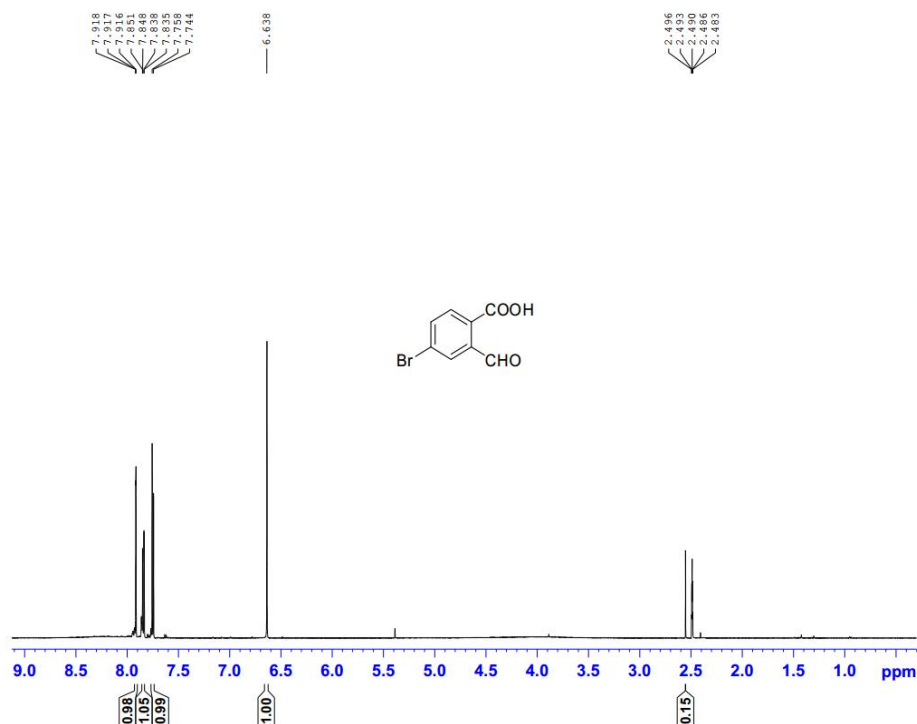
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WSC-0827 C



NMR of compound 2d

wsc-0867



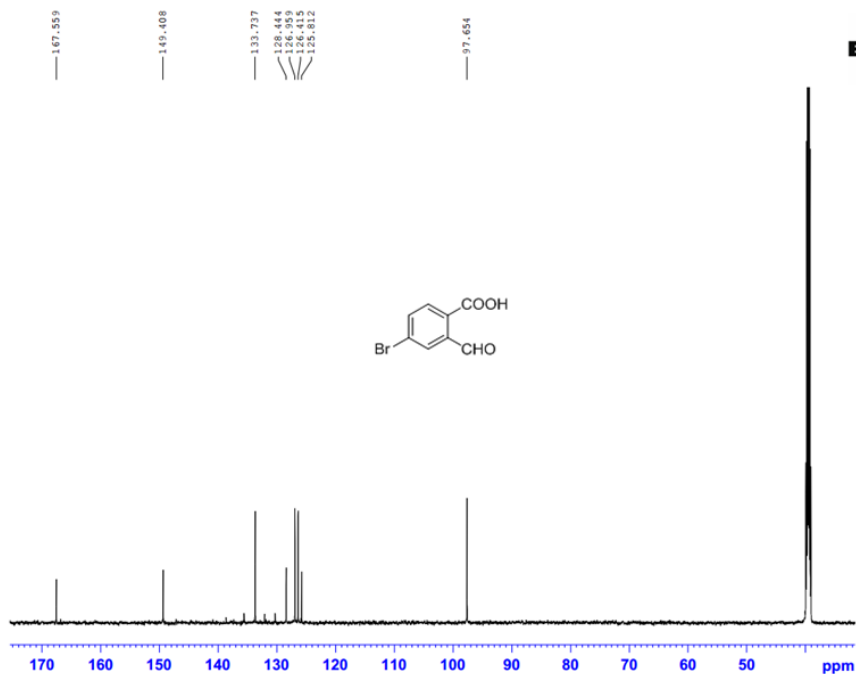
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TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 13.70 usec
PL1 19.34000015 dB

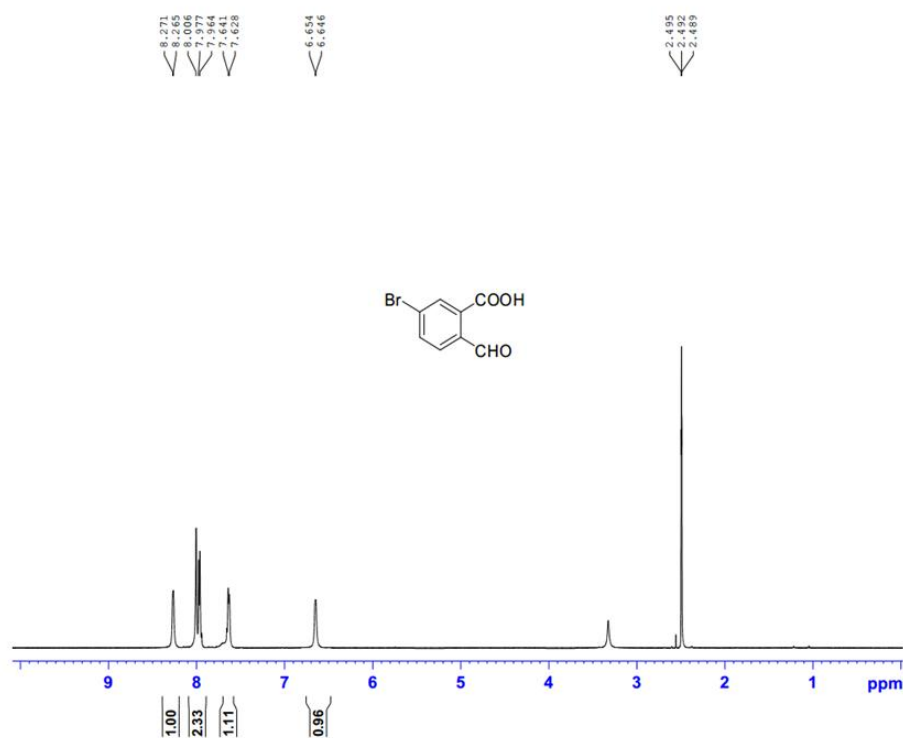
F2 - Processing parameters
SI 65536
SF 600.1300126 MHz
WDW EM
SSB 0
LB 0 Hz
GB 0
PC 1.00

wsc-0867 C



NMR of compound **2e**

wsc-6Br-CC



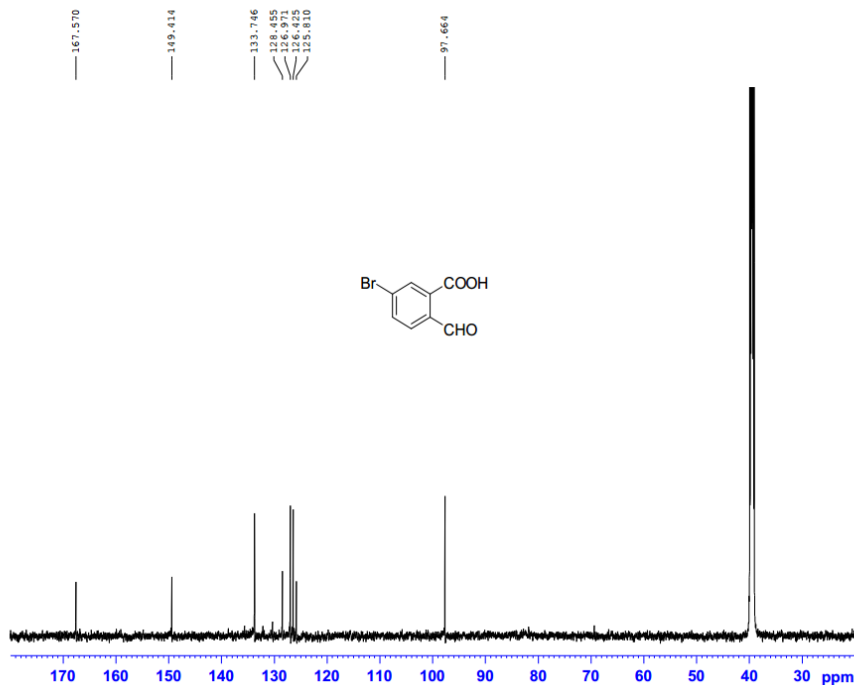
Current Data Parameters
NAME wsc-6Br-CC
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160526
Time_ 10.05
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 0
SWH 12019.230 Hz
FIDRES 0.183399 Hz
AQ 2.7263477 sec
RG 234
DM 41.600 usec
DE 6.50 usec
TE 298.7 K
D1 1.00000000 sec
TD 1

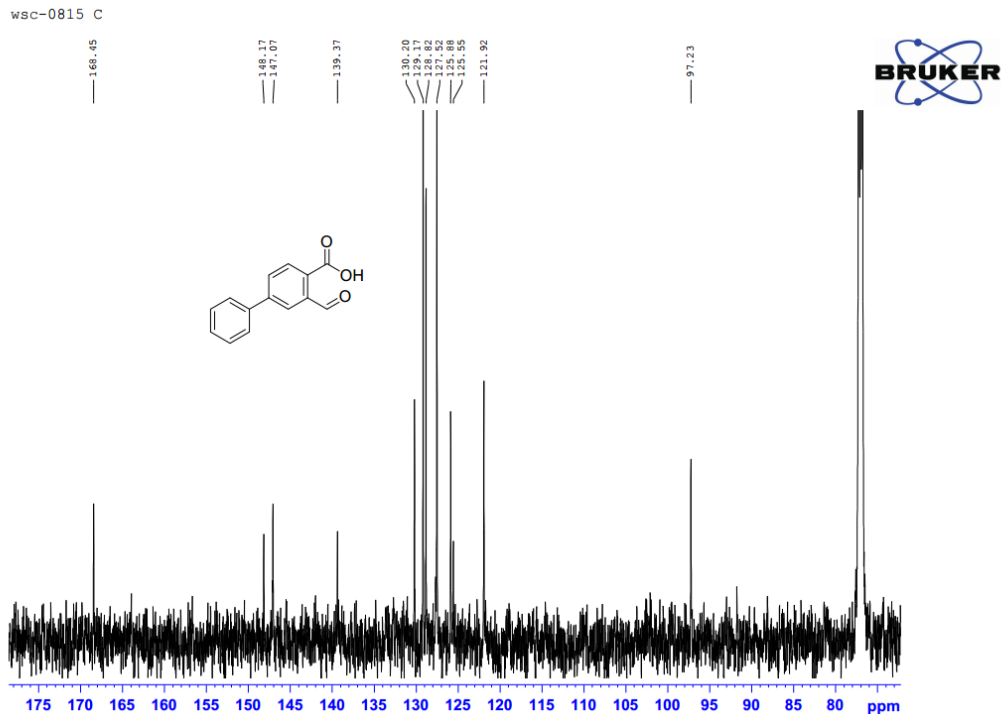
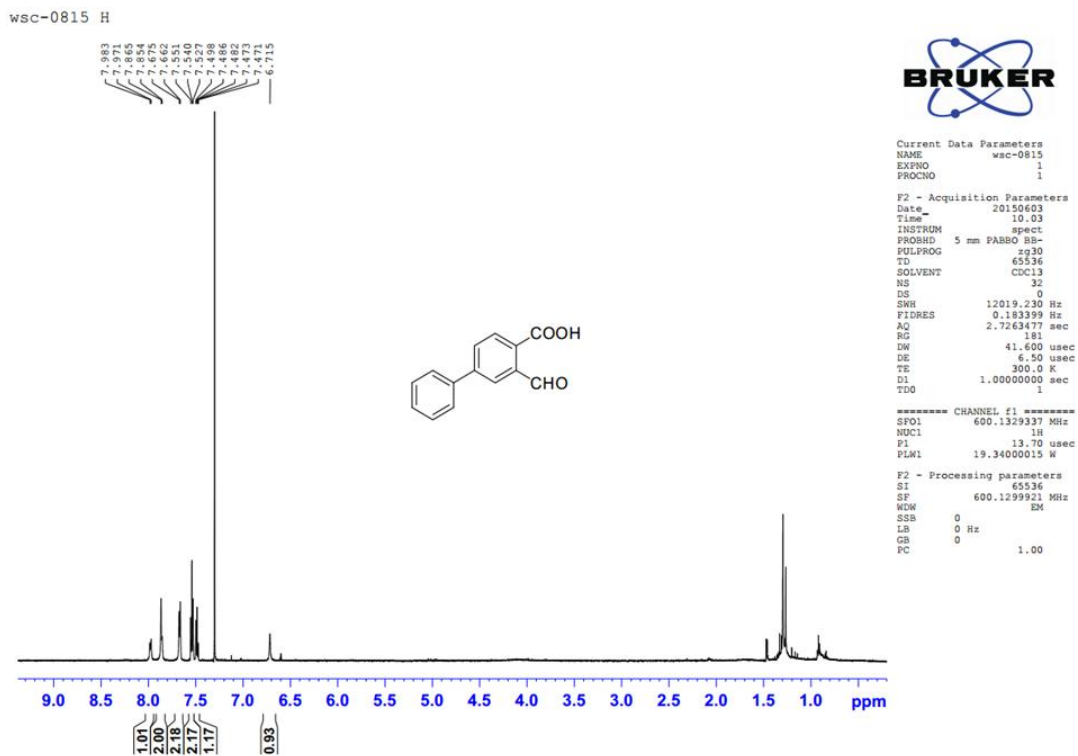
CHANNEL f1 -----
SF01 600.1329337 MHz
NUC1 1H
P1 13.70 usec
PLW1 19.34000015 W

F2 - Processing parameters
SI 65536
SF 600.1300106 MHz
WDW EM
SSB 0
LB 0 Hz
GB 0
PC 1.00

wsc-0876second C

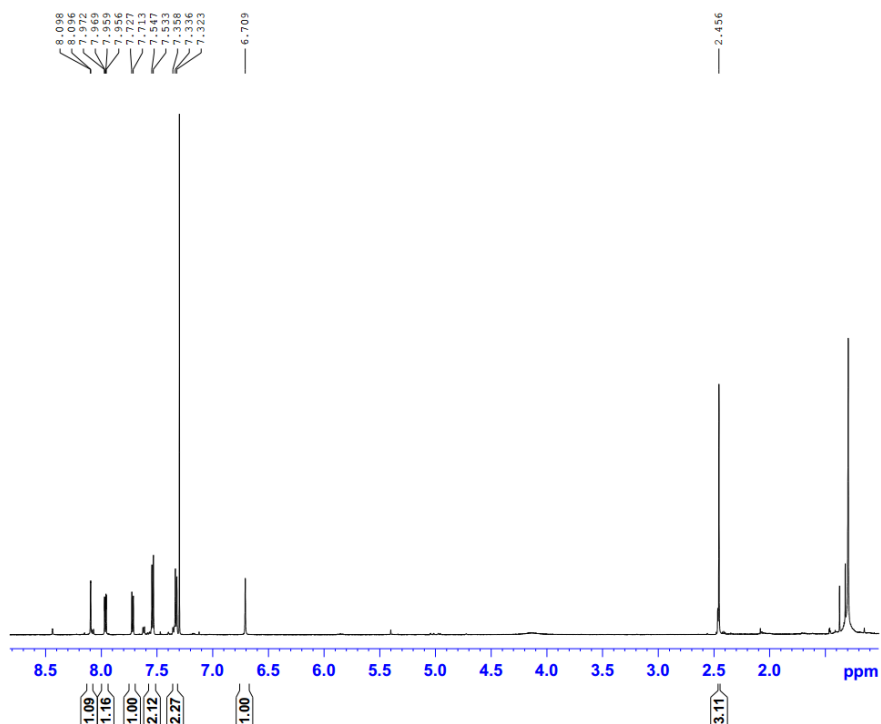


NMR of compound **2g**

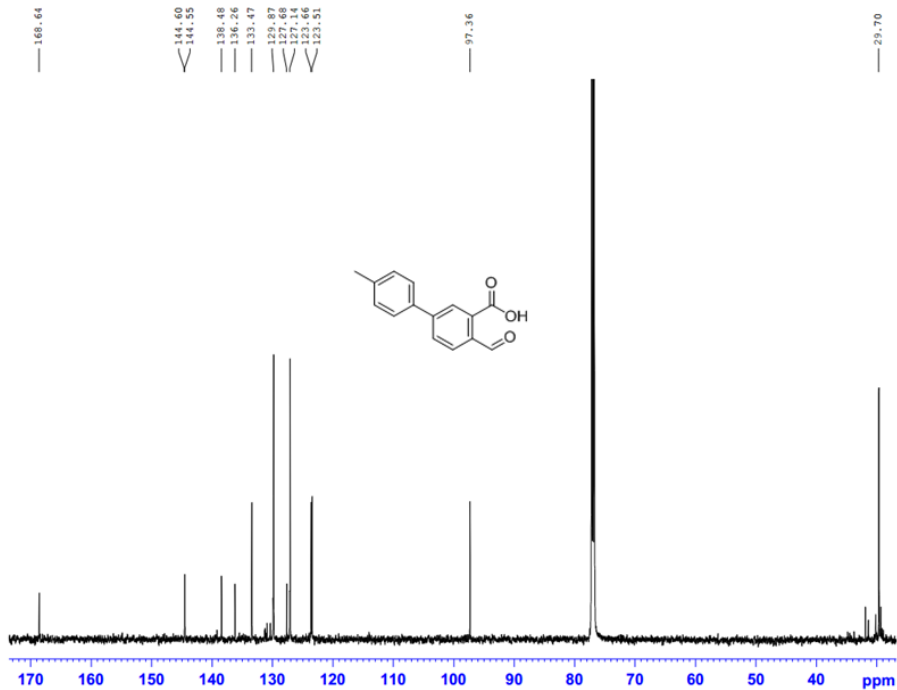


NMR of compound 2i

wsc-08442new H

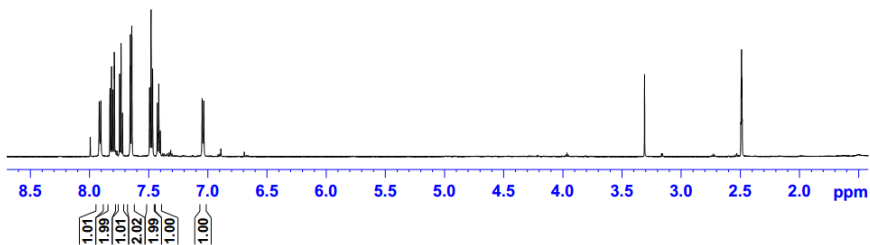
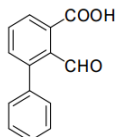


wsc-08442new C



NMR of compound **2j**

wsc-0866ne



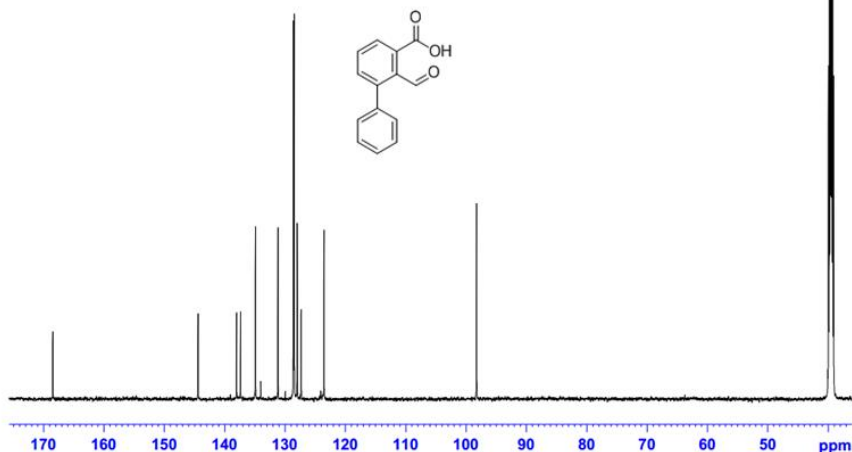
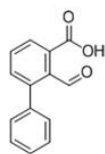
Current Data Parameters
 NAME wsc-0866new
 EXPRO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20150904
 Time 7.01
 INSTRUM spect
 PROHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 0
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7263477 sec
 RG 256
 DW 41.600 usec
 DE 6.50 usec
 TE 299.5 K
 D1 1.00000000 sec
 TDO

