Electronic Supplementary Information for:

Putting Chromium on the Map for N₂ Reduction: Production of Hydrazine and Ammonia. A Study of *cis*-M(N₂)₂ (M= Cr, Mo, W) Bis(diphosphine) Complexes

Jonathan D. Egbert,[‡] Molly O'Hagan,[‡] Eric S. Wiedner,[‡] R. Morris Bullock,[‡] Nicholas A. Piro,[†] W. Scott Kassel,[†] and Michael T. Mock[‡]*

[‡]Center for Molecular Electrocatalysis, Physical Sciences Division, Pacific Northwest National Laboratory, Richland, Washington 99352, United States

[†]Department of Chemistry, Villanova University, Villanova, Pennsylvania 19085, United States

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General Procedures

All synthetic procedures were performed under an atmosphere of N2 using standard Schlenk or glovebox techniques. Unless described otherwise, all reagents were purchased from commercial sources and were used as received. Protio solvents were dried by passage through activated alumina columns in an Innovative Technology, Inc., PureSolv solvent purification system and stored under N_2 or argon until use. THF- d_8 and benzene- d_6 were purchased from Cambridge Isotope Laboratories and dried over NaK and vacuum transferred before use. Isotopically labelled ¹⁵N₂ was purchased from Cambridge Isotope Laboratories. Magnesium powder, triflic acid, and 1,3,5-trimethoxybenzene were purchased from Sigma-Aldrich and used as received. CrCl₃(THF)₃ was purchased from Strem Chemicals Inc. MoBr₃(THF)₃,¹ WCl₄(PPh₃)₂,² and the diphosphine ligand PEtN2,6-F2-BnPEt,3 were prepared by following literature procedures. The ¹H NMR spectra were collected in thin walled NMR tubes on a Varian NMR S 300 or 500 MHz spectrometer. ¹H and ¹³C NMR chemical shifts are referenced to TMS using the residual protio solvent resonances in the deuterated solvent. ³¹P chemical shifts are proton decoupled unless otherwise noted and referenced to H₃PO₄ as an external reference. ¹⁵N NMR chemical shifts were externally referenced to CH_3NO_2 ($\delta = 0$). Infrared spectra were recorded on a Thermo Scientific Nicolet iS10 FT-IR spectrometer at ambient temperature and under a purge stream of nitrogen gas. Cyclic voltammetry was performed in a Vacuum Atmospheres Nexus II glovebox under an N₂ atmosphere using a CH Instruments model 620D or 660C potentiostat. Measurements were performed using standard three-electrode cell containing a 1 mm PEEKencased glassy carbon working electrode, Cypress Systems EE040, a 3 mm glassy carbon rod (Alfa) as the counter electrode, and a silver wire suspended in electrolyte solution and separated from the analyte solution by a Vycor frit (CH Instruments 112) as the pseudo-reference electrode

in THF with 0.20 M [Bu₄N][B(C₆F₅)₄] as the supporting electrolyte. Prior to the acquisition of each voltammogram, the working electrode was polished using 0.1 μ m γ -alumina (BAS CF-1050), and rinsed with THF. Ferrocene or decamethylferrocene was used as an internal reference, and all potentials are reported versus the ferrocenium/ferrocene couple at 0.0 V. Elemental analysis was performed by Atlantic Microlabs, Norcross, GA.

