# 4-Methylcoumarin-bridged Fluorescent Responsive Cryptand: from [2+2]Photodimerization to Supramolecular Polymer

Ming Cheng,<sup>a</sup> Chenhao Yao,<sup>a</sup> Yihan Cao,<sup>a</sup> Qi Wang,<sup>a</sup> Yi Pan,<sup>b</sup> Juli Jiang,<sup>\*a</sup> and Leyong Wang<sup>\*a</sup>

<sup>*a*</sup> Key Laboratory of Mesoscopic Chemistry of MOE, School of Chemistry and Chemical Engineering, Nanjing University, Nanjing 210093, China, Fax: (+86) 25-83597090; <sup>*b*</sup> State Key Laboratory of Coordination Chemistry, School of Chemistry and Chemical Engineering, Nanjing University, Nanjing 210093, China \*Corresponding Author: jjl@nju.edu.cn; lywang@nju.edu.cn

# SUPPORTING INFORMATION

### **Table of Contents**

1. Materials and methods	··· <b>§</b> 2
2. Experimental procedures	··· <del>§</del> 2
3. Association constants and Job plots of $1 \supset 2$	••• <del>\$</del> 7
4. Concentration-dependent <sup>1</sup> H NMR spectroscopy of $3 \supset 4$	• <del>S</del> 10
5. X-ray crystal data for cryptand 1 ·····	• <del>\$</del> 10
6. References ······	• <del>\$</del> 11

#### 1. Materials and methods

All reactions were performed in open atmosphere unless otherwise stated. All reagents, unless otherwise indicated, were obtained from commercial sources. Anhydrous  $CH_3CN$  was obtained by distillation from  $CaH_2$  under  $N_2$  atmosphere. Melting points (M.p.) were determined using a Focus X-4 apparatus and were not corrected. All yields were given as isolated yields. NMR spectra were recorded on a Bruker DPX 300 MHz or 400 MHz spectrometer with internal standard tetramethylsilane (TMS) and solvent signals as internal references, and the chemical shifts ( $\delta$ ) were expressed in ppm and *J* values were given in Hz. Low-resolution electrospray ionization mass spectra (LR-ESI-MS) were obtained on Finnigan MatTSQ 7000 instruments. High-resolution electrospray ionization mass spectra (HR-ESI-MS) were recorded on an Agilent 6540Q-TOF LCMS equipped with an electrospray ionization (ESI) probe operating in positive-ion mode with direct infusion.

#### 2. Experimental procedures.



Scheme S1. Synthesis of cryptand 1 and dimer-cryptand 3.

General procedure for the synthesis of cryptand 1: A mixture of A<sup>1</sup> (0.20 g, 0.28 mmol) and

compound **B** (0.053 g, 0.28 mmol) in anhydrous CH<sub>3</sub>CN was stirred and refluxed for 24 h. The reaction mixture was filtered. The filtrate was removed in vacuum and the residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub>. The organic phase was washed with water, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated under vacuum. The residue was purified by silica-gel column chromatography using (CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH, 100:1) to afford **1** (0.063 g, 30.0%) as a white solid. M.p.178-180 °C ; The <sup>1</sup>H NMR & <sup>13</sup>C NMR spectra of **1** are shown in Fig. S1-S2. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN, 298K) *δ* (ppm): 7.23 (s, 1H), 7.04 (s, 1H), 6.57-6.56 (m, 4H), 6.45-6.42 (m, 2H), 6.15 (s, 1H), 4.89 (s, 4H), 4.13-3.93 (m, 8H), 3.88-3.67 (m, 8H), 3.58-3.54 (m, 16H), 2.42 (s, 3H).; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>, 298 K) *δ* (ppm): 161.4, 160.0, 159.9, 153.5, 152.4, 150.3, 145.9, 138.9, 137.6, 113.2, 112.6, 111.6, 106.2, 105.8, 102.4, 102.1, 102.0, 73.6, 71.5, 71.1, 70.7, 69.8, 69.7, 67.6, 67.4.; LR-ESI-MS: calcd. for [**1** + H]<sup>+</sup>: 753.31, found *m*/*z* = 775.20; [**1** + NH<sub>4</sub>]<sup>+</sup>: 775.29, found *m*/*z* = 775.15; HR-MS (ESI): calcd. for [**1** + H]<sup>+</sup>: 753.317, found *m*/*z* = 770.3387; [**1** + Na]<sup>+</sup>: 775.2936, found *m*/*z* = 775.2940.

General procedure for the synthesis of dimer-cryptand **3**: Cryptand **1** (0.7 g, 0.92 mmol) was dissolved in 30 mL dichloromethane. Then the solution was exposed to a 500 W mercury lamp for 96 h. The residue was purified by silica-gel column chromatography using (CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH, 50:1) to afford **3** (0.030 g, 4.3%) as a white solid. M.p.93-95 °C ; The <sup>1</sup>H NMR & <sup>13</sup>C NMR spectra of **3** are shown in Fig. S5-S6. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN, 298K)  $\delta$  (ppm): 7.00 (s, 2H), 6.85 (s, 2H), 6.65-6.49 (m, 8H), 6.44-6.40 (m, 4H), 4.89 (d, *J* = 19.6 Hz, 8H), 4.05-3.92 (m, 16H), 3.71 (br, 16H), 3.55 (br, 32H), 3.42 (s, 2H), 1.36 (s, 6H).; <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>CN, 298 K)  $\delta$  (ppm): 165.9, 159.7, 159.6, 148.9, 145.7, 144.4, 138.6, 138.2, 114.4, 111.6, 107.6, 107.5, 107.5, 107.2, 102.1, 100.3, 100.2, 71.5, 71.0, 70.1, 69.9, 68.9, 67.3, 67.2, 46.7, 44.2, 25.5..; LR-ESI-MS: calcd. for [**3** + H]<sup>+</sup>: 1523.63, found *m*/*z* = 1523.30; [**3** + Na]<sup>+</sup>: 1527.60, found *m*/*z* = 1527.5980, found *m*/*z* = 1527.5983.