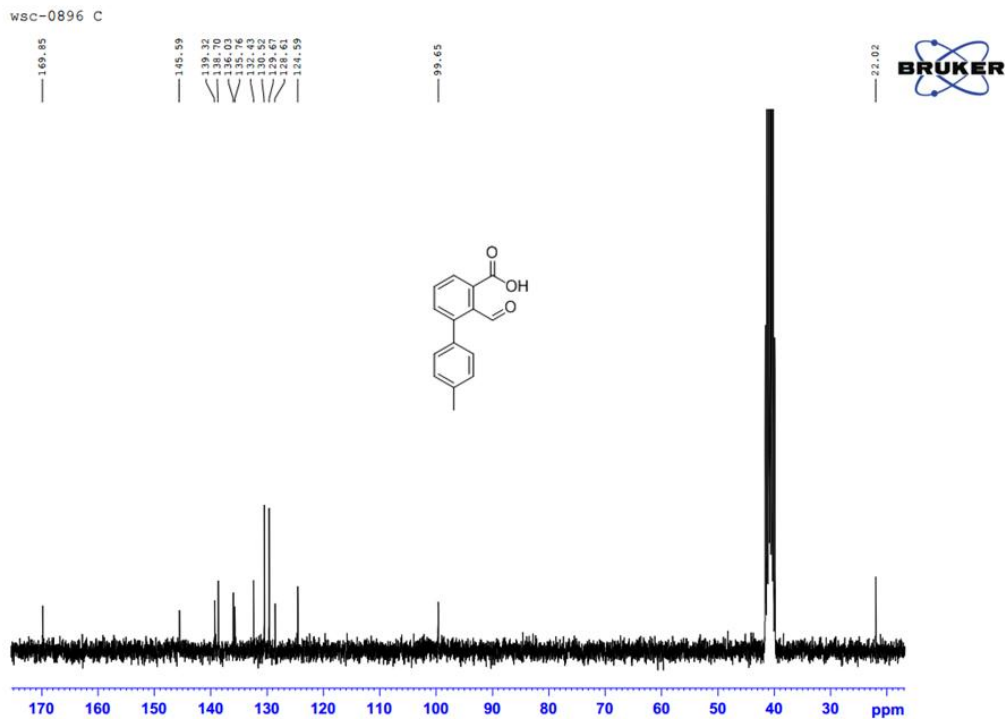
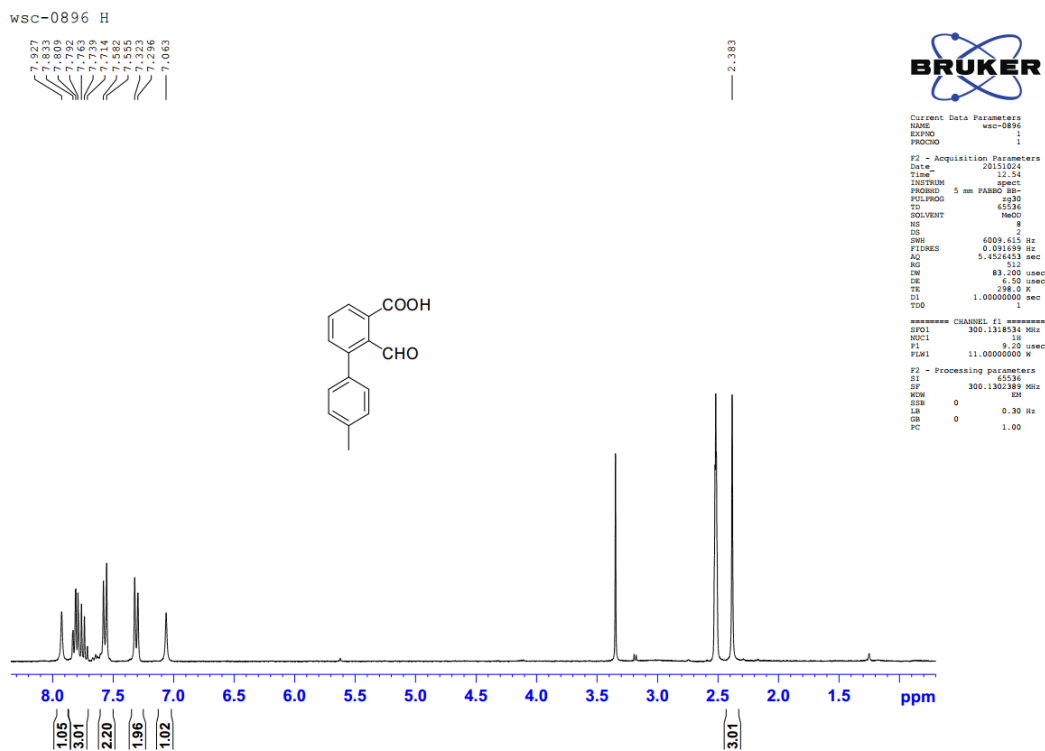
===== CHANNEL f1 =====
 SF01 600.1332000 MHz
 NUC1 1H
 F1 13.70 usec
 PLW1 19.34000015 W

F2 - Processing parameters
 SI 65536
 SF 600.1300121 MHz
 WDW EM
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

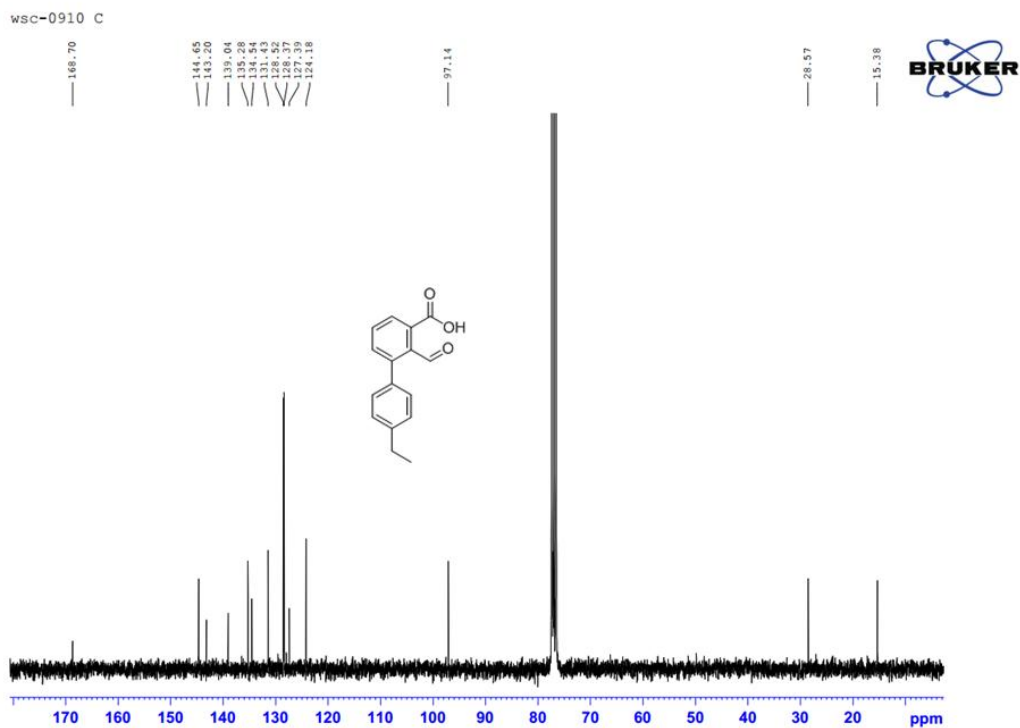
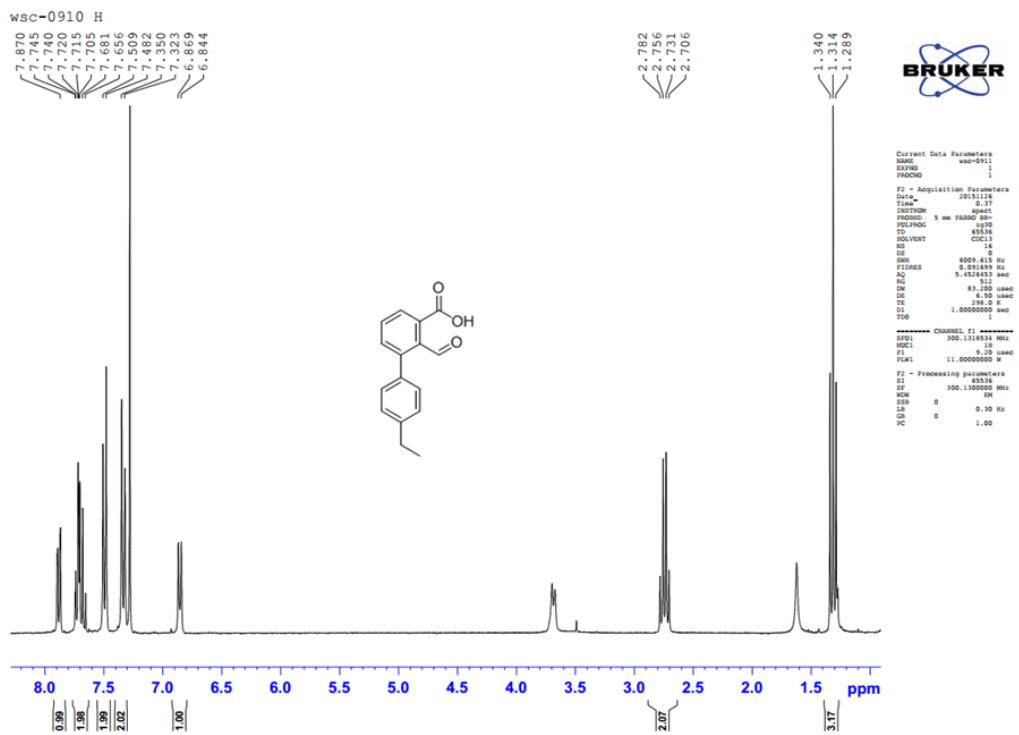
wsc-0866new



NMR of compound **2k**

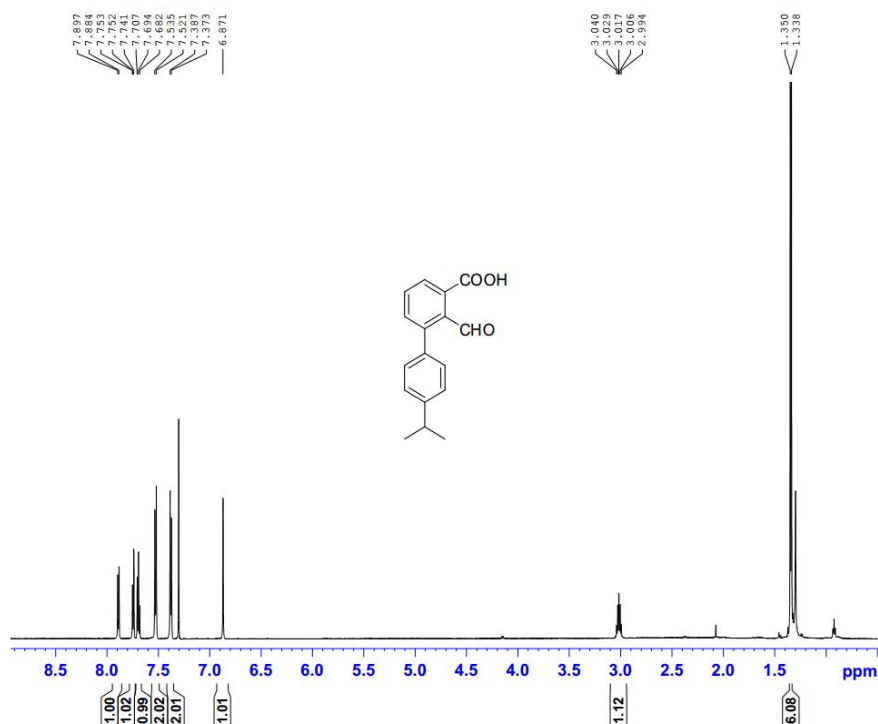


NMR of compound **2l**



NMR of compound **2m**

wsc-cc-2m



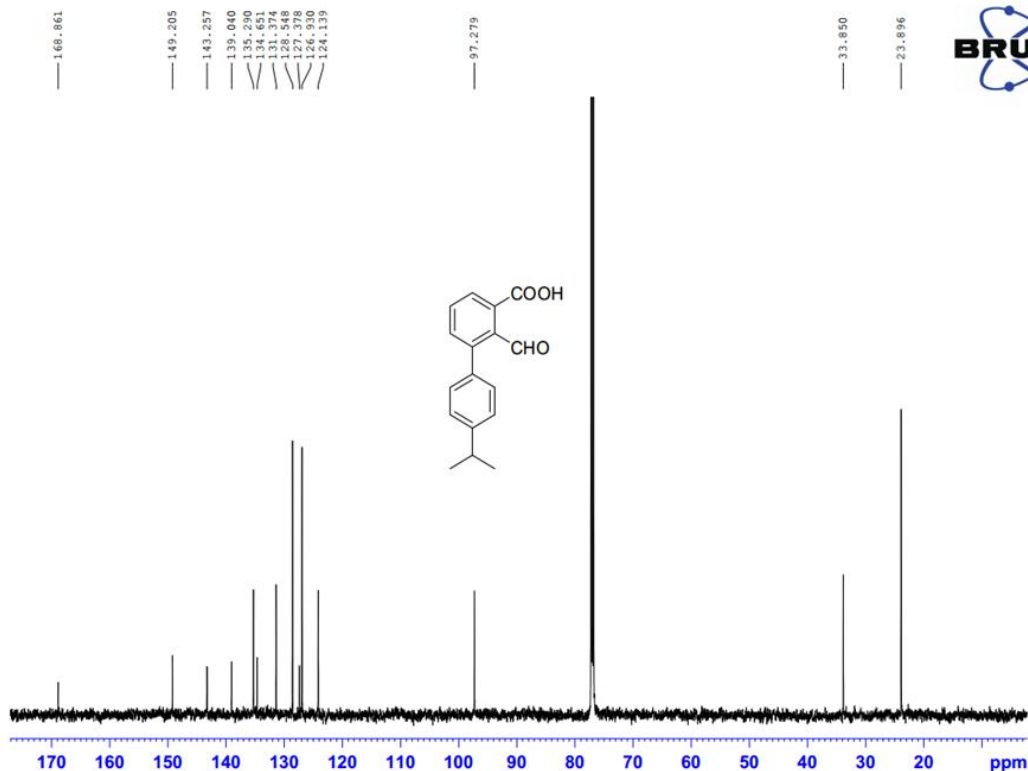
Current Data Parameters
NAME wsc-cc-3m
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160522
Time 12.41
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 18516
SOLVENT CDCl3
NS 16
DS 0
SWH 9259.259 Hz
FIDRES 0.500068 Hz
AQ 0.9999140 sec
RG 228
DW 54.000 usec
DE 6.50 usec
TE 296.7 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====
SP01 600.1325000 MHz
NUC1 1H
P1 13.70 usec
PLW1 19.34000015 W

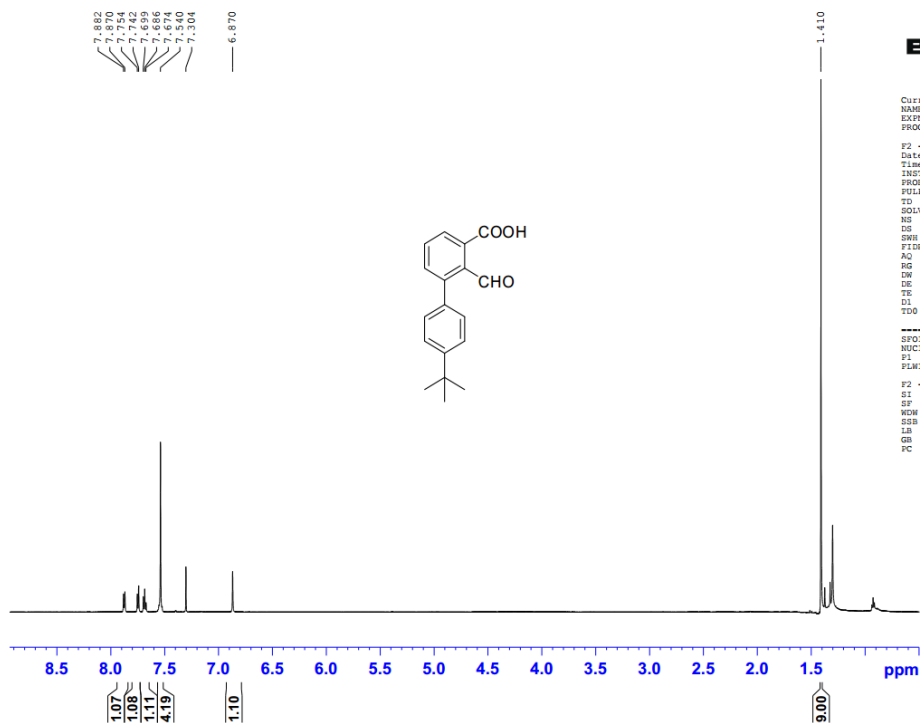
F2 - Processing parameters
SI 65536
SF 600.1299907 MHz
WDW EM
SSB 0
LB 0 Hz
GB 0
PC 1.00

wsc-cc-2m



NMR of compound **2n**

wsc-cc-2n H



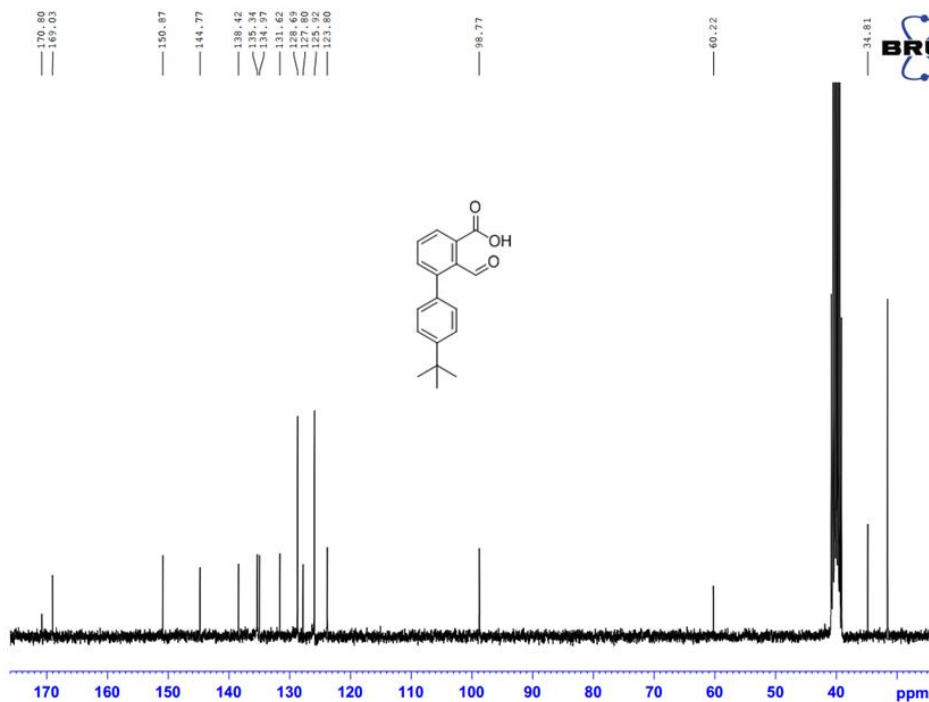
Current Data Parameters
NAME wsc-cc-2n
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160511
Time 20.03
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 183.6
SOLVENT CDCl3
NS 16
DS 0
SWH 9259.259 Hz
FIDRES 0.500068 Hz
AQ 0.9999140 sec
RG 101
DW 54.000 usec
DE 6.50 usec
TE 296.6 K
D1 1.00000000 sec
TD0 1