Synthesis of the dinitrogen complexes 1-3

 $cis-[Cr(N_2)_2(P^{Et}N^{2,6-F2-Bn}P^{Et})_2]$ (1): $P^{Et}N^{2,6-F2-Bn}P^{Et}$ (0.71 g, 2.0 mmol) was dissolved in ~2 mL of THF and added to a purple solution of $CrCl_3(THF)_3$ (0.38 g, 1.0 mmol) in ~ 50 mL THF that was cooled to -5 °C, resulting in a dark blue solution. After the reaction was stirred for 2 h at -5 °C excess magnesium powder (1.7 g, 70 mmol) was added to the reaction mixture. The reaction was stirred at -5 °C and was periodically monitored for 72 h by ³¹P NMR for the formation of 1. After 24 h, ³¹P NMR analysis showed the formation of 1 but a significant amount of free PNP ligand was also present. After stirring for 72 h the reaction mixture was filtered through a plug of alumina (2 in x $\frac{1}{2}$ in) and the volume of the clear orange solution was reduced to 3 mL under reduced pressure. Cold pentane (stored at -5 °C) (~10 mL) was added to precipitate an orange solid that was collected by filtration and washed with cold pentane to remove unreacted PEtN2,6-F2-BnPEt ligand. The yellow-orange solid was dried briefly under reduced pressure. Yield: 0.031 g (0.039 mmol, 4%). ¹H NMR (500 MHz, THF-d₈, 298 K): δ 7.33 (m, 2H, ArH), 6.97 (m, 4H, ArH), 3.62 (s, 4H, NCH₂Ar), 3.3 (m, 2H, PCH₂N), 3.2 (m, 2H, PCH₂N), 2.38 (m, 2H, PCH₂N), 2.31 (m, 4H, PCH₂CH₃), 2.20 (m, 2H, PCH₂N), 1.89 (m, 2H, PCH₂CH₃), 1.69 (m, 2H, PCH₂CH₃), 1.48 (m, 2H, PCH₂CH₃), 1.41 (m, 4H, PCH₂CH₃), 1.16 (m, 2H, PCH₂CH₃), 0.89-1.10 (m, 24H, PCH₂CH₃). ¹³C{¹H} NMR (125.7 MHz, THF- d_8 , 298 K): δ 163.2 (m, ArCF, ¹J_{CF} = 251 Hz), 130.7 (m, ArC, ${}^{1}J_{CF}$ = 167 Hz, ${}^{3}J_{CF}$ = 11 Hz), 114.8 (m, ArC), 111.9 (m, ArC, ${}^{1}J_{CF}$ =

165 Hz), 58 (m, NCH₂P), 54 (m, NCH₂Ar), 17-30 (m, PCH₂CH₃), 6-11 (m, PCH₂CH₃). ³¹P{¹H} NMR (202.4 MHz, THF-*d*₈, 298 K): δ 37.9 (dd, 2P, ²J_{PP} = 23.4, 27.9 Hz), 32.2 (dd, 2P, ²J_{PP} = 23.4, 27.9 Hz). ¹⁹F NMR (470.5 MHz, THF-*d*₈, 298 K): δ -116.7 (m, 4F). ¹⁵N{¹H} NMR (50.7 MHz, THF-*d*₈, 298 K): δ -7.3 (d, 2N, Cr-N≡*N*, ¹J_{NN} = 7 Hz), -11.5 (dm, 2N, Cr-*N*≡N, ¹J_{NN} = 7 Hz). IR (THF): ν_{NN} cm⁻¹ = 1990, 1911. Anal. Calcd. for C₃₄H₅₈CrF₄N₆P₄: C, 50.9; H, 7.3; N, 10.5. Found: C, 50.5; H, 7.3; N, 9.95. Complex 1 is remarkably stable in THF at 40 °C for up to three days with a small amount of free PNP ligand being detected in solution after this time. Heating slowly converts 1 to *trans*-1, and after 3 days ~20% is converted to the trans isomer.

trans-[Cr(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (*trans*-1): Following the synthesis of complex 1, the material removed in the pentane wash performed in the final purification step slowly precipitates a small quantity of *trans*-1, a yellow solid. This material was identified as *trans*-1 by NMR and IR spectroscopies, and X-ray diffraction analysis (see Fig. S1). ¹H NMR (500 MHz, THF-*d*₈, 298 K): δ 7.33 (m, 2H, Ar*H*), 6.97 (m, 4H, Ar*H*), 3.64 (s, 4H, NC*H*₂Ar), 2.82 (s, 8H, PC*H*₂N), 1.83 (m, 16H, PC*H*₂CH₃), 1.02 (m, 24H, PCH₂C*H*₃). ³¹P{¹H} NMR (202.4 MHz, THF-*d*₈, 298 K): δ 37.1 (s, 4P). ¹⁹F NMR (470.5 MHz, THF-*d*₈, 298 K): δ -116.5 (m, 4F). ¹⁵N{¹H} NMR (50.7 MHz, THF-*d*₈, 298 K): δ -22.6 (dm, 2N, Cr-*N*=N, ¹*J*_{NN} = 6 Hz), -28.0 (d, 2N, Cr-N=*N*, ¹*J*_{NN} = 6 Hz). IR (THF): *v*_{NN} cm⁻¹ = 1906.

cis-[Mo(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (2): P^{Et}N^{2,6-F2-Bn}P^{Et} (0.43 g 1.2 mmol) was dissolved in ~2 mL of THF and added to a solution of MoBr₃(THF)₃ (0.34 g, 0.62 mmol) in ~ 50 mL THF that was cooled to -5 °C, resulting in a blue solution. After the reaction mixture was stirred for 17 h at -5 °C and after this time magnesium powder (1.5 g, 62 mmol) was added to the reaction mixture.