## $\begin{array}{c} -7.23\\ 6.57\\ 6.57\\ 6.56$





PositiveLine#:1 R.Time:0.200(Scan#:13) MassPeaks:97(Positive) Spectrum Mode:Averaged 0.133-0.267(9-17) BasePeak:775.15(1403427) BG Mode:Averaged 0.033-0.533(3-33) Segment 1 - Event 1 Intensity

0.

 


Fig. S4 HR-ESI-MS of cryptand 1

Э  



Fig. S6  $^{13}$ C NMR spectrum (100 MHz, CD<sub>3</sub>CN, 298 K) of dimer-cryptand 3.



Fig. S8 HR-ESI-MS of dimer-cryptand 3.

#### **3.** Association constants and Job plots of $1 \supset 2$ .





**Fig. S9** Partial <sup>1</sup>H NMR spectra (400 MHz, CD<sub>3</sub>CN, 298 K): (a) **2**; (b) 10.00 mM **1** and 10.00 mM **2**; (c) **1**.



**Fig. S10** Partial <sup>1</sup>H NMR spectra changes (300 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>CN = 1/1(v/v), 298 K) of **2** (guest, 2.00 mM) upon addition of **1** (host): (a) 0.00, (b) 0.40, (c) 0.80, (d) 1.20, (e) 1.60, (f) 2.00, (g) 3.00, (h) 5.00, (i) 7.00, (j) 9.00, (k) 13.00, (l) 17.00, (m) 20.00, (n) 24.00, (o) 30.00, (p)36.00, (q)40.00 mM.



**Fig. S11** The non-linear curve-fitting (NMR titrations) for the complexation of **2** (guest) with **1** (host) in CDCl<sub>3</sub>/CD<sub>3</sub>CN = 1/1 (v/v) at 298 K. Using the signal of **2** at  $\delta$  8.8281 (proton H<sub>2a</sub>) as the reference. The association constant ( $K_a$ ) of **2**  $\subset$  **1** in CDCl<sub>3</sub>/CD<sub>3</sub>CN = 1/1(v/v) was estimated to be about (4.5  $\pm$ 0.7)  $\times$  10<sup>3</sup> M<sup>-1</sup>.



**Fig. S12** The Job Plot (NMR titrations) for the complexation of **1** (host) with **2** (guest) in  $CD_3CN$  at 298 K. ([H] + [G] = 8 mM).

# 4. Concentration-dependent <sup>1</sup>H NMR spectroscopy of $3 \supset 4$ .



**Fig. S13** <sup>1</sup>H NMR spectra (400 MHz, 298 K) of (a) individual **4** (CDCl<sub>3</sub>/CD<sub>3</sub>CN, 1/1, v/v); (b) individual **3** (CD<sub>3</sub>CN); mixtures of **3** and **4** in a 1 : 1 molar ratio at different **3** concentrations (CDCl<sub>3</sub>/CD<sub>3</sub>CN, 1/1, v/v): (c) 5, (d) 10, (e) 20, (f) 40, (g) 60, (h) 74 mM

#### 5. X-ray crystal data for cryptand 1.

CCDC number	1449565
Empirical formula	C <sub>40</sub> H <sub>48</sub> O <sub>14</sub>
Formula weight	752.81
Temperature	296(2)
Wavelength	0.71073 Å
Crystal system	Triclinic
Space group	P-1
a	13.5950(13) Å
b	20.8235(19) Å
c	23.687(2) Å

**Table 1.** Crystal data and structure refinement for cryptand 1.

α	74.639(2) °
β	84.656(2) °
γ	76.364(2) °
Volume	6280.4(10) Å <sup>3</sup>
Ζ	4
Density (calculated)	1.300
Absorption coefficient	0.164
F(000)	2552
Crystal size	$0.26 \times 0.24 \times 0.22 \text{ mm}^3$
Theta range for data collection	2.03 to 25.01 °
Index ranges	$-16 \le h \le 11, -24 \le k \le 24,$ $-28 \le 1 \le 28$
Reflections collected	39583
Independent reflections	21877 [R(int) = 0.0365]
Completeness to theta = $25.01^{\circ}$	98.7%
Absorption correction	multi-scan
Refinement method	Full-matrix least-squares on $F^2$
Goodness-of-fit on F2	1.184
Final R indices $[I > 2 \text{sigma}(I)]$	R1 = 0.0893, wR2 = 0.1740
<i>R</i> indices (all data)	R1 = 0.1375, wR2 = 0.1904
Largest diff. peak and hole	1.440 and -1.021 e Å <sup>-3</sup>

## 6. References

1. X. Ji, M. Zhang, X. Yan, J. Li and F. Huang, *Chem. Commun.*, 2013, **49**, 1178-1180.