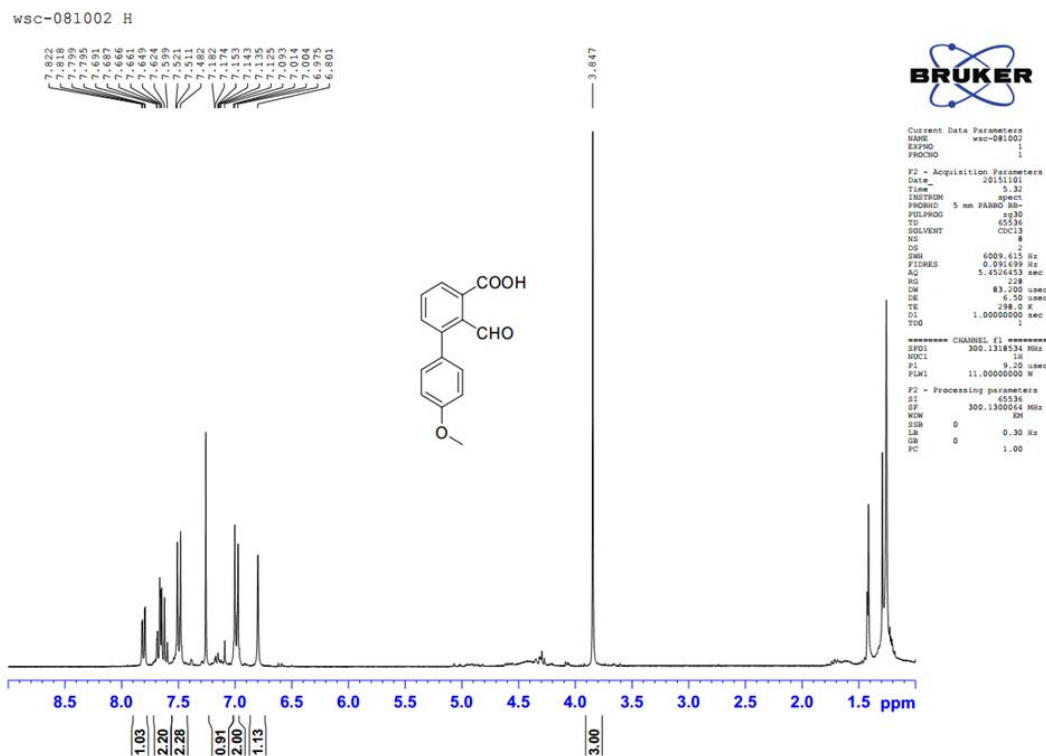
CHANNEL f1
SFO1 600.1325000 MHz
NUC1 13
P1 13.70 usec
PL1 19.34000015 W

F2 - Processing parameters
SI 653.6
SF 600.1299907 MHz
WOW 2M
SBB 0
LA 0 Hz
GB 0
PC 1.00

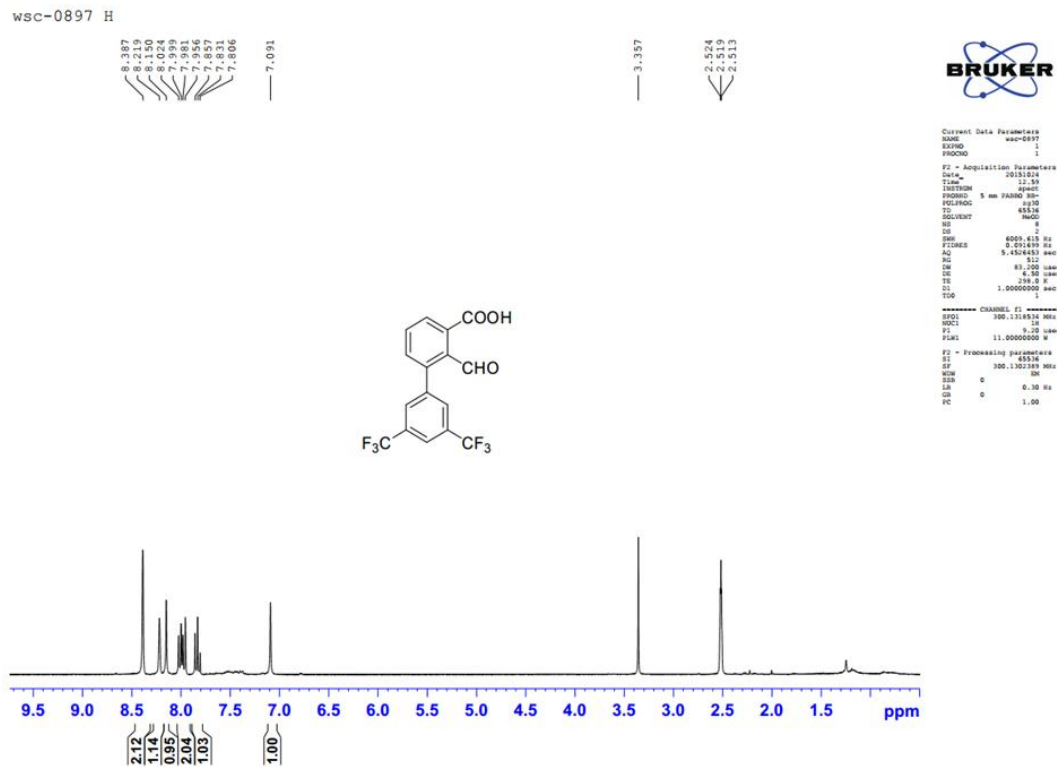
wsc-0919 C



NMR of compound 2o



NMR of compound 2p



wsc-0897 C

169.43

146.19

141.16

136.80

136.08

132.75

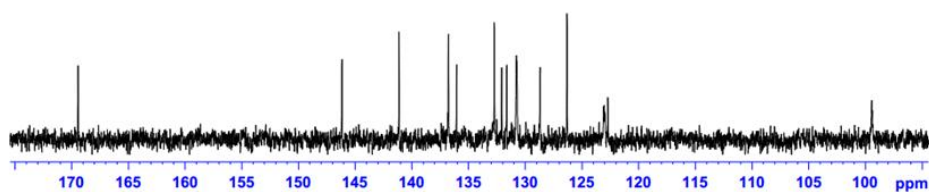
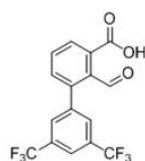
132.10

130.72

128.73

126.35

99.47



NMR of compound 2q

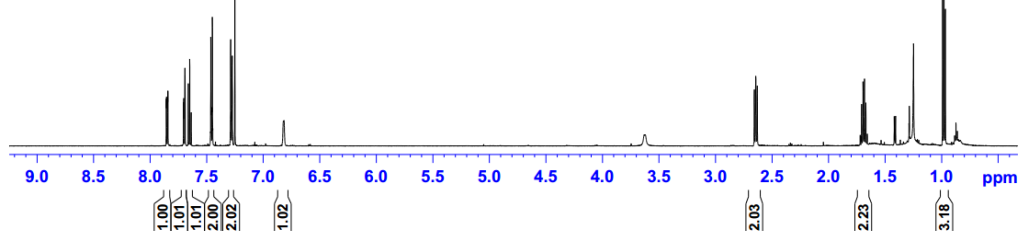
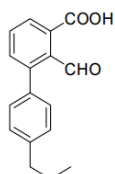
wsc-0928 H

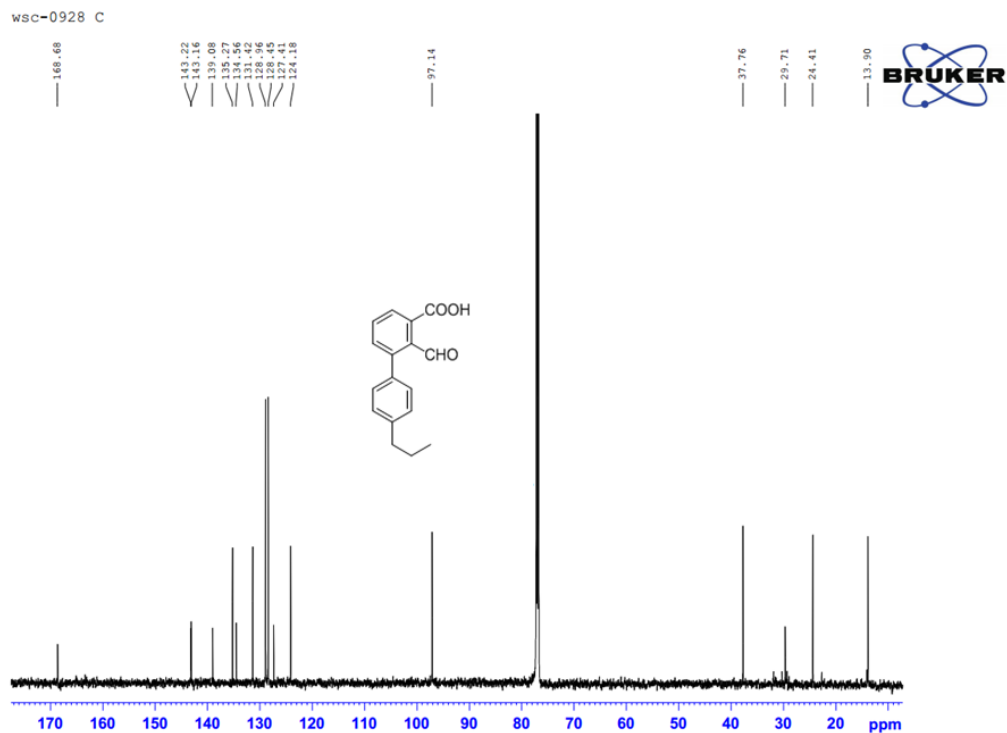
7.858
7.856
7.846
7.844
7.706
7.705
7.692
7.664
7.651
7.648
7.465
7.461
7.451
7.447
7.289
6.823
6.817