The reaction was stirred at -5 °C and was periodically monitored by ³¹P NMR for the formation of 2. After stirring for 8 h, ³¹P NMR indicated that the free PNP ligand was consumed and 2 was the only observable product in solution. The yellow reaction mixture was filtered through a plug of alumina (2 in x $\frac{1}{2}$ in). The solvent was removed under reduced pressure affording a yellowbrown solid. The solid was washed with cold pentane (3 x 15 ml) and was dried briefly under vacuum affording a yellow powder. Yield: 0.22 g (0.26 mmol, 41%). ¹H NMR (500 MHz, THFd₈, 298 K): δ 7.32 (m, 2H, ArH), 6.96 (m, 4H, ArH), 3.62 (s, 4H, NCH₂Ar), 3.27 (m, 2H, PCH₂N), 3.18 (m, 2H, PCH₂N), 2.40 (m, 2H, PCH₂N), 2.27 (m, 2H, PCH₂N), 2.14 (m, 2H, PCH₂CH₃), 2.02 (m, 2H, PCH₂CH₃), 1.93 (m, 2H, PCH₂CH₃), 1.55 (m, 2H, PCH₂CH₃), 1.50 (m, 2H, PCH₂CH₃), 1.42 (m, 4H, PCH₂CH₃), 1.38 (m, 2H, PCH₂CH₃), 0.99 (m, 18H, PCH₂CH₃), 0.92 (m, 6H, PCH₂CH₃). ³¹P{¹H} NMR (202.4 MHz, THF- d_8 , 298 K): δ 16.1 (dd, 2P, ¹J_{PP} = 24.6 Hz), 12.8 (dd, 2P, ${}^{1}J_{PP}$ = 24.6 Hz). ${}^{19}F$ NMR (470.5 MHz, THF- d_8 , 298 K): δ -116.5 (m, 4F). ¹⁵N{¹H} NMR (50.7 MHz, THF-*d*₈, 298 K): δ -23.6 (d, 2N, Mo-N≡*N*, ¹*J*_{NN} = 6 Hz), -39.1 (m, 2N, Mo- $N \equiv N$). IR (THF): ν_{NN} cm⁻¹ = 2012, 1950. Anal. Calcd. for C₃₄H₅₈MoF₄N₆P₄: C, 48.2; H, 6.9; N, 9.9. Found: C, 48.8; H, 7.1; N, 8.75. Complex 2 has a short lifetime in THF at 40 °C and readily isomerizes to the trans isomer upon heating over several hours.

cis-[W(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (**3**): WCl₄(PPh₃)₂ (0.37 g, 0.43 mmol) was added to a stirring THF (50 mL) solution of P^{Et}N^{2,6-F2-Bn}P^{Et} (0.30 g, 0.86 mmol). Magnesium (1g, 41 mmol) was added and the reaction was stirred for 24 h turning orange. The reaction was filtered through a plug of alumina (2 in x $\frac{1}{2}$ in). The solution was reduced to 7 mL and 50 mL of pentane was added to precipitate a beige powder. The powder was filtered off. The remaining orange-yellow solution was reduced to 3 mL under vacuum and layered with pentane and placed in a -35 °C freezer. A