2.658
2.655
2.632

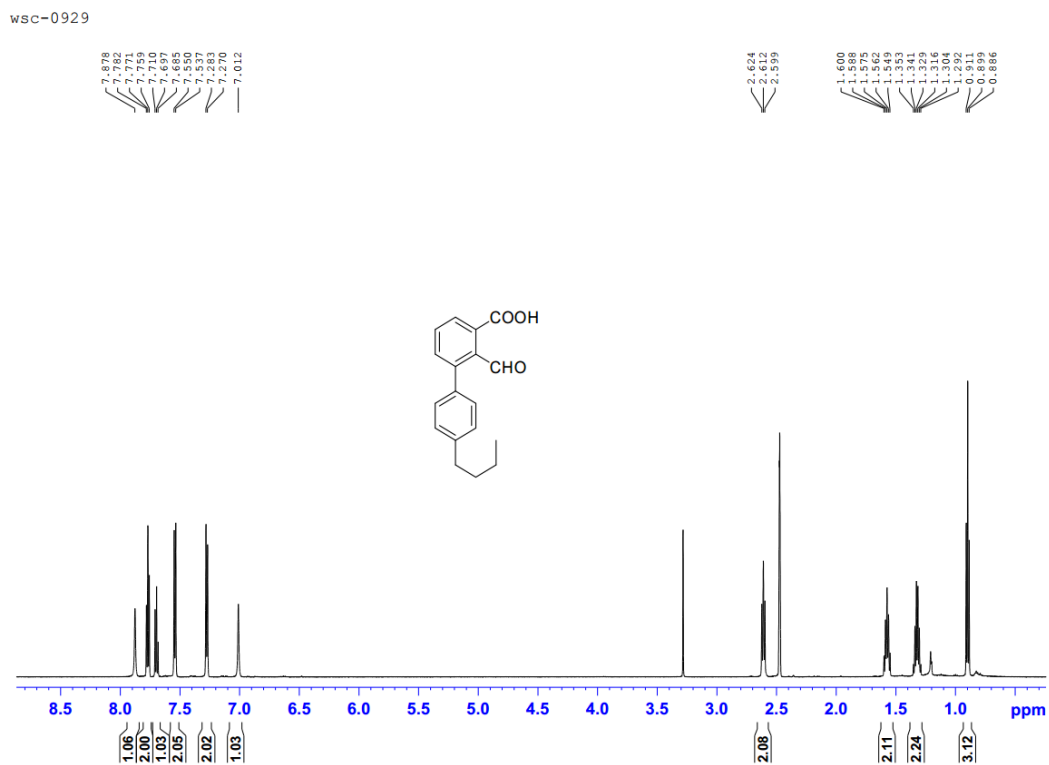
1.721
1.709
1.696
1.671
1.659

0.993
0.980
0.968

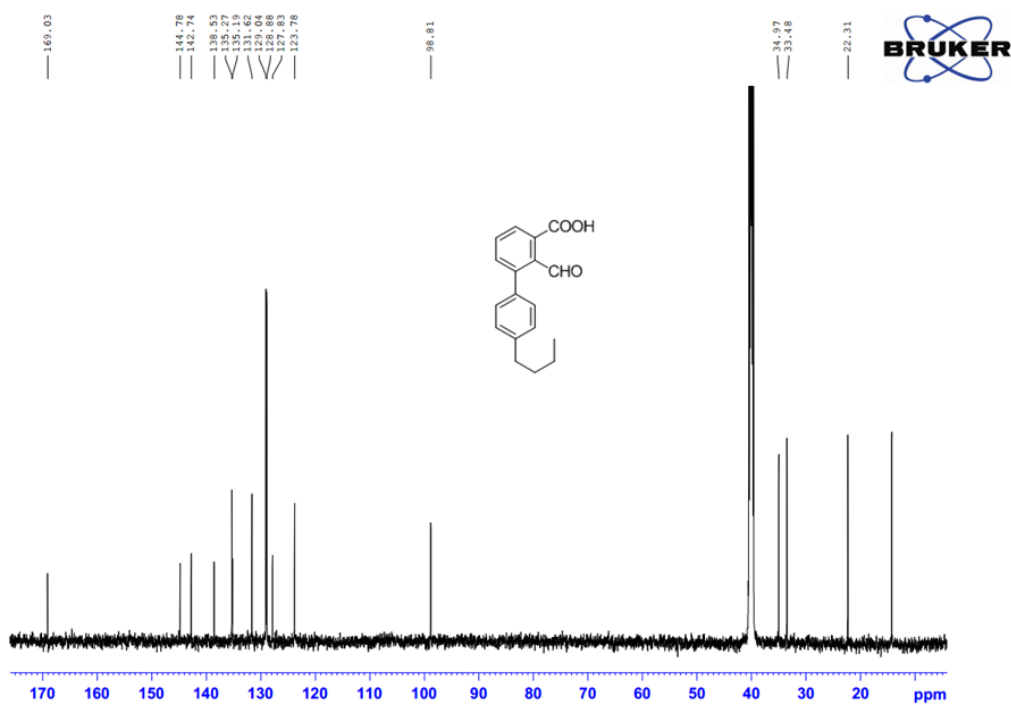




NMR of compound 2r

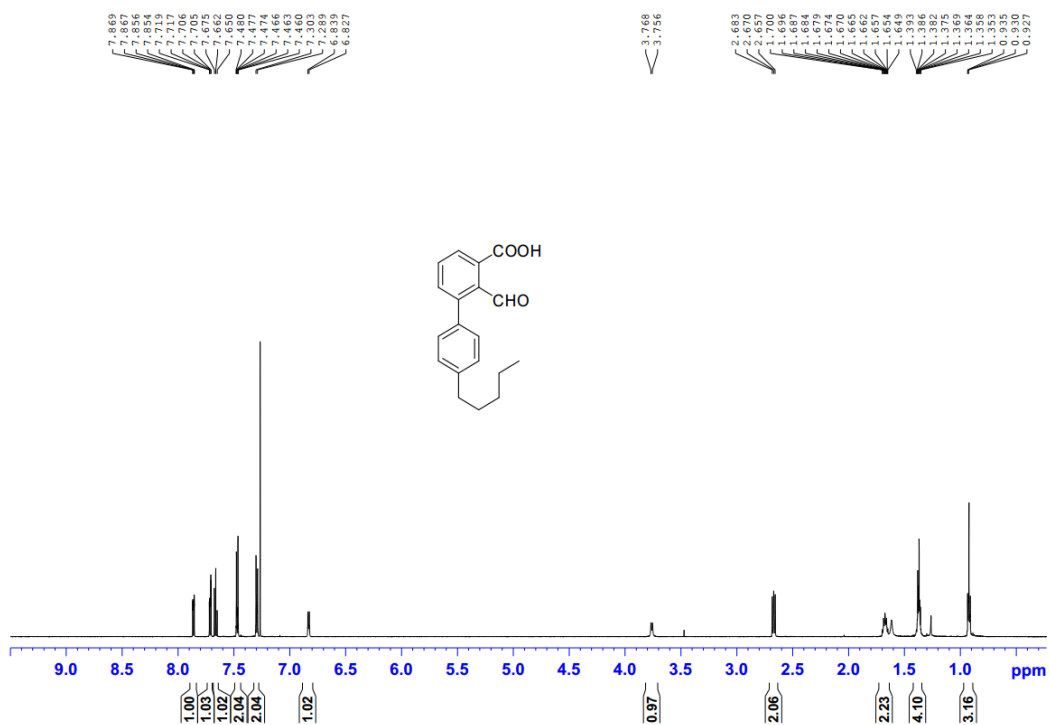


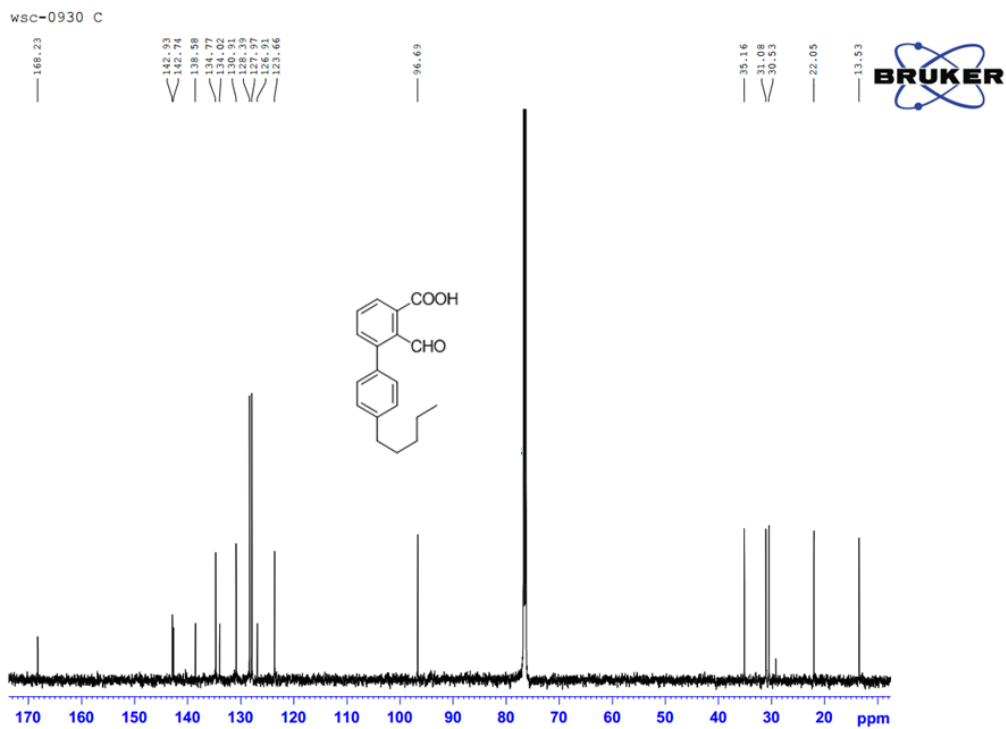
wsc-0929 C



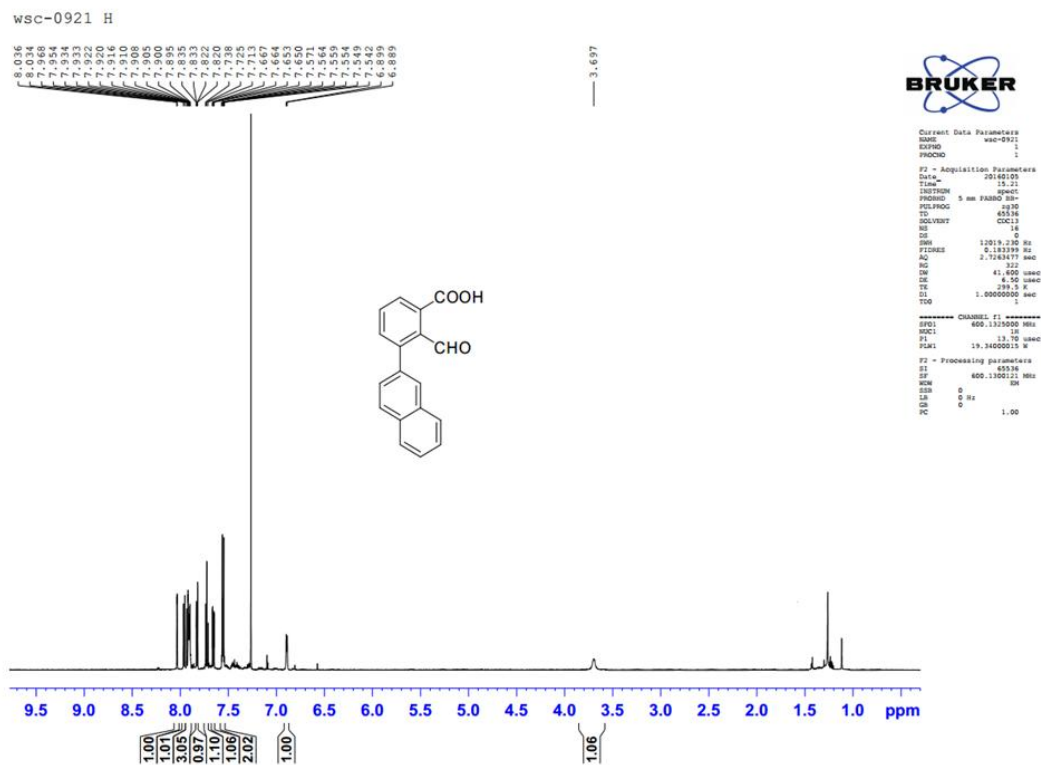
NMR of compound 2s

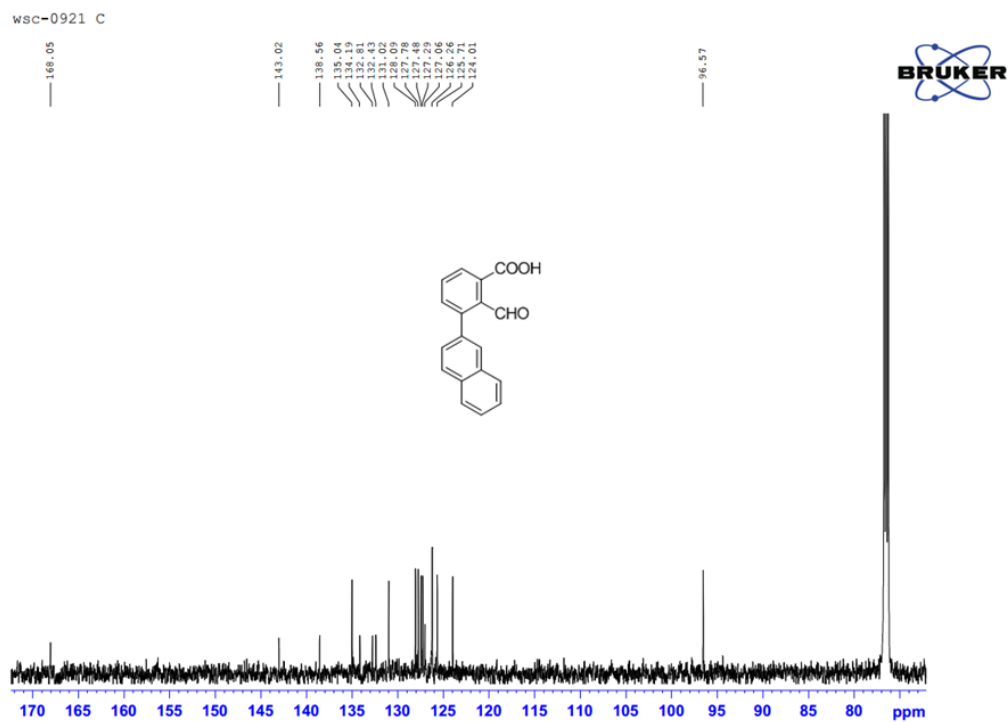
wsc-0930 H



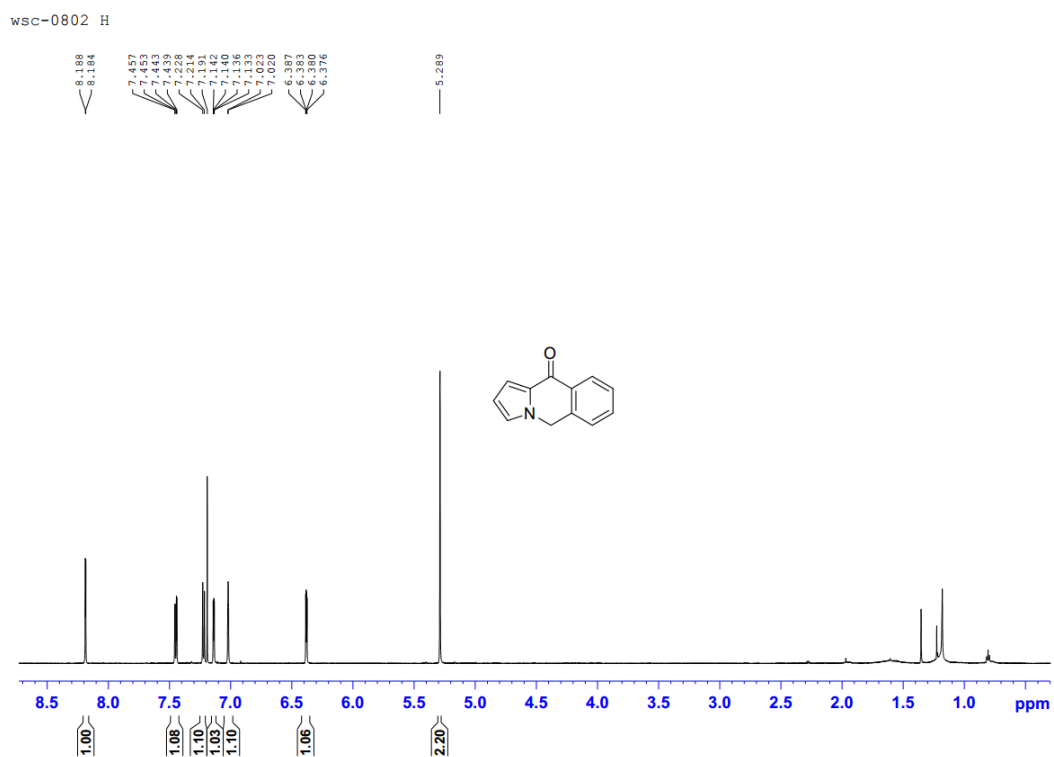


NMR of compound 2t

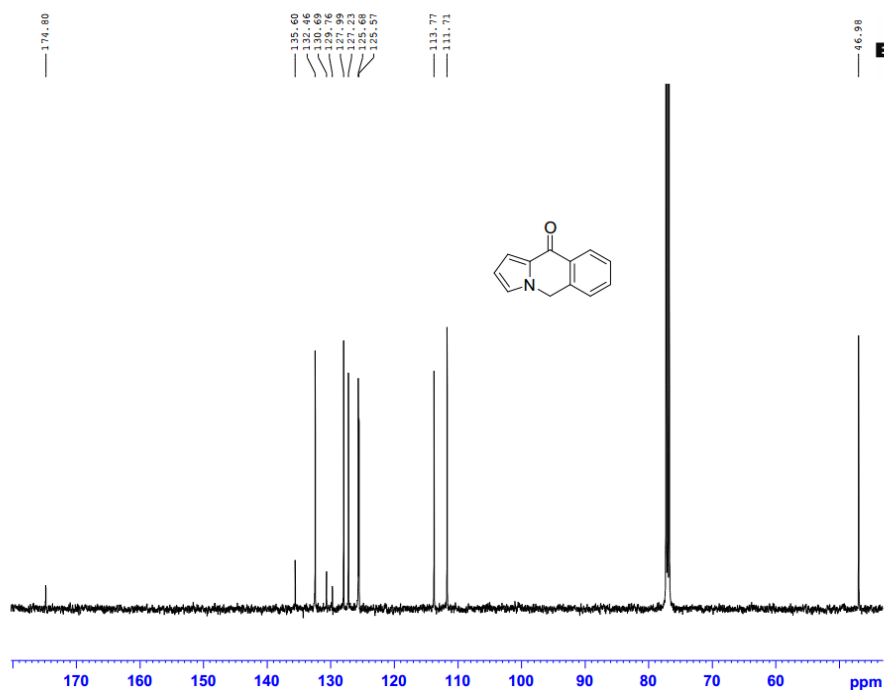




NMR of compound 3a

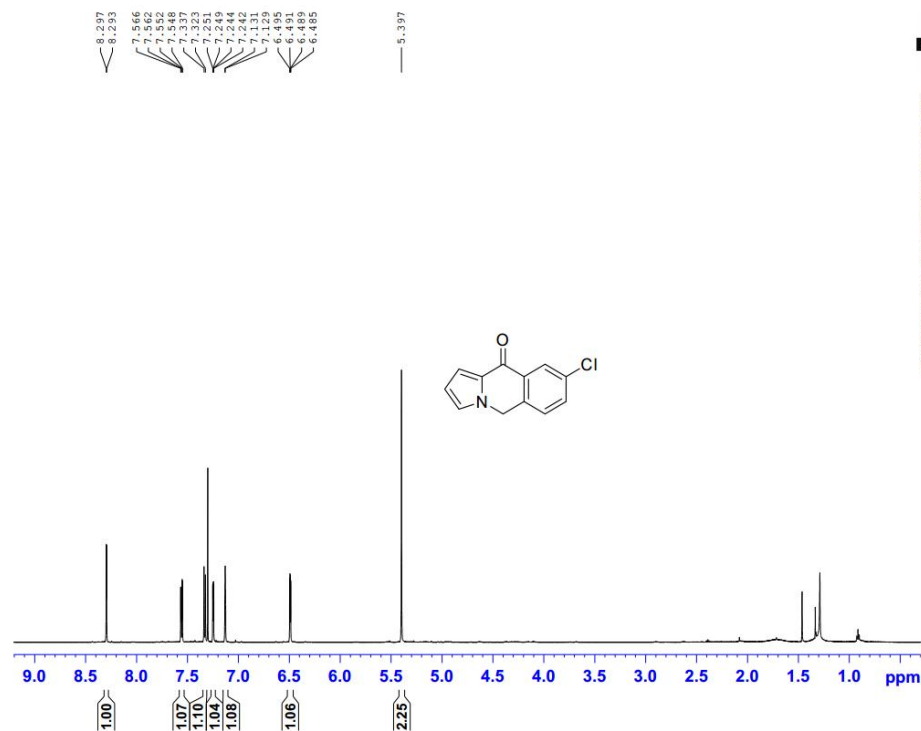


wsc-0802 C



NMR of compound 3b

wsc-0804 H

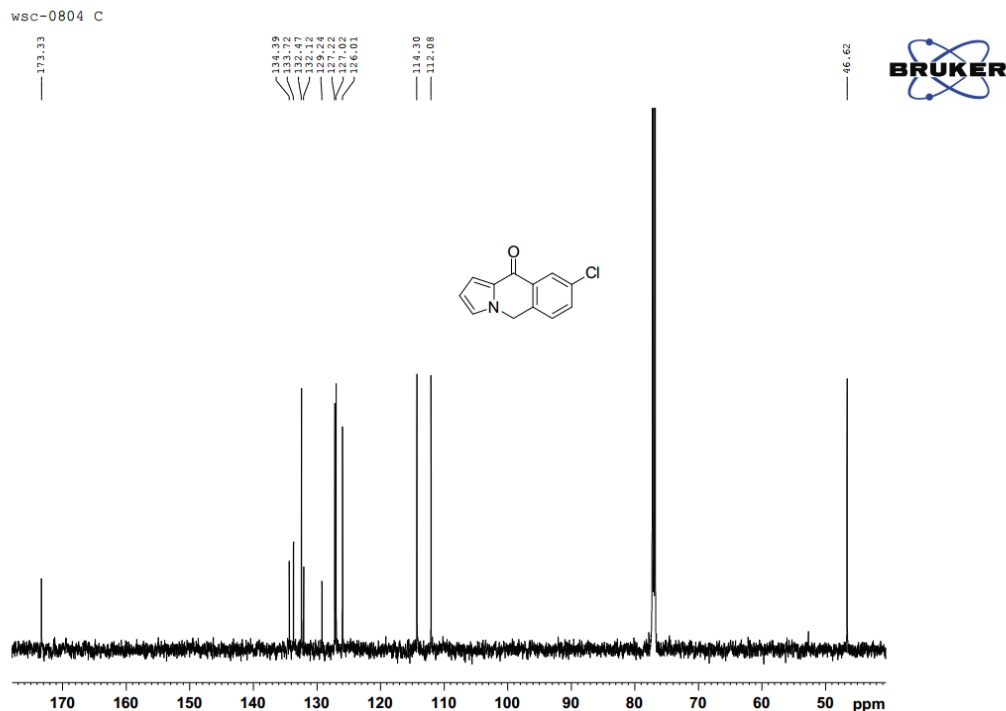


Current Data Parameters
NAME wsc-0804
DATE 1
PROCNO 1

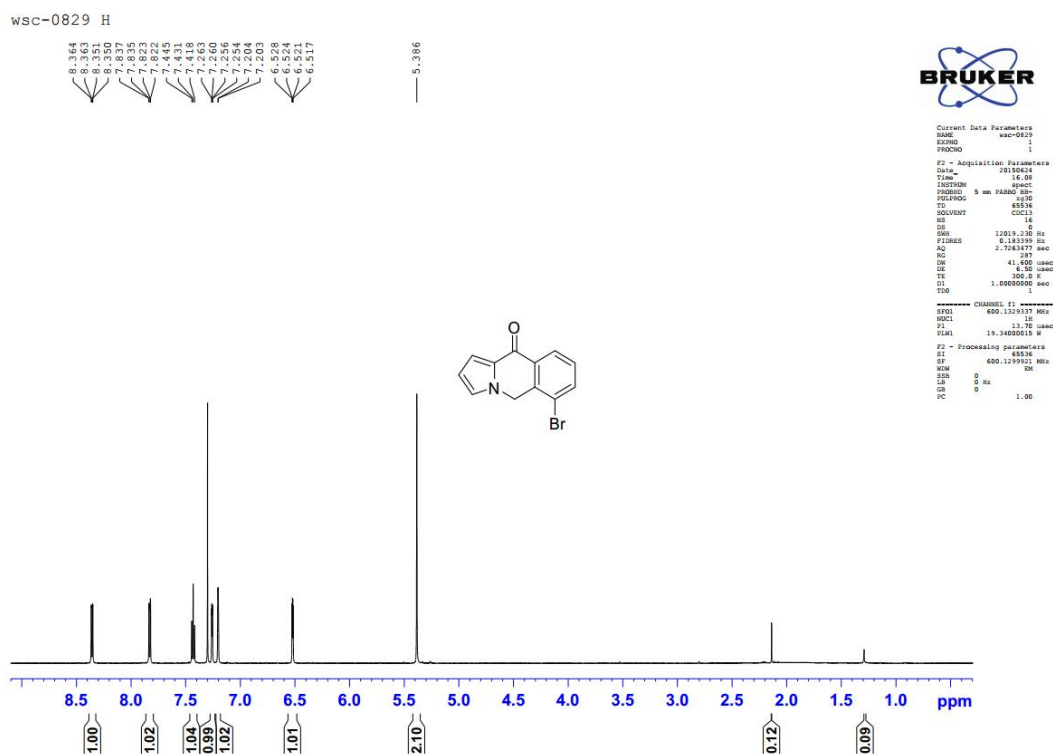
F2 - Acquisition Parameters
Date 20100529
Time 16.59
INSTRUM spect
PROBHD 5 mm HBBBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 92
DS 9
SWH 12019.230 Hz
FIDRES 0.213378 Hz
AQ 2.7243477 sec
RG 181
WM 41.400 umsec
DE 6.50 umsec
TE 296.3 K
D1 1.00000000 sec
D06 1

===== CHANNEL f1 =====
NUC1 13C
P1 19.34000013 sec
PL1 0.00

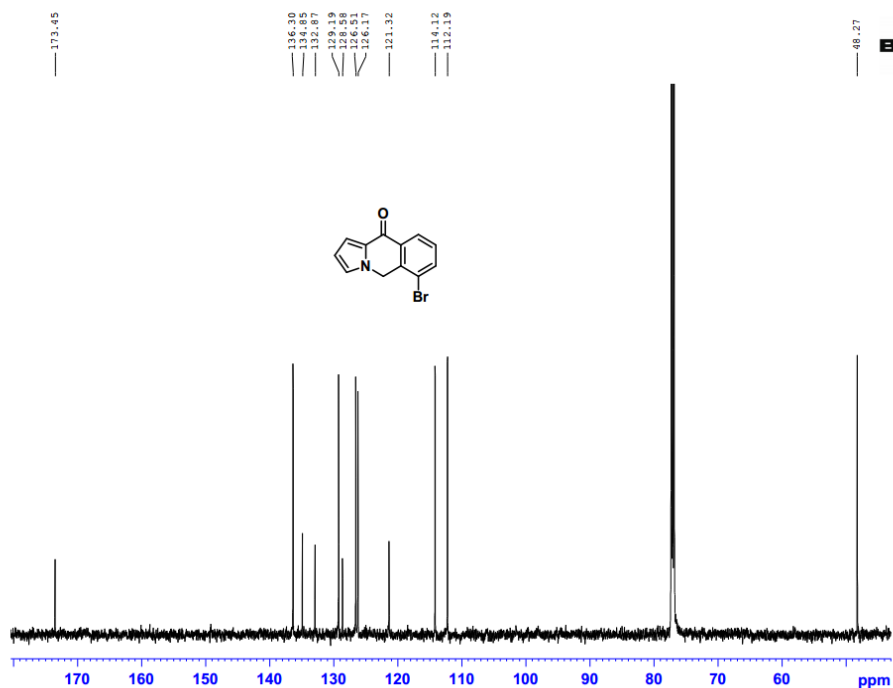
F2 - Processing parameters
SI 32768
SF 600.1259921 MHz
WDW EM
SSB 0
LB 0 Hz
GB 0
PC 1.00