yellow solid precipitated over 16 h that was collected and washed with pentane. The solid was dried briefly under vacuum affording a yellow powder. Yield: 0.25 g (0.27 mmol, 62%). ¹H NMR (500 MHz, THF-d₈, 298 K): 8 7.32 (m, 2H, ArH), 6.97 (m, 4H, ArH), 3.60 (s, 4H, NCH₂Ar), 3.37 (m, 2H, PCH₂N), 3.31 (m, 2H, PCH₂N), 2.41 (m, 2H, PCH₂N), 2.29 (m, 4H, PCH₂N), 2.24 (m, 2H, PCH₂CH₃), 2.15 (m, 2H, PCH₂CH₃), 2.05 (m, 2H, PCH₂CH₃), 1.64 (m, 4H, PCH₂CH₃), 1.50 (m, 4H, PCH₂CH₃), 1.39 (m, 2H, PCH₂CH₃), 0.97 (m, 18H, PCH₂CH₃), 0.90 (m, 6H, PCH₂CH₃). ¹³C{¹H} NMR (125.7 MHz, THF- d_8 , 298 K): δ 163.3 (m, ArCF, ¹J_{CF} = 247 Hz), 130.6 (m, ArC, ${}^{1}J_{CF} = 165$ Hz, ${}^{3}J_{CF} = 11$ Hz), 114.9 (s, ArC), 112.0 (m, ArC, ${}^{1}J_{CF} = 166$ Hz), 59.4 (m, NCH₂P), 55.4 (m, NCH₂Ar), 29.9 (m, PCH₂CH₃), 18.2-23.6 (m, PCH₂CH₃) 6.2-11.6 (m, PCH₂CH₃). ³¹P{¹H} NMR (202.4 MHz, THF- d_8 , 298 K): δ -12.1 (dd, 2P, ¹J_{PP} = 14.9 Hz, ${}^{1}J_{183_{WP}} = 297$ Hz), -14.5 (dd, 2P, ${}^{1}J_{PP} = 14.9$ Hz, ${}^{1}J_{183_{WP}} = 303$ Hz). ${}^{19}F$ NMR (470.5 MHz, THF-*d*₈, 298 K): δ-116.4 (m, 4F). ¹⁵N{¹H} NMR (50.7 MHz, THF-*d*₈, 298 K): δ-24.9 (d, 2N, W-N=N, ${}^{1}J_{NN} = 6$ Hz), -60.4 (m, 2N, W-N=N). IR (THF): v_{NN} cm⁻¹ = 1987, 1925. Anal. Calcd. for C₃₄H₅₈WF₄N₆P₄: C, 43.7; H, 6.3; N, 9.0. Found: C, 43.9; H, 6.3; N, 8.8. Complex **3** is remarkably stable in THF at 40 °C, and after 5 days only ~30% of 3 has isomerized to the trans isomer, a singlet at -16.5 ppm with ¹⁸³W sidebands (${}^{1}J_{183}_{WP}$ = 301 Hz) in the ³¹P NMR spectrum.

Protonolysis Procedure for 1-3

In a typical experiment, a known amount of complex 1-3, $(cis-[M(N_2)_2(P^{Et}N^{2,6-F-Bn}P^{Et})_2])$ (ca. 3-12 mmol) and 1,3,5-trimethoxybenzene was weighed into a rubber septum capped NMR tube and dissolved in THF-*d*₈. The solution was cooled to -40 °C in the NMR probe. Once the solution had temperature equilibrated for several minutes the tube was ejected and excess triflic acid (50-100 eq.) was added by microliter syringe to the cooled sample. The contents of the reaction were mixed by inversion of the NMR tube, and the sample was quickly returned to the

cold NMR probe. The sample was allowed to equilibrate to the temperature of the -40 °C NMR probe before initiating data collection. Room temperatures experiments were performed using an identical procedure with an NMR probe maintained at 25 °C.

Quantification of N₂H₅⁺ and NH₄⁺ by ¹H NMR spectroscopy.

To accurately quantify $N_2H_5^+$ (broad singlet at 10.8 ppm) and NH_4^+ (1:1:1 triplet, 7.0 ppm, $J_{\rm NH}$ = 51 Hz) formed in the protonolysis studies with HOTf, these product resonances were integrated versus 1,3,5-trimethoxybenzene resonances at 6.03 ppm. ¹H NMR spectra were collected using a 5 or 10 s delay time between scans to ensure complete relaxation of the sample for accurate integration. Manual peak integration of the ¹H NMR spectra was carried out after performing a full spectrum baseline correction. Due to the overlap of NH₄⁺ resonances with aromatic resonances of the PNP ligand, the NH₄⁺ resonances and the N₂H₅⁺ resonance were also analyzed by peak shape analysis to compare to the values obtained my manual peak integration. Peak fitting of the ¹H NMR resonances for N₂H₅⁺ and NH₄⁺ was preformed using the CRAFT (Complete Reduction to Amplitude Frequency Table) analysis software package included in the VNMRJ 4.2 software suite.⁴ Processing parameters include 0.2 Hz line broadening with maximum line widths defined in CRAFT as 35 Hz. Signal amplitudes were determined relative to the internal standard of the aromatic protons of 1,3,5-trimethoxybenzene at 6.03 ppm. Tabulated results for protonolysis studies of complexes 1, 2, and 3 are listed in Table S3. CRAFT simulated ¹H NMR data are shown in Fig. S7.