NMR of compound 3c

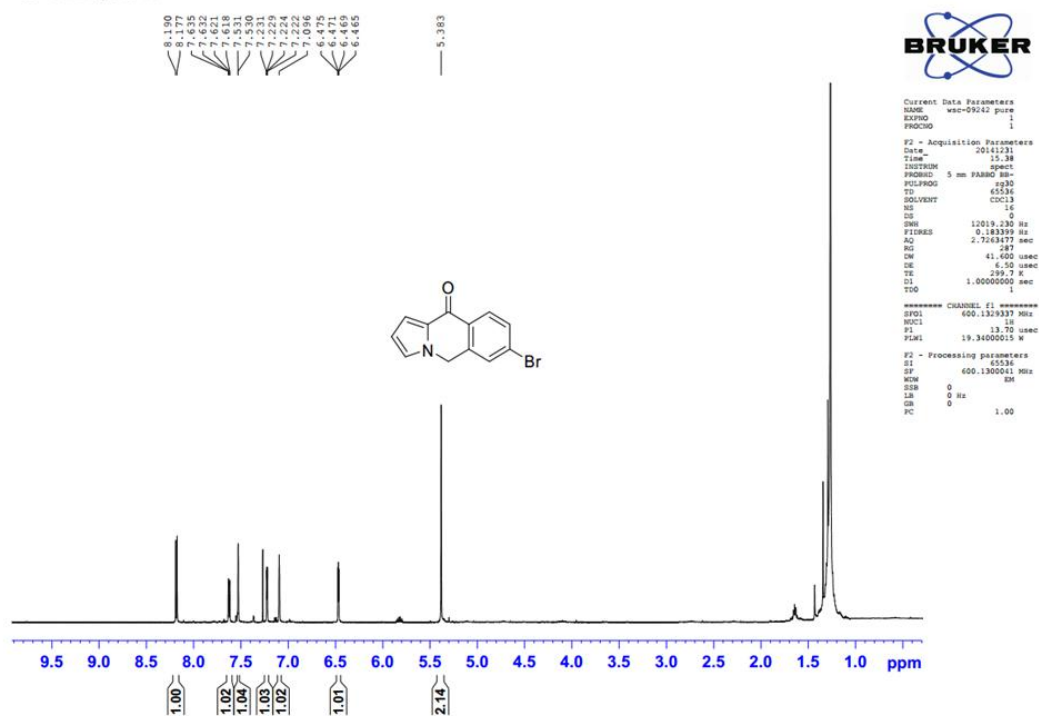


wsc-0829 C

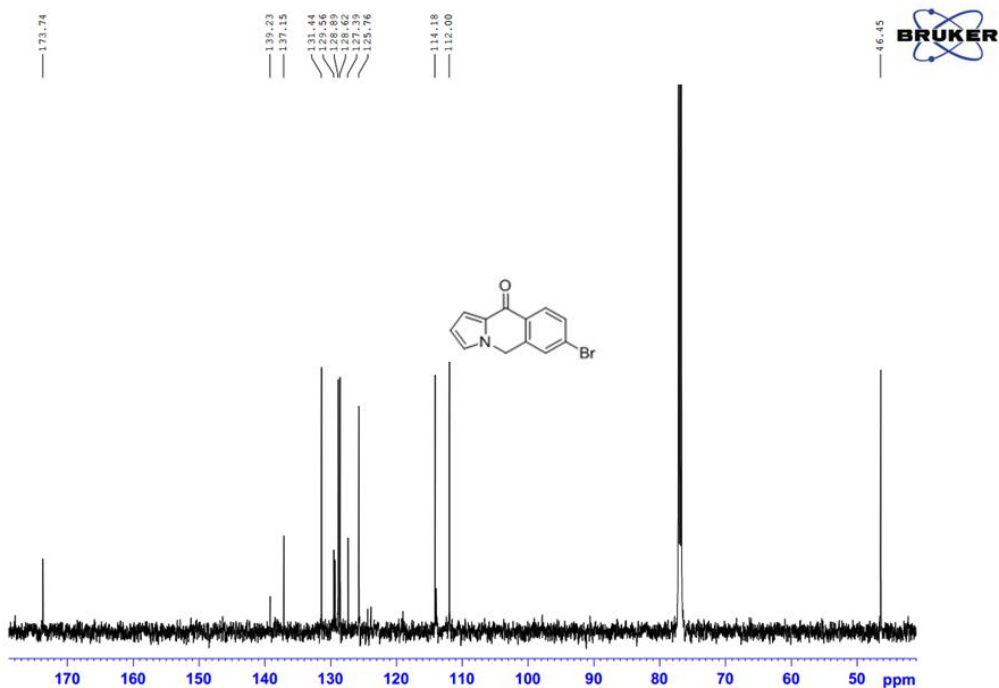


NMR of compound 3d

wsc-0924 pure H

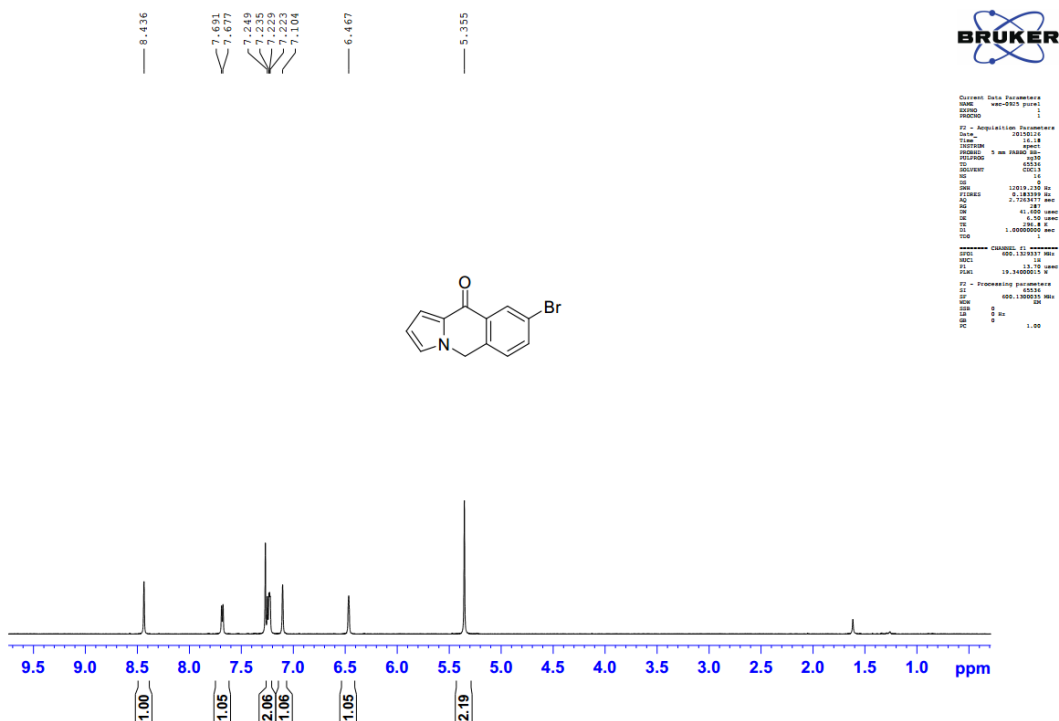


wsc-0924 pure H

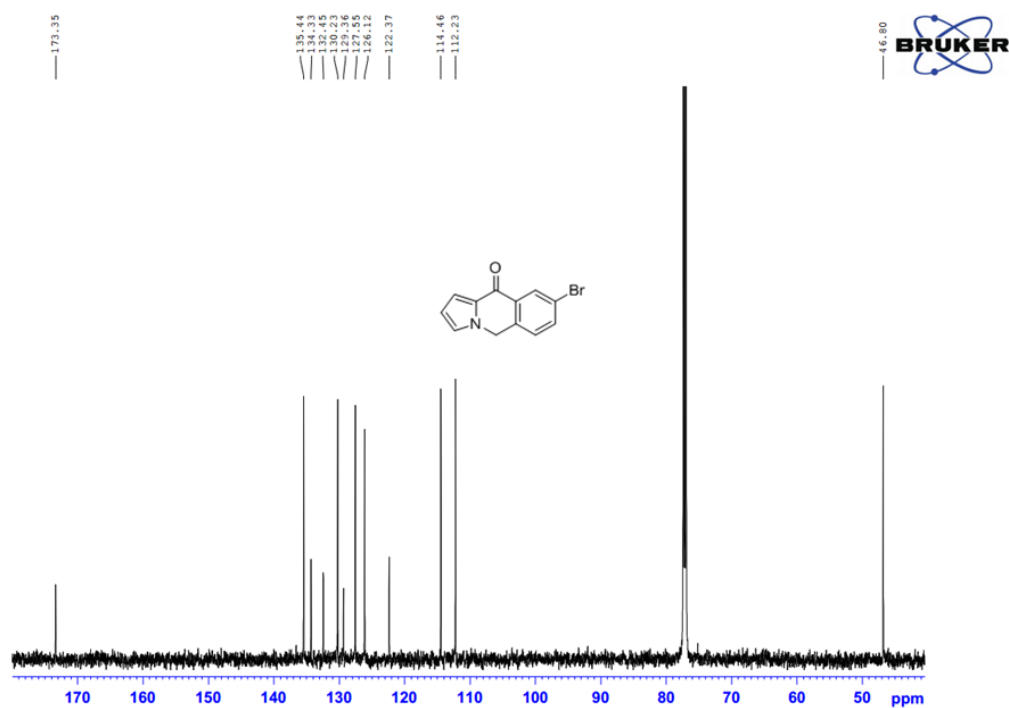


NMR of compound 3e

wsc-0925 pure1 H

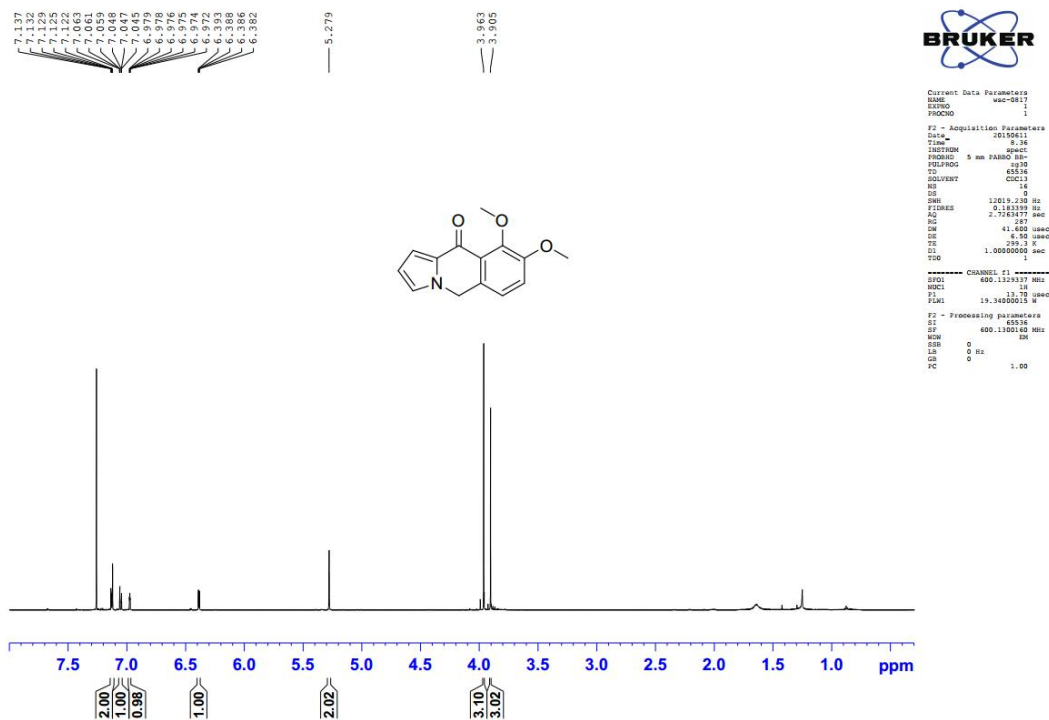


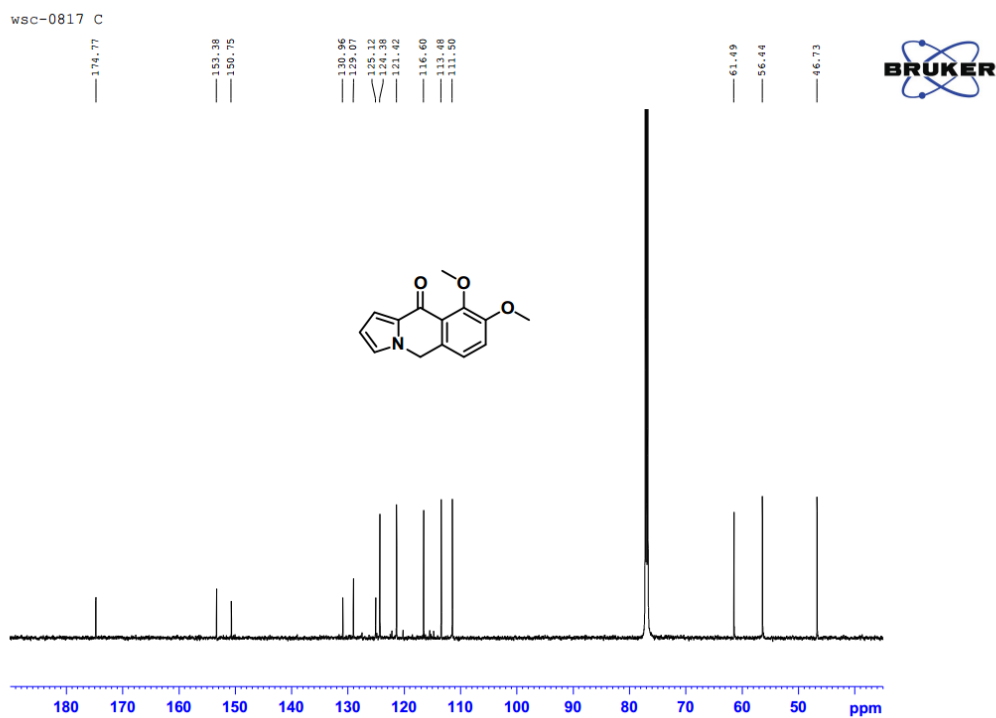
wsc-0925pure 1 C



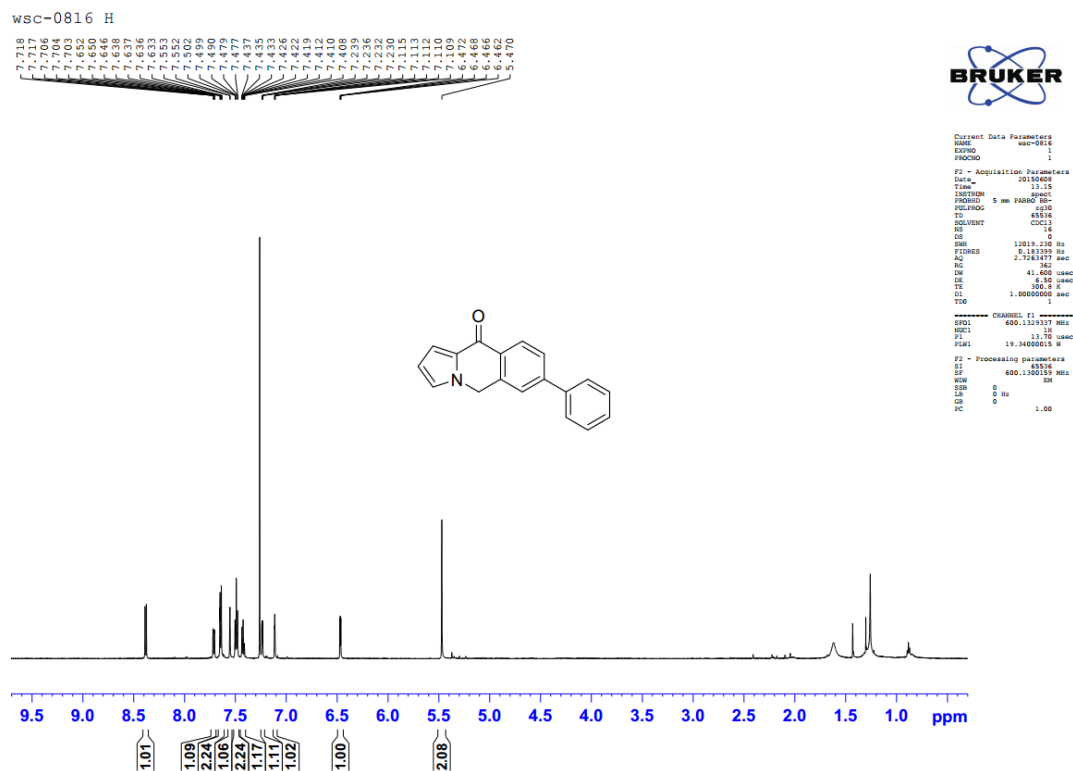
NMR of compound 3f

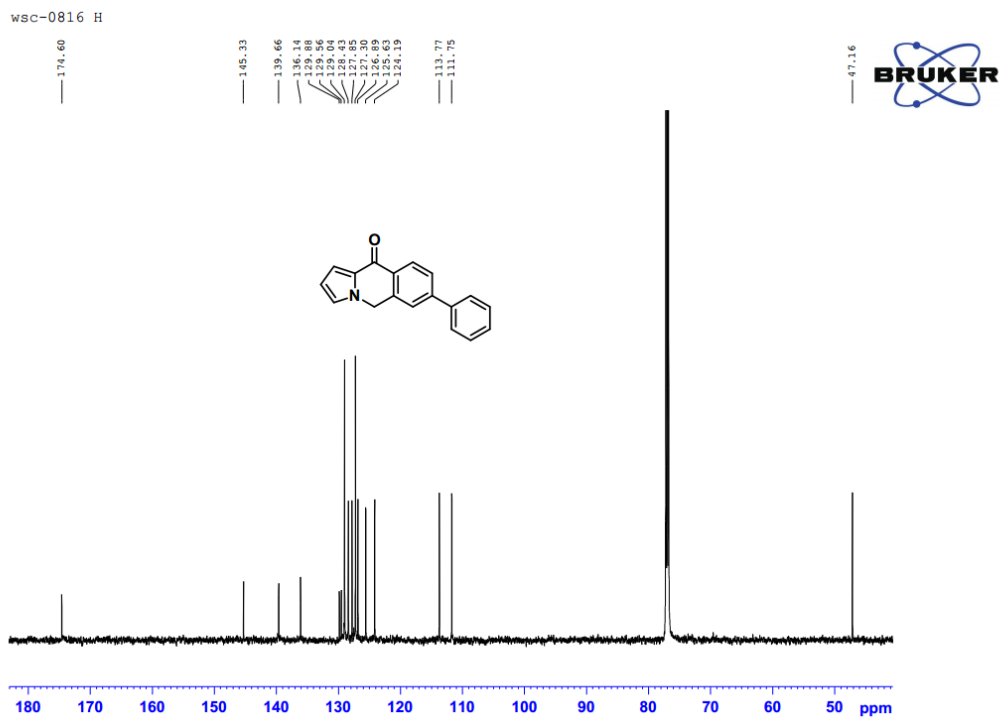
wsc-0817 H



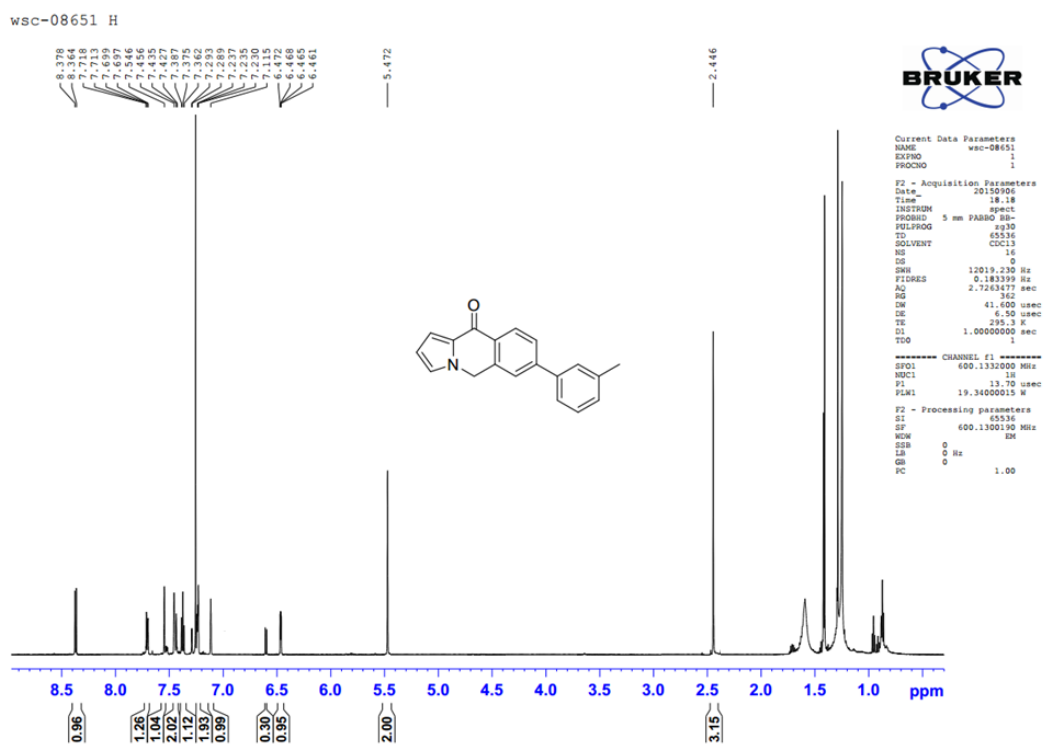


NMR of compound 3g

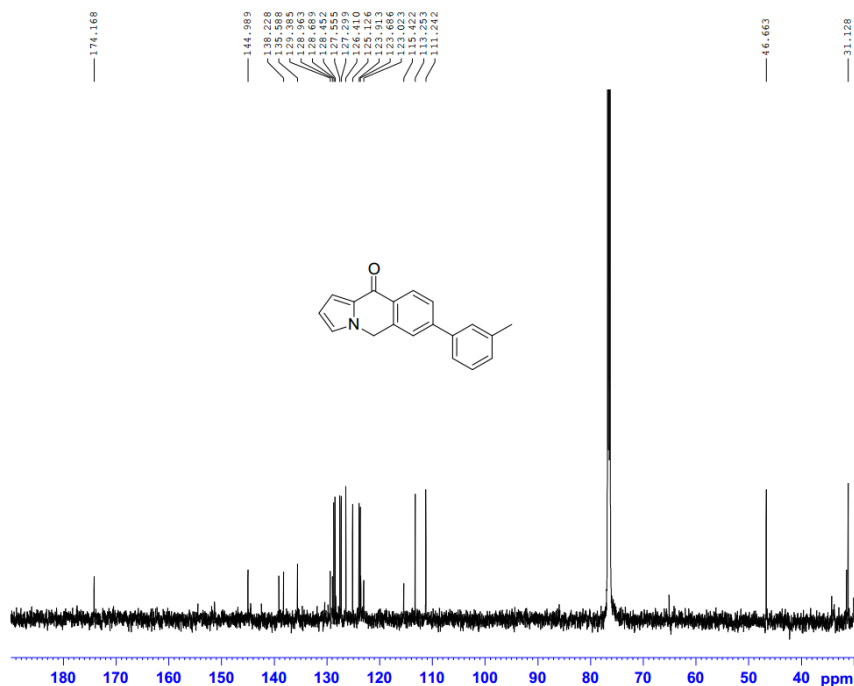




NMR of compound 3h

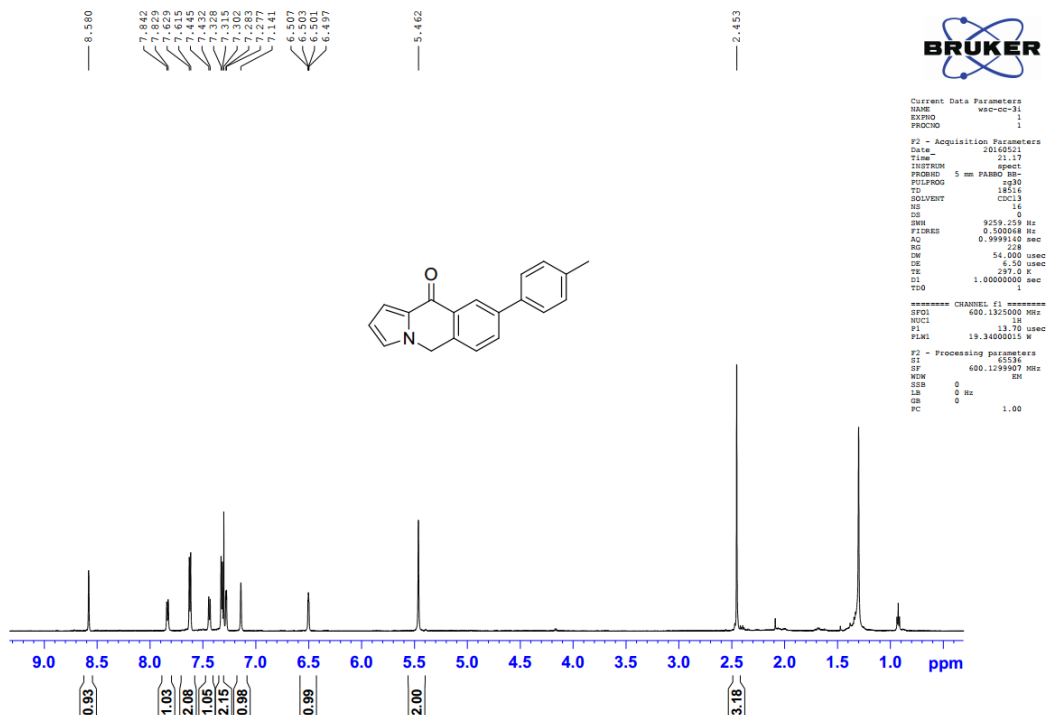


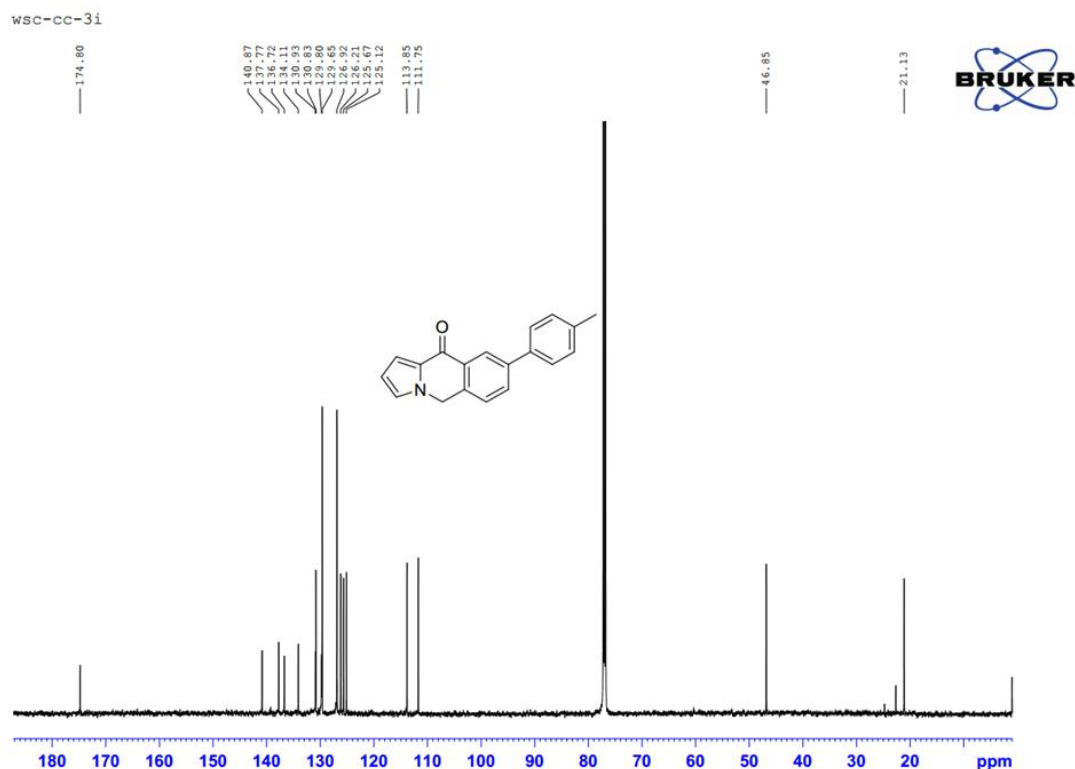
wsc-08651 C



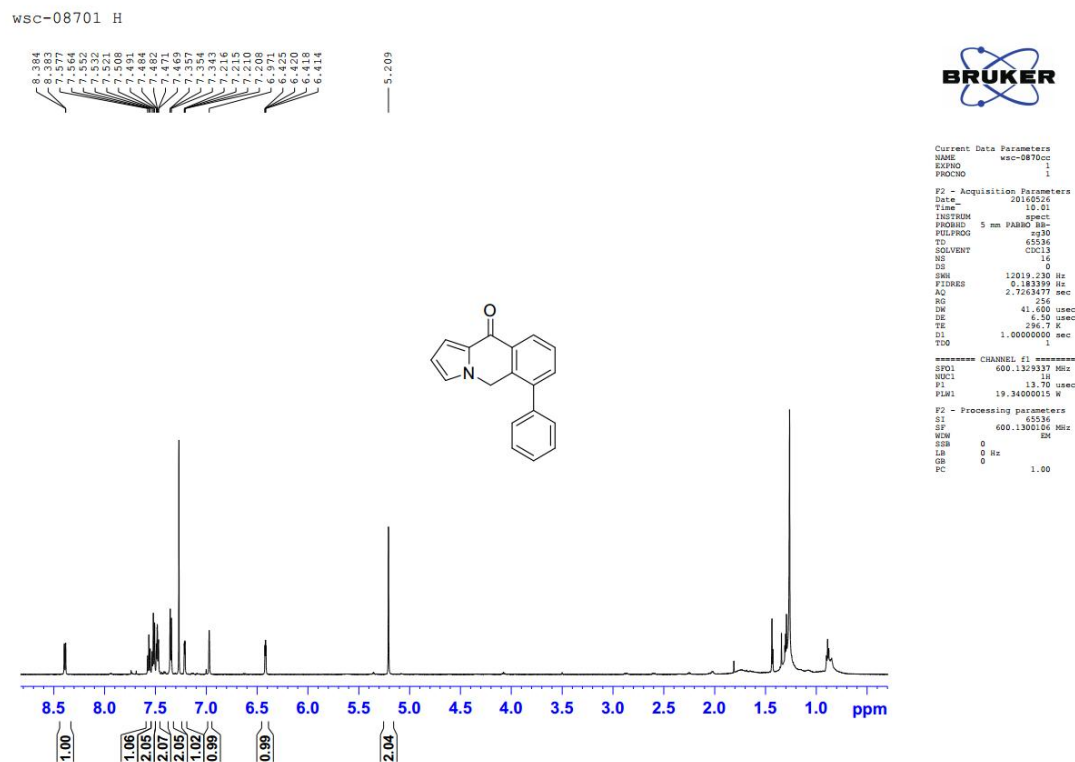
NMR of compound 3i

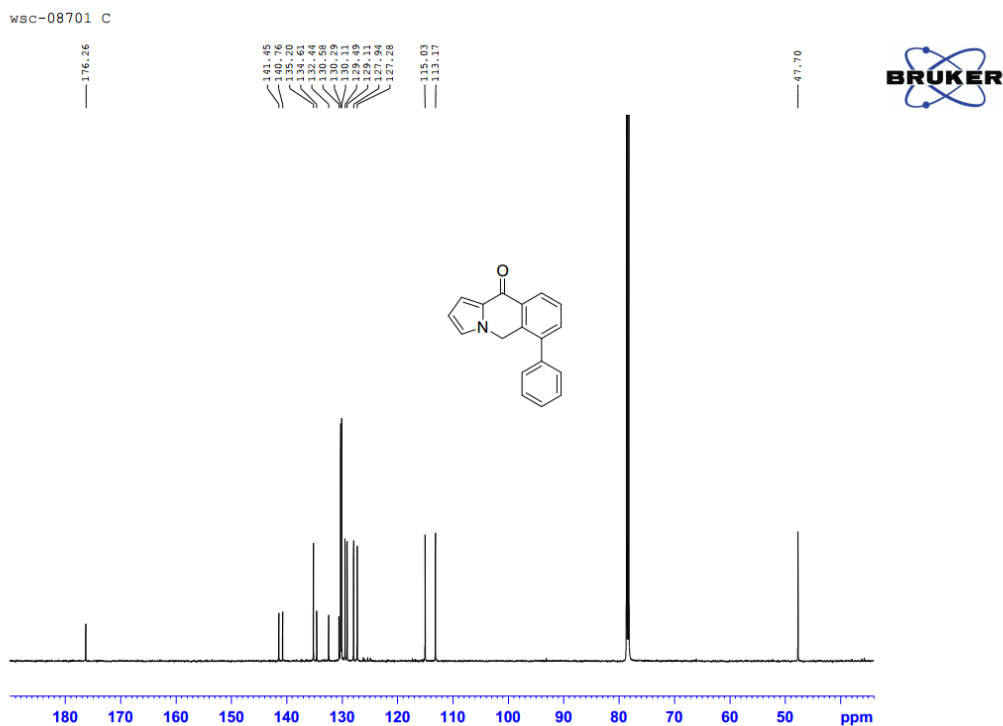
wsc-cc-3i H



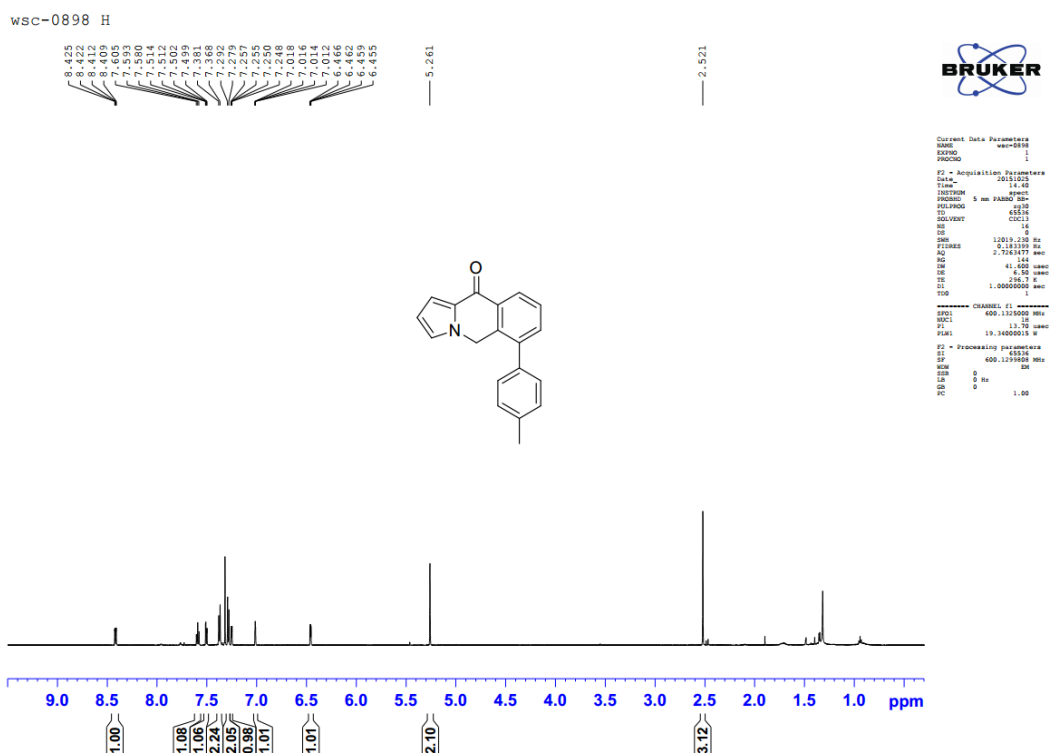


NMR of compound 3j

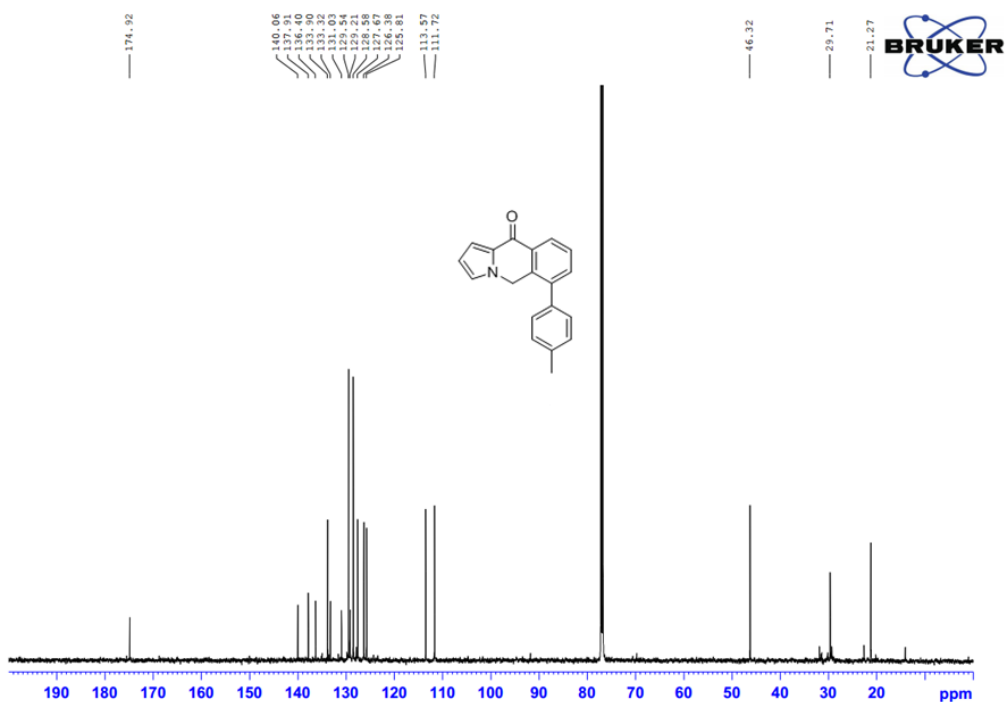




NMR of compound 3k

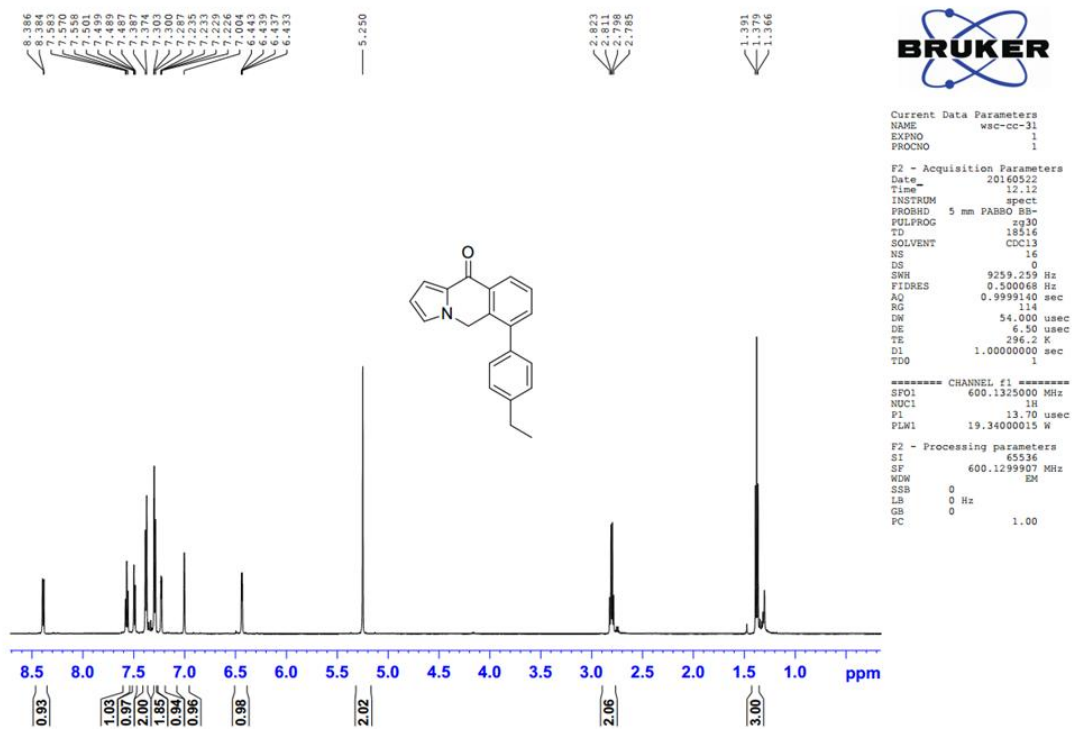


wsc-0898 C

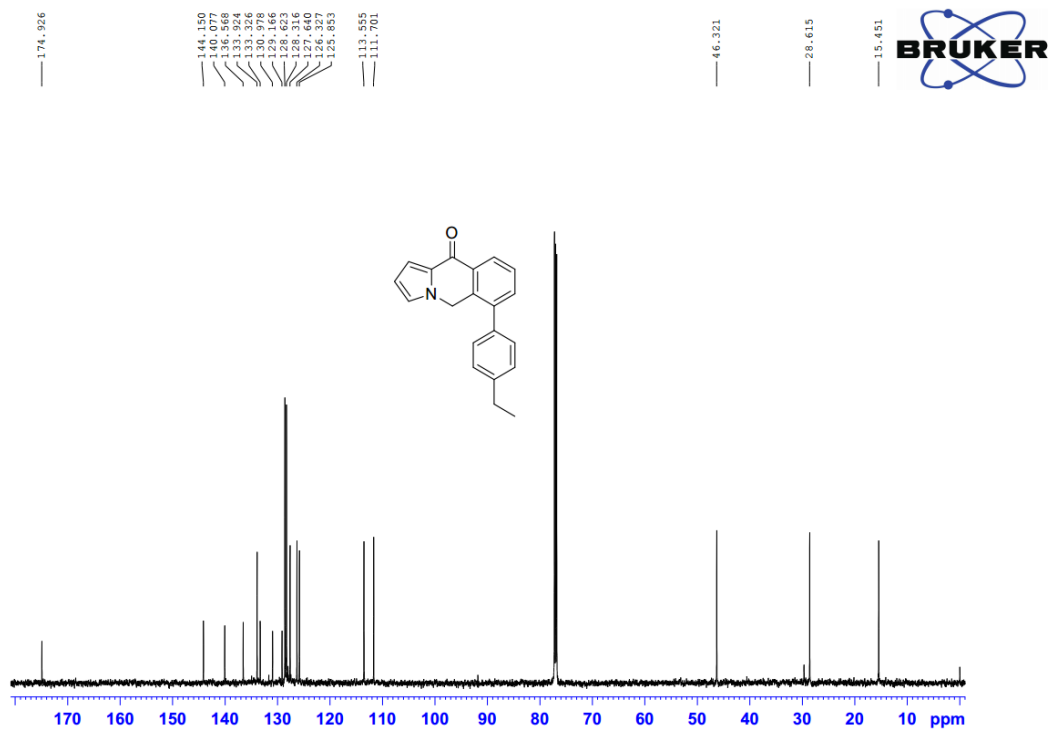


NMR of compound 31

wsc-cc-31

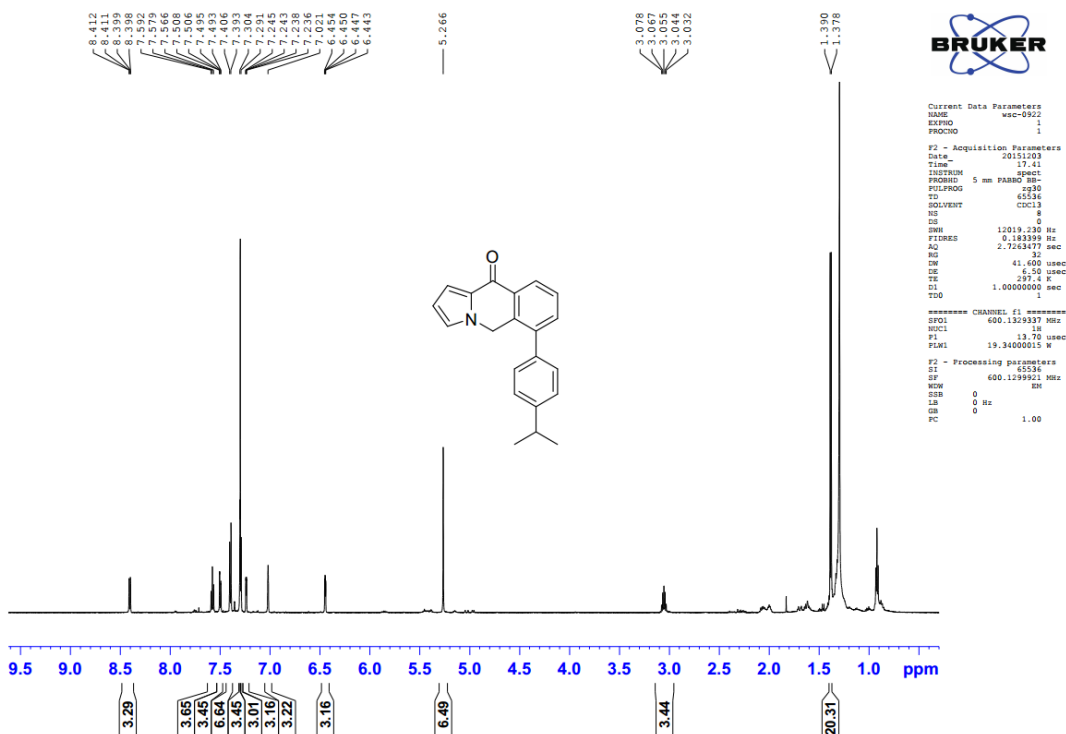


wsc-cc-31

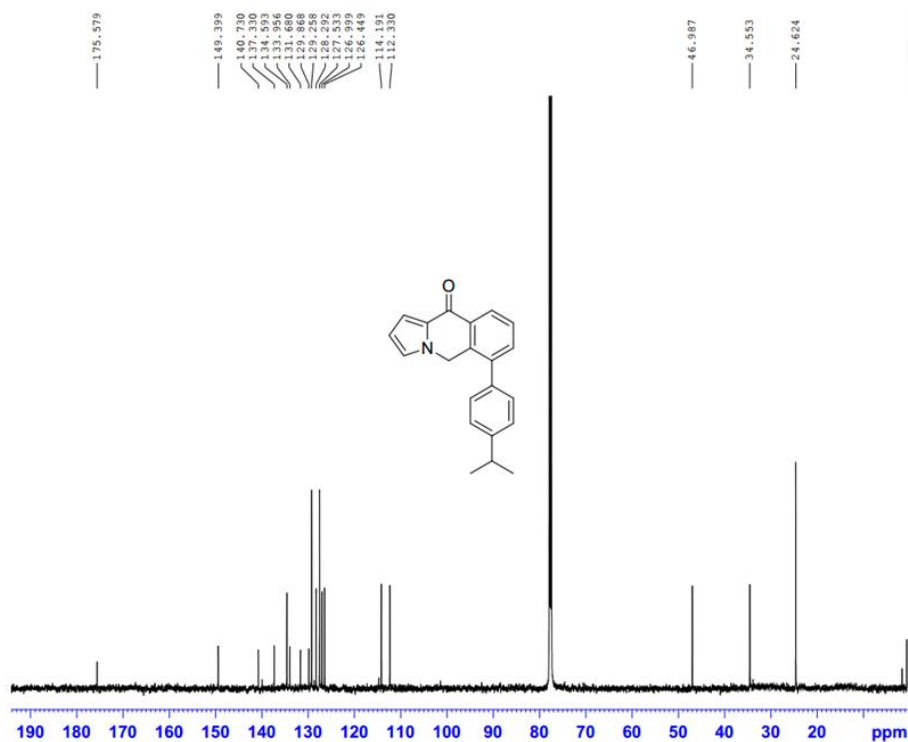


NMR of compound 3m

wsc-0922 H

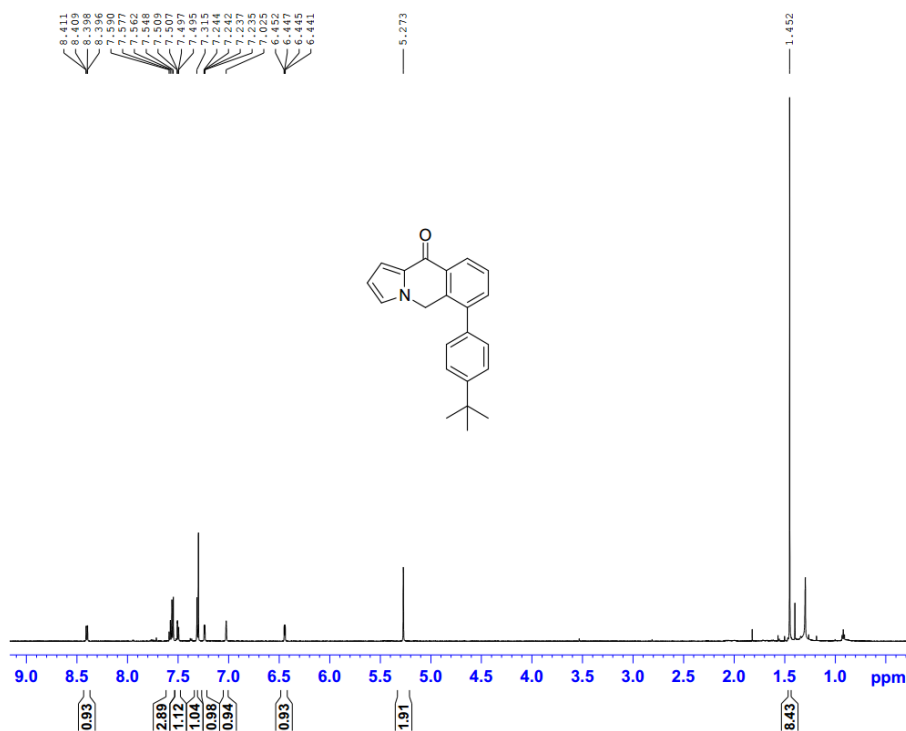