Fig. S1. Molecular structure of *trans*-[$Cr(N_2)_2(P^{Et}N^{2,6-F2-Bn}P^{Et})_2$] (*trans*-1). Thermal ellipsoids are drawn at 50% probability. Hydrogen atoms are omitted for clarity. Selected bond distances (Å): (1) Cr-P1 = 2.3546(4); Cr-P2 = 2.3535(4); Cr-N1 = 1.8858(14); N1-N2 = 1.1274(19).



Fig. S2. FTIR spectra collected in THF showing the ${}^{14}N_2$ bands of *trans*-[Cr(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (*trans*-1; green), *cis*-[Cr(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (1; black), *cis*-[Mo(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (2; blue), *cis*-[W(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (3; red).



Fig. S3. ³¹P NMR spectra collected in THF at 25 °C showing the isomerization of *trans*- $[Cr(N_2)_2(P^{Et}N^{2,6-F2-Bn}P^{Et})_2]$ (*trans*-1) to *cis*- $[Cr(N_2)_2(P^{Et}N^{2,6-F2-Bn}P^{Et})_2]$ (1). Trace 1: Initial ³¹P spectrum; Trace 2: after 1 h; Trace 3: after 2.5 h; Trace 4: after 4.5 h; Trace 5: after 21.5 h. The resonance at 33.6 ppm is free P^{Et}N^{2,6-F2-Bn}P^{Et} ligand.



Fig. S4. FTIR spectra collected in THF at room temperature showing the isomerization of *trans*- $[Cr(N_2)_2(P^{Et}N^{2,6-F2-Bn}P^{Et})_2]$ (*trans*-1) to *cis*- $[Cr(N_2)_2(P^{Et}N^{2,6-F2-Bn}P^{Et})_2]$ (1) over 75 h.



Fig. S5. ³¹P NMR spectra collected in THF at 25 °C showing the isomerization of *cis*-[Cr(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (1) to *trans*-[Cr(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (*trans*-1) after prolonged heating of the sample at 40 °C. Trace 1: Initial ³¹P spectrum; Trace 2: after 1 day at 40 °C; Trace 3: after 2 days at 40 °C; Trace 4: after 3 days at 40 °C. The resonance at 33.6 ppm is free P^{Et}N^{2,6-F2-Bn}P^{Et} ligand.

$\frac{1}{100}$							
	ν_1	v_2	μ	λ_1	λ_2	k (mdynes/Å)	k₀ (mdynes/Å)
Cr	1990	1911	0.1428	233210.2	215061.6	15.69	0.64
Мо	2012	1950	0.1428	238395.2	223929.2	16.18	0.51
W	1987	1925	0.1428	232507.6	218224.2	15.78	0.50

Table S1. Determination of N-N force constants for 1, 2, and 3. The method and equations to obtain the values of k and k_c is described in *J. Am. Chem. Soc.* **1962**, 84, 4432-4438.



Fig. S6 Cyclic voltammograms of (a) **1**; (b) **2**; (c) **3** recorded at v = 0.10 V s⁻¹. Conditions: 0.2 M [Bu₄N][B(C₆F₅)₄] THF solution, glassy carbon working electrode, 25 °C, 0.6 mM **1**; 0.9 mM **2**; 1.4 mM **3**.

Kinetic analysis of the irreversible Cr^{1/0} wave in the cyclic voltammetry experiment

Assuming the $Cr^{1/0}$ couple of **1** is irreversible due to rapid loss of N₂ from the oxidized species, the oxidation wave can be analyzed according to an electrochemical E_rC_i mechanism (a reversible electron transfer followed by an irreversible chemical reaction). The oxidation wave of **1** is irreversible at all scan rates examined (0.1 to 20 V s⁻¹), indicating that the wave is controlled by the kinetics of the chemical step according to Equation S1.1:

$$E_{pa} = E^{\circ'} + 0.78 \frac{RT}{F} - 1.15 \frac{RT}{F} \log\left(\frac{RTk}{Fv}\right)$$
 (eq.S1.1)

where E_{pa} is the anodic peak potential (V), $E^{\circ'}$ is the formal potential of the Cr^{1/0} couple (V), *F* is the Faraday constant, *R* is the gas constant, *T* is the temperature (K), v is the scan rate (V s⁻¹), and *k* is the first-order rate constant (s⁻¹) for N₂ dissociation from Cr^{1.5,6} This equation cannot be solved accurately for **1** since two of the variables ($E^{\circ'}$ and *k*) are unknown. However, an upper limit on $E^{\circ'}$ can be estimated by assuming values of *k*. To this purpose, the value of $E_{pa} = -1.36$ V at v = 0.1 V s⁻¹ was used to generate estimated values of $E^{\circ'}$ for a range of *k* values (Table S2).

Note that the experimental E_{pa} value is also affected by *i*R drop resulting from uncompensated resistance of the 0.2 M [Bu₄N][B(C₆F₅)₄] THF electrolyte solution. In our

experiments, the Cp₂Fe^{+/0} and Cp*₂Fe^{+/0} couples displayed a peak-to-peak separation ($E_{pa}-E_{pc}$) of ~130 mV, which is much larger than the value expected for an ideal Nernstian wave (57 mV at 25 °C), as well as the values we typically observe for the Cp₂Fe^{+/0} wave in acetonitrile solvent at similar scan rates (65-70 mV). An estimate of the potential error due to *i*R drop can be made by taking the difference in $E_{pa}-E_{pc}$ of the Cp₂Fe^{+/0} couple in THF and acetonitrile and dividing by two. This conservative estimate suggests a corrected E_{pa} value of **1** is 30 mV more negative than observed, i.e. $E_{pa} = -1.39$ V at v = 0.1 V s⁻¹. Similarly, consideration of *i*R drop also results in the estimated $E^{\circ\prime}$ values in Table S2 being more negative by 30 mV, e.g. $E^{\circ\prime} = -1.12$ V when $k = 1 \times 10^9$ s⁻¹.

Table S2. Estimated Values of $E^{\circ\prime}$ for 1.^{*a*}

k (s ⁻¹)	<i>E</i> °′ (V)
1×10^3	-1.27
1×10^4	-1.24
1×10^5	-1.21
1×10^{6}	-1.18
1×10^7	-1.15
1×10^8	-1.12
1×10^{9}	-1.09

^{*a*} Calculated according to Equation S1.1 using $E_{pa} = -1.36$ V (uncorrected for *iR*-drop) and v = 0.1 V s⁻¹.

	Metal (M)	(M) mass (mg)	Acid	Acid equiv	Temp °C	equi per N	v NH4 ⁺ M atom	equiv 1 per N	$\rm NH_2 \rm NH_3^+$ M atom
						CRAFT	manual integration	CRAFT	manual integration
1	Cr (1)	2.7	HOTf	100	-40	0.13	0.13	0.28	0.34
2	Cr (1)	4.9	HOTf	100	-40	0.10	0.12	0.28	0.29
3	Cr (1)	2.0	HOTf	100	-40	0.06	0.11	0.19	0.20
4	Cr (1)	7.3	HOTf	100	-40	0.05	0.09	0.18	0.18
5	Cr (1)	3.6	HOTf	100	-40	0.06	0.12	0.20	0.20
6	Cr (1)	3.6	HOTf	100	RT	0	0	0	0
7	Cr (trans-1)	3.3	HOTf	100	-40	0.10	0.07	0.19	0.22
8	Cr (1)	1.0	H ₂ SO ₄	100	RT	0	0	0	0
9	Mo (2)	11	HOTf	100	-40	0	0	0	0
10	Mo (2)	7.4	HOTf	100	RT	0	0	0	0
11	Mo (2)	5.3	H ₂ SO ₄	100	RT	0	0	0	0
12	W (3)	2.9	HOTf	100	-40	0	0	0	0
13	W (3)	12.1	H ₂ SO ₄	100	RT	0	0	0	0
14	W (3)	15.4	HOTf	35	-40	0	0	0	0
15	W (3)	8.7	HOTf	61	-40	0	0	0	0

 Table S3. Tabulated Results for protonolysis studies of complexes 1, trans-1, 2, 3



Fig. S7. Measured ¹H NMR data (blue) and simulated CRAFT ¹H NMR data (red) of the resonances for $N_2H_5^+$ (top spectrum) and NH_4^+ (bottom spectrum).

General procedure for X-ray diffraction studies: Single crystals were selected and mounted using NVH immersion oil onto a glass fiber, a nylon fiber and cooled to the data collection temperature of 100(2) K with a stream of dry nitrogen gas. Data were collected on a Brüker-AXS Kappa APEX II CCD diffractometer with 0.71073 Å Mo-K α radiation. Unit cell parameters were obtained from 60 data frames, 0.5° ϕ , from three different sections of the Ewald sphere and complete data collection strategies were determined for each crystal using the APEX2 suite.⁷ Each data set was treated with SADABS absorption corrections based on redundant multi-scan data.⁸ The structures were solved by direct methods (XS) or intrinsic phasing (XT) and refined by least squares method on *F*² using the XL program package interfaced through OLEX2.⁹⁻¹¹ All non-hydrogen atoms were refined with anisotropic displacement parameters and all hydrogen atoms were treated as idealized contributions, unless otherwise noted. Details regarding specific solution refinement for each compound are provided below.

Refinement of 1: Data were collected on a yellow prism (0.25 x 0.20 x 0.06 mm³) grown by evaporation of a diethyl ether solution. The data set consisting of 92748 reflections (11730 unique, $R_{int} = 7.03\%$) was collected over $2\theta = 2.928$ to 60.068° . The metric symmetry and systematic absences were consistent with the monoclinic space group P2₁/*n*. The asymmetric unit contains one molecule of *cis*-[Cr(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] that is bisected by a non-crystallographic pseudo-2-fold axis parallel to the crystallographic *b* axis. The goodness of fit on *F*² was 1.027 with $R_1 = 3.87\%$ [*I*>2 σ (*I*)], *wR*₂ = 9.69% (all data) and with a largest difference peak and hole of 0.50 and -0.38 *e*/Å³.

Refinement of *trans*-1: Data were collected on an orange block (0.15 x 0.15 x 0.06 mm³) grown by rapid evaporation of a diethyl ether solution. The data set consisting of 48915 reflections (5435 unique, $R_{int} = 7.00\%$) was collected over $2\theta = 3.812$ to 59.148°. The metric symmetry and systematic absences were consistent with the monoclinic space group P2₁/*c*. The asymmetric unit contains one molecule of *trans*-[Cr(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂]. The goodness of fit on F^2 was 1.024with $R_1 = 3.44\%$ [*I*>2 σ (*I*)], $wR_2 = 8.30\%$ (all data) and with a largest difference peak and hole of 0.37 and -0.35 *e*/Å³. Refinement of **2**: Data were collected on a yellow shard (0.15 x 0.10 x 0.08 mm³) grown by evaporation of a diethyl ether solution. The data set consisting of 68516 reflections (19824 unique, $R_{int} = 8.37\%$) was collected over $2\theta = 2.92$ to 56.566°. The metric symmetry and systematic absences were consistent with the monoclinic space group Pn. The asymmetric unit contains two molecules of *cis*-[Mo(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂], which are nearly related by translation along the crystallographic *b* axis, but differ in the orientation of an ethyl group. A non-zero-Flack parameter indicated racemic twinning, and the data were treated as such with a freely refined twin ratio converging on ~78:22. The goodness of fit on *F*² was 1.003 with $R_1 = 5.28\%$ [*I*>2 σ (*I*)], $wR_2 = 10.71\%$ (all data) and with a largest difference peak and hole of 1.03 and -0.44 $e/Å^3$. The largest residuals are near the heavy Mo atoms.

Refinement of **3**: Data were collected on a yellow block (0.23 x 0.22 x 0.15 mm³) grown by evaporation of a diethyl ether solution. The data set consisting of 206157 reflections (39026 unique, $R_{int} = 5.03\%$) was collected over $2\theta = 2.922$ to 72.636° . The metric symmetry and systematic absences were consistent with the monoclinic space group P*n*. The asymmetric unit contains two molecules of *cis*-[W(N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂], which are nearly related by translation along the crystallographic *b* axis, but differ in the orientation of an ethyl group. A non-zero-Flack parameter indicated racemic twinning, and the data were treated as such with a freely refined twin ratio converging on ~52:48. The goodness of fit on *F*² was 1.043 with $R_1 = 3.12\%$ [*I*>2 σ (*I*)], $wR_2 = 6.30\%$ (all data) and with a largest difference peak and hole of 5.08 and -1.28 *e*/Å³. The large residuals are near the heavy W atoms.

	1	trans-1	2	
Formula	$C_{34}H_{58}CrF_4N_6P_4$	$C_{34}H_{58}CrF_4N_6P_4$	C34H58M0F4N6P4	
Crystal System	Monoclinic	Monoclinic	Monoclinic	
Space Group	$P2_1/n$	$P2_1/c$	Pn	
<i>a</i> , Å	10.3010(2)	10.8985(6)	10.2417(2)	
b, Å	18.8047(4)	13.4410(7)	18.3288(3)	
<i>c</i> , Å	20.7192(4)	13.4741(7)	21.5881(4)	
a, deg	90	90	90	
β, deg	93.7700(10)	101.392(3)	95.3341(11)	
γ, deg	90	90	90	
V, Å ³	4004.77(14)	1934.89(18)	4034.93(13)	
Ζ	4	2	4	
Radiation (λ, Å)	Μο-Κα, 0.71073	Μο-Κα, 0.71073	Μο-Κα, 0.71073	
ρ (calcd.), g/cm ³	1.331	1.378	1.394	
μ , mm ⁻¹	0.496	0.514	0.534	
Temp, K	100.0	100.0	100.0	
Size, mm ³	0.25 x 0.2 x 0.06	0.15 x 0.15 x 0.06	0.15 x 0.1 x 0.08	
Color	Yellow	Orange	Yellow	
Habit	Prism	Block	Irregular	
No. Reflections	92748	48915	68516	
No. Ind. Ref. (R _{int})	11730 (0.0703)	5435 (0.0700)	19824 (0.0837)	
2θ range, deg	2.928 - 60.068	3.812 - 59.148	2.92 - 56.566	
R_1	0.0387	0.0344	0.0528	
wR_2	0.0859	0.0743	0.0908	
$GOF(F^2)$	1.027	1.024	1.003	
F(000)	1696.0	848.0	1768.0	

Table S4. Crystallographic data for complexes 1, *trans*-1, and 2.

	3
Formula	C34H58WF4N6P4
Crystal System	Monoclinic
Space Group	Pn
<i>a</i> , Å	10.2469(2)
<i>b</i> , Å	18.3292(3)
<i>c</i> , Å	21.5498(4)
α, deg	90
β, deg	95.4260(10)
γ, deg	90
V, Å ³	4029.29(13)
Ζ	4
Radiation (λ, Å)	Μο-Κα, 0.71073
ρ (calcd.), g/cm ³	1.541
μ, mm ⁻¹	3.077
Temp, K	100
Size, mm ³	0.23 x 0.22 x 0.15
Color	Yellow
Habit	Block
No. Reflections	206157
No. Ind. Ref. (R _{int})	39026 (0.0503)
20 range, deg	2.922 - 72.636
R_{I}	0.0312
wR_2	0.0601
$GOF(F^2)$	1.043
F(000)	1896.0

Table S5. Crystallographic data for complex 3.

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Fig. S8. ¹H NMR spectrum of **1** recorded in THF- d_8 .



Fig. S9. ³¹P{¹H} NMR spectrum of 1 recorded in THF- d_8 .



Fig. S10. ¹⁵N{¹H} NMR spectrum of 1^{15N} recorded in THF- d_8 .



Fig. S11. ¹H NMR spectrum of 2 recorded in THF- d_8 .



Fig. S12. ³¹P{¹H} NMR spectrum of 2 recorded in THF- d_8 .



Fig. S13. ¹⁵N{¹H} NMR spectrum of 2^{15N} recorded in THF- d_8 .



Fig. S14. ¹H NMR spectrum of **3** recorded in THF- d_8 .



Fig. S15. ³¹P $\{^{1}H\}$ NMR spectrum of **3** recorded in THF-*d*₈.



Fig. S16. ¹⁵N{¹H} NMR spectrum of 3^{15N} recorded in THF- d_8 .



Fig. S17. ¹H-¹⁵N HSQC NMR spectrum recorded in THF- d_8 at -40 °C from the addition of HOTf to *cis*-[Cr(¹⁵N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (**1**^{15N}). The cross-peak 10.8/-326 corresponds to ¹⁵N₂H₅⁺. The cross-peak 7.0/-364 corresponds to ¹⁵NH₄⁺.



Fig. S18. ¹H NMR spectrum recorded in THF- d_8 at -40 °C from the addition of HOTf to *cis*-[Cr(¹⁵N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (1^{15N}), showing the doublet for ¹⁵N₂H₅⁺ (J_{NH} = 75 Hz). The doublet for ¹⁵NH₄⁺ at 7.0 ppm is obscured by aromatic resonances of PNP ligand.



Fig. S19. ¹⁵N {¹H} NMR spectrum recorded in THF- d_8 at -40 °C from the addition of HOTf to cis-[Cr(¹⁵N₂)₂(P^{Et}N^{2,6-F2-Bn}P^{Et})₂] (1^{15N}), showing the singlet for ¹⁵N₂H₅⁺ at -326 ppm. The singlet for ¹⁵NH₄⁺ is at -364 ppm.