WSC-CC-3m C

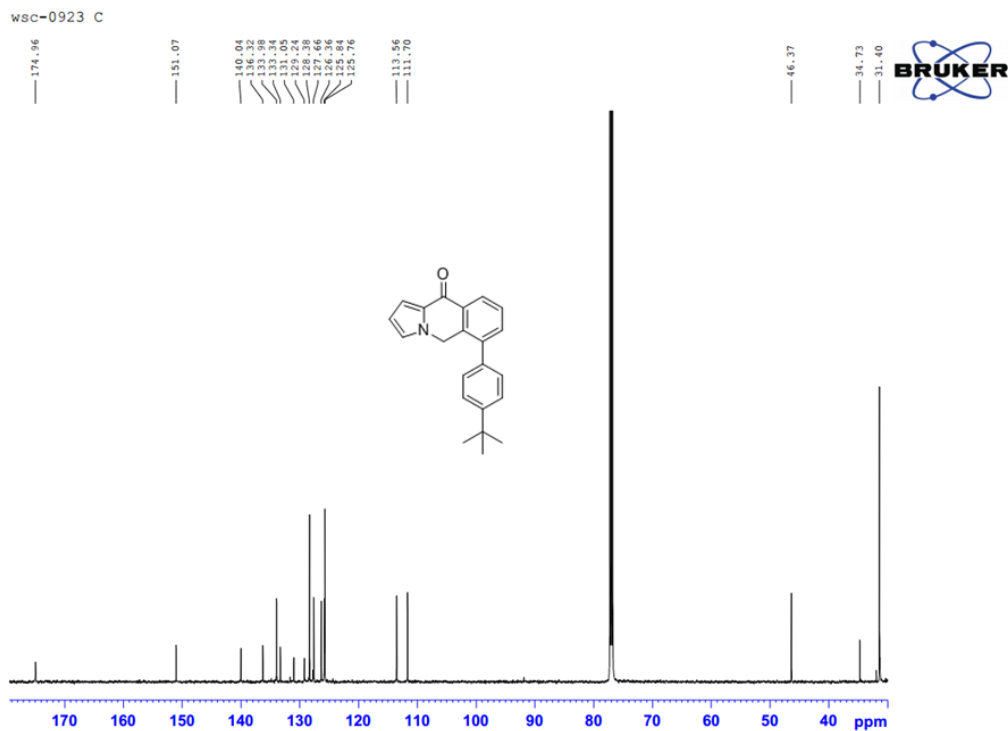


NMR of compound 3n

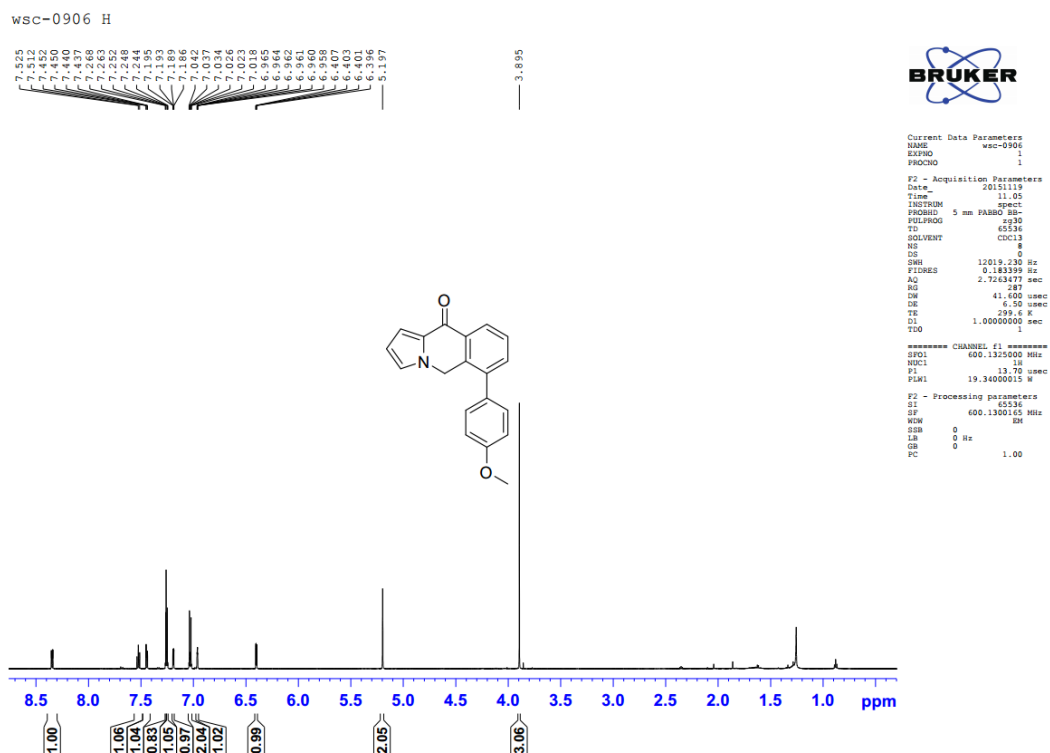
WSC-0923 H

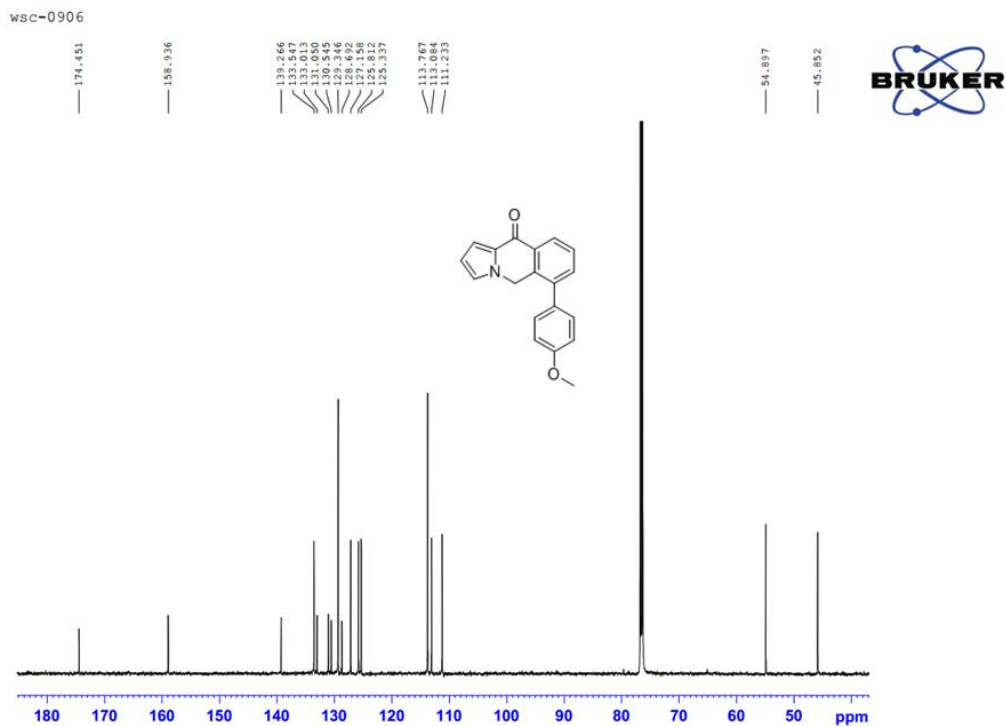


Current Data Parameters
NAME WSC-0923
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20151023
Time 17.45
INSTRUM spect
PROBHD 5 mm PABBO 2H
PULPROG zgpg30
PC 45.54
SOLVENT CDCl3
NS 8
DS 0
SWH 12019.230 Hz
FIDRES 0.181399 Hz
AQ 2.726177 sec
RG 32
RM 41.600 usec
DE 6.50 usec
TE 297.2 K
D1 1.00000000 sec
TD 1
===== CHANNEL f1 =====
NUC1 13C
P1 12.19 usec
PLM1 19.34000015 W
F2 - Processing parameters
SI 65536
SF 400.139921 MHz
WDW EM
SSB 0
LA 0 Hz
GB 0
PC 1.00

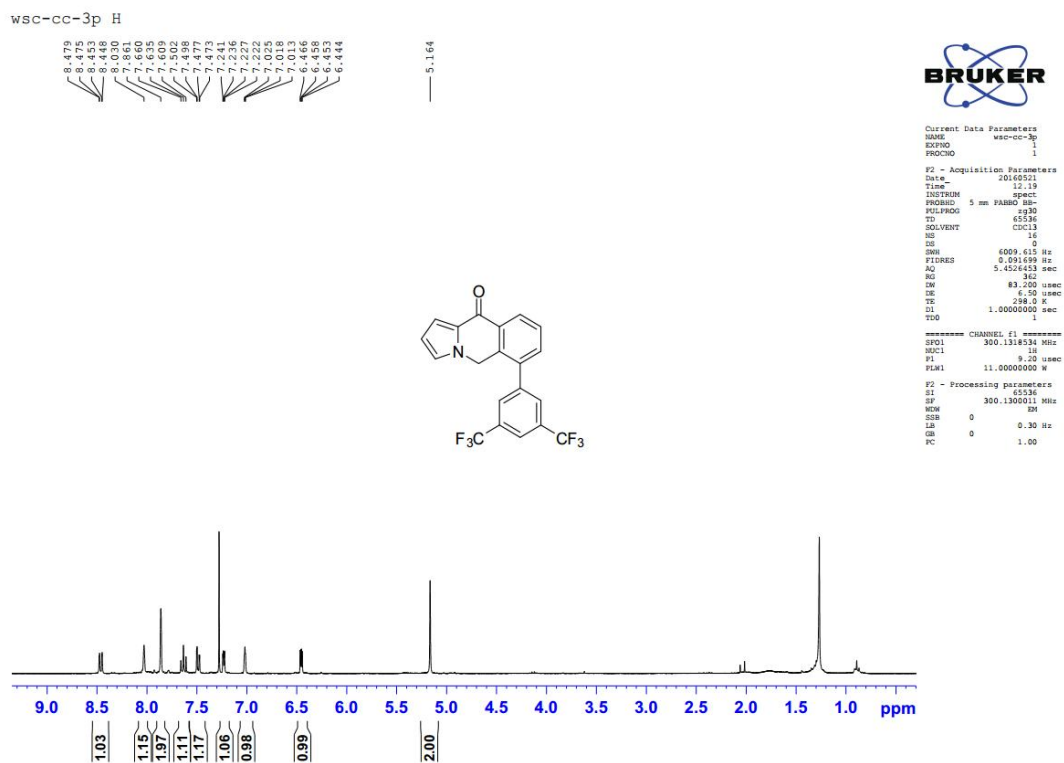


NMR of compound **3o**

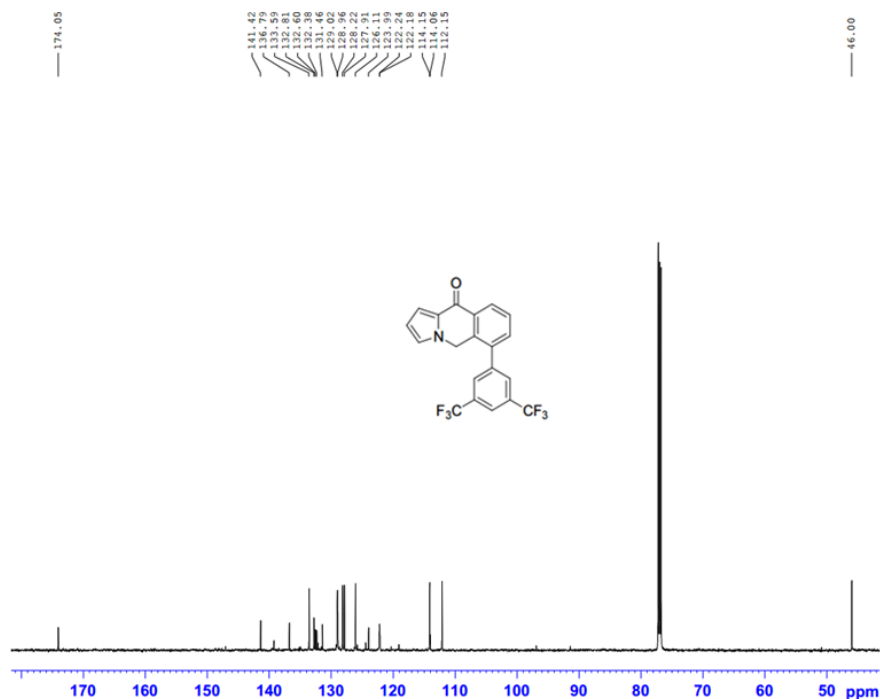




NMR of compound 3p

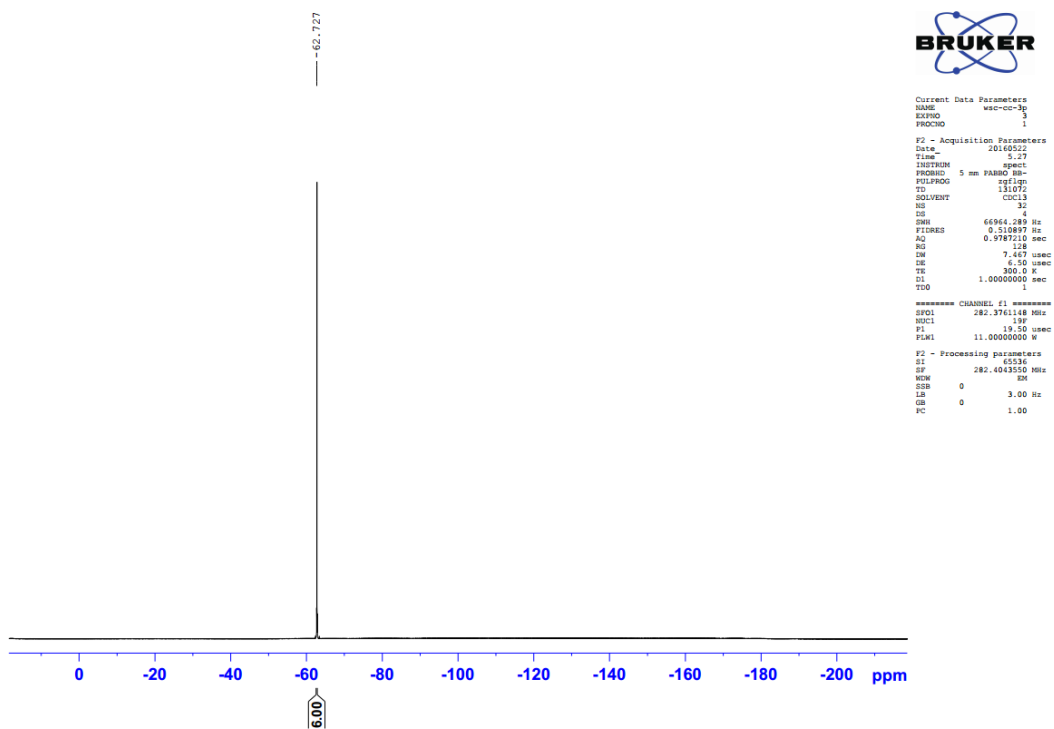


wsc-0899



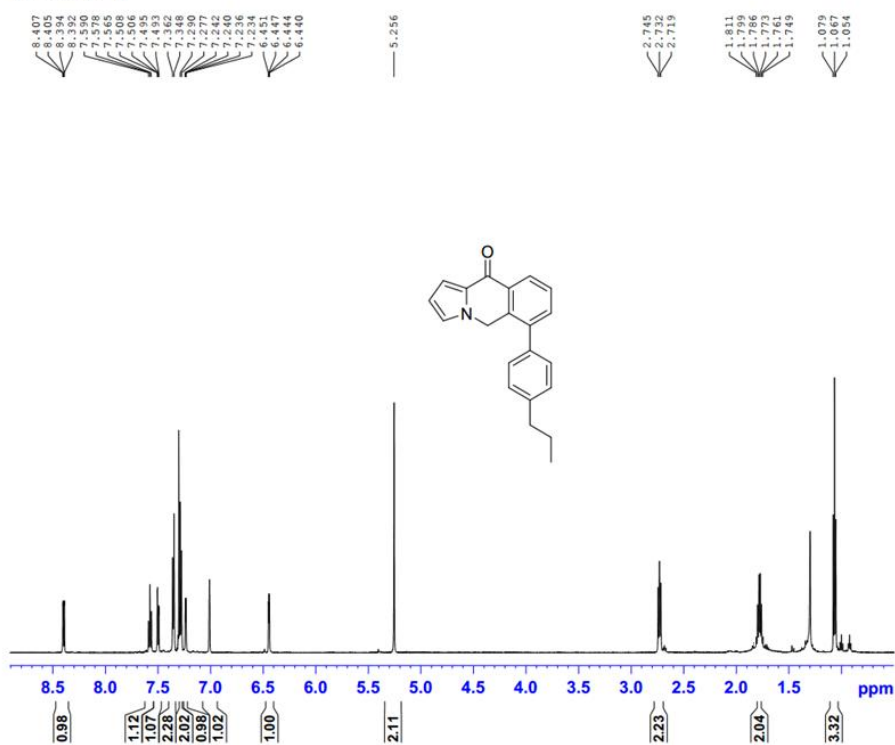
¹⁹F NMR of compound 3p

wsc-cc-3p F



NMR of compound 3q

wsc-cc-3q-H



BRUKER

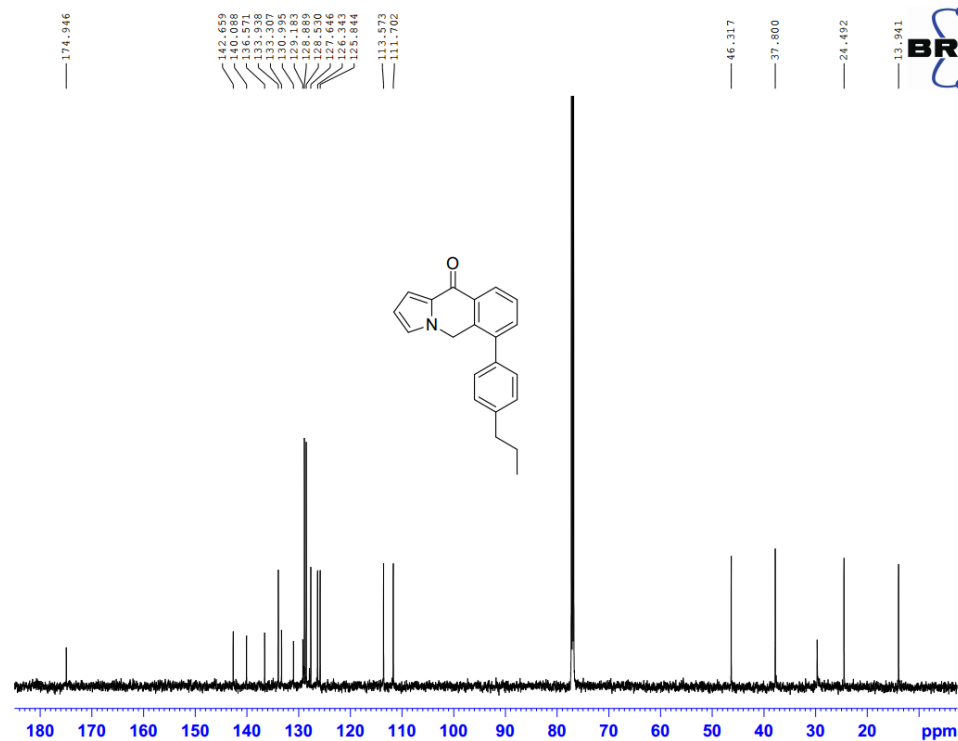
Current Data Parameters
NAME wsc-cc-3q
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160522
Time 12.24
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 18516
SOLVENT CDCl3
NS 16
DS 0
SWH 9259.259 Hz
FIDRES 0.500068 Hz
AQ 0.9999140 sec
RG 228
SW 54.000 usec
DE 6.50 usec
TE 296.6 K
D1 1.00000000 sec
TD0 1

----- CHANNEL f1 -----
SFO1 600.1325000 MHz
NUC1 1H
P1 13.70 usec
PLW1 19.34000015 W

F2 - Processing parameters
SI 65536
SF 600.1299907 MHz
WDW EM
SSB 0
LB 0 Hz
GB 0
PC 1.00

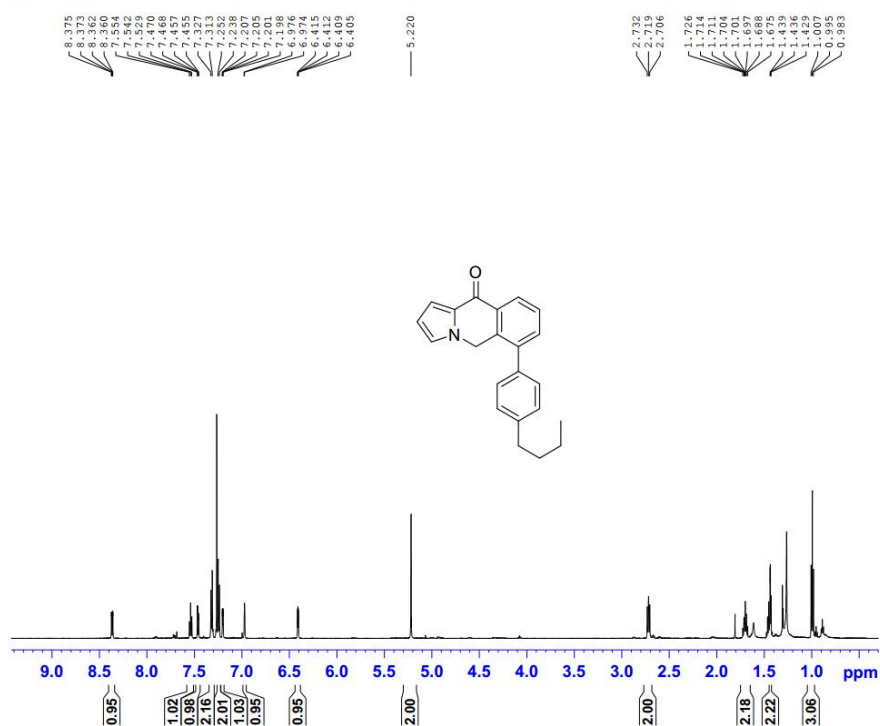
wsc-cc-3q-C



BRUKER

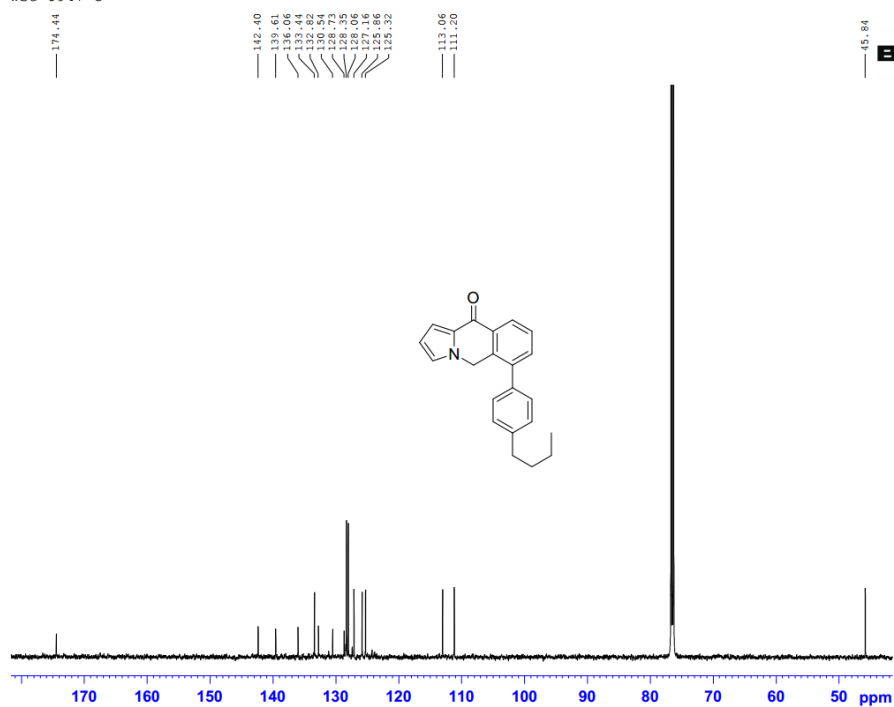
NMR of compound 3r

wsc-0947 H



Current Data Parameters
 NAME wsc-0947
 EXPNO 1
 PROCNO 1
 P2 - Acquisition Parameters
 Date_ 20160105
 Time 10.21
 INSTRUM spect
 PROBRD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
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 FIDRES 0.183399 Hz
 AQ 2.7263477 sec
 RG 144
 DW 41.400 usec
 DE 6.50 usec
 TE 300.2 K
 D1 1.00000000 sec
 T20 1
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 NUC1 1H
 P1 13.70 usec
 PLW1 19.34000015 W
 P2 - Processing parameters
 SI 65536
 SF 600.1300121 MHz
 WHW 8M
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

wsc-0947 C



NMR of compound 3s

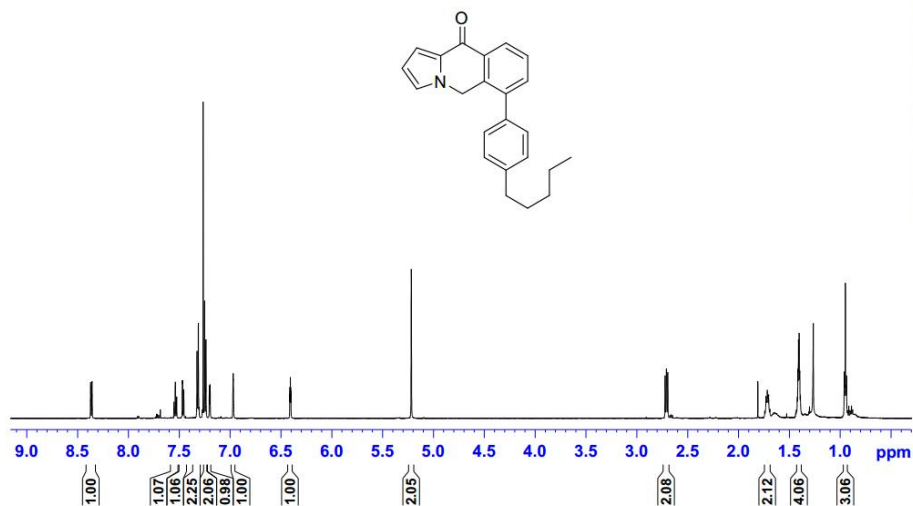
wsc-0949

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7.527
7.526
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7.457
7.320
7.313
7.306
7.249
7.242
7.236
7.205
7.200
7.198
7.196
6.972
6.971
6.969
6.968
6.415
6.413
6.411
6.408
6.406
6.404
5.219

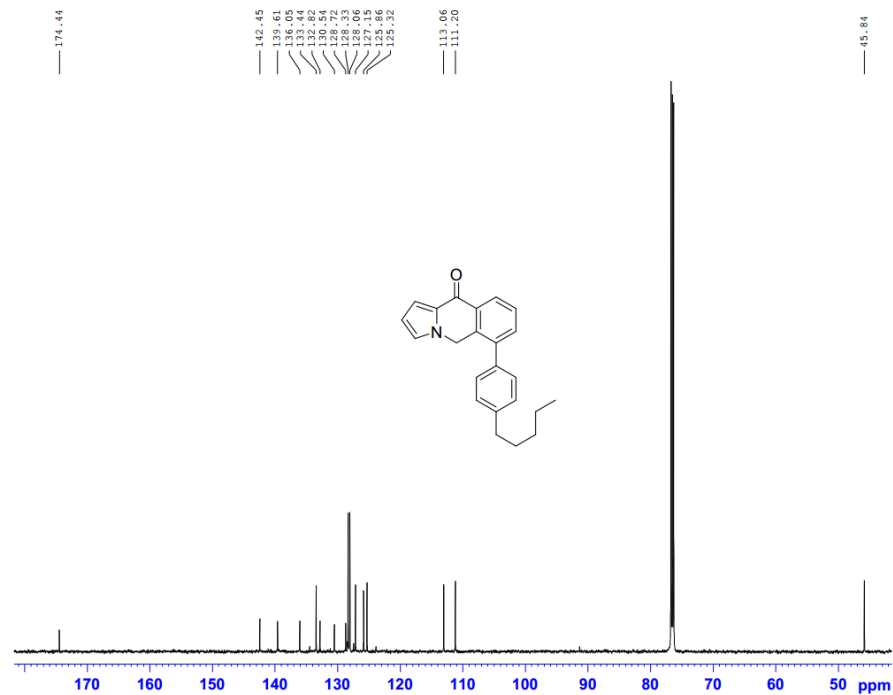
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2.696
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1.710
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1.695
1.692
1.437
1.432
1.422
1.416
1.410
1.398
1.392
1.383
0.961



Current Data Parameters
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EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
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Time_ 19.07
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PULPROG zgpg30
TD 65536
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DS 0
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FIDRES 0.183399 Hz
AQ 2.7263477 sec
RG 161
DM 41.600 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
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NUC1 1H
P1 13.70 usec
PLW1 19.34000015 W
F2 - Processing parameters
SI 65536
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SSB 0
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GB 0
PC 1.00



wsc-0949 C



NMR of compound **3t**

wsc-0950 H



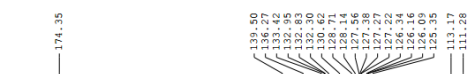
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SIMP: 1
PROCNO: 1

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NS: 16
DS: 4
SWH: 12019.230 Hz
FIDRES: 0.153399 Hz
AQ: 2.7283477 sec
RG: 281
SR: 41.400 usec
DE: 6.50 usec
TE: 300.2 K
D1: 1.00000000 sec
YD0: 1

***** CHANNEL f1 *****
NUC1: 13C
P1: 13.70 usec
PL1: 19.34000013 V

F2 - Processing parameters
SI: 65536
SF: 600.1300121 MHz
WDW: EM
SSB: 0
LB: 0 Hz
GB: 0
PC: 1.00

wsc-0950 C



Top1 and Top2 Inhibitory Activity Assay

Top1 Inhibitory Activity Assay

DNA relaxation assays were employed according to the procedure described in previous studies¹. The reaction mixture contained 35 mM Tris-HCl (pH 8.0), 72 mM KCl, 5 mM MgCl₂, 5 mM dithiothreitol, 5 mM spermidine, 0.1% bovine serum albumin (BSA), pBR322 plasmid DNA (0.25 µg), the indicated drug concentrations (1% DMSO), and 1 unit of Top1 (TaKaRa Biotechnology Co., Ltd., Dalian) in a final volume of 20 µL. Reaction mixtures were incubated for 15 min at 37 °C and stopped by addition of 2 µL of 10× loading buffer (0.9% sodium dodecyl sulfate (SDS), 0.05% bromophenol blue, and 50% glycerol). Electrophoresis was carried out in a 0.8% agarose gel in TAE (Tris-acetate-EDTA) at 8 V/cm for 1 h. Gels were stained with ethidium bromide (0.5 µg/mL) for 60 min. The DNA band was visualized over UV light and photographed with Gel Doc Ez imager (Bio-Rad Laboratories Ltd.)

Top2 Inhibitory Activity Assay

DNA Top2 α inhibitory activity of the compounds was measured using Topoisomerase II Drug Screening Kit (TopoGEN, Inc.). The reaction mixture contained 50 mM Tris-HCl (pH 8.0), 150 mM NaCl, 10 mM MgCl₂, 5 mM dithiothreitol, 30 µg/mL bovine serum albumin (BSA), 2 mM ATP, pBR322 plasmid DNA (0.25), the indicated drug concentrations (1% DMSO), and 0.75 unit of Top2 α (TopoGEN, Inc.) in a final volume of 20 µL. Reaction mixtures were incubated for 30 min at 37°C and stopped by addition of 2 µL of 10% SDS. After that, 2 µL of 10× gel loading buffer (0.25% bromophenol blue, 50% glycerol) was added. The reaction products were analyzed on 1% agarose gel at 8 V/cm for 1 h with TAE (Tris acetate-EDTA) as the running buffer. Gels were stained with ethidium bromide (0.5µg/mL) for 60 min. The DNA band was visualized over UV light and photographed with Gel Doc Ez imager (Bio-Rad Laboratories Ltd.).

Biological assays

Cell cultures

Hela cells and HepG2 cells were cultured in DMEM medium (Hyclone), HCT116 cells were cultured in McCoy's 5A medium (Gibco) and A549 cells were cultured in RPMI 1640 medium (Gibco) with 10% fetal calf serum (Gibco), 100 U/ml penicillin

and 100 µg/ml streptomycin (Invitrogen-Gibco, Karlsruhe, Germany). Cells were maintained in a humidified incubator with 5% CO₂ at 37 °C.

***In vitro* cytotoxicity assay**

5×10³ cells were seeded on a 96-well culture transwell apparatus (Costar, Cambridge, MA, USA) and cultured in DMEM medium, RPMI 1640 medium or McCoy's 5A medium with 10% (v/v) fetal bovine serum (FBS), respectively. The tested compounds were diluted to different final concentration and the cells were incubated for 48 h in a CO₂ incubator at 37 °C. Above loading media were removed, and the cells were fed new medium, followed by 10 µL of Cell Counting Kit-8 (CCK-8) per 100 µL of medium and incubated for 1 h. The cell viability was then assessed by Cell-Counting Kit-8 assay (Dojindo Laboratories, Japan) following the manufacturers' protocol. The absorbance of each well was monitored by a spectrophotometer (Tecan, Switzerland) at 450 nm.

References

1. G. Dong, C. Sheng, S. Wang, Z. Miao, J. Yao and W. Zhang, *J. Med. Chem.*, 2010, **53**, 7